

Exuberant endothelial C5b-9 formation in recurrent and de-novo post-transplant TMA

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Supplementary material :

- Supplementary methods;
- Supplementary results;
- Supplementary references;
- Supplementary Table 1: Characteristics and clinical parameters of aHUS patients with recurrence after kidney transplant at the time of sample collection;
- Supplementary Table 2: Characteristics and clinical parameters of transplanted aHUS patients with no sign of recurrence at the time of sample collection;
- Supplementary Table 3: Characteristics and clinical parameters of patients with de-novo TMA at the time of sample collection;
- Supplementary Table 4: Characteristics and clinical parameters of aHUS patients in chronic dialysis at the time of sample collection;
- Supplementary Table 5: Characteristics, clinical and complement parameters of stable renal transplant patients at the time of sample collection;
- Supplementary Table 6: Characteristics, clinical and complement parameters of control dialyzed patients at the time of sample collection;
- Supplementary Table 7: Complement parameters and genetic characteristics of aHUS patients with recurrence after kidney transplant at the time of sample collection;
- Supplementary Table 8: Complement parameters and genetic characteristics of transplanted aHUS patients with no sign of recurrence at the time of sample collection;
- Supplementary Table 9: Complement parameters and genetic characteristics of patients with de-novo TMA at the time of sample collection;
- Supplementary Table 10: Complement parameters and genetic characteristics of aHUS patients in chronic dialysis at the time of sample collection;
- Appendix - International Registry of Recurrent and Familial HUS-TTP.

Supplementary Methods

Study participants

We evaluated three groups of patients: 1) 23 aHUS patients recipients of a kidney transplant of which 12 with recurrence and 11 with no sign of recurrence; 2) 22 transplant recipients who had other diseases in the native kidneys of which 15 developed de-novo post-transplant TMA and 7 with stable graft function; and 3) 39 patients on dialysis, including 28 patients with aHUS (4 of them were also studied after transplant) and 11 non-aHUS patients (Table 1 and Supplementary Tables S1-6) .

Patients and controls consented to participate in the “Mario Negri”, Biobank “Malattie Rare e Malattie Renali”. The patients’ main characteristics are shown in Table 1 (individual values are detailed in Suppl. Tables 1-6).

Participants were patients with aHUS or de-novo TMA post-kidney transplant, included in the International Registry of Recurrent and Familial Haemolytic Uremic Syndrome/Thrombotic Thrombocytopenic Purpura (HUS/TTP), who were analysed with the serum-induced C5b-9 formation assay in our laboratories between 2014 and 2023. The Registry was established in 1996 at the Aldo e Cele Daccò Clinical Research Center for Rare Diseases (Ranica, Bergamo) (villacamozzi.marionegri.it/seu).

Inclusion criteria were diagnosis of aHUS, or de novo post-transplant TMA; analysis with the serum-induced C5b-9 formation assay on HMEC-1; kidney transplant or renal replacement therapy; no C5-inhibitors for at least 8 weeks prior to the time of serum collection; informed consent to participate in the “Malattie Rare e Malattie Renali” Biobank.

aHUS was diagnosed in patients who have had one or more episodes of microangiopathic hemolytic anemia and thrombocytopenia, with hematocrit (Ht) <30%, hemoglobin (Hb) <10 g/dL, serum lactate dehydrogenase (LDH) >500 IU/L, undetectable haptoglobin, fragmented erythrocytes in the peripheral blood smear, and platelet count <150,000/ μ L, associated with acute renal failure (serum creatinine >1.3 mg/dL for adults, >0.5 mg/dL for children below 5 years of age and >0.8 mg/dL for

children aged 5-10, and/or urinary protein/creatinine ratio >200 mg/g; or an increase in serum creatinine or a urinary protein/creatinine ratio >15% compared to baseline levels).⁶ Recurrence in the graft in patients with a history of aHUS in the native kidneys was defined as exhibiting hematological and/or histologic markers of TMA: glomerular and/or arteriolar microthrombi, glomerular fibrinoid necrosis, amorphous material and endothelial swelling in glomerular capillaries and arterioles, glomerular fibrin, thickening and double contours of the glomerular basement membrane, and fragmented erythrocytes in microvessels and/or the interstitium.¹

De-novo post-transplant TMA was defined based on the following criteria: hematological and/or histologic markers of TMA in kidney transplant patients who never had any evidence of TMA before transplantation. Diagnoses were made by the physicians who referred the patients for the C5b-9 assay and were revised by an independent clinician at the Clinical Research Center for Rare Diseases (E.B.). Only patients with a confirmed diagnosis were included in the study.

Patients had not taken C5 inhibitory drugs for at least 8 weeks prior to blood draws.

TTP was ruled out on the basis of ADAMTS13 activity >10% and no anti-ADAMTS13 antibodies. Stx-*E.Coli* infection was ruled out based on negative assays for *stx* and *eae* genes (by PCR) or Shiga-toxin (Vero cell assay) in the stool and/or anti-Shiga-toxin antibodies (ELISA) and/or LPS O157, O26, O111, or O145 (ELISA) in serum.

Full remission was defined as the normalisation of both hematological parameters (Ht >30%; Hb >10 g/dL; LDH <500 IU/L; platelets >150,000/ μ L) and renal function (serum creatinine <1.3 mg/dL for adults, <0.5 mg/dL for children below 5 years of age and <0.8 mg/dL for children aged 5-10; urinary protein/creatinine ratio <200 mg/g). Hematological remission in patients was defined as the normalisation of hematological parameters but with residual renal dysfunction.

The protocol was approved by the Ethical Committee of the Azienda Sanitaria Locale Bergamo, Italy.

Genetic and biochemical complement abnormalities and anti-CFH autoantibodies

The screening of coding sequences of aHUS-associated genes (*CFH*, *CFHR1*, *MCP*, *CFI*, *CFB*, *C3*, *THBD* and *DGKE*) and candidate genes (*CFHR2*, *CFHR3*, *CFHR4*, *CFHR5*, and *C5*) was performed using amplicon-based next-generation sequencing (S1). Rare functional variants (missense, nonsense, indel, or splicing variants) with minor allele frequency (MAF) <0.001 in the Genome Aggregation Database (gnomAD, <https://gnomad.broadinstitute.org/>) were selected. Stop-gain, frameshift and splicing variants, and missense variants with published functional studies, were categorised as pathogenic variants (PV). The other variants were categorised as likely pathogenic variants (LPV), variants of uncertain significance (VUS), likely benign or benign, using guidelines from the American College of Medical Genetics and Genomics (ACMG) and from the KDIGO conference on aHUS and C3G (S2-S4). We also used the Combined Annotation Dependent depletion tool (CADD), version 1.6, which is a tool for scoring the deleteriousness of single nucleotide variants as well as insertion/deletion variants, and which integrates multiple annotations into one metric (<https://cadd.gs.washington.edu/info>) (S5). We selected the PHRED-scaled C-score ranking. We searched for genomic abnormalities that affect the *CFH* and *CFHR* genes (S6) using multiplex ligation-dependent probe amplification, as reported (S1). We also genotyped the *CFH* single-nucleotide polymorphisms (SNPs) (c.1-331C>T, rs3753394; c.184G>A, p.V62I, rs800292; c.1204C>T, p.H402Y, rs1061170; c.2016A>G, p.Q672Q, rs3753396; c.2237-543G>A, rs1410996; c.2808G>T, p.E936D, rs1065489) that define the disease risk haplotype *CFH*_{TGTGGT} (known as *CFH*-H3 haplotype) and one SNP in *MCP* (rs7144, c.*897T>C) that tags the risk *MCP*_{GGAAC} haplotype (S7-S8). Anti-FH autoantibodies were measured in plasma using ELISA (S9).

Complement C3 was measured using nephelometry. Normal ranges (defined as mean±2SD) for C3: 83-180 mg/dL, n=50. SC5b-9 levels were evaluated in EDTA plasma using MicroVue sC5b-9 Plus EIA (SC5b-9 Plus; Quidel). The normal range of plasma sC5b-9 in our laboratory is 127-400 ng/mL; n=50.

Serum-induced C5b-9 deposition on HMEC-1

The assay was performed as described previously,² with minor modifications. The human microvascular endothelial cells (HMEC-1 cell line) were a gift from Dr Edwin Ades and Francisco J. Candal of the CDC and Dr Thomas Lawley of Emory University, Atlanta, GA. The growth medium we used is MCDB 131 (Gibco, Grand Island, NY) supplemented with 10% foetal bovine serum (Gibco), 10 µg/mL hydrocortisone, 100 U/mL penicillin, 100 µg/mL streptomycin, 2 mM glutamine (Gibco), and 50 µg/mL endothelial cell growth factor. HMEC-1 (about 75,000 cells) were plated on glass coverslips and used when confluent. Cells were washed 3 times with test medium (HBSS: 137 mM NaCl, 5.4 mM KCl, 0.7 mM Na₂HPO₄, 0.73 mM KH₂PO₄, 1.9 mM CaCl₂, 0.8 mM MgSO₄, 28 mM Trizma base pH 7.3, 0.1% dextrose; with 0.5% BSA) and activated with 10 µM ADP (Sigma) for 10 minutes. Thereafter, cells were exposed for 2 hours to serum from patients or healthy subjects, 50% in test medium (HBSS with 0.5% BSA). HMEC-1 were then fixed in 3% paraformaldehyde and stained with rabbit anti-human complement C5b-9 complex antibody (Calbiochem) followed by FITC-conjugated secondary antibody (Jackson Immuno Research Laboratories).

Sera from 10 controls were pooled and the pool was run in each experiment as a reference (100%) for C5b-9 staining. We also analysed single sera from additional controls and calculated the percentages of C5b-9 deposits vs. control serum pool for each to set the normal range (mean±2SD of the percentage of C5b-9 deposits of the single control sera vs. control serum pool: ADP-activated HMEC-1, n=35 control sera, 60-149%; unstimulated HMEC-1, n=22 control sera, 50-150%). We verified the integrity of the cell monolayer after exposure to serum samples in parallel slides in which cells were stained with May-Grunwald Giemsa.

The fluorescent staining on the endothelial cell surface was acquired using the Apotome Axio Imager Z2 (Zeiss) microscope. Fifteen fields, which were systematically digitised along the surface,

were acquired using a computer-based image analysis system. The area occupied by the fluorescent staining was evaluated with automatic edge detection using the built-in specific functions of the software Image J (NIH, Bethesda, MD), and calculated as pixel² per field analysed.² For each sample, 15 fields were analysed; the highest and the lowest values were discarded and the mean of the other 13 fields was calculated. We expressed the results as the percentage of the staining compared to the control serum pool, run in parallel.²

Statistical analysis

Normality of distributions was visually inspected by box-plot representation and formally assessed with the D'Agostino-Pearson test. Between-groups differences were analysed using the t test, Mann-Whitney U test, or Fisher exact test, as appropriate. All data were analysed using MedCalc 10.0.1 statistical software. P values of less than 0.05 were considered statistically significant. Positive predictive values of serum induced C5b-9 assay were calculated as “true positive values / true positive + false positive values”; negative predictive values were calculated as “true negative values / true negative + false negative values”.

Supplementary results

Calculations and results of positive and negative predictive values for either recurrent aHUS or de-novo TMA are as follows:

aHUS Recurrence vs aHUS No Recurrence			
	aHUS-r	aHUS-nr	
Positive results	11	1	
Negative results	1	10	
Positive predictive value: 11 / (11+1): 0.916			
Negative predictive value: 10 / (10+1): 0.909			

De-Novo TMA vs Stable TX			
	De Novo	Stable	
Positive results	15	1	
Negative results	0	6	
Positive predictive value: 15 / (15+1): 0.937			
Negative predictive value: 6 / (6+0): 1			

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Supplementary Table 1. Characteristics and clinical parameters of aHUS patients with recurrence after kidney trasplant at the time of sample collection.

patient (n°)	gender	age (years)	native kidney disease	phase	time from trasplant (days)	kidney donor	immunosuppression therapy	hystology	eculizumab* (response)	platelets	LDH	hemoglobin	haptoglobin	schistocytes	C3	C4	serum creatinin
										150-400x10 ³ /μL	266-500 IU/L	14-18 g/dL	30-200 mg/dL		70-152 mg/dL	15-40 mg/dL	0.55 - 1.25 mg/dL
patient 1	M	45	aHUS	acute	3	cadaver	TA - MMF - ST	TMA	YES (CR)	108	1240	9.6	67	n.a.	102	32	3.5
patient 2	M	26	aHUS	acute	1	cadaver	TA - MMF - ST	TMA	YES (CR)	52	1060	13.5	<30	YES	72	22	9.56
patient 3	M	48	aHUS	acute	9189	cadaver	TA - MMF - ST	n.a.	NO	102	165	10.1	40	n.a.	46	15	2.5
patient 4	M	72	aHUS	acute	4227	n.a.	TA - MMF - ST	n.a.	NO	234	622	9	<7	YES	62	n.a.	4.4
patient 5	M	48	aHUS	acute	9	cadaver	ST	n.a.	NO	160	435	8.5	53	n.a.	n.a.	n.a.	4.97
patient 6	F	48	aHUS	acute - post pex	5	cadaver	TA - MMF - ST	n.a.	NO	37	1977	10.4	<10	n.a.	79	18	6
patient 7	F	29	aHUS	acute	69	n.a.	sirolimus - ST	n.a.	NO	107	309	9.4	40	n.a.	n.a.	n.a.	2.91
patient 8	F	35	aHUS	acute	2560	cadaver	TA - MMF - ST	TMA	NO	170	532	9	95	n.a.	77	15	1.5
patient 9	M	56	aHUS	acute	121	cadaver	TA - ST	n.a.	n.a.	90	567	8.7	n.a.	n.a.	n.a.	n.a.	2.2
patient 10	M	61	aHUS	acute	2920	cadaver	TA - ST	n.a.	NO	161	567	10.7	15	n.a.	65	12	1.9
patient 11	F	45	aHUS	acute	2561	cadaver	TA - MMF - ST	TMA	YES (HR)	275	372	9.4	67	YES	n.a.	n.a.	6.3
patient 12	M	48	aHUS	acute	135	cadaver	TA - MMF	TMA	NO	100	607	9.3	<1	YES	109	23	3.3

n.a. not available. TA, Tacrolimus. MMF, mycophenolate. ST, steroids. Pex, plasma exchange. CR, complete remission. HR, hematological remission.

* treatment started after blood sample collection.

Supplementary Table 2. Characteristics and clinical parameters of transplanted aHUS patients with no sign of recurrence at the time of sample collection.

patient (n°)	gender	age (years)	native kidney disease	phase	time from trasplant (days)	kidney donor	immunosuppression therapy	hystology	platelets 150-400x10 ³ /μL	LDH 266-500 IU/L	hemoglobin 14-18 g/dL	schistocytes	C3 83-180 mg/dL	C4 15-40 mg/dL	serum creatinin 0.55-1.25 mg/dL
patient 13	F	29	aHUS	remission	1090	cadaver	TA - MMF	n.a.	172	n.a.	11.6	n.a.	n.a.	n.a.	0.76
patient 14	F	39	aHUS	remission	2289	cadaver	TA - MMF - ST	n.a.	169	201	13	n.a.	105	27	2.07
patient 15	F	18	aHUS	remission	1585	n.a.	n.a.	n.a.	n.a.	115	n.a.	n.a.	84	25	n.a.
patient 16	F	33	aHUS	remission	1717	cadaver	TA - MMF - ST	n.a.	440	n.a.	12	n.a.	n.a.	n.a.	1
patient 17	F	60	aHUS	remission	2057	cadaver	TA - MMF - ST	n.a.	215	n.a.	12.2	n.a.	112	54	2.23
patient 18	F	56	aHUS	remission	7	cadaver	TA - MMF - ST	n.a.	155	n.a.	13	n.a.	n.a.	n.a.	1.41
patient 19	F	44	aHUS	remission	506	cadaver	TA -MMF - ST	n.a.	355	n.a.	12.4	n.a.	n.a.	n.a.	1.53
patient 20	M	75	aHUS	remission	4383	cadaver	TA - MMF - ST	n.a.	150	n.a.	15	n.a.	125	41	1.5
patient 21	M	45	aHUS	remission	1167	cadaver	TA -MMF	MPGN	275	372	12.4	NO	21	n.a.	2
patient 22	M	13	aHUS	remission	2728	cadaver	TA - MMF - ST	chronic rejection	222	327	6.9	n.a.	106	54	3.85
patient 23	F	43	aHUS	remission	1040	cadaver	n.a.	n.a.	275	327	15	n.a.	78.2	35.4	0.8

n.a. not available. TA, Tacrolimus. MMF, mycophenolate. ST, steroids. MPGN, membrano-proliferative-glomerulonephritis.

Supplementary Table 3. Characteristics and clinical parameters of patients with de-novo TMA at the time of sample collection.

patient (n°)	gender	age (years)	native kidney disease	phase	time from trasplant (days)	kidney donor	immunosuppression therapy	hystology	eculizumab* response	platelets 150-400x10 ³ /μL	LDH 266-500 IU/L	hemoglobin 14-18 g/dL	haptoglobin 30-200 mg/dL	schistocytes	C3 70-152 mg/dL	C4 15-40 mg/dL	serum creatinin 0.55 - 1.25 mg/dL
patient 59	F	63	pyelonephritis	acute	4	cadaver	TA - MMF - ST	TMA	YES (HR)	150	620	11	97	n.a.	104	26	5.6
patient 60	M	65	IgA N	acute	96	cadaver	TA - MMF - ST	TMA	NO	93	628	9.3	<20	n.a.	84	21	1.76
patient 61	F	64	Good-pasture	acute	1	cadaver	TA - MMF - ST	TMA	YES (CR)	133	1162	13.6	45	YES	n.a.	n.a.	7
patient 62	M	59	ADPKD	acute	12	cadaver	TA - MMF - ST	TMA	YES (HR)	178	751	7.7	<30	YES	n.a.	n.a.	7.4
patient 63	F	66	hypertension	acute	4	cadaver	TA - MMF - ST	n.a.	YES (HR)	36	1746	7.9	<30	n.a.	n.a.	n.a.	9
patient 64	M	59	diabetic N	acute	8	cadaver	TA - MMF - ST	n.a.	YES (CR)	79	317	8.7	<30	YES	n.a.	n.a.	4.28
patient 65	M	58	obstructive N	acute	8	cadaver	MMF - ST	n.a.	NO	90	1249	9.3	<20	n.a.	104	27	3.48
patient 66	M	81	tubulo-interstitial N	acute	6047	cadaver	TA - MMF - ST	n.a.	YES (NR)	98	506	9.4	<5	YES	59	11	1.4
patient 67	M	51	IgA N	acute	6595	cadaver	TA - ST	n.a.	n.a.	110	n.a.	n.a.	n.a.	n.a.	72	26	4.55
patient 68	M	60	ANCA-vasculitis	acute	3	cadaver	TA - MMF - ST	TMA	YES (CR)	70	1790	7.7	<30	n.a.	n.a.	n.a.	11
patient 69	F	46	lupus N	acute	5	cadaver	TA - MMF - ST	TMA	YES (CR)	76	369	7.5	<30	YES	81	35	5.93
patient 70	M	60	ADPKD	acute	3	cadaver	TA - MMF - ST	TMA	YES (HR)	71	951	9	49	YES	n.a.	n.a.	9.4
patient 71	M	57	PAH / CRI	acute	1497	n.a.	n.a.	n.a.	NO	95	624	10	<1	n.a.	79.5	20.5	5.3
patient 72	M	73	IgA N	acute	3	cadaver	TA - MMF - ST	n.a.	YES (CR)	39	627	8.3	<30	YES	n.a.	n.a.	4.35
patient 73	M	55	diabetic N	acute	3	cadaver	TA -MMF - ST	TMA	n.a.	63	n.a.	11.9	<30	n.a.	n.a.	n.a.	12.4

n.a. not available. TA, Tacrolimus. MMF, mycophenolate. ST, steroids. N, nephropathy. ADPKD, autosomal dominant polycystic kidney disease. PAH/CRI, pulmonary arterial hypertension/chronic renal insufficiency.

* treatment started after blood sample collection. CR, complete remission. HR, hematological remission. NR, no remission.

Supplementary Table 4. Characteristics and clinical parameters of aHUS patients in chronic dialysis at the time of sample collection.

patient (n°)	gender	age (years)	native kidney disease	dialysis	platelets 150-400x10 ³ /μL	LDH 266-500 IU/L	hemoglobin 14-18 g/dL	haptoglobin 30-200 mg/dL	C3 70-152 mg/dL	C4 15-40 mg/dL	serum creatinin 0.55 - 1.25 mg/dL
patient 24	F	74	aHUS	HD	302	488	14.6	n.a.	105	35.4	3.81
patient 25	F	39	aHUS	HD	163	653	10.4	n.a.	n.a.	n.a.	4.59
patient 22	M	14	aHUS	HD	150	611	8	n.a.	67	34	4.15
patient 26	F	35	aHUS	HD	302	303	11.5	n.a.	47.7	27.5	8.9
patient 27	M	51	aHUS	PD	188	489	13.5	232	111	41	14.9
patient 28	F	43	aHUS	HD	177	277	8	91	79	25	9.95
patient 29	F	27	aHUS	HD	228	309	12.7	n.a.	45	20	11
patient 30	F	54	aHUS	HD	105	339	9.75	n.a.	72	32	4.87
patient 31	M	41	aHUS	HD	189	409	8	n.a.	n.a.	n.a.	12
patient 15	F	11	aHUS	PD	254	339	9.2	n.a.	n.a.	n.a.	3.5
patient 17	F	52	aHUS	PD	291	352	11.1	n.a.	112	54	11
patient 32	F	71	aHUS	HD	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
patient 33	F	43	aHUS	PD	164	394	12.2	n.a.	86	n.a.	3.2
patient 34	F	43	aHUS	HD	260	345	14.5	n.a.	58	27	8.4
patient 35	F	46	aHUS	HD	168	447	12.8	n.a.	n.a.	n.a.	8.8
patient 36	M	57	aHUS	HD	130	418	13.2	n.a.	51	n.a.	9
patient 37	F	37	aHUS	HD	328	453	11.4	n.a.	63	33	8.11
patient 38	F	49	aHUS	HD	238	n.a.	11.6	n.a.	n.a.	n.a.	11.6
patient 39	F	48	aHUS	HD	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
patient 40	M	51	aHUS	HD	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.	n.a.
patient 41	M	52	aHUS	HD	175	401	13.6	n.a.	111	n.a.	6.6
patient 42	M	53	aHUS	n.a.	176	n.a.	13.6	n.a.	67	47	7.58
patient 43	F	37	aHUS	HD	269	n.a.	9.6	n.a.	48.6	25	11
patient 20	M	60	aHUS	PD	n.a.	n.a.	n.a.	n.a.	125	41	8
patient 44	F	23	aHUS	HD	209	n.a.	11.9	n.a.	36.4	28.3	5.7
patient 45	M	2	aHUS	PD	395	n.a.	12.6	n.a.	n.a.	n.a.	11.4
patient 46	F	46	aHUS	HD	179	136	10.6	37	n.a.	n.a.	n.a.
patient 47	F	25	aHUS	HD	166	397	9.3	n.a.	67	19	n.a.

n.a. not available. HD, hemodialysis. PD, peritoneal dialysis.

Supplementary Table 5. Characteristics, clinical and complement parameters of stable renal transplant patients at the time of sample collection.

patient (n°)	gender	age (years)	native kidney disease	time from trasplant (days)	Kidney donor	immunosuppression therapy	platelets 150-400x10 ³ /μL	hemoglobin 14-18 g/dL	serum creatinin 0.55-1.25 mg/dL	C5b-9 on HMEC-1	
										unstimulated (50-150%)	ADP-activated (60-149)%
patient 74	M	81	FSGS	182	cadaver	TA - MMF	200	13.4	2.29	106	109
patient 75	F	59	proteinuric N	730	cadaver	TA - MMF	343	14.9	1.21	100	113
patient 76	M	49	IgA N	730	cadaver	TA - MMF	191	12.9	1.16	95	97
patient 77	M	61	ADPKD	1460	cadaver	TA - MMF	227	13.4	1.33	101	108
patient 78	M	56	hypertensive N	2190	cadaver	TA - MMF	206	14.5	1.58	113	122
patient 79	M	36	IgA N	6	living	TA - MMF - ST	196	10.7	1.03	217	221
patient 80	M	39	IgA N	182	cadaver	TA - MMF	139	13.4	1.39	98	99

TA, Tacrolimus. MMF, mycophenolate. ST, steroids. FSGS, focal segmental glomerulosclerosis. N, nephropathy. ADPKD, autosomal dominant polycystic kidney disease.

Supplementary Table 6. Characteristics, clinical and complement parameters of control dialyzed patients at the time of sample collection.

patient (n°)	gender	age (years)	native kidney disease	dialysis	platelets 150-400x10 ³ /μL	LDH 266-500 IU/L	hemoglobin 14-18 g/dL	C3 70-152 mg/dL	C4 15-40 mg/dL	plasma sC5b-9 140-280 ng/mL	serum creatinin 0.55 - 1.25 mg/dL	C5b-9 on HMEC-1		rare gene variants
												unstimulated (50-150%)	ADP-activated (60-149)%	
patient 48	M	48	ADPKD	HD	179	320	13	111	23	n.a.	15.22	110	148	n.a.
patient 49	M	26	proteinuric N	PD	264	436	12.4	149	28	n.a.	9.4	115	116	n.a.
patient 50	M	47	ADPKD	HD	166	n.a.	12.3	n.a.	n.a.	n.a.	13.7	95	99	n.a.
patient 51	M	32	CRI	HD	267	353	11.9	n.a.	n.a.	n.a.	14.74	91	121	n.a.
patient 52	F	27	ADPKD	PD	359	269	9	113	25	n.a.	12.91	65	78	n.a.
patient 53	M	46	fabry disease	PD	256	257	11.8	93	19	n.a.	10.3	98	130	n.a.
patient 54	M	46	MPGN	PD	250	353	11.7	96	22	n.a.	10.36	91	102	n.a.
patient 55	M	28	nephroangio sclerosis	HD	277	277	12	64	43	n.a.	17.47	105	119	NO
patient 56	M	39	CRI	PD	268	440	13.4	119	35	175	7.24	113	138	NO
patient 57	M	62	ADPKD	HD	240	n.a.	11.4	78	27	n.a.	7.73	94	159	NO
patient 58	F	59	horse kidney	HD	251	n.a.	11.2	n.a.	n.a.	n.a.	8	115	127	NO

n.a. not available. N, nephropathy. ADPKD, autosomal dominant polycystic kidney disease. MPGN, membrano-proliferative-glomerulonephritis. CRI, chronic renal insufficiency. HD, hemodialysis. PD, peritoneal dialysis.

Supplementary Table 7. Complement parameters and genetic characteristics of aHUS patients with recurrence after kidney transplant at the time of sample collection.

patient (n°)	plasma sC5b-9 140-280 ng/mL	C5b-9 on HMEC-1			rare gene variants					risk haplotypes			
		unstimulated (50-150%)	ADP-activated (60-149)%		gene	nucleotide change	aminoacid change	gnomAD frequency	CADD	classification	CFH-H3 (n)	MCPggaac (n)	total (n)
patient 1	n.a.	161	151	YES	CFI	c.905-3T>C	-	1.4x10 ⁻⁴	13	VUS	2	0	2
patient 2	n.a.	298	365	NO	-	-	-	-	-	-	1	0	1
patient 3	102	173	182	n.a.	-	-	-	-	-	-	n.a.	n.a.	n.a.
patient 4	n.a.	196	197	YES	MCP	c.286+2T>G	-	1x10 ⁻⁴	14.5	PV	1	2	3
patient 5	n.a.	255	283	NO	-	-	-	-	-	-	0	2	2
patient 6*	n.a.	84	86	NO	-	-	-	-	-	-	0	0	0
patient 7	n.a.	204	219	NO	-	-	-	-	-	-	0	n.a.	0
patient 8	96	209	212	NO	-	-	-	-	-	-	2	1	3
patient 9	1043	165	180	NO	-	-	-	-	-	-	1	2	3
patient 10	n.a.	187	187	NO	-	-	-	-	-	-	2	1	3
patient 11	269	182	204	YES	MCP CFI	c.286+2T>G c.G950A	- p.R317Q	1x10 ⁻⁴ 5.8x10 ⁻⁵	14.5 15	PV VUS	2	2	4
patient 12	242	169	178	YES	MCP	c.C1148T	p.T383I	1.9x10 ⁻³	4	VUS	2	1	3

n.a. not available. VUS, variant of uncertain significance. PV, pathogenic variant.

Supplementary Table 8. Complement parameters and genetic characteristics of transplanted aHUS patients with no sign of recurrence at the time of sample collection.

patient (n°)	plasma sC5b-9 140-280 ng/mL	C5b-9 on HMEC-1			gene	rare gene variants					risk haplotypes		
		unstimulated (50-150%)	ADP-activated (60-149)%			nucleotide change	aminoacid change	gnomAD frequency	CADD	classification	CFH-H3 (n)	MCPggaac (n)	total (n)
patient 13	n.a.	105	173	YES	CFI	c.A779G	p.Q260R	2.4x10 ⁻⁵	1	VUS	0	0	0
patient 14	n.a.	145	180	NO	-	-	-	-	-	-	0	0	0
patient 15	219	123	159	NO	-	-	-	-	-	-	0	1	1
patient 16	n.a.	107	138	NO	-	-	-	-	-	-	0	2	2
patient 17	295	78	256	NO	-	-	-	-	-	-	1	1	2
patient 18	n.a.	135	172	NO	-	-	-	-	-	-	n.a.	n.a.	n.a.
patient 19	n.a.	95	198	NO	-	-	-	-	-	-	2	1	3
patient 20	676	132	264	YES	CFI	c.C148G	p.P50A	1x10 ⁻³	24.9	VUS	0	1	1
patient 21	2092	131	161	YES	CFI	c.G1075T	p.D359Y	8.7x10 ⁻⁶	24.4	VUS	0	0	0
patient 22	n.a.	133	135	YES	C3 <i>CFHR1/CFH hybrid</i>	c.G2284A reverse hybrid <i>CFHR1</i> ₁₋₅ - <i>CFH</i> ₂₃	p.V762I	0.0002	0.25	VUS PV	0	2	2
patient 23	n.a.	215	225	NO	-	-	-	-	-	-	1	0	1

n.a. not available. VUS, variant of uncertain significance. PV, pathogenic variant.

Supplementary Table 9. Complement parameters and genetic characteristics of patients with de-novo TMA at the time of sample collection.

patient (n°)	plasma sC5b-9 140-280 ng/mL	C5b9 on HMEC-1			rare gene variants					risk haplotypes			
		unstimulated (50-150%)	ADP-activated (60-149)%		gene	nucleotide change	aminoacid change	gnomAD frequency	CADD	classification	CFH-H3 (n)	MCPggaac (n)	total (n)
patient 59	n.a	171	247	NO	-	-	-	-	-	-	0	1	1
patient 60	n.a	196	230	NO	-	-	-	-	-	-	0	0	0
patient 61	235	181	213	NO	-	-	-	-	-	-	1	0	1
patient 62	252	245	262	NO	-	-	-	-	-	-	1	0	1
patient 63	n.a	175	201	NO	-	-	-	-	-	-	0	0	0
patient 64	974	197	242	NO	-	-	-	-	-	-	0	1	1
patient 65	n.a	378	384	NO	-	-	-	-	-	-	0	0	0
patient 66	n.a	188	235	YES	CFH	c.C1511T	p. T504M	3x10 ⁻⁴	6.3	VUS	0	1	1
patient 67	n.a	303	311	NO	-	-	-	-	-	-	0	0	0
patient 68	831	193	209	NO	-	-	-	-	-	-	0	0	0
patient 69	n.a	230	268	YES	CFH	c.C3626T	p. S1209L	-	9	VUS	1	0	1
patient 70	833	301	352	YES	C3	c.G1909C	p. G637R	5x10 ⁻⁴	23.5	LPV	1	1	2
patient 71	n.a	241	244	NO	-	-	-	-	-	-	0	2	2
patient 72	277	232	273	NO	-	-	-	-	-	-	1	0	1
patient 73	358	194	225	NO	-	-	-	-	-	-	1	0	1

n.a. not available.

Supplementary Table 10. Complement parameters and genetic characteristics of aHUS patients in chronic dialysis at the time of sample collection.

patient (n°)	plasma sC5b-9 140-280 ng/mL	C5b 9 on HMEC-1			gene	rare gene variants				classification	risk haplotypes		
		unstimulated (50-150%)	ADP-activated (60-149)%			nucleotide change	aminoacid change	gnomAD frequency	CADD		CFH-H3 (n)	MCPggaac (n)	total (n)
patient 24	233	97	246	YES	CFH	c.C3628T	p.R1210C	0.0003	11.8	PV	1	1	2
patient 25	380	129	193	YES	CFH	c.A232G	p.R78G	-	15.8	PV	0	1	1
patient 22	127	114	213	YES	C3 <i>CFHR1/CFH hybrid</i>	c.G2284A reverse hybrid <i>CFHR1</i> ₁₋₅ - <i>CFH</i> ₂₃	p.V762I -	0.0002 -	0.25 -	VUS PV	0	2	2
patient 26	n.a	120	154	YES	C3	c.G181A	p.D61N	1x10 ⁻⁴	19	LPV	2	1	3
patient 27	260	145	201	NO	-	-	-	-	-	-	0	0	0
patient 28	n.a	111	173	NO	-	-	-	-	-	-	1	0	1
patient 29	1436	78	229	YES	C3	c.1774C>T	p.R592W	1.5x10 ⁻⁵	34	LPV	2	2	4
patient 30	n.a	102	199	n.a.	-	-	-	-	-	-	n.a.	n.a.	n.a.
patient 31	n.a	139	235	NO	-	-	-	-	-	-	0	1	1
patient 15	183	148	504	NO	-	-	-	-	-	-	0	1	1
patient 17	295	n.a.	409	NO	-	-	-	-	-	-	1	1	2
patient 32	n.a	n.a.	336	YES	CFI	c.G1234A	p.V412M	0.0013	25	VUS	0	2	2
patient 33	444	128	266	YES	C3	c.A3187C	p.S1063R	3.7x10 ⁻⁵	23.4	LPV	0	n.a.	n.a.
patient 34	n.a	104	155	NO	-	-	-	-	-	-	1	0	1
patient 35	n.a	142	164	NO	-	-	-	-	-	-	0	0	0
patient 36	n.a	n.a.	403	YES	CFH	c.G3587T	p.E1172X	0.0002	-	PV	0	n.a.	n.a.
patient 37	n.a	n.a.	590	YES	CFH	c.C3572T	p.S1191L	-	18.6	PV	0	2	2
patient 38	91	173	177	YES	<i>CFHR1/CFH hybrid</i>	reverse hybrid <i>CFHR1</i> ₁₋₄ - <i>CFH</i> ₂₂₋₂₃	-	-	-	PV	1	0	1
patient 39	n.a	155	276	NO	-	-	-	-	-	-	1	1	2
patient 40	n.a	191	197	YES	C3	c.4973T>C	p.V1658A	-	1.45	VUS	n.a.	n.a.	n.a.
patient 41	236	n.a.	605	YES	<i>CFHR1/CFH hybrid</i>	reverse hybrid <i>CFHR1</i> ₁₋₅ - <i>CFH</i> ₂₃	-	-	-	PV	1	2	3
patient 42	467	249	286	NO	-	-	-	-	-	-	1	1	2
patient 43	n.a	n.a.	1219	YES	CFH	c.3493+1G>C	-	-	24.3	PV	2	0	2
patient 20	676	n.a.	250	YES	CFI	c.C148G	p.P50A	1x10 ⁻³	24.9	VUS	0	1	1
patient 44	294	137	170	YES	C3	c.T544C	p.S182P	-	7.12	VUS	0	0	0
patient 45	n.a	120	157	NO	-	-	-	-	-	-	0	0	0
patient 46	n.a	140	189	YES	CFI	c.C148G	p.P50A	1x10 ⁻³	24.9	VUS	1	1	2
patient 47	n.a	193	207	NO	-	-	-	-	-	-	1	1	2

n.a. not available. PV, pathogenic variant. LPV, likely pathogenic variant. VUS, variant of uncertain significance.

International Registry of Recurrent and Familial HUS/ TTP

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