DOI: 10.1111/ivim.17221

STANDARD ARTICLE

Journal of Veterinary Internal Medicine AC

American College Veterinary Internal Medicine

Open Access

Familial osteochondrodysplastic and cardiomyopathic syndrome in Chianina cattle

¹Department of Veterinary Medical Sciences, University of Bologna, Bologna, Italy

²Institute of Genetics, Department of Clinical Research and Veterinary Public Health, Vetsuisse Faculty, University of Bern, Bern, Switzerland

Correspondence

Cord Drögemüller, Institute of Genetics, Department of Clinical Research and Veterinary Public Health, Vetsuisse Faculty, University of Bern, Bern, Switzerland. Email: cord.droegemueller@unibe.ch

Joana G. P. Jacinto^{1,2} | Tolulope G. Ogundipe¹ | Cinzia Benazzi¹ Irene M. Häfliger² | Luisa V. Muscatello¹ | Marilena Bolcato¹ Riccardo Rinnovati¹ | Arcangelo Gentile¹ | Cord Drögemüller²

Abstract

Background: Skeletal dysplasia encompasses a heterogeneous group of genetic disorders characterized by an abnormal development of bones, joints, and cartilage. Two Chianina half-sibling calves from consanguineous mating with congenital skeletal malformations and cardiac abnormalities were identified.

Hypothesis/Objectives: To characterize the disease phenotype, to evaluate its genetic cause, and to determine the prevalence of the deleterious alleles in the Chianina population.

Animals: Two affected calves, their parents and 332 Chianina bulls.

Methods: The affected animals underwent clinicopathological investigation. Wholegenome sequencing trio-approach and PCR-based assessment of the frequency of TDP-glucose 4,6-dehydratase (TGDS) and laminin subunit alpha 4 (LAMA4) alleles were performed.

Results: The cases presented with retarded growth, poor nutritional status associated with muscular atrophy and angular deformities of the hindlimbs. Radiologic examination identified generalized osteopenia and shortening of the limb long bones. Necropsy showed osteochondrodysplastic limbs and dilatation of the heart right ventricle. On histological examination, the physeal cartilages were characterized by multifocal mild to moderate loss of the normal columnar arrangement of chondrocytes. Osteopenia also was observed. Genetic analysis identified a missense variant in TGDS and a splice-site variant in LAMA4, both of which were homozygous in the 2 cases. Parents were heterozygous and allele frequency in the Chianina population for the TGDS variant was 5% and for the LAMA4 variant was 2%.

Conclusions and Clinical Importance: Genetic findings identified 2 potentially pathogenic alleles in TGDS and LAMA4, but no clear mode of inheritance could be determined.

Abbreviations: Al. artificial insemination; BTV, bluetongue virus; BVDV, bovine viral diarrhea virus; ECM, extracellular matrix; FAK, focal adhesion pathwav; H&E, hematoxylin and eosin; IF, ichthyosis fetalis; IGV, Integrative Genomics Viewer; LAMA4, laminin subunit alpha 4; PMT, pseudomyotonia congenita; RI, reference intervals; SBV, Schmallenberg virus; SDR, short chain dehydrogenase-reductases family; TGDS, TDP-glucose 4,6-dehydratase; VBG, venous blood gas analysis; WGS, whole-genome sequencing,

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2024 The Author(s). Journal of Veterinary Internal Medicine published by Wiley Periodicals LLC on behalf of American College of Veterinary Internal Medicine.

KEYWORDS

bovine, inbreeding, lethal allele frequency, precision medicine, recessive disorders

1 | INTRODUCTION

Skeletal dysplasia comprises a heterogeneous group of rare genetic disorders characterized by abnormal development of bones, joints, and cartilage.¹ Lethal dominantly inherited skeletal dysplasia syndromes in cattle include achondrogenesis type II COL2A1-related in Holstein and crossbred (OMIA 001926-9913),²⁻⁵ chondrodysplasia FGFR3-related (OMIA 001703-9913),⁶ and vertebral and spinal dysplasia T-related in Holstein (OMIA 001951-9913),⁷ frontonasal dysplasia ZIC2-related in Limousine (OMIA 002307-9913).⁸ osteogenesis imperfecta type II COL1A1-related in Red Angus and Holstein (OMIA 002127-9913),^{5,9,10} Marfan syndrome FBN1-related in Japanese Black (OMIA 000628-9913).¹¹ and skeletal-cardioenteric dysplasia MAP2K2-related in Romagnola (OMIA 002381-9913).¹² Lethal recessively inherited skeletal dysplasias also have been described in cattle. They include arachnomelia MOCS1- and SUOX-related in Fleckvieh and Brown Swiss respectively (OMIA 001541-9913; OMIA 000059-9913),^{13,14} brachyspina FANCI-related (OMIA 000151-9913)¹⁵ and complex vertebral malformation SLC35A3-related (OMIA 001340-9913)¹⁶ in Holstein, hereditary perinatal weak calf syndrome IARS-related in Japanese Black (OMIA 001817-9913).¹⁷ osteopetrosis SLC4A2-related in Red Angus (OMIA 002443-9913)¹⁸ and lethal multiorgan developmental dysplasia KDM2B-related in Romagnola and Marchigiana (OMIA 001722-9913).^{19,20} Most of the skeletal dysplasias described in cattle are associated with malformations or defects in other organs such as the heart.

In recent decades, molecular investigations allowed the discovery of the etiology at the DNA level of an increasing number of diseases in cattle. This discovery also has occurred with respect to Chianina, 1 of the most important Italian autochthonous beef cattle breeds. Chianina cattle are mainly reared for beef production, and in recent years intensive selective breeding programs have been carried out to improve performance, leading to a worrisome increase in inbreeding rates.²¹ These practices have led to the emergence of 3 recessively inherited disorders: pseudomyotonia congenita (PMT; OMIA 001464-9913),²² ichthyosis fetalis (IF; OMIA 002238-9913),²³ and ichthyosis congenita (OMIA 002450-9913).²⁴ Currently, PMT and IF genetic testing is routinely included in the breeding program of the Italian Association of Italian Beef Cattle Breeders.

Two half-sibling Chianina calves with congenital skeletal malformations and cardiac abnormalities were identified. The aim of our study was to report the clinicopathological phenotype of a familial osteochondrodysplastic and cardiomyopathic syndrome in Chianina cattle. Moreover, genome analysis was carried out and the prevalence of possible deleterious alleles in the Chianina population was estimated.

2 | MATERIALS AND METHODS

2.1 | Animals

Our study did not require official or institutional ethical approval because it was not experimental, but rather part of clinical and pathological veterinary diagnostic testing. All animals in the study were examined with the consent of their owners and handled according to ethical standards. A total of 337 Chianina cattle, including 2 affected calves, their dams and their sire and 332 healthy control bulls were studied.

2.2 | Clinical and pathological investigations

Two Chianina calves (cases 1 and 2) that were presented with obvious congenital skeletal malformations were referred to the Clinic for Ruminants of the University of Bologna. Both calves were the result of natural mating.

Case 1 was a 3-month-old female calf. It had a history of weakness, impaired growth and abnormal skeletal development. The calf underwent a physical examination and skeletal radiological investigation. In addition, venous blood gas analysis (VBG), CBC, serum biochemical analysis were performed. The calf was euthanized after the clinical investigations because of severe emaciation, lameness and renal disease, and submitted for necropsy and histological examination.

Case 2 was a stillbirth male calf reported because of skeletal abnormalities. After having carried out a postmortem skeletal radio-logical investigation, the animal was submitted for necropsy and histo-logical examination.

The cases were tested for bovine viral diarrhea virus (BVDV), Schmallenberg virus (SBV), bluetongue virus (BTV), *Neospora caninum*, and *Toxoplasma gondii* using PCR and ELISA for detecting antigens and antibodies, respectively.

Tissues were collected, formalin-fixed and paraffin embedded. Three-micron thick sections were cut and routinely stained with hematoxylin and eosin (H&E). Bone tissue was further stained with toluidine blue.

2.3 | Genetic analysis and DNA extractions

A pedigree analysis of the affected calves was performed.

Genomic DNA was obtained from the affected animals (ear cartilage tissue samples), their respective dams (EDTA blood samples) and the common sire (semen) using Promega Maxwell RSC DNA system (Promega, Dübendorf, Switzerland). Furthermore, genomic DNA also

3347

Open Access American Colleg

was obtained from semen of Chianina artificial insemination (AI) top sires' semen as well as from EDTA blood samples of Chianina young bull calves shortlisted for admission to the performance test at the testing station in the years 2017, 2018, 2019, and 2020.

2.4 | Whole-genome sequencing and variant calling

A whole-genome sequencing (WGS) trio approach using the Illumina NovaSeq6000 (Illumina Inc., San Diego, CA, USA) was performed on the genomic DNA extracted from the 2 affected calves (cases 1 and 2), their 2 dams and the common sire. The sequenced reads were mapped to the ARS-UCD1.2 reference genome, resulting in an average read depth of approximately $18.2 \times$ in case 1 and $17.9 \times$ in case 2, and single nucleotide variants and small indel variants were called.²⁵ The applied software and steps to process fastg files into binary alignment map (BAM) and genomic variant call format files were in accordance with the 1000 Bull Genomes Project processing guidelines of run 7,²⁶ except for the trimming, which was performed using fastp.²⁷ Further preparation of the genomic data was performed as reported previously.²⁸ The effects of the above variants were functionally evaluated with snpeff v4.3,²⁹ using the National Center for Biotechnology Information Annotation Release 106 (https://www.ncbi.nlm.nih.gov/ genome/annotation_euk/Bos_taurus/106/; accessed on 20 September 2022). This resulted in the final VCF file, comprising individual variants and their functional annotations. To identify private variants, we compared the genotypes of the cases with 942 cattle genomes of different breeds sequenced as part of the ongoing Swiss Comparative Bovine Resequencing project. Integrative Genomics Viewer (IGV)³⁰ software version 2.0 was used for visual evaluation of genome regions containing potential candidate genes.

2.5 | Homozygosity analysis

Biallelic variants were selected from the vcf file using PLINK v1.9³¹ command –biallelic-only as a common quality control step. A genomewide search for homozygous regions shared by the 2 cases was performed using the R package detectRUNS v.0.9.6³² with the commands –homozyg-kb 1000 (considering homozygous segments of at least 1000 kb), –homozyg-match 0.95 (for allelic matching between both cases) and –homozyg-group (for generating an overlap file), resulting in shared runs of homozygosity (ROH) indicating chromosomal region of identity-by-descent (IBD).

2.6 | Variant validation and selection of candidate variants

The comprehensive variant catalog of run 9 of the 1000 Bull Genomes Project was used to investigate the allele distribution of variants within a global control cohort allowing the exclusion of common variants.²⁶ The entire data set consists of 5116 cattle genomes including 576 from the Swiss Comparative Bovine Resequencing project, from a variety of breeds (>130 breeds indicated).

PredictSNP1,³³ PolyPhen-2,³⁴ SIFT,³⁵ and Mutpred2³⁶ were used to predict the biological consequences of the detected missense variant. For cross-species sequence alignments, the following NCBI protein accessions were considered: NP_001094629.1 (*Bos taurus*), NP_055120.1 (*Homo sapiens*), XP_522697.2 (*Pan troglodytes*), XP_001083495.1 (*Macaca mulatta*), XP_542640.3 (*Canis lupus*), NP_083854.3 (*Mus musculus*), XP_416988.1 (*Gallus gallus*), NP_956111.1 (*Danio rerio*), and XP_002939327.2 (*Xenopus tropicalis*).

The GeneMANIA tool³⁷ was used to predict the interaction between TDP-glucose 4,6-dehydratase (*TGDS*) and laminin subunit alpha 4 (*LAMA4*).

2.7 | Investigation of structural variants

To assess possible larger structural variants, including chromosomal, numerical, and structural abnormalities, the read depth along all chromosomes was calculated as previously described.³⁸ In addition, in shared ROH identified regions (Table S3) the read depth also was calculated.

2.8 | Sequence accessions

All references to the bovine *TGDS* gene correspond to the NCBI accessions NC_037339.1 (chromosome 12, ARS-UCD1.2), NM_001101159.1 (*TGDS* mRNA), and NP_001094629.1 (*TGDS* protein). For the protein structure of TGDS the Uniprot database³⁹ and Inter-Pro⁴⁰ with accession number A6QLW2 was used.

All references for the bovine LAMA4 gene correspond to the NCBI accessions NC_037336.1 (chromosome 9, ARS-UCD1.2), XM_024996678.1 (LAMA4 mRNA), and XP_024852446.1 (LAMA4 protein). For the protein structure of LAMA4 the Uniprot database³⁹ and InterPro⁴⁰ with accession number A0A3Q1N9E9 were used.

2.9 | Genotyping

Polymerase chain reaction and Sanger sequencing were used to confirm the WGS results and to perform targeted genotyping for the identified TGDS missense variant (12:69092831A>T). The 2 cases, their dams and the sire were genotyped. Additionally, Chianina AI top sires (n = 113) as well as Chianina young males shortlisted for admission to the performance testing at the testing station in the years 2017, 2018, 2019, and 2020 (n = 219) were genotyped. Primers were designed using the Primer-BLAST tool. After amplification with Ampli-TaqGold360Mastermix (Thermo Fisher Scientific), the purified PCR products were directly sequenced on an ABI3730 capillary sequencer (Thermo Fisher Scientific). The primer sequences used for genotyping the Chr12:69092831A>T varinat in TGDS were the following: 5'-TCCAAGCCTATATCACACAGGT-3' (forward primer) and 5'-GCCTACCACAGACAGTAAGT-3' (reverse primer). The primer

American College of terinary Internal Medicine 3349

3 | RESULTS

3.1 | Clinical phenotype

On physical examination, case 1 had retarded growth and poor nutritional status associated with muscular atrophy and skeletal abnormalities (Figure 1A). These were characterized by pronounced varus of the hindlimbs and wide stance of the forelimbs. Severe angular deformities of the hindlimbs mainly in the region of the hock joint and in the distal portion of the tibia were noted (Video S1). In particular, the left hindlimb was more severely affected and consequently the calf only placed it with the hoof tip (Figure 1B). Limited motion of the hock join bilaterally but more severe in the left hindlimb was perceived on passive motion testing. The gait was impaired and characterized by reluctance to move and with complete weight transfer of the left to the contralateral hindlimb. The haircoat was dull with presence of *Linognathus* spp. The oculoconjunctival mucous membranes were pale and the episcleral vessels were visible. The heart rate was slightly increased at 90 beats/minute. Spontaneously voided urine was macroscopically pale yellow and transparent. The urine specific gravity was 1.010 and the pH 8.5.

Venous blood gas analysis yielded a temperature corrected pO₂ of 37 mmHg and O₂ saturation of 51.4%. A CBC showed moderate erythrocytosis (10.2×10^{12} /L; reference intervals [RI]: 5-7.2), with microcytosis (mean cell volume: 29.1 fL; RI: 38-51), decreased mean corpuscular hemoglobin (10 pg; RI: 11-17), but with normal hematocrit (31.9%; RI: 24-35) and mean cell hemoglobin concentration (34.9 g/dL; RI: 34-38). Echinocytes also were noticed in the blood smear. On plasma biochemistry, marked hyperlactatemia (7.2 mmol/L; RI: 0-2), hypocholesterolemia (46 mg/dL; RI: 163-397), increased BUN concentration (67 mg/dL; RI: 4.2-20.4), increased uric acid concentration (1 mg/dL; RI: 0.1-0.4) and mild hyperkalemia (6 mmol/L; RI: 3.9-5.75) were identified.

In both cases, tests for BVDV, SBV, BTV, *Neospora caninum*, and *Toxoplasma gondii* were negative using PCR and ELISA.



FIGURE 1 Osteochondrodysplasia in Chianina cattle. (A) and (B), Case 1 showing severe angular deformities of the hindlimbs mainly in the region of the hock joint and in the distal portion of the tibia. Note that the left hindlimb is more severely affected and consequently the calf only places it with the hoof tip. Note the pronounced varus posture of the hindlimbs. (C) Case 2 showing severe angular deformities of the hindlimbs in the region of the hock joint, in the distal portion of the tibia and proximal portion of the metatarsal bone. Note the inferior brachygnathia. (D) Case 2, detail of the hindlimbs.

3.2 Skeletal radiological investigation

The radiological examination showed similar findings in both cases. The distal femur and proximal tibia secondary ossification centers appeared to be decreased and with slower development. The metaphysis of the same segments was enlarged and with sharp medial and lateral angles. The metaphyses maintained their internal areas of trabecular bone with vertical striations and, in the tibia, varus deformity was present (Figure 2A,B). Generalized osteopenia with cortical thinning also was present. The radius showed bone shortening with morphologic changes, and generalized osteopenia and cortical thinning were present. The metacarpal and metatarsal bones were shorter than normal, asymmetrically misshapen, with enlargement of proximal and distal ends and bending of the shafts. Cortical thinning was present.

3.3 Pathological phenotype

Macroscopically, case 1 had localized hindlimb chondrodysplasia, with severe volume increase of the left tibio-tarsal joint, and on the right a similar modest volume increase of the tibio-tarsal joint accompanied by curvature of the tibia and metatarsus. In the left limb, there was evidence of edema infiltrating the gluteal muscles. The capsule of the left tibio-tarsal joint had a thickness of 1 cm, and yielded approximately 10 mL of clear, colorless synovial fluid. The left tibia showed an angle of approximately 90° at the lower 3rd in the dorsoventral direction (Figure 3A). In sagittal section, the diaphyseal medulla was interrupted at the angle. The red pith typical of young subjects was present in the proximal segment for a length of 4 cm near the

interruption, formed by a 0.5 cm layer of connective tissue. The diaphyseal metatarsal was 2.5 cm thick, and the centrally-located hematopoietic medulla was replaced by adipose tissue (Figure 3B).

The kidneys had a light hazel cortex (Figure 3C). The heart had a dilated and ectatic right ventricle, whereas the epicardium lining the left ventricle showed ectatic lymphatics (Figure 3D).

Case 2 showed marked prognathism and micrognathia (Figure 1C). All 4 limbs were osteochondrodysplastic: the forelimbs curved and rotated inward, both at the height of the humerus; the hindlimbs also were intrarotated, T forming a 30° angle at the tarsometatarsal joint (Figure 1D). On section, the distal epiphysis of the femur showed a rounded portion of red hematopoietic marrow approximately 3 cm in diameter in a central position, as did the tibia. On sectioning, the tibia had hematopoietic marrow for approximately 2 cm in the central portion of the proximal epiphysis whereas the remainder appeared to consist of spongiosa. In the middle, the diaphysis had an angle of approximately 30° in the caudal position with an apposition of connective tissue of 1.5 cm. The macroscopic findings of the heart and kidneys were similar to those of case 1.

On histological examination of the heart, the connective tissue between the groups of cardiomyocytes was edematous with ectatic lymphatic vessels. Leakage of blood from small vessels with erythrocytes had accumulated in the spaces between the bundles of heart muscle fibers and infiltrated between individual fibers. Small foci of lymphoid cell myocarditis surrounded the capillaries and some of the arterioles, and were more prominent in thickness in the epicardium. The histomorphological features of the heart resembled a form of cardiomyopathy.

In the long bones, the physeal cartilage, more severe in case 2, was characterized by multifocal mild to moderate loss of the columnar



Radiology of the hindlimbs of Chianina cattle with osteochondrodysplasia. (A) Note the 90° angular deformity in the tibia. In the FIGURE 2 proximal tibia, secondary ossification centers appear to be narrowed and with slower development in Case 2. (B) Note the 90° angular deformity in the metatarsus in Case 2.



FIGURE 3 Gross pathological findings of Chianina cattle with osteochondrodysplastic and cardiomyopathic syndrome. (A) Sagittal section. The left tibia showing an angle of approximately 90° at the lower 3rd in the dorsoventral direction; the diaphyseal medulla is interrupted at the angle mentioned above in Case 1. (B) Sagittal section. The diaphyseal metatarsal is 2.5 cm thick, and the centrally located hematopoietic medulla has been replaced by adipose tissue in case 1. (C) Kidney showing a light hazel cortex in Case 2. (D) Heart showing a dilated and ectatic right ventricle and epicardium lining the left ventricle with ectatic lymphatics in Case 1.

arrangement of chondrocytes. The chondrocytes of the proliferative zone were multifocal and arranged in clusters with loss of columnar polarity. The primary spongiosa was decreased in thickness with occasional retention of hypertrophic chondrocytes. In both cases, the metaphyseal trabeculae were severely thin and spaced apart, with retention of islands of cartilage in the center of the trabeculae, morphologically suggestive of bone rarefaction (osteopenia; Figure 4A). No cartilage matrix degeneration was observed with toluidine blue staining (Figure 4B). The morphological features of the bones were indicative of physeal osteochondrodysplasia.

3.4 | Genetic analysis

Pedigree analysis indicated multiple inbreeding loops between the parents and a common male ancestor born in 2005 occurring 2-3 generations previously (Figure 5A). Given the apparent consanguinity, the fact that both sexes are affected and that no parents showed clinically

visible defects, we hypothesized that the current cases could be explained by a rare recessive inherited variant. In addition, because both cases were sired by the same bull, we also considered possible dominant inheritance. We therefore hypothesized 2 different possible scenarios: (i) a spontaneous, fully penetrant dominant trait arising from a de novo mutation inherited from the sire's germline or (ii) an autosomal recessive trait that was present in the homozygous state and inherited from both (heterozygous) parents.

Filtering of WGS for private shared heterozygous variants, present in the sequenced genomes of cases 1 and 2 and absent in 942 available control genomes (including the 3 parents), identified no variant common to both cases.

Filtering of WGS for private shared heterozygous variants present in the sequenced genomes of cases 1 and 2 and sire and absent in 942 available control genomes (including the 2 dams), identified 5 private protein-changing variants predicted as moderate. Analysis of the occurrence of these variants in the global control cohort of 4540 genomes from a variety of breeds²⁶ identified 3 remaining protein-



FIGURE 4 Histological findings of Chianina cattle with osteochondrodysplasia. (A) Physeal cartilage, 4×, hematoxylin and eosin: Loss of the columnar arrangement of chondrocytes, more severe in the proliferative zone in Case 2. (B) Physeal cartilage, toluidine blue, 4×: Uniform staining indicating no cartilage degeneration in Case 2.

changing variants with predicted moderate impact that were heterozygous only in the genomes of cases 1 and 2 and their sire (Table 1). These 3 variants then were visually inspected using the IGV software (Broad institute, Cambridge, MA, USA), which confirmed 1 as a true variant. This heterozygous variant at Chr 7:72238027C>T was a missense variant in exon 19 of the *ATPase phospholipid transporting 10B* (*ATP10B*) gene (XM_015472347.2:c.3574G>A). It exchanges the encoded amino acid of ATP10B at position 1192 (XP_015327833.2: p.Asp1192Asn), located in the P-type ATPase, C-terminal domain. Its effect was predicted to be neutral (PredictSNP1 score 60% neutral; PolyPhen-2 score 64% neutral; SIFT score 46% neutral).

Filtering of WGS for private shared homozygous variants, present in the sequenced genomes of cases 1 and 2, heterozygous in the 3 parents, and allowing the 942 available control genomes to be either heterozygous or homozygous wild type, identified 45 private proteinchanging variants with predicted moderate or high impact. Analysis of the occurrence of these variants in the global control cohort of 4540 genomes from a variety of breeds²⁶ identified 13 remaining proteinchanging variants with predicted moderate or high impact that were homozygous only in the genomes of cases 1 and 2. These 13 variants then were visually inspected using the IGV software (Broad institute, Cambridge, MA, USA), which confirmed 12 as true variants (Table S1). Among these 12 remaining private variants involving 11 different genes, 2 variants affected interesting functional candidate genes for the studied phenotype. One was a homozygous variant at Chr12:69092831A>T that is a missense variant in exon 3 of the TGDS gene (NM_001101159.1: c.160T>A; Figure S1). It exchanges the encoded amino acid of TGDS at position 54 (NP_001094629.1: p.Tyr54Asn), located in the NAD(P)-

binding domain (Figure 5C). Furthermore, the tyrosine-to- asparagine substitution affects a highly conserved residue (Figure 5D) and was predicted to be deleterious by 4 different tools (PredictSNP1 score 72% deleterious; PolyPhen-2 score 68% deleterious; SIFT score 79% deleterious; Mutpred2 score 0.922 deleterious). Indeed, it is predicted to alter the ordered interface, the metal binding, the DNA binding and transmembrane protein, to cause loss of the allosteric site at p.Tyr54 and loss of staining, and to cause gain of relative solvent accessibility, of catalytic site at p.Asp50 and of acetylation at p.Lys59.³⁶ The 2nd variant affecting an interesting functional candidate gene for the studied phenotype was a homozygous splice-site variant (Chr9:38176716GAGAAAGTGAGAG AGGGAAACAGAGGGGAGAGAGAA>G) affecting exon 1 of the LAMA4 gene (Figure 5B; Figure S2). Based on the used NCBI annotation it is predicted to be a splice-site variant that most likely leads to the expression of an altered protein-coding sequence (XM 024996678.1:c.153 153 + 34delAGTGAGAGAGGGAAACAGAGGGGAGAGAGAAAGAA).

Analysis of the remaining 9 identified variants, taking into account the known function of the genes, the reported association with Mendelian diseases, and the in silico assessment of the molecular consequences of the variants in the proteins, did not identify any other plausible cause for the observed phenotype (Table S2).

Homozygosity mapping identified 70 IBD segments >1 Mb and up to 3.1 Mb on 15 different chromosomes shared by the 2 sequenced cases and for which the 3 sequenced parents were heterozygous (Table S3). Two of these regions, a 1.8 Mb segment on chromosome 9 and a 2.1 Mb segment on chromosome 12, contained the genes LAMA4 and TGDS, respectively. No evidence of structural variants was found when analyzing the read depth coverage obtained in the



FIGURE 5 Homozygous laminin subunit alpha 4 (*LAMA4*) splice-site and TDP-glucose 4,6-dehydratase (*TGDS*) missense variants in the 2 affected Chianina calves. (A) Pedigree of the 2 cases affected by the osteochondrodysplasitc and cardiomyopathic syndrome. Note the inbreeding loops between the parents. The arrow indicates the common ancestor. DNA samples available from animals with genotypes for both variants are given below the symbols. (B) Schematic representation of the bovine LAMA4 protein and its functional domains. Note that the *LAMA4* splice-site variant is situated at the very beginning of the protein (red line and arrow). (C) Schematic representation of the bovine TGDS protein and its functional domains. Multiple sequence alignment of TGDS protein encompassing the region of the p.Tyr54Asn variant demonstrates complete evolutionary conservation across species.

ROH regions, nor were any chromosomal abnormalities detected when analyzing the read depth coverage across the chromosomes.

Analysis of *TGDS* genotyping data confirmed that cases 1 and 2 were homozygous and the dams and sire were heterozygous for the detected *TGDS* variant (Figure S1). Furthermore, the genotyping of the Chianina top sires and young bulls shortlisted for admission to performance testing at the testing station identified 32 heterozygous carriers (Table 2; Figure S3). In addition, 1 top sire was homozygous for the variant allele in *TGDS* but homozygous for the reference allele at *LAMA4*.

Analysis of LAMA4 genotyping data confirmed that cases 1 and 2 were homozygous and the dams and sire were heterozygous for the detected LAMA4 variant. Furthermore, genotyping of the same Chianina males identified 11 heterozygous carriers, but no animals homozygous for the variant LAMA4 allele (Table 2; Figure S3).

The total allele frequency in the male Chianina population was 5% for the *TGDS* variant and 2% for the *LAMA4* variant.

Using the GeneMANIA tool,³⁷ TGDS and LAMA4 were predicted to interact through SYMD3 and TP53 (Figure S4). In particular, TGDS was predicted to interact and be coexpressed with SMYD3. In addition, SMYD3 showed physical interactions with TP53. Finally, TP53 had physical interactions and colocalized with LAMA4.

4 | DISCUSSION

We completed a comprehensive clinical, histologic, and genetic evaluation of 2 Chianina calves with a familial osteochondrodysplastic and cardiomyopathic syndrome. We present evidence for the occurrence of a rare, possibly recessively inherited congenital syndrome that



TABLE 1 Results of variant filtering of the affected calves using the whole-genome sequence data.

Filtering Step	Homozygous Variants	Heterozygous Variants
All variants in case 1	3 529 543	4 668 770
All variants in case 2	4 023 845	3 808 248
Private variants in cases 1 and 2	6743	0
Private variants in cases 1 and 2 and their sire	NA	558
Private variants in cases 1 and 2 with obligatory carrier parents (protein-changing)	6180 (245)	NA
Private protein-changing variants in cases 1 and 2 with obligatory carrier parents and absent in homozygous state in 942 controls	45	NA
Protein-changing private variants in cases 1 and 2 with obligatory carrier parents and absent in homozygous state in a global cohort of 5279 controls	5	NA
Protein-changing private variants absent in both parents (de novo mutations) and absent in 942 controls	NA	0
Protein-changing private variants absent in the dams and present in the sire (de novo germinal in the sire) and absent in 942 controls	NA	5
Protein-changing private variants absent in the dams and present in the sire (de novo germinal in the sire) and in a global cohort of 5279 controls	NA	3

Abbreviation: NA, not applicable.

TABLE 2 Association of the missense variant in TGDS and of the splice-site variant in LAMA4 with the osteochondrodysplastic and cardiomyopathic syndrome in Chianina cattle.

	Genotype TGDS			Genotype LAMA4		
	ref/ref	ref/var	var/var	ref/ref	ref/var	var/var
Affected calves	0	0	2	0	0	2
Obligate carriers ^a	0	3	0	0	3	0
Chianina bulls	296	32	1	321	11	0
Normal control cattle from various breeds	5279	0	0	5278	1 ^b	0

Abbreviations: ref, reference allele; var, variant allele.

^aParents of the affected calves;

^bFrom Shorthorn breed.

could be explained by the 2 identified homozygous loss-of-function variants in TGDS and LAMA4. The trio-based WGS approach identified 12 protein-changing variants that simultaneously were uniformly homozygous in the genome of the affected calves. After gene function analysis, taking into account the occurrence of the variant alleles in a global control cohort, the rarity and breed specificity of the variants, and in silico effect predictions, only the TGDS and LAMA4 alleles were considered to be potential candidates to explain the observed congenital syndrome. We speculate that this congenital syndrome could have a recessive inheritance either caused by the TGDS or the LAMA4 allele. Alternatively, a parallel homozygosity for the TGDS and LAMA4 alleles could be considered a potential candidate genetic cause of the observed phenotype. The TGDS variant was identified only in Chianina cattle, whereas the LAMA4 variant was identified in both Chianina and Shorthorn cattle. Alternatively, evaluation of possible paternal germinal mosaicism identified only a single heterozygous missense variant in ATP10B with no evidence of deleterious effects. This variant was present in the genomes of the 2 affected paternal half siblings as well as in the sperm-derived DNA of their sire. Taking this into

account, the heterozygous ATP10B variant seems less plausible as a cause of the observed phenotype.

TGDS encodes the dTDP-D-glucose 4,6-dehydratase protein that is a member of the short chain dehydrogenase-reductases family (SDR), which plays a pivotal role in nucleotide-sugar metabolism.⁴¹ It is 1 of the 70 SDR genes the roles of which include the metabolism of retinoids, steroid hormones, and prostaglandins, as well as formation of the extracellular matrix (ECM).⁴² In humans, pathogenic variants in TGDS are associated with the recessive inherited Catel Mankze syndrome (OMIM 616145) that is characterized by bilateral clinodactyly, micrognathia digital syndrome, and in some cases cleft palate and heart malformations.⁴³ Affected patients are either homozygous mutant or compound heterozygous for the TGDS variants.^{43,44} We observed some similarities of Catel Mankze syndrome with the phenotype of our 2 cases, such as severe bone dysplasia and micrognathia.

LAMA4 encodes the laminin subunit alpha 4 protein, which is a member of the most abundant noncollagenous protein laminins that are components of the ECM of the basement membrane.45,46 LAMA4 is considered a signaling molecule and a multidomain glycoprotein.47

Journal of Veterinary Internal Medicine AC VIM

American College of terinary Internal Medicir 3355

Additionally, it plays an important role in tissue development, cellular adhesion, and migration. LAMA4 adhesion to integrins triggers cell signaling, regulation of pathways, and regulation of tissue formation during embryogenesis such as in cartilage development.⁴⁸⁻⁵¹ Furthermore, 1 of the pathways in which LAMA4 is involved is the focal adhesion pathway (FAK). Interestingly, FAK signaling influences Wnt/β-catenin pathways, impacting chondrogenesis and bone development.⁵²⁻⁵⁴ Disruption of this pathway may result in osteoarthritis, intervertebral disc degeneration, and rheumatoid arthritis.⁵⁴⁻⁵⁶ In humans, a missense variant in LAMA4 has been associated with a dominant inherited form of dilated cardiomyopathy (OMIM 600133).⁴⁹ We observed cardiac abnormalities sharing some similarities with dilated cardiomyopathy in humans such as dilatation of the heart and ectatic right ventricles in our cases. In addition, disruption in the FAK pathway could be caused by the observed splice-site variant in LAMA4 and impact bone development, contributing to the observed bone malformations.

We have described an inherited familial syndrome in cattle that may be associated with 2 independent deleterious recessive alleles in 2 different genes on different chromosomes. The central hypothesis of the Human Undiagnosed Diseases Network is that many of these rare genetic disorders remain undiagnosed because they are caused by multiple variants in >1 gene.⁵⁷ Recent studies have identified a role for digenic inheritance of Mendelian diseases in humans⁵⁸ and similar examples may exist in domestic animals.⁵⁹ In humans, digenic autosomal recessively inherited diseases include the severe dilated cardiomyopathy, which occurs because of variants in MYH7 and TNNT2.60 People who are homozygous for only 1 of the 2 pathogenic alleles do not have the disease. However, the disease is manifested when both variant alleles are present in parallel.⁶⁰ Additionally, Bardet Biedl syndrome (OMIM 617119), a recessively inherited multiorgan syndrome. also can be digenically inherited. Clinical features of this syndrome include polydactyly, neurodevelopmental disabilities, obesity, rodcone dystrophy, speech deficit, and renal disease.⁶¹⁻⁶³ In our study, only the 2 affected calves were homozygous for both the TGDS and LAMA4 variants. Although 1 top sire was identified as a homozygous carrier for the TGDS variant, the bull was homozygous wild type for the LAMA4 variant. We therefore hypothesized that, as in the abovementioned diseases of humans, possibly homozygosity for both TGDS and LAMA4 variants would be required to manifest the congenital syndrome in cattle. Therefore, a combined or additive effect of both variants leading to an osteochondrodysplastic and cardiomyopathic syndrome might be plausible. Digenic forms of inheritance involving TGDS and LAMA4 have never been reported. However, with the evidence presented in our study, it cannot be excluded that the presence of the homozygous variant in LAMA4 or alternatively in TGDS itself might be causative. Therefore, mating of carriers should be avoided to prevent further similarly affected offspring. Future research will be needed to provide additional insight into the biological effects of the identified TGDS and LAMA4 variants.

Currently, the use of short-read WGS still has some limitations, including methodological limitations such as structural variant detection, in addition to the still incomplete annotation of the bovine genome. In fact, the annotation of bovine LAMA4 is not entirely clear, as the alternative annotation available in the Ensembl genome browser shows that the deletion is in the 5' untranslated region of the gene. Nonetheless, as RNA sequencing data shows reads in the region of the deletion, we believe that the variant allele might be functionally important, with a regulatory effect, or as a true coding variant as mentioned above. Other possible limitations are the focus on coding variants in selected candidate genes based on literature searches, challenges associated with the short-read WGS approach, inaccuracies in read alignment and variant calling,⁶⁴ and potential influences of epigenetics factors.

Moreover, collecting additional similarly affected calves could help confirm or rule out the possible causality of the 2 identified variants of *TGDS* and *LAMA4*.

Our study alerts veterinarians and breeders of Chianina cattle to the potential emergence of rare diseases in the future. Reporting of additional similarly affected calves would help in understanding the underlying possible genetic causes and better clarify the mode of inheritance and role of the identified variants.

ACKNOWLEDGMENT

The authors acknowledge Isabella Aebi-Huber for expert technical assistance and the National Association of Italian Beef Cattle Breeders for providing the material. The WGS data are available under the study accession no. PRJEB18113 at the European Nucleotide Archive (www.ebi.ac.uk/ena; SAMEA111531430 is case 1, SAMEA111531431 is case's 1 dam, SAMEA111531432 is case 2, SAMEA111531433 is case's 2 dam, SAMEA111531434 is cases' 1 and 2 sire).

CONFLICT OF INTEREST DECLARATION

Authors declare no conflicts of interests.

OFF-LABEL ANTIMICROBIAL DECLARATION

Authors declare no off-label use of antimicrobials.

INSTITUTIONAL ANIMAL CARE AND USE COMMITTEE (IACUC) OR OTHER APPROVAL DECLARATION

This case description was not based on an invasive animal experiment but on clinical examination and diagnostic evaluation of spontaneous occurring cases; therefore, there are no associated authorization numbers. The control cattle sampling was performed for diagnostic purposes at admission at the testing station.

HUMAN ETHICS APPROVAL DECLARATION

Authors declare human ethics approval was not needed for this study.

ORCID

Joana G. P. Jacinto https://orcid.org/0000-0002-6438-7975 Tolulope G. Ogundipe https://orcid.org/0000-0002-3976-6391 Cinzia Benazzi https://orcid.org/0000-0003-1726-0442 Irene M. Häfliger https://orcid.org/0000-0002-5648-963X Luisa V. Muscatello https://orcid.org/0000-0003-4653-0624 Marilena Bolcato https://orcid.org/0000-0002-0605-3344 Riccardo Rinnovati https://orcid.org/0000-0002-4997-2340 American College of Veterinary Internal Medicine

Arcangelo Gentile D https://orcid.org/0000-0002-6091-8978 Cord Drögemüller D https://orcid.org/0000-0001-9773-522X

REFERENCES

- 1. Krakow D. Skeletal dysplasias. Clin Perinatol. 2015;42(2):301-319.
- Jacinto JGP, Häfliger IM, Letko A, Drögemüller C, Agerholm JS. A large deletion in the COL2A1 gene expands the spectrum of pathogenic variants causing bulldog calf syndrome in cattle. *Acta Vet Scand*. 2020;62(1):1-8.
- Agerholm JS, Menzi F, McEvoy FJ, Jagannathan V, Drogemuller C. Lethal chondrodysplasia in a family of Holstein cattle is associated with a de novo splice site variant of COL2A1. BMC Vet Res. 2016;12(1):1-9.
- Jacinto JGP, Häfliger IM, Gentile A, Drögemüller C, Bolcato M. A 6.7 kb deletion in the COL2A1 gene in a Holstein calf with achondrogenesis type II and perosomus elumbis. *Anim Genet*. 2021;52(2): 244-245.
- Bourneuf E, Otz P, Pausch H, et al. Rapid discovery of de novo deleterious mutations in cattle enhances the value of livestock as model species. *Sci Rep.* 2017;7(1):1-19.
- Häfliger IM, Letko A, Murgiano L, Drögemüller C. De novo stop-lost germline mutation in FGFR3 causes severe chondrodysplasia in the progeny of a Holstein bull. *Anim Genet*. 2020;51(3):466-469.
- Kromik A, Ulrich R, Kusenda M, et al. The mammalian cervical vertebrae blueprint depends on the T (brachyury) gene. *Genetics*. 2015; 199(3):873-883.
- Braun M, Lehmbecker A, Eikelberg D, et al. De novo ZIC2 frameshift variant associated with frontonasal dysplasia in a Limousin calf. BMC Genomics. 2021;22(1):1-9.
- Jacinto JGP, Häfliger IM, McEvoy FJ, Drögemüller C, Agerholm JS. A de novo mutation in COL1A1 in a Holstein calf with osteogenesis imperfecta type II. Animals. 2021;11(2):561.
- Petersen JL, Tietze SM, Burrack RM, Steffen DJ. Evidence for a de novo, dominant germ-line mutation causative of osteogenesis imperfecta in two red Angus calves. *Mamm Genome*. 2019;30(3–4):81-87. doi:10.1007/s00335-019-09794-4
- Hirano T, Matsuhashi T, Kobayashi N, Watanabe T, Sugimoto Y. Identification of an FBN1 mutation in bovine Marfan syndrome-like disease. *Anim Genet*. 2012;43(1):11-17.
- Jacinto JGP, Häfliger IM, Gentile A, Drögemüller C. A heterozygous missense variant in MAP2K2 in a stillborn romagnola calf with skeletal-cardio-enteric dysplasia. *Animals*. 2021;11(7):1931.
- Seichter D, Russ I, Förster M, Medugorac I. SNP-based association mapping of Arachnomelia in Fleckvieh cattle. *Anim Genet*. 2011;42(5): 544-547.
- Drögemüller C, Tetens J, Sigurdsson S, et al. Identification of the bovine arachnomelia mutation by massively parallel sequencing implicates sulfite oxidase (SUOX) in bone development. *PLoS Genet*. 2010; 6(8):e1001079.
- Charlier C, Agerholm JS, Coppieters W, et al. A deletion in the bovine FANCI gene compromises fertility by causing fetal death and brachyspina. *PLoS One.* 2012;7(8):e43085.
- Thomsen B, Horn P, Panitz F, et al. A missense mutation in the bovine SLC35A3 gene, encoding a UDP-N-acetylglucosamine transporter, causes complex vertebral malformation. *Genome Res.* 2006;16(1): 97-105.
- Hirano T, Kobayashi N, Matsuhashi T, et al. Mapping and exome sequencing identifies a mutation in the IARS gene as the cause of hereditary perinatal weak calf syndrome. *PLoS One.* 2013;8(5):e64036.
- Meyers SN, McDaneld TG, Swist SL, et al. A deletion mutation in bovine SLC4A2 is associated with osteopetrosis in red Angus cattle. BMC Genomics. 2010;11(1):337.
- Murgiano L, Militerno G, Sbarra F, et al. KDM2B-associated paunch calf syndrome in Marchigiana cattle. J Vet Intern Med. 2020;34(4): 1657-1661.

- Testoni S, Bartolone E, Rossi M, et al. KDM2B is implicated in bovine lethal multi-organic developmental dysplasia. *PLoS One*. 2012;7(9): 1-8.
- Mastrangelo S, Ciani E, Ajmone Marsan P, et al. Conservation status and historical relatedness of Italian cattle breeds. *Genet Sel Evol.* 2018;50(1):1-16.
- Drögemüller C, Drögemüller M, Leeb T, et al. Identification of a missense mutation in the bovine ATP2A1 gene in congenital pseudomyotonia of Chianina cattle: an animal model of human Brody disease. *Genomics.* 2008;92(6):474-477. doi:10.1016/j.ygeno.2008.07.014
- Charlier C, Coppieters W, Rollin F, et al. Highly effective SNP-based association mapping and management of recessive defects in livestock. Nat Genet. 2008;40(4):449-454.
- Jacinto JGP, Häfliger IM, Veiga I, Letko A, Gentile A, Drögemüller C. A frameshift insertion in FA2H causes a recessively inherited form of ichthyosis congenita in Chianina cattle. *Mol Genet Genomics*. 2021; 296(6):1313-1322.
- Rosen BD, Bickhart DM, Schnabel RD, et al. De novo assembly of the cattle reference genome with single-molecule sequencing. *Gigascience*. 2020;9(3):1-9.
- Hayes BJ, Daetwyler HD. 1000 bull genomes project to map simple and complex genetic traits in cattle: applications and outcomes. *Annu Rev Anim Biosci.* 2019;7:89-102.
- Chen S, Zhou Y, Chen Y, Gu J. Fastp: An ultra-fast all-in-one FASTQ preprocessor. *Bioinformatics*. 2018;34(17):i884-i890.
- Häfliger IM, Wiedemar N, Švara T, et al. Identification of small and large genomic candidate variants in bovine pulmonary hypoplasia and anasarca syndrome. *Anim Genet*. 2020;51(3):382-390.
- 29. Cingolani P, Platts A, Wang LL, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of Drosophila melanogaster strain w1118; iso-2; iso-3. *Fly* (*Austin*). 2012;6(2):80-92.
- Robinson JT, Thorvaldsdóttir H, Wenger AM, Zehir A, Mesirov JP. Variant review with the Integrative Genomics Viewer. *Cancer Res.* 2017;77(21):e31-e34.
- Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. *Gigascience*. 2015;4(1):7.
- Biscarini F, Cozzi P, Gaspa G, Marras G. DetectRUNS: an R package to detect runs of homozygosity and heterozygosity in diploid genomes. CRAN (the Comprehensive R Archive Network). 2019 Acceseed 4 Jul, 2023. https://CRAN.R-project.org/package= detectRUNS
- Bendl J, Stourac J, Salanda O, et al. PredictSNP: robust and accurate consensus classifier for prediction of disease-related mutations. *PLoS Comput Biol.* 2014;10(1):1-11.
- Adzhubei IA, Schmidt S, Peshkin L, et al. A method and server for predicting damaging missense mutations. Nat Methods. 2010;7:248-249.
- Ng PC, Henikoff S. SIFT: predicting amino acid changes that affect protein function. *Nucleic Acids Res.* 2003;31(13):3812-3814.
- Pejaver V, Urresti J, Lugo-Martinez J, et al. MutPred2: inferring the molecular and phenotypic impact of amino acid variants. *Nat Comm.* 2017;11:5918.
- Warde-Farley D, Donaldson SL, Comes O, et al. The GeneMANIA prediction server: biological network integration for gene prioritization and predicting gene function. *Nucleic Acids Res.* 2010;38-(Suppl_2):W214-W220.
- Jacinto JGP, Häfliger IM, Letko A, et al. Multiple independent de novo mutations are associated with the development of schistosoma reflexum, a lethal syndrome in cattle. Vet J. 2024;304:106069.
- Bateman A, Martin MJ, Orchard S, et al. UniProt: the universal protein knowledgebase in 2023. Nucleic Acids Res. 2023;51(D1):D523-D531.
- Paysan-Lafosse T, Blum M, Chuguransky S, et al. InterPro in 2022. Nucleic Acids Res. 2023;51(D1):D418-D427.

Journal of Veterinary Internal Medicine AC M |

n College of nternal Medicine 3357

- 41. Allard STM, Beis K, Giraud MF, et al. Toward a structural understanding of the dehydratase mechanism. *Structure*. 2002;10(1):81-92.
- Persson B, Kallberg Y, Bray JE, et al. The SDR (short-chain dehydrogenase/reductase and related enzymes) nomenclature initiative. Chem Biol Interact. 2009;178(1-3):94-98.
- Ehmke N, Caliebe A, Koenig R, et al. Homozygous and compoundheterozygous mutations in TGDS cause Catel-Manzke syndrome. *Am J Hum Genet*. 2014;95(6):763-770.
- 44. Schoner K, Bald R, Horn D, Rehder H, Kornak U, Ehmke N. Mutations in TGDS associated with additional malformations of the middle fingers and halluces: atypical Catel-Manzke syndrome in a fetus. *Am J Med Genet A*. 2017;173(6):1694-1697.
- Singh B, Fleury C, Jalalvand F, Riesbeck K. Human pathogens utilize host extracellular matrix proteins laminin and collagen for adhesion and invasion of the host. *FEMS Microbiol Rev.* 2012;36(6): 1122-1180.
- 46. Aumailley M. The laminin family. Cell Adh Migr. 2013;7(1):48-55.
- 47. Yue B. Biology of the extracellular matrix. J Glaucoma. 2014;23: S20-S23.
- Yousif LF, Di Russo J, Sorokin L. Laminin isoforms in endothelial and perivascular basement membranes. *Cell Adh Migr.* 2013;7(1):101-110.
- Knöll R, Postel R, Wang J, et al. Laminin-α4 and integrin-linked kinase mutations cause human cardiomyopathy via simultaneous defects in Cardiomyocytes and endothelial cells. *Circulation*. 2007;116(5): 515-525.
- Vinod E, Parameswaran R, Manickam Amirtham S, Livingston A, Ramasamy B, Kachroo U. Comparison of the efficiency of laminin versus fibronectin as a differential adhesion assay for isolation of human articular cartilage derived chondroprogenitors. *Connect Tissue Res.* 2021;62(4):427-435.
- Wilusz RE, Sanchez-Adams J, Guilak F. The structure and function of the pericellular matrix of articular cartilage. *Matrix Biol.* 2014;39: 25-32.
- Dong Y, Soung DY, Schwarz EM, O'Keefe RJ, Drissi H. Wnt induction of chondrocyte hypertrophy through the Runx2 transcription factor. *J Cell Physiol*. 2006;208(1):77-86.
- Später D, Hill TP, O'Sullivan RJ, Gruber M, Conner DA, Hartmann C. Wnt9a signaling is required for joint integrity and regulation of *lhh* during chondrogenesis. *Development*. 2006;133(15): 3039-3049.
- 54. Yuan C, Gou X, Deng J, Dong Z, Ye P, Hu Z. FAK and BMP-9 synergistically trigger osteogenic differentiation and bone formation of adipose derived stem cells through enhancing Wnt-β-catenin signaling. *Biomed Pharmacother*. 2018;105:753-757.

- 55. Li Q, Lan T, Xie J, Lu Y, Zheng D, Su B. Integrin-mediated tumorigenesis and its therapeutic applications. *Front Oncol.* 2022;11:12.
- Chen S, He T, Zhong Y, et al. Roles of focal adhesion proteins in skeleton and diseases. *Acta Pharm Sin B*. 2023;13(3):998-1013.
- Mukherjee S, Cogan JD, Newman JH, et al. Identifying digenic disease genes via machine learning in the undiagnosed diseases network. *Am J Hum Genet*. 2021;108(10):1946-1963.
- Lupski JR. Digenic inheritance and Mendelian disease. Nat Genet. 2012;44(12):1291-1292.
- 59. Knaust J, Hadlich F, Weikard R, Kuehn C. Epistatic interactions between at least three loci determine the "rat-tail" phenotype in cattle. *Genet Sel Evol.* 2016;48(1):26.
- Petropoulou E, Soltani M, Firoozabadi AD, et al. Digenic inheritance of mutations in the cardiac troponin (TNNT2) and cardiac beta myosin heavy chain (MYH7) as the cause of severe dilated cardiomyopathy. *Eur J Med Genet.* 2017;60(9):485-488.
- Gnanasekaran H, Chandrasekhar SP, Kandeeban S, et al. Mutation profile of Bardet-Biedl syndrome patients from India: implicative role of multiallelic rare variants and oligogenic inheritance pattern. *Clin Genet*. 2023;104(4):443-460.
- 62. Badano JL, Ansley SJ, Leitch CC, Lewis RA, Lupski JR, Katsanis N. Identification of a novel Bardet-Biedl syndrome protein, BBS7, that shares structural features with BBS1 and BBS2. *Am J Hum Genet*. 2003;72(3):650-658.
- Melluso A, Secondulfo F, Capolongo G, Capasso G, Zacchia M. Bardet-Biedl syndrome: current perspectives and clinical outlook. *Ther Clin Risk Manag.* 2023;19:115-132.
- Caspar SM, Dubacher N, Kopps AM, Meienberg J, Henggeler C, Matyas G. Clinical sequencing: from raw data to diagnosis with lifetime value. *Clin Genet*. 2018;93(3):508-519.

SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

How to cite this article: Jacinto JGP, Ogundipe TG, Benazzi C, et al. Familial osteochondrodysplastic and cardiomyopathic syndrome in Chianina cattle. *J Vet Intern Med*. 2024;38(6): 3346-3357. doi:10.1111/jvim.17221