REVIEW

Open Access

Tracking neuroinfammatory biomarkers in Alzheimer's disease: a strategy for individualized therapeutic approaches?

Simone Lista^{1*†}, Bruno P. Imbimbo^{2†}, Margherita Grasso³, Annamaria Fidilio³, Enzo Emanuele⁴, Piercarlo Minoretti⁵, Susana López-Ortiz¹, Juan Martín-Hernández¹, Audrey Gabelle⁶, Giuseppe Caruso^{3,7}, Marco Malaguti⁸, Daniela Melchiorri⁹, Alejandro Santos-Lozano^{1,10}, Camillo Imbimbo¹¹, Michael T. Heneka^{12*†} and Filippo Caraci^{3,7*†}

Abstract

Background Recent trials of anti-amyloid-β (Aβ) monoclonal antibodies, including lecanemab and donanemab, in early Alzheimer disease (AD) showed that these drugs have limited clinical benefits and their use comes with a signifcant risk of serious adverse events. Thus, it seems crucial to explore complementary therapeutic approaches. Genome-wide association studies identifed robust associations between AD and several AD risk genes related to immune response, including but not restricted to CD33 and *TREM2*. Here, we critically reviewed the current knowledge on candidate neuroinfammatory biomarkers and their role in characterizing the pathophysiology of AD.

Main body Neuroinfammation is recognized to be a crucial and contributing component of AD pathogenesis. The fact that neuroinfammation is most likely present from earliest pre-stages of AD and co-occurs with the deposition of Aβ reinforces the need to precisely defne the sequence and nature of neuroinfammatory events. Numerous clini‑ cal trials involving anti-infammatory drugs previously yielded unfavorable outcomes in early and mild-to-moderate AD. Although the reasons behind these failures remain unclear, these may include the time and the target selected for intervention. Indeed, in our review, we observed a stage-dependent neuroinfammatory process in the AD brain. While the initial activation of glial cells counteracts early brain Aβ deposition, the downregulation in the functional state of microglia occurs at more advanced disease stages. To address this issue, personalized neuroinfammatory modulation therapy is required. The emergence of reliable blood-based neuroinfammatory biomarkers, particularly glial fbrillary acidic protein, a marker of reactive astrocytes, may facilitate the classifcation of AD patients based on the ATI(N) biomarker framework. This expands upon the traditional classification of Aβ ("A"), tau ("T"), and neurodegeneration ("N"), by incorporating a novel infammatory component ("I").

[†]Simone Lista and Bruno P. Imbimbo contributed equally to this work.

† Michael T. Heneka and Filippo Caraci are co-senior authorship.

*Correspondence: Simone Lista slista@uemc.es Michael T. Heneka michael.heneka@uni.lu Filippo Caraci fcaraci@unict.it Full list of author information is available at the end of the article

© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit [http://creativecommons.org/licenses/by/4.0/.](http://creativecommons.org/licenses/by/4.0/)

Conclusions The present review outlines the current knowledge on potential neuroinfammatory biomarkers and, importantly, emphasizes the role of longitudinal analyses, which are needed to accurately monitor the dynamics of cerebral infammation. Such a precise information on time and place will be required before anti-infammatory therapeutic interventions can be considered for clinical evaluation. We propose that an effective anti-neuroinflammatory therapy should specifcally target microglia and astrocytes, while considering the individual ATI(N) status of patients.

Keywords Alzheimer's disease, Neuroinfammation, Biomarkers, GFAP, YKL-40, ATI(N) classifcation system, Microglia, Astrocytes, Longitudinal studies, Clinical trials

Introduction

Alzheimer's disease (AD), the predominant type of dementia, accounts for approximately two-thirds of all dementia cases in individuals aged 60 years and older [\[1](#page-19-0)]. At present, it afects a staggering 33 million people globally and continues to grow at an alarming rate, with its incidence doubling every 5–10 years [[1\]](#page-19-0). Notably, developing countries play a substantial role in the increasing incidence of new AD cases $[2]$ $[2]$. This trend may be attributed to the rapid growth of the older population in these regions, which is increasingly afected by dementia. Recently, there has been a signifcant increase in the focus on disease-modifying therapies that utilize monoclonal antibodies. One notable example is aducanumab, an anti-amyloid agent that received conditional approval from the U.S. Food and Drug Administration (FDA) in 2021 for the treatment of early-stage AD [\[3\]](#page-19-2). In 2023, the FDA approved lecanemab, a monoclonal antibody designed to target soluble amyloid-β (Aβ) protofbrils. This groundbreaking approval came after the successful outcomes of a phase III randomized, controlled clinical trial. The results demonstrated, for the first time, that reducing cerebral Aβ plaques through lecanemab led to a noticeable deceleration in cognitive decline over an 18-month treatment period [\[4](#page-19-3)]. A recent phase III clinical trial evaluated the efficacy of donanemab, a monoclonal antibody targeting a pyroglutamate form of Aβ, in individuals with prodromal AD and mild dementia due to AD $[5]$. The study revealed that donanemab efectively slowed both cognitive and functional decline. However, the magnitude of the clinical efects observed with donanemab, along with other similar drugs, such as aducanumab and lecanemab, was limited. This suggests that additional mechanisms, including neuroinfammation [[6\]](#page-19-5), tau processing [\[7](#page-19-6)], apolipoprotein E (APOE) isoforms imbalance [[8\]](#page-19-7), mitochondrial dysfunction [[9\]](#page-19-8), and synaptic degeneration $[10]$ $[10]$, should be explored to fully understand and address the pathogenesis of AD. Neuroinfammation refers to the activation of the brain's innate immune system in response to infammatory challenges such as injury, infection, toxin exposure, neurodegenerative diseases, or aging. Microglia, the innate immune cells of the central nervous system (CNS), are pivotal in mediating these neuroinfammatory responses [\[11\]](#page-19-10). Activated microglia and reactive astrocytes can phagocytize senile plaques or dystrophic neurites, induce intraneuronal infammatory reactions towards neurofbrillary tangles, and activate the complement cascade in response to vascular amyloid, thereby contributing to cerebral amyloid angiopathy.

Currently, there are 164 ongoing clinical trials in phase I, II, and III, evaluating the efectiveness of 127 distinct drugs. Interestingly, in phase II, approximately 23% of these compounds are specifcally targeting infammatory mechanisms $[12]$ $[12]$. These efforts highlight the urgent requirement for innovative pharmacological treatments that can efectively prevent or delay the onset of dementia, while also signifcantly slowing down the progression of the disease. However, infammatory mechanisms may cycle between infammation and resolution, and also convert into a chronic type which means that any intervention will require a precise knowledge on the nature and site of infammation to target.

In this context, the identifcation of reliable biomarkers of the initial pathological processes assumes paramount signifcance in disease management during all stages. Conducting biomarker studies becomes imperative in unraveling the intricate interplay between specifc immune and/or infammatory molecules in the development and progression of AD clinical manifestations. Employing longitudinal biomarker studies can unveil varying expression patterns in the initial stages of AD pathology and thus potentially shed light on variances in treatment response $[13]$. Efforts are currently underway to identify and validate innovative blood-based biomarkers that can efectively refect the pathophysiological mechanisms associated with AD at a peripheral level. These biomarkers offer several advantages, such as being non-invasive and well-tolerated compared to brain imaging techniques and cerebrospinal fuid (CSF) biomarkers [[14](#page-19-13)], which together will ease longitudinal assessment over years and decades. Plasma levels of hyperphosphorylated tau at position 217 (p-tau₂₁₇) have demonstrated clinical performance that is equivalent to or superior to the FDA-approved CSF tests in detecting brain

Aβ pathology [\[15,](#page-19-14) [16\]](#page-19-15). Plasma neuroflament light chain (NfL) protein, which is a scafolding cytoskeleton protein released when neurons are damaged. Research has shown that plasma NfL concentrations can efectively predict brain imaging biomarkers of neurodegeneration and initial cognitive decline in middle-aged individuals [[17](#page-19-16)].

Currently, there are several fuid biomarkers available for detecting AD dementia development. The $AT(N)$ biomarker framework, which assesses the brain deposition of Aβ ("A"), tau pathology ("T"), and neurodegeneration ("N"), can be further expanded to include neuroinfammatory $('T")$ candidate biomarkers, resulting in an ATI(N) system [[18](#page-19-17), [19](#page-19-18)]. By monitoring activated microglia and reactive astrocytes, CSF and blood "I" biomarkers enable the tracking of neuroinfammatory processes [\[20\]](#page-19-19).

In this review, we will provide an overview of the involvement of microglia and astrocytes in the neuroinfammatory processes that impact the AD brain. Furthermore, we will conduct a thorough evaluation of the key studies focusing on biomarkers that track the activation of microglial cells and reactive astrocytes. We will also assess the ability of longitudinal neuroinfammatory biomarker studies to predict the onset of AD and cognitive decline (see Table [1](#page-3-0)). Lastly, we will analyze the signifcance of neuroinfammatory biomarkers in the diagnosis of AD and their role in AD clinical trials.

Search strategy and selection criteria

This non-systematic literature review aims to provide an informative overview of the current state of biomarkers for neuroinflammation in AD. The manuscript is based on a selective analysis of high-quality, contemporary articles on neuroinflammation biomarkers in AD. The primary objective is to identify trends and enhance understanding of the current landscape of neuroinfammation biomarkers in AD. References for this review were identifed through searches of PubMed databases for peer-reviewed articles published in English between January 1, 2013, and December 31, 2023. The search terms employed included "Neuroinfammation" and "Alzheimer," "longitudinal studies" and "Alzheimer," "TREM2" and "Alzheimer," "GFAP" and "Alzheimer," and "YKL40" and "neurodegeneration." Additionally, bibliographies of relevant papers were reviewed. Only papers published in English were considered for inclusion. The final list of references was selected by SL, MG, and AF, and validated by BPI.

The role of microglia and astrocytes in Alzheimer's disease pathophysiology

Astrocytes and microglia, the brain-resident macrophages, are vital components in the development of neural circuits. These dynamic cells establish bidirectional communications with synapses, exerting a

profound infuence on synaptic function. Contrary to popular belief, synaptic information processing is not solely dependent on neurons. Astrocytes envelop synapses and microglia interact with synapses in an activitydependent manner $[21]$ $[21]$ $[21]$, collectively contributing to the intricate network of neural connectivity. Prior studies [[22,](#page-19-21) [23\]](#page-19-22) have also demonstrated that astrocytes, microglia, and synapses interact in a "quad-partite" model, where the axon terminal and dendritic spine communicate directly with microglial and astrocytic processes. Disruptions to this quad-partite arrangement can lead to abnormal plasticity, which consequently afects the encoding of information in neuronal circuits (Fig. [1\)](#page-6-0).

Astrocytes play a crucial role in the formation of synapses and regulating the release of neurotransmitters, thereby maintaining the balance of glutamate in the brain. This function of astrocytes is essential for promoting various physiological activities associated with synaptic plasticity and, consequently, cognitive function [\[24](#page-19-23)]. Moreover, astrocytes can facilitate neuroinfammatory processes through the release of infammatory cytokines and chemokines. These cells are also involved in the clearance of Aβ, which subsequently activates them, leading them to encircle senile plaques. This, in turn, contributes to Aβ-induced damage to the BBB, ultimately depriving neurons of their metabolic supply [[25\]](#page-19-24).

Microglia cells, which represent approximately 5–20% of all glial cells, serve as the main type of macrophages in the CNS. Their primary function is to regularly survey brain regions for pathogens and cellular debris, ensuring the preservation of neuronal circuits. Additionally, microglia protect and remodel synapses to support brain function $[26]$ $[26]$. These cells express various receptors that detect both internal and external insults to the CNS. When triggered by pathological factors such as protein aggregates or neuronal death, microglia migrate to the site of injury and initiate innate immune responses [[27\]](#page-19-26).

During the inflammatory processes involved in the pathogenesis of AD, there is a transition from the resting to the active functional state of microglia. Inflammation is primarily triggered by the accumulation of Aβ aggregates, including soluble oligomers and insoluble fibrils $[28]$. In the early stages of AD, A β oligomers and fibrils build up in the extracellular space, initiating a pathological cascade that leads to neuronal apoptosis and depletion. Microglia also play a crucial role in clearing Aβ oligomers, fibrils, and dead cells through phagocytosis and by secreting proteolytic enzymes. Additionally, microglia surround plaques and fibrils, forming a barrier that prevents their spread and limits their toxicity [[29\]](#page-19-28). As $\text{A}\beta$ deposition becomes increasingly severe, microglia undergo a transition from their normal, homeostatic state to a dysfunctional

Table 1 Main characteristics of selected longitudinal studies on neuroinfammatory biomarkers in AD (n Table 1 Main characteristics of selected longitudinal studies on neuroinflammatory biomarkers in AD (n=27)

Lista *et al. Journal of Neuroinflammation (2024) 21:187* Page 6 of 24

Fig. 1 A Pathological interactions between glia and neurons in AD. Amyloid-beta species can be recognized by and activate microglia, which contribute to activate astrocytes. These cells release pro-infammatory cytokines and chemokines and lose their homeostatic functions, leading to disruption of the BBB and neuronal damage. Defcit of anti-infammatory cytokines (e.g. TGF-β1) also contributes to microglial activation and synaptic dysfunction. Disease associated microglia can also directly eliminate synaptic structures. **B** "Eat-me" signals and the quad-partite synapse in AD. Neuronal eat-me signals are recognized by several phagocytic receptors on both astrocytes and microglia. Some of these signals include milk fat globule-EGF factor 8 protein (MFGE8) and Protein S, both bound to exposed phosphatidylserine, and complement component 3 (C3) and 1q (C1q). Microglia use as cell-surface receptors the complement receptor 3 (CR3), vitronectin receptor (VNR), MER receptor tyrosine kinase (MERTK) and triggering receptor expressed on myeloid cells 2 (TREM2) among others. Astrocytes may also contribute to synaptic engulfment via the binding of C1q to multiple EGF-like domains 10 (MEGF10). Metabotropic glutamate receptor 5 (mGluR5) may also induce synaptotoxicity via calcium dysregulation and complement activation, making the synapses a target for removal by astrocytes and microglia. Interactions between α7 nicotinic acetylcholine receptors (α7nAChRs) and soluble amyloid-β (Aβ) can also contribute to reactive astrogliosis and neuronal death, through the excessive release of glutamate from astrocytes in a Ca2+ -dependent manner. *α7nAChRs* α7 nicotinic acetylcholine receptors, *Aβ* amyloid-β, *AD* Alzheimer disease, *BBB* blood–brain barrier, *C1q* complement component, *C3* complement component 3, *CR3* complement receptor 3 (CR3), *IL* interleukin, *MEGF10* multiple EGF-like domains 10, *MERTK* MER, receptor tyrosine kinase, *MFGE8* milk fat globule-EGF factor 8 protein, *mGluR5* Metabotropic glutamate receptor 5, *TGF-β1* transforming growth factor-beta 1, *TNF* tumor necrosis factor, *TREM2* triggering receptor expressed on myeloid cells 2, *VNR* vitronectin receptor

phenotype. This shift prompts the release of proinflammatory factors, oxidative stress, and neuroinflammation, while also reducing the secretion of neurotrophic factors. Ultimately, this leads to synaptic impairment and the exacerbation of neuronal damage [[30](#page-19-29), [31](#page-19-30)]. Despite the potential benefits of early activation, chronic activation of microglia by Aβ can be detrimental. This leads to amplified Aβ deposition and prolonged inflammatory processes, ultimately accelerating neurodegeneration. In advanced stages of the disease, microglia can exacerbate AD by circulating proinflammatory cytokines—such as interleukin-1β (IL-1 β) and tumor necrosis factor-alpha (TNF- α) which cause neuronal cell death, as well as by stimulating astrocytes, which can impact neuronal survival [[26](#page-19-25)].

Biomarkers of neuroinfammation in Alzheimer's disease

Biomarkers play a crucial role in enhancing our understanding of the molecular mechanisms underlying the onset and progression of AD. Specifcally, plasma biomarkers offer a convenient method to evaluate individuals at various stages of the disease continuum, including healthy individuals, those at risk of developing AD, and patients with AD. By monitoring these biomarkers over time, we can gain valuable insights into the individual's longitudinal progression within the AD spectrum.

As of 2009, genome-wide association studies (GWAS) led to the identifcation of novel genetic associations, revealing genome-wide statistically signifcant links between AD and variants within the *CLU*, *PICALM*, and *CR1* genes. Since then, more than 50 risk loci and over

70 gene variants associated with an increased risk of developing sporadic late-onset AD (LOAD) have been identified $[32]$ $[32]$ $[32]$. These findings underscore the interconnected network of molecular and cellular pathways that signifcantly infuence the progression and pathogenesis of AD. Interestingly, several of these identifed variants are intertwined with genes that play a crucial role in immune responses and inflammation. These include, but are not limited to, *TREM2, CD33, PILRA, CR1, MS4A, CLU, ABCA7, EPHA1*, and *HLA-DRB1* [[23,](#page-19-22) [33](#page-19-32)]. The majority of these genes play signifcant roles in various functions such as proinfammatory intracellular signaling, cytokines/interleukins/cell turnover, synaptic activity, lipid metabolism, and vesicle trafficking. The crucial involvement of neuroinfammation is supported by extensive GWAS studies that reveal a notable rise in the likelihood of developing LOAD in individuals carrying rare variants of microglial immunoreceptors.

CD33 encodes a transmembrane receptor expressed on myeloid lineage cells, functioning as a sialic acid-binding immunoglobulin-like lectin that modulates innate immunity. The minor allele of the CD33 SNP rs3865444, which confers protection against AD, has been associated with reduced levels of insoluble Aβ42 in AD brains. In microglial cell cultures, CD33 inhibits the uptake and clearance of $Aβ42$. Therefore, CD33 inactivation has been shown to mitigate Aβ pathology, suggesting that CD33 inhibition could represent a novel therapeutic strategy for AD [\[34](#page-19-33)]. *PILRA* encodes the paired immunoglobulin-like type 2 receptor alpha, a cell surface inhibitory receptor that recognizes specifc O-glycosylated proteins. PILRA is expressed on various innate immune cell types, including microglia. It has been proposed that the common *PILRA* missense variant (G78R, rs1859788) protects individuals from AD risk by reducing inhibitory signaling in microglia and microglial infection during herpes simplex virus 1 (HSV-1) recurrence [\[35](#page-19-34)]. Complement component (3b/4b) receptor 1 (*CR1*) is a notable candidate gene with a signifcant connection to AD. Polymorphisms in *CR1* have been reported to be associated with LOAD susceptibility. A recent review identifed that the rs6656401 variant in CR1 increases the risk of LOAD [[36\]](#page-19-35). Additionally, a common variant in the membrane-spanning 4-domains subfamily A (*MS4A*) gene, specifcally MS4A4A, has been linked to increased CSF sTREM2 concentrations and a reduced risk of AD [\[37](#page-19-36)]. The *CLU* gene, which contains several AD-associated intronic SNPs, encodes clusterin, a secretory protein predominantly synthesized in astrocytes. Clusterin levels are elevated in the brain tissues, CSF, and plasma of AD patients and may play antiamyloidogenic roles [[38\]](#page-19-37).

Rare variants of the ATP-binding cassette sub-family A member 7 (*ABCA7*) gene are signifcantly enriched in patients with AD, while a common *ABCA7* missense variant may confer protection against the disease. Methylation at several CpG sites within the *ABCA7* locus is signifcantly associated with AD and correlates with amy-loid deposition and brain morphology [[39](#page-20-3)]. The EPHA1 gene encodes ephrin type-A receptor 1 (EphA1), a member of the ephrin receptor subfamily within the proteintyrosine kinase family. Ephrin receptors and related proteins have been implicated in mediating immune cell recruitment. The P460L variant of the *EPHA1* gene is associated with an increased risk of LOAD and has recently been shown to impact immune responses and blood vessel function in the brain [[40\]](#page-20-4). HLA-DRB1 is part of the human leukocyte antigen (HLA) class II beta chain paralogues. This class II molecule is a heterodimer composed of an alpha (DRA) and a beta (DRB) chain, both of which are membrane-anchored. HLA-DRB1 plays a pivotal role in the immune system by presenting peptides derived from extracellular proteins. Specifc HLA-DRB1*04 alleles have been shown to confer protection against AD [[41](#page-20-5)]. Below, we provide an in-depth description of the TREM2 protein and other signifcant neuroinfammatory proteins.

Soluble triggering receptor expressed on myeloid cells 2 (sTREM2)

TREM2, a cell surface receptor, is predominantly expressed on microglial cells. Its primary function includes enhancing the phagocytic capabilities of microglia and macrophages while also modulating infammatory signaling. TREM2 transduces its intracellular signaling through the adapter protein DAP12 $[42]$ $[42]$. The binding of TREM2 to ligands, including anionic lipids, lipoproteins, and Aβ, initiates downstream signaling cascades that promote microglial survival, proliferation, chemotaxis, and phagocytosis $[43, 44]$ $[43, 44]$ $[43, 44]$ $[43, 44]$ $[43, 44]$. The significance of TREM2 in the pathogenesis of AD was underscored by the discovery of a specifc heterozygous mutation, R47H, which substantially elevates the risk of developing LOAD [[45,](#page-20-9) [46](#page-20-10)]. Subsequent research has established that TREM2 activation in microglia plays a crucial role in the pathological processes underlying AD [\[47\]](#page-20-11). This receptor also plays a pivotal role in regulating microglial-related activities, particularly in response to the presence of Aβ plaques and tau tangles [[48\]](#page-20-12). A soluble form of TREM2 (sTREM2), which is generated through the proteolytic cleavage of the receptor found on the surface of cells, has the ability to promote microglial activation. Proper receptor ligation is anti-infammatory and facilitates microglial survival.

TREM2 gene variants have been found to increase the risk of developing AD by impairing the ability of microglia cells to efectively clear Aβ and disrupting the normal proinfammatory response of these immune cells [\[49](#page-20-13)]. Currently, approximately 50 variants of the *TREM2* gene have been studied in relation to AD [[50](#page-20-14)]. The rare *R47H* variant of *TREM2* (rs75932628) has been linked to a two- to three-fold increase in the risk of developing AD in European and North Ameri-can (Caucasian) populations [\[51](#page-20-15)[–53](#page-20-16)]. The *R47H* variant is believed to contribute to a loss of TREM2 function, leading to a reduction of microglia and an increase in neuritic dystrophy at the site of Aβ deposition [[54\]](#page-20-17). A meta-analysis involving more than 73,000 participants found that individuals carrying this variant had a four-fold higher risk of developing AD compared to non-carriers, which is similar to the efect size of the *APOE ε4* variant [[55\]](#page-20-18). Additionally, it has been suggested that the *R47H* variant may be associated with an earlier age at AD onset [[56\]](#page-20-19). However, this variant is extremely rare or undetectable in African-American [[57](#page-20-20)] and Asian populations [[58,](#page-20-21) [59](#page-20-22)]. Another variant, *R62H* (rs143332484) [[60\]](#page-20-23), has been associated with an increased risk of AD in individuals of European descent [[61](#page-20-24)]. Interestingly, patients with AD harboring *TREM2* risk variants show an abundance of autophagic vesicles in their microglia [[62\]](#page-20-25).

An additional variant, *H157Y* (rs2234255), is located at the TREM2 site and is cleaved by two α-secretases (a disintegrin and metalloproteinase 10 and 17 [ADAM-10 and ADAM-17]), resulting in increased shedding of TREM2 and reduced cell surface expression of the receptor $[63]$ $[63]$. Unlike the *R47H* and *R62H* variants, *H157Y* is more frequently observed in Asians and is particularly associated with an increased risk of AD in a Han Chinese cohort [[64,](#page-20-27) [65](#page-20-28)]. Prior studies conducted on Caucasian, Japanese, and African-American cohorts did not fnd any signifcant association between this variant and AD. However, a comprehensive meta-analysis revealed a strong correlation with an odds ratio of 3.65 [\[65](#page-20-28)]. Additionally, further analyses conducted on Chinese cohorts identifed the presence of the p.Ala130Val and p.Ala192Thr variants, specifcally observed in cases of LOAD. Another variant, p.Ser183Cys, was found to be more prevalent among Chinese patients with AD $[50]$ $[50]$. The presence of multiple variants associated with a higher susceptibility to developing AD in Chinese and African-American populations highlights the presence of diverse mutations across different cohorts and ethnic groups. This underscores the need to investigate various ethnic populations to discover specifc disease risk variants and explore potential associations between these variants and specifc disease phenotypes [[50\]](#page-20-14). Notably, individuals carrying *TREM2* variants exhibit an earlier onset of the disease and experience faster cerebral atrophy. Therefore, identifying *TREM2* carriers can be valuable in improving patient stratifcation for clinical trials and supporting the development of personalized therapeutic approaches [\[33](#page-19-32)].

In general, the published literature has shown a slight increase in CSF sTREM2 concentrations in patients with AD [\[66](#page-20-29)[–68](#page-20-30)] and individuals with mild cognitive impairment (MCI) [\[69](#page-20-31)] compared to CU controls. However, other studies did not fnd signifcant diferences in CSF sTREM2 levels between AD or MCI participants and CU controls [[70](#page-20-32)]. Notably, increasing CSF sTREM2 concentrations were associated with changes in brain structure, specifcally an increase in grey matter volume in regions such as the bilateral inferior and middle temporal cortices, precuneus, supramarginal and angular gyri, in individuals with MCI. This suggests that sTREM2 may play a role in modulating neuroinfammatory responses to early neurodegeneration [[71\]](#page-20-33).

Despite the fact that CSF sTREM2 values are higher in patients with AD compared to CU controls, this potential biomarker lacks sufficient discriminatory power. Specifically, it falls short of the 80–90% range required in clinical diagnostic practice to efectively diferentiate between patient groups [[69](#page-20-31)]. However, the increase in CSF sTREM2 concentrations during the early stages of AD may indicate a change in microglial activation status due to neurodegeneration. This suggests that sTREM2 exhibits a dynamic response linked to microglial activity as the disease progresses. These findings highlight the potential of CSF sTREM2 as a biomarker for tracking the progression from preclinical AD/MCI to AD dementia [\[68,](#page-20-30) [72](#page-20-34)]. Consequently, it may also serve as a valuable biomarker in clinical trials focusing on secondary AD prevention. The dynamics of CSF sTREM2 should be investigated in relation to the key pathophysiological mechanisms of AD, namely Aβ and tau pathologies. Decreased levels of CSF sTREM2 are linked to Aβ pathology [[73](#page-20-35)] while increased levels are associated with tau-related neurodegeneration [\[74\]](#page-20-36). In a cross-sectional study of 127 individuals with autosomal dominant AD (ADAD) mutations, it was observed that CSF sTREM2 concentrations began to rise fve years before symptom onset, but quite a long time after the accumulation of $Aβ$ in the brain [[72](#page-20-34)].

Signifcant insights have also been gained from longitudinal studies on CSF sTREM2. For instance, a 1.5-year study involving 268 CU individuals with initial brain Aβ deposition revealed that a decline in basal forebrain volume was associated with a greater accumulation of CSF sTREM2 over time [[75](#page-20-2)]. In another longitudinal study spanning 4 years and including 72 individuals with MCI, it was found that CSF sTREM2 levels correlated with the progression of CSF Aβ and tau [\[76](#page-20-1)]. Additionally, a 3-year study on 231 AD patients showed that higher CSF sTREM2 levels at baseline were linked to slower clinical progression [\[77\]](#page-20-0). Finally, a 3-year study on 148

pre-symptomatic ADAD patients indicated that higher annual rates of increase in CSF sTREM2 corresponded to a reduced rate of cognitive decline $[78]$. These studies suggest that sTREM2 should be considered not an early marker, but indicative of the MCI to AD conversion, although further studies are needed to validate its role as biomarker in this scenario.

Limited reports have investigated the dynamics of TREM2 in blood. However, the available research suggests that both TREM2 mRNA and protein expression levels are elevated in monocytes of patients with AD compared to controls. Furthermore, these increased levels were found to be inversely correlated with cognitive performance, as measured by the Mini-Mental State Examination (MMSE). A positive correlation between TREM2 mRNA and protein expression was observed in monocytes. In addition, a tendency towards upregulation of TREM2 protein was also noticed in granulocytes and plasma [\[79](#page-21-11)]. Other studies have demonstrated higher expression levels of TREM2 mRNA in leukocytes of patients with AD than in controls, which were found to be associated with cognitive decline and hippocampal atrophy [[80,](#page-21-12) [81\]](#page-21-13). In a longitudinal study involving 57 individuals with MCI, higher plasma TREM2 levels and mRNA expression were detected in peripheral blood mononuclear cells (PBMCs) of *APOE ε4* positive individuals who later developed AD during the two-year follow-up period [\[82\]](#page-21-0). Another investigation focusing on peripheral leukocytes revealed higher expression levels of TREM2 in LOAD compared to early-onset AD. Interestingly, the expression of this receptor was markedly increased in LOAD individuals who carried the *APOE ε4* allele [\[83\]](#page-21-14).

Chitinase-3-like protein 1 (YKL-40)

The glycoprotein YKL-40, also known as chitinase-3-like protein 1 (CHI3L1), is detected primarily in various cell types, including macrophages, chondrocytes, fbroblasts, vascular smooth muscle cells, endothelial cells, and even certain cancer cells [[84\]](#page-21-15). Expression is most prominent in reactive astrocytes [[85\]](#page-21-16). While the exact physiological role of YKL-40 is still debated, there is consensus regarding its involvement in tissue remodeling and renovation during infammation, as well as in angiogenic mechanisms that afect the infltration, diferentiation, and maturation of macrophages. Consequently, YKL-40 is considered a candidate biomarker for infammation and endothelial dysfunction [\[84](#page-21-15)], a specifc biomarker for human macrophage activation/diferentiation and its expression is induced in reactive astrocytes by the presence of activated macrophages [[86\]](#page-21-17). A critical review of the literature has identifed YKL-40 as a pathophysiological biomarker indicating the activation of glial cells,

including astrocytes and microglia, which are associated with tau pathology [\[87](#page-21-18), [88](#page-21-19)]. In addition, prior studies have shown that CSF levels of YKL-40 are linked to biomarkers of neurodegeneration (total tau, t-tau), damage to large-caliber myelinated axons (NfL), tau-mediated toxicity (p-tau), and synaptic damage (neurogranin, SNAP-25) in various neurodegenerative diseases [\[89\]](#page-21-20).

Interest in YKL-40 in AD was sparked by initial observations that CSF levels of YKL-40 were signifcantly higher in individuals with AD and MCI compared to controls [\[90](#page-21-21), [91](#page-21-3)]. Notably, early histopathological studies have demonstrated that YKL-40 expression is localized near $\text{A}\beta$ plaques and tau neurofibrillary tangles [[92\]](#page-21-2). YKL-40 can effectively differentiate patients with overt dementia from CU individuals [\[91–](#page-21-3)[94\]](#page-21-22). Secondly, YKL-40 serves as a predictor of cognitive decline, allowing for the identifcation of individuals progressing from preclinical AD to prodromal AD and later stages of dementia. Finally, it aids in distinguishing individuals with MCI who will convert to AD from those who will remain stable at fve years [[86\]](#page-21-17). However, it should be noted that CSF YKL-40 is not clinically useful in diferentiating the characteristic AD phenotype (i.e., amnestic syndrome of hippocampal type) from other atypical presentations of AD [[95\]](#page-21-23). In addition, patients exhibited a signifcant correlation with the corresponding CSF values. Signifcantly, patients in a 6-year longitudinal study by Craig-Schapiro and colleagues (2010) exhibited a strong association between YKL 40 levels in CSF and plasma [[92\]](#page-21-2). Moreover, plasma YKL-40 concentrations were found to be elevated in individuals with very mild and mild AD dementia ($CDR = 0.5$ and $CDR = 1$, respectively) compared to control individuals $(CDR=0)$ [\[92](#page-21-2)]. Another study discovered a signifcant increase in plasma YKL-40 levels in patients with early AD than in individuals with MCI and healthy older controls. Moreover, YKL-40 demonstrated a positive correlation with neuropsychological test results in both MCI and early AD [[96\]](#page-21-24). In a longitudinal study conducted by Vergallo et al. (2020) [\[97](#page-21-5)], it was found that plasma YKL-40 concentration can serve as a valuable biomarker to assess the severity of AD. The investigation focused on a cohort of CU individuals at risk for AD and revealed a positive association between plasma YKL-40 and episodic memory performance (assessed using the Free and Cued Selective Rating Test). Conversely, a negative association was observed between plasma YKL-40 and brain Aβ accumulation [[97\]](#page-21-5). These fndings suggest that glia activation, refected by elevated YKL-40 levels, may have a potentially protective efect on initial brain Aβ deposition and neuronal homeostasis, without causing clinical harm. Moreover, the study also observed a sexual dimorphism, with men exhibiting higher YKL-40 concentrations than women [[97\]](#page-21-5). Another

study investigating biomarker diferences across diferent ethnicities reported elevated plasma YKL-40 concentrations specifcally in Hispanic women with prodromal AD compared to both CU controls and prodromal AD participants of African or Caucasian origins. This emphasizes the impact of potential modulating factors on the variability of YKL-40 levels [\[98](#page-21-25)].

YKL-40 has also emerged as a compelling candidate biomarker for investigating the clinical evolution of AD. Its potential role in clinical trials lies in its ability to track the dynamics of glial neuroinfammatory mechanisms in relation to neurodegeneration. According to Baldacci et al. (2017), CSF YKL-40 exhibits a correlation with elevated levels of CSF t-tau, even in asymptomatic and preclinical AD individuals. This suggests an early association between YKL-40 and tau protein during the course of neurodegeneration [\[86](#page-21-17)].

In conclusion, exploring the connection between YKL-40 and AD features could shed light on the intricate relationship between Aβ dysmetabolism, neuronal activity, and neuroinfammation. YKL-40 thus represents a promising biomarker for accurate classifcation of neuroinfammatory phenotypes, facilitating advancements in neuroinfammatory clinical trials [\[97](#page-21-5)].

Glial fbrillary acidic protein (GFAP)

Astrogliosis refers to the abnormal activation and proliferation of astrocytes that occurs in response to initial brain damage [[99\]](#page-21-26). This process leads to significant cellular, molecular, and functional changes. In acute brain injuries—as well as in AD, other neurodegenerative diseases, and low-grade astrocytoma—astrocytes adopt a reactive phenotype [\[99\]](#page-21-26). Transcriptomic analyses of human patients and disease models have demonstrated the presence of multiple putative reactive astrocyte substates. In CNS disorders, there is a marked upregulation and reorganization of intermediate flament proteins, leading to the formation of an intricate network comprising various isoforms of GFAP (ten isoforms), vimentin, synemin, and nestin [\[100\]](#page-21-27). GFAP, a type III intermediate flament protein, serves as a key cytoskeletal component in astrocytes. In AD, astrocytes exhibit a complex response to both neurofbrillary tangles and Aβ plaques, which may exert either neuroprotective or deleterious effects $[101]$ $[101]$ $[101]$. The capacity of astrocytes to colocalize with Aβ plaques in the AD brain has been demonstrated using labeled tracers [[102](#page-21-29)]. Moreover, studies have reported a signifcant correlation between GFAP expression levels and the density of Aβ plaques in the hippocampus and entorhinal cortex of AD brains [[103](#page-21-30)]. Interestingly, elevated concentrations of GFAP within the CSF have been reported in patients with AD and other forms of dementia compared to healthy individuals [[104](#page-21-31)]. An increase in plasma GFAP concentrations was also noted in both early-onset AD and LOAD [\[105,](#page-21-32) [106\]](#page-21-33). This increase was positively correlated with the extent of white matter injury, as determined through the quantifcation of white matter hyperintensities [[105](#page-21-32), [106\]](#page-21-33), whereas cognitive function assessed with MMSE showed an inverse association [\[106](#page-21-33)]. When combined with plasma $Aβ_{1-42}$ / Aβ₁₋₄₀ ratio, the *APOE ε4* status, and/or age, the diagnostic value of plasma GFAP was found to be increased [[107–](#page-21-34)[109](#page-21-8)].

The potential of blood GFAP as a biomarker for AD has been signifcantly highlighted by longitudinal investigations. A study conducted over 3 years on 23 asymptomatic AD patients revealed an increase in plasma GFAP concentrations in mutation carriers compared to noncarrier controls. This suggests that plasma GFAP alterations can be detected up to a decade before the onset of AD clinical symptoms [\[110](#page-21-10)]. Additionally, a 6-year longitudinal study on 106 individuals with MCI found that baseline serum GFAP levels were signifcantly higher in patients who progressed to AD at follow-up [\[111](#page-21-9)]. In a cross-sectional analysis conducted in preclinical AD, a plasma biomarker panel consisting of GFAP, p-tau₁₈₁, and p-tau₂₃₁ exhibited increased levels in CU Aβ-positron emission tomography (PET)**-**positive individuals compared to those who tested negative [\[112](#page-21-7)]. The study subsequently confirmed the longitudinal predictive value of two of the three biomarkers (GFAP and p-tau₁₈₁), emphasizing their diagnostic and monitoring potential in preclinical AD [[112\]](#page-21-7). In a 2-year, longitudinal study, researchers examined 288 CU individuals and 141 patients. The study found that plasma GFAP levels were associated with both longitudinal Aβ-PET deposition and cognitive decline [[113\]](#page-21-6). Another extensive 2-year longitudinal study involving 1,106 CU participants revealed that Aβ-dependent tau accumulation occurred only in individuals who tested positive for astrocyte reactivity—which was defned by plasma GFAP levels above a specified cutoff (mean $+2.0$ standard deviations of con-trols without Aβ pathology) [\[114\]](#page-22-2). These data suggest that secondary astrocytosis caused by Aβ aggregation might promoting tau accumulation, although further longitudinal studies are needed. A 5-year longitudinal study involving 169 individuals with MCI showed that higher baseline plasma GFAP concentrations were associated with the progression to AD and faster rates of cognitive decline [\[109](#page-21-8)]. Another 3-year longitudinal study conducted on 300 CU older individuals found that higher serum GFAP levels at baseline were linked to an increased risk of incident dementia [[115](#page-22-0)]. An analysis conducted within the ESTHER cohort, a German population-based study of older individuals living in the community, revealed a substantial early association

(spanning between 9 and 17 years prior to clinical diagnosis) of plasma GFAP with the incidence of AD. Notably, this association was found to be signifcantly earlier than that of NfL and p -tau₁₈₁, which were typically associated within approximately 9 years of diagnosis [\[116](#page-22-1)]. These results suggested that GFAP may serve as a more efective prognostic biomarker for incident AD dementia compared to NfL $[115]$. The diverse prognostic values of these cytoskeletal proteins are believed to stem from different underlying mechanisms. Notably, NfL is released into the bloodstream following axonal degeneration, while increased levels of GFAP are a response to damage triggered by Aβ and tau aggregates $[101]$ $[101]$. Continual activation of astrocytes in response to this damage leads to a pro-infammatory neurotoxic state, which further exacerbates neurodegeneration [\[117](#page-22-8)].

A recent 17-year longitudinal study involving 1712 CU participants found that serum GFAP levels at baseline were associated with a hazard ratio of 1.38 (95% confdence interval=1.15–1.66) for incident dementia and 2.76 (95% confidence interval $=$ 1.73–4.40) for dementiaspecifc mortality, supporting the notion that circulating GFAP can be a valuable tool for assessing dementia risk and prognosis [[118\]](#page-22-3). In another recent study involving 318 CU participants, including 158 individuals who later converted to AD and 160 who remained cognitively unimpaired, the authors observed elevated plasma levels of GFAP in AD-converters up to 10 years before the onset of cognitive impairment $[119]$ $[119]$. This finding suggests that increased astrocyte reactivity, as indicated by higher GFAP levels, is an early event in the progression of blood biomarker changes during the preclinical stage of AD. Taken as a whole, these results indicate that GFAP holds promise an early blood-based biomarker for reactive astrogliosis associated with Aβ pathology in preclinical AD. Consequently, this marker could be used to identify individuals at risk of AD before the onset of clinical symptoms [\[108](#page-21-35)].

Transforming growth factor-beta 1 (TGF-β1)

Recent studies indicate that a defciency in anti-infammatory cytokines, particularly transforming growth factor-β1 (TGF-β1), in the brains of patients with AD signifcantly contributes to microglia activation and neuroinfammation, thereby playing a crucial role in the pathophysiological mechanisms underlying cognitive decline in AD. [\[120](#page-22-9), [121](#page-22-10)].

Building on this evidence, researchers have explored the potential of TGF-β1 as a novel biomarker for AD [[120\]](#page-22-9). The deficit of TGF- β 1 can contribute to neurodegeneration through multiple mechanisms. Notably, TGF $β1$ plays a constitutive role in suppressing inflammation and regulates the degree of microglial activation in the CNS in an age-dependent manner [[122](#page-22-11)]. TGF-β1 also plays a pivotal role in synaptic plasticity and memory formation by facilitating the transition from early to late hippocampal long-term potentiation [\[123](#page-22-12)] and stimulating the uptake of Aβ by microglia [\[122](#page-22-11)]. Notably, numerous studies have shown that the TGF-β1 signaling pathway is selectively impaired in the early stages of AD, leading to microglia activation, neuroinfammation, increased neuronal vulnerability to Aβ oligomers, hippocampal atrophy, and cognitive decline [\[121,](#page-22-10) [124\]](#page-22-13). In addition, the AD brain exhibits a reduced expression of TGF-βR2, a specifc receptor which correlates with the pathological hallmarks of the disease [\[125](#page-22-14)]. When evaluating the potential of TGF-β1 as a novel biomarker for early AD, it is essential to consider the difering results obtained from its measurement in the plasma *versus* the CSF of AD patients. AD patients display decreased concentrations of active and inactive forms of TGF-β1 in their plasma [[126](#page-22-15)] as well as a decline in its secretion from PBMCs [\[127](#page-22-16)]. TGF-β1 levels were found to be elevated in the CSF and brain of AD patients compared to non-demented individuals, and positively correlated with the extent of cerebrovascular $\text{A}\beta$ deposition [\[128\]](#page-22-17).

Consequently, patients with AD exhibit elevated levels of TGF-β1 in their CSF [\[129](#page-22-18)], while decreased concentrations of both total and cleaved (active) forms of this mol-ecule have been observed in their plasma [\[130](#page-22-19)]. These seemingly contradictory fndings can be clarifed by longitudinally assessing TGF-β1 levels at various stages of AD in future prospective long-term observational studies. We hypothesize that elevated levels of TGF-β1 may act as a neuroprotective factor in the early phases of AD pathogenesis, while decreased levels contribute to neurodegeneration and cognitive decline in individuals with MCI [[121](#page-22-10)]. Interestingly, a study on patients with dementia found lower CSF concentrations of TGF-β1 in individuals with fast disease progression compared to those with slower progression [\[131\]](#page-22-7). Genetic investigations have provided only preliminary and partial evidence regarding the deficit of TGF-β1 in AD $[121]$.

The TGF- β 1 gene is located on chromosome 19q13.1–3 and contains multiple functional single nucleotide polymorphisms (SNPs) in the upstream and transcript regions [[132](#page-22-20)]. Two studies demonstrated that the SNPs at codons + 10 (T/C) and + 25 (G/C), as well as the CC genotype of the *TGF-β1* gene, which are associated with reduced TGF-β1 levels, have been linked to an increased conversion from MCI to AD [\[133,](#page-22-5) [134](#page-22-21)], whereas another research involving oldest-old individuals aged over 75 demonstrated that carriers of at least one minor T allele displayed a signifcant decline in cognitive and functional performance in the short-term, while those harboring the CC genotype of the TGF-β1 codon + 10 T > C

polymorphism remained stable [\[135](#page-22-6)]. Building on the evidence obtained from AD patients, various studies have been conducted in animal models of AD to validate the role of TGF-β1 as both a novel biomarker and a potential pharmacological target [[136\]](#page-22-22). It has been hypothesized that the selective deficit of the canonical TGF- $β1/S$ mad pathway in AD may impair the cross-talk between astrocytes and microglia, subsequently leading to microgliamediated neurodegeneration [\[136](#page-22-22)]. *APOE ε4* impairs the microglial response in AD by inducing TGFβ1-mediated checkpoints, suggesting a neurobiological link between *APOE ε4* and the deficit of TGFβ1 signaling in the disease process [\[137\]](#page-22-23). Despite conficting fndings regarding TGF-β1 levels at various stages of AD, this anti-infammatory cytokine has emerged as one of the leading 20 CSF candidate biomarkers associated with the rate of cognitive decline in dementia patients, as demonstrated in longitudinal studies [[131](#page-22-7)]. To validate the role of TGFβ1 as a novel biomarker in early AD, future prospective long-term observational studies are essential.

Other neuroinfammatory biomarkers

Numerous cytokines and chemokines, commonly linked to infammation, vascular injury, and angiogenesis, have emerged as potential neuroinfammatory biomarkers. Among these, eight can be measured in serum—including basic fbroblast growth factor (bFGF), C-reactive protein (CRP), interleukin-16 (IL-16), soluble fms-like tyrosine kinase-1 (sFLT-1), soluble intercellular adhesion molecule-1 (sICAM1), the Tie-2 receptor tyrosine kinase, vascular endothelial growth factor-C (VEGF-C), and vascular endothelial growth factor-D (VEGF-D). Three others, interleukin-15 (IL-15), monocyte chemoattractant protein-1 (MCP-1), and sFLT-1, are quantifable in the $CSF[138]$ $CSF[138]$ $CSF[138]$. The addition of these neuroinflammatory biomarkers to traditional AD biomarkers improved diagnostic accuracy by 13.9% and 12.5%, respectively, in older individuals with cognitive decline [\[139](#page-22-25)]. However, further studies are needed to confrm their utility in clinical settings.

A multicenter study has highlighted the signifcance of complement dysregulation as a potential predictor of disease progression in MCI [[140\]](#page-22-26). Specifcally, the results revealed higher levels of Factor B enzyme and lower levels of Factor H regulator in MCI progressors as compared to non-progressors. Collectively, these fndings suggest that the dysregulation of the complement system's amplifcation loop may act as an early event that predisposes to AD progression [[140](#page-22-26)].

Multiple meta-analyses have explored the potential of various molecules as biomarkers for AD. In one metaanalysis, which included 54 studies measuring cytokine concentrations (40 in peripheral blood and 14 in the CSF), patients with AD exhibited higher concentrations of IL-6, TNF-α, IL-1β, TGF-β, IL-12, and IL-18 in peripheral blood, and elevated levels of TGF-β in CSF, compared to healthy controls [\[141\]](#page-22-27). Another meta-analysis comprising 175 studies on peripheral blood revealed increased levels of IL-1β, IL-2, IL-6, IL-18, interferon-γ, homocysteine, high-sensitivity CRP (hs-CRP), C-X-C motif chemokine-10, epidermal growth factor, vascular cell adhesion molecule-1, TNF-α converting enzyme, soluble TNF receptors 1 and 2, α 1-antichymotrypsin, as well as decreased concentrations of IL-1 receptor antagonist and leptin in patients with AD compared to controls [[142\]](#page-22-28).

A comprehensive meta-analysis of 170 studies revealed signifcantly elevated blood concentrations of various proteins including hs-CRP, IL-6, soluble tumor necrosis factor receptors 1 and 2 (sTNFR1 and TNFR2), α1-antichymotrypsin (α1-ACT), IL-1β, and soluble CD40 ligand (sCD40L) in patients with AD compared to controls. Additionally, the CSF concentrations of certain proteins such as IL-10, MCP-1, TGF-β1, sTREM2, YKL-40, α1-ACT, nerve growth factor, and visinin-like protein-1 (VILIP-1) were also found to be higher in AD [\[143](#page-22-29)]. Furthermore, patients with MCI exhibited increased peripheral blood concentrations of sTNFR2, IL-6, MCP-1, and decreased concentrations of IL-8. The authors also observed elevated CSF concentrations of YKL-40, VILIP-1, and sTREM2 in MCI patients compared to controls. Finally, patients with AD were found to have increased peripheral blood concentrations of sTNFR1 and sTNFR2 compared to those with MCI [[143](#page-22-29)]. Another meta-analysis comprising 88 studies found increased levels of CRP, IL-1β, IL-2, IL-6, IL-12, IL-18, MCP-1, MCP 3, IL-8, and interferon-γ-inducible protein 10 (IP-10) in patients with AD $[144]$. These findings were at least in part consistent with a separate meta-analysis involving 13 studies that indicated an association between infammatory candidate proteins—including CRP, IL-6, α1-ACT, lipoproteinassociated phospholipase A2, and fbrinogen—and an increased risk of all-cause dementia, although these bio-markers were not specific to AD [[145](#page-22-31)].

The role of neuroinfammatory biomarkers in Alzheimer's disease diagnosis and therapy Role of neuroinfammatory biomarkers in the diagnostic work-up of Alzheimer's disease

The definitive diagnosis of AD—in the absence of neuropathological confrmation—continues to pose a signifcant challenge. Despite extensive research into the molecular and biological mechanisms underlying the disease, the efective identifcation of AD remains an elusive task. This poses limitations on therapeutic interventions, as they are often initiated after the onset of symptoms. In this scenario, there is a growing interest in studying and treating the prodromal stages of AD. One unresolved question in understanding AD pathophysiology is why a considerable percentage of brain Aβ-positive CU individuals do not develop detectable downstream tau pathology and subsequent clinical decline. Recent research has shown that elevated levels of phosphorylated tau in the blood are associated with Aβ accumulation in the brain only in individuals with abnormally high blood levels of GFAP $[114]$. These findings suggest that astrocyte reactivity, evaluated through the measurement of plasma GFAP, is a signifcant precursor event that connects brain Aβ accumulation to the onset of tau pathology. This connection may have important implications for the biological characterization of preclinical AD. Furthermore, considering that neurotoxic reactive astrocytes are stimulated by activated microglia [[117\]](#page-22-8), the availability of a biomarker for activated microglia becomes essential in identifying individuals who are at a higher risk of developing AD. While CSF sTREM2 and 18 kDa translocator protein (TPSO)-PET imaging are efective for identifying activated microglia, they do not inform about the precise molecular and functional cellular status. Furthermore, a reliable blood biomarker remains elusive. The availability of blood-based biomarkers for both activated microglia and reactive astrocytes could assist in the clinical recognition of MCI and AD, or even earlier stages[[138,](#page-22-24) [146](#page-22-32)]. Moreover, these biomarkers could facilitate tracking disease progression over time in patients as a part of therapeutic strategies and potentially provide personalized drug targets for early intervention in MCI and AD cases.

Role of neuroinfammatory biomarkers in anti-Alzheimer's disease trials and individualized therapy

Targeting neuroinfammation may prove to be an extremely efective strategy for AD prevention and therapy during the preclinical stage before signifcant neuronal loss occurs. Figure [2](#page-13-0) shows the number of ongoing AD clinical trials with anti-neuroinfammatory agents [[12\]](#page-19-11).

Overall, there are 25 ongoing clinical trials targeting neuroinfammation in AD: four in phase 1, 19 in phase 2, and two in phase 3 [masitinib (tyrosine-kinase inhibitor) and NE3107 (insulin-sensitizing agent)]. Promising results have been observed in several phase I/II clinical trials that targeted TNF-α, TREM2, or CD33. We strongly advocate for the utilization of neuroinfammatory biomarkers, such as blood GFAP for tracking reactive astrocytes and CSF-sTREM2 for monitoring microglial activation, throughout clinical trials [\[147\]](#page-22-33). In our view, these biomarkers present a signifcant potential for tracking disease progression within the AD continuum in the feld of drug development, including trials with compounds not directly impacting biological infammatory targets. Accordingly, blood GFAP concentrations have already been successfully employed as a biomarker in clinical trials evaluating anti-Aβ monoclonal antibodies, such as lecanemab $[148]$ $[148]$ and donanemab [[149\]](#page-22-35). In general, we are confident that adopting a biomarker-guided strategy for AD treatment, which tailors specifc interventions to relevant molecular pathways, will enhance therapeutic efectiveness, as witnessed in the field of oncology. This approach has already seen

Fig. 2 Presently ongoing clinical trials in AD by mechanism of action of the tested agents. Twenty-fve trials are testing anti-neuroinfammatory agents. *AD* Alzheimer disease, *APOE* Apolipoprotein E. Modifed from Cummings et al. Alzheimers Dement (N Y). 2024 [\[12](#page-19-11)]

application in the NSAID treatment of AD. A *post-hoc* analysis of a trial of naproxen and rofecoxib for mild-tomoderate AD demonstrated that those who responded favorably exhibited a distinct plasma neuroinfammatory profile (TNF- α , CRP, IL-6 and IL-10) [\[150\]](#page-23-5). These results imply that the use of anti-infammatory drugs for AD should be reserved for patients who show clear signs of systemic infammation. A Phase II trial, conducted more recently, with donanemab—a potent anti- $A\beta(p3-42)$ monoclonal antibody—exclusively enrolled patients who, as confrmed by [18F]fortaucipir-PET scans, exhibited pathologic tau deposition $(>1.10$ SUVR), but with quantitative tau levels below a specifc upper threshold (1.46 SUVR) [\[151](#page-23-6)]. This strategic approach was undertaken to address concerns surrounding the limited efficacy of donanemab in advanced disease situations, as suggested by the presence of extensive tau pathology. Another signifcant investigation is the AHEAD prevention study, currently in its fourth year. This research is analyzing the efects of lecanemab in CN participants at risk of developing AD due to the presence of brain Aβ accumulation, as evidenced by Aβ-PET scans $[152]$ $[152]$. This investigation is divided into two sub-studies (AHEAD 45 and AHEAD 3). AHEAD 45 focuses on participants exhibiting elevated brain Aβ-PET pathology, specifcally above 40 Centiloids, measured during the screening phase. In contrast, AHEAD 3 is being conducted on individuals with intermediate brain Aβ pathology levels, defned as 20 to 40 Centiloids, also measured during the screening process. The primary purpose of this comprehensive study is twofold. Firstly, it aims to determine if lecanemab treatment outperforms a placebo in modifying baseline Preclinical Alzheimer Cognitive Composite 5 (PACC5) after 216 weeks of treatment (A45 Trial). Secondly, it seeks to establish if lecanemab treatment is superior to a placebo in mitigating brain $Aβ$ accumulation, as measured by PET scans, following 216 weeks of treatment (A3 Trial) [[152\]](#page-23-7). We posit that the integration of a reliable blood biomarker of neuroinfammation is essential for the efective and predictive ATN(I) categorization of the AD continuum. During the pathological progression of AD, a pivotal moment occurs when innate immune and glial cells begin to sustain an excessively expressed chronic inflammatory response. This process acts in synergy with the accumulation of Aβ and tau proteins, driving synaptotoxicity and neurodegeneration in a self-perpetuating cycle. The precise timing of this neuroinflammatory shift in individual cases remains elusive, possibly explaining why past clinical trials exploring anti-infammatory compounds have failed to yield successful results. Plasma GFAP displays a compelling ability to predict individual clinical AD risk and is thus suggested as a potential preliminary screening tool for AD risk stratifcation in the

older adult population $[153]$ $[153]$ $[153]$. The presence of plasma GFAP "positivity" may be a straightforward indicator for initiating a comprehensive therapy. This treatment which would combine anti-infammatory drugs with agents that modulate either Aβ or tau—may be particularly applicable to those with preclinical AD or individuals at risk of developing AD.

Risk stratifcation tools tailored to each individual are crucial for applying precision medicine principles in AD. Blood biomarkers for AD offer a promising strategy that is both time and cost-effective. They hold potential to identify and categorize patients at risk of developing AD, thereby enhancing the screening procedures for potential participants in AD clinical trials. Additionally, these biomarkers can signifcantly improve patient management in clinical settings. This includes making informed decisions about treatment, such as choosing a diseasemodifying therapy based on altered biomarker profles, or referring patients to specialized memory clinics for focused care [\[19](#page-19-18)].

Tracking neuroinfammatory biomarkers could also be crucial for monitoring amyloid-related imaging abnormalities (ARIA), a signifcant adverse event associated with anti-Aβ monoclonal antibodies, including lecanemab and donanemab $[154]$ $[154]$. These ARIA manifest as brain edema (ARIA-E), microbleeds, and occasionally large brain hemorrhages (ARIA-H), and have been associated with some fatalities in clinical trials. ARIA are considered an infammatory reaction to cerebral amyloid angiopathy. Specifcally, ARIA-E resembles cerebral amyloid angiopathy-related infammation, a rare and serious condition caused by auto-antibodies to Aβ. Anti-Aβ antibodies may bind to vascular amyloid, triggering the complement cascade to attack cerebral blood vessels. This process can create small holes, leading to fluid leaks and microbleeds. It is recommended to identify preexisting medical disorders that may predispose individuals to ARIA or increase the likelihood of ARIA-related complications. Such conditions include pre-existing autoimmune or infammatory disorders, seizures, transient ischemic attacks, cerebrovascular disease, or signifcant changes in brain white matter.

AD is frequently associated with cerebrovascular disorders, which may contribute to neuronal dysfunction and death. Notably, both conditions share common risk factors, including *APOE ε4*, hyperlipidemia, and obesity [[155\]](#page-23-10). Several lines of evidence support the role of neuroinfammation and cerebrovascular dysfunction in AD. A study involving 508 CU older individuals and 313 patients with MCI and AD found that CSF levels of fve biomarkers of neuroinfammation and cerebrovascular dysfunction (YKL-40, ICAM-1, VCAM-1, IL-15, and Flt-1) were elevated in AD, even during the preclinical

and prodromal stages, and were associated with CSF tau. Additionally, longitudinal data suggested that higher levels of these neuroinfammatory and cerebrovascular biomarkers were linked to cognitive decline and an increased risk of subsequent development of AD [\[156\]](#page-23-11).

Regulatory perspectives on neuroinfammatory biomarkers in Alzheimer's disease

In the past decade, the identifcation of biomarkers relevant to AD has become a crucial tool in the development of disease-modifying therapies. Regulatory bodies such as the FDA [\(https://www.fda.gov/regulatory-infor](https://www.fda.gov/regulatory-information/search-fda-guidance-documents/qualification-process-drug-development-tools-guidance-industry-and-fda-staff) [mation/search-fda-guidance-documents/qualificat](https://www.fda.gov/regulatory-information/search-fda-guidance-documents/qualification-process-drug-development-tools-guidance-industry-and-fda-staff) [ion-process-drug-development-tools-guidance-indus](https://www.fda.gov/regulatory-information/search-fda-guidance-documents/qualification-process-drug-development-tools-guidance-industry-and-fda-staff) [try-and-fda-staf\)](https://www.fda.gov/regulatory-information/search-fda-guidance-documents/qualification-process-drug-development-tools-guidance-industry-and-fda-staff) and the European Medicines Agency (EMA) [\(https://www.ema.europa.eu/en/human-regul](https://www.ema.europa.eu/en/human-regulatory/research-development/scientific-advice-protocol-assistance/novel-methodologies-biomarkers/opinions-letters-support-qualification-novel-methodologies-medicine-development) [atory/research-development/scientific-advice-proto](https://www.ema.europa.eu/en/human-regulatory/research-development/scientific-advice-protocol-assistance/novel-methodologies-biomarkers/opinions-letters-support-qualification-novel-methodologies-medicine-development) [col-assistance/novel-methodologies-biomarkers/opini](https://www.ema.europa.eu/en/human-regulatory/research-development/scientific-advice-protocol-assistance/novel-methodologies-biomarkers/opinions-letters-support-qualification-novel-methodologies-medicine-development) [ons-letters-support-qualifcation-novel-methodologies](https://www.ema.europa.eu/en/human-regulatory/research-development/scientific-advice-protocol-assistance/novel-methodologies-biomarkers/opinions-letters-support-qualification-novel-methodologies-medicine-development)[medicine-development](https://www.ema.europa.eu/en/human-regulatory/research-development/scientific-advice-protocol-assistance/novel-methodologies-biomarkers/opinions-letters-support-qualification-novel-methodologies-medicine-development)) have established pathways for the qualifcation of these biomarkers to facilitate drug development. A qualifed biomarker may be defned as a"*tool that, within the stated context-of-use, can be relied upon to have a specifc interpretation and application in medical product development and regulatory review*" [[157\]](#page-23-12). However, despite the initial EMA opinion on the CSF biomarkers positive signature, which includes low $A\beta_{1-42}$ and high p-tau concentrations, as a predictor for dementia evolution in individuals with MCI [\(https://](https://www.ema.europa.eu/en/documents/regulatory-procedural-guideline/qualification-opinion-alzheimers-disease-novel-methodologies/biomarkers-use-cerebrospinal-fluid-amyloid-beta-1-42-t-tau-signature/positron-emission-tomography-amyloid-imaging-positive) [www.ema.europa.eu/en/documents/regulatory-proce](https://www.ema.europa.eu/en/documents/regulatory-procedural-guideline/qualification-opinion-alzheimers-disease-novel-methodologies/biomarkers-use-cerebrospinal-fluid-amyloid-beta-1-42-t-tau-signature/positron-emission-tomography-amyloid-imaging-positive) [dural-guideline/qualifcation-opinion-alzheimers-disea](https://www.ema.europa.eu/en/documents/regulatory-procedural-guideline/qualification-opinion-alzheimers-disease-novel-methodologies/biomarkers-use-cerebrospinal-fluid-amyloid-beta-1-42-t-tau-signature/positron-emission-tomography-amyloid-imaging-positive) [se-novel-methodologies/biomarkers-use-cerebrospinal](https://www.ema.europa.eu/en/documents/regulatory-procedural-guideline/qualification-opinion-alzheimers-disease-novel-methodologies/biomarkers-use-cerebrospinal-fluid-amyloid-beta-1-42-t-tau-signature/positron-emission-tomography-amyloid-imaging-positive)[fuid-amyloid-beta-1-42-t-tau-signature/positron-emiss](https://www.ema.europa.eu/en/documents/regulatory-procedural-guideline/qualification-opinion-alzheimers-disease-novel-methodologies/biomarkers-use-cerebrospinal-fluid-amyloid-beta-1-42-t-tau-signature/positron-emission-tomography-amyloid-imaging-positive) [ion-tomography-amyloid-imaging-positive\)](https://www.ema.europa.eu/en/documents/regulatory-procedural-guideline/qualification-opinion-alzheimers-disease-novel-methodologies/biomarkers-use-cerebrospinal-fluid-amyloid-beta-1-42-t-tau-signature/positron-emission-tomography-amyloid-imaging-positive), only a small number of biomarkers have undergone a formal regulatory process for qualifcation. Signifcantly, the absence of qualifed biomarkers for diagnosing AD, predicting disease prognosis, and evaluating treatment efficacy remains a notable issue. This can be attributed to our limited understanding of the neurobiology of AD and its connection to cognitive and behavioral decline over time. The disease progresses along a continuum of states, which are not fully characterized presently, and exhibits considerable variability among patients. Consequently, the identifcation and validation of prognostic and predictive biomarkers are urgently required, but their achievement poses substantial challenges.

The FDA recent approval of aducanumab for the treatment of AD through the accelerated approval pathway was met with criticism due to the lack of demonstrated clinical beneft. While the drug induced a reduction in the Aβ biomarker, the EMA did not replicate the FDA approval. The recent FDA full approval of lecanemab, which reduces brain Aβ burden but shows limited cognitive and clinical beneft, further highlights the challenge of using individual biomarkers as efficacy endpoints in AD. The results of recent clinical trials in AD indicate the importance of identifying a broader range of positive biomarker signatures. These should include inflammatory biomarkers, as well as other markers of brain damage or susceptibility to damage. Such biomarkers can be used to monitor and anticipate disease progression across various stages, and to measure the efectiveness of new disease-modifying drugs.

Discussion

In the AD brain, neuroinflammation is a multifaceted biological process that entails the recruitment of peripheral immune cells, the activation of intracellular signaling pathways, and the release of various proinfammatory cytokines. The key contributors to the neuroinflammatory process are microglia and astrocytes. Their involvement exhibits distinct phenotypic variations, both spatially and temporally, which can be observed at diferent stages of disease progression [[99,](#page-21-26) [158](#page-23-13)]. Recent GWAS have provided compelling evidence supporting the signifcant involvement of the innate immune system and neuroinfammation in the pathogenesis of AD. A comprehensive GWAS conducted on over 1 million participants has specifcally highlighted the relevance of microglia and immune cells in the pathogenesis of LOAD [[159\]](#page-23-14). Furthermore, the identifcation of several AD risk genes associated with immune response and microglia, such as CD33 and TREM2, through GWAS has shed light on their role in the disease [[160\]](#page-23-15). In addition to these genes, sTREM2 and YKL-40, as well as other emerging cytokines, such as IL-6, MCP-1, and TGF-β1 are showing promising potential as infammatory candidate biomarkers. However, to fully comprehend the clinical implications of these neuroinfammatory biomarkers, it is paramount to conduct large-scale longitudinal studies across the entire AD continuum [\[13](#page-19-12)].

sTREM2 has emerged as a promising biomarker of activated microglia and has been validated in longitudinal studies in both pre-clinical and early AD. However, its widespread use is impeded by the fact that it can be easily measured in CSF, but not in blood. Despite this limitation, TREM2 is still considered a promising therapeutic target for AD. One such investigational therapy is AL002, a humanized monoclonal IgG1 antibody that enhances TREM2 signaling to improve microglia survival and pro-liferation [[161](#page-23-16)]. AL002 is currently undergoing investigation in a 96-week, double-blind, placebo-controlled study that involves 265 patients with early AD (INVOKE-2 study, *ClinicalTrials.gov* NCT04592874). The primary objective of this phase 2 study is to evaluate the impact of AL002 on disease progression, measured through the Clinical Dementia Rating Sum Boxes (CDR-SB).

Numerous cross-sectional and longitudinal studies have consistently shown the potential of YKL-40 as a reliable biomarker of neuroinfammation in AD. YKL-40 serves as an indicator of both activated microglia and reactive astrocytes and can be detected in both CSF and blood samples. Notably, several studies in the feld of AD have highlighted the diagnostic signifcance of plasma YKL-40 levels in the early stages of dementia, such as MCI and mild clinical AD [[92,](#page-21-2) [96](#page-21-24), [97\]](#page-21-5). In addition, elevated plasma concentrations of YKL-40 have been found to be positively associated with cognitive performance in individuals with subjective cognitive complaints [\[97](#page-21-5)]. However, it is important to note that increased YKL-40 concentrations in CSF or plasma do not exclusively indicate an infammatory biomarker specifc to AD or other neurodegenerative diseases. Accordingly, elevated YKL-40 concentrations have also been observed in other conditions such as stroke, atrial fbrillation, hypertension, and diabetes, as well as in association with vascular risk factors $[86]$ $[86]$. The non-specificity of YKL-40 expression in various age-related pathological conditions, including neoplastic and cardiovascular diseases, as well as infammatory disorders of diferent etiologies, poses a constraint on its future application as a biomarker in the older adult population [[84,](#page-21-15) [162](#page-23-17)]. Moreover, while CSF YKL-40 concentrations have shown a moderately positive correlation with p-tau and t-tau, there was no correlation with Aβ, further substantiating its lack of specificity for AD $[163]$ $[163]$. Therefore, when utilizing YKL-40 in diagnostic examinations, it is crucial to gather a comprehensive medical history of patient comorbidities to avoid misinterpretation of biomarker values [\[164\]](#page-23-19).

IL-6 shows promise as a potential peripheral infammatory biomarker for evaluating the severity of cognitive decline. However, there is currently a lack of a standardized molecular panel of fuid infammatory biomarkers that can be efectively used for screening purposes [\[33](#page-19-32)].

It is important to note that the literature on infammatory biomarkers and their ability to track the progression of AD contains some seemingly contradictory fndings. A recent 10-year longitudinal study involving CU older individuals identifed a positive correlation between CSF sTREM2 levels and the risk of CDR conversion [\[165](#page-23-20)]. Conversely, other studies have found that elevated CSF sTREM2 levels are associated with a slower cognitive and clinical decline in AD $[166]$ and in Aβ-PET-positive MCI [167]. Therefore, TREM2-related immune activation may infuence the progression of age-related cognitive decline and AD symptoms diferently, depending on the disease status and amyloid pathology. It has been hypothesized that throughout the AD continuum, neuroinfammatory biomarkers in blood and CSF exhibit a complex temporal progression, with distinct profles for CSF and blood sTREM2, GFAP, and YKL-40 [[168\]](#page-23-23) (Fig. [3](#page-17-0)).

This complexity accounts for the apparent discrepancies in the results of some neuroinfammatory biomarker studies and may also partially explain the failure of antiinfammatory therapies across the AD continuum to date. It is reasonable to assume that anti-infammatory drugs were tested without considering the infammatory status of the trial participants. This situation is analogous to the initial studies with anti-Aβ drugs, which were tested in AD patients without assessing their brain Aβ deposition and tau load status. Ideally, future anti-infammatory candidates should be tested in homogeneous subject populations, characterized by reactive astrocytosis (estimated via plasma GFAP levels) and microglial activation (estimated via CSF sTREM2 levels).

A crucial consideration when utilizing AD neuroinfammatory biomarkers is their lack of specifcity. For instance, it is well-established that blood GFAP elevations are not exclusive to AD, as they are also observed in other acute CNS conditions such as ischemic stroke or traumatic brain injury [[169](#page-23-24)]. Similarly, while low plasma sTREM2 has been associated with Aβ accumulation and CSF p-tau levels, a comparable decrease has been reported in vascular dementia [[170](#page-23-25)]. To enhance specifcity, a combination of infammatory biomarkers might be a viable option [\[139\]](#page-22-25).

However, we posit that the lack of disease specifcity in blood biomarkers should not be viewed as a limitation. Instead, it could serve as a valuable initial screening tool. A positive result for a specifc blood biomarker test could be interpreted as a non-specifc signal of neuroinfammation, neurodegeneration, or Aβ brain deposition, underscoring the need for additional confrmatory testing and further clinical examinations [[171](#page-23-26)].

Currently, the $AT(N)$ classification system is extensively utilized as a biological staging model for AD. It assesses three specifc classes of biomarkers, i.e., Aβ, tau pathology, and neurodegeneration/neuronal injury. Recent advancements have identifed promising blood-based biomarkers for each category—including $A\beta_{1-42}/A\beta_{1-40}$ ratio, phosphorylated tau, and NfL. The Alzheimer's Association has published recommendations for bloodbased biomarkers, emphasizing the need for longitudinal and observational clinical trials to establish changes in peripheral biomarkers over time in patients with AD. These trials should also monitor changes in clinically relevant outcomes, such as cognition and motor functions [[172,](#page-23-27) [173\]](#page-23-28). Recently, there has been a proposal to enhance the $AT(N)$ matrix by introducing the $ATI(N)$ system, with the addition of a neuroinfammatory biomarker denoted as "I". The Alzheimer's Association workgroup

Alzheimer's disease continuum

Fig. 3 Hypothetical time course profles of main fuid neuroinfammatory biomarkers (GFAP, sTREM2, and YKL-40) across the AD continuum. The biomarker levels presented in the graph are referred to their normal range and should not be compared with each other. *AD* Alzheimer disease, *CSF* cerebrospinal fuid, *GFAP* glial fbrillary acidic protein, *MCI* mild cognitive impairment, *sTREM2* soluble triggering receptor expressed on myeloid cells 2, *YKL-40* chitinase-3-like protein 1

is proposing the inclusion of glial GFAP as a biomarker of infammation and astrocyte activation in their revised diagnostic criteria for AD. This proposal is based on the growing evidence supporting the role of astrocyte reactivity in the pathogenesis of AD. In the near future, it is anticipated that specifc "threshold" serum GFAP levels will be established to defne "astrogliosis positivity" along the AD continuum [\[171\]](#page-23-26), providing a standardized approach for assessing astrocyte activation in AD patients. To support this endeavor, a study involving 371 healthy Danish volunteers aged between 21 and 90 years has already determined the normal range of serum GFAP levels across different age groups $[174]$. This expanded plasma ATI(N) system, in combination with *APOE* genotyping and cognitive testing, presents an opportunity for personalized assessment, enabling a therapy approach tailored to the specifc biomarker profles of patients with AD.

Conclusion and future directions

Recent longitudinal studies have successfully recruited large cohorts of individuals with accurate clinical characterizations, leading to the identifcation of potentially reliable blood-based candidate biomarkers of neuroinflammation in AD. These markers offer a practical and non-invasive means of screening and monitoring the infammatory status of the brain in the AD continuum and hold potential to identify and categorize patients at risk of developing AD, thereby enhancing the screening procedures for potential participants in AD clinical trials and choosing a disease-modifying therapy based on altered biomarker profles [[19](#page-19-18)].

Longitudinal studies suggest that CSF sTREM-2 exhibits a dynamic response linked to microglial activity as the disease progresses and increased CSF sTREM-2 concentrations are associated with high levels of NfL, indicating axonal injury [\[76](#page-20-1)]. CSF sTREM-2 might be validated as a new biomarker for tracking the progression from preclinical AD/MCI to AD dementia that correlates with the progression of CSF Aβ and tau. However, the widespread use of sTREM-2 is impeded by the fact that it can be easily measured in CSF, but not in blood. Despite this limitation, TREM-2 is still considered a promising therapeutic target for AD. One such investigational therapy is AL002, a humanized monoclonal IgG1 antibody that enhances TREM-2 signaling to improve microglia survival and pro-liferation [[161](#page-23-16)]. AL002 is currently undergoing investigation in a 96-week, double-blind, placebo-controlled study

that involves 265 patients with early AD (INVOKE-2 study, *ClinicalTrials.gov* NCT04592874). The primary objective of this phase 2 study is to evaluate the impact of AL002 on disease progression, measured through the Clinical Dementia Rating Sum Boxes (CDR-SB).

According to cross-sectional and longitudinal studies YKL-40 possesses a good potential as a reliable biomarker of neuroinfammation in AD, because its levels are increased in preclinical AD [\[92](#page-21-2)] and linked to biomarkers of neurodegeneration (total tau, t-tau), taumediated toxicity (p-tau) [\[89](#page-21-20)]. YKL-40 might be helpful in distinguishing individuals with MCI who will convert to AD from those who will remain stable at 5 years [\[86](#page-21-17)]. Additionally, serum concentrations of YKL-40 can efectively distinguish between CU individuals and those with mild dementia, with a sensitivity and specifcity of 85% [[164\]](#page-23-19), although it is not clinically useful in differentiating the characteristic AD phenotype and increased YKL-40 concentrations in CSF or plasma have been found in various age-related pathological conditions, posing a constraint on its future application as a biomarker in the older population [\[84](#page-21-15)].

When considering all neuroinfammatory biomarkers and all recent longitudinal studies, GFAP emerges as the most promising biomarker, efectively tracking reactive astrocytes and enabling the identifcation of Aβ-positive CU individuals who exhibit early signs of p-tau pathology, with its plasma concentrations being indicative of early-stage dementia and in MCI who progress into AD [[175\]](#page-23-30). By assessing a combination of plasma biomarkers, such as the ratio between the $A\beta_{1-42}$ and $A\beta_{1-40}$, p-tau₂₁₇, NfL, and GFAP concentrations, it may be possible to create a novel efective panel for assessing the risk of developing AD [[19,](#page-19-18) [116](#page-22-1)].

According to the evidence discussed in the present review, we believe that enhancing the AT(N) matrix by introducing the ATI(N) system, with the addition of neuroinfammatory biomarkers denoted as "I" (CSF and blood "I" biomarkers), such as GFAP, might signifcantly improve, in combination with the other biomarkers and cognitive testing, both the early diagnosis of AD and the development of disease-modifying drugs in future AD clinical trials.

Abbreviations

Acknowledgements

Not applicable.

Author contributions

S.L., B.P.I., M.T.H., and F.C.: project administration and conceptualization of the manuscript. S.L., B.P.I., M.G., A.F., C.I., and F.C.: literature search. S.L., B.P.I., M.G., A.F., A.S.-L., and M.T.H.: writing—original draft. S.L., B.P.I., and C.I.: preparation of the fgures. E.E., P.M., S.L.-O., J.M.-H., A.G., G.C., M.M., and D.M.: writing—review & editing. S.L., B.P.I., M.T.H., and F.C.: supervision of the manuscript. All Authors reviewed and approved the fnal version of the manuscript.

Funding

Research by S.L.-O. is funded by the Spanish Ministry of Science and Innovation (Grant number FPU19/02117).

Availability of data and materials

Not applicable.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

B.P.I. is an employee at Chiesi Farmaceutici. He is listed among the inventors of a number of Chiesi Farmaceutici's patents of anti-Alzheimer drugs. E.E. is the unique owner of 2E Science, a for-proft private scientifc company. Neither E.E. nor 2E Science have any commercial interest or fnancial tie in relation with this article. S.L., M.G., A.F., P.M., S.L.-O., J.M.-A., A.G., G.C., M.M., D.M., A.S.-L., C.I., M.T.H., and F.C. declare no competing interests.

Author details

¹i+HeALTH Strategic Research Group, Department of Health Sciences, Miguel de Cervantes European University (UEMC), 47012 Valladolid,

Spain. ² Department of Research and Development, Chiesi Farmaceutici, 43122 Parma, Italy. ³Oasi Research Institute-IRCCS, 94018 Troina, Italy. ⁴2E Science, 27038 Robbio, Italy. ⁵Studio Minoretti, 23848 Oggiono, LC, Italy. ⁶CMRR, Memory Resources and Research Center, Montpellier University of Excellence i-site, 34295 Montpellier, France. ⁷ Department of Drug and Health Sciences, University of Catania, 95125 Catania, Italy. ⁸Department for Life Quality Studies, Alma Mater Studiorum, University of Bologna, 40126 Bologna, Italy. ⁹Department of Physiology and Pharmacology, Sapienza University, 00185 Rome, Italy.
¹⁰Physical Activity and Health Research Group (PaHerg), Research Institute of the Hospital, 12 de Octubre ('imas12'), 28041 Madrid, Spain. ¹¹ Department of Brain and Behavioral Sciences, University of Pavia, 27100 Pavia, Italy.
¹²Luxembourg Centre for Systems Biomedicine, University of Luxembourg, 4367 Esch‑Belval, Luxembourg.

Received: 29 May 2024 Accepted: 28 June 2024 Published online: 30 July 2024

References

- 1. Kumar A, Sidhu J, Goyal A, Tsao JW. Alzheimer Disease. In StatPearls. 2023.
- 2. Chávez-Fumagalli MA, Shrivastava P, Aguilar-Pineda JA, Nieto-Montesinos R, Del-Carpio GD, Peralta-Mestas A, Caracela-Zeballos C, Valdez-Lazo G, Fernandez-Macedo V, Pino-Figueroa A, et al. Diagnosis of Alzheimer's disease in developed and developing countries: systematic review and meta-analysis of diagnostic test accuracy. J Alzheimers Dis Rep. 2021;5:15–30.
- 3. Dhillon S. Aducanumab: frst approval. Drugs. 2021;81:1437–43.
- 4. Reardon S. FDA approves Alzheimer's drug lecanemab amid safety concerns. Nature. 2023;613:227–8.
- 5. Sims JR, Zimmer JA, Evans CD, Lu M, Ardayfo P, Sparks J, Wessels AM, Shcherbinin S, Wang H, Monkul Nery ES, et al. Donanemab in early symptomatic alzheimer disease: the TRAILBLAZER-ALZ 2 randomized clinical trial. JAMA. 2023;330:512–27.
- 6. Melchiorri D, Merlo S, Micallef B, Borg JJ, Dráf F. Alzheimer's disease and neuroinfammation: will new drugs in clinical trials pave the way to a multi-target therapy? Front Pharmacol. 2023;14:1196413.
- 7. Imbimbo BP, Ippati S, Watling M, Balducci C. A critical appraisal of tautargeting therapies for primary and secondary tauopathies. Alzheimers Dement. 2022;18:1008–37.
- 8. Lozupone M, Imbimbo BP, Balducci C, Lo Vecchio F, Bisceglia P, Latino RR, Leone M, Dibello V, Solfrizzi V, Greco A, et al. Does the imbalance in the apolipoprotein E isoforms underlie the pathophysiological process of sporadic Alzheimer's disease? Alzheimers Dement. 2023;19:353–68.
- 9. Venkataraman AV, Mansur A, Rizzo G, Bishop C, Lewis Y, Kocagoncu E, Lingford-Hughes A, Huiban M, Passchier J, Rowe JB, et al. Widespread cell stress and mitochondrial dysfunction occur in patients with early Alzheimer's disease. Sci Transl Med. 2022;14:eabk1051.
- 10. Tzioras M, McGeachan RI, Durrant CS, Spires-Jones TL. Synaptic degeneration in Alzheimer disease. Nat Rev Neurol. 2023;19:19–38.
- 11. Streit WJ, Mrak RE, Griffin WS. Microglia and neuroinflammation: a pathological perspective. J Neuroinfammation. 2004;1:14.
- 12. Cummings J, Zhou Y, Lee G, Zhong K, Fonseca J, Cheng F. Alzheimer's disease drug development pipeline: 2024. Alzheimers Dement (N Y). 2024;10: e12465.
- 13. Morgan DG, Mielke MM. Knowledge gaps in Alzheimer's disease immune biomarker research. Alzheimers Dement. 2021;17:2030–42.
- 14. Blennow K, Galasko D, Perneczky R, Quevenco FC, van der Flier WM, Akinwonmi A, Carboni M, Jethwa A, Suridjan I, Zetterberg H. The potential clinical value of plasma biomarkers in Alzheimer's disease. Alzheimers Dement. 2023.
- 15. Barthélemy NR, Salvadó G, Schindler SE, He Y, Janelidze S, Collij LE, Saef B, Henson RL, Chen CD, Gordon BA, et al. Highly accurate blood test for Alzheimer's disease is similar or superior to clinical cerebrospinal fuid tests. Nat Med. 2024;30:1085–95.
- 16. Ashton NJ, Brum WS, Di Molfetta G, Benedet AL, Arslan B, Jonaitis E, Langhough RE, Cody K, Wilson R, Carlsson CM, et al. Diagnostic accuracy of a plasma phosphorylated tau 217 immunoassay for Alzheimer disease pathology. JAMA Neurol. 2024;81:255–63.
- 17. Malek-Ahmadi M, Su Y, Ghisays V, Luo J, Devadas V, Chen Y, Lee W, Protas H, Chen K, Zetterberg H, et al. Plasma NfL is associated with the APOE ε4 allele, brain imaging measurements of neurodegeneration, and lower recall memory scores in cognitively unimpaired late-middle-aged and older adults. Alzheimers Res Ther. 2023;15:74.
- 18. Rosenberg A, Öhlund-Wistbacka U, Hall A, Bonnard A, Hagman G, Rydén M, Thunborg C, Wiggenraad F, Sandebring-Matton A, Solomon A, Kivipelto M. β-Amyloid, tau, neurodegeneration classifcation and eligibility for anti-amyloid treatment in a memory clinic population. Neurology. 2022;99:e2102–13.
- 19. Imbimbo BP, Watling M, Imbimbo C, Nisticò R. Plasma ATN(I) classification and precision pharmacology in Alzheimer's disease. Alzheimers Dement. 2023;19:4729–34.
- 20. Gaetani L, Paolini Paoletti F, Bellomo G, Mancini A, Simoni S, Di Filippo M, Parnetti L. CSF and blood biomarkers in neuroinfammatory and neurodegenerative diseases: implications for treatment. Trends Pharmacol Sci. 2020;41:1023–37.
- 21. Liu Y, Shen X, Zhang Y, Zheng X, Cepeda C, Wang Y, Duan S, Tong X. Interactions of glial cells with neuronal synapses, from astrocytes to microglia and oligodendrocyte lineage cells. Glia. 2023;71:1383–401.
- 22. Hufels CFM, Middeldorp J, Hol EM. Aß pathology and neuron-glia interactions: a synaptocentric view. Neurochem Res. 2023;48:1026–46.
- 23. Villegas-Llerena C, Phillips A, Garcia-Reitboeck P, Hardy J, Pocock JM. Microglial genes regulating neuroinfammation in the progression of Alzheimer's disease. Curr Opin Neurobiol. 2016;36:74–81.
- 24. Jensen CJ, Massie A, De Keyser J. Immune players in the CNS: the astrocyte. J Neuroimmune Pharmacol. 2013;8:824–39.
- 25. Jiwaji Z, Tiwari SS, Avilés-Reyes RX, Hooley M, Hampton D, Torvell M, Johnson DA, McQueen J, Baxter P, Sabari-Sankar K, et al. Reactive astrocytes acquire neuroprotective as well as deleterious signatures in response to Tau and Aß pathology. Nat Commun. 2022;13:135.
- 26. Wang S, Colonna M. Microglia in Alzheimer's disease: a target for immunotherapy. J Leukoc Biol. 2019;106:219–27.
- 27. Heneka MT, Carson MJ, El Khoury J, Landreth GE, Brosseron F, Feinstein DL, Jacobs AH, Wyss-Coray T, Vitorica J, Ransohoff RM, et al. Neuroinfammation in Alzheimer's disease. Lancet Neurol. 2015;14:388–405.
- 28. Forloni G, Balducci C. Alzheimer's disease, oligomers, and infammation. J Alzheimers Dis. 2018;62:1261–76.
- 29. Condello C, Yuan P, Schain A, Grutzendler J. Microglia constitute a barrier that prevents neurotoxic protofbrillar Aβ42 hotspots around plaques. Nat Commun. 2015;6:6176.
- 30. Cai Y, Liu J, Wang B, Sun M, Yang H. Microglia in the neuroinfammatory pathogenesis of Alzheimer's disease and related therapeutic targets. Front Immunol. 2022;13: 856376.
- 31. Merlo S, Spampinato SF, Caruso GI, Sortino MA. The ambiguous role of microglia in Aβ toxicity: chances for therapeutic intervention. Curr Neuropharmacol. 2020;18:446–55.
- Sims R, Hill M, Williams J. The multiplex model of the genetics of Alzheimer's disease. Nat Neurosci. 2020;23:311–22.
- Hampel H, Caraci F, Cuello AC, Caruso G, Nisticò R, Corbo M, Baldacci F, Toschi N, Garaci F, Chiesa PA, et al. A path toward precision medicine for neuroinfammatory mechanisms in Alzheimer's disease. Front Immunol. 2020;11:456.
- 34. Griciuc A, Serrano-Pozo A, Parrado AR, Lesinski AN, Asselin CN, Mullin K, Hooli B, Choi SH, Hyman BT, Tanzi RE. Alzheimer's disease risk gene CD33 inhibits microglial uptake of amyloid beta. Neuron. 2013;78:631–43.
- 35. Rathore N, Ramani SR, Pantua H, Payandeh J, Bhangale T, Wuster A, Kapoor M, Sun Y, Kapadia SB, Gonzalez L, et al. Paired Immunoglobulinlike Type 2 Receptor Alpha G78R variant alters ligand binding and confers protection to Alzheimer's disease. PLoS Genet. 2018;14: e1007427.
- 36. Lu L, Yao QY, Ruan SS, Hu JW, Long WJ, Dai WZ, Ma T, Zhu XC. Explore the role of CR1 genetic variants in late-onset Alzheimer's disease susceptibility. Psychiatr Genet. 2021;31:216–29.
- 37. Deming Y, Filipello F, Cignarella F, Cantoni C, Hsu S, Mikesell R, Li Z, Del-Aguila JL, Dube U, Farias FG, et al. The MS4A gene cluster is a key modulator of soluble TREM2 and Alzheimer's disease risk. Sci Transl Med. 2019; 11.
- 38. Liu X, Che R, Liang W, Zhang Y, Wu L, Han C, Lu H, Song W, Wu Y, Wang Z. Clusterin transduces Alzheimer-risk signals to amyloidogenesis. Signal Transduct Target Ther. 2022;7:325.
- 39. De Roeck A, Van Broeckhoven C, Sleegers K. The role of ABCA7 in Alzheimer's disease: evidence from genomics, transcriptomics and methylomics. Acta Neuropathol. 2019;138:201–20.
- 40. Owens HA, Thorburn LE, Walsby E, Moon OR, Rizkallah P, Sherwani S, Tinsley CL, Rogers L, Cerutti C, Ridley AJ, et al. Alzheimer's diseaseassociated P460L variant of EphA1 dysregulates receptor activity and blood-brain barrier function. Alzheimers Dement. 2024;20:2016–33.
- 41. James LM, Christova P, Lewis SM, Engdahl BE, Georgopoulos A, Georgopoulos AP. Protective effect of human leukocyte antigen (HLA) Allele DRB1*13:02 on age-related brain gray matter volume reduction in healthy women. EBioMedicine. 2018;29:31–7.
- 42. Klesney-Tait J, Turnbull IR, Colonna M. The TREM receptor family and signal integration. Nat Immunol. 2006;7:1266–73.
- 43. Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Szternfeld R, Ulland TK, David E, Baruch K, Lara-Astaiso D, Toth B, et al. A unique microglia type associated with restricting development of Alzheimer's disease. Cell. 2017;169:1276-1290.e1217.
- 44. Zhao Y, Wu X, Li X, Jiang LL, Gui X, Liu Y, Sun Y, Zhu B, Piña-Crespo JC, Zhang M, et al. TREM2 is a receptor for β-amyloid that mediates microglial function. Neuron. 2018;97:1023-1031.e1027.
- 45. Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E, Cruchaga C, Sassi C, Kauwe JS, Younkin S, et al. TREM2 variants in Alzhei‑ mer's disease. N Engl J Med. 2013;368:117–27.
- 46. Jonsson T, Stefansson H, Steinberg S, Jonsdottir I, Jonsson PV, Snaedal J, Bjornsson S, Huttenlocher J, Levey AI, Lah JJ, et al. Variant of TREM2 associated with the risk of Alzheimer's disease. N Engl J Med. 2013;368:107–16.
- 47. Li JT, Zhang Y. TREM2 regulates innate immunity in Alzheimer's disease. J Neuroinfammation. 2018;15:107.
- 48. Zheng H, Cheng B, Li Y, Li X, Chen X, Zhang YW. TREM2 in Alzheimer's disease: microglial survival and energy metabolism. Front Aging Neurosci. 2018;10:395.
- 49. Hickman SE, El Khoury J. TREM2 and the neuroimmunology of Alzheimer's disease. Biochem Pharmacol. 2014;88:495–8.
- 50. Carmona S, Zahs K, Wu E, Dakin K, Bras J, Guerreiro R. The role of TREM2 in Alzheimer's disease and other neurodegenerative disorders. Lancet Neurol. 2018;17:721–30.
- 51. Ruiz A, Dols-Icardo O, Bullido MJ, Pastor P, Rodríguez-Rodríguez E, López de Munain A, de Pancorbo MM, Pérez-Tur J, Alvarez V, Antonell A, et al. Assessing the role of the TREM2 pR47H variant as a risk factor for Alzheimer's disease and frontotemporal dementia. Neurobiol Aging. 2014;35:444.
- 52. Cuyvers E, Bettens K, Philtjens S, Van Langenhove T, Gijselinck I, van der Zee J, Engelborghs S, Vandenbulcke M, Van Dongen J, Geerts N, et al. Investigating the role of rare heterozygous TREM2 variants in Alzheimer's disease and frontotemporal dementia. Neurobiol Aging. 2014;35(726):e711-729.
- 53. Condello C, Yuan P, Grutzendler J. Microglia-mediated neuroprotection, TREM2, and Alzheimer's disease: evidence from optical imaging. Biol Psychiatry. 2018;83:377–87.
- 54. Cheng-Hathaway PJ, Reed-Geaghan EG, Jay TR, Casali BT, Bemiller SM, Puntambekar SS, von Saucken VE, Williams RY, Karlo JC, Moutinho M, et al. The Trem2 R47H variant confers loss-of-function-like phenotypes in Alzheimer's disease. Mol Neurodegener. 2018;13:29.
- 55. Li R, Wang X, He P. The most prevalent rare coding variants of TREM2 conferring risk of Alzheimer's disease: a systematic review and metaanalysis. Exp Ther Med. 2021;21:347.
- 56. Slattery CF, Beck JA, Harper L, Adamson G, Abdi Z, Uphill J, Campbell T, Druyeh R, Mahoney CJ, Rohrer JD, et al. R47H TREM2 variant increases risk of typical early-onset Alzheimer's disease but not of prion or frontotemporal dementia. Alzheimers Dement. 2014;10:602-608.e604.
- 57. Jin SC, Carrasquillo MM, Benitez BA, Skorupa T, Carrell D, Patel D, Lincoln S, Krishnan S, Kachadoorian M, Reitz C, et al. TREM2 is associated with increased risk for Alzheimer's disease in African Americans. Mol Neurodegener. 2015;10:19.
- 58. Miyashita A, Wen Y, Kitamura N, Matsubara E, Kawarabayashi T, Shoji M, Tomita N, Furukawa K, Arai H, Asada T, et al. Lack of genetic associa· tion between TREM2 and late-onset Alzheimer's disease in a Japanese population. J Alzheimers Dis. 2014;41:1031–8.
- Huang M, Wang D, Xu Z, Xu Y, Xu X, Ma Y, Xia Z. Lack of genetic association between TREM2 and Alzheimer's disease in East Asian population:

a systematic review and meta-analysis. Am J Alzheimers Dis Other Demen. 2015;30:541–6.

- 60. Sims R, van der Lee SJ, Naj AC, Bellenguez C, Badarinarayan N, Jakobs‑ dottir J, Kunkle BW, Boland A, Raybould R, Bis JC, et al. Rare coding variants in PLCG2, ABI3, and TREM2 implicate microglial-mediated innate immunity in Alzheimer's disease. Nat Genet. 2017;49:1373–84.
- 61. Jin SC, Benitez BA, Karch CM, Cooper B, Skorupa T, Carrell D, Norton JB, Hsu S, Harari O, Cai Y, et al. Coding variants in TREM2 increase risk for Alzheimer's disease. Hum Mol Genet. 2014;23:5838–46.
- 62. Ulland TK, Song WM, Huang SC, Ulrich JD, Sergushichev A, Beatty WL, Loboda AA, Zhou Y, Cairns NJ, Kambal A, et al. TREM2 maintains microglial metabolic ftness in Alzheimer's disease. Cell. 2017;170:649-663. e613.
- 63. Schlepckow K, Kleinberger G, Fukumori A, Feederle R, Lichtenthaler SF, Steiner H, Haass C. An Alzheimer-associated TREM2 variant occurs at the ADAM cleavage site and afects shedding and phagocytic function. EMBO Mol Med. 2017;9:1356–65.
- 64. Jiang T, Tan L, Chen Q, Tan MS, Zhou JS, Zhu XC, Lu H, Wang HF, Zhang YD, Yu JT. A rare coding variant in TREM2 increases risk for Alzheimer's disease in Han Chinese. Neurobiol Aging. 2016;42(217):e211-213.
- 65. Jiang T, Hou JK, Gao Q, Yu JT, Zhou JS, Zhao HD, Zhang YD. TREM2 p.H157Y variant and the risk of Alzheimer's disease: a meta-analysis involving 14,510 subjects. Curr Neurovasc Res. 2016;13:318–20.
- 66. Heslegrave A, Heywood W, Paterson R, Magdalinou N, Svensson J, Johansson P, Öhrfelt A, Blennow K, Hardy J, Schott J, et al. Increased cerebrospinal fuid soluble TREM2 concentration in Alzheimer's disease. Mol Neurodegener. 2016;11:3.
- 67. Piccio L, Deming Y, Del-Águila JL, Ghezzi L, Holtzman DM, Fagan AM, Fenoglio C, Galimberti D, Borroni B, Cruchaga C. Cerebrospinal fuid soluble TREM2 is higher in Alzheimer disease and associated with mutation status. Acta Neuropathol. 2016;131:925–33.
- 68. Suárez-Calvet M, Kleinberger G, Araque Caballero M, Brendel M, Rominger A, Alcolea D, Fortea J, Lleó A, Blesa R, Gispert JD, et al. sTREM2 cerebrospinal fuid levels are a potential biomarker for microglia activity in early-stage Alzheimer's disease and associate with neuronal injury markers. EMBO Mol Med. 2016;8:466–76.
- 69. Brosseron F, Traschütz A, Widmann CN, Kummer MP, Tacik P, Santarelli F, Jessen F, Heneka MT. Characterization and clinical use of infammatory cerebrospinal fuid protein markers in Alzheimer's disease. Alzheimers Res Ther. 2018;10:25.
- 70. Henjum K, Almdahl IS, Årskog V, Minthon L, Hansson O, Fladby T, Nilsson LN. Cerebrospinal fuid soluble TREM2 in aging and Alzheimer's disease. Alzheimers Res Ther. 2016;8:17.
- 71. Gispert JD, Suárez-Calvet M, Monté GC, Tucholka A, Falcon C, Rojas S, Rami L, Sánchez-Valle R, Lladó A, Kleinberger G, et al. Cerebrospinal fuid sTREM2 levels are associated with gray matter volume increases and reduced difusivity in early Alzheimer's disease. Alzheimers Dement. 2016;12:1259–72.
- 72. Suárez-Calvet M, Araque Caballero M, Kleinberger G, Bateman RJ, Fagan AM, Morris JC, Levin J, Danek A, Ewers M, Haass C. Early changes in CSF sTREM2 in dominantly inherited Alzheimer's disease occur after amyloid deposition and neuronal injury. Sci Transl Med. 2016;8:369ra178.
- 73. Ma LZ, Tan L, Bi YL, Shen XN, Xu W, Ma YH, Li HQ, Dong Q, Yu JT. Dynamic changes of CSF sTREM2 in preclinical Alzheimer's disease: the CABLE study. Mol Neurodegener. 2020;15:25.
- 74. Suárez-Calvet M, Morenas-Rodríguez E, Kleinberger G, Schlepckow K, Araque Caballero M, Franzmeier N, Capell A, Fellerer K, Nuscher B, Eren E, et al. Early increase of CSF sTREM2 in Alzheimer's disease is associated with tau related-neurodegeneration but not with amyloid-β pathology. Mol Neurodegener. 2019;14:1.
- 75. Schmitz TW, Soreq H, Poirier J, Spreng RN. Longitudinal basal forebrain degeneration interacts with TREM2/C3 biomarkers of infammation in presymptomatic Alzheimer's disease. J Neurosci. 2020;40:1931–42.
- 76. Winfree RL, Dumitrescu L, Blennow K, Zetterberg H, Gifford KA, Pechman KR, Jeferson AL, Hohman TJ. Biological correlates of elevated soluble TREM2 in cerebrospinal fuid. Neurobiol Aging. 2022;118:88–98.
- 77. Edwin TH, Henjum K, Nilsson LNG, Watne LO, Persson K, Eldholm RS, Saltvedt I, Halaas NB, Selbæk G, Engedal K, et al. A high cerebrospinal fuid soluble TREM2 level is associated with slow clinical progression of Alzheimer's disease. Alzheimers Dement (Amst). 2020;12: e12128.
- 78. Morenas-Rodríguez E, Li Y, Nuscher B, Franzmeier N, Xiong C, Suárez-Calvet M, Fagan AM, Schultz S, Gordon BA, Benzinger TLS, et al. Soluble TREM2 in CSF and its association with other biomarkers and cognition in autosomal-dominant Alzheimer's disease: a longitudinal observational study. Lancet Neurol. 2022;21:329–41.
- 79. Hu N, Tan MS, Yu JT, Sun L, Tan L, Wang YL, Jiang T, Tan L. Increased expression of TREM2 in peripheral blood of Alzheimer's disease patients. J Alzheimers Dis. 2014;38:497–501.
- Tan YJ, Ng ASL, Vipin A, Lim JKW, Chander RJ, Ji F, Qiu Y, Ting SKS, Hameed S, Lee TS, et al. Higher peripheral TREM2 mRNA levels relate to cognitive defcits and hippocampal atrophy in Alzheimer's disease and amnestic mild cognitive impairment. J Alzheimers Dis. 2017;58:413–23.
- 81. Mori Y, Yoshino Y, Ochi S, Yamazaki K, Kawabe K, Abe M, Kitano T, Ozaki Y, Yoshida T, Numata S, et al. TREM2 mRNA expression in leukocytes is increased in Alzheimer's disease and schizophrenia. PLoS ONE. 2015;10: e0136835.
- 82. Casati M, Ferri E, Gussago C, Mazzola P, Abbate C, Bellelli G, Mari D, Cesari M, Arosio B. Increased expression of TREM2 in peripheral cells from mild cognitive impairment patients who progress into Alzheimer's disease. Eur J Neurol. 2018;25:805–10.
- 83. Guven G, Bilgic B, Samanci B, Gurvit H, Hanagasi H, Donmez C, Aslan R, Lohmann E, Erginel-Unaltuna N. Peripheral TREM2 mRNA levels in early and late-onset Alzheimer disease's patients. Mol Biol Rep. 2020;47:5903–9.
- 84. Llorens F, Thüne K, Tahir W, Kanata E, Diaz-Lucena D, Xanthopoulos K, Kovatsi E, Pleschka C, Garcia-Esparcia P, Schmitz M, et al. YKL-40 in the brain and cerebrospinal fuid of neurodegenerative dementias. Mol Neurodegener. 2017;12:83.
- 85. Bonneh-Barkay D, Wang G, Starkey A, Hamilton RL, Wiley CA. In vivo CHI3L1 (YKL-40) expression in astrocytes in acute and chronic neurological diseases. J Neuroinfammation. 2010;7:34.
- 86. Baldacci F, Lista S, Cavedo E, Bonuccelli U, Hampel H. Diagnostic function of the neuroinfammatory biomarker YKL-40 in Alzheimer's disease and other neurodegenerative diseases. Expert Rev Proteomics. 2017;14:285–99.
- 87. Arranz AM, De Strooper B. The role of astroglia in Alzheimer's disease: pathophysiology and clinical implications. Lancet Neurol. 2019;18:406–14.
- 88. Stephenson J, Nutma E, van der Valk P, Amor S. Inflammation in CNS neurodegenerative diseases. Immunology. 2018;154:204–19.
- 89. Baldacci F, Lista S, Palermo G, Giorgi FS, Vergallo A, Hampel H. The neuroinfammatory biomarker YKL-40 for neurodegenerative diseases: advances in development. Expert Rev Proteomics. 2019;16:593–600.
- 90. Antonell A, Mansilla A, Rami L, Lladó A, Iranzo A, Olives J, Balasa M, Sánchez-Valle R, Molinuevo JL. Cerebrospinal fluid level of YKL-40 protein in preclinical and prodromal Alzheimer's disease. J Alzheimers Dis. 2014;42:901–8.
- 91. Kester MI, Teunissen CE, Sutphen C, Herries EM, Ladenson JH, Xiong C, Scheltens P, van der Flier WM, Morris JC, Holtzman DM, Fagan AM. Cerebrospinal fuid VILIP-1 and YKL-40, candidate biomarkers to diagnose, predict and monitor Alzheimer's disease in a memory clinic cohort. Alzheimers Res Ther. 2015;7:59.
- 92. Craig-Schapiro R, Perrin RJ, Roe CM, Xiong C, Carter D, Cairns NJ, Mintun MA, Peskind ER, Li G, Galasko DR, et al. YKL-40: a novel prognostic fuid biomarker for preclinical Alzheimer's disease. Biol Psychiatry. 2010;68:903–12.
- 93. Lleó A, Alcolea D, Martínez-Lage P, Scheltens P, Parnetti L, Poirier J, Simonsen AH, Verbeek MM, Rosa-Neto P, Slot RER, et al. Longitudinal cerebrospinal fuid biomarker trajectories along the Alzheimer's disease continuum in the BIOMARKAPD study. Alzheimers Dement. 2019;15:742–53.
- 94. Baldacci F, Toschi N, Lista S, Zetterberg H, Blennow K, Kilimann I, Teipel S, Cavedo E, Dos Santos AM, Epelbaum S, et al. Two-level diagnostic classifcation using cerebrospinal fuid YKL-40 in Alzheimer's disease. Alzheimers Dement. 2017;13:993–1003.
- 95. Paterson RW, Toombs J, Slattery CF, Nicholas JM, Andreasson U, Magdalinou NK, Blennow K, Warren JD, Mummery CJ, Rossor MN, et al. Dissecting IWG-2 typical and atypical Alzheimer's disease: insights from cerebrospinal fuid analysis. J Neurol. 2015;262:2722–30.
- 96. Choi J, Lee HW, Suk K. Plasma level of chitinase 3-like 1 protein increases in patients with early Alzheimer's disease. J Neurol. 2011;258:2181–5.
- 97. Vergallo A, Lista S, Lemercier P, Chiesa PA, Zetterberg H, Blennow K, Potier MC, Habert MO, Baldacci F, Cavedo E, et al. Association of plasma YKL-40 with brain amyloid-β levels, memory performance, and sex in subjective memory complainers. Neurobiol Aging. 2020;96:22–32.
- 98. Grewal R, Haghighi M, Huang S, Smith AG, Cao C, Lin X, Lee DC, Teten N, Hill AM, Selenica MB. Identifying biomarkers of dementia prevalent among amnestic mild cognitively impaired ethnic female patients. Alzheimers Res Ther. 2016;8:43.
- 99. Patani R, Hardingham GE, Liddelow SA. Functional roles of reactive astrocytes in neuroinfammation and neurodegeneration. Nat Rev Neurol. 2023;19:395–409.
- 100. Hol EM, Pekny M. Glial fbrillary acidic protein (GFAP) and the astrocyte intermediate flament system in diseases of the central nervous system. Curr Opin Cell Biol. 2015;32:121–30.
- 101. Garwood CJ, Ratclife LE, Simpson JE, Heath PR, Ince PG, Wharton SB. Review: astrocytes in Alzheimer's disease and other age-associated dementias: a supporting player with a central role. Neuropathol Appl Neurobiol. 2017;43:281–98.
- 102. Carter SF, Schöll M, Almkvist O, Wall A, Engler H, Långström B, Nordberg A. Evidence for astrocytosis in prodromal Alzheimer disease provided by 11C-deuterium-L-deprenyl: a multitracer PET paradigm combining 11C-Pittsburgh compound B and 18F-FDG. J Nucl Med. 2012;53:37–46.
- 103. Muramori F, Kobayashi K, Nakamura I. A quantitative study of neurofibrillary tangles, senile plaques and astrocytes in the hippocampal subdivisions and entorhinal cortex in Alzheimer's disease, normal controls and non-Alzheimer neuropsychiatric diseases. Psychiatry Clin Neurosci. 1998;52:593–9.
- 104. Ishiki A, Kamada M, Kawamura Y, Terao C, Shimoda F, Tomita N, Arai H, Furukawa K. Glial fbrillar acidic protein in the cerebrospinal fuid of Alzheimer's disease, dementia with Lewy bodies, and frontotemporal lobar degeneration. J Neurochem. 2016;136:258–61.
- 105. Elahi FM, Casaletto KB, La Joie R, Walters SM, Harvey D, Wolf A, Edwards L, Rivera-Contreras W, Karydas A, Cobigo Y, et al. Plasma biomarkers of astrocytic and neuronal dysfunction in early- and late-onset Alzheimer's disease. Alzheimers Dement. 2020;16:681–95.
- 106. Oeckl P, Halbgebauer S, Anderl-Straub S, Steinacker P, Huss AM, Neugebauer H, von Arnim CAF, Diehl-Schmid J, Grimmer T, Kornhuber J, et al. Glial fbrillary acidic protein in serum is increased in Alzheimer's disease and correlates with cognitive impairment. J Alzheimers Dis. 2019;67:481–8.
- 107. Verberk IMW, Thijssen E, Koelewijn J, Mauroo K, Vanbrabant J, de Wilde A, Zwan MD, Verfaillie SCJ, Ossenkoppele R, Barkhof F, et al. Combination of plasma amyloid beta((1-42/1-40)) and glial fibrillary acidic protein strongly associates with cerebral amyloid pathology. Alzheimers Res Ther. 2020;12:118.
- 108. Chatterjee P, Pedrini S, Stoops E, Goozee K, Villemagne VL, Asih PR, Verberk IMW, Dave P, Taddei K, Sohrabi HR, et al. Plasma glial fbrillary acidic protein is elevated in cognitively normal older adults at risk of Alzheimer's disease. Transl Psychiatry. 2021;11:27.
- 109. Cicognola C, Janelidze S, Hertze J, Zetterberg H, Blennow K, Mattsson-Carlgren N, Hansson O. Plasma glial fbrillary acidic protein detects Alzheimer pathology and predicts future conversion to Alzheimer dementia in patients with mild cognitive impairment. Alzheimers Res Ther. 2021;13:68.
- 110. O'Connor A, Abel E, Benedet AL, Poole T, Ashton N, Weston PSJ, Heslegrave AJ, Ryan N, Barker S, Polke JM, et al. Plasma GFAP in presymptomatic and symptomatic familial Alzheimer's disease: a longitudinal cohort study. J Neurol Neurosurg Psychiatry. 2023;94:90–2.
- 111. Silva-Spínola A, Lima M, Leitão MJ, Bernardes C, Durães J, Duro D, Tábuas-Pereira M, Santana I, Baldeiras I. Blood biomarkers in mild cognitive impairment patients: relationship between analytes and progression to Alzheimer disease dementia. Eur J Neurol. 2023;30:1565–73.
- 112. Chatterjee P, Pedrini S, Ashton NJ, Tegg M, Goozee K, Singh AK, Karikari TK, Simrén J, Vanmechelen E, Armstrong NJ, et al. Diagnostic and prognostic plasma biomarkers for preclinical Alzheimer's disease. Alzheimers Dement. 2022;18:1141–54.
- 113. Pereira JB, Janelidze S, Smith R, Mattsson-Carlgren N, Palmqvist S, Teunissen CE, Zetterberg H, Stomrud E, Ashton NJ, Blennow K, Hansson O. Plasma GFAP is an early marker of amyloid-β but not tau pathology in Alzheimer's disease. Brain. 2021;144:3505–16.
- 114. Bellaver B, Povala G, Ferreira PCL, Ferrari-Souza JP, Lefa DT, Lussier FZ, Benedet AL, Ashton NJ, Triana-Baltzer G, Kolb HC, et al. Astrocyte reactivity infuences amyloid-β efects on tau pathology in preclinical Alzheimer's disease. Nat Med. 2023;29:1775–81.
- 115. Verberk IMW, Laarhuis MB, van den Bosch KA, Ebenau JL, van Leeuwenstijn M, Prins ND, Scheltens P, Teunissen CE, van der Flier WM. Serum markers glial fbrillary acidic protein and neuroflament light for prognosis and monitoring in cognitively normal older people: a prospective memory clinic-based cohort study. Lancet Healthy Longev. 2021;2:e87–95.
- 116. Stocker H, Beyer L, Perna L, Rujescu D, Holleczek B, Beyreuther K, Stockmann J, Schöttker B, Gerwert K, Brenner H. Association of plasma biomarkers, p-tau181, glial fbrillary acidic protein, and neuroflament light, with intermediate and long-term clinical Alzheimer's disease risk: results from a prospective cohort followed over 17 years. Alzheimers Dement. 2023;19:25–35.
- 117. Liddelow SA, Guttenplan KA, Clarke LE, Bennett FC, Bohlen CJ, Schirmer L, Bennett ML, Münch AE, Chung WS, Peterson TC, et al. Neurotoxic reactive astrocytes are induced by activated microglia. Nature. 2017;541:481–7.
- 118. Cronjé HT, Liu X, Odden MC, Moseholm KF, Seshadri S, Satizabal CL, Lopez OL, Bis JC, Djoussé L, Fohner AE, et al. Serum NfL and GFAP are associated with incident dementia and dementia mortality in older adults: the cardiovascular health study. Alzheimers Dement. 2023.
- 119. Varma VR, An Y, Kac PR, Bilgel M, Moghekar A, Loeffler T, Amschl D, Troncoso J, Blennow K, Zetterberg H, et al. Longitudinal progression of blood biomarkers reveals a key role of astrocyte reactivity in preclinical Alzheimer's disease. medRxiv 2024.
- 120. Kapoor M, Chinnathambi S. TGF-β1 signalling in Alzheimer's pathology and cytoskeletal reorganization: a specialized Tau perspective. J Neuroinfammation. 2023;20:72.
- 121. Caraci F, Spampinato SF, Morgese MG, Tascedda F, Salluzzo MG, Giambirtone MC, Caruso G, Munafò A, Torrisi SA, Leggio GM, et al. Neurobiological links between depression and AD: the role of TGF-β1 signaling as a new pharmacological target. Pharmacol Res. 2018;130:374–84.
- 122. Tichauer JE, von Bernhardi R. Transforming growth factor-β stimulates β amyloid uptake by microglia through Smad3-dependent mechanisms. J Neurosci Res. 2012;90:1970–80.
- 123. Caraci F, Gulisano W, Guida CA, Impellizzeri AA, Drago F, Puzzo D, Palmeri A. A key role for TGF-β1 in hippocampal synaptic plasticity and memory. Sci Rep. 2015;5:11252.
- 124. Caraci F, Spampinato S, Sortino MA, Bosco P, Battaglia G, Bruno V, Drago F, Nicoletti F, Copani A. Dysfunction of TGF-β1 signaling in Alzheimer's disease: perspectives for neuroprotection. Cell Tissue Res. 2012;347:291–301.
- 125. Tesseur I, Zou K, Esposito L, Bard F, Berber E, Can JV, Lin AH, Crews L, Tremblay P, Mathews P, et al. Defciency in neuronal TGF-beta signaling promotes neurodegeneration and Alzheimer's pathology. J Clin Invest. 2006;116:3060–9.
- 126. Juraskova B, Andrys C, Holmerova I, Solichova D, Hrnciarikova D, Vankova H, Vasatko T, Krejsek J. Transforming growth factor beta and soluble endoglin in the healthy senior and in Alzheimer's disease patients. J Nutr Health Aging. 2010;14:758–61.
- 127. Luppi C, Fioravanti M, Bertolini B, Inguscio M, Grugnetti A, Guerriero F, Rovelli C, Cantoni F, Guagnano P, Marazzi E, et al. Growth factors decrease in subjects with mild to moderate Alzheimer's disease (AD): potential correction with dehydroepiandrosterone-sulphate (DHEAS). Arch Gerontol Geriatr. 2009;49(Suppl 1):173–84.
- 128. Wyss-Coray T. Tgf-Beta pathway as a potential target in neurodegeneration and Alzheimer's. Curr Alzheimer Res. 2006;3:191–5.
- 129. Tarkowski E, Issa R, Sjögren M, Wallin A, Blennow K, Tarkowski A, Kumar P. Increased intrathecal levels of the angiogenic factors VEGF and TGFbeta in Alzheimer's disease and vascular dementia. Neurobiol Aging. 2002;23:237–43.
- 130. Mocali A, Cedrola S, Della Malva N, Bontempelli M, Mitidieri VA, Bavazzano A, Comolli R, Paoletti F, La Porta CA. Increased plasma levels of soluble CD40, together with the decrease of TGF beta 1, as possible differential markers of Alzheimer disease. Exp Gerontol. 2004;39:1555–61.
- 131. Gogishvili D, Vromen EM, Koppes-den Hertog S, Lemstra AW, Pijnenburg YAL, Visser PJ, Tijms BM, Del Campo M, Abeln S, Teunissen CE,

Vermunt L. Discovery of novel CSF biomarkers to predict progression in dementia using machine learning. Sci Rep. 2023;13:6531.

- 132. Bosco P, Ferri R, Salluzzo MG, Castellano S, Signorelli M, Nicoletti F, Nuovo SD, Drago F, Caraci F. Role of the transforming-growth-factor-β1 gene in late-onset Alzheimer's disease: implications for the treatment. Curr Genomics. 2013;14:147–56.
- 133. Arosio B, Bergamaschini L, Galimberti L, La Porta C, Zanetti M, Calabresi C, Scarpini E, Annoni G, Vergani C. +10 T/C polymorphisms in the gene of transforming growth factor-beta1 are associated with neurodegeneration and its clinical evolution. Mech Ageing Dev. 2007;128:553–7.
- 134. Caraci F, Bosco P, Signorelli M, Spada RS, Cosentino FI, Toscano G, Bonforte C, Muratore S, Prestianni G, Panerai S, et al. The CC genotype of transforming growth factor-β1 increases the risk of late-onset Alzheimer's disease and is associated with AD-related depression. Eur Neuropsychopharmacol. 2012;22:281–9.
- 135. Fraga VG, Guimarães HC, Lara VP, Teixeira AL, Barbosa MT, Carvalho MG, Caramelli P, Gomes KB. TGF-β1 Codon 10 T>C polymorphism infuences short-term functional and cognitive decline in healthy oldest-old individuals: the Pietà study. J Alzheimers Dis. 2015;48:1077–81.
- 136. Estrada LD, Oliveira-Cruz L, Cabrera D. Transforming growth factor beta type I role in neurodegeneration: implications for Alzheimer's disease. Curr Protein Pept Sci. 2018;19:1180–8.
- 137. Yin Z, Rosenzweig N, Kleemann KL, Zhang X, Brandão W, Margeta MA, Schroeder C, Sivanathan KN, Silveira S, Gauthier C, et al. APOE4 impairs the microglial response in Alzheimer's disease by inducing TGFβmediated checkpoints. Nat Immunol. 2023;24:1839–53.
- 138. Brosseron F, Maass A, Kleineidam L, Ravichandran KA, González PG, McManus RM, Ising C, Santarelli F, Kolbe CC, Häsler LM, et al. Soluble TAM receptors sAXL and sTyro3 predict structural and functional protection in Alzheimer's disease. Neuron. 2022;110:1009-1022.e1004.
- 139. Popp J, Oikonomidi A, Tautvydaitė D, Dayon L, Bacher M, Migliavacca E, Henry H, Kirkland R, Severin I, Wojcik J, Bowman GL. Markers of neuroinfammation associated with Alzheimer's disease pathology in older adults. Brain Behav Immun. 2017;62:203–11.
- 140. Morgan AR, Touchard S, Leckey C, O'Hagan C, Nevado-Holgado AJ, Barkhof F, Bertram L, Blin O, Bos I, Dobricic V, et al. Inflammatory biomarkers in Alzheimer's disease plasma. Alzheimers Dement. 2019;15:776–87.
- 141. Swardfager W, Lanctôt K, Rothenburg L, Wong A, Cappell J, Herrmann N. A meta-analysis of cytokines in Alzheimer's disease. Biol Psychiatry. 2010;68:930–41.
- 142. Lai KSP, Liu CS, Rau A, Lanctôt KL, Köhler CA, Pakosh M, Carvalho AF, Herrmann N. Peripheral infammatory markers in Alzheimer's disease: a systematic review and meta-analysis of 175 studies. J Neurol Neurosurg Psychiatry. 2017;88:876–82.
- 143. Shen XN, Niu LD, Wang YJ, Cao XP, Liu Q, Tan L, Zhang C, Yu JT. Inflammatory markers in Alzheimer's disease and mild cognitive impairment: a meta-analysis and systematic review of 170 studies. J Neurol Neurosurg Psychiatry. 2019;90:590–8.
- 144. Su C, Zhao K, Xia H, Xu Y. Peripheral inflammatory biomarkers in Alzheimer's disease and mild cognitive impairment: a systematic review and meta-analysis. Psychogeriatrics. 2019;19:300–9.
- 145. Darweesh SKL, Wolters FJ, Ikram MA, de Wolf F, Bos D, Hofman A. Inflammatory markers and the risk of dementia and Alzheimer's disease: a meta-analysis. Alzheimers Dement. 2018;14:1450–9.
- 146. Brosseron F, Maass A, Kleineidam L, Ravichandran KA, Kolbe CC, Wolfsgruber S, Santarelli F, Häsler LM, McManus R, Ising C, et al. Serum IL-6, sAXL, and YKL-40 as systemic correlates of reduced brain structure and function in Alzheimer's disease: results from the DELCODE study. Alzheimers Res Ther. 2023;15:13.
- 147. Liu P, Wang Y, Sun Y, Peng G. Neuroinflammation as a potential therapeutic target in Alzheimer's disease. Clin Interv Aging. 2022;17:665–74.
- 148. van Dyck CH, Swanson CJ, Aisen P, Bateman RJ, Chen C, Gee M, Kanekiyo M, Li D, Reyderman L, Cohen S, et al. Lecanemab in early Alzheimer's disease. N Engl J Med. 2023;388:9–21.
- 149. Pontecorvo MJ, Lu M, Burnham SC, Schade AE, Dage JL, Shcherbinin S, Collins EC, Sims JR, Mintun MA. Association of donanemab treatment with exploratory plasma biomarkers in early symptomatic alzheimer disease: a secondary analysis of the TRAILBLAZER-ALZ randomized clinical trial. JAMA Neurol. 2022;79:1250–9.
- 150. O'Bryant SE, Zhang F, Johnson LA, Hall J, Edwards M, Grammas P, Oh E, Lyketsos CG, Rissman RA. A precision medicine model for targeted NSAID therapy in Alzheimer's disease. J Alzheimers Dis. 2018;66:97–104.
- 151. Mintun MA, Lo AC, Duggan Evans C, Wessels AM, Ardayfo PA, Andersen SW, Shcherbinin S, Sparks J, Sims JR, Brys M, et al. Donanemab in early Alzheimer's disease. N Engl J Med. 2021;384:1691–704.
- 152. Rafi MS, Sperling RA, Donohue MC, Zhou J, Roberts C, Irizarry MC, Dhadda S, Sethuraman G, Kramer LD, Swanson CJ, et al. The AHEAD 3-45 study: design of a prevention trial for Alzheimer's disease. Alzheimers Dement. 2023;19:1227-33.
- 153. Beyer L, Stocker H, Rujescu D, Holleczek B, Stockmann J, Nabers A, Brenner H, Gerwert K. Amyloid-beta misfolding and GFAP predict risk of clinical Alzheimer's disease diagnosis within 17 years. Alzheimers Dement. 2022;19:1020.
- 154. Hampel H, Elhage A, Cho M, Apostolova LG, Nicoll JAR, Atri A. Amyloidrelated imaging abnormalities (ARIA): radiological, biological and clinical characteristics. Brain. 2023;146:4414–24.
- 155. Love S, Miners JS. Cerebrovascular disease in ageing and Alzheimer's disease. Acta Neuropathol. 2016;131:645–58.
- 156. Janelidze S, Mattsson N, Stomrud E, Lindberg O, Palmqvist S, Zetterberg H, Blennow K, Hansson O. CSF biomarkers of neuroinflammation and cerebrovascular dysfunction in early Alzheimer disease. Neurology. 2018;91:e867–77.
- 157. Group F-NBW: In *BEST (Biomarkers, EndpointS, and other Tools) Resource.* Silver Spring (MD) Bethesda (MD): Food and Drug Administration (US) National Institutes of Health (US); 2016.
- 158. Sun N, Victor MB, Park YP, Xiong X, Scannail AN, Leary N, Prosper S, Viswanathan S, Luna X, Boix CA, et al. Human microglial state dynamics in Alzheimer's disease progression. Cell. 2023;186:4386-4403.e4329.
- 159. Wightman DP, Jansen IE, Savage JE, Shadrin AA, Bahrami S, Holland D, Rongve A, Børte S, Winsvold BS, Drange OK, et al. A genome-wide association study with 1,126,563 individuals identifes new risk loci for Alzheimer's disease. Nat Genet. 2021;53:1276–82.
- 160. Griciuc A, Tanzi RE. The role of innate immune genes in Alzheimer's disease. Curr Opin Neurol. 2021;34:228–36.
- 161. Wang S, Mustafa M, Yuede CM, Salazar SV, Kong P, Long H, Ward M, Siddiqui O, Paul R, Gilfllan S, et al. Anti-human TREM2 induces microglia proliferation and reduces pathology in an Alzheimer's disease model. J Exp Med. 2020;217:e20200785.
- 162. Yeo IJ, Lee CK, Han SB, Yun J, Hong JT. Roles of chitinase 3-like 1 in the development of cancer, neurodegenerative diseases, and infammatory diseases. Pharmacol Ther. 2019;203: 107394.
- Alcolea D, Carmona-Iragui M, Suárez-Calvet M, Sánchez-Saudinós MB, Sala I, Antón-Aguirre S, Blesa R, Clarimón J, Fortea J, Heó A, Relationship between β-Secretase, infammation and core cerebrospinal fuid biomarkers for Alzheimer's disease. J Alzheimers Dis. 2014;42:157–67.
- 164. Wilczyńska K, Maciejczyk M, Zalewska A, Waszkiewicz N. Serum amyloid biomarkers, tau protein and YKL-40 utility in detection, differential diagnosing, and monitoring of dementia. Front Psychiatry. 2021;12: 725511.
- 165. Shue F, White LJ, Hendrix R, Ulrich J, Henson RL, Knight W, Martens YA, Wang N, Roy B, Starling SC, et al. CSF biomarkers of immune activation and Alzheimer's disease for predicting cognitive impairment risk in the elderly. Sci Adv. 2024;10:eadk3674.
- 166. Ewers M, Franzmeier N, Suárez-Calvet M, Morenas-Rodriguez E, Caballero MAA, Kleinberger G, Piccio L, Cruchaga C, Deming Y, Dichgans M, et al. Increased soluble TREM2 in cerebrospinal fuid is associated with reduced cognitive and clinical decline in Alzheimer's disease. Sci Transl Med. 2019; 11.
- 167. Zhao A, Jiao Y, Ye G, Kang W, Tan L, Li Y, Deng Y, Liu J. Soluble TREM2 levels associate with conversion from mild cognitive impairment to Alzheimer's disease. J Clin Invest. 2022; 132.
- 168. Bieger A, Rocha A, Bellaver B, Machado L, Da Ros L, Borelli WV, Therriault J, Macedo AC, Pascoal TA, Gauthier S, et al. Neuroinflammation biomarkers in the AT(N) framework across the Alzheimer's disease continuum. J Prev Alzheimers Dis. 2023;10:401–17.
- 169. Abdelhak A, Foschi M, Abu-Rumeileh S, Yue JK, D'Anna L, Huss A, Oeckl P, Ludolph AC, Kuhle J, Petzold A, et al. Blood GFAP as an emerging biomarker in brain and spinal cord disorders. Nat Rev Neurol. 2022;18:158–72.
- 170. Wang Q, Xu Y, Qi C, Liu A, Zhao Y. Association study of serum soluble TREM2 with vascular dementia in Chinese Han population. Int J Neurosci. 2020;130:708–12.
- 171. Limberger C, Zimmer ER. Blood GFAP reflects astrocyte reactivity to Alzheimer's pathology in post-mortem brain tissue. Brain. 2024;147:1598–600.
- 172. Hampel H, Hu Y, Cummings J, Mattke S, Iwatsubo T, Nakamura A, Vellas B, O'Bryant S, Shaw LM, Cho M, et al. Blood-based biomarkers for Alzheimer's disease: current state and future use in a transformed global healthcare landscape. Neuron. 2023;111:2781–99.
- 173. Hansson O, Edelmayer RM, Boxer AL, Carrillo MC, Mielke MM, Rabinovici GD, Salloway S, Sperling R, Zetterberg H, Teunissen CE. The Alzheimer's Association appropriate use recommendations for blood biomarkers in Alzheimer's disease. Alzheimers Dement. 2022;18:2669–86.
- 174. Tybirk L, Hviid CVB, Knudsen CS, Parkner T. Serum GFAP—reference interval and preanalytical properties in Danish adults. Clin Chem Lab Med. 2022;60:1830–8.
- 175. Benedet AL, Milà-Alomà M, Vrillon A, Ashton NJ, Pascoal TA, Lussier F, Karikari TK, Hourregue C, Cognat E, Dumurgier J, et al. Differences between plasma and cerebrospinal fuid glial fbrillary acidic protein levels across the Alzheimer disease continuum. JAMA Neurol. 2021;78:1471–83.
- 176. Olsson B, Hertze J, Lautner R, Zetterberg H, Nägga K, Höglund K, Basun H, Annas P, Lannfelt L, Andreasen N, et al. Microglial markers are elevated in the prodromal phase of Alzheimer's disease and vascular dementia. J Alzheimers Dis. 2013;33:45–53.
- 177. Yakoub Y, Ashton NJ, Strikwerda-Brown C, Montoliu-Gaya L, Karikari TK, Kac PR, Gonzalez-Ortiz F, Gallego-Rudolf J, Meyer PF, St-Onge F, et al. Longitudinal blood biomarker trajectories in preclinical Alzheimer's disease. Alzheimers Dement. 2023;19:5620.
- 178. Yaffe K, Lindquist K, Penninx BW, Simonsick EM, Pahor M, Kritchevsky S, Launer L, Kuller L, Rubin S, Harris T. Infammatory markers and cognition in well-functioning African-American and white elders. Neurology. 2003;61:76–80.
- 179. Tan ZS, Beiser AS, Vasan RS, Roubenoff R, Dinarello CA, Harris TB, Benjamin EJ, Au R, Kiel DP, Wolf PA, Seshadri S. Infammatory markers and the risk of Alzheimer disease: the Framingham Study. Neurology. 2007;68:1902–8.
- 180. Caldwell JZK, Kinney JW, Ritter A, Salazar A, Wong CG, Cordes D, Slavich GM. Infammatory cytokine levels implicated in Alzheimer's disease moderate the efects of sex on verbal memory performance. Brain Behav Immun. 2021;95:27–35.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional afliations.