## **GENOME RESOURCE PAPER (GRP)**



## Draft genome sequence of the keylime (*Citrus* × *aurantiifolia*) pathogen *Colletotrichum limetticola*

Andrea Menicucci<sup>1</sup> · Isis Tikami<sup>1,2</sup> · Tiziano Benocci<sup>3</sup> · Antonio Zapparata<sup>4</sup> · Nelson Sidnei Massola Júnior<sup>2</sup> · Natalia Aparecida Peres<sup>5</sup> · Lavern Wayne Timmer<sup>5</sup> · Antonio Prodi<sup>1</sup> · Riccardo Baroncelli<sup>1,6</sup>

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Many species belonging to the genus Colletotrichum are causal agents of plant diseases, generally referred to as anthracnose, in a wide range of hosts worldwide. Colletotrichum spp. are responsible for impacting numerous economically important crops on a global scale. This genus comprises approximately 257 distinct species, which are further organized into at least 15 major phylogenetic lineages known as species complexes (Talhinhas and Baroncelli 2021). Virtually every crop grown in the world is susceptible to one or more species of Colletotrichum (Baroncelli et al. 2014). Among these, the *Colletotrichum acutatum* species complex stands out as a diverse group of closely related plant pathogenic fungi within the genus (Baroncelli et al. 2017). Members of the Colletotrichum acutatum species complex have a wide host range in both domesticated and wild plant species, and their capability to infect insects has also been described (Damn et al. 2012, Marcelino et al. 2008). In this species complex, Colletotrichum limetticola (formerly known as Gloeosporium limetticola; Clausen

Andrea Menicucci and Isis Tikami contributed equally to this work.

Riccardo Baroncelli riccardo.baroncelli@unibo.it

- <sup>1</sup> Department of Agricultural and Food Sciences (DISTAL), University of Bologna, Bologna, Italy
- <sup>2</sup> Escola Superior de Agricultura Luiz de Queiroz (ESALQ), University of São Paulo (USP), Piracicaba, São Paulo, Brazil
- <sup>3</sup> Center for Health and Bioresources, AIT Austrian Institute of Technology, Tulln, Austria
- <sup>4</sup> "Federigo Enriques" High School, Livorno, Italy
- <sup>5</sup> Plant Pathology Department, Gulf Coast Research and Education Center, University of Florida, Wimauma, USA
- <sup>6</sup> Center for Studies on Bioinspired Agro-Environmental Technology, University of Naples Federico II, Portici, Italy

1912) was initially described in 2012 as a species predominantly associated with wither tip symptoms on sour lime (Citrus aurantiifolia) in Cuba and the USA during the 1910s (Damm et al. 2012). Later descriptions associated the disease with strains of C. gloeosporioides (Brown et al. 1996) or C. acutatum (Peres et al. 2008). Recent findings in Brazil have revealed the presence of C. limetticola causing Glomerella leaf spot on apples, although its prevalence remains low while displaying high virulence (Moreira et al. 2019). To the best of our knowledge, no further occurrences of C. *limetticola* have been documented, despite the presence of other known Colletotrichum species that infect citrus and apples (Talhinhas and Baroncelli 2021). This raises concerns regarding the conservation status of C. limetticola considering the scarcity of records on its original hosts and the occurrence of cross-infections.

In the present study, Colletotrichum limetticola strain KLA-Anderson was isolated from a leaf tissue of Citrus x aurantiifolia commonly known as the Key lime or Mexican lime in the Lake Alfred region (Florida, USA). C. limetticola genome was sequenced using the Illumina NovaSeq 6000 150 bp paired-end sequencing system. Illumina sequences were analyzed with FastQC (Babraham Bioinformatics) to assess the quality of the reads. Sequences adapters and low-quality reads were trimmed with TrimGalore! v0.6.10 (Krueger et al. 2021). Pairend reads were merged with FLASH v1.2.11 (Magoc and Salzberg 2011). Merged and unmerged reads were then assembled using SPAdes v3.15.1 (Bankevich et al. 2012). Scaffolds with low coverage were removed as possible contaminations. Scaffolds corresponding to the mitochondrial DNA (mtDNA) and ribosomal DNA (rDNA) genome were identified by BLASTN v2.9.0 (Camacho et al. 2009) using queries of the closely related species Colletotrichum lupini (Baroncelli et al. 2021) which was

 
 Table 1
 Summary statistics of the Collectorichum limetticola KLA-Anderson strain genome

Genome variables	Statistics							
Assembly statistics								
Number of scaffolds	1750							
Assembly length (Mb)	50.48							
Maximum scaffold size (Mb)	0.38							
N50	68638							
N90	17501							
L50	229							
L90	752							
Guanine-Cytosine content (%)	52.77							
BUSCO completeness								
Complete (%)	97.7							
Single copy (%)	97.6							
Duplicate (%)	0.1							
Fragmented (%)	1.1							
Missing (%)	1.2							
Protein encoding genes								
Number of predicted proteins	15248							
Number of secreted proteins	1981							
Number of predicted effector proteins	624							
Number of Predicted cytoplasmic effectors	172							
Number of Predicted apoplastic effectors	452							

the closest complete genome to *C. limetticola*. The completeness of the assembly was assessed using BUSCO v5.4.7 (Simão et al. 2015) while statistics were evaluated

with OUAST v5.2.0 (Gurevich et al. 2013). The total size of the nuclear genome assembly was 50,48 Mb, with an N50 contig length of 68638 kb and a L50 of 229. The nuclear genome assembly resulted in 1750 contigs with an average coverage of 90X and it was assessed to be 97.7% complete (Table 1). A total of 15248 protein-coding genes were predicted to be encoded using MAKER v3.01.02 pipeline (Holt and Yandell 2011) with both self-trained GeneMark-ES v4.10 (Borodovsky and Lomsadze 2011) and AUGUSTUS v3.3 prediction using the "Colletotrichum" model (Becerra et al. 2023). SignalP v5.0 (Almagro Armenteros et al. 2019) revealed that 1981 proteins in C. limetticola are secreted and among those 624 have been predicted by EffectorP v3.0 (Sperschneider and Dodds 2022) to be candidate effectors. A comparative analysis of the newly sequenced genome with those publicly available (Baroncelli et al. 2016, 2021, 2022; Goulin et al. 2023) revealed similar genomic features and gene content within closely related species (Fig. 1).

In this study we presented a draft genome sequence of *C. limetticola*, obtained using Illumina sequencing technology, providing a range of new resources that serve as a useful platform for further research in the field of comparative genomics of fungi. Further analysis of these genomes will enhance our understanding of the molecular mechanisms underlying the pathogenicity and virulence of *Colletotrichum* species facilitating the exploration of potential targeted and environmentally friendly strategies for its control.

	Species	Strain	Contigs	Size	N50	L50	GC%	Busco	Gene content	
	C. simmondsii	CBS 122122	929	50474234 🔴	292136 •	55 🔵	51.84 🔴	98.1 🔵	12013	1871
	C. paranaense	IMI 384185	36	49177916 🔴	3374443 🔵	6	52.23 🔴	98.3	14636	2139
∎––	C. limetticola	KLA-Anderson	1750	50483870 🛑	68638 •	229	52.77 🛑	97.7	13267	1981
0.90/61/51	C. melonis	CBS 134730	92 •	50135825 🛑	2830390 🔵	7 •	52.15 🔴	98.3	14668	2155
0.002	C. cuscutae	IMI 304802	325 🔴	80453496 🔵	514068 •	47 🔴	41.34 🔴	98.0	15409	1880
	C. filicis	CBS 101611	367 🔴	62965231 🔵	325017 •	60	45.98 🔴	98.1	15295	2096
1.00/89/-	C. abscissum	IMI 504890	560 🔵	53936108 🛑	962862 🔵	19	51.11 🛑	98.3 🔵	15126	2154
	C. abscissum	LGMF1258	423 🌑	54001896 🛑	321765 •	46 🔴	51.12 🔴	98.3 🔵	13479	2020
1.00/95/92	C. costaricense	IMI 309622	70 •	51625248 🛑	2423109 🔵	8 •	51.36 🛑	98.4	14965	2161
1.00/100/94	C. tamarilloi	CBS 129955	604 🔵	52077224 🔴	410822 •	30 🔴	51.2 🔴	98.2	15270	2129
0.94/-/55	C. lupini	CBS 109225	116 •	58756634 🔴	918987 👤	20	47.58 🔴	97.4	13410	1955
	C. lupini	IMI 504893	11 ·	63407421 🔴	7871922	4 •	46.71 🔴	98.2	16557	1767

**Fig. 1** Comparative analysis between the newly sequenced genome of *C. limetticola* and a selection of closely related species publicly available. The *C. limetticola* genome is highlighted in bold. On the left side, multilocus sequence typing (MLST) tree based on the concatenation of the partial sequences of following loci: actin [ACT], beta-tubulin 2 [TUB2], calmodulin [CAL], glyceraldehyde-3-phosphate dehydrogenase [GAPDH], chitin synthase [CHS-1], glutamine synthetase [GS], histone-3 [HIS3], superoxide dismutase 2 [SOD2]

mating type 1–2 [MAT1-2] and the Apn2-Mat1-2 intergenic spacer [ApnMat]. Numbers next to the nodes represent in order: Bayesian posterior probability, FastTree and RAxML bootstrap support values. Bubble plots report on assembly fragmentation, genome size, N50 and L50, GC content and completeness. Bubble sizes have been scaled to each panel and are not comparable across panels. Horizontal histograms report on secreted and non-secreted protein coding gene content **Funding** Open access funding provided by Alma Mater Studiorum - Università di Bologna within the CRUI-CARE Agreement. This study was carried out within the Agritech National Research Center and received funding from the European Union Next-GenerationEU (PIANO NAZIONALE DI RIPRESA E RESILIENZA (PNRR) – MISSIONE 4 COMPONENTE 2, INVESTIMENTO 1.4 – D.D. 1032 17/06/2022, CN00000022). This manuscript reflects only the authors' views and opinions, neither the European Union nor the European Commission can be considered responsible for them and the Coordination for the Improvement of Higher Education Personnel (CAPES), Brazil, Grant/ Award Number: 8887.695424/2022–00.

**Data availability** The data generated in this study are publicly available from the NCBI GenBank database at Bioproject ID PRJNA952538 and Biosample ID SAMN34075281. This Whole Genome Shotgun project has been deposited at DDBJ/ENA/GenBank under the accession JARUPT000000000. The version described in this paper is version JARUPT010000000.

## Declarations

**Conflict of interest** The authors have no relevant financial or non-financial interests to disclose.

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