



ALMA MATER STUDIORUM
UNIVERSITÀ DI BOLOGNA

ARCHIVIO ISTITUZIONALE
DELLA RICERCA

Alma Mater Studiorum Università di Bologna Archivio istituzionale della ricerca

Evaluation of polycrystalline cerium oxide electrodes for electrochemiluminescent detection of sarcosine

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Bahrami, H., Kosto, Y., Santo, C.I., Yakovlev, Y., Khalakhan, I., Mousavi, M., et al. (2023). Evaluation of polycrystalline cerium oxide electrodes for electrochemiluminescent detection of sarcosine. MICROCHEMICAL JOURNAL, 187, 1-6 [10.1016/j.microc.2022.108362].

Availability:

This version is available at: <https://hdl.handle.net/11585/958943> since: 2024-02-18

Published:

DOI: <http://doi.org/10.1016/j.microc.2022.108362>

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>).
When citing, please refer to the published version.

(Article begins on next page)

This is the final peer-reviewed accepted manuscript of:

Hengameh Bahrami, Yuliia Kosto, Claudio Ignazio Santo, Yurii Yakovlev, Ivan Khalakhan, Mehdi Mousavi, Vladimir Matolin, Iva Matolinová, Francesco Paolucci, Giovanni Valenti, Nataliya Tsud, Alessandra Zanut

Evaluation of polycrystalline cerium oxide electrodes for electrochemiluminescent detection of sarcosine

Microchemical Journal 187 (2023) 108362

The final published version is available online at:

<https://doi.org/10.1016/j.microc.2022.108362>

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>)

When citing, please refer to the published version.

Evaluation of polycrystalline cerium oxide electrodes for electrochemiluminescent detection of sarcosine

Hengameh Bahrami¹, Yuliia Kosto^{2†}, Claudio Ignazio Santo³, Yurii Yakovlev², Ivan Khalakhan², Mehdi Mousavi¹, Vladimir Matolin², Iva Matolinová², Francesco Paolucci³, Giovanni Valenti³, Nataliya Tsud^{2*}, Alessandra Zanut^{3*††}

¹ Shahid Bahonar University of Kerman, Faculty of Sciences, Department of Chemistry, 76169-14111 Kerman, Iran

² Charles University, Faculty of Mathematics and Physics, Department of Surface and Plasma Science, V Holešovičkách 2, 18000 Prague, Czech Republic

³ University of Bologna, Department of Chemistry, F. Selmi 2, 40126 Bologna, Italy

† Current affiliation: Brandenburg University of Technology Cottbus-Senftenberg, Applied Physics & Semiconductor Spectroscopy, Konrad-Zuse Str 1, D-03046 Cottbus, Germany

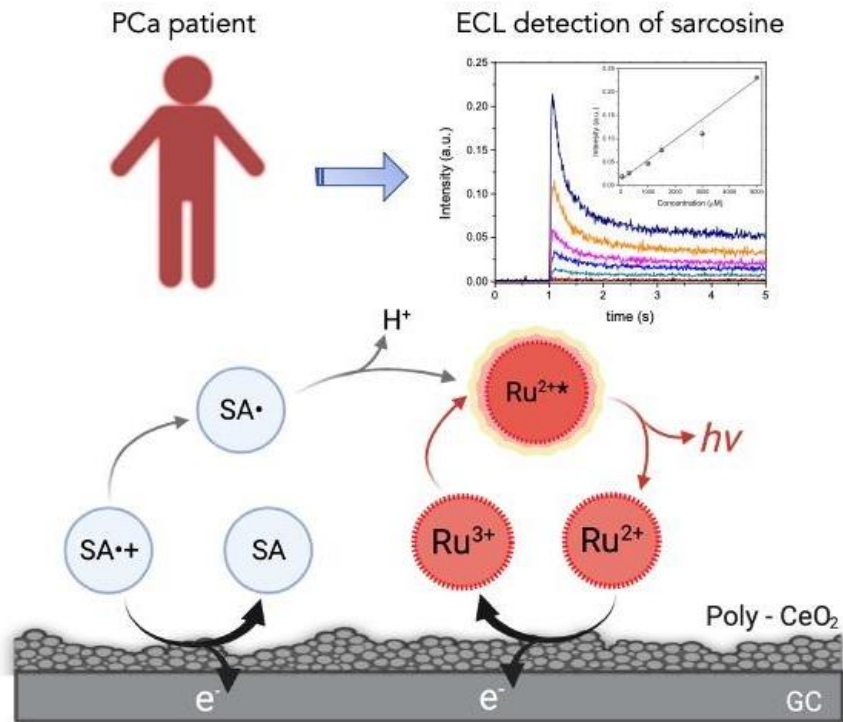
†† Current affiliation: University of Padova, Department of Chemical Sciences, F. Marzolo 1, 35131 Padova, Italy

* Corresponding author: alessandra.zanut@unipd.it, tsud@mbox.troja.mff.cuni.cz

Abstract

Prostate cancer (PCa) is widely spread in male population, especially over 65 years. Currently used medical methods of PCa diagnosis often lead to false-positive results thus new non-invasive methods for PCa detection, such as urine tests for cancer metabolites, are actively studied. Herein, nanostructured polycrystalline cerium oxide thin films (CeO_2/GC) prepared by magnetron sputtering on a glassy carbon substrate are tested for electrochemiluminescent (ECL) detection of sarcosine exploiting the oxidative-reduction mechanism using $\text{Ru}(\text{bpy})_3^{2+}$ as luminophore. Non-functionalized CeO_2/GC electrodes revealed a higher ECL signal stability compared to bare glassy carbon electrodes. Moreover, CeO_2/GC electrodes were successfully applied for rapid and sensitive detection of different sarcosine concentrations ranging from 50 to 5000 μM . These results open new possibilities for developing sensing platforms for sarcosine detection based on the CeO_2/GC working electrode via surface modification and functionalization, aiming to further investigate and improve their sensitivity and selectivity.

Graphical abstract:



Introduction

According to World Health Organization, prostate cancer (PCa) is the fourth most occurred cancer overall and the second in men part of the Earth human population.¹ Actual common screening procedures for PCa include prostate-specific antigen (PSA) blood testing and digital rectal examination (DRE).² However, PSA test may often lead to false-positive results as increased PSA concentration may occur also in benign prostatic hyperplasia or prostatitis. Moreover, DRE is an invasive and thus uncomfortable procedure with limited sensitivity and specificity.³ In order to reduce medical expenses and improve patients' quality of life, non-invasive diagnostic methods with high precision able to detect alternative and more accurate biomarkers for PCa diagnosis are needed.² In the last decades, non-invasive urine tests for cancer metabolites became a promising direction for PCa screening. In particular, the mono-peptide sarcosine (SA) has been recently studied as a potential biomarker molecule in PCa as its urine concentration increases during cancer progression.^{4,5} SA is an N-methyl glycine metabolite and it is involved in methylation processes, occurring during the progression of prostate cancer, and in the metabolism of amino acids.⁶

SA detection is nowadays performed through standard analytical methods, such as high-performance liquid-chromatography-mass-spectrometry (HPLC/MS) and gas-chromatography-mass-spectrometry (GC/MS), which ensure high sensitivity (in range of nM), but need expensive instrumentation and are time-consuming.⁷ In recent years, many research groups have focused on the design and development of biosensing platforms for rapid and easy SA detection, exploiting mainly fluorescent,⁸⁻¹⁰ luminescent,¹¹ and electrochemical^{12,13} schemes. However, determination of SA in urine samples is challenging because it requires high sensitivity toward SA molecules to discriminate it from the rest of the organic matrix. In this context, electrochemiluminescence (ECL) is an efficient analytical technique widely employed for the development of biosensing platforms for biomarkers detection.^{14,15} In the ECL luminescence process the excited states of luminophores are electrochemically generated in proximity of the electrode surface and then relax to a lower state emitting light.¹⁶⁻¹⁸ Thanks to its temporal and spatial control of light emission, ECL possess high sensitivity, wide dynamic range, and good stability even in complex matrixes.^{14,19,20} The most efficient ECL scheme for (bio)analytical applications involves the use of ruthenium(II)tris(2,2'-bipyridine) complex [Ru(bpy)₃]²⁺ as luminophore and tri-n-propylamine (TPrA) as co-reactant.^{17,21} In general,

tertiary amines are the most used because they provide the highest ECL intensity and have the lowest limit of detection (LOD).^{22,23}

Since SA is a secondary amine, it can also be potentially used in the ECL systems interacting with $\text{Ru}(\text{bpy})_3^{2+}$ and assisting in light emission. Indeed, it has been shown that SA acts as a good ECL co-reactant using a supramolecular approach to create a sensor for SA determination in urine samples, providing good selectivity in a concentration range used for medical diagnosis of PCa .²⁴

It is well known that the ECL efficiency is highly dependent on the electrochemical properties of electrode materials and many strategies have been proposed so far to make suitable electrodes and construct elaborate interfaces.²⁵ A good electrode material should be stable for extended use, while maintaining good kinetics for co-reactant oxidation. In this context, carbon-based materials, noble materials, and doped materials such as boron-doped diamond possess high chemical stability and high conductivity.^{26,27} However, almost all these materials are characterized by scarce porosity plus modification of such electrodes can strongly influence the ECL signal.²⁸

Among materials, cerium oxide is one of the most promising for biological sensing devices thanks to its catalytic activity, good biocompatibility, oxygen storage capacity, and electron transfer capability.²⁹⁻³¹

Recently, we reported the use of compact polycrystalline cerium oxide thin film deposited on a glassy carbon substrate (CeO_2/GC) as electrode for the electrochemical detection of hydrogen peroxide.³² Herein, we explore the capability of non-functionalized polycrystalline CeO_2/GC oxide film as a working electrode for ECL generation through an “oxidative-reduction” based scheme using SA as a co-reactant. The ECL detection of SA in aqueous solution (PBS, pH 7.2) was tested and compared with the one at bare glassy carbon electrodes, demonstrating advantages of nanostructured film over flat surfaces. Moreover, polycrystalline CeO_2/GC electrodes demonstrated the ability to distinguish different concentrations of SA in a range suitable for analytical purposes.

Experimental

Materials

All chemicals were of analytical grade and were purchased from Sigma-Aldrich (St. Louis, MO) unless otherwise indicated. Glassy carbon substrates (>99.9%, 1 mm thick, type 2) were

purchased from Alfa Aesar (USA). The phosphate buffer solution (PBS, pH 7.2) was prepared by dissolving 15.9 g of NaH_2PO_4 and 13.9 g of Na_2HPO_4 in 1000 mL of ultrapure water (18 M Ω cm). Total concentration of PBS was 200 mM. SA solutions (0 – 5000 μM) were prepared in PBS 200 mM (pH 7.2) and stored at 4°C.

Polycrystalline cerium oxide fabrication and characterization

Polycrystalline cerium oxide films were deposited onto glassy carbon substrate by non-reactive magnetron sputtering of a CeO_2 target (99.999%, Kurt J. Lesker) under Ar atmosphere (4×10^{-3} mbar). The magnetron sputtering was done using a RF power of 65 W, providing 1 nm min^{-1} growth rate of cerium oxide film. The thickness of the oxide films was estimated to be about 15 nm. The surface morphology and structure were characterized by a Bruker MultiMode 8 atomic force microscope (AFM) and a Tescan Mira 3 scanning electron microscope (SEM).

The chemical state of the CeO_2/GC electrode on the surface and throughout the whole thickness was examined by means of X-ray photoelectron spectroscopy (XPS) technique. The Ce 3d core level spectrum was recorded by XPS, providing information about oxidative state of Ce cations from about 7 nm deep layers of the film. The XPS experiment was conducted at the Materials Science beamline at Elettra synchrotron in Trieste, Italy.

Electrochemiluminescent detection of SA

ECL and electrochemical (EC) measurements were carried out with an AUTOLAB electrochemical station (Ecochemie, Mod. PGSTAT 30) in a home-made transparent three electrode cell with Ag/AgCl used as reference electrode, a platinum wire as counter electrode, and bare GC or CeO_2/GC as working electrode. During the EC measurements, the surface of CeO_2/GC electrodes in contact with the working solution was delimited by 0.5 cm^2 (circle of 0.8 cm diameter) o-ring. ECL signals were generated in chronoamperometry (CA) mode at potential of 1.4 V and measured with a photomultiplier tube (Acton PMT PD471, Hamamatsu R4220p) placed at a constant distance in front of the cell and inside a dark box. A voltage of 750 V was supplied to the PMT. The light/current/voltage curves were recorded by collecting the pre-amplified PMT output signal (using an ultralow-noise Acton research model 181) with the second input channel of an ADC module of the AUTOLAB instrument.

Results and discussion

The structure and morphology of polycrystalline CeO₂/GC electrodes were characterized using SEM and AFM microscopies (Fig. 1 a-b). From Figure 1a the surface mean roughness was estimated to be 1.2-1.3 nm and the grain size between 10-30 nm. The obtained XPS results showed a minor contribution of the Ce³⁺ cations within the measured thickness range in the as prepared samples (Fig. 1 c).

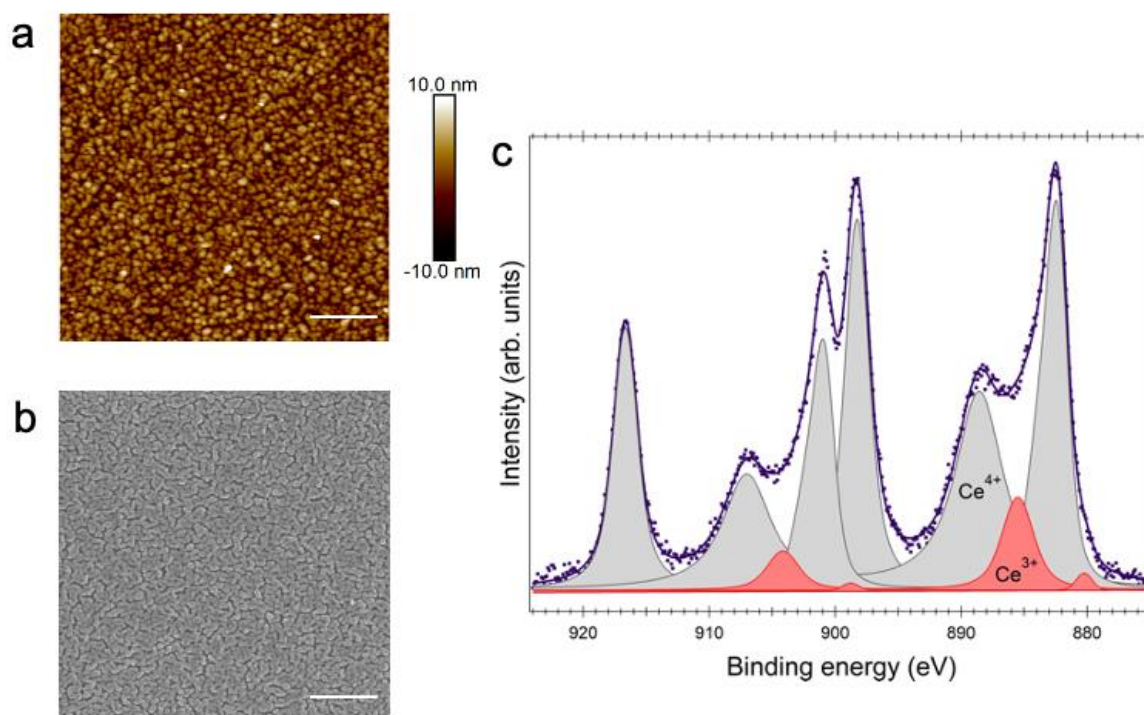


Figure 1. Polycrystalline CeO₂/GC electrodes characterization. a) AFM and b) SEM images of the 15 nm thick CeO₂/GC thin film. Scale bar 200 nm. c) Ce 3d XPS spectrum taken from the as prepared oxide film with photon energy 1486.7 eV. Violet dots and solid line correspond to the measured data and fit result, respectively. Gray peaks are attributed to the Ce⁴⁺ cations, and red peaks to the Ce³⁺ cations.

CV experiments to evaluate the electrocatalytic behaviour of CeO₂/GC and bare GC working electrodes, were conducted in pure 200 mM PBS solution and in the presence of 1 mM potassium ferricyanide (Fig. S1), confirming good conductivity of the CeO₂/GC substrate. Our group previously investigated the electrochemical properties of CeO₂/GC electrodes in the same conditions, observing good conductivity and larger electro-active area compared to bare GC.³² Moreover, it was proved that the higher total surface area of the electrode, modified by

polycrystalline ceria film, bring to an improved reaction rate for heterogeneous catalysts and the electrode transfer kinetics compared to bare GC electrodes.³²

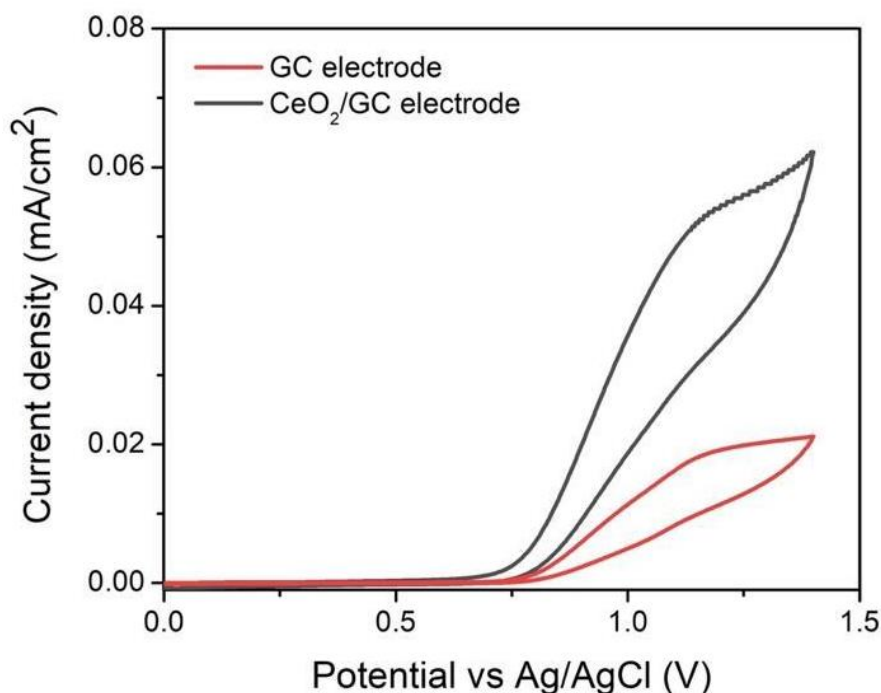


Figure 2. Cyclic voltammogram of GC (red) and CeO₂/GC (black) electrode recorded in 200 mM PBS (pH 7.2) containing 30 mM TPrA and 10 μ M Ru(bpy)₃²⁺. Scan rate of 100 mV/s, potential range from 0 to 1.4 V.

ECL efficiency of the CeO₂/GC electrode was first tested using Ru(bpy)₃²⁺ as luminophore and TPrA as co-reactant, applying a potential from 0 to 1.4 V vs. Ag/AgCl to ensure oxidation of both species.³³ The CV of CeO₂/GC and bare GC (Fig. 2) shows a higher current density of the Ru(bpy)₃²⁺ oxidation peak on the CeO₂/GC electrode compared to the bare GC electrode. Good conductivity of the CeO₂ thin film gives the ability to act as a conducting agent between the Ru(bpy)₃²⁺ and the GC substrate surface, facilitating electron-transfer in the ECL reaction. It was already demonstrated that surface modification of the working electrode surface by inorganic thin films or nanomaterials, polymer coatings, etc. can increase intensity of the ECL response.¹⁷ These results confirm the compatibility of CeO₂/GC electrodes with the Ru(bpy)₃²⁺/TPrA co-reactant ECL system.

As previously mentioned, tertiary amines are the most productive co-reactants in combination with $\text{Ru}(\text{bpy})_3^{2+}$ complex for the generation of ECL signals. However, secondary amines (where the amino group is directly bonded to two carbon atoms of any hybridization), such as SA, have the potential to provide sufficient effectivity for light generation and thus to be used as a co-reactant in ECL experiments. Indeed, SA shows an irreversible oxidation peak at 1 V satisfying the energy requirements for generating the excited state of the chromophore.²⁴ In this work, the ability of SA to act as a co-reactant for ECL generation has been tested according to the “oxidative-reduction” mechanism in 200 mM PBS using $10\mu\text{M}$ $\text{Ru}(\text{bpy})_3^{2+}$ on the CeO_2/GC electrode. Figure 3a shows the ECL signals generated at the CeO_2/GC electrode before and after the addition of 5mM SA as co-reactant.

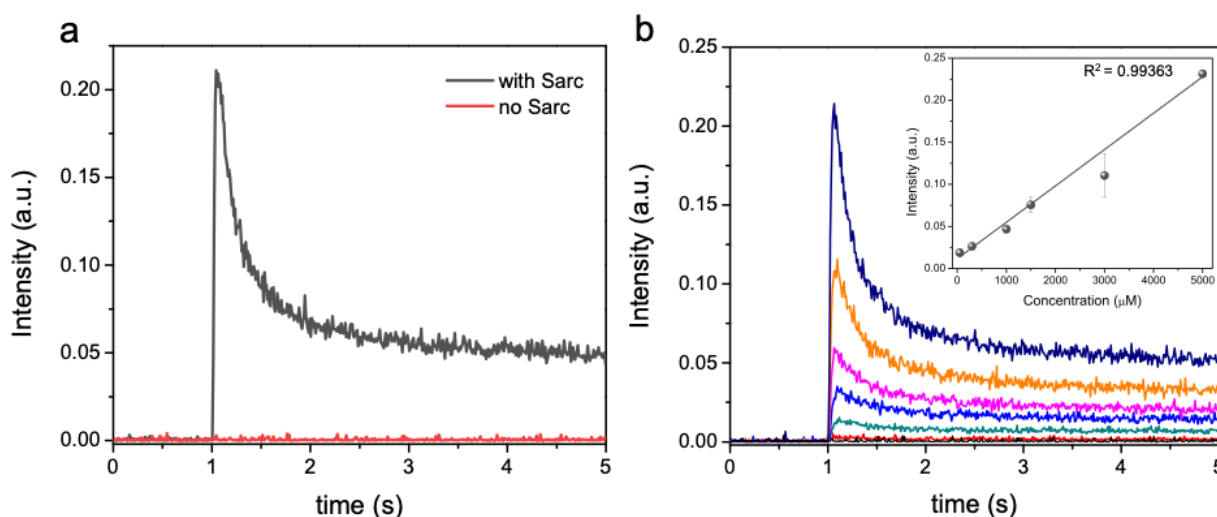
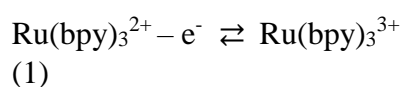


Figure 3. ECL generation using sarcosine as a co-reactant. ECL signal recorded in chronoamperometry mode using CeO_2/GC electrodes measured at 1.4 V in 200 mM PBS (pH 7.2) containing $10\mu\text{M}$ $\text{Ru}(\text{bpy})_3^{2+}$ **a)** with (black) and without (red) 5 mM SA as co-reactant and **b)** after addition of 50 (red), 300 (green), 1000 (blue), 1500 (pink), 3000 (orange), and 5000 μM (dark blue) of sarcosine. Inset shows the max ECL value as a function of SA concentration. The error is represented as the standard deviation ($n=3$) and is equal to 0.00065.

The proposed mechanism for ECL generation involves the reaction between SA and $\text{Ru}(\text{bpy})_3^{2+}$ to form the excited state $\text{Ru}(\text{bpy})_3^{2+*}$, which relaxes to the ground state generating light. The reaction can be described as follow:





To evaluate the sensing ability of the CeO₂/GC electrode to different concentrations of SA, the experiment was conducted in 10 μM Ru(bpy)₃²⁺ PBS solution with stepwise addition of SA (50-5000 μM) (Fig. 3 b). Concentrations of sarcosine were chosen to investigate a wide range of values useful for analytical applications.

Inset of Fig. 3b shows the maximum ECL intensity profile at different concentrations of sarcosine in the working solution (hereinafter called calibration curve) taken with CeO₂/GC electrodes. For comparison and estimation of the CeO₂/GC electrode efficiency, the same measurements were repeated with the bare GC electrode (Fig. S2). The results reveal a direct dose-response correlation between the sarcosine concentration and the ECL signal intensity.

The LOD parameter was calculated from the obtained calibration lines (Fig. S3) using the following formula:

$$LOD = \frac{(K \times S_b)}{m},$$

where *K* equals 3 (confidence level 98.3%), *S_b* is the standard deviation of the blank solution, and *m* is the calibration sensitivity (slope of the calibration curve). Under the optimized experimental conditions, the LOD was found to be 45.9 and 63.10 μM for CeO₂/GC and bare GC electrodes, respectively. It confirms that GC electrodes modified with polycrystalline cerium oxide thin film is able to detect lower concentrations of SA.

It is worth noting that on the CeO₂/GC electrode the intensity stability, measured as the total area under the ECL peak, is higher than on the bare GC (Fig. 4), suggesting that the polycrystalline CeO₂/GC electrode possess better electroanalytical performance and higher electrochemical stability.

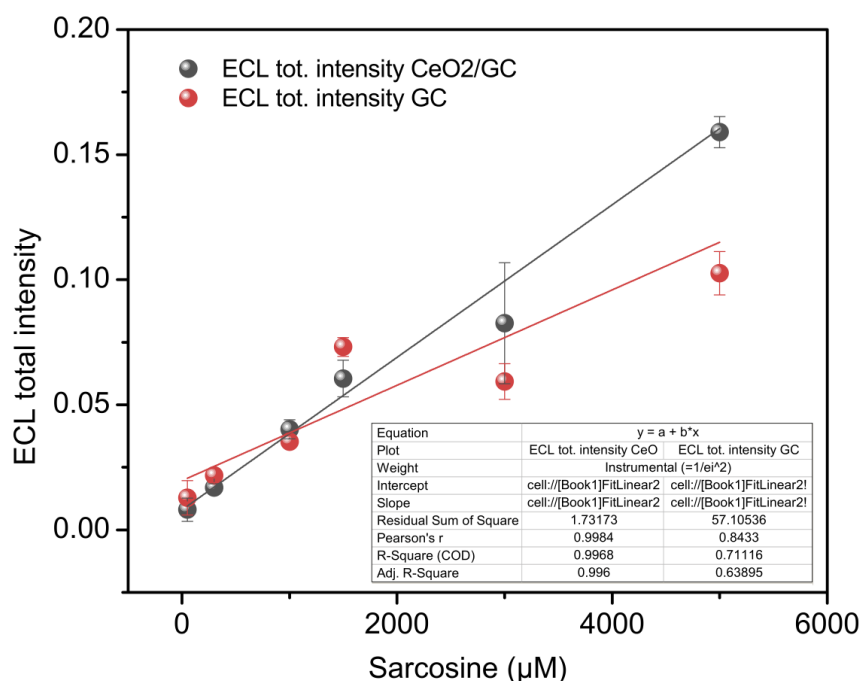


Figure 4. The ECL total intensity as a function of sarcosine concentration is measured on bare GC and CeO₂/GC electrodes in 200 mM PBS (pH 7.2) containing 10 µM Ru(bpy)₃²⁺. The error is represented as standard deviation (n=3).

This represents another advantage of CeO₂/GC electrodes over bare GC ones, on which the oxygen-containing surface species, which decrease the efficiency of coreactant oxidation are usually formed.^{20,34,35}

To evaluate the effect of possible interferences on ECL signals, we tested three representative Sarcosine concentrations (0, 50, 3000 µM) using CeO₂/GC electrodes in a solution of 10 µM Ru(bpy)₃²⁺ containing BSA 10 mg/ml which mimicking the concentration present in human urine (Fig. S4a). Obtained results confirm the ability of CeO₂/GC electrodes to discriminate between different SA concentrations even in presence of BSA 10mg/ml. Moreover, signals are reproducible as confirmed by the calculated RSD% which is ≤ 5% for both concentrations (Fig. S4b).

Conclusions

Polycrystalline cerium oxide thin films prepared by magnetron sputtering on the glassy carbon substrate were studied as an electrode for ECL detection of sarcosine, using SA as a co-reactant

in oxidative-reduction mechanism with $\text{Ru}(\text{bpy})_3^{2+}$ as luminophore. Obtained results revealed that modification of the bare glassy carbon electrode by polycrystalline cerium oxide layer increases the electrode's electroactive area and enhances its catalytic properties. Moreover, the comparative study with bare glassy carbon electrodes revealed a higher ECL stability and thus higher signal stability on the CeO_2/GC electrode due to its higher surface area and porosity. The CeO_2/GC electrode was successfully applied for rapid and sensitive detection of different SA concentrations (50 – 5000 μM). CeO_2/GC was shown to be a more efficient electrode material for ECL applications compared to the commercial GC electrode. LOD of CeO_2/GC electrode was estimated to be 45.9 μM which makes it suitable for SA detection in the concentration range used in clinical assays for PCa diagnostics. Preliminary results demonstrated the ability of CeO_2/GC electrodes to detect lower concentration of SA also in presence of BSA 10 mg/ml with a good reproducibility ($\text{RSD} \leq 5\%$). Further investigations are ongoing in our laboratories to better assess the analytical strength of this system in complex matrices (i.e. urine). These results open new possibilities for the development of sensors for SA detection based on the CeO_2/GC working electrode via surface modification and functionalization, with the aim to further investigate and improve their sensitivity and selectivity.

Acknowledgement

The CERIC-ERIC consortium and the Czech Ministry of Education, Youth and Sports (project LM2018116) are acknowledged for financial support. This work is supported by the Italian Ministero dell'Istruzione, Università e Ricerca (PRIN-2017FJCPEX) and University of Bologna and Fondazione CarisBo (project #18668).

References

- (1) World Cancer Research Fund International <https://www.wcrf.org/cancer-trends/prostate-cancer-statistics/>.
- (2) Chan, K. M.; Gleadle, J. M.; O'Callaghan, M.; Vasilev, K.; MacGregor, M. Prostate Cancer Detection: A Systematic Review of Urinary Biosensors. *Prostate Cancer Prostatic Dis.* **2022**, No. August, 1–8. <https://doi.org/10.1038/s41391-021-00480-8>.
- (3) Klotz, L. Prostate Cancer Overdiagnosis and Overtreatment. *Curr. Opin. Endocrinol. Diabetes Obes.* **2013**, *20* (3), 204–209.

<https://doi.org/10.1097/MED.0b013e328360332a>.

- (4) Sreekumar, A.; Poisson, L. M.; Rajendiran, T. M.; Khan, A. P.; Cao, Q.; Yu, J.; Laxman, B.; Mehra, R.; Lonigro, R. J.; Li, Y.; Nyati, M. K.; Ahsan, A.; Kalyana-Sundaram, S.; Han, B.; Cao, X.; Byun, J.; Omenn, G. S.; Ghosh, D.; Pennathur, S.; Alexander, D. C.; Berger, A.; Shuster, J. R.; Wei, J. T.; Varambally, S.; Beecher, C.; Chinnaiyan, A. M. Metabolomic Profiles Delineate Potential Role for Sarcosine in Prostate Cancer Progression. *Nature* **2009**, *457* (7231), 910–914. <https://doi.org/10.1038/nature07762>.
- (5) Couzin, J. BIOMARKERS: Metabolite in Urine May Point To High-Risk Prostate Cancer. *Science*. **2009**, *323* (5916), 865a-865a. <https://doi.org/10.1126/science.323.5916.865a>.
- (6) Dereziński, P.; Klupczynska, A.; Sawicki, W.; Pałka, J. A.; Kokot, Z. J. Amino Acid Profiles of Serum and Urine in Search for Prostate Cancer Biomarkers: A Pilot Study. *Int. J. Med. Sci.* **2017**, *14* (1), 1–12. <https://doi.org/10.7150/ijms.15783>.
- (7) Jiang, Y.; Cheng, X.; Wang, C.; Ma, Y. Quantitative Determination of Sarcosine and Related Compounds in Urinary Samples by Liquid Chromatography with Tandem Mass Spectrometry. *Anal. Chem.* **2010**, *82* (21), 9022–9027. <https://doi.org/10.1021/ac1019914>.
- (8) Biavardi, E.; Tudisco, C.; Maffei, F.; Motta, A.; Massera, C.; Condorelli, G. G.; Dalcanale, E. Exclusive Recognition of Sarcosine in Water and Urine by a Cavitand-Functionalized Silicon Surface. *Proc. Natl. Acad. Sci.* **2012**, *109* (7), 2263–2268. <https://doi.org/10.1073/pnas.1112264109>.
- (9) Luo, Y.; Wang, J.; Yang, L.; Gao, T.; Pei, R. Sensors and Actuators B : Chemical In Vitro Selection of DNA Aptamers for the Development of Fluorescent Aptasensor for Sarcosine Detection. *Sensors Actuators B. Chem.* **2018**, *276* (April), 128–135. <https://doi.org/10.1016/j.snb.2018.08.105>.
- (10) Li, W.; Li, T.; Chen, S.; Deng, D.; Ji, Y.; Li, R. Sensors and Actuators : B . Chemical Nanozyme-Mediated Cascade Reaction System for Ratiometric Fluorescence Detection of Sarcosine. *Sensors Actuators B. Chem.* **2022**, *355* (October 2021), 131341. <https://doi.org/10.1016/j.snb.2021.131341>.
- (11) Zhao, L.; Yang, J.; Gong, M.; Li, K.; Gu, J. Specific Screening of Prostate Cancer Individuals Using an Enzyme-Assisted Substrate Sensing Platform Based on Hierarchical MOFs with Tunable Mesopore Size. **2021**.
- (12) Hu, J.; Wei, W.; Ke, S.; Zeng, X.; Lin, P. Electrochimica Acta A Novel and Sensitive Sarcosine Biosensor Based on Organic Electrochemical Transistor. *Electrochim. Acta* **2019**, *307*, 100–106. <https://doi.org/10.1016/j.electacta.2019.03.180>.
- (13) Liu, T.; Fu, B.; Chen, J.; Li, K. An Electrochemical Sarcosine Sensor Based on Biomimetic Recognition. **2019**.
- (14) Faatz, E.; Finke, A.; Josel, H. P.; Prencipe, G.; Quint, S.; Windfuhr, M. Chapter 15: Automated Immunoassays for the Detection of Biomarkers in Body Fluids. *RSC Detect. Sci.* **2020**, *2020-January* (15), 443–470. <https://doi.org/10.1039/9781788015776-00443>.

- (15) Zanut, A.; Fiorani, A.; Canola, S.; Saito, T.; Ziebart, N.; Rapino, S.; Rebecani, S.; Barbon, A.; Irie, T.; Josel, H.-P.; Negri, F.; Marcaccio, M.; Windfuhr, M.; Imai, K.; Valenti, G.; Paolucci, F. Insights into the Mechanism of Coreactant Electrochemiluminescence Facilitating Enhanced Bioanalytical Performance. *Nat. Commun.* **2020**, *11* (1), 2668. <https://doi.org/10.1038/s41467-020-16476-2>.
- (16) Bouffier, L.; Sojic, N. Chapter 1: Introduction and Overview of Electrogenerated Chemiluminescence. *RSC Detect. Sci.* **2020**, *2020-Janua* (15), 1–28. <https://doi.org/10.1039/9781788015776-00001>.
- (17) Forster, J. R.; Bertocello, P.; Keyes E. Tia. Electrogenerated Chemiluminescence. *Annu. Rev. Anal. Chem.* **2009**, *2*, 359–385. <https://doi.org/10.1146/annurev-anchem-060908-155305>.
- (18) Richter, M. M. Electrochemiluminescence (ECL). *Chem. Rev.* **2004**, *104*, 3003–3036. <https://doi.org/10.1021/cr020373d>.
- (19) Zhang, J.; Arbault, S.; Sojic, N.; Jiang, D. Electrochemiluminescence Imaging for Bioanalysis. *Annu. Rev. Anal. Chem.* **2019**, *12* (1), 275–295. <https://doi.org/10.1146/annurev-anchem-061318-115226>.
- (20) Rebecani, S.; Zanut, A.; Santo, C. I.; Valenti, G.; Paolucci, F. A Guide Inside Electrochemiluminescent Microscopy Mechanisms for Analytical Performance Improvement. *Anal. Chem.* **2022**, *94* (1), 336–348. <https://doi.org/10.1021/acs.analchem.1c05065>.
- (21) Kerr, E.; Doeven, E. H.; Barbante, G. J.; Hogan, C. F.; Bower, D. J.; Donnelly, P. S.; Connell, T. U.; Francis, P. S. Annihilation Electrogenerated Chemiluminescence of Mixed Metal Chelates in Solution: Modulating Emission Colour by Manipulating the Energetics. *Chem. Sci.* **2015**, *6* (1), 472–479. <https://doi.org/10.1039/C4SC02697G>.
- (22) Liu, X.; Shi, L.; Niu, W.; Li, H.; Xu, G. Environmentally Friendly and Highly Sensitive Ruthenium(II) Tris(2,2'-Bipyridyl) Electrochemiluminescent System Using 2-(Dibutylamino)Ethanol as Co-Reactant. *Angew. Chem. Int. Ed.* **2007**, *46* (3), 421–424. <https://doi.org/10.1002/anie.200603491>.
- (23) Yuan, Y.; Han, S.; Hu, L.; Parveen, S.; Xu, G. Coreactants of Tris(2,2'-Bipyridyl)Ruthenium(II) Electrogenerated Chemiluminescence. *Electrochim. Acta* **2012**, *82*, 484–492. <https://doi.org/10.1016/j.electacta.2012.03.156>.
- (24) Valenti, G.; Rampazzo, E.; Biavardi, E.; Villani, E.; Fracasso, G.; Marcaccio, M.; Bertani, F.; Ramarli, D.; Dalcanale, E.; Paolucci, F.; Prodi, L. An Electrochemiluminescence-Supramolecular Approach to Sarcosine Detection for Early Diagnosis of Prostate Cancer. *Faraday Discuss.* **2015**, *185*, 299–309. <https://doi.org/10.1039/C5FD00096C>.
- (25) Valenti, G.; Fiorani, A.; Li, H.; Sojic, N.; Paolucci, F. *Essential Role of Electrode Materials in Electrochemiluminescence Applications*; 2016. <https://doi.org/10.1002/celc.201600602>.
- (26) Fiorani, A.; Merino, J. P.; Zanut, A.; Criado, A.; Valenti, G.; Prato, M.; Paolucci, F. Advanced Carbon Nanomaterials for Electrochemiluminescent Biosensor Applications. *Current Opinion in Electrochemistry*. 2019.

<https://doi.org/10.1016/j.coelec.2019.04.018>.

- (27) Sakanoue, K.; Fiorani, A.; Santo, C. I.; Irkham; Valenti, G.; Paolucci, F.; Einaga, Y. Boron-Doped Diamond Electrode Outperforms the State-of-the-Art Electrochemiluminescence from Microbeads Immunoassay. *ACS Sensors* **2022**, *7* (4), 1145–1155. <https://doi.org/10.1021/acssensors.2c00156>.
- (28) Nikolaou, P.; Valenti, G.; Paolucci, F. Nano-Structured Materials for the Electrochemiluminescence Signal Enhancement. *Electrochim. Acta* **2021**, *388*, 138586. <https://doi.org/10.1016/j.electacta.2021.138586>.
- (29) Karimi, A.; Husain, S. W.; Hosseini, M.; Azar, P. A.; Ganjali, M. R. Rapid and Sensitive Detection of Hydrogen Peroxide in Milk by Enzyme-Free Electrochemiluminescence Sensor Based on a Polypyrrole-Cerium Oxide Nanocomposite. *Sensors Actuators, B Chem.* **2018**, *271* (January), 90–96. <https://doi.org/10.1016/j.snb.2018.05.066>.
- (30) Valenti, G.; Melchionna, M.; Montini, T.; Boni, A.; Nasi, L.; Fonda, E.; Criado, A.; Zitolo, A.; Voci, S.; Bertoni, G.; Bonchio, M.; Fornasiero, P.; Paolucci, F.; Prato, M. Water-Mediated ElectroHydrogenation of CO₂ at Near-Equilibrium Potential by Carbon Nanotubes/Cerium Dioxide Nanohybrids. *ACS Appl. Energy Mater.* **2020**, *3* (9), 8509–8518. <https://doi.org/10.1021/acsaem.0c01145>.
- (31) Alizadeh, N.; Salimi, A.; Sham, T.-K.; Bazylewski, P.; Fanchini, G. Intrinsic Enzyme-like Activities of Cerium Oxide Nanocomposite and Its Application for Extracellular H₂O₂ Detection Using an Electrochemical Microfluidic Device. *ACS Omega* **2020**, *5* (21), 11883–11894. <https://doi.org/10.1021/acsomega.9b03252>.
- (32) Kosto, Y.; Zanut, A.; Franchi, S.; Yakovlev, Y.; Khalakhan, I.; Matolín, V.; Prince, K. C.; Valenti, G.; Paolucci, F.; Tsud, N. Electrochemical Activity of the Polycrystalline Cerium Oxide Films for Hydrogen Peroxide Detection. *Appl. Surf. Sci.* **2019**, *488*, 351–359. <https://doi.org/10.1016/j.apsusc.2019.05.205>.
- (33) Miao, W.; Choi, J.-P.; Bard, A. J. Electrogenenerated Chemiluminescence 69: The Tris(2,2'-Bipyridine)Ruthenium(II), (Ru(Bpy)₃²⁺)/Tri-n-Propylamine (TPrA) System Revisited A New Route Involving TPrA^{•+} Cation Radicals. *J. Am. Chem. Soc.* **2002**, *124* (48), 14478–14485. <https://doi.org/10.1021/ja027532v>.
- (34) Dutta, P.; Han, D.; Goudeau, B.; Jiang, D.; Fang, D.; Sojic, N. Reactivity Mapping of Luminescence in Space: Insights into Heterogeneous Electrochemiluminescence Bioassays. *Biosens. Bioelectron.* **2020**, *165* (May), 112372. <https://doi.org/10.1016/j.bios.2020.112372>.
- (35) Han, D.; Goudeau, B.; Jiang, D.; Fang, D.; Sojic, N. Electrochemiluminescence Microscopy of Cells: Essential Role of Surface Regeneration. *Anal. Chem.* **2021**, *93* (3), 1652–1657. <https://doi.org/10.1021/acs.analchem.0c05123>.