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Identification of the most effective serovars to be included in the MAT antigen panel to optimize the serodiagnosis of *Leptospira* infection in Northern Italy

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1    **Short Communication / Brief Report**

2

3    **Identification of the most effective serovars to be included in the MAT antigen panel to**  
4    **optimize the serodiagnosis of *Leptospira* infection in Northern Italy**

5

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23   **Abstract**

24   The microscopic agglutination test (MAT) assay is adopted as a world reference test for the  
25   serodiagnosis of leptospirosis in humans and animals. The incapability of MAT to detect antibodies  
26   against leptospiral serogroups not included in the assay antigen panel is one of the main limitations  
27   of this test and serodiagnostic antigens should be periodically updated with locally circulating  
28   serovars in order to optimise its performance. The aim of this study was to determine the need to  
29   implement the antigen panel currently adopted in Northern Italy for the diagnosis of *Leptospira*  
30   infection in dogs. For this purpose, a group of 288 dogs with and without clinical signs potentially  
31   associated with *Leptospira* infection or increased C-reactive protein (CRP) serum concentration,  
32   sampled in 2013-2016 in Northern Italy, were tested by MAT comparing the results obtained with a  
33   nine antigens panel (Australis-Bratislava, Ballum-Ballum, Canicola-Canicola, Grippotyphosa-  
34   Grippotyphosa, Icterohaemorrhagiae-Copenhageni, Icterohaemorrhagiae-Icterohaemorrhagiae,  
35   Sejroe-Hardjo, Pomona-Pomona and Tarassovi-Tarassovi serovars) routinely adopted and a panel  
36   expanded to 27 antigens. In general, the antigen panel currently adopted in Northern Italy for the  
37   routine MAT assay resulted adequate for the diagnosis of *Leptospira* infection in dogs. The main  
38   exception concerns the Sejroe serogroup, with the Saxkoebing and Sejroe serovars that were more  
39   effective than Hardjo for diagnosis in dogs and whose inclusion in the antigen panel is  
40   recommended. Among other antigens evaluated in this study, Cynopteri serovar was detected with  
41   high frequency but its pathogenic role in dogs and as public health threat deserve further  
42   investigation.

43

44   *Keywords:* antigen, diagnosis, dog, *Leptospira*, microscopic agglutination test, serology

45     **Introduction**

46     Leptospirosis is a worldwide zoonosis affecting numerous wild and domestic mammalian species  
47     (Bharti et al., 2003), sustained by pathogenic Gram-negative and highly motile spirochete bacteria  
48     of the genus *Leptospira*. Some host species act as reservoirs, representing the natural source of  
49     infection and of environmental contamination (Gomard et al., 2021; Levett, 2001). Reservoir hosts  
50     are persistently infected, normally with no clinical signs, and shed bacteria through their urine even  
51     lifelong (Schuller et al., 2015). Differently, incidental hosts can develop acute and severe disease  
52     (Levett, 2001; Schuller et al., 2015). The dog is usually an incidental host, showing a wide range of  
53     clinical manifestations, from subclinical to severe (Sykes et al., 2011; Schuller et al., 2015), but can  
54     represent an important sentinel species as well as a potential reservoir host for some serovars  
55     (Balboni et al., 2022).

56     *Leptospira* spp. are classified in hundreds of serovars due to variable epitopes in the  
57     lipopolysaccharide (LPS) structure; furthermore, different serovars are grouped into serogroups  
58     (Faine et al., 1999; Ko et al., 2009). Most of the known serovars have close relationships with  
59     specific reservoir hosts and the epidemiology of canine leptospirosis can vary by geographic area  
60     and over time, in relation to the spread of maintenance hosts and vaccination (Bharti et al., 2003;  
61     Schuller et al., 2015; Sykes et al., 2010). In Europe, dogs are apparently more exposed to  
62     Icterohaemorrhagiae, Grippotyphosa, Australis, Canicola and Sejroe serogroups (Ellis, 2010), but  
63     many other serogroups such as Autumnalis, Ballum, Bataviae, Cynopteri, Pomona, Pyrogenes and  
64     Tarassovi were reported worldwide in dogs (Costa et al., 2022; Pinto et al., 2017; Sykes et al.,  
65     2011), also in association with clinical manifestations, and reservoirs of some serovars still remain  
66     unknown.

67     The vaccination evokes a serovar-specific and partially serogroup-specific immune protection  
68     (Bouvet et al., 2020; Klaasen et al., 2022). To date, trivalent or tetravalent vaccines containing  
69     antigens from up to four different serovars belonging to Canicola, Icterohaemorrhagiae, Australis,

70 and Grippotyphosa serogroups are available in Europe and Italy (Ellis, 2010; Klaasen et al., 2014;  
71 Schuller et al., 2015).

72 Diagnosis of leptospirosis should be multifaceted and not rely on any one single test (Sykes et al.,  
73 2022). To achieve a definitive diagnosis many factors should be considered, such as potential  
74 exposure, clinical presentation and laboratory values, and the results of multiple direct or indirect  
75 diagnostic approaches must be evaluated (Sykes et al., 2022). The microscopic agglutination test  
76 (MAT) involves incubation of serial dilutions of patient sera with a panel of live leptospiral  
77 organisms as antigens and reading the resulting agglutination under a darkfield microscope (Sykes  
78 et al., 2022). MAT is a serogroup rather than a serovar-specific test (Levett, 2001); nevertheless,  
79 different responses are detectable between serovars belonging to the same serogroup. Although  
80 MAT is subjected to a number of limitations (Barr et al., 2005; Kohn et al., 2010; Martin et al.,  
81 2014; Schuller et al., 2015; Sykes et al., 2010; Sykes et al., 2022), it is still being adopted as a world  
82 reference test for the serodiagnosis of leptospirosis in humans and animals (World Organisation for  
83 Animal Health, 2022). The incapability of MAT to detect antibodies against leptospiral serogroups  
84 not included in the assay antigen panel is one of the main limitations of this test. For this reason,  
85 live antigen panels should include locally circulating serovars and serodiagnostic antigens should be  
86 periodically updated as new strains emerge in order to optimise its performance (Sykes et al., 2022).

87 In this study, a group of dogs showing clinical signs potentially associated with *Leptospira* infection  
88 or increased C-reactive protein (CRP) serum concentration and a group of apparently healthy dogs  
89 were tested by MAT comparing the results obtained with a nine antigens panel routinely used in  
90 Northern Italy and a panel expanded to 27 antigens, in order to determine the need to implement the  
91 antigen panel currently adopted in Northern Italy for the diagnosis of *Leptospira* infection in dogs.

92

93 **Materials and Methods**

94 ***Study design, population and sampling***

95 This retrospective study was carried out at the Istituto Zooprofilattico Sperimentale delle Venezie,  
96 (IZSVE, Legnaro, Padova, Italy) and the Italian Reference Centre for Animal Leptospirosis (Istituto  
97 Zooprofilattico Sperimentale della Lombardia e dell'Emilia Romagna, IZSLER, Brescia, Italy). In  
98 the study, serum samples from dogs were selected and sent to the IZSVE laboratory for diagnostic  
99 purposes, and the implementation of the research study was supported by funding from the RC IZS  
100 VE 16/12, from August 2013 to July 2016. The study population was composed by owned dogs  
101 showing clinical signs potentially associated with *Leptospira* infection or increased CRP serum  
102 concentration (Ceron et al., 2005; Schuller et al., 2015) sampled by veterinary practitioners,  
103 apparently healthy kennel dogs undergoing neutering surgery sampled to perform pre-operative  
104 profile tests and apparently healthy blood donor dogs sampled to perform pre-donation screening  
105 tests. No dogs were sampled exclusively for the purposes of this study. Only samples taken for  
106 diagnostic purposes following owner or legal manager of the kennel consent were used.  
107 Blood sampling was carried out by venepuncture and serum samples were stored at -20 °C until  
108 analysis. Signalment data and vaccination status were retrieved from medical records. Vaccination  
109 status was compared to international guidelines for the vaccination of dogs (Day et al., 2016).  
110 All dogs included in the study were tested by MAT both with a nine antigens panel routinely  
111 adopted in Northern Italy, in line with the eight antigen panel fixed at national level by the National  
112 Reference Laboratory for Leptospirosis to which the Icterohaemorrhagiae serovar was added  
113 (Tagliabue, 2016), and a panel expanded to 27 antigens selected on the basis of epidemiological  
114 data from Europe and Mediterranean basin (Arent et al., 2013; Benkirane et al., 2016; Goris et al.,  
115 2013; Mayer-Scholl et al., 2013). The results obtained with the two panels were compared to detect  
116 seroreactions against antigens not included in the routine test.

117

#### 118 ***Microscopic agglutination test (MAT)***

119 Dog serum samples were tested for antibodies against *Leptospira* using the MAT following the  
120 World Organisation for Animal Health (WOAH) method (Chap 3.1.12) (World Organisation for

Animal Health, 2022). In the assay routinely adopted in Northern Italy, the antigen panel included eight serogroups and nine serovars (**Table 1**). In the expanded assay, the antigen panel included 20 serogroups and 27 serovars (**Table 1**), including those of the routine assay. Serum samples were pre-tested at the final dilution of 1:100. Serum with 50% agglutination were retested to determine an endpoint using dilutions of serum beginning at 1:100 through to 1:6400. Serum samples with the widely accepted minimum significant titre of 1:100 (reciprocal of the final dilution of serum with 50% agglutination) were assessed positive. Positive antibody titres  $\geq 1:800$  against at least one *Leptospira* serogroup were recognised as of potential infectious origin, excluding most vaccine responses.

Addition to the antigen panel has been suggested for serovars that showed a titre  $\geq 1:100$  in dogs tested negative by the routine MAT assay or a titre equal to or higher than that obtained for serovars used routinely in Northern Italy.

#### ***Statistical analysis***

The data were evaluated using standard descriptive statistics and reported as median and range. Categorical data were analysed using the Chi-squared test. Statistical significance was set at  $P < 0.05$ . Not available data was excluded to statistical analysis. Statistical analysis was carried out using commercially available software (MedCalc Statistical Software version 16.8.4).

### **Results**

#### ***Study population***

During the study period, 288 dogs were included in the study: 217/288 (75.3%) were owned dogs showing clinical signs potentially associated with *Leptospira* infection or increased CRP serum concentration and 71/288 (24.7%) dogs were apparently healthy kennel dogs (N: 20) or blood donor dogs (N: 51). Among the study population, 108/288 (37.5%) dogs were males and 94/288 (32.6%) were females, for the remaining 86/288 (29.9%) dogs this data was not available. The median age



147 of dogs (N: 155) was five years (range 1 month – 16 years), whereas, 133/288 (46.2%) were  
148 purebred, 60/288 (20.8%) were mixed breed and for the remaining 95/288 (33%) this data was not  
149 available. Ninety-four out of 288 (32.6%) dogs were regularly vaccinated against leptospirosis with  
150 bivalent (N: 84; Canicola and Icterohaemorrhagiae serogroups) or tetravalent (N: 10; Canicola,  
151 Icterohaemorrhagiae, Australis and Grippotyphosa serogroups) vaccines, 59/288 (20.5%) were not  
152 or not regularly vaccinated and for the remaining 135/288 (46.9%) this data was not available  
153 (**Table 2**).

154

### 155 ***Routine and expanded MAT assays results***

156 The sera of 128/288 (44.4%) dogs included in the study were tested positive with a cut-off  $\geq 1:100$   
157 to antibodies against at least one of the pathogenic *Leptospira* serovars included in the nine antigens  
158 panel of the routine MAT assay (**Table 3**). Adopting a cut-off  $\geq 1:800$ , the number of dogs tested  
159 positive was 46/288 (16%).

160 The sera of 141/288 (49%) dogs included in the study were tested positive with a cut-off  $\geq 1:100$  to  
161 antibodies against at least one of the *Leptospira* serovars included in the 27 antigens panel of the  
162 expanded MAT assay (**Table 3**). Adopting a cut-off  $\geq 1:800$ , the number of dogs tested positive was  
163 52/288 (18.1%).

164 The majority of the seropositive dogs reported multiple titres against different serovars and  
165 serogroups. The frequency of detection of the different serovars is summarized in **Table 1**.

166 Considering a cut-off  $\geq 1:100$ , five of the 10 most frequently detected serovars were not included in  
167 the routine MAT assay (Australis-Jalna, Autumnalis-Autumnalis, Cynopteri-Cynopteri, Lyme-  
168 Lyme and Pyrogenes-Pyrogenes), whereas, considering a cut-off  $\geq 1:800$ , four of the 10 most  
169 frequently detected serovars were not included in the routine MAT assay (Australis-Jalna,  
170 Autumnalis-Autumnalis, Cynopteri-Cynopteri, and Pomona-Mozdok). The Australis-Bratislava,  
171 Canicola-Canicola, Grippotyphosa-Grippotyphosa, Icterohaemorrhagiae-Copenhageni,  
172 Icterohaemorrhagiae- Icterohaemorrhagiae and Pomona-Pomona serovars, included in the routine

173 MAT assay, were among the most frequently detected using both cut-offs **Table 1**. Antibodies  
 174 against Celledoni-Celledoni, Mini-Mini, Shermani-Shermani and Tarassovi-Tarassovi serovars  
 175 were not found in this study, whereas Bataviae-Bataviae and Hebdomadis-Hebdomadis serovars  
 176 were sporadically detected (in one and two dogs, respectively) with low titres ( $< 1:800$ , **Table 1**).  
 177 Using the MAT assay with the expanded 27 antigens panel, 13/288 (4.5%) dogs were tested  
 178 seropositive only against at least one of the 18 *Leptospira* antigens not included in the routine MAT  
 179 assay (**Table 3**). Of these dogs, 11 showed antibody titre values  $< 1:800$  against Cynopteri-  
 180 Cynopteri (N: 1), Hurstbridge-Hurstbridge (N: 3), Lyme-Lyme (N: 5) and Pyrogenes-Pyrogenes (N:  
 181 1) serovars, and two showed antibody titre value  $\geq 1:800$  against Sejroe-Saxkoebing serovar (1:800  
 182 and 1:3200, respectively). The dog tested positive to Sejroe-Saxkoebing serovar with titre 1:3200  
 183 also showed seropositivity against Lyme-Lyme (1:200) and Sejroe-Sejroe (1:100) serovars. In  
 184 addition, four dogs had MAT titres  $\geq 1:800$  against at least one of the 18 *Leptospira* antigens not  
 185 included in the routine assay (Cynopteri-Cynopteri N: 1, Lyme-Lyme N: 1 and Sejroe-Sejroe N: 2)  
 186 and titres  $< 1:800$  against antigens included in the routine assay, for a total of 6/288 (2.1%) dogs  
 187 detected positive only with the expanded MAT assay using a cut-off  $\geq 1:800$  (**Table 3**).  
 188 Furthermore, other 35/288 (12.1%) dogs had prevalent MAT titre against at least one of the 18  
 189 *Leptospira* antigens not included in the routine MAT assay, with values higher (N: 14) or equal (N:  
 190 21) to those obtained against serovars included in the routine MAT assay: Australis-Jalna (N: 7),  
 191 Cynopteri-Cynopteri (N: 6), Hurstbridge-Hurstbridge (N: 4), Javanica-Javanica (N: 1), Lyme-Lyme  
 192 (N: 4), Pyrogenes-Pyrogenes (N: 4), Sejroe-Saxkoebing (N: 2), Sejroe-Sejroe (N: 3), Australis-Jalna  
 193 with Cynopteri-Cynopteri (N: 1), Autumnalis-Autumnalis with Cynopteri-Cynopteri (N: 2) and  
 194 Cynopteri-Cynopteri with Sejroe-Sejroe (N: 1) (**Online Resource 1**).  
 195 Considering the results obtained by the expanded MAT assay with a cut-off  $\geq 1:100$ , the frequency  
 196 of seropositivity was significantly higher in apparently healthy kennel dogs, followed by owned  
 197 dogs showing clinical signs potentially associated with *Leptospira* infection or increased CRP  
 198 serum concentration and lower in apparently healthy blood donor dogs ( $P = 0.0124$ , **Table 2**),

199 whereas no significant association was found between seropositivity and clinical status. Frequency  
200 of seropositivity was also significantly higher in mixed breed dogs (35/60, 58.3%) compared to  
201 purebred ones and in regularly vaccinated dogs compared to not regularly vaccinated ones ( $P =$   
202 0.0198 and 0.0009 respectively, **Table 2**). No other significant association was found between the  
203 seropositivity to *Leptospira* and the variables analysed (**Table 2**). Differently, no significant  
204 association was found between the seropositivity with a cut-off  $\geq 1:800$  and all the variables  
205 analysed, including the clinical and vaccination status (**Online Resource 2**).

206

## 207 **Discussion**

208 The MAT assay with the expanded 27 antigens panel detected 13/288 (4.5%) more seropositive  
209 dogs than the routine MAT assay with the nine antigens panel. Furthermore, 35/288 (12.1%) dogs  
210 had prevalent MAT titre against at least one of the 18 *Leptospira* antigens not included in the  
211 routine MAT assay.

212 In the routine nine antigens panel, the Australis-Bratislava, Canicola-Canicola, Grippotyphosa-  
213 Grippotyphosa, Icterohaemorrhagiae-Copenhageni, Icterohaemorrhagiae-Icterohaemorrhagiae and  
214 Pomona-Pomona were the most frequently detected serovars, confirming that these are the most  
215 effective antigens for the diagnosis of *Leptospira* infection in dogs in Northern Italy. Indeed, these  
216 variants are the most widespread in Europe (Ellis, 2010), and are included in the vaccines currently  
217 adopted (Ellis, 2010; Klaasen et al., 2014; Schuller et al., 2015). The remaining three serovars  
218 included in the routine nine antigens panel were detected with low frequency and low titres  
219 (Ballum-Ballum and Sejroe-Hardjo), or undetected (Tarassovi-Tarassovi), in this study,  
220 highlighting its limited importance for the diagnosis of *Leptospira* infection in dogs. However, their  
221 inclusion in the antigen panel is justified by the use of the same MAT assay for the diagnosis of  
222 leptospirosis in other animal hosts (Tagliabue et al., 2016).

223 Among the serovars not included in the routine nine antigens panel, Australis-Jalna and Pomona-  
224 Mozdok serovars were positive only when Australis-Bratislava and Pomona-Pomona, respectively,

were positive and normally with a lower titre, suggesting a probable cross-reaction. Differently, the Sejroe-Saxkoebing and Sejroe-Sejroe serovars were positive more frequently and with higher titre than Sejroe-Hardjo serovar, suggesting that they are more effective for the diagnosis of *Leptospira* infection in dogs and its inclusion in the antigen panel of the MAT assay could be useful. A recent study conducted in Italy reported the usefulness of using the Saxkoebing and Sejroe serovars, in addition to Hardjo, for the MAT assay, identifying a higher number of seropositive dogs in a leptospirosis outbreak in a kennel (Balboni et al., 2022). The Autumnalis-Autumnalis, Cynopteri-Cynopteri, Hurstbridge-Hurstbridge, Lyme-Lyme and Pyrogenes-Pyrogenes serovars were not included in the routine antigen panel and showed a high frequency of positivity. Among these, the Cynopteri serovar is the most interesting as it often showed the highest titre, alone or in association with other serovars. The other four serovars had antibody titres usually less than 1:800 and, when associated with positivity to other serovars, they were rarely those with the highest titre, therefore, although their role in dogs cannot be ruled out with certainty, it is plausible to speculate that they could be the results of non-specific or cross reactions. Serovar Cynopteri and its serogroup are poorly documented in the literature and available data are usually limited to serological reactivity. A seroprevalence of 59% in dogs in Buenos Aires (Argentina) was reported by Tealdo and colleagues (Tealdo et al., 2007). Cynopteri serovar was also reported in dogs in Peru (Siuze et al., 2015) and in Portugal, where it was found to be among the most common reactivity in pigs, sheep and horses (Rocha, 1998). Bats are the maintenance hosts of the Cynopteri serovar (Bharti et al., 2003). While its pathogenicity is not clearly documented in the dog, its infection is clinically relevant in humans (Bharti et al., 2003), with a case reported in Poland (Zwierz et al., 1964). The remaining eight serovars not included in the routine MAT assay evaluated in this study (Bataviae-Bataviae, Celledoni-Celledoni, Hebdomadis-Hebdomadis, Javanica-Javanica, Mini-Mini, Panama-Panama, Ranarum-Ranarum, and Shermani-Shermani) were not detected or sporadically detected with low titres not exceeding 1:800 in association with other serovars, resulting negligible for the diagnosis of *Leptospira* infection in dogs in Northern Italy.

251 This study confirms that kennels may represent high-risk environments for the diffusion of  
252 leptospiral infection in dogs (Balboni et al., 2022), as demonstrated by the significantly higher  
253 frequency of seropositivity  $\geq 1:100$  for kennel dogs than for owned dogs, also confirmed by a  
254 significantly higher frequency of seropositivity in mixed breed dogs, mainly coming from kennels.  
255 Differently, the significantly higher frequency of seropositivity  $\geq 1:100$  in regularly vaccinated dogs  
256 than in not regularly vaccinated ones is a probable consequence of the positive reaction to the MAT  
257 test due to antibodies of vaccine origin.

258 The main limitation of this study was the lack of a second paired MAT test on the enrolled dogs.  
259 This aspect, associated with the typical paradoxical reactions and cross-reactivity that characterise  
260 the MAT assay (Levett, 2003; Murray et al., 2011; Smythe et al., 2009; Sykes et al., 2022), could  
261 determine a misinterpretation of the highest titres obtained. Indeed, especially if an animal is tested  
262 only once, the antigen with the highest titre cannot be considered with certainty the infecting  
263 serovar. The lack of a second paired MAT test would have been an important limitation if the main  
264 aim of the study was to assess the diffusion of the different serovars in Northern Italy. Otherwise, as  
265 the aim of this study was to identify the most effective serovars to include in the MAT antigen panel  
266 for the diagnosis of *Leptospira* infection in dogs and because the antibody titre was only interpreted  
267 as a measure of seroreactivity, this can be considered a negligible limitation. Nevertheless,  
268 seroepidemiological studies involving the analysis of paired serum samples collected at appropriate  
269 times (acute and convalescent) in dogs with clinical signs potentially associated with *Leptospira*  
270 infection should be performed to more accurately detect which serogroups circulate and cause  
271 disease in the canine population.

272

## 273 **Conclusions**

274 In general, the antigen panel currently adopted for the routine MAT assay in Northern Italy resulted  
275 adequate for the diagnosis of *Leptospira* infection in dogs, both for the serogroups detected and for  
276 the choice of the serovar within the serogroup. The main exception concerns the Sejroe serogroup,

277 with the Saxkoebing and Sejroe serovars that were more effective than Hardjo for diagnosis in dogs  
278 and whose inclusion in the antigen panel is recommended. Among other antigens evaluated in this  
279 study, Cynopteri serovar was detected with high frequency but, as it was usually in association with  
280 other serogroups, its pathogenic role in dogs and public health threats deserve further investigation.

281

## 282 **Statements and Declarations**

283 **Funding:** This work was supported by the Italian Ministry of Health (Grant numbers RC IZS VE  
284 16/12).

285 **Competing interests:** The authors have no relevant financial or non-financial interests to disclose.

286 **Authors' contributions:** All authors contributed to the study conception and design. Material  
287 preparation, data collection and laboratory analysis were performed by Mario D'Incau, Silvia  
288 Marchione and Alda Natale. The analysis of the results was performed by Andrea Balboni, Mario  
289 D'Incau and Silvia Zamagni. The first draft of the manuscript was written by Andrea Balboni and  
290 Silvia Zamagni. Mario D'Incau, Mara Battilani, Laura Lucchese, Elisa Mazzotta and Alda Natale  
291 supervised and commented on previous versions of the manuscript. Funding acquisition was made  
292 by Alda Natale. All authors read and approved the final manuscript.

293 **Availability of data and material:** The datasets generated during and/or analysed during the current  
294 study are available from the corresponding author on reasonable request.

295 **Ethics approval:** Ethical approval were waived for this study, because only blood samples taken by  
296 clinicians from owned dogs for diagnostic purposes and surplus material derived from blood  
297 samples taken by clinicians from kennel dogs undergoing neutering surgery sampled to perform  
298 pre-operative profile tests or from donor dogs sampled to perform pre-donation screening tests were  
299 used. For all dogs, blood sampling was performed following owner or legal manager of the kennel  
300 consent.

301 **Consent to participate:** Not applicable.

302 **Consent to publish:** Not applicable.

303

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413 **Table 1** Panel of eleven *Leptospira* spp. used as live antigens for MAT assay and results obtained.

| Serogroup           | Serovar             | Strain                            | Routine<br>MAT <sup>a</sup> | Expanded<br>MAT <sup>b</sup> | Dogs tested positive<br>with cut-off ≥ 1:100 | Dogs tested positive<br>with cut-off ≥ 1:800 | Highest MAT titre |
|---------------------|---------------------|-----------------------------------|-----------------------------|------------------------------|--|--|-------------------|
| Australis           | Bratislava          | Riccio 2                          |                             |                              | 54 (18.7%)                                   | 26 (9%)                                      | 1:6400            |
| Australis           | Jalna               | Jalna                             |                             |                              | 42 (14.6%)                                   | 19 (6.6%)                                    | 1:6400            |
| Autumnalis          | Autumnalis          | Akiyami A                         |                             |                              | 25 (8.7%)                                    | 9 (3.1%)                                     | 1:6400            |
| Ballum              | Ballum              | Mus 127                           |                             |                              | 9 (3.1%)                                     | 1 (0.3%)                                     | 1:800             |
| Bataviae            | Bataviae            | Swart                             |                             |                              | 1 (0.3%)                                     | 0 (0%)                                       | 1:100             |
| Canicola            | Canicola            | Alarik                            |                             |                              | 53 (18.4%)                                   | 5 (1.7%)                                     | 1:1600            |
| Celledoni           | Celledoni           | Celledoni                         |                             |                              | 0 (0%)                                       | 0 (0%)                                       | 0                 |
| Cynopteri           | Cynopteri           | 3522 C                            |                             |                              | 27 (9.4%)                                    | 13 (4.5%)                                    | 1:6400            |
| Grippotyphosa       | Grippotyphosa       | Moskva V                          |                             |                              | 25 (8.7%)                                    | 14 (4.9%)                                    | 1:6400            |
| Hebdomadis          | Hebdomadis          | Hebdomadis                        |                             |                              | 2 (0.7%)                                     | 0 (0%)                                       | 1:400             |
| Hurstbridge         | Hurstbridge         | BUT 6                             |                             |                              | 14 (4.9%)                                    | 2 (0.7%)                                     | 1:1600            |
| Icterohaemorrhagiae | Copenhageni         | Wijnberg                          |                             |                              | 87 (30.2%)                                   | 21 (7.3%)                                    | 1:6400            |
| Icterohaemorrhagiae | Icterohaemorrhagiae | Bianchi                           |                             |                              | 59 (20.5%)                                   | 14 (4.9%)                                    | 1:6400            |
| Javanica            | Javanica            | Veldrat Bataviae 46               |                             |                              | 7 (2.4%)                                     | 3 (1%)                                       | 1:800             |
| Lyme                | Lyme                | 10                                |                             |                              | 20 (6.9%)                                    | 3 (1%)                                       | 1:3200            |
| Mini                | Mini                | Sari                              |                             |                              | 0 (0%)                                       | 0 (0%)                                       | 0                 |
| Panama              | Panama              | CZ 214 K                          |                             |                              | 5 (1.7%)                                     | 1 (0.3%)                                     | 1:800             |
| Pomona              | Mozdok              | 5621                              |                             |                              | 15 (5.2%)                                    | 5 (1.7%)                                     | 1:3200            |
| Pomona              | Pomona              | Pomona                            |                             |                              | 15 (5.2%)                                    | 8 (2.8%)                                     | 1:6400            |
| Pyrogenes           | Pyrogenes           | Salinem                           |                             |                              | 21 (7.3%)                                    | 3 (1%)                                       | 1:1600            |
| Ranarum             | Ranarum             | ICF                               |                             |                              | 2 (0.7%)                                     | 1 (0.3%)                                     | 1:800             |
| Sejroe              | Hardjo              | Hadjoprajitno/g.t. hardjoprajitno |                             |                              | 6 (2.1%)                                     | 1 (0.3%)                                     | 1:1600            |
| Sejroe              | Hardjo              | Sponselee/g.t. hardjobovis        |                             |                              | 5 (1.7%)                                     | 1 (0.3%)                                     | 1:3200            |
| Sejroe              | Saxkoebing          | Mus24                             |                             |                              | 9 (3.1%)                                     | 3 (1%)                                       | 1:6400            |
| Sejroe              | Sejroe              | M84                               |                             |                              | 16 (5.6%)                                    | 3 (1%)                                       | 1:3200            |
| Shermani            | Shermani            | LT 821                            |                             |                              | 0 (0%)                                       | 0 (0%)                                       | 0                 |
| Tarassovi           | Tarassovi           | Mitis-Johnson                     |                             |                              | 0 (0%)                                       | 0 (0%)                                       | 0                 |

414 <sup>a</sup> MAT assay with antigen panel composed by eight serogroups and nine serovars.

415 <sup>b</sup> MAT assay with antigen panel composed by 20 serogroups and 27 serovars.

416 **Table 2** Descriptive statistics of the dogs included in the study population and comparison between  
 417 dogs tested positive and dog tested negative by MAT assay with expanded 27 antigens panel  
 418 adopting a cut-off  $\geq 1:100$ .

| Variables  | Total       | Positive      | Negative        | P value       |
|--|-------------|---------------|-----------------|---------------|
| Number of dogs   | 288         | 141 (49%)     | 147 (51%)       |               |
| Origin   |             |               |                 |               |
| Owned dogs showing clinical signs potentially associated with <i>Leptospira</i> infection or increased CRP serum concentration | 217         | 110 (50.7%)   | 107 (49.3%)     | <b>0.0124</b> |
| Apparently healthy kennel dogs   | 20          | 14 (70%)      | 6 (30%)         |               |
| Apparently healthy blood donor dogs  | 51          | 17 (33.3%)    | 34 (66.7%)      |               |
| Sex  |             |               |                 |               |
| Male   | 108         | 50 (46.3%)    | 58 (53.7%)      | 0.9342        |
| Female   | 94          | 45 (47.9%)    | 49 (52.1%)      |               |
| NA   | 86          |               |                 |               |
| Age <sup>a</sup>   | 5y [1m-16a] | 5y6m [1m-16y] | 4y1m [1m-14y4m] |               |
| <1   | 25          | 12 (48%)      | 13 (52%)        | 0.5451        |
| 1-5  | 66          | 33 (50%)      | 33 (50%)        |               |
| 6-10   | 49          | 29 (59.2%)    | 20 (40.8%)      |               |
| >10  | 15          | 6 (40%)       | 9 (60%)         |               |
| NA   | 155         |               |                 |               |
| Breed  |             |               |                 |               |
| Purebred   | 133         | 52 (39.1%)    | 81 (60.9%)      | <b>0.0198</b> |
| Mixed breed  | 60          | 35 (58.3%)    | 25 (41.7%)      |               |
| NA   | 95          |               |                 |               |
| Vaccination  |             |               |                 |               |
| Yes  | 94          | 59 (62.8%)    | 35 (37.2%)      | <b>0.0009</b> |
| No <sup>b</sup>  | 59          | 20 (33.9%)    | 39 (66.1%)      |               |
| NA   | 135         |               |                 |               |
| Clinical status  |             |               |                 |               |
| Sick   | 217         | 110 (50.7%)   | 107 (49.3%)     | 0.3725        |
| Apparently healthy   | 71          | 31 (43.7%)    | 40 (56.3%)      |               |

419 The Chi-squared test were carried out on the positive and negative dogs. Not available data was  
 420 excluded to statistical analysis. Data are reported as n (%).

421 <sup>a</sup> Data are reported as median [range] and in four age classes, statistical analysis was carried out on  
 422 categorical age classes.

423 <sup>b</sup> Dog not vaccinated or vaccinated for more than 12 months (Day et al., 2016).

424 Values in bold indicate statistical significance. m: months. NA: not available. y: years.

425 **Table 3** Comparison between routine MAT results and expanded MAT results.

| Cut-off ≥ 1:100           |          |             |           |             | Cut-off ≥ 1:800           |          |            |             |            |
|---------------------------|----------|-------------|-----------|-------------|---------------------------|----------|------------|-------------|------------|
| Expanded MAT <sup>b</sup> |          |             |           |             | Expanded MAT <sup>b</sup> |          |            |             |            |
|                           |          | positive    | negative  | total       |                           |          | positive   | negative    | total      |
| Routine                   | positive | 128 (44.5%) | 0 (0%)    | 128 (44.5%) | Routine                   | positive | 46 (16%)   | 0 (0%)      | 46 (16%)   |
| MAT <sup>a</sup>          | negative | 13 (4.5%)   | 147 (51%) | 160 (55.5%) | MAT <sup>a</sup>          | negative | 6 (2.1%)   | 236 (81.9%) | 242 (84%)  |
|                           | total    | 141 (49%)   | 147 (51%) | 288 (100%)  |                           | total    | 52 (18.1%) | 236 (81.9%) | 288 (100%) |

426 <sup>a</sup> MAT assay with antigen panel composed by eight serogroups and nine serovars.

427 <sup>b</sup> MAT assay with antigen panel composed by 20 serogroups and 27 serovars.