Supporting Information

Conformational disorder analysis of the conditionally disordered protein CP12 from *Arabidopsis thaliana* in its different redox states

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Figure S1. EOM optimizations. Results of the EOM optimizations to fit the SAXS profile of oxidized (left; AtCP12ox) and reduced (right; AtCP12red) AtCP12. For each pool of conformers (*dis* in purple, *bin* in light blue, *bis* in green, *tef* in red, *ter* in black, *bis-tef* in orange), the plots in the first and second columns of panels show R_g and D_{max} frequency distributions for the whole generated initial pool (black solid lines) and for the final selected ensembles to fit SAXS data (coloured shaded areas, overlap of the final distributions from eight independent optimizations). The pool codes reported are explained in the main text and in Figure 2, and here reminded by visualization of an example model. In the last column of panels the theoretical profile (solid line) obtained by the optimization with the lowest χ^2 among the repetitions is shown superimposed to the experimental data points (coloured dots) in Kratky plot representation, and below the corresponding normalized residuals $\frac{I_{calc}(q)-I_{exp}(q)}{I_{exp}(q)}$ are shown.

CrCP12	4	SGQPAVDLNKKVQDAVKEAEDACAKG-TSADCA	VAWDTVEELSAAVSHKKDAV:	52
AtCP12	2	APEGGISDVVEKSIKEAQETCAGDPVSGECV	YAAWDEVEELSAAASHARDKK	51
CrCP12	53	KADVTLTDPLEAFCKDAPDADECRVYED	80	
AtCP12	52	KADGSDPLEEYCKDNPETNECRTYDN	77	

Figure S2. Sequence alignment between AtCP12 and CrCP12. The alignment was performed with EMBOSS Matcher highlighting the N-terminal (in red) and C-terminal (in blue) possibly structured regions assumed when building conformational pools.



Figure S3. Experimental SAXS data collected for the dilution series of samples of oxidized and reduced AtCP12. Dilution series of oxidized AtCP12 (**a**) and reduced AtCP12 (**d**). The values of R_g estimated by Guinier fit in different q ranges for all concentrations are reported in (**b**) for oxidized AtCP12 and (e) for reduced AtCP12. The final merged data (black solid lines in (**a**) and (**d**)) are also shown in (**c**) and (**f**) together with the portion of the original experimental data at low (blue line) and high (red line) concentration.

Guinier fit	Oxidized AtCP12	Reduced AtCP12
R _g (Å)	2.02	2.32
$\sigma(R_g)$ (Å)	0.03	0.07
I(0) [kDa]	9.77	9.98
$\sigma(I(0))$	0.11	0.26
q range (nm ⁻¹)	0.44-0.65	0.44-0.57
Fidelity	1.0	0.92
Indirect Fourier transform		
Rg (nm)	2.29	2.9
$\sigma(R_g)$ (nm)	0.03	0.1
I(0) [kDa]	10.4	11.5
$\sigma(I(0))$	0.1	0.3
q range (nm ⁻¹)	0.36-3.5	0.36-3.5
D _{max} imposed for P(r) (nm)	9	11.6
D _{max} variability estimate (nm)	2	3
GNOM quality estimate	0.565	0.547
MW Bayesian		
q minimum (nm ⁻¹)	0.27	0.44
estimate (kDa)	7.3	7.3
estimate probability (%)	99.05	90.23
credibility interval (kDa)	6.65-9.05	6.65-9.05
interval probability (%)	99.35	99.62
MW from sequence (kDa)	8.26	8.26

 Table S1. Summary of model independent SAXS data analysis of oxidized and reduced AtCP12.

Table S2. Summary of the EOM analysis of AtCP12 SAXS experimental data. (a) average structural parameters of the generated random pools of conformers: the radius of gyration (R_a), the maximum particle size (D_{max}) , and the flexibility index (R_{flex}) . The R_{flex} index (between 0 and 1) is a metric obtained from the entropy (information content) of the distribution and can be used to approximate the flexibility [37]. Using this metric the size distributions of the selected ensembles after fitting can be numerically compared to that of the initial random pool, which is supposed to represent the reference for the randomness of the distribution under the specific structural constraints. The average R_a of each pool is also compared to the experimental R_g obtained from the Guinier fit and reported as a % relative deviation. (b) Structural parameters and fitting results obtained by eight repetitions of the fitting algorithm applied to the SAXS data of AtCP12 in oxidized form (top) and in reduced form (bottom) considering each of the generated pools. The average value and standard deviation for each quantity are reported reported. From left to right, for each pool these parameters are shown: average Rg of the selected ensemble of conformers; average D_{max} of the selected ensemble of conformers; flexibility parameter of the selected distribution; check parameter representing the ratio between the standard deviations of the selected and starting size histograms distributions; experimental vs. calculated SAXS intensity agreement index χ^2 ; optimized constant background subtracted from the data while fitting.

(a)						
Pool	ool Pool average		Flexibility index	%Rg deviation		
	Rg (nm)	D _{max} (nm)	R _{flex}	Oxidized (R _g = 2.03 nm)	Reduced (R _g = 2.32 nm)	
dis	2.46	7.41	0.86	-21	-6	
bin	2.43	7.39	0.88	-20	-5	
bis	2.35	7.11	0.85	-16	-1	
tef	1.94	6.06	0.89	5	17	
ter	2.20	7.41	0.30	-8	5	
bis-tef	2.14	6.59	0.84	-5	8	

(b)

Fitting data	Select	ected ensemble average		Flexibility index		Distribution width ratio		Goodness of fit		Constant background		
	<r<sub>g> (nm)</r<sub>	σ	<d<sub>max> (nm)</d<sub>	σ	<r<sub>flex></r<sub>	σ	< <i>R</i> ₀>	σ	< χ²>	σ	<bg> (kDa)</bg>	σ
Oxidized AtCP12		_										
dis	2.61	0.04	8.05	0.17	0.91	0.001	1.21	0.005	0.85	0.017	0.12	0.007
bin	2.57	0.05	8.00	0.13	0.93	0.001	1.32	0.005	0.69	0.007	0.14	0.007
bis	2.44	0.04	7.54	0.14	0.91	0.001	1.30	0.007	0.66	0.007	0.14	0.006
tef	2.13	0.01	6.62	0.04	0.90	0.001	1.04	0.005	0.63	0.002	0.16	0.003
ter	2.20	0.00	7.41	0.00	0.23	0.002	0.73	0.008	1.65	0.004	0.14	0.001
bis-tef	2.28	0.03	7.05	0.12	0.86	0.001	1.15	0.008	0.63	0.001	0.15	0.004
Reduced AtCP12												
dis	2.69	0.03	8.14	0.17	0.90	0.001	1.18	0.005	0.66	0.001	0.121	0.004
bin	2.64	0.02	8.07	0.11	0.91	0.001	1.15	0.005	0.66	0.001	0.135	0.003
bis	2.56	0.02	7.75	0.11	0.89	0.001	1.18	0.007	0.66	0.001	0.128	0.004
tef	2.06	0.01	6.30	0.07	0.84	0.001	0.81	0.005	1.18	0.013	0.193	0.003
ter	2.20	0.00	7.41	0.00	0.23	0.003	0.73	0.007	4.35	0.013	0.171	0.001
bis-tef	2.44	0.02	7.38	0.10	0.86	0.001	1.07	0.005	0.68	0.002	0.142	0.002

Table S3. Summary of SAXS data acquisition information, sample details, and data analysis software used.

	Oxidized AtCP12	Reduced AtCP12			
Concentration range (mg ml ⁻¹)	0.25-5.35	0.27-5.57			
Storage and dilution buffer composition	25 mM K-phosphate, pH 7.5	25 mM K-phosphate, pH 7.5			
Extinction coefficient for concentration	8730	8480			
determination from UV absorbance at					
280 nm (M ⁻¹ cm ⁻¹)					
(b) SAXS data collection parameters					
Source, instrument	ESRF	F, BM29			
Detector	PILA	TUS 1M			
Beam geometry (mm ²)	0.7	x 0.7			
Wavelength (Å)	0.9919				
sample-to-detector distance (m)	2.872				
q-measurement range (nm ⁻¹)	0.05-0.45				
Absolute scaling method	water scattering I(0)= 0.01632 cm ⁻¹ ,				
	protein partial specific volume 0.735 cr	m ³ g ⁻¹			
Capillary path length (mm)		1.8			
Injection volume (µI)		60			
Exposure time (s)		1			
Number of exposures		10			
Extra flow time (s)		10			
Sample temperature (°C)		4			
(c) Software employed for SAS data reduc	tion, analysis, and interpretation				
Solvent subtraction, averaging and basic a	nalysis (Guinier fit, ATSAS 3.2				
P(R), MW)					
Theoretical intensity calculations	CRYSOL 2.8				
Ensemble optimization	EOM 3.0				
Molecular graphics	PyMOL 1.8				