

Alma Mater Studiorum Università di Bologna Archivio istituzionale della ricerca

Production of biocrude from organic waste: Influence of feedstock composition on hydrodenitrogenation reactivity in biocrude upgrading

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Matayeva A., Fasolini A., Bianchi D., Chiaberge S., De Maron J., Basile F. (2023). Production of biocrude from organic waste: Influence of feedstock composition on hydrodenitrogenation reactivity in biocrude upgrading. FUEL, 335, 1-8 [10.1016/j.fuel.2022.126981].

Availability: This version is available at: https://hdl.handle.net/11585/915068 since: 2024-04-04

Published:

DOI: http://doi.org/10.1016/j.fuel.2022.126981

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (https://cris.unibo.it/). When citing, please refer to the published version.

(Article begins on next page)

1	Production of biocrude from organic waste: influence of feedstock composition on
2	hydrodenitrogenation reactivity in biocrude upgrading.
3	
4	Aisha Matayeva ^{a,b} , Andrea Fasolini ^a , Daniele Bianchi ^c , Stefano Chiaberge ^c , Jacopo De Maron ^a , Francesco
5	Basile ^{a, *}
6	^a Dipartimento di Chimica industriale "Toso Montanari", University of Bologna, 40136 Bologna, Italy
7	^b Department of Biological and Chemical Engineering, Aarhus University, Hangøvej 2, 8200 Aarhus N,
8	Denmark
9	^c Eni S.p.A, Renewable, New Energies and Material Science Research Center, Via Giacomo Fauser, 4 -
10	28100 Novara
11	* <u>f.basile@unibo.it</u>
12	
13	Hydrothermal liquefaction (HTL) process of protein-rich biomass produces biocrude with a
14	high nitrogen content that hinders its use as a biofuel intermediate. Nitrogen compounds in
15	the biocrudes are mainly present as fatty acid amides (FAAs) and nitrogen-containing
16	polycyclic aromatic compounds (NAs), having different hydrodenitrogenation reactivity
17	during the biocrude upgrading process. Therefore, it is crucial to shift reaction pathways
18	towards the formation of less recalcitrant compounds and reduce the nitrogen content of the
19	biocrudes. Herein, the impact of the chemical composition, in particular a lipid content, on
20	the biocrude yield, nitrogen content and types of nitrogen species has been studied using
21	macromolecular model compounds. By comparing the hydrodenitrogenation (HDN)
22	reactivity of FAAs and NAs, it was found that the latter are more recalcitrant compounds
23	compared to the FAAs and that the higher lipid content inhibits the generation of NAs. These
24	results suggest that lipids-rich waste can be supplemented with food waste to produce the
25	biocrudes highly suitable for conventional hydrotreatment processes.
26	Keywords: hydrothermal liquefaction, hydrotreatment, food waste, nitrogen, bio-oil, fatty acid

amides.

29 1 Introduction

30 About one-third of food produced for human consumption is wasted globally [1], while 31 approximately 88 million of food waste is accumulated only in the European Union per year [2]. 32 Majority of the food waste are mainly ended up in landfills [3] and this has led to serious 33 environmental concerns, as different microorganisms cause serious bacterial contaminations and 34 infectious diseases due to their nutritional composition [4]. Moreover, the food waste causes the 35 greenhouse gas emissions of about 4.4 Gt CO_2 equivalent per year [5], accounting for 12.9% of 36 the global CO₂ emissions for energy production reported by the IEA World Energy Outlook 2021 37 $(34.156 \text{ Gt CO}_2 \text{ equivalent per year})$ [6].

38 Hydrothermal liquefaction (HTL) is a promising approach to minimize the generation of organic 39 food waste and recover energy [7]. Generally, HTL operates at the sub- and supercritical region 40 of water (250–400 °C; 200–300 bar) for a short reaction time [8]. The HTL process provides two 41 main advantages: (i) no requirement for drying the wet feedstock, which is an energy intensive 42 process; (ii) unique attributes of subcritical water, such as a high ionic product and lower dielectric 43 constant [9]. Under HTL conditions the biomass is depolymerized into four different products, 44 namely biocrude oil, aqueous, solid residue, and gas phases [9, 10]. Among these fractions, the 45 biocrude oil is of great interest, as it can be used as drop-in fuels after catalytic upgrading process. 46 Food waste consists of three main macromolecular fractions, namely carbohydrates, proteins and 47 lipids [11]. It has been reported that about 20-40 wt.% of nitrogen in the protein can migrate into 48 the biocrude fraction [12], causing a high nitrogen content (>10 wt.%). In comparison, the crude 49 fossil fuels are characterized by much lower one (>1 wt.%) [12]. Several studies on real petroleum 50 feeds showed that lower than 50 ppm of nitrogen in the feedstock can deactivate catalytic active 51 sites in conventional refinery units [13-15]. Consequently, the existing conventional refinery 52 infrastructures are not suitable for the upgrading of waste-derived oils, especially for 53 hydrodenitrogenation (HDN), while constructing new biorefinery units for the biocrudes is not

54 economically feasible due to the high capital investments, longer implementation period, higher 55 risks and uncertainty regarding the process efficiency [16]. In addition, little is known about the 56 HDN reactivity of nitrogen compounds of the biocrudes [17]. Optimization of the HTL reaction 57 parameters, such as reaction temperature, operating reaction time, heating rate, etc., is typically 58 not effective, as the low nitrogen content always accompanies with the lower bio-oil yield [18-59 20]. On the other hand, engineering the feedstock composition for balancing the biocrude quantity 60 and quality (lower O and N contents) is highly relevant to meet the minimum specifications of 61 the upgrading process at the existing refinery units.

62 In this context, it is important to comprehend the behavior of proteins under hydrothermal 63 conditions and their contribution into the biocrude qualities. In our previous work [21] we 64 investigated the mechanism of the nitrogen moieties generation under HTL conditions employing 65 amino acids (leucine, phenylalanine) as protein model compounds in the presence of organic acids 66 and alkali catalysts. Our results showed that the addition of acetic acid into the system greatly 67 increases the biocrude yield as well as the carbon yield due to the transference of hydrophilic 68 amino moieties from the aqueous phase after acetylation reaction forming more hydrophobic 69 amides. Most importantly, we reported that with the addition of lipids the generation of fatty acid 70 amides (FAAs) competes with the formation of nitrogen- containing polycyclic aromatic 71 compounds (NAs) by Maillard reaction, observed in the presence of polysaccharides [21, 22]. 72 Likewise, Fan, Y. et al. (2020) [23] confirmed the existence of the strong counteraction between 73 amidation and Maillard reaction products, using lactose, lysine and palmitic acid as model 74 compounds of carbohydrate, protein and lipid, respectively. As both studies have been performed 75 with only surrogate model mixtures, further investigations are still needed using more complex 76 mixtures in order to gain the exhaustive understanding about how the chemical composition of 77 biomass affect the specimens of the nitrogen in the final biocrude and their HDN reactivity during 78 the hydrotreatment process of the food waste-derived biocrudes.

In the present study we performed the HTL experiments by employing macromolecular modelcompounds (albumin, starch, tripalmitin) to demonstrate the competition between the formation

81 FAAs and NAs. Moreover, we studied the influence of lipid content on the biocrude yield and 82 nature of nitrogen species in the biocrude by quantifying the relative contents of FAAs and NAs. 83 To the best of our knowledge, the impact of chemical composition of biomass waste on the nature 84 of nitrogen species (FAAs and NAs) has not been previously semi-quantified. Last but not least, 85 we also evaluated the HDN reactivity of the two main nitrogen compound classes, FAAs and 86 NAs, in the biocrude produced from the food waste. The results obtained in the present study are 87 important to build better fundamentals for formulating the feedstock composition to produce the 88 biocrude with the higher yield as well as quality, and, most importantly, make the biocrude 89 components less problematic for the existing refinery infrastructures, thus avoiding the additional 90 investment costs.

91

104

92 **2. Materials and methods**

93 2.1 HTL experiments

94 HTL experiments were carried out in a Parr 2L batch reactor (4520 series) at 300 °C for 60 95 minutes with a heating rate of 2.5 °C/min. The detailed experiment procedure has been described 96 elsewhere [21, 22]. The elemental composition of the albumin, starch and tripalmitin is given 97 Table S1. Shortly, 7 g of the feedstock was mixed with 300 g of water solution and placed into 98 the reactor. For the binary and ternary mixtures of albumin with starch and tripalmitin, equivalent 99 masses of each component (4 g) were employed with the same ratio of the feedstock to water. 100 Before running experiments, the reactor was purged with nitrogen to remove any residual air. 101 After 60 minutes of reaction time, the reactor was cooled with a cooling rate of 11 °C/min by 102 running cold water in an internal coil of the reactor. 103 The gas yield was determined by the ideal gas law taking into account the residual pressure. The

105 Chromatograph (GC). The gas phase was found to consist mainly of CO₂. The biocrude extraction

gas fraction was then collected in a gas sample bag and its composition was analyzed by Gas

has been done using ethyl acetate and the solvent was removed by rotary evaporation. The
biocrude and aqueous phases were stored at 5 °C until further analytical characterization.

108 **2.2 Characterization of HTL products**

The composition of the gas fraction was analyzed by Gas Chromatograph (Agilent 7890A)
equipped with a carboxen 1006 Plot column (30 m x 0.32 mm, average thickness 30 µm). The
method used for the gas analysis has been described elsewhere [21, 22].

112 The chemical composition of biocrudes were determined using a GC-MS with a Finnigan Trace 113 DSQ (Thermo) quadrupole mass spectrometry, interfaced to a Finnigan Trace GC Ultra. The 114 separation of biocrude components has been done using a DB-5 MS (Agilent J&W) fused silica, 115 non-polar capillary column (30 m x 0.25 mm ID x 0.25 m). Helium was used as a carrier gas (1 116 ml/min). The method was described well in previous studies [21, 22]. Shortly, about 1 mg/ml of 117 biocrude samples in diethyl ether were prepared. The oven temperature was set at the temperature 118 of 60 °C for 2 minutes, followed by a heating ramp of 10 °C/min to the final temperature of 320 119 °C. The acquisition of mass spectra was done in electronic ionization mode with a mass range of 120 50-650 Da.

121 The content of free fatty acids was quantified by GC-FID equipped with Supelco Petrocol 122 EX2887 (5m x 0.53 mm ID, thickness 0.1 µm). Helium was employed as a carrier gas (40 cm/sec). 123 BSTFA (N,O-bis(trimethylsilyl)trifluoroacetamide) was used as a derivitization agent for 124 silvlation reaction of fatty acids (palmitic, stearic, mono- and dioleic acids) present in the 125 biocrudes. After the addition of about 100 µl of BSTFA and 200 µl of pyridine to biocrude 126 samples, the solutions were heated at 70°C for 40 min. After silvlation of fatty acids, the samples 127 were diluted with 1 ml of dichloromethane, followed by addition of $50 \,\mu$ l of tetradecane (4 mg/ml 128 in hexane) and 25 µl of tricaprin (8000 ppm in pyridine) as internal standards. The oven 129 temperature was set at the temperature of 50 °C for 2 minutes and heated up to the final 130 temperature of 350 °C with a heating rate of 10 °C/min. Elemental composition of biocrude and 131 solid fractions was analyzed using an elemental analyzer Flash 2000 Thermo Fisher.

132 Fourier transform ion cyclotron resonance mass spectrometry was utilized to analyze the 133 composition of biocrude samples. The analysis was carried out using a 7 T FTICR MS (LTQ-FT 134 Ultra Thermo Scientific), equipped with ESI (Electrospray) ion source. The mass spectra were 135 mainly acquired in positive mode (ESI+). The typical conditions for the ESI+ analysis were 136 described in details elsewhere [21, 22, 24]. Generally, the spectra were collected simultaneously 137 with a low resolution linear ion trap (m/z 100-1000) and with a 7 T ultrahigh resolution FTICR 138 cell with a mass range of m/z 100-1000. The resolution was fixed at 400.000 (m/z 400) and the 139 ion accumulation time was defined by the automatic gain control, which was set to 10^6 . About 140 100 scans were collected and averaged for each analysis for the improvement of the signal-to-141 noise ratio. Data acquired were processed by Xcalibur software (Thermo Scientific). Since a 142 single accurate mass could generate the great amount of potential different combinations of 143 elements, some restrictions are required to the element ranges. In the present study, the 144 restrictions were selected based on the elemental analysis of biocrude samples with the error range 145 of ± 2.5 ppm: 0-60 12 C, 0-2 13 C, 10-100 H, 0-6 N, 0-1 31 S, 0-6 O. The assignment of molecular 146 formula was performed first below 400 Da due to the lower number of possible combinations for 147 a single mass and then the mass peaks above 400 Da were assigned through the Kendrick mass 148 [25]. The list of corresponding molecular formulas with the masses were categorized into groups 149 using a custom-built software ISOMASS [26] and the mass peaks related to isotopes were 150 determined and removed [24].

Then relevant peaks were grouped based on several parameters, in particular, the number of heteroatoms (N, O, S) and the unsaturation degree, assigned as DBE (double bond equivalents) [27, 28]. The DBE value for each molecular formula $C_cH_hN_nO_oS_s$ was defined as DBE=ch/2+n/2+1. The molecular formulas were specified to about 90 % of the peaks with the relative intensities higher than 0.1 %.

156

157 **2.3. Biocrude hydrocracking upgrade**

158	The biocrude samples were upgraded by a hydrocracking treatment carried out under the reacti	on
159	conditions typically used for the fossil heavy oil treatment [29]. The biocrude sample was add	ed
160	into a stirred reactor together with an oleo-soluble catalyst precursor (Mo ^{IV} 2-ethylhexanoat	e).
161	During the upgrading process, the catalyst precursor converts to highly dispersed molybdenu	ım
162	sulfide (MoS ₂) nanoparticles [29].	
163	The reaction occurs at 430°C and 160 bar of hydrogen for 3 hours with 3000 wppm MoS_2 cataly	yst
164	concentration. At the end of the reaction, the mixture was extracted with tetrahydrofuran (TH	F),
165	filtered and directly analyzed.	
166		
167	2.4. Product yield determination	
168	The yields of biocrude and solid fractions were determined according to Equation 1:	
169	$X_{fraction} = Mass_{fraction} / Mass_{feedstock} * 100\% $ (Eq	.1)
170	High heating values of biocrudes and upgraded oils were defined by Dulong's formula, where	C,
171	H and O represent the weight percentage of the carbon, hydrogen and oxygen, respectively.	
172	$HHV(MJ/kg) = 0.338 \cdot C + 1.428 \cdot (H - O/8)$ (Eq.	2)
173	Energy Recovery (ER) in the biocrude samples was determined by the following formula:	
174	$ER = X_{biocrude} \cdot HHV_{biocrude} / HHV_{feedstock}$ (Eq	3)
175	The nitrogen and oxygen removal are calculated by the following formula:	
176	$O_{removal} = (O_{initial} - O_{final}) / O_{initial} \cdot 100\% $ (Eq.	4)
177	$N_{removal} = (N_{initial} - N_{final}) / N_{initial} \cdot 100\% $ (Eq.	5)
178		
179	3 Results and discussions	
180	3.1 HTL of binary mixtures with starch and tripalmitin	
181		
182	The nitrogen content in biocrudes is mainly originated from the protein source. Still, the present	ce
183	of lipids or carbohydrates can promote or inhibit the migration of nitrogen into the biocru	de

184 fraction. To investigate the interaction mechanisms between biomass fractions, representative 185 macromolecular model compounds, such as albumin, starch and tripalmitin, were tested in both 186 water and water-acetic binary solvent system. Table 1 lists the biocrude yields, element analysis, 187 HHV and ER values of biocrudes produced from binary mixtures HTL.

188

Table 1. Biocrude yields, elemental analysis, HHV and ER values of biocrudes from albumin
and binary mixtures of albumin/starch and albumin/tripalmitin.

	Albumin	Albumin/Starch	Albumin/Tripalmitin
Biocrude yield, wt.%	7.4±1.5	6.3±0.21	70.5±3.6
	Element a	nalysis, wt.%	
С	72.6	71.0	74.3±0.6
Ν	6.6	6.2	2.2±0.07
Н	9.1	7.5	11.4±0.3
S	3.5	2.6	<0.1
O*	7.9	12.7	12.1
Solid yield, wt.%	0.11±0.01	0.1 ± 0.01	0.15 ± 0.07
HHV, MJ/kg	35.6	32.7	38.2
ER, %	10.3	9.06	82.9

191 ** calculated by difference, %*

- 192
- 193

The biocrude yield produced from albumin and its nitrogen content was in agreement with the previous study performed with the egg albumin at 300°C [30]. The solid yield was negligible, thus implying a full conversion of albumin under hydrothermal conditions. It is worth noting that the yield of the biocrude fraction produced from protein fraction of different feedstocks vary widely based on its amino acid distribution. When the peptide bonds in the protein undergo hydrolysis, some amino acids with hydrophobic side chains (phenylalanine, tryptophan, etc.) are
partitioned to the biocrude phase. On the other hand, hydrophilic amino acids (serine, asparagine,
etc.) contribute to the generation of water soluble compounds [22].

The Albumin/Starch binary mixture produced lower biocrude yield than that from the albumin alone. However, the nitrogen content of the biocrude was not reduced correspondingly even if the albumin content in the binary mixture was twice less compared to the individual experiment with the albumin. This indicates the occurrence of the interaction reactions between starch and albumin decomposition products, causing the generation of more nitrogen compounds in the biocrude fraction.

It should be pointed out that the experiments with albumin and albumin/starch binary mixture have been also performed in an acetic acid-water solution (5/95 % wt.%) (Table S2). As previously reported for model amino acids [21], also in the present study a significant improvement of the biocrude yield was observed due to the selective acetylation of water soluble amines forming more hydrophobic amides.

The binary mixture of albumin/tripalmitin resulted in the highest biocrude yield due to the contribution of fatty acids produced from the tripalmitin under HTL conditions and their derivatives into the biocrude phase. In this case, the decreased nitrogen content can be attributed to two main factors: the dilution of protein derived fragments by long carbon chains of fatty acids that are quantitatively partitioned into the biocrude phase and the reaction between the acidic moiety of palmitic acid and water soluble amines to give hydrophobic amides [32].

Energy recovery is an important indicator for the efficiency of the HTL process. In this context, the greatest ER was observed for the albumin/tripalmitin binary mixture due to the highest biocrude yield and HHV values. In contrast, co-feeding albumin with starch resulted in the slightly reduced ER compared to that from albumin only owing to its lower biocrude yield and HHV, which is consistent with previous literature [34, 35]. The presence of starch also negatively affected both oxygen and nitrogen contents of the biocrude. In general, the biocrude with higher heteroatom contents is more difficult to upgrade into transport fuels.

3.2 Characterization of main chemical species in biocrudes produced from albumin andbinary mixtures.

229

230 The biocrude samples produced from albumin and albumin/starch mixture were analyzed by 231 FTICR MS ESI+, as basic nitrogen species are more efficiently ionized in this ionization 232 condition. The mass spectra of biocrudes contained thousands of different peaks and each of them 233 is related to a specific molecular formula. The spectra were then elaborated and the molecular 234 formulas were categorized into groups based on their heteroatoms using a petroleomic approach. 235 Fig.1 depicts the main class components of biocrude products produced from the albumin and 236 albumin/starch mixture. The abundance of one or two nitrogen atom containing compounds (N1 237 and N2, respectively) was observed for the biocrude produced from the albumin HTL. 238 For the albumin and starch mixture HTL the N2 class was clearly predominant in the biocrude 239 followed by N1, O1N2 and O1N1 class components.

240



241



243

In order to determine the possible structures of the biocrude components the abundance plots of double bonds equivalents DBE (the sum of rings and double bonds) versus carbon number (Cn) for the main heteroatom classes are depicted in Fig. 2. In the case of the biocrude obtained from albumin, the main N1 class distribution corresponded at DBE= 4 and Cn=17-20; DBE=8 and
Cn=20-23. Concerning N2 class, a broad distribution of species ranging from Cn=20-26 and
DBE=7 was observed. The N1 species could be tentatively assigned to aniline and pyridine
derivatives with a single ring (DBE=4) or functionalized with a benzene moiety (DBE=8). The
N2 class could be assigned to condensed pyrazine or pyrimidine derivatives (DBE=7).

Generally, NAs are generated through Maillard reaction; however, their formation was also observed when amino acids were treated solely [39, 40]. For instance, the degradation of ¹³Clabelled serine in the absence of any sugars led to the generation of different α-aminocarbonyl intermediates that are common precursors to pyrrols, pyrazines and pyridines via Knorr pyrrole synthesis [39]. The other mechanism for the formation of pyrazines can also occur through the intra- and intermolecular cyclization of amino acids followed by dehydration reactions [12, 34, 38].

259 The biocrude from the albumin/starch mixture contained mainly N2 homologues series of 260 compounds with the same DBE values of 7 and 11 with Cn=16-26, indicating a higher degree of 261 unsaturation and functionalization. These species could be indicative for guinazoline and 262 naphthyridine derivatives. Based on the FTICR MS analysis, it was shown that NAs, mainly N1 263 and N2 containing compounds, can be produced from the albumin without any carbohydrate 264 source; however, in the presence of starch, the formation of N2 class components with higher 265 DBE values is enhanced via reactions of amines with oxygenated derivatives (aldehydes, ketones) 266 generated by sugars. Some hypothesized structures based on the elemental analysis and DBE 267 values are illustrated in Fig. S1.

268



270

Fig. 2. C_n vs DBE plots for main classes found in biocrudes from a) N1 classes for albumin; b)
N2 classes for albumin; c) N1 class for albumin/starch; d) N2 classes for albumin/starch.

273

The predominant nitrogen species in the biocrude produced from albumin/tripalmitin mixture were mainly in the form of fatty acid amides (FAAs). Therefore, GC-FID was employed for the quantification of FAAs. Under hydrothermal conditions, tripalmitin underwent hydrolytic cleavage to form palmitic acid, corresponding to 72 wt.% of the biocrude. The amidation reaction took place between palmitic acid and ammonia and amines, resulting in the formation of about 4 wt.%FAAs (Fig. S2).

These results confirm that Maillard reaction is the main reaction pathway that takes place in the binary mixture of protein and carbohydrate, thus generating NAs, while the presence of lipids shifts the reaction pathway towards the formation of FAs and FAAs, which are expected to be less resistant to upgrading process compared to the NAs.

285 **3.3 HTL of ternary mixture**

286

287 In order to verify the influence of the lipid content on the nitrogen distribution in the biocrudes, 288 comparative experiments were carried out with the ternary mixture containing two different lipid 289 contents of 20 wt.% and 33 wt.%. Two ternary mixtures of albumin/starch/triolein with the weight 290 ratios of 1:1:0.5 and 1:1:1 were processed under identical HTL conditions. Triolein was selected 291 herein in replace of tripalmitin, as oleic acid is the main component of waste cooking oils [41]. 292 Table 2 represents the yields, HHV and ER values of biocrudes.

293

_

294 Table 2. Biocrude yields, elemental analysis, HHV and ER of biocrudes from HTL of ternary mixtures

	Albumin/Starch/Triolein 20	Albumin/Starch/Triolein 33
	wt.%	wt.%
Biocrude yield, %	25.0±2.9	36.1±1.5
Biocrude yield without triolein contribution, %	7.1	6.4
Solid yield, %	2.1	1.0
	Element analysis, %	
C%	74.0±0.7	74.5±0.6
N %	3.8±0.1	3.0±0.05
Н, %	10.5±0.4	11.1±0.2
O, %	11.4	11.4
S, %	0.3±0.04	0.1
HHV, MJ/kg	36.7	38.0

ER, % 37.17 51.2

295

296 As expected, the higher biocrude yield and lower nitrogen content were obtained with the higher 297 lipid content mainly due to the direct contribution of fatty acids to the biocrude. As result of this, 298 the ER was substantially improved as well. In this case, the synergistic effect of the addition of 299 lipid to albumin on the biocrude yield, previously reported for the binary mixture, was not 300 prominent. In fact, most of the biocrude is derived from the hydrolysis of the lipid fraction with 301 formation of oleic acid. If it is subtracted, the yields decrease to 6-7%, which are similar to the 302 cases of the binary albumin/starch or albumin alone tests. This can be explained by the occurrence 303 of hydrolysis of three macromolecular fractions, albumin, starch and triolein, at different 304 temperatures. In fact, starch is rapidly hydrolyzed to produce oligosaccharides and 305 monosaccharides at 180-250 °C [42], while the hydrolysis of lipids, for instance, soybean, 306 linseed, and coconut oils proceeds at 260–280 °C [43]. Thus, starch-derived products are present 307 in the reaction mixture since the first stages of the process and they limit their effect on the 308 formation of amides, that would have increased the biocrude yield due to their solubility in the 309 oil phase.

Deep characterization of the products was thus carried out. Given the poor resolution of the GC-MS analysis for polar nitrogen compounds, FTICR ESI+ was utilized to characterize the FAAs and NAs species of the biocrudes. It should be pointed out that by ESI+ the presence of FAs was not detected, as they are selectively ionized in the ESI- mode.

In order to determine the effect of the lipid content on the formation of the NAs and FAAs in the biocrudes produced from the ternary mixtures, the relative contents of FAAs and NAs were determined by FTICR-MS, taking into account their relative intensities in the mass spectra normalized with respect to that of an internal standard (caffeine) and also considering dilution factors. The distribution of FAAs in the biocrude products from ternary mixtures is illustrated in Fig.3. As it can be seen from Fig. 3, the biocrude produced from the 33% lipid containing ternary mixture contained more FAAs compared to that from the mixture with the 20% lipid content. The
ratio of the total relative peak intensity of FAAs in the biocrudes from the mixtures with 33 %
and 20 % lipid contents was estimated as 1.51, thus indicating that the high-lipid mixture produced
a higher amount of FAAs.

324



325

Fig. 3. FTICR ESI+ based distribution of main FAAs species as protonated ions for ternary
 mixture with 20 % and 33 % triolein contents.

328

329 For the semi-quantitative analysis of NAs, the mass spectrum peaks that are relevant to the 330 nitrogen-containing heterocyclic and aromatic compounds were considered by elaborating the 331 compounds only with a DBE value higher than 4. Fig.4 illustrates the abundance plots of DBE 332 versus Cn for the NAs. It can be seen that the NAs in the biocrudes from the ternary mixtures 333 were present in forms of N1, N2 and O1N1 class components at DBE=4-9 and Cn=8-20 and their 334 relative abundances were more significant for the mixture with the 20% of lipid content. In fact, 335 the ratio of the total peak intensity of NAs between 20% and 33% triolein containing mixtures 336 was 0.47 (Total Peak area of NAs for 20% triolein/Total Peak area for 33% triolein), thereby 337 confirming the lower content of the NAs in the biocrude obtained from the mixture with the higher lipid content. These results confirm the positive impact of the lipids on the promotion of amidation
reactions, forming FAAs and inhibiting the formation of NAs, the latter being more recalcitrant
for the hydrotreatment process.

341





Fig. 4. Cn vs DBE plots for aromatic nitrogen classes (DBE >4) for biocrudes from ternary
mixtures: a) 20% and b) 33% triolein content.

345

346 **3.4 Hydrotreatment of biocrude produced from organic food waste**

347

348 In order to validate the hypotheses elaborated using model compounds, the composition of the 349 biocrudes produced from the ternary mixture of albumin/cellulose/tripalmitin and from a real 350 municipal organic food waste were characterized by FTICR MS with APPI and compared 351 considering the number of common products and their intensities. According to the results, only 352 about 44% common products were present in both model and biomass waste biocrudes [44]. 353 Even though the employment of the ternary mixtures can help to understand the interaction 354 mechanisms that occur under HTL conditions and optimize the process, they may not be fully 355 reliable for the next upgrading stage via hydrodeoxygenation and hydrodenitrogenation.

Therefore, in order to accurately evaluate the hydrodenitrogenation reactivity under relevant hydrotreatment conditions, and particular the effect of FAAs and NAs content, the two biocrude samples were produced starting from real food wastes with the different lipid contents. A standard sorted household organic food waste was selected with the dry matter of 35%, carbon content of 42% and lipid content of 20%. The comparison was made with a supermarket food packaging and expiring waste with the dry matter of 50%, 55% of carbon content, and 31% of lipid content (due to the presence of meat, cured meats and dairy products packaging).

363 The HTL of sorted household and supermarket wastes resulted in the biocrude yields of 35% and 364 58%, respectively. Afterwards, the HTL oils were upgraded by conventional hydrocracking 365 process that is typically employed for the refinery upgrading of fossil heavy oil, such as the 366 vacuum distillation residue, also referred to as the bottom-of-the-barrel [29]. The yields of 367 upgraded oils were 92 and 93 % for the biocrudes obtained from organic waste with 20 and 33 % 368 lipid contents, respectively. As shown in Fig. 5, the upgrading of the biocrude from the 33% lipid 369 containing food waste resulted in the increased yields of diesel (170-350 °C) fraction as expense 370 of vacuum gas oil VGO (350-500 °C) and heavy oil (+500 °C) fractions compared to those from 371 the food waste with the 20% lipid content.



Fig. 5. Distillation cuts for upgraded oils from: a) household food waste; b) supermarket food
waste.

372

376 The characterization of the biocrudes before and after the upgrading process is given in Table 3377 [45]. It can be seen that the upgrading of the household waste HTL oil resulted in the removal of

around 75% of oxygen, 64% of sulphur and 60% of nitrogen and, as a result of the removal of
heteroatoms, the HHV increased from 34.8 to 43. MJ/kg, similar to that of fossil fuels. In the case
of the household waste HTL oil, the main difference was the denitrification efficiency that
increased up to 71%.

382

383

Table 3. Element analysis of biocrudes produced form the organic food waste [45].

HTL oil from	Upgraded HTL	HTL oil from	Upgraded HTL
sorted	oil	supermarket food	oil
household		waste	
organic waste			
	Element analysis,	, %	
72.0	82.0	72.5	83.0
9.0	12.3	9.3	12.2
12.5	3.1	12.3	3.1
5.8	2.3	5.2	1.5
0.75	0.27	0.68	0.27
34.8	43.3	35.6	44.9
	HTL oil from sorted household organic waste 72.0 9.0 12.5 5.8 0.75 34.8	HTL oil fromUpgraded HTLsortedoilhouseholdorganic waste72.082.09.012.312.53.15.82.30.750.2734.843.3	HTL oil fromUpgraded HTLHTL oil fromsortedoilsupermarket foodhouseholdwasteorganic wasteT2.082.072.59.012.39.312.53.112.35.82.35.20.750.270.6834.843.335.6

³⁸⁴

The upgraded oil produced from household waste was analyzed by GC-MS to determine the type of residual nitrogen containing species (Fig.S4). The native biocrude was found to contain FAs, FAAs and some NAs. After the hydrotreatment, the main peaks were related to the linear saturated hydrocarbons with the carbon numbers ranging from C14 to C30 were detected; however, the elemental analysis clearly showed that nitrogen was still present in the upgraded oil.

The nitrogen compounds that are not detectable by GC-MS due to the low volatility and high polarity, were additionally characterized by FTICR MS with the APPI (Atmospheric pressure photoionization) in positive ion mode (Fig. 6). The classes were determined and compared for the biocrudes and the upgraded oils, normalizing the abundances according to the elemental analysis

394 results. The class distribution confirmed the efficient oxygen removal, as only trace amounts of 395 compounds related to the O1 and O1Nx classes (FAAs) were detected. On the other hand, the 396 amount of NAs belonging to N1, N2, N3 classes, decreased slightly, thereby confirming that the 397 NAs are more difficult to remove by the hydrotreatment process. Similar results were obtained 398 with the supermarket food waste upgraded oil, confirming the NAs belonging to N1, N2, N3 399 classes are the most recalcitrant, under the given hydrocracking conditions. In this case, the 400 relative abundance of the Nx species was lower than that from the oil with the lower lipid content 401 (even if by a semi-quantitative comparison, due to complexity of the mixture), thus confirming 402 the effect of lipids on the distribution of nitrogenous products, which are shifted more towards 403 the FAAs components. This is in fully agreement with the lower nitrogen content measured with 404 the elemental analysis.

Based on these results, blending of the protein-rich biomass with acyl donor species, such as oleaginous biomass, microalgae, waste cooking oils and fat fractions from animal waste, can be one of the most optimal solutions to inhibit the Maillard reactions and promote the formation of nitrogen compounds with the higher HDN reactivity, thereby producing the biocrudes with specifications that can meet the requirements of the existing refinery facilities.





411

412

Fig. 6. Main molecular classes distribution of HTL oil and upgraded oil.

The relative abundances of the classes have been normalized according to the elemental analysis
results for a) household food waste; b) supermarket food waste [45].

416 **4. Conclusion**

417 HTL tests carried out on model molecules have given general indications that interactions 418 between proteins and lipids lead to an improved biocrude yield and a lower content of nitrogen-419 containing aromatic compounds, compared to that observed with protein and carbohydrate 420 mixtures. The presence of lipids in the HTL feedstock favors the formation of fatty acid amides, 421 which can be converted into hydrocarbons, thereby resulting in the more efficient removal of 422 nitrogen in the biocrude upgrading process. These results suggest that HTL feedstock should be 423 supplemented with a lipids-rich waste biomass to produce a biocrude more suitable for upgrading 424 by conventional hydrotreatment processes. 425 Acknowledgements 426 We would like to acknowledge the financial support from SINCHEM Grant. SINCHEM is a Joint 427 Doctorate program under the Erasmus Mundus Action 1 (framework agreement N° 2013-0037) 428 of the European Union. 429 430 Fundings 431 This work was supported by the SINCHEM Grant [framework agreement N° 2013-0037]. 432 433 **References:** 434 1. Aierzhati, A., et al., *Experimental and model enhancement of food waste* 435 hydrothermal liquefaction with combined effects of biochemical composition and reaction conditions. Bioresour Technol, 2019. 284: p. 139-147. 436 437 Scherhaufer, S., et al., Environmental impacts of food waste in Europe. Waste 2.

- 438 Management, 2018. **77**: p. 98-113.
- Aierzhati, A., et al., Development of a mobile, pilot scale hydrothermal *liquefaction reactor: Food waste conversion product analysis and techno- economic assessment.* Energy Conversion and Management: X, 2021. 10: p.
 100076.
- 443 4. Ng, H.S., et al., *Recent advances on the sustainable approaches for conversion*444 *and reutilization of food wastes to valuable bioproducts.* Bioresource
 445 Technology, 2020. **302**: p. 122889.

446	5.	Fujii, H. and Y. Kondo, Decomposition analysis of food waste management with
447		explicit consideration of priority of alternative management options and its
448		application to the Japanese food industry from 2008 to 2015. Journal of Cleaner
449		Production, 2018. 188 : p. 568-574.
450	6.	Agency, I.E., World Energy Outlook 2021. 2021.
451	7.	Déniel, M., et al., Modelling and Predictive Study of Hydrothermal
452		Liquefaction: Application to Food Processing Residues. Waste and Biomass
453		Valorization, 2017. 8(6): p. 2087-2107.
454	8.	Matayeva, A., S.R. Rasmussen, and P. Biller, Distribution of nutrients and
455		phosphorus recovery in hydrothermal liquefaction of waste streams. Biomass
456		and Bioenergy, 2022. 156 : p. 106323.
457	9.	Beims, R.F., et al., Hydrothermal liquefaction of biomass to fuels and value-
458		added chemicals: Products applications and challenges to develop large-scale
459		operations. Biomass and Bioenergy, 2020. 135: p. 105510.
460	10.	Katakojwala, R., et al., Hydrothermal liquefaction of biogenic municipal solid
461		waste under reduced H2 atmosphere in biorefinery format. Bioresource
462		Technology, 2020. 310 : p. 123369.
463	11.	Li, Y., et al., Correlations between the physicochemical properties of hydrochar
464		and specific components of waste lettuce: Influence of moisture, carbohydrates,
465		proteins and lipids. Bioresource Technology, 2019. 272: p. 482-488.
466	12.	Leng, L., et al., Nitrogen in bio-oil produced from hydrothermal liquefaction of
467		biomass: A review. Chemical Engineering Journal, 2020. 401: p. 126030.
468	13.	Furimsky, E. and F.E. Massoth, Hydrodenitrogenation of Petroleum. Catalysis
469		Reviews, 2005. 47 (3): p. 297-489.
470	14.	Bellussi, G., et al., The role of MoS2 nano-slabs in the protection of solid
471		cracking catalysts for the total conversion of heavy oils to good quality
472		distillates. Catalysis Science & Technology, 2013. 3(1): p. 176-182.
473	15.	Almarri, M., et al., Adsorptive Pretreatment of Light Cycle Oil and Its Effect on
474		Subsequent Hydrodesulfurization, in Production and Purification of Ultraclean
475		Transportation Fuels. 2011, American Chemical Society. p. 33-54.
476	16.	Borugadda, V.B., R. Chand, and A.K. Dalai, Screening suitable refinery
477		distillates for blending with HTL bio-crude and evaluating the co-processing
478		potential at petroleum refineries. Energy Conversion and Management, 2020.
479		222 : p. 113186.
480	17.	Zeuthen, P., K.G. Knudsen, and D.D. Whitehurst, Organic nitrogen compounds
481		in gas oil blends, their hydrotreated products and the importance to
482		hydrotreatment. Catalysis Today, 2001. 65(2): p. 307-314.
483	18.	Leng, L., et al., A review on pyrolysis of protein-rich biomass: Nitrogen
484		transformation. Bioresource Technology, 2020. 315: p. 123801.
485	19.	Matayeva, A., et al., Development of Upgraded Bio-Oil Via Liquefaction and
486		Pyrolysis, in Studies in Surface Science and Catalysis. 2019. p. 231-256.
487	20.	Fan, Y., et al., Hydrothermal liquefaction of protein-containing biomass: study
488		of model compounds for Maillard reactions. Biomass Conversion and
489	_ .	Biorefinery, 2018. 8 (4): p. 909-923.
490	21.	Matayeva, A., et al., Elucidation of reaction pathways of nitrogenous species by
491		hydrothermal liquefaction process of model compounds. Fuel, 2019. 240: p.
492		169-178.

493	22.	Matayeva, A., Development of innovative processes and catalysts for the
494		valorisation of bio-oil. 2019, Bologna University: Bologna.
495	23.	Fan, Y., et al., The influence of lipids on the fate of nitrogen during
496		hydrothermal liquefaction of protein-containing biomass. Journal of Analytical
497		and Applied Pyrolysis, 2020. 147.
498	24.	Chiaberge, S., et al., Bio-Oil from Waste: A Comprehensive Analytical Study by
499		Soft-Ionization FTICR Mass Spectrometry. Energy & Fuels, 2014. 28(3): p.
500		2019-2026.
501	25.	Kendrick, E., A Mass Scale Based on CH2 = 14.0000 for High Resolution Mass
502		Spectrometry of Organic Compounds. Analytical Chemistry, 1963. 35(13): p.
503		2146-2154.
504	26.	Chiaberge, S., et al., Investigation of Asphaltene Chemical Structural
505		Modification Induced by Thermal Treatments. Energy & Fuels, 2009. 23(9): p.
506		4486-4495.
507	27.	Panda, S.K., J.T. Andersson, and W. Schrader, <i>Characterization of</i>
508		Supercomplex Crude Oil Mixtures: What Is Really in There? Angewandte
509		Chemie International Edition, 2009, 48 (10); p. 1788-1791.
510	28.	Conte. A., et al., <i>Dealing with complex contamination: A novel approach with a</i>
511		combined bio-phytoremediation strategy and effective analytical techniques.
512		Journal of Environmental Management, 2021, 288 : p. 112381.
513	29.	Bellussi, G., et al., Hydroconversion of heavy residues in slurry reactors:
514	_>.	Developments and perspectives. Journal of Catalysis. 2013. 308 : p. 189-200.
515	30.	Dote, Y., et al., <i>Studies on the direct liquefaction of protein-contained biomass:</i>
516	201	<i>The distribution of nitrogen in the products.</i> Biomass and Bioenergy, 1996.
517		11 (6): n 491-498
518	31.	Merli, G., et al., Acetic acid bioproduction: The technological innovation
519	011	<i>change</i> Science of The Total Environment, 2021, 798 : p. 149292
520	32.	Déniel, M., et al., Hydrothermal liquefaction of blackcurrant pomace and model
521	021	molecules: understanding of reaction mechanisms. Sustainable Energy & Fuels
522		2017 1(3): p. 555-582.
523	33.	Deluca, P.P., L. Lachman, and H.G. Schroeder, <i>Physical-chemical Properties of</i>
524	001	Substituted Amides in Aqueous Solution and Evaluation of Their Potential Use
525		as Solubilizing Agents Journal of Pharmaceutical Sciences, 1973, 62 (8): p
526		1320-1327
520	34	Yang W et al Understanding low-lipid aloge hydrothermal liquefaction
528	51.	characteristics and pathways through hydrothermal liquefaction of algal major
520		components: Crude polysaccharides crude proteins and their hinary mixtures
530		Bioresource Technology 2015 196 : n 99-108
531	35	Biller P and A B Ross Potential yields and properties of oil from the
532	55.	hydrothermal liquefaction of microalgae with different biochemical content
533		Bioresource Technology 2011 102 (1): p 215-225
534	36	Vang I et al Hydrothermal liquefaction of biomass model components for
535	50.	nroduct yield prediction and reaction pathways exploration Applied Epergy
536		2018 228 n 1618-1628
530	37	He S et al Hydrothermal liquefaction of low linid alage Nannochloronsis sn
539	57.	and Saraassum sn : Effect of feedstock composition and temperature Science of
520		The Total Environment 2020 712 n 125677
559		The Total Environment, 2020. 112 . p. 1550/7.

540	38.	Zhang, C., et al., Enhancing the performance of Co-hydrothermal liquefaction
541		for mixed algae strains by the Maillard reaction. Green Chemistry, 2016. 18(8):
542		p. 2542-2553.
543	39.	Yaylayan, V.A. and A. Keyhani, Elucidation of the mechanism of pyrrole
544		formation during thermal degradation of 13C-labeled l-serines. Food
545		Chemistry, 2001. 74 (1): p. 1-9.
546	40.	Yaylayan, V.A., A. Keyhani, and A. Wnorowski, Formation of Sugar-Specific
547		Reactive Intermediates from 13C-Labeled l-Serines. Journal of Agricultural and
548		Food Chemistry, 2000. 48 (3): p. 636-641.
549	41.	Mohammed, S.T., et al., Evaluation and optimal design of a high stability
550		hydrothermal deoxygenation process for production of green diesel fuel via
551		deoxygenation of waste cooking oil. Process Safety and Environmental
552		Protection, 2022. 159: p. 489-499.
553	42.	Changi, S.M., et al., Hydrothermal Reactions of Biomolecules Relevant for
554		Microalgae Liquefaction. Industrial & Engineering Chemistry Research, 2015.
555		54 (47): p. 11733-11758.
556	43.	Holliday, R.L., J.W. King, and G.R. List, Hydrolysis of Vegetable Oils in Sub-
557		and Supercritical Water. Industrial & Engineering Chemistry Research, 1997.
558		36 (3): p. 932-935.
559	44.	Croce, A., et al., A Model Study to Unravel the Complexity of Bio-Oil from
560		Organic Wastes. ChemSusChem, 2017. 10(1): p. 171-181.
561	45.	Bianchi, D., A. Bosetti, and S. Chiaberge, Production of Advanced Biofuels by
562		Hydrothermal Liquefaction of Sorted Organic Domestic Waste. DGMK
563		Tagungsbericht, 2019. 2019: p. 49-59.
564		