



ALMA MATER STUDIORUM
UNIVERSITÀ DI BOLOGNA

ARCHIVIO ISTITUZIONALE DELLA RICERCA

Alma Mater Studiorum Università di Bologna Archivio istituzionale della ricerca

Survey on the presence of *Leishmania* sp. in peridomestic rodents from the Emilia-Romagna Region (North-Eastern Italy)

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Magri A., Galuppi R., Fioravanti M., Caffara M. (2023). Survey on the presence of *Leishmania* sp. in peridomestic rodents from the Emilia-Romagna Region (North-Eastern Italy). *VETERINARY RESEARCH COMMUNICATIONS*, 47(1), 291-296 [10.1007/s11259-022-09925-4].

Availability:

This version is available at: <https://hdl.handle.net/11585/906128> since: 2024-01-29

Published:

DOI: <http://doi.org/10.1007/s11259-022-09925-4>

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>).
When citing, please refer to the published version.

(Article begins on next page)

This is the peer reviewed version of the following article:

Magri A., Galuppi R., Fioravanti ML., Caffara M. (2022). Survey on the presence of *Leishmania* sp. in peridomestic rodents from Emilia-Romagna region (North-Eastern Italy). Veterinary Research Communication. doi:10.1007/s11259-022-09925-4

which has been published in final form at:

<https://doi.org/10.1007/s11259-022-09925-4>

This article may be used for non-commercial purposes in accordance with Springer Terms and Conditions for Use of Self-Archived Versions.

1 **Survey on the presence of *Leishmania* sp. in peridomestic rodents from the**
2 **Emilia-Romagna Region (North-Eastern Italy)**
3 **Alice Magri *, Roberta Galuppi, Marialetizia Fioravanti and Monica Caffara**

4 Department of Veterinary Medical Sciences – *Alma Mater Studiorum* University of Bologna,
5 Ozzano Emilia (Bologna, Italy)

6 *Correspondence: alice.magri3@unibo.it

7 RG: roberta.galuppi@unibo.it

8 MF: marialeti.fioravanti@unibo.it

9 MC: monica.caffara@unibo.it

10

11

12 **Abstract**

13 Leishmaniasis is a neglected vector-borne parasitic disease caused in Italy only by the
14 species *Leishmania infantum* of the *Leishmania donovani* complex, which is the causative agent
15 of the zoonotic visceral leishmaniasis (VL), and the sporadic cutaneous leishmaniasis (CL)
16 in humans, and of the canine leishmaniasis (CanL). The disease is considered endemic in
17 southern, central, and insular Italian regions and recognizes phlebotomine sand flies as
18 vector and dogs as main reservoir. However, a specific north-eastern region, namely Emilia-
19 Romagna, always showed a peculiar epidemiological situation when compared to the other
20 northern Italian regions and recent studies are indeed questioning the role of dog as main
21 reservoir of *L. infantum*. Due to their synanthropic relationship with humans, rodents have
22 been tested for *Leishmania* spp. in several European countries. The aim of this study was to
23 assess the presence of *Leishmania* spp. in peridomestic rodents in the Emilia-Romagna

24 Region. The study was carried out on 136 peridomestic rodents collected by professional
25 rodent control services: 47 brown rats (*Rattus norvegicus*), 39 black rats (*Rattus rattus*) and 50
26 mice (*Mus musculus*). Specimens of earlobe skin, spleen, liver and prescapular lymph nodes
27 were tested with a real-time PCR. Fifteen (11 %) rodents, tested positive for *L. infantum*.
28 Positivity was obtained from different target organs; notably 33% of the rodents tested
29 positive in earlobe skin samples. These findings revealed the presence of *Leishmania* spp. in
30 peridomestic rodents of the Emilia-Romagna Region, also in two species never tested before
31 in Italy, namely *R. norvegicus* and *M. musculus*.

32 **Keywords:** Leishmaniasis, Italy, *Mus musculus*, *Rattus norvegicus*, *Rattus rattus*

33

34 **Background**

35 Leishmaniasis is a neglected vector-borne parasitic disease endemic in southwestern
36 Europe. With reference to Italy, *Leishmania infantum* of the *Leishmania donovani* complex is
37 the only species responsible for visceral leishmaniasis (VL), for sporadic cutaneous
38 leishmaniasis (CL) in humans and for canine leishmaniasis (CanL) (Gramiccia and Gradoni
39 2005; Rugna et al. 2018). The parasite is transmitted by phlebotomine sand flies, and in Italy
40 dogs have long been recognized as the major reservoir in southern, central and insular
41 regions, where the disease is considered endemic. Among the northern Italian regions,
42 Emilia-Romagna has always had a different epidemiological scenario: CL has been widely
43 reported since 1934, and between 1950-1958 up to 2,670 cases were diagnosed in the
44 province of Forlì (Pampiglione 1975). In contrast, until the early 1970's, in this region VL
45 appeared sporadically, with only 4 autochthonous cases observed, one in the province of
46 Bologna and 3 in the province of Forlì. In the same period and within the same area, no
47 ascertained autochthonous cases of CanL were reported (Pampiglione 1975). In 1971-1972,
48 in two municipalities of Bologna province located in a foothill area a dramatic outbreak of
49 VL was reported, involving 60 patients with a lethality of 21.7% (Pampiglione 1975). Since
50 then, the geographic distribution of human and canine leishmaniasis has notably increased
51 and the disease spread even in other regions of northern Italy, where many autochthonous
52 cases of VL, CL and CanL have been reported (Gaspari et al. 2017; Mendoza-Roldan et al.
53 2020). This epidemiological change may be due to environmental issues, occurrence of
54 competent insect vectors and movement of infected dogs from endemic areas (Santi et al.
55 2014). However, such changes might not be sufficient to explain the recurrent VL and CL

56 foci recorded in Emilia-Romagna Region (Gaspari et al. 2017), especially considering that
57 molecular studies carried out on strains isolated from autochthonous cases of VL are
58 questioning the role of dogs as reservoirs of *L. infantum* in this region, as earlier suggested
59 (Pampiglione 1975; Rugna et al. 2018).

60 The role of wildlife has long been recognized as crucial in the transmission and maintenance
61 of zoonotic agents and several sylvatic species are known to be susceptible to leishmaniasis.
62 Considering their synanthropic relationship with humans and their abundance the role of
63 rodents as possible leishmaniasis reservoirs has been questioned in different European
64 countries (Alcover et al. 2021). Several studies established the presence of *L. infantum* in
65 these hosts in Greece (Papadogiannakis et al. 2009; Tsakmakidis et al. 2017), Portugal
66 (Helhazar et al. 2013) and Spain (Navea-Pérez et al. 2015; Galán-Puchades et al. 2019;
67 Martín-Sánchez et al. 2020).

68 In Italy, a study performed in Sicily detected *L. infantum* by PCR in 45% of black rats, even
69 if in this region the role of the dog as reservoir has been well established (Di Bella et al.
70 2003). However, a study performed in Montecristo Island (Tuscany), revealed the presence
71 of *L. infantum* in the 15.5% of rodents examined, even in the absence of domestic carnivores
72 (Zanet et al. 2014).

73 The aim of this survey was to assess the presence of *Leishmania* spp. in peridomestic rodents
74 collected in the Emilia-Romagna Region, Italy.

75

76

77

78 **Materials and Methods**

79 From June 2019 to June 2021, 136 peridomestic rodent carcasses were sampled during pest
80 control programs from the provinces of Ferrara, Forlì-Cesena and Ravenna (Emilia
81 Romagna) (figure 1): 47 brown rats, *Rattus norvegicus* (20 females and 27 males), 39 black
82 rats, *R. rattus* (21 females and 18 males), 50 mice, *Mus musculus* (22 females and 28 males)
83 were collected from the territory by professional rodent control services and stored at -20
84 °C before processing.

85 The entire carcass was examined; species, sex and age classes were identified by
86 morphological and metrical evaluation (CDC). Necropsies and samples collection were
87 performed with sterile surgical instruments and when possible, according to the state of the
88 carcasses, 25 mg of tissue were collected from earlobe skin, prescapular lymph node and
89 liver, and 10 mg from the spleen (Helhazar et al. 2013). Due to the corruption of the remains,
90 lymph nodes were not collected from 16 subjects. Samples were placed in sterile 1.5 ml tubes
91 and stored at -20 °C.

92 DNA extraction was performed with PureLink® Genomic DNA Mini Kit (Invitrogen,
93 ThermoFisher Scientific) following the manufacturer's instructions. For DNA amplification
94 a real-time PCR protocol was performed targeting a 71-bp region of the minicircle
95 kinetoplast DNA using primer pair Leish71Up (5'-
96 CCAAACCTTTTCTGGTCCTYCGGGTAG-3') and Leish71Do (5'-
97 TGAACGGGATTTCTGCACCCATTTTTC -3') (Tsakmakidis et al. 2017). Reactions were
98 carried out in a total volume of 20 µL with 10 µL of PowerUP™ SYBR™ Green master mix
99 (2X), 0.3 µM of each primer and 2 µL of DNA. The amplification was performed in

100 StepOnePlus Real-Time PCR System (Applied Biosystems) and the thermal cycling profile
101 was as follows: 95 °C for 5 min, followed by 40 cycles at 95 °C for 5 sec., 60 °C for 30 sec. At
102 the end of the amplification, a melting curve analysis was performed from 60 °C to 95 °C,
103 with a slope of 0.3 °C to monitor primer dimers of non-specific product formation. Each
104 sample was amplified in triplicate, the average temperature of melting (T_m) observed was
105 79.39 ± 0.15 °C and the average standard deviation observed in cts was 0.65. The standard
106 curve was created with serial dilution of *L. infantum* DNA ranging from 10,000 to 0.1
107 parasites per reaction. Each reaction was carried out by three replicates per dilution, in three
108 independent experiments. The ct value cut-off was settled at mean ct value of 39.3 which
109 corresponds to 1 parasite per mL of the original parasite suspension.

110 As a positive control the reference strain *L. infantum* MHOM/TN/80/IPT1, kindly provided
111 by the Unit of Clinical Microbiology, Regional Reference Centre for Microbiological
112 Emergencies (CRREM), St. Orsola-Malpighi University Hospital, Bologna, Italy, was used.
113 Confidence intervals were calculated by R Studio (RStudio Team 2020).

114 **Results and Discussion**

115 Out of 136 subjects examined, 15 (11 %; 95% CI=5.7-16.3) were positive for *Leishmania* spp.
116 In particular, 10.6% (95% CI=1.8-19.4) of brown rats, 12.8% (95% CI=2.5-23.7) of black rats
117 and 10% (95% CI=1.7-18.3) of mice (Table 1). Of the five positive mice, three tested positive
118 in two target organs - spleen and earlobe skin or spleen and liver or spleen and lymph nodes
119 - the remaining two subjects tested positive only in lymph nodes or liver, respectively. The
120 geographical distribution of the positive subjects appears homogeneous between the
121 sampled sites (figure 1).

122 The present survey assessed the presence of *Leishmania* spp. in synanthropic rodents of the
123 Emilia-Romagna Region. The conditions settled by the WHO (2010) for a species to be
124 recognized as reservoir is the prevalence of infection > 20% and the availability of the
125 parasite in blood and skin in sufficient amount to be ingested by a sand fly. In the
126 Mediterranean area such conditions were globally assessed only for *M. musculus*, while *R.*
127 *norvegicus* and *R. rattus* showed lower prevalence of infections (16.4% and 9.9%,
128 respectively) (Alcover et al. 2021).

129 The prevalence values observed in the current study are below the average found in
130 Portugal or Spain (Barcelona) where the 33.3% of examined rodents (*M. musculus* and *R.*
131 *norvegicus*, Helhazar et al. 2013; *R. norvegicus*, Galán-Puchades et al. 2019) resulted positive,
132 or the one reported in Granada (Spain) in mice (88.9%) (Martín-Sánchez et al. 2020) or in
133 different rodent species (*R. rattus*, *M. musculus* and *Apodemus sylvaticus*) (27%) by Navea-
134 Pérez et al. (2015), whilst it is higher than the prevalence observed in brown rats (5.9%) in
135 Greece (Papadogiannakis et al. 2009). Further studies performed in Greece by Tsakmakidis
136 et al. (2017) on spleen of *R. norvegicus*, *R. rattus* and *M. musculus* revealed a prevalence of
137 19.58% comparable to the one herein reported. The majority of the studies evaluated the
138 presence of the parasite in more than one target organ including skin, liver, spleen and blood
139 (Helhazar et al. 2013; Martín-Sánchez et al. 2020; Navea-Pérez et al. 2015; Tsakmakidis et al.
140 2017) while few studies examined only the spleen as target organ (Galán-Puchades et al.
141 2019; Papadogiannakis et al. 2009). Testing more than one target tissue allow to increase the
142 possibility to detect *Leishmania* spp. as observed also in our study. Three *M. musculus* here
143 examined showed the presence of the parasite DNA in two different target organs (spleen

144 + lymph nodes and spleen + liver). Although the spleen is traditionally recognized as
145 *Leishmania* spp. target organ for PCR in different animal species (Papadogiannakis et al.
146 2009), our results showed the presence of *Leishmania* spp. in the earlobe skin samples from
147 33 % of the positive rodents pointing out that this tissue should be also considered. In fact,
148 wild animals are frequently collected in decomposition state and the putrefaction of the
149 target tissues, like visceral organs, may affect the integrity of the kinetoplast DNA (Múnoz-
150 Madrid et al. 2013).

151 In Italy, the role of black rats in the transmission of *L. infantum* has long been investigated,
152 starting from surveys performed in Tuscany in the 1980's (Pozio et al. 1985). Further studies
153 showed that *Phlebotomus perniciosus* and *P. perfiliewi* are attracted to *R. rattus* and that these
154 sand fly species become infected when they feed on black rats experimentally infected with
155 *L. infantum* (Pozio et al. 1985). More recent study carried out in Calabria (Italy) by Di Bella
156 et al. (2003) showed 45% positivity in the spleen of 22 *R. rattus* although in this region the
157 role of dogs as reservoirs has long been established. Zanet et al. (2014), reported 15.5%
158 prevalence in black rats examined in the Montecristo Island (Tuscany, Italy) where *L.*
159 *infantum* was recorded even in absence of domestic carnivore hosts. This value is similar to
160 the one (12.8%) obtained in the same host in our study, that moreover provided also data
161 on *R. norvegicus* and *M. musculus* (10.6% and 10% respectively) species not previously tested
162 for *L. infantum* in Italy.

163 Leishmaniasis in Emilia-Romagna has a peculiar epidemiological scenario compared to the
164 other Northern Italian regions. Recently Rugna et al. (2018), by Multilocus Microsatellite
165 Typing (MLMT) detected differences between *Leishmania* strains from men and sand flies to

166 the ones from dogs. The MLMT profiles showed all canine samples belonged to one group
167 genetically related to Mediterranean MON-1 strain and similar to the VL samples from other
168 Italian regions, while all but one VL Emilia-Romagna case, and the isolates from sand fly
169 fell into a different group. Therefore, in this region the co-circulation of two distinct groups
170 of *L. infantum* seems to occur, and the VL in humans could have different cycles involving
171 *P. perfiliewi* as a vector (Rugna et al. 2018; Calzolari et al. 2019) and might include other
172 vertebrates, besides dogs, as reservoirs.

173 In two of the three provinces studied, Ravenna and Forlì-Cesena, foci of VL, usually located
174 in hilly areas, were historically described. The rodent samples analyzed were collected in
175 an area not higher than 50 m above sea level, where the density of phlebotomines is scant
176 and, according to leishmaniasis regional control plan, in 2020 only CL cases have been
177 reported (Santi et al. 2021). Further research should focus on studying which strains
178 circulate in this area.

179 Also notable is the presence of a positive brown rat in the province of Ferrara, where
180 autochthonous cases of leishmaniasis in both dogs and humans have never been recorded:
181 the specimen was collected in a locality on the border between the provinces of Ferrara and
182 Ravenna where the phlebotomine population is recorded as being moderate (Santi et al.
183 2021). This finding, considering the consistent increase in geographical distribution of the
184 disease and its vector, will require further investigation.

185 *L. infantum* is a vector-borne parasite and in its epidemiology many mammal species are
186 involved, hence identifying which one may act as a reservoir in the Emilia-Romagna Region
187 is an ambitious task due to the presence of different environments i.e. hilly or flatlands and

188 different distribution of sylvatic and peridomestic animals, which may possibly be involved
189 in the parasite cycle. Even if the presence of the parasite in mammalian hosts is crucial, in
190 order to fully understand his meaning as main reservoir or epiphenomena it should be
191 associated with studies on the blood preferences of the phlebotomine vector.

192 The total prevalence observed in the present study (11%), despite being lower to the one
193 required from WHO (2010) to establish a role of reservoir is comparable to the
194 Mediterranean's one. As reported in previous studies, this value is far from being trivial:
195 considering their close relationship with humans, their ability to colonize new environments
196 and their impact on human health, rodents should not be neglected for their potential role
197 in the transmission of Leishmaniasis, especially in urban areas (Alcover et al. 2021).

198 Although these preliminary findings are not sufficient to prove the role of peridomestic
199 rodents as reservoirs of *L. infantum*, they nevertheless indicate the opportunity to further
200 investigate their possible role in the epidemiology of different strains of *L. infantum*
201 circulating in the Emilia-Romagna Region.

202

203 **List of abbreviations**

204 VL = Visceral Leishmaniasis

205 CanL = Canine Leishmaniasis

206 CL = Cutaneous Leishmaniasis

207

208 **Declarations:**

209 *-Ethics approval and consent to participate*

210 No ethical approval is officially required since the rodents examined had been subjected to
211 pest control are considered pest species.

212 - *Consent for publication*

213 Not applicable

214 - *Availability of data and materials*

215 The datasets generated during and/or analyzed during the current study are available from
216 the corresponding author on reasonable request

217 - *Competing interests*

218 The authors declare that they have no competing interests

219 - *Funding*

220 This study received no external funding

221 - *Authors' contributions*

222 MF and RG conceived the study. AM performed field work. AM and MC performed
223 laboratory work and analyzed data. AM and MC wrote the first draft of the manuscript. MF
224 and RG reviewed the manuscript. All authors read and approved the final manuscript

225 - *Acknowledgements*

226 Authors would like to thank social cooperative For.B and dr. Guglielmo Pampiglione for
227 kindly providing rodent carcasses from the provinces of Ravenna and Forlì-Cesena of
228 Emilia-Romagna Region.

229

230

231

232 **References**

- 233 Alcover MM, Riera MC and Fisa R (2021). Leishmaniosis in rodents caused by *Leishmania*
234 *infantum*: a review of studies in the Mediterranean area. *Frontiers in Veterinary Science*
235 8:702687.
- 236 Calzolari M, Carra E, Rugna G, Bonilauri P, Bergamini F, Bellini R, et al. (2019) Isolation and
237 molecular typing of *Leishmania infantum* from *Phlebotomus perfiliewi* in a re-
238 emerging focus of Leishmaniasis, Northeastern Italy. *Microorganisms* 7:644.
239 <https://doi.org/10.3390/microorganisms7120644>.
- 240 CDC. Domestic rodent field identification. CDC Pictorial Keys. Atlanta, USA. Available
241 from: https://www.cdc.gov/nceh/ehs/docs/pictorial_keys/rodents.pdf.
- 242 Di Bella C, Vitale E, Russo G, Greco A, Millazzo C, Aloise G, et al. (2003) Are rodents a
243 potential reservoir for *Leishmania infantum* in Italy? *J Mt Ecol* 7(Suppl.):125-129.
- 244 Galán-Puchades MT, Gómez-Samblás M, Suárez-Morán JM, Osuna A, Sanxis-Furló J,
245 Pascual J, et al. (2019) Leishmaniasis in norway rats in Sewers, Barcelona, Spain. *Emerg*
246 *Infect Dis* 25(6):1222-1224. <https://doi.org/10.3201/eid2506.181027>.
- 247 Gaspari V, Ortalli M, Foschini MP, Baldovini C, Lanzoni A, Cagarelli R, et al. (2017) New
248 Evidence of Cutaneous Leishmaniasis in North-Eastern Italy. *JEADV* 31(9): 1534–40.
249 <https://doi.org/10.1111/jdv.14309>.
- 250 Gramiccia M, Gradoni L (2005) The current status of zoonotic Leishmaniases and
251 approaches to disease control. *Int J Parasitol* 35(11-12):1169-80.
252 <https://doi.org/10.1016/j.ijpara.2005.07.001>.

253 Helhazar M, Leitão J, Duarte A, Tavares L, Pereira da Fonseca I (2013) Natural infection of
254 synanthropic rodent species *Mus musculus* and *Rattus norvegicus* by *Leishmania*
255 *infantum* in Sesimbra and Sintra – Portugal. *Parasites Vectors* 6:88.
256 <https://doi.org/10.1186/1756-3305-6-88>.

257 Martín-Sánchez J, Torres-Medina N, Corpas-López V, Morillas-Márquez F, Díaz-Sáez V.
258 2020 Vertical transmission may play a greater role in the spread of *Leishmania infantum*
259 in synanthropic *Mus musculus* rodents than previously believed. *Transbound Emerg*
260 *Dis.* 67:1113–1118. <https://doi.org/10.1111/tbed.13436>.

261 Mendoza-Roldan J, Benelli G, Panarese R, Iatta R, Furlanello T, et al. (2020) *Leishmania*
262 *infantum* and *Dirofilaria immitis* infections in Italy, 2009–2019: changing distribution
263 patterns. *Parasites Vectors.* 13:193. <https://doi.org/10.1186/s13071-020-04063-9>.

264 Muñoz-Madrid R, Belinchón-Lorenzo S, Iniesta V, Fernández-Cotrina J, Parejo JC, Monroy
265 I, et al. (2013) First detection of *Leishmania infantum* kinetoplast DNA in hair of wild
266 mammals: Application of qPCR method to determine potential parasite reservoirs.
267 *Acta Trop* 128:706-709. <http://dx.doi.org/10.1016/j.actatropica.2013.08.009>.

268 Navea-Pérez HM, Díaz-Sáez V, Corpas-Lóez V, Merino-Espinosa G, Morillas-Márquez F,
269 Martín-Sánchez J (2015). *Leishmania infantum* in wild rodents: reservoirs or just
270 irrelevant incidental hosts? *Parasitol Res:* 114:2363-2370
271 <https://doi.org/10.1007/s00436-015-4434-y>

272 Pampiglione S (1975) La Leishmaniosi viscerale in Italia. *Ann San pubbl* 35(6)1021-1028.

273 Papadogiannakis E, Spannakos G, Kontos V, Menounos PG, Tegos N, Vakalis N (2009)
274 Molecular detection of *Leishmania infantum* in wild rodents (*Rattus norvegicus*) in

275 Greece. Zoonoses and Public Health 57:23-25. <https://doi.org/10.1111/j.1863->
276 [2378.2009.01264.x](https://doi.org/10.1111/j.1863-2378.2009.01264.x).

277 Pozio E, Maroli M, Gradoni L, Gramiccia M (1985) Laboratory transmission of *Leishmania*
278 *infantum* to *Rattus rattus* by the bite of experimentally infected *Phlebotomus perniciosus*.
279 Trans R Soc Trop Med Hyg 79(4):524–526. [https://doi.org/10.1016/0035-9203\(85\)90085-](https://doi.org/10.1016/0035-9203(85)90085-)
280 [9](https://doi.org/10.1016/0035-9203(85)90085-9).

281 RStudio Team (2020) RStudio: Integrated Development for R. RStudio, PBC, Boston, MA
282 URL <http://www.rstudio.com/>.

283 Santi A, Renzi M, Baldelli R, Calzolari M, Caminiti A, Dell'Anna S, et al. (2014) A
284 surveillance program on canine Leishmaniasis in the public kennels of Emilia-
285 Romagna Region, Northern Italy. Vector Borne Zoonotic Dis 14(3):206-11.
286 <https://doi.org/10.1089/vbz.2013.1362>.

287 Santi A, Rossi A, Galletti G, Casadei G, Tamba M (2021) Piano Regionale di controllo della
288 leishmaniosi risultati anno 2020. Ordine dei veterinari di Reggio Emilia
289 [http://www.ordineveterinariereggioemilia.it/userfiles/files/Relazione Piano Leishma](http://www.ordineveterinariereggioemilia.it/userfiles/files/Relazione_Piano_Leishma)
290 [nia_2020.pdf](http://www.ordineveterinariereggioemilia.it/userfiles/files/Relazione_Piano_Leishmania_2020.pdf)

291 Tsakmakidis I, Angelopoulou K, Dovas CI, Dokianakis E, Tamvakis A, Symeonidou I,
292 Antoniou M, Diakou A (2017) Leishmania infection in rodents in Greece. Trop Med Int
293 Health 22(12):1523-1532. <https://doi.org/10.1111/tmi.12982>

294 Rugna G, Carra E, Bergamini F, Calzolari M, Salvatore D, Corpus F, et al. (2018) Multilocus
295 microsatellite typing (MLMT) reveals host-related population structure in *Leishmania*

296 *infantum* from northeastern Italy. PLoS Negl Trop Dis 12(7):e0006595.
297 <https://doi.org/10.1371/journal.pntd.0006595>.
298 Zanet S, Sposimo P, Trisciuoglio A, Giannini F, Strumia F, Ferroglia E (2014) Epidemiology
299 of *Leishmania infantum*, *Toxoplasma gondii*, and *Neospora caninum* in *Rattus rattus* in
300 absence of domestic reservoir and definitive host. Vet Parasitol 199:247- 249.
301 <https://doi.org/10.1016/j.vetpar.2013.10.023>.

Table 1: Real time PCR positive samples

ID	Specimen	Locality	Real-Time PCR			
			Earlobe Skin	Spleen	Liver	Lymph Node
57	<i>Mus musculus</i>	Bizzuno (RA)	ct= 32.7 (87)	ct= 29.68 (676)	Negative	NA
59	<i>Mus musculus</i>	Bizzuno (RA)	Negative	ct= 30.77 (316)	ct= 31.97 (143)	NA
67	<i>Mus musculus</i>	Bizzuno (RA)	Negative	Negative	Negative	ct= 33.61 (47)
98	<i>Mus musculus</i>	S. Alberto (RA)	Negative	ct= 36.71 (5.8)	Negative	ct= 37.07 (4.5)
111	<i>Mus musculus</i>	Bizzuno (RA))	Negative	Negative	ct= 35.9 (10)	Negative
4	<i>Rattus norvegicus</i>	Ravenna (RA)	ct= 34.25 (30.9)	Negative	Negative	Negative
86	<i>Rattus norvegicus</i>	Godo (RA)	Negative	Negative	ct= 36.47 (6.8)	Negative
141	<i>Rattus norvegicus</i>	Ravenna (RA)	ct= 37.75 (2.9)	Negative	Negative	Negative
175	<i>Rattus norvegicus</i>	Forlì (FC)	Negative	Negative	ct= 36.27 (7.8)	Negative
178	<i>Rattus norvegicus</i>	Argenta (FE)	Negative	Negative	ct= 36.67 (5.8)	Negative
37	<i>Rattus rattus</i>	Forlì (FC)	Negative	ct= 36.47 (6.8)	Negative	Negative
60	<i>Rattus rattus</i>	San Pietro in Campiano (RA)	ct= 36.86 (6.2)	Negative	Negative	Negative
95	<i>Rattus rattus</i>	Montaletto di Cervia (RA)	Negative	ct= 37.44 (6.2)	Negative	Negative
179	<i>Rattus rattus</i>	Montaletto di Cervia (RA)	Negative	Negative	Negative	ct= 36.63
206	<i>Rattus rattus</i>	Longastrino (RA)	ct= 37.89 (2.6)	Negative	Negative	Negative

Legend: Ct values are reported as mean ct of observed in different target organs with the estimated quantity of parasites/ml (mean standard deviation observed ± 0.65). Localities are as well reported with reference to the province: Ferrara (FE), Forlì-Cesena (FC) and Ravenna (RA).

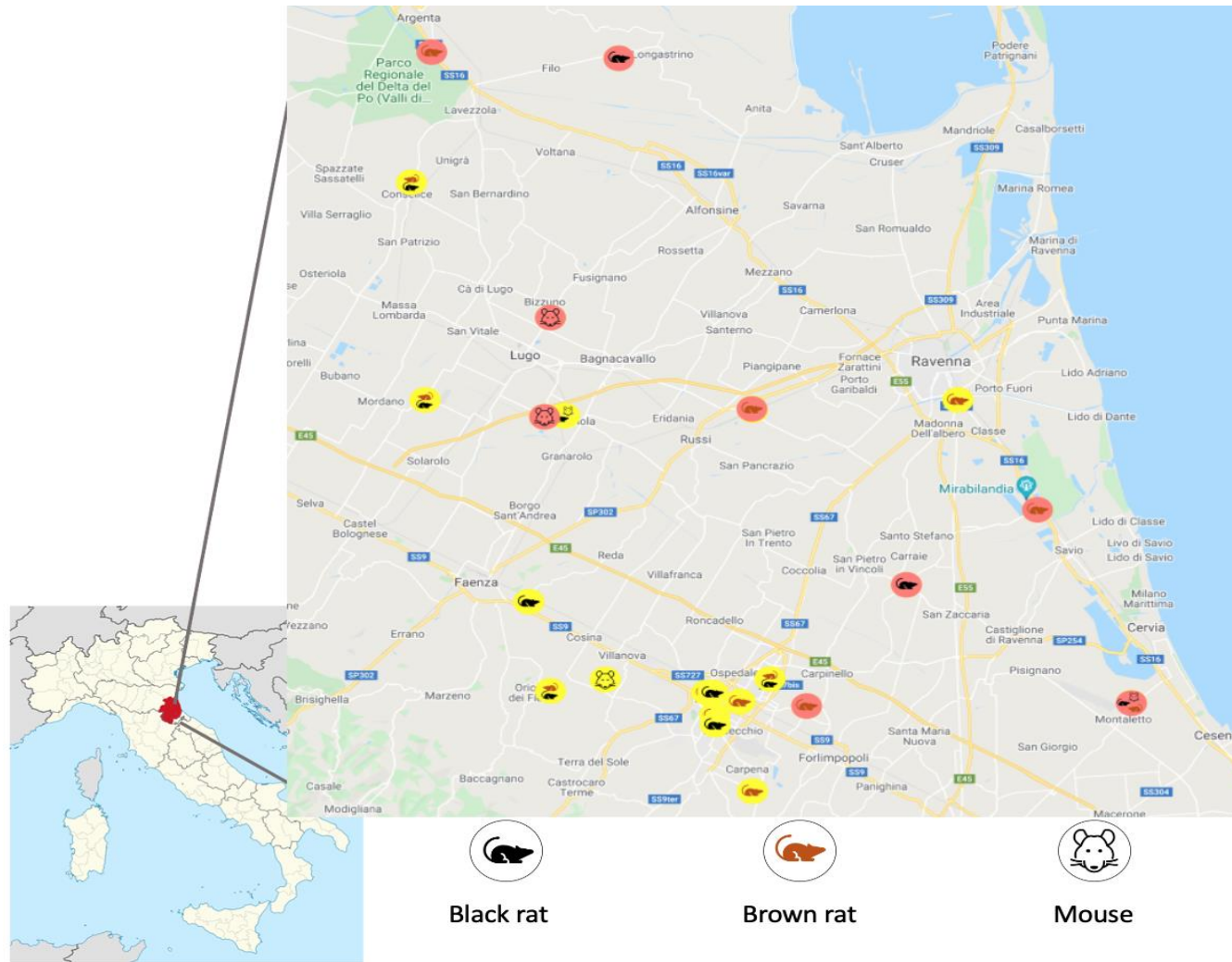


Figure 1. Map of the sampling area in the Emilia-Romagna Region. Dots are representative for sampling sites; red dots: at least one specimen positive, yellow dots: all the specimen negative.