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In-field and non-destructive monitoring of grapes maturity by hyperspectral imaging

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Abstract

Monitoring the quality attributes of grapes is a practice that allows the state of ripeness to be checked and the optimal harvest time to be identified. A non-destructive method based on hyperspectral imaging (HSI) technology was developed. Analyses were carried out directly in the field on a ‘Sangiovese’ (*Vitis vinifera* L.) vineyard destined for wine production, by using a Vis/NIR (400–1000 nm) hyperspectral camera. One vineyard row was analysed on 13 different days during the pre-harvest and harvest time. The soluble solids content (SSC) expressed in terms of °Brix was measured by a portable digital refractometer. Afterwards, the grape samples were split in two classes: the first one composed by the samples characterised by a °Brix lower than 20 (not-ripe), while the second one by the samples with a °Brix higher than 20 (ripe). Grape mean spectra were extracted from each hyperspectral image and used to predict the SSC by partial least squares regression (PLS), and to classify the samples into the two classes by PLS discriminant analysis (PLS-DA). SSC was predicted with a $R^2=0.77$ (RMSECV=0.79 °Brix), and the samples were correctly classified with a percentage from 86 to 91%. Even if the number of wavelengths was limited, the percentages of correctly classified samples were again within the above-mentioned range. The present study shows the

potential of the use of HSI technology directly in the field by proximal measurements under natural light conditions for the prediction of the harvest time of the ‘Sangiovese’ red grape.

Keywords: hyperspectral, in-field, grape, wine, harvest, classification

1. Introduction

Italy is the largest wine producer in the world, with a production in 2018 of 5,480 million litres (18.8% of world production) (OIV International Organisation of Vine and Wine, 2019). In an industrialised wine growing system, monitoring the quality attributes, such as soluble solids content (SSC), acidity and anthocyanin content of grapes is extremely important: well-planned monitoring allows to check the growth and ripening of the grapes, and finally to decide when to proceed with the harvest (Delrot, Medrano, Or, Bavaresco, Grando, 2010). For instance, by managing irrigation through the use of techniques such as the regulated deficit irrigation, significant increases in SSC and anthocyanins can be achieved which together with a decrease in yield and berry size can lead to substantial improvements in grape quality (Acevedo-Opazo, Ortega-Farias, Fuentes, 2010; Pellegrino, Lebon, Simonneau, Wery, 2005).

Monitoring of grape quality attributes can be carried out directly in the field using traditional destructive techniques. Alternatively, quality attributes can be estimated by non-destructive techniques, such as near-infrared (NIR) spectroscopy. Portable NIR instruments were used to determine the following quality attributes of grapes: water content, SSC, reductant sugars, pH, titratable acidity, maturity index (sugar/acidity ratio), extractable anthocyanins, potential anthocyanins (Teixeira Dos Santos, Lopo, Páscoa, Lopes, 2013).

Over the last two decades the use of HSI technology in the quality assessment of fruits and vegetables has become of increasing interest (Chandrasekaran, Panigrahi, Ravikanth, Singh, 2019; Liu, Zeng, Sun, 2015). HSI was initially limited to controlled environments such as laboratories but gradually,

53 thanks to the miniaturisation and improved computing and data storage capabilities, it began to be
54 used directly in the field (Benelli, Cevoli, Fabbri, 2020). Hyperspectral images can be captured either
55 remotely by airborne vehicles and unmanned aerial vehicles (Ishida et al., 2018; Matese & Di
56 Gennaro, 2015; Zarco-Tejada et al., 2013) or by ground vehicles (Deery, Jimenez-Berni, Jones,
57 Sirault, Furbank, 2014; Gutiérrez, Fernández-Novales, Diago, Tardaguila, 2018a; Huang, Lee,
58 Thomson, Reddy, 2016; Jay et al., 2017; Underwood, Wendel, Schofield, McMurray, Kimber, 2017;
59 Wendel, Underwood, Walsh, 2018; Whetton, Waine, Mouazen, 2018), which produces proximal
60 hyperspectral images with high spatial resolution. Proximal HSI could therefore allow non-
61 destructive, contactless, and automated monitoring of grape quality attributes. Moreover, the
62 acquisition of hyperspectral images can be performed continuously, enabling the rapid scanning of
63 large areas. Hyperspectral data is characterised by two spatial and one spectral dimension, therefore
64 only specific regions of interest (ROIs) can be selected, and the residual regions can be excluded. In
65 this sense, hyperspectral analysis is useful to guide the choice of true multispectral less expensive
66 solutions.

67 Concerning in-field grape studies, Gutiérrez et al. (2018a) adopted an on-the-go HSI system for the
68 classification of 30 grapevine varieties directly acquired in the field. Through the development of
69 classification models based on support vector machines (SVM) and multilayer perceptron (MLP),
70 prediction performance (F1 score) up to 0.99 was achieved. The same on-the-go HSI system
71 described above, combined with SVM, was adopted to estimate SSC and anthocyanin concentration
72 of wine grapes (Gutiérrez et al., 2018b). Determination coefficients (R^2) of 0.92 (RMSE=1.274 °Brix)
73 and 0.83 (RMSE=0.211 mg g⁻¹) were obtained for the prediction of SSC and anthocyanin
74 concentration, respectively. These two studies highlight the potential of HSI to monitor the indices
75 of grape ripening directly in the field and therefore to improve vineyard decisions and management.
76 Non-linear statistical methods were used in both studies. Considering the results just mentioned, it
77 would be interesting to investigate whether, even combining HSI with linear methods and reducing
78 the number of wavelengths, it is possible to monitor the degree of grape maturity directly in the field.

Furthermore, considering a possible application of this technique, a simple binary model able to simply determine whether grapes are ripe for harvesting could be interesting.

Thus, the present study aims to identify the proper degree of ripeness, suitable for harvesting wine grapes, through the observation of the SSC evolution by means of HSI technology applied directly in the field. The mean spectra were extracted from each hyperspectral image and used to predict the SSC and to classify the samples into the two classes (not-ripe and ripe), by partial least squares regression (PLS) and discriminant analysis (PLS-DA), respectively.

2. Materials and methods

2.1 Samples

One side of a row of ‘Sangiovese’ (*Vitis vinifera* L.) grape vineyards, located near Cesena, Italy, was analysed on 13 different days in the period between August 20th and October 4th, 2019 (from pre-harvest to harvest time). The row was divided into 11 sections; from each section 3 grapes were taken for each day of analysis, for a total of 429 samples.

2.2 Hyperspectral acquisitions

The adopted push-broom hyperspectral camera (Nano-Hyperspec VNIR, Headwall Photonics, Inc., Fitchburg, MA, USA) scans single lines in a sequence, each one consisting of 640 voxels: the image is created by moving the camera along the scanning direction (Fig. 1a). Each voxel contains, in addition to the two spatial dimensions, a Vis/NIR spectrum (400–1000 nm) characterized by 272 spectral bands, with a nominal spectral resolution of 2.2 nm. The mounted lens has an effective focal length of 17 mm, with the optical axis perpendicular to the side of the vineyard row (scanned surface) analysed (Figure 1b). The camera was installed on a garden cart (Fig. 1b) 120 cm above the ground and it was powered by a 12 V, 45 Ah automotive battery through a DC to AC power inverter. The scans were performed at about 1.6 m from the side of the vineyard row.

104 Direct sunlight with clear sky conditions, was used as a light source. To reduce the fluctuation of the
105 sample temperature, all the acquisitions were carried during the same period of the day, from
106 10:30 a.m. to 12:00 p.m.

107 The frame rate was set to approximately 100 frames s⁻¹. The exposure time was set from 6 to 8 ms,
108 depending on the light intensity, and was achieved through calibration, by framing a white high-
109 reflectance matter panel, placed at the same distance as the vineyard row, to cover the entire angle of
110 view of the camera. Given the clear sky conditions and the short time required, about 10 min,
111 acquisition of hyperspectral images of the 11 vineyard row sections and calibration was carried out
112 only once per day.

113 The raw diffuse reflectance spectrum (R_R) was extracted from the HS images. The calibrated diffuse
114 reflection spectrum (R_C) was calculated by applying the following equation:(Guo et al., 2019):

$$115 \quad R_C = \frac{R_R - R_D}{R_W - R_D} \quad (1)$$

116 where R_D is the reflectance spectrum of dark reference, obtained by applying the cap on the lens; R_W
117 is the reflectance spectrum of white reference, obtained by means calibration with the white high-
118 reflectance matte panel reported above.

119 A hyperspectral image from each of the 11 sections obtained from the vineyard row, was acquired
120 per day of analysis (Fig. 1c): therefore, during the 13 d of analysis, a total of 143 vineyard row sections
121 were scanned.

122

123 *2.3 Soluble solids content measurement*

124 After the acquisition of the images, the SSC, expressed in °Brix, was measured on 3 grape berries
125 (randomly selected) from each of the 11 sections by using a portable digital refractometer (PR-101
126 Digital Refractometer, ATAGO CO., LTD, Tokyo, Japan). Subsequently, the mean for each section
127 was calculated.

128 One-way analysis of variance (ANOVA) with Tukey-HSD post-hoc test (p-level < 0.05) was applied
129 to evaluate significant differences between SSC means over the different days of analysis.

2.4 Hyperspectral images elaboration

ROI selection was made using the software HyperCube, v. 11.52 (U.S. Army Engineer Research and Development Center (ERDC), USA). For each image, 5 points and a maximum of a further 120 adjacent points (11 x 11 voxels matrix, with the selected point in the centre of the matrix) were selected on 5 different berries directly illuminated by the sun, not in the shade. The (reference) points included in the classification were characterized by a metric distance from the mean (signature) spectrum of the manually selected points within the range (tolerance threshold) [0,0.04] (Fig. 2). The metric distance was calculated by applying the absolute difference (Manhattan) function (Eq. (2)) (Deborah, Richard, Hardeberg, 2015) and normalised in the range [0,1].

$$d(R_1, R_2) = \sum_{\lambda} |R_{1,\lambda} - R| \quad (2)$$

where R_1, R_2 are two reflectance spectra.

From the spectra of the classified points, the mean spectrum for each hyperspectral image was calculated.

The spectral bands between 400–424 nm were omitted as a result of the low signal-to-noise ratio produced by the sensor, as reported in Wendel et al. (2018). The spectra were smoothed (Savitzky-Golay method; polynomial order: 2; smoothing points: 15) to reduce noise from the spectra and following pre-treated by the standard normal variate (SNV) method, first derivative (D1) and finally mean centred (MC). The SNV is one of the most common pre-processing method used to correct spectra for changes in optical path length and light scattering, while the derivatives have the capability to remove both additive and multiplicative effects in the spectra (Rinnan, van den Berg, Engelsen, 2009). After SNV, each spectrum will have a mean of 0 and a standard deviation of 1.

Principal component analysis (PCA) was applied to the mean spectra as exploratory technique to visualize the data according to °Brix and time evolution. Subsequently, a preliminary PLS regression model was built to estimate the SSC. The validation was carried out by the venetian blind cross-validation method (segments: 10).

PLS-DA models were built to classify the samples according to °Brix. In particular, classification models with 2 categories (not-ripe and ripe) were developed: according to Bucelli, Costantini and Storchi (2010), the first class was composed by the samples characterized by a °Brix lower than 20 (0), while the second one by the samples with a °Brix equal or higher than 20 (1). The sample dataset (n = 143) was split in calibration (venetian blinds cross-validation, including 75% of the samples) and external validation set (25% of the samples) by using the Onion method (Gallagher et al., 2004). The threshold value, able to identify the belonging category of each sample into one of the groups, was defined by using a probabilistic approach based on Bayes's rule. To avoid the model over-fitting, the optimal number of latent variables were chosen by plotting the root mean square error of cross-validation (RMSECV) as a function of the number of components and by identifying where the curve reaches a local minimum. The receiver operating characteristic (ROC) curves in prediction were evaluated to assess the goodness of the models. All the chemometrics models were developed by using PLS Toolbox for Matlab2018a.

3. Results and discussion

Means and standard deviation of the °Brix measured during the 13 d of analysis are reported in Table 1. The increase between the first and last day was of 27.5%, from 17.8 °Brix (day I) to 22.7 °Brix (day XIII). Several significant differences between the °Brix mean values were achieved over the different days of analysis. The whole data set was characterized by a mean value of 20.6 ± 1.7 °Brix, which make suitable to split it in two subsets with a threshold of 20 °Brix. Raw and pre-treated (smoothing and SNV) mean spectra of all the samples by day of analysis are presented in Fig. 3. In the Vis/NIR region (400–1000 nm), the visible spectrum (400–700 nm) presents the absorption bands of some substances used as ripening indexes of fruit: anthocyanins at around 500 nm, carotenoids at 570–590 nm, and chlorophyll *a* at 680–710 nm (ElMasry, Wang, ElSayed, Ngadi, 2007; Munera et al., 2017). In the NIR region (700–1000 nm), absorption bands of water at 760 nm and 960–970 nm are characterised by the overtone of O-H bonds (McGlone &

181 Kawano, 1998; Nicolaï et al., 2007): since the water content of a ripe wine grape is 70–80% (FAO
182 Food and Agriculture Organization, 2009), it can be expected that the water related absorption band
183 will prevail. Absorption band around 840 nm was associated with sugar (Pu, Liu, Wang, Sun, 2016);
184 moreover, peaks observed in the 950–1000 nm region were related to both water and carbohydrates,
185 as the second overtone of O-H and N-H, a combination band of O-H bonds and the third overtone of
186 C-H, were found in the region (Camps & Christen, 2009). As observed, the water absorption peaks
187 in the NIR (700–1000 nm) spectral region are not very marked and wide. Therefore, spectral
188 information from SSC in the 800–1000 nm range will tend to be less covered by water (Camps &
189 Christen, 2009; Manley, Joubert, Myburgh, Kidd, 2007).

190 The score plot of the first two principal components (PC1: 59%; PC2: 28%) resulting from PCA
191 shows the samples distribution according to the day of analysis. A tendency to place the samples from
192 left to right can be observed on PC1, starting from day I (quadrant III) to day XIII (quadrant IV)
193 (Figure 4a).

194 A similar distribution can be observed according to the SSC: the samples are distributed along PC1,
195 mainly on the left those with $\text{SSC} < 20^\circ\text{Brix}$ (quadrants II and III), on the right those with
196 $\text{SSC} \geq 22^\circ\text{Brix}$ (quadrants I and IV) (Fig. 4b).

197 The best PLS results were obtained pre-treating the spectra by SNV + MC. Particularly, $R^2 = 0.768$
198 and $\text{RMSECV} = 0.79^\circ\text{Brix}$ were achieved in cross-validation with 7 latent variables. Figure 5 shows
199 the predicted versus measured $^\circ\text{Brix}$ values.

200 These results agree with those present in literature and developed by using spectra in the same
201 wavelength range and acquired directly in-field. Furthermore, regardless of the R^2 values, the RMSE
202 is substantially lower. Diezma-Iglesias, Barreiro, Blanco and García-Ramos (2008) predicted SSC on
203 480 samples of ‘Cabernet Sauvignon’ by using a portable hand-held spectrometer (590–1090 nm),
204 reporting a $R^2 = 0.72$, while Guidetti, Beghi and Bodria (2010) working with a simple Vis/NIR system
205 in the range from 400 to 1000 nm, obtained in prediction $r=0.82$ ($R^2=0.67$) and $\text{RMSEP}=1.48^\circ\text{Brix}$.
206 Gutiérrez et al. (2018b) used the HSI technique (400–1000 nm) to measure SSC in wine grapes in

207 real time. R^2 of 0.91 (RMSE = 1.358 °Brix) and 0.92 (RMSE = 1.274 °Brix) were achieved in cross
208 validation and prediction, respectively, by using the SVM techniques.

209 PLS-DA results, in terms of percentage of correctly classified samples, are reported in Table 2. The
210 percentages ranged from 86 to 91%. Considering the prediction set, the best result was obtained
211 applying as pre-treatment the SNV + MC. The results obtained with the hyperspectral technique
212 improved those achieved by Guidetti et al. (2010) by using of a portable contact system. The authors
213 combined Vis/NIR spectroscopy in the wavelength range 450 – 980 nm with PLS-DA to classify
214 grape samples in two groups based on SSC (threshold = 21 °Brix), obtaining a percentage of samples
215 correctly classified (in prediction) of 77.1% (Guidetti et al., 2010).

216 The ROC curves in prediction (Fig. 6) summarise the trade-off between specificity (number of
217 samples predicted to not be in the class divided by the actual number not in the class) and sensitivity
218 (number of samples predicted to be in the class divided by number actually in the class) for the PLS-
219 DA classification models. The area under the curves (AUC = 0.9855 for SNV + MC and
220 AUC = 0.9578 for SNV + D1 + MC) suggest that the models were characterised by a high degree of
221 discrimination, confirming that the best model was those developed considering only the SNV + MC
222 as pre-treatment.

223 Results, in terms of probability (Bayes's rule) of belonging to the class °Brix < 20, are shown in Fig.
224 7. The higher a sample is placed, the higher the probability that it will be classified as a member of
225 the °Brix < 20 class. Consequently, samples classified as members of the other class (°Brix \geq 20)
226 are placed at the bottom of the graph. The threshold value (dotted red line) was set at 0.5 (probability
227 of 50%); samples with a probability lower than this value are considered improperly classified.

228 Considering the SNV + MC and SNV + D1 + MC pre-treatments, 89% and 77% of the samples (in
229 prediction) were classified with a probability higher than 70%.

230 Figure 8 shows the VIP (variable importance in projection) scores obtained by the PLS-DA models.
231 These scores estimate the importance of each variable in the projection used in the PLS-DA model.
232 A variable with a VIP score close to or higher than 1 can be considered important in a given model.

233 For both the pre-treatments, similar regions with VIP score higher than 1 were obtained, suggesting
234 that the wavelengths with the highest contribution are in the NIR region of the spectrum (from 700
235 to 1000 nm). Consequently, the variable selection method based on the VIP scores higher than 1 was
236 used to reduce the original data set.

237 PLS-DA results (percentage of correctly classified samples) obtained by using the reduced number
238 of variables (wavelengths), are reported in Table 3. In particular, 93 (SNV + MC) and 88
239 (SNV + D1 + MC) x-variables were used to develop the new PLS-DA. The percentages of correctly
240 classified samples are slightly lower (from 86 to 91%) than those obtained considering the whole
241 spectrum. However, the results are still completely acceptable. This confirms also by the high AUC
242 values (0.939 and 0.942).

243 This work presents a solution for in-field and non-destructively determination of grape maturity
244 degree by using HSI combined with linear chemometric techniques. Particularly, the results confirm
245 the suitability for estimating the soluble solid content of red grapes.

246 The positioning of the vineyard row did not create any obstacles regarding the direct solar lighting of
247 the grape bunches. The images along the vineyard row were acquired in the NNE-SSO direction, with
248 the row on the right-hand side of the scanning direction. Consequently, the suitable period for analysis
249 was during the late morning and not later than midday. To scan the other side of the row, it would
250 have been necessary to proceed after midday. Shadows also did not present any problems, as the sun
251 was behind the camera at the time of the acquisitions. To correctly acquire hyperspectral images, the
252 vines should be stripped to ensure that the upper leaves do not overshadow the grapes below. Grapes
253 often had reflection and shaded areas, which need to be excluded during the selection of the ROIs,
254 along with fully or partially shaded grape bunches. In the presence of clouds or even just a slight
255 cloud cover, a significant variation in light intensity was observed: this means that the camera would
256 need to be recalibrated every time a section of the row was scanned. In addition, it is possible that the
257 brightness conditions change soon after calibration, so it would be necessary to recalibrate and
258 immediately proceed with the scan of the row section.

259 The variability grape of a vineyards, in terms of SCC, often is quite high. This depends on many
260 factors, such us the vineyards orientation, unevenness of the land and meteorological phenomena. For
261 this reason, the grapes are harvested at different times even along the same row. Consequently, to
262 optimise the harvest, a technique that allows to have a mapping of the SCC for all grapes, would
263 certainly be an advantage.

264

265 **4. Conclusions**

266 Hyperspectral imaging technology, usually adopted in laboratories with auxiliary artificial lighting,
267 was used in-field under natural lighting conditions to monitor the maturity degree of ‘Sangiovese’
268 (*Vitis vinifera* L.) grapes. The results achieved confirm that it is possible to predict the soluble solid
269 content and to classify grape samples into two classes (not-ripe and ripe) using a linear technique to
270 elaborate the spectral data. Furthermore, the classification performance remained substantially
271 unchanged by reducing the number of wavelengths, so it is expected that a less expensive
272 multispectral camera in the 400–1000 nm range can work just as well. The implementation of a
273 hyperspectral imaging system on an agricultural vehicle coupled to a gimbal stabilisation system,
274 together with the development of hyperspectral image segmentation techniques, would allow
275 on-the-go analysis of large vineyard extensions. Attention should be paid to the presence of water on
276 the surface of the sample under analysis, to the presence of variable cloudiness and, if the leaves are
277 analysed, to the presence of wind.

278

279 **Declarations of interest: none.**

280

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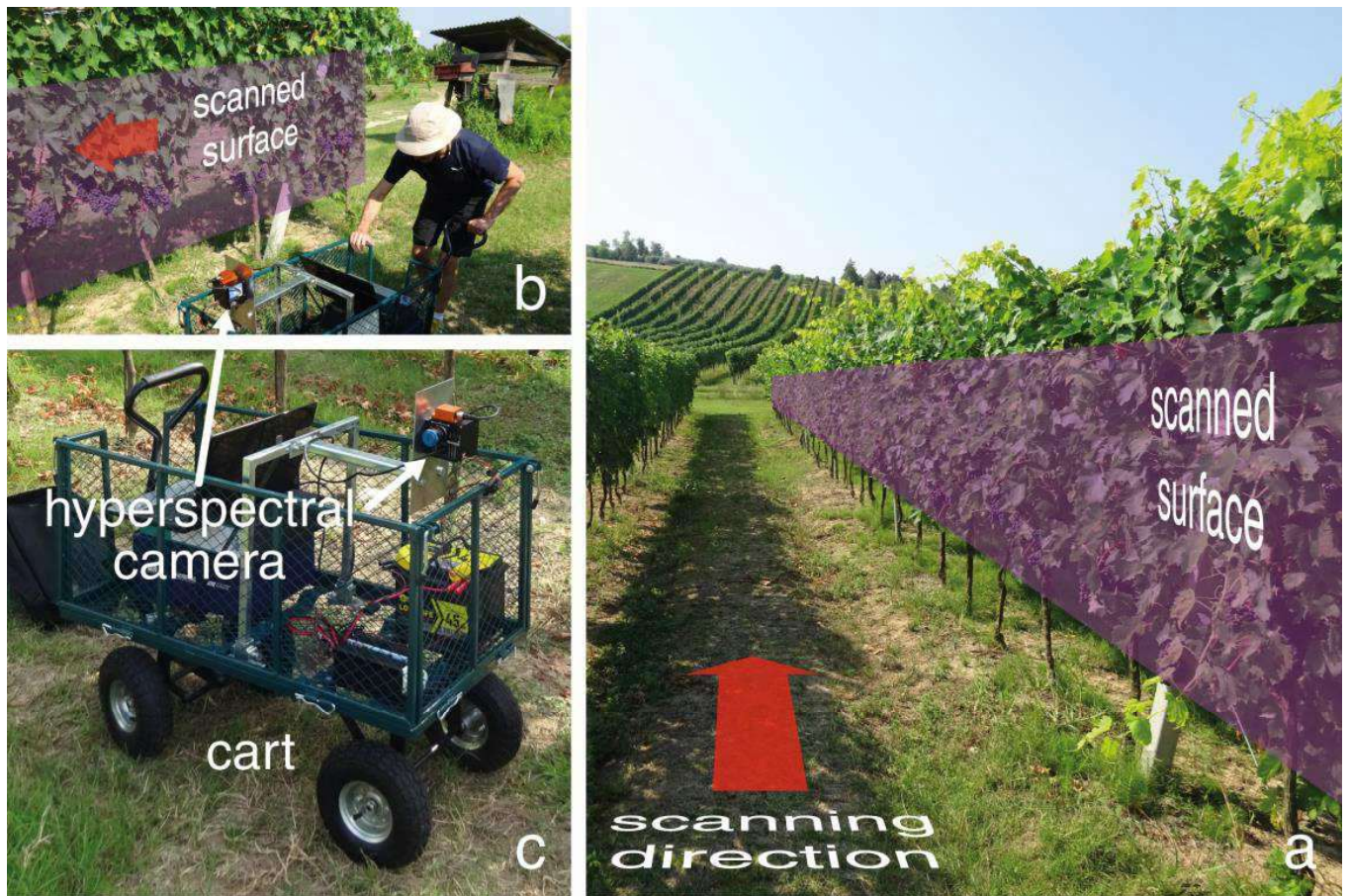


Fig.1

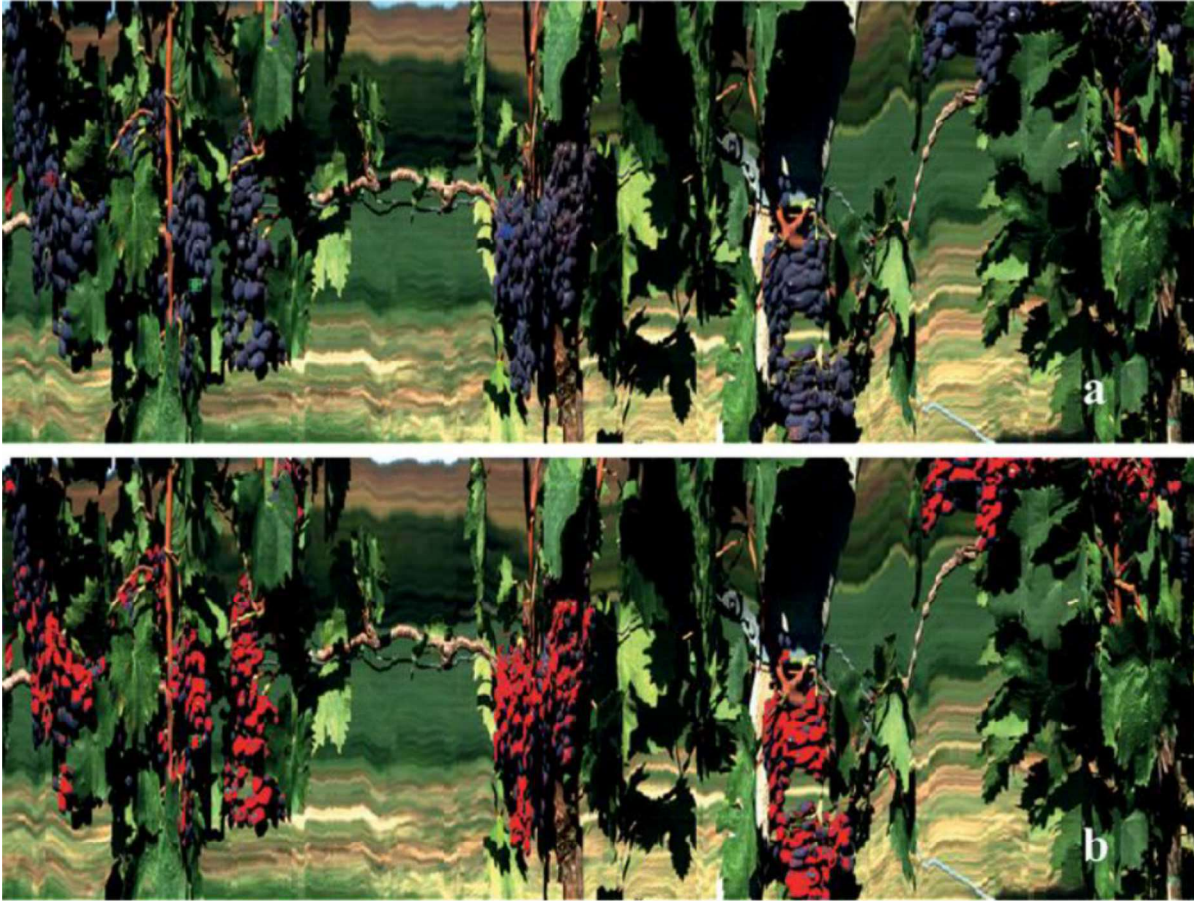


Fig. 2

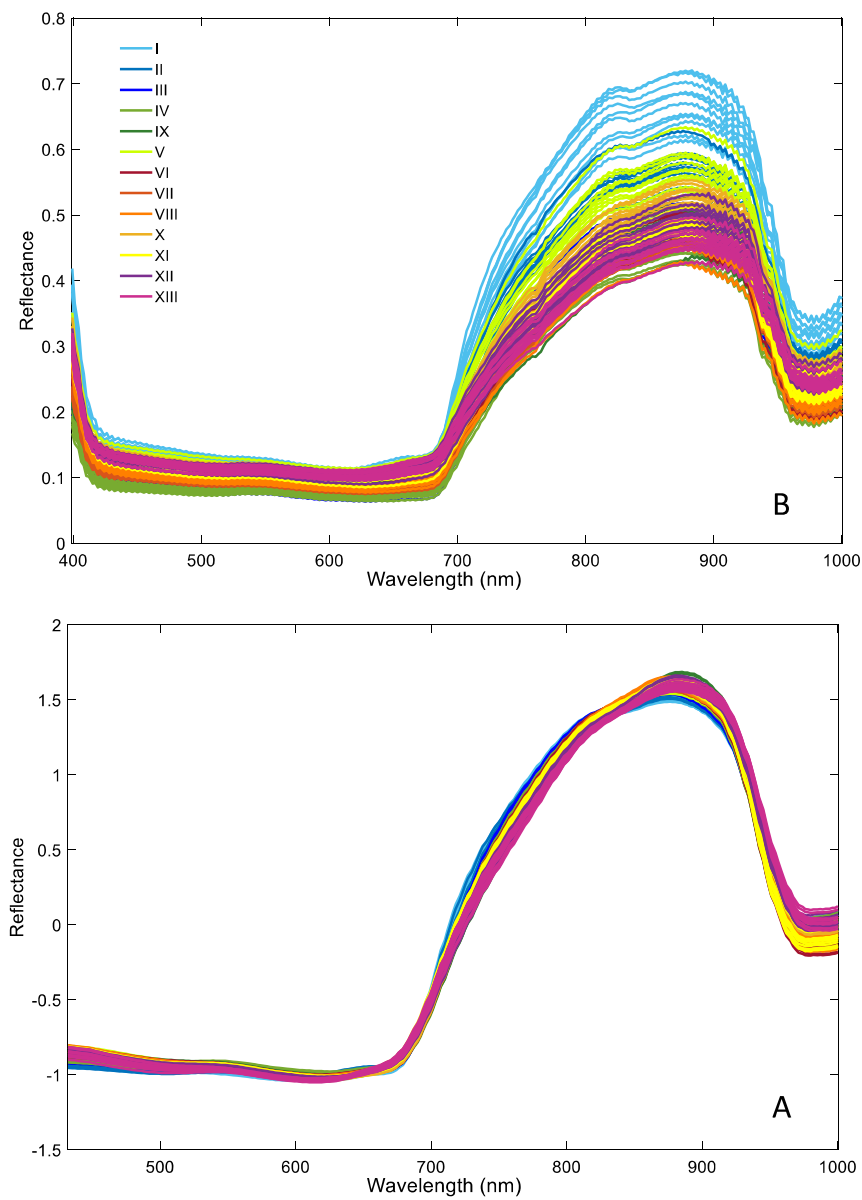


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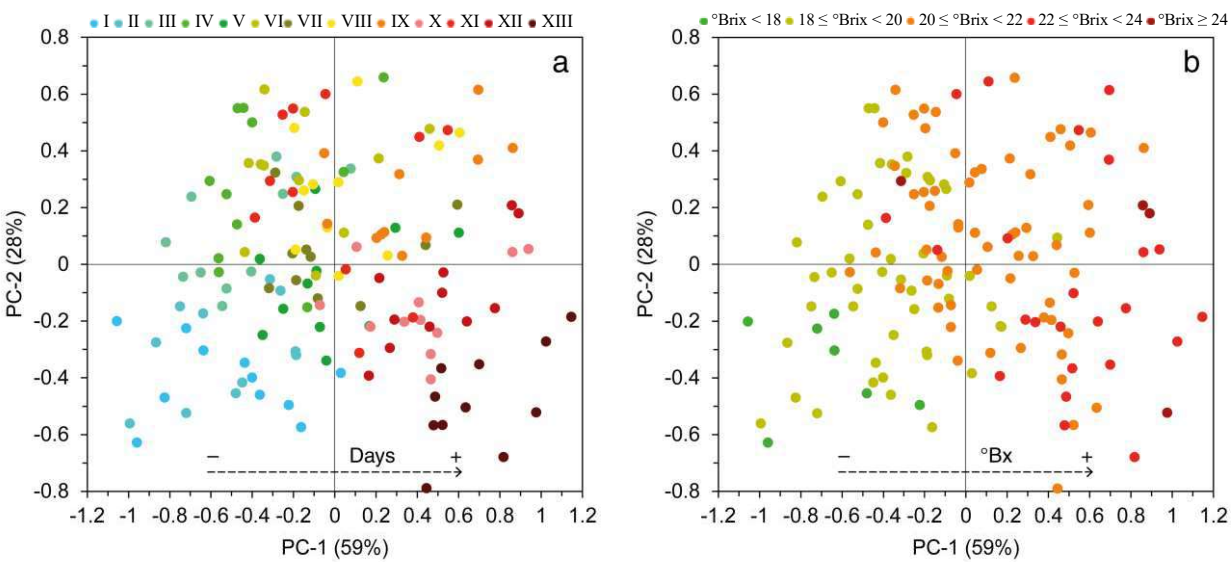


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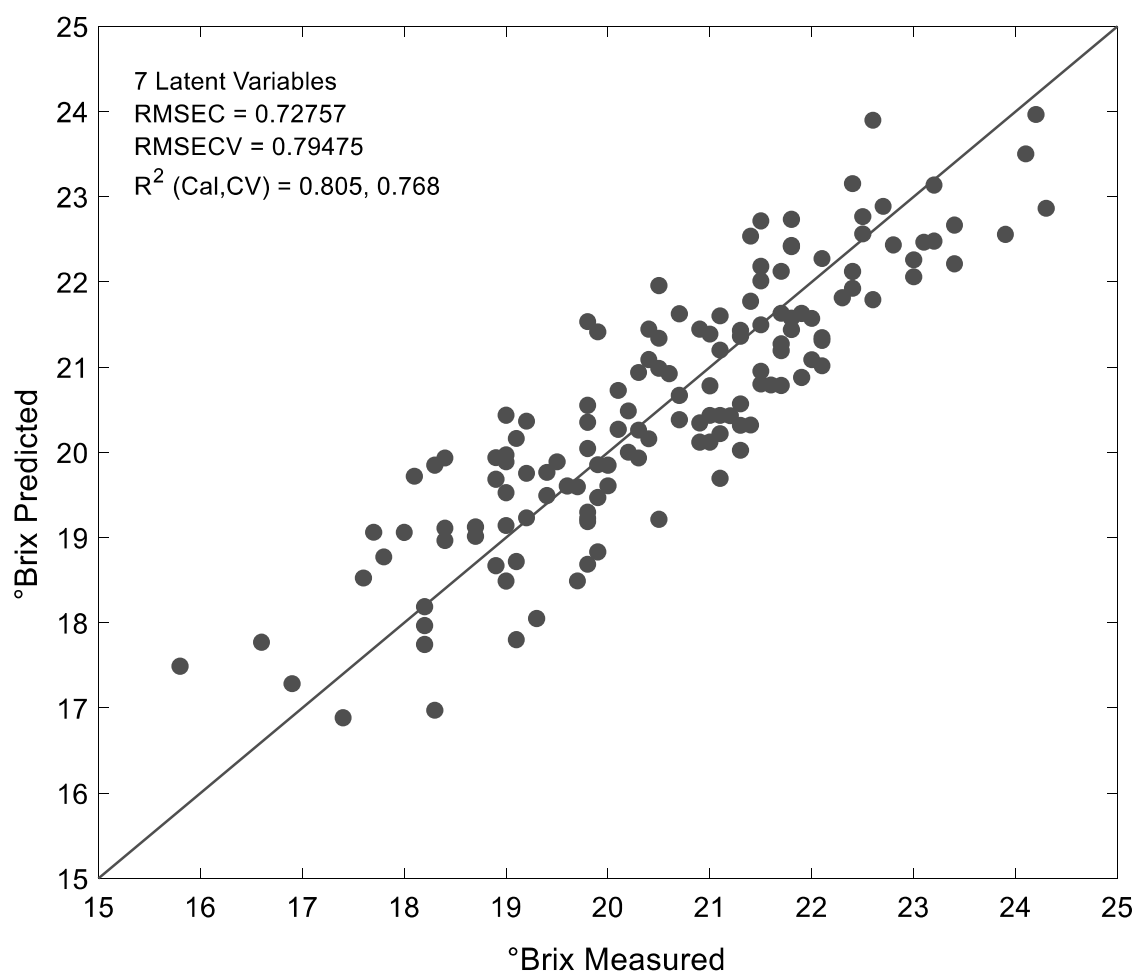


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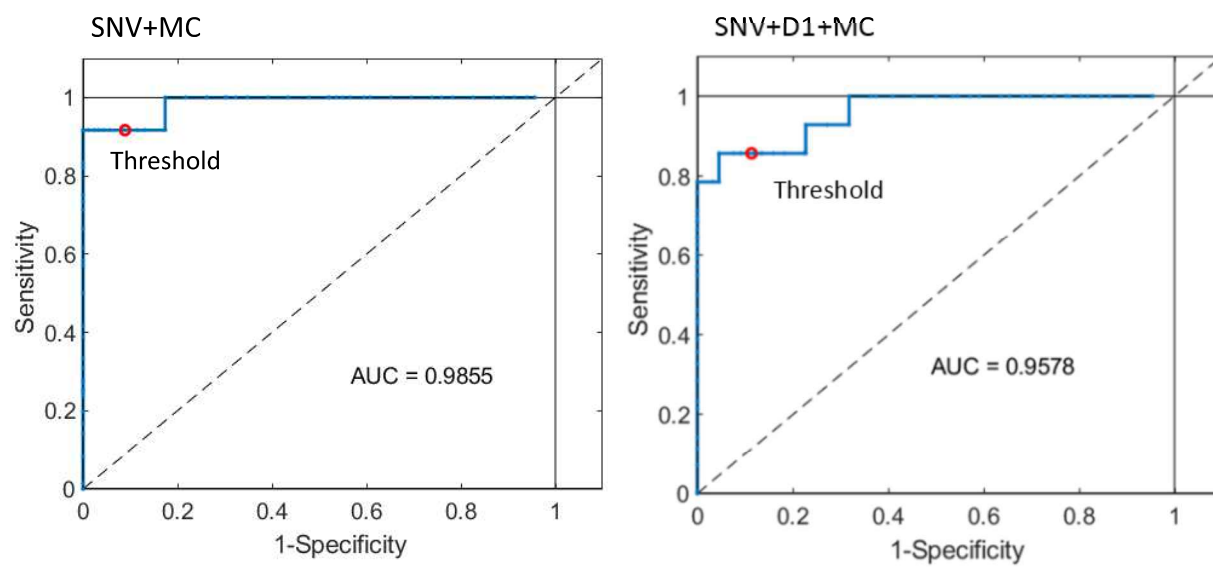


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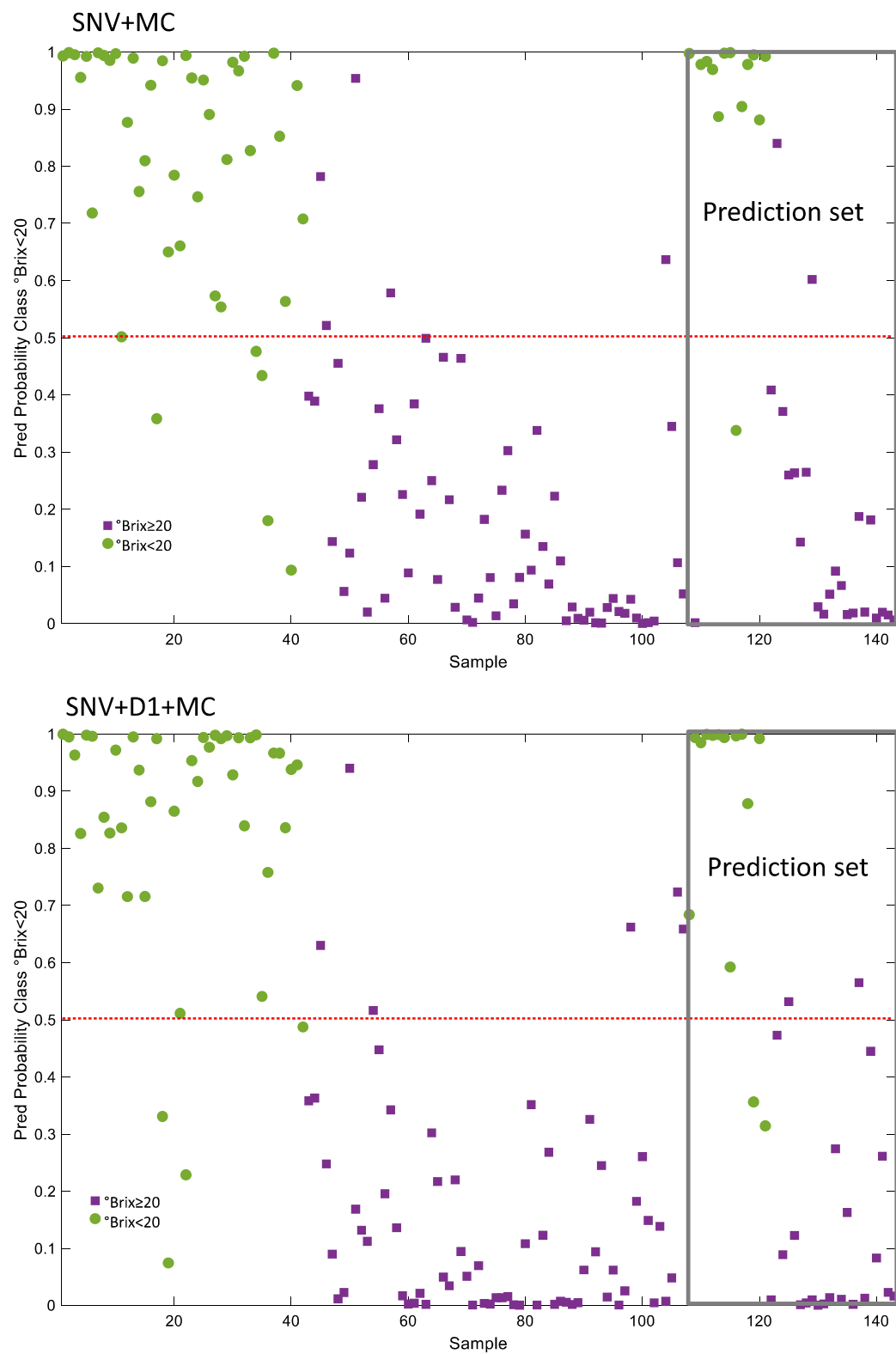


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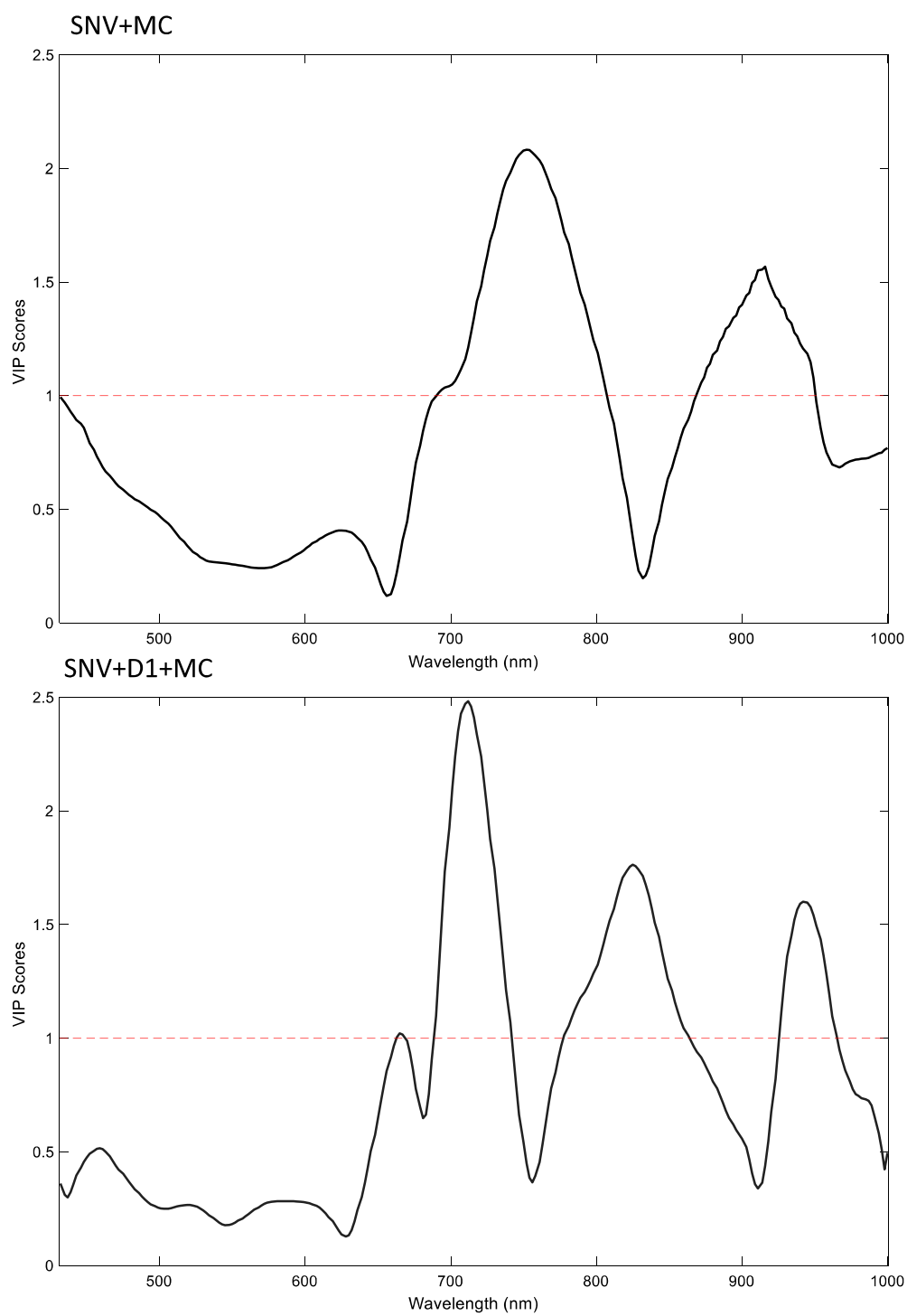


Fig. 8

1 **Figure captions**

2 **Fig. 1.** (a) Vineyard row, highlighted by the purple surface, scanned with the Vis/NIR hyperspectral
3 camera; red arrow indicates the direction of scanning; (b) Garden cart mounted with hyperspectral
4 camera. (c) In-field hyperspectral imaging to measure soluble solids content of wine grape berries
5 during ripening.

6 **Fig. 2.** (a) RGB image from hyperspectral image of a scanned vineyard row section. (b)
7 Representation of the ROI (in red) resulting from the classification obtained by the Manhattan
8 function.

9 **Fig. 3.** Raw (A) and pre-treated by smoothing and SNV (B) spectra of all samples on different days
10 of analysis (from I to XIII).

11 **Fig. 4.** Score plot obtained by the PCA according to: (a) days of analysis (from I to XIII); (b) soluble
12 solids content (°Brix).

13 **Fig. 5.** Measured vs predicted values of solid soluble content (°Brix) obtained by PLS regression
14 (cross validation).

15 **Fig. 6.** ROC curve of the PLS-DA models in prediction.

16 **Fig. 7.** Probability values of belonging to the class °Brix<20.

17 **Fig. 8.** VIP scores of the PLS-DA.

Table 1. Mean and standard deviation values of soluble solids content (°Brix) as a function of days of analysis.

Day of analisys	Mean (°Brix)	Standard deviation (°Brix)
I	17.8 ^a	1.05
II	19.0 ^{a,b}	0.81
III	19.2 ^{b,c}	0.75
IV	19.9 ^{b,c,d}	1.1
V	20.1 ^{b,c,d}	1.16
VI	20.2 ^{b,c,d,e}	0.95
VII	20.5 ^{c,d,e,f}	1.11
VIII	20.7 ^{d,e,f}	0.82
IX	21.5 ^{e,f,g}	0.76
X	21.6 ^{e,f,g}	0.89
XI	21.7 ^{f,g}	1.17
XII	22.7 ^g	0.94
XIII	22.7 ^g	0.88

Note: means with the same letter are not significant different at p-level < 0.05.

Table 2. PLS-DA results in terms of percentages of correctly classified samples (whole spectral range).

Spectra pretreatment	Class	Calibration (n=107)	Cross-validation (n=107, 10 segments)	Prediction (n=36)	LV
SNV+MC	°Brix<20	89%	89%	91%	4
	°Brix≥20	91%	91%	91%	
SNV+D1+ MC	°Brix<20	91%	89%	86%	4
	°Brix≥20	91%	91%	91%	

Note: SNV=Standard Normal Variate; MC=Mean Cantered; D1=first derivative, LV=Latent Variable.

Table 3. PLS-DA results in terms of percentages of correctly classified samples (reduced spectral range).

Spectra pretreatment	Class	Calibration (n=107)	Cross-validation (n=107, 10 segments)	Prediction (n=36)	LV
SNV+MC	°Brix<20	84%	83%	86%	4
	°Brix≥20	93%	92%	91%	
SNV+D1+ MC	°Brix<20	89%	89%	88%	3
	°Brix≥20	90%	87%	87%	

Note: SNV=Standard Normal Variate; MC=Mean Cantered; D1=first derivative, LV=Latent Variable.

<i>Nomenclature</i>	
AUC	area under the curve
CV	cross validation
D1	first derivative
HSI	hyperspectral imaging
MC	mean centring
NIR	near-infrared
PCA	principal component analysis
PLS	partial least squares
PLS-DA	partial least squares discriminant analysis
R	reflectance spectrum
RMSE	root mean square error
ROC	receiver operating characteristic
ROI	region of interest
SNV	standard normal variate
SSC	soluble solids content
SVM	support vector machines
VIP	variable importance in projection

Declaration of interests

☒ The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

☐The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: