

Melanins

Hydrogen Atom Transfer from HOO^\bullet to *ortho*-Quinones Explains the Antioxidant Activity of Polydopamine

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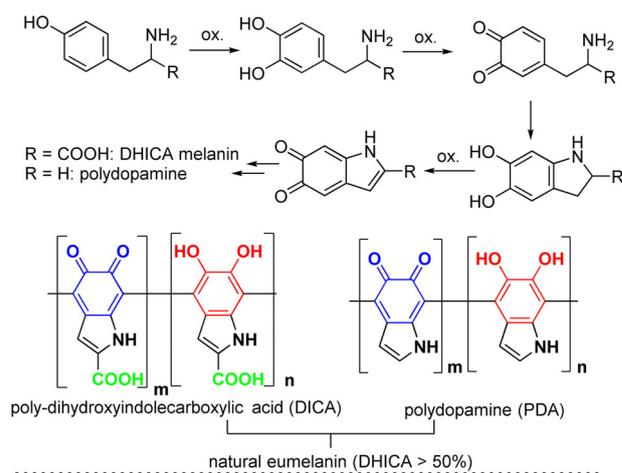
Abstract: Melanins are stable and non-toxic biomaterials with a great potential as chemopreventive agents for diseases connected with oxidative stress, but the mechanism of their antioxidant action is unclear. Herein, we show that polydopamine (PDA), a well-known synthetic melanin, becomes an excellent trap for alkylperoxyl radicals (ROO^\bullet ; typically formed during autoxidation of lipid substrates) in the presence of hydroperoxyl radicals (HOO^\bullet). The key reaction explaining this peculiar antioxidant activity is the reduction of the *ortho*-quinone moieties present in PDA by the reaction with HOO^\bullet . This reaction occurs via a H-atom transfer mechanism, as demonstrated by the large kinetic solvent effect of the reaction of a model quinone (3,5-di-*tert*-butyl-1,2-benzoquinone) with HOO^\bullet ($k = 1.5 \times 10^7$ and $1.1 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$ in PhCl and MeCN). The chemistry disclosed herein is an important step to rationalize the redox-mediated bioactivity of melanins and of quinones.

Melanins are a family of intensely colored polymers derived from oxidative polymerization of phenols such as tyrosine, DOPA and dopamine.^[1,2] Although the structure of melanins is not fully clarified, there is evidence that it consists of reduced (catechol) and oxidized (*ortho*-quinone) units (see Scheme 1 A).^[3,4] Beside their important role in living organisms,^[5–7] melanins are currently being investigated for many applications, including energy storage, biocompatible adhesion or coating systems, drug delivery, and skin protection.^[5,6,8] Melanins have been proposed to possess a “multi-antioxidative” activity,^[4,5,8,9] that has been related, for instance, to their anti-inflammatory,^[10] wound regeneration^[11] and anti-ischemic activity.^[12] Natural eumelanin incorporating large amounts of 5,6-dihydroxyindole-2-carboxylic acid, DHICA (see Scheme 1 A), has been reported to have the

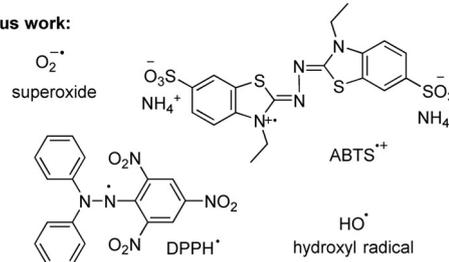
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strongest capability to quench radicals, as compared to synthetic melanins obtained by dopamine polymerization.^[5] Polydopamine traps HO^\bullet radicals and has superoxide dismutase (SOD)-like activity (see Scheme 1 B) presumably involving the stable radicals “hosted” inside melanins.^[8,9] Despite the number of publications reporting the antioxidant activity of melanins is steadily increasing, the mechanism of radical trapping remains unclear. Most mechanistic studies have been performed by using stable radicals such as DPPH $^\bullet$ and ABTS $^{+\bullet}$ (Scheme 1 B).^[7,9] While these pioneering works have triggered the interest on melanins’ redox properties, these artificial radicals have limited similarity to transient alkylperoxyl radicals (ROO^\bullet) (Scheme 1 C).^[13] The structure of melanins at the molecular level largely depends on the

A. Biosynthesis of melanins



B. Previous work:



C. This work:



Scheme 1. (A) (Bio)synthetic pathway leading to the formation of melanins; (B,C) radical used to investigate the antioxidant activity of melanins.

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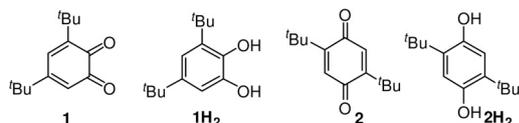
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nature of the phenolic monomer,^[7,14] on the polymerization conditions^[14,15] and on post-synthetic functionalization.^[12] Despite the efforts in this direction,^[16] there is currently no accepted simplified model for the redox properties of melanins. A good starting point to face this problem would be having a clear understanding of the radical trapping behavior of the basic structural units that are present in all melanins, namely the 1,2-dihydroxy benzene (catechol) and the 1,2-benzoquinone moieties.^[17–19]

Herein we report the results of our studies of controlled radical chain oxidations carried on by mixed alkylperoxy (ROO[•]) and hydroperoxy (HOO[•]) radicals^[20–23] to understand the reactivity and the mechanisms of antioxidant action of polydopamine (PDA, Scheme 1 A) and of two model *ortho* (**1**) and *para* (**2**) quinones (Scheme 2).



Scheme 2. Model compounds investigated in this study.

The chain-breaking antioxidant activity of **1** and **2** was evaluated by measuring, at 30 °C and in a low-polarity solvent (chlorobenzene), the rate of O₂ consumption during the azobis(isobutyronitrile) (AIBN) initiated oxidation of styrene, a reference organic substrate whose autoxidation at low temperature is propagated by ROO[•].^[24,25] 1,4-Cyclohexadiene (CHD) was used as oxidizable co-substrate to produce HOO[•] [(Scheme 3, Eqs. (8) and (9)].^[20–23] As expected,^[17–19] **1** and **2** had no effect on the peroxidation of styrene alone (Figure 1 A). When CHD was added to styrene, both quinones suppressed the peroxidation, as shown in Figures 1 A and S1.

Control experiments (styrene containing CHD but without quinones) indicated neither inhibition nor retardation, thus the strong inhibition observed for styrene/CHD/quinone can be explained by the reactions reported in Scheme 3. Comparing the kinetic traces in Figure 1, we should notice that **1** generates longer induction period than **2** used at the same concentration (see τ_1 and τ_2) and, additionally, the rate of O₂ uptake during the inhibition period is smaller for **1** than for **2**, demonstrating a superior antioxidant activity of the *ortho*-isomer. To better clarify the mechanism proposed in Scheme 3, we completely replaced styrene with CHD. Figure 1 B presents the kinetic traces recorded for the system, with the autoxidation propagated exclusively by HOO[•] radicals.^[22] Quinone **1** almost completely stopped the autoxidation of CHD (i.e. it trapped HOO[•] with high efficiency), while **2** was less effective even if used at much higher concentration. We also employed the reduced species, catechol/ hydroquinone, **1H₂** and **2H₂** to test their behavior during autoxidation of CHD. Results, reported in Figure 1 B, indicated a smaller and a higher HOO[•] trapping ability than the corresponding quinones for **1H₂** and **2H₂** respectively.^[26]

The use of CHD as the only oxidizable substrate simplifies the analysis of autoxidation kinetics, propagated only by HOO[•] [X = H in Eqs. (8) and (10)], since reactions (5) and (6)

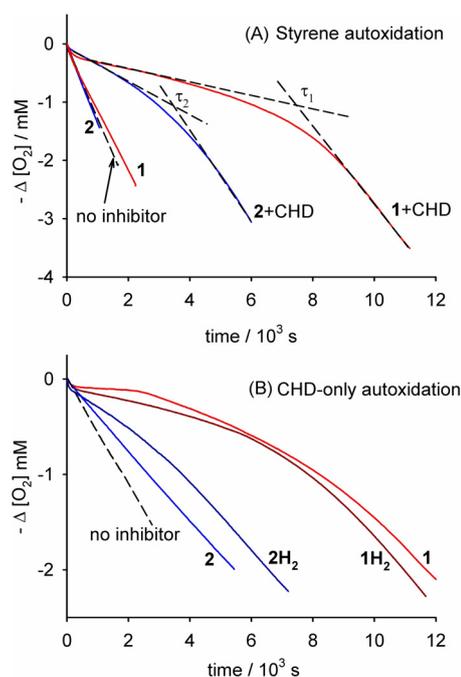
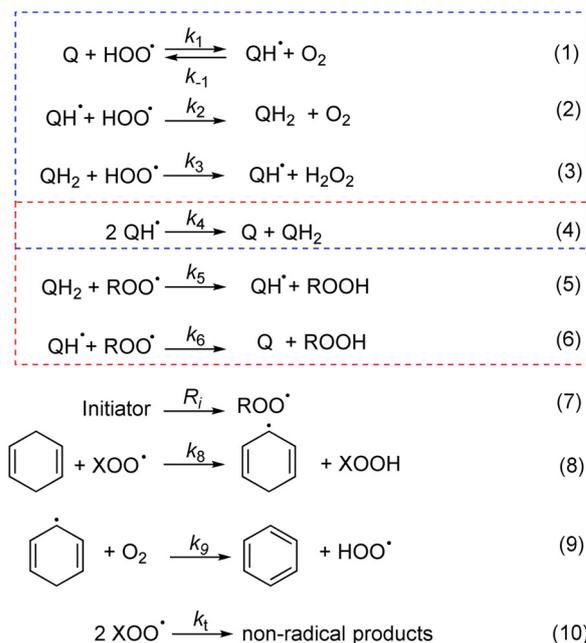


Figure 1. A) O₂ consumption during the autoxidation of styrene in chlorobenzene at 30 °C initiated with 25 mM AIBN in the absence of inhibitors (dashed line), and in the presence of quinones **1** and **2** (5 μM) with or without 0.023 M CHD. B) O₂ consumption during the autoxidation of CHD (0.23 M) initiated by 25 mM AIBN at 30 °C in chlorobenzene with no antioxidants (dashed line), or with added 40 μM of **2** or **2H₂**, and with 5 μM **1H₂** or **1**.



Scheme 3. Key reactions explaining the antioxidant activity of *ortho* and *para*-quinones, Q, in the presence of 1,4-cyclohexadiene (X = H or R).

become unimportant. Having clarified that the antioxidant activity of *ortho*-benzoquinone **1** stems from its reaction with HOO[•] [Eq. (1)], in the initial stages of autoxidation when the inhibitor is **1**, before a substantial accumulation of reduced **1H₂** [Eq. (2)], reaction (3) also becomes unimportant. There-

fore, the rate constant for reaction (1) can be determined in first approximation by using Equation (11), which relates the rates of the inhibited and non-inhibited autoxidation (R_{inh} and R_0 , respectively) to the rate constant for the reaction of the antioxidant with the chain-carrying radicals and the stoichiometry of radical trapping (n).^[25]

$$(R_0/R_{\text{in}}) - (R_{\text{in}}/R_0) = nk_1[\text{AH}]/(2k_t R_i)^{1/2} \quad (11)$$

The rate of initiation (R_i) was determined experimentally as $3.1 \times 10^{-9} \text{ Ms}^{-1}$, while $2k_t$ for CHD in chlorobenzene is $1.2 \times 10^9 \text{ M}^{-1} \text{ s}^{-1}$.^[22] With the assumptions that the rate of the back reaction k_{-1} is negligible and that $n=2$, k_1 was obtained as $(1.4 \pm 0.2) \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$. Numerical modeling of O_2 consumption traces of CHD and styrene autoxidations, inhibited either by **1** or **1H₂**, was then performed by using the full array of kinetic equations and the COPASI kinetic simulation software (see Supporting Information)^[27,28] as illustrated in Figure 2A. The optimized values of k_1 and k_3 obtained with this procedure were 1.5×10^7 and $1.5 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$, respectively, while for k_{-1} an upper limit of $65 \text{ M}^{-1} \text{ s}^{-1}$ was determined. Hence our results show that, counterintuitively, the oxidized quinone **1** is a far better antioxidant than catechol **1H₂**, but only in the presence of HOO^\bullet . Results also show that a substantial quantity of catechol **1H₂** is formed from **1** (Figure 2A). This finding is again counterintuitive, as it means that the antioxidant is being reduced, albeit transiently, during the inhibition of CHD autoxidation. Formation of **1H₂** was confirmed by ESI-MS analysis performed on the reaction mixture containing **1**, CHD and AIBN (Figure 2B,C). Quinone **1** was visible only in positive ion mode mainly as **1** + Na^+ peak, while catechol **1H₂** could be detected in negative ion mode as **1H⁻**. The same result was obtained by GC-MS, after derivatization of the sample with trimethylsilyl-N,N-dimethylcarbamate (TMSDMC) as silylating agent to protect **1H₂** from decomposition during the analysis (see Figures S5–S7).

Analysis of the kinetic solvent effect (KSE) offered further insight in the reaction of **1** with HOO^\bullet . In acetonitrile, the inhibiting effect of **1** during the autoxidation of CHD was

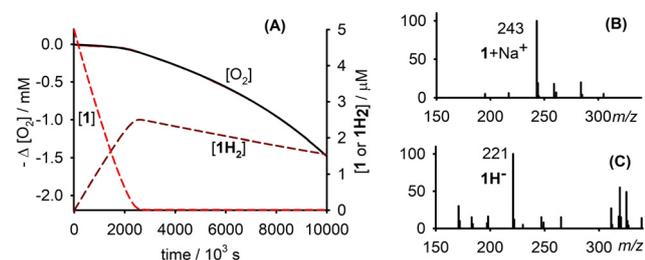
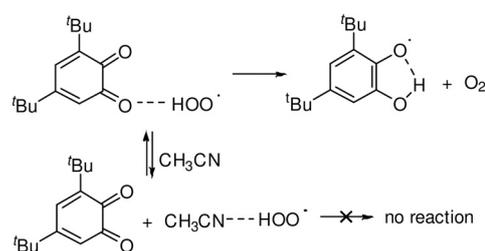


Figure 2. (A) Numerical fitting of the O_2 consumption traces during the autoxidation of CHD initiated by AIBN in PhCl at 30°C inhibited by **1**. Experimental results for oxygen consumption (black, solid line) and the simulated results (red line), perfectly overlapping each other. The transient concentrations of the quinone or hydroquinone species obtained during the simulations are reported. (B,C) ESI-MS spectra showing **1H₂** formation during the reaction of **1** (0.1 mM) with CHD (0.23 M) and AIBN (50 mM) in MeCN at 30°C ; after 1 hour of reaction in positive (B) and negative (C) ion mode.

weaker than in PhCl (see Figures S8 and S9). Numerical fitting provided $k_1 = 1.1 \times 10^5 \text{ M}^{-1} \text{ s}^{-1}$, that is 180-folds smaller than in PhCl. This type of KSE is well established for antioxidants whose mechanism relies on a rate-limiting formal H-atom transfer, which is hampered by occurrence of H-bonding to the solvent.^[13] Normally, when the antioxidant is a phenol transferring the H-atom to a peroxy radical, phenol itself is the H-bond donor (HBD), complexing to the solvent.^[13,29,30] Here, instead, the role is reverted and the HOO^\bullet radicals act as the HBD. From the observed magnitude of this KSE, the ability of HOO^\bullet as H-bond donor, H-bond acidity parameter α_2^{H} , can be calculated^[29] as 0.78, in reasonable agreement with previous estimates.^[20,31] KSE is a proof that the reaction of **1** with HOO^\bullet involves a H-atom transfer (HAT) and that this process starts with formation of a pre-reaction H-bond complex competing with the solvent (Scheme 4).



Scheme 4. Kinetic solvent effect for H-atom donation from HOO^\bullet to quinones.

Quantitative understanding of the reactions of these model structures could then be transferred to rationalize the behavior of PDA as a representative member of melanin biopolymers. Polydopamine (**PDA**) nanoparticles were synthesized by the oxidative polymerization of dopamine in alkaline water/ethanol solution and were accurately purified by repeated centrifugation cycles (characterization by DLS, TEM and FT-IR is reported in Figure 3, S10–S12).^[32,33] The nanoparticles were concentrated, dispersed in acetonitrile and then used as inhibitor in autoxidation studies. As shown in Figure 3C, **PDA** itself had a modest antioxidant activity toward styrene autoxidation (solvent MeCN), presumably due to the low concentration of catechol moieties, or their engagement in strong intramolecular H-bonds.^[34] Actually, on further extending the purification cycles, **PDA** showed even lower inhibition of autoxidation (Figure S13), suggesting that some catechol moieties could be physically trapped into the nanoparticle matrix, instead of being covalently bound, or could exist as small (extractable) oligomers or covalent adducts, such as the pyranoacridinetrione recently proposed.^[35] Interestingly, after the addition of CHD the antioxidant activity of **PDA** became very prominent (Figure 3C trace d). Under the same conditions, CHD had a negligible effect on **1H₂** (Figure 3D traces d and e), while it greatly increased the activity of **1** (traces c and f). The modest reactivity of **PDA** toward ROO^\bullet can be explained by considering that the catechol moieties are strongly bonded to H-bond acceptors, such as carbonyl groups, present into the

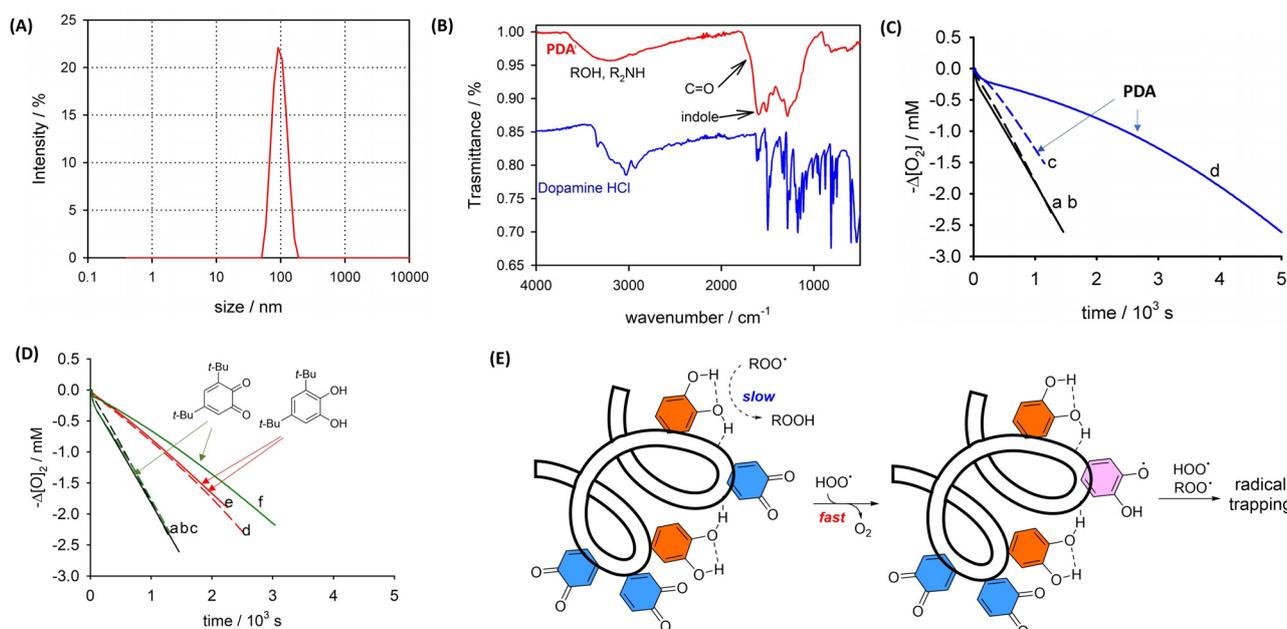


Figure 3. (A) Dynamic light scattering of **PDA** nanoparticles in water. (B) ATR-FTIR spectrum of dried **PDA**. (C) O_2 consumption recorded during the styrene (2.1 M) autoxidation initiated by AIBN (25 mM) in MeCN without inhibitors (a) and in the presence of: (b) CHD; (c) **PDA** ($25 \mu\text{g mL}^{-1}$); (d) **PDA** ($25 \mu\text{g mL}^{-1}$) + CHD. (D) Same conditions as C, without inhibitors (a) and in the presence of: (b) CHD; (c) **1** ($5 \mu\text{M}$); (d) **1H₂** ($5 \mu\text{M}$); (e) **1H₂** ($5 \mu\text{M}$) + CHD; (f) **1** ($5 \mu\text{M}$) + CHD. In all cases, [CHD] = 23 mM. (E) Catechol and quinone units in the polydopamine polymer are unreactive toward alkylperoxy radicals (ROO^\bullet) but upon the reaction with HOO^\bullet the quinones are converted to *ortho*-semiquinone radicals with enhanced ability to trap both species, HOO^\bullet or ROO^\bullet .

polymer (Figure 3E).^[36] Not H-bonded catechols which would be more reactive toward radicals than H-bonded ones,^[13] are most probably oxidized during the preparation of **PDA**. Instead, upon reaction with HOO^\bullet , *ortho*-quinones can form “exposed” semiquinones and catechol groups able to trap both HOO^\bullet and ROO^\bullet .

To rationalize the antioxidant activity of **PDA**, the driving force for the H-atom transfer reaction of HOO^\bullet to **1**, **2** and to the quinones relevant to the chemistry of **PDA** (**3–6**) was calculated (see Figure 4, S14–S17). The reaction of HOO^\bullet with **1** is much more exothermic than with **2**, in agreement with the results from CHD autoxidation studies (see Figure 1) and with previous reports.^[37,38] Dopaminochrome (**4**) (tran-

siently formed during **PDA** synthesis)^[7] doesn't react as efficiently with HOO^\bullet as dopaminoquinone (**3**) and indole-5,6-quinone (**5**). Interestingly, the indole-5,6-quinone-5,6-dihydroxyindole dimer (**6**), proposed as building block of **PDA**, shows a high reactivity toward HOO^\bullet , fully consistently with experimental results. Moreover, *ortho*-semiquinones are stabilized by a strong intramolecular H-bond and are therefore generally unreactive toward O_2 .^[37] These calculations further support our proposed mechanism to explain the antioxidant behavior of **PDA**. Regarding the semiquinone-type radicals naturally hosted in **PDA** (and in other melanins), contributing to the persistent EPR signal,^[4,5] their rapid reduction to catechols by HOO^\bullet [Eq. (2)]—a reaction expected to be diffusion controlled from our numerical fittings—is also likely to have a role in the enhanced antioxidant behavior in the presence of HOO^\bullet , although its actual contribution might deserve further investigation.

The simultaneous presence of alkylperoxy and hydroperoxy radicals is a common feature in biological systems.^[39] Alkylperoxy radicals are responsible for lipid peroxidation of unsaturated membranes,^[24] while the HOO^\bullet radicals are constantly produced by protonation of superoxide leaking from the mitochondrial respiratory chain and by the autoxidation of alcohols, aliphatic amines and alkenes.^[21,23,40] The chemistry disclosed herein is an important step to rationalize the redox-mediated bioactivity of **PDA** and the prominent antioxidant chemistry of *ortho* and *para*-quinones. With the caution imposed by their distinctive chemical diversity,^[14] it could also serve as a rational basis to understand the properties of other melanins and it might be implemented

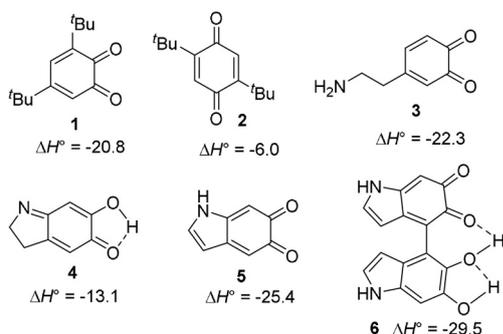


Figure 4. Calculated enthalpy variation for the reaction of H-atom transfer from HOO^\bullet to quinones [reaction (1)] in kcal mol^{-1} . The most stable tautomers of the quinones and of the semiquinone radicals are considered (level: CBS-QB3 or B3LYP/6-311 + + g(d,p) for **6**, gas phase).

for the rational design of novel antioxidant biomaterials that can be selectively activated by hydroperoxyl radicals.

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Conflict of interest

The authors declare no conflict of interest.

Keywords: antioxidants · peroxy radicals · polydopamine · quinone · radical reactions

- [1] J. H. Ryu, P. B. Messersmith, H. Lee, *ACS Appl. Mater. Interfaces* **2018**, *10*, 7523–7540.
- [2] Y. Liu, K. Ai, L. Lu, *Chem. Rev.* **2014**, *114*, 5057–5115.
- [3] J. Liebscher, R. Mrówczyński, H. A. Scheidt, C. Filip, N. D. Hädade, R. Turcu, A. Bende, S. Beck, *Langmuir* **2013**, *29*, 10539–10548.
- [4] P. Manini, V. Lino, P. Franchi, G. Gentile, T. Sibillano, C. Giannini, E. Picardi, A. Napolitano, L. Valgimigli, C. Chiappe, M. D'Ischia, *ChemPlusChem* **2019**, *84*, 1331–1337.
- [5] L. Panzella, G. Gentile, G. D'Errico, N. F. D. Vecchia, M. E. Errico, A. Napolitano, C. Carfagna, M. D'Ischia, *Angew. Chem. Int. Ed.* **2013**, *52*, 12684–12687; *Angew. Chem.* **2013**, *125*, 12916–12919.
- [6] I. Carballo-Carbajal, A. Laguna, J. Romero-Giménez, T. Cuadros, J. Bové, M. Martínez-Vicente, A. Parent, M. González-Sepulveda, N. Peñuelas, A. Torra, B. Rodríguez-Galván, A. Ballabio, T. Hasegawa, A. Bortolozzi, E. Gelpi, M. Vila, *Nat. Commun.* **2019**, *10*, 973.
- [7] E. Monzani, S. Nicolis, S. Dell'Acqua, A. Capucciati, C. Bacchella, F. A. Zucca, E. V. Mosharov, D. Sulzer, L. Zecca, L. Casella, *Angew. Chem. Int. Ed.* **2019**, *58*, 6512–6527; *Angew. Chem.* **2019**, *131*, 6580–6596.
- [8] T. Feng, W. Ji, Y. Zhang, F. Wu, Q. Tang, H. Wei, L. Mao, M. Zhang, *Angew. Chem. Int. Ed.* **2020**, *59*, 23445–23449; *Angew. Chem.* **2020**, *132*, 23651–23655.
- [9] P. Yang, Z. Gu, F. Zhu, Y. Li, *CCS Chem.* **2020**, *2*, 128–138.
- [10] H. Zhao, Z. Zeng, L. Liu, J. Chen, H. Zhou, L. Huang, J. Huang, H. Xu, Y. Xu, Z. Chen, Y. Wu, W. Guo, J. H. Wang, J. Wang, Z. Liu, *Nanoscale* **2018**, *10*, 6981–6991.
- [11] Y. Liang, X. Zhao, T. Hu, Y. Han, B. Guo, *J. Colloid Interface Sci.* **2019**, *556*, 514–528.
- [12] Y. Liu, K. Ai, X. Ji, D. Askhatova, R. Du, L. Lu, J. Shi, *J. Am. Chem. Soc.* **2017**, *139*, 856–862.
- [13] G. Litwinienko, K. U. Ingold, *Acc. Chem. Res.* **2007**, *40*, 222–2.
- [14] M. d'Ischia, A. Napolitano, V. Ball, C.-T. Chen, M. J. Buehler, *Acc. Chem. Res.* **2014**, *47*, 3541–3550.
- [15] N. F. Della Vecchia, A. Luchini, A. Napolitano, G. D'Errico, G. Vitiello, N. Szekely, M. d'Ischia, L. Paduano, *Langmuir* **2014**, *30*, 9811–9818.
- [16] C.-T. Chen, M. J. Buehler, *Phys. Chem. Chem. Phys.* **2018**, *20*, 28135–28143.
- [17] K. Jodko-Piórecka, G. Litwinienko, *J. Free Radicals Biol. Med.* **2015**, *83*, 1–11.
- [18] R. Amorati, A. Baschieri, G. Morroni, R. Gambino, L. Valgimigli, *Chem. Eur. J.* **2016**, *22*, 7924–7934.
- [19] R. Amorati, L. Valgimigli, L. Panzella, A. Napolitano, M. d'Ischia, *J. Org. Chem.* **2013**, *78*, 9857–9864.
- [20] J. Cedrowski, G. Litwinienko, A. Baschieri, R. Amorati, *Chem. Eur. J.* **2016**, *22*, 16441–16445.
- [21] A. Baschieri, L. Valgimigli, S. Gabbanini, G. A. DiLabio, E. Romero-Montalvo, R. Amorati, *J. Am. Chem. Soc.* **2018**, *140*, 10354–10362.
- [22] J. A. Howard, K. U. Ingold, *Can. J. Chem.* **1967**, *45*, 785–792.
- [23] E. T. Denisov, *Russ. Chem. Rev.* **1996**, *65*, 505–520.
- [24] G. W. Burton, K. U. Ingold, *J. Am. Chem. Soc.* **1981**, *103*, 472–6477.
- [25] R. Amorati, A. Baschieri, L. Valgimigli, *J. Chem.* **2017**, *2017*, 1–12.
- [26] H-atom abstraction from CHD by the semiquinone radicals of the investigated compounds (i.e. $\text{QH}^\bullet + \text{CHD} \rightarrow \text{QH}_2 + \text{CHD}_{\cdot\text{H}}$) was ruled out because when performing inhibited autoxidation studies at different CHD concentrations, the same profile of O_2 consumption was obtained. The direct reaction of **1** with CHD was found to be marginal because of the low [CHD] used. See: A. Baschieri, R. Amorati, L. Valgimigli, L. Sambri, *J. Org. Chem.* **2019**, *84*, 13655–13664.
- [27] S. Hoops, S. Sahle, R. Gauges, C. Lee, J. Pahle, N. Simus, M. Singhal, L. Xu, P. Mendes, U. Kummer, *Bioinformatics* **2006**, *22*, 3067–3074.
- [28] R. Amorati, P. T. Lynett, L. Valgimigli, D. A. Pratt, *Chem. Eur. J.* **2012**, *18*, 6370–6379.
- [29] D. W. Snelgrove, J. Lusztyk, J. T. Banks, P. Mulder, K. U. Ingold, *J. Am. Chem. Soc.* **2001**, *123*, 469–477.
- [30] M. Jha, D. A. Pratt, *Chem. Commun.* **2008**, 1252–1254.
- [31] M. C. Foti, S. Sortino, K. U. Ingold, *Chem. Eur. J.* **2005**, *11*, 1942–1948.
- [32] Y. Huang, Y. Li, Z. Hu, X. Yue, M. T. Proetto, Y. Jones, N. C. Gianneschi, *ACS Cent. Sci.* **2017**, *3*, 564–5699.
- [33] X. Jiang, Y. Wang, M. Li, *Sci. Rep.* **2015**, *4*, 6070.
- [34] M. C. Foti, L. R. C. Barclay, K. U. Ingold, *J. Am. Chem. Soc.* **2002**, *124*, 12881–12888.
- [35] M. L. Alfieri, R. Micillo, L. Panzella, O. Crescenzi, S. L. Oscurato, P. Maddalena, A. Napolitano, V. Ball, M. d'Ischia, *ACS Appl. Mater. Interfaces* **2018**, *10*, 7670–7680.
- [36] X. Zhang, C. Erb, J. Flammer, W. M. Nau, *Photochem. Photobiol.* **2000**, *71*, 524–533.
- [37] L. Valgimigli, R. Amorati, M. G. Fumo, G. A. DiLabio, G. F. Pedulli, K. U. Ingold, D. A. Pratt, *J. Org. Chem.* **2008**, *73*, 1830–1841.
- [38] H. G. Korth, P. Mulder, *J. Org. Chem.* **2020**, *85*, 2560–2574.
- [39] J.-F. Poon, O. Zilka, D. A. Pratt, *J. Am. Chem. Soc.* **2020**, *142*, 14331–14342.
- [40] K. A. Harrison, E. A. Haidasz, M. Griesser, D. A. Pratt, *Chem. Sci.* **2018**, *9*, 6068–6079.

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