

Supporting Information

PHOTOREDOX PROPARGYLATION OF ALDEHYDES CATALYTIC IN TITANIUM.

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Reaction set-up

Figure S1. Reaction setup

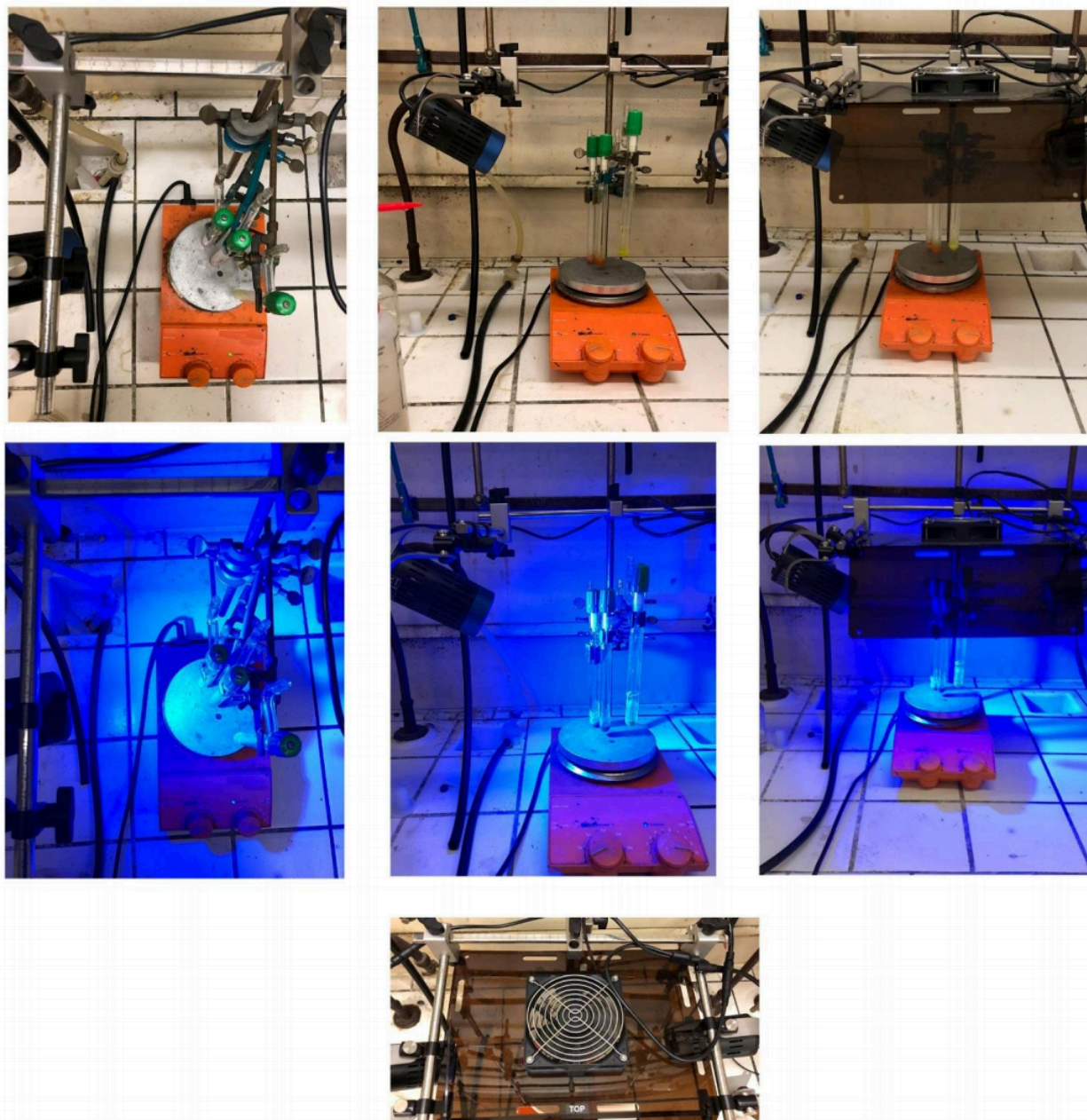
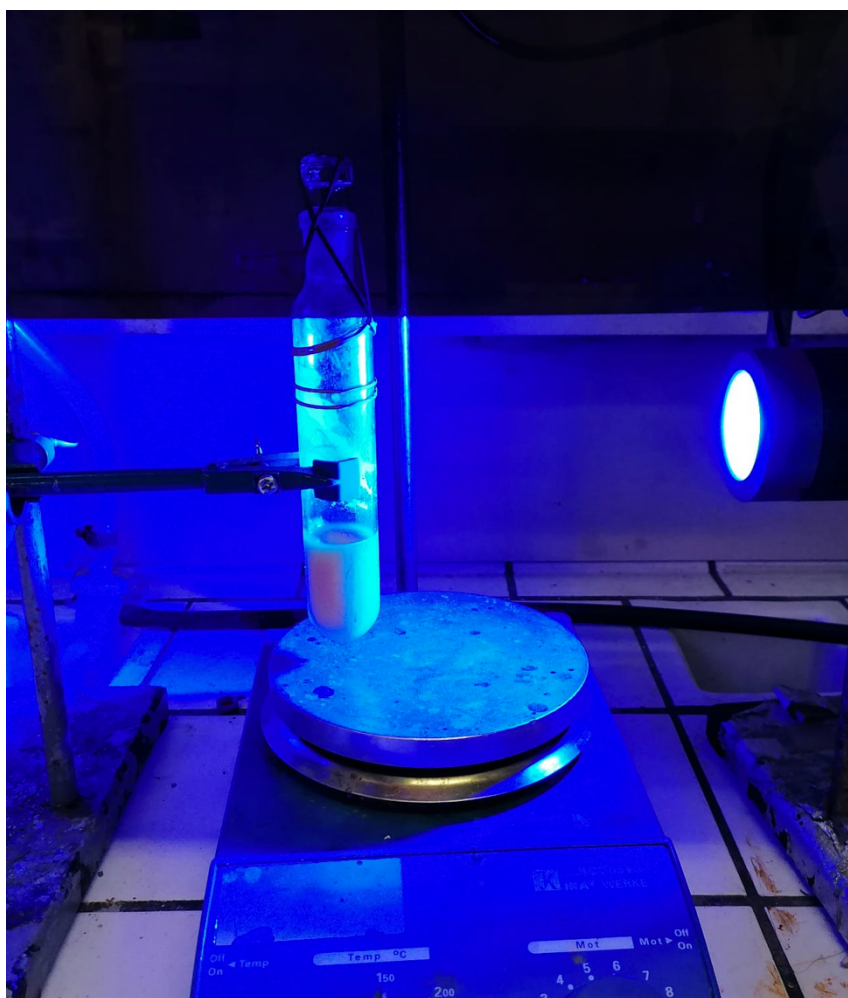


Figure S2. Reaction set-up with Kessil® PR160L@456 nm lamp for 1 mmol scale.



The reaction flasks were positioned approximately at 10 cm from the light source and Kessil® PR160 Rig with Fan Kit¹ was used to control the temperature. The reaction temperature was close to room temperature during the irradiation as measured with a thermometer at 1 cm from reaction flask.

Photophysical and mechanistic studies

All the photophysical analyses were carried out in air-equilibrated tetrahydrofuran at 298 K unless otherwise specified. UV-vis absorption spectra were recorded with a PerkinElmer λ 40 spectrophotometer using quartz cells with optical path length of 1.0 cm. Degassed solutions are obtained by means of repeated pump-freeze-thaw cycles (ca. $4 \cdot 10^{-6}$ mbar) in sealed quartz cuvettes. Luminescence spectra were performed with a PerkinElmer LS-50, a Varian Cary Eclipse or an Edinburgh FLS920 spectrofluorimeter equipped with a Hamamatsu R928 phototube. Lifetimes shorter than 10 μ s were measured by the above-mentioned Edinburgh FLS920 spectrofluorimeter equipped with a TCC900 card for data acquisition in time-correlated single-photon counting experiments (0.5 ns time resolution). The estimated experimental errors are 2 nm on the band maximum, 5% on the molar absorption coefficient and luminescence lifetime.

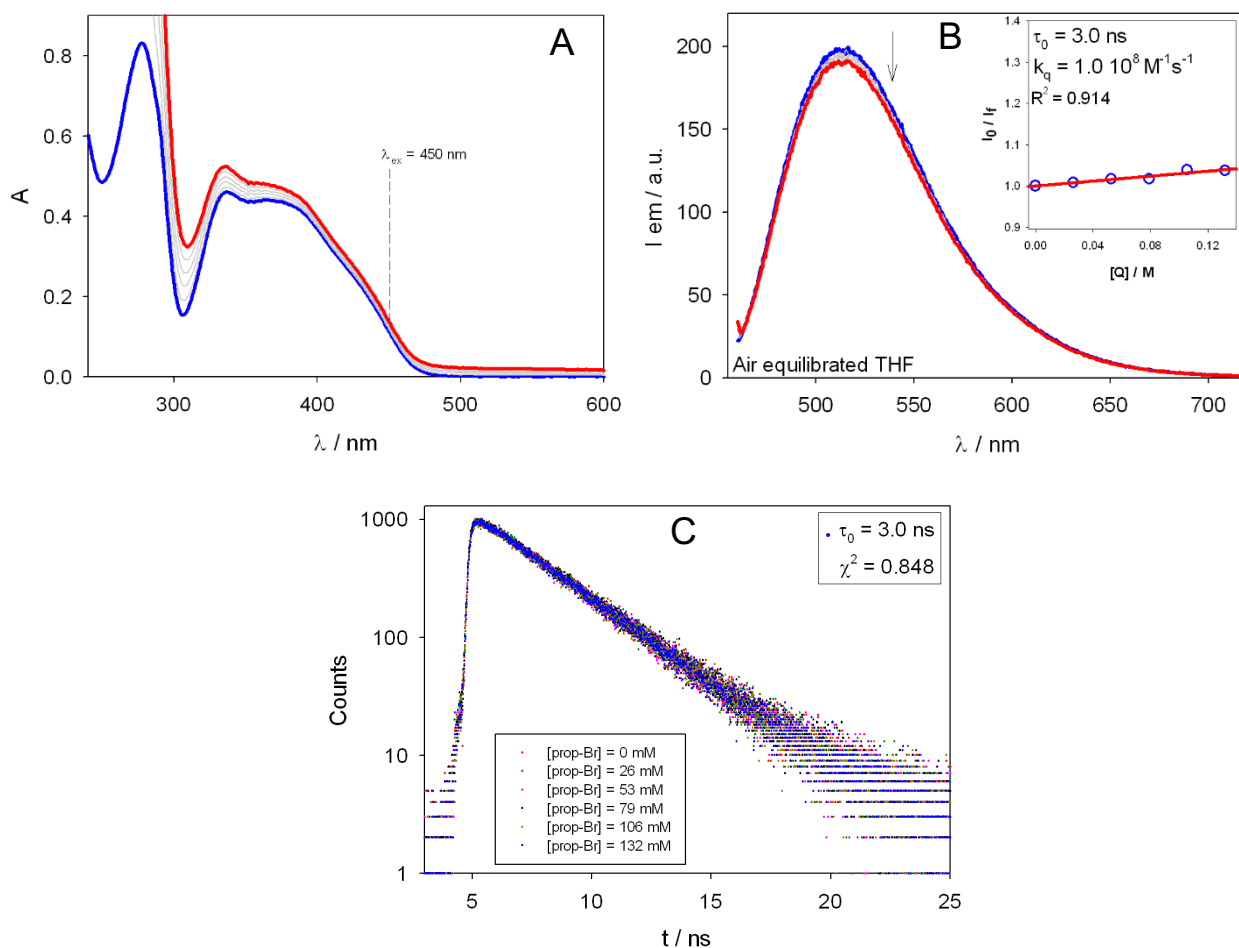


Figure S3. A: absorption spectra of solutions of 3-DPAFIPN in air-equilibrated THF (blue line) obtained upon addition of increasing amounts of propargyl bromide (prop-Br, up to ca. 0.13 M, red line). B: fluorescence spectra of 3-DPAFIPN obtained from the same solutions at $\lambda_{em} = 510$ nm. Inset: Stern-Volmer diagram relative to the fluorescence spectra shown in B. C: fluorescence decays of 3-DPAFIPN obtained from the same solutions as B, at $\lambda_{em} = 510$ nm ($\lambda_{ex} = 405$ nm). Since the fluorescence lifetime of 3-DPAFIPN is not significantly changing upon increasing additions of propargyl bromide, no Stern-Volmer constant has been calculated.

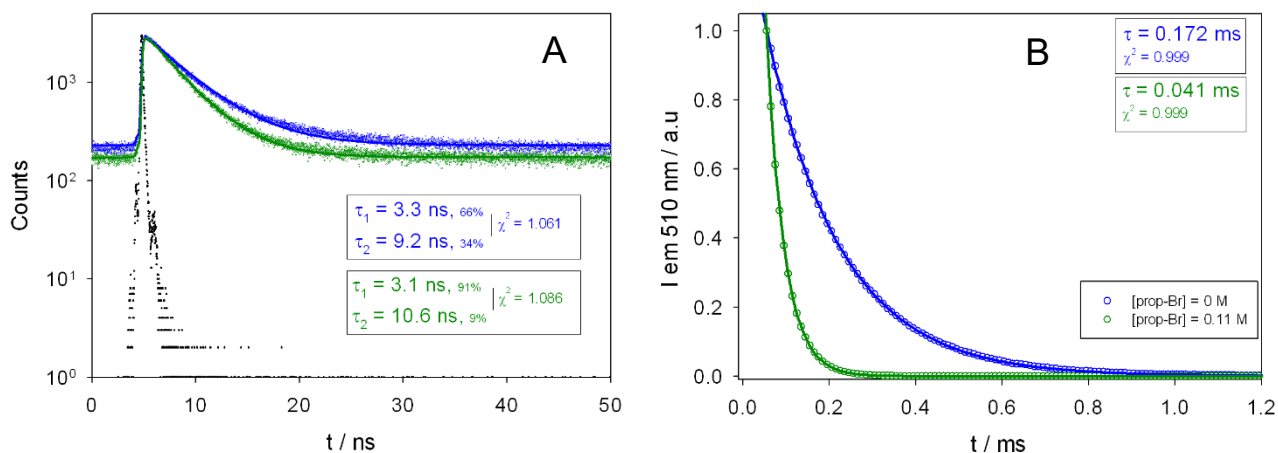
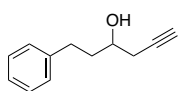
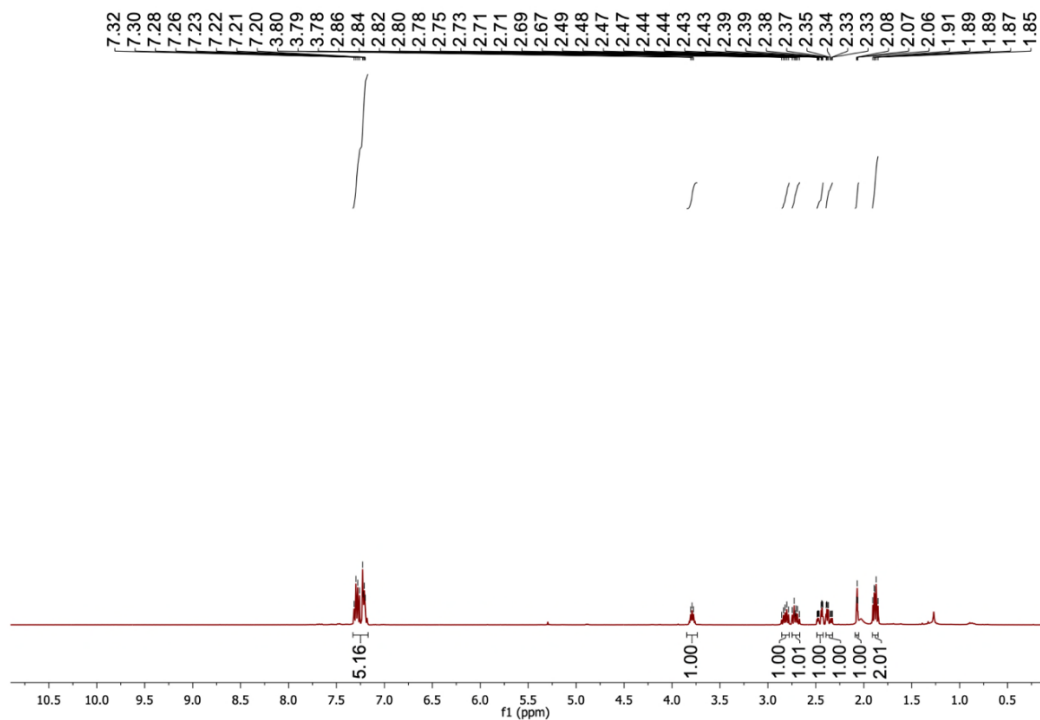


Figure S4. A: prompt fluorescence decays of **3-DPAFIPN** obtained from degassed THF solutions at $\lambda_{em}=510$ nm (pump-freeze-thaw, 3 cycles; $\lambda_{ex}=405$ nm) in the absence and in the presence of **prop-Br** 0.11 M (blue and green dots, respectively). The corresponding biexponential fitting curves are shown as the blue and green solid lines. The instrument response function is also shown (black dots). **B:** normalised emission decays of **3-DPAFIPN** in degassed THF solutions (pump-freeze-thaw, 3 cycles) in the absence (blue dots) and in the presence of **prop-Br** 0.11 M (green dots). The decays are fitted with monoexponential functions (blue and green solid lines, respectively). $\lambda_{ex}=350$ nm, $\lambda_{em}=510$ nm; delay=0.05 ms; gate=0.01 ms. Based on these emission lifetimes, the quenching constant k_q is estimated as ca. $1.8 \cdot 10^5 \text{ M}^{-1}\text{s}^{-1}$.

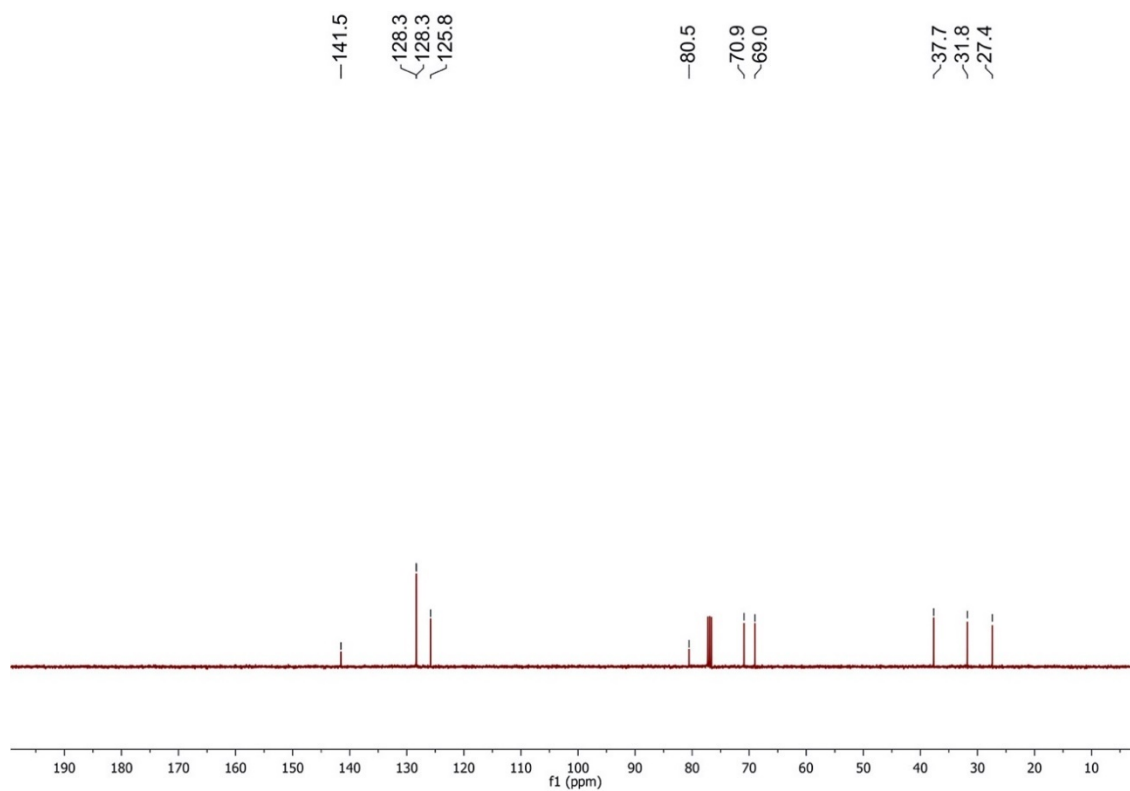
Copies of NMR spectra

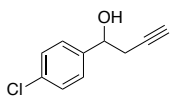


3a ^1H NMR (401 MHz, CDCl_3)

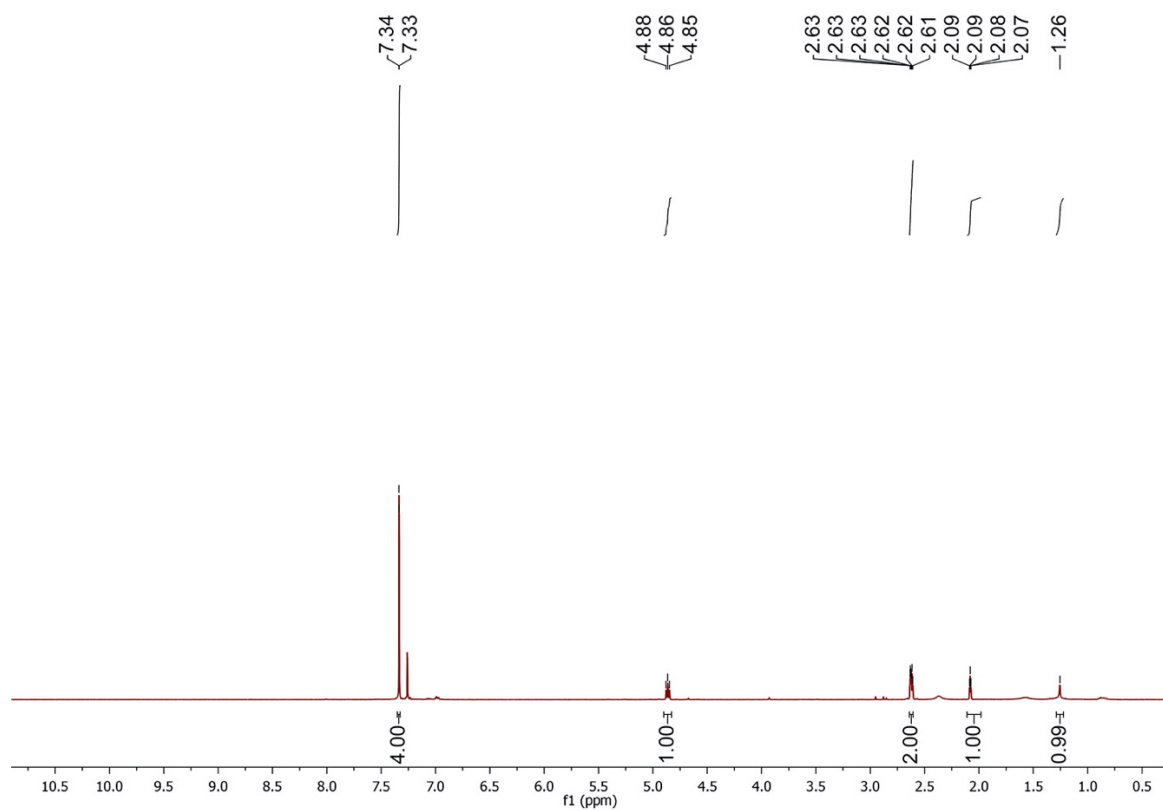


$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)

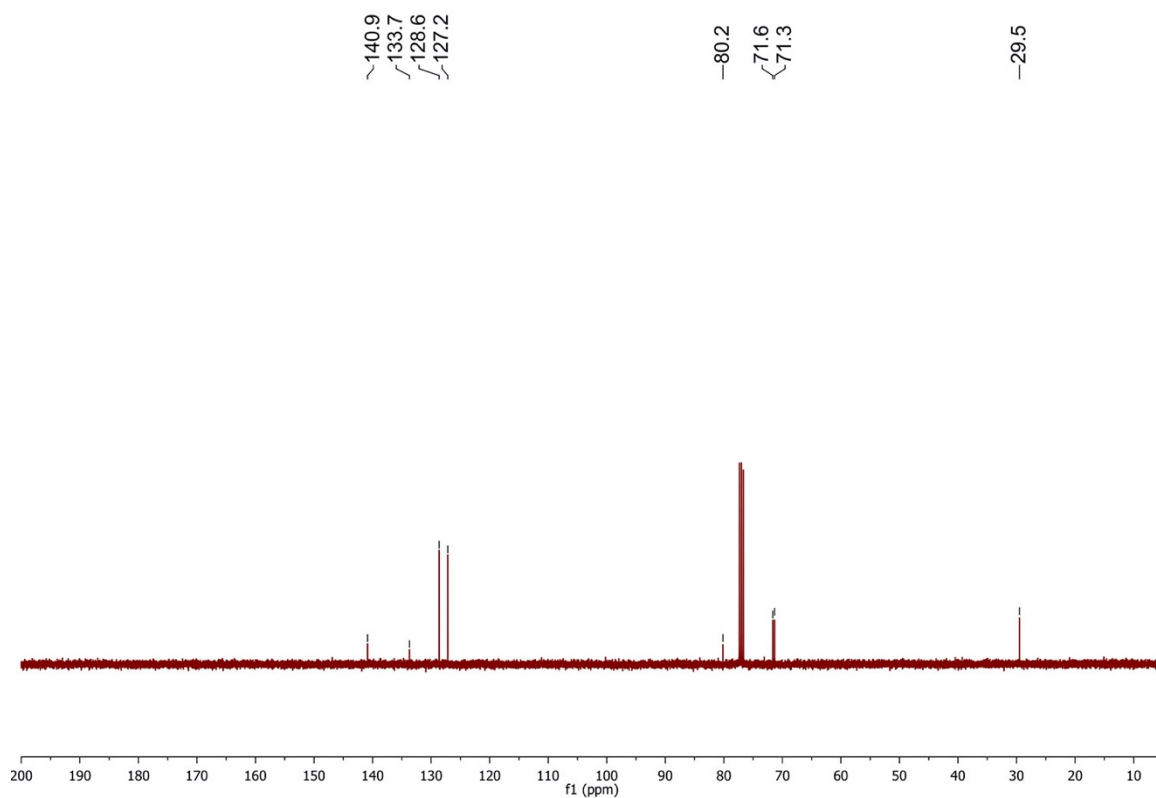


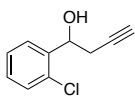


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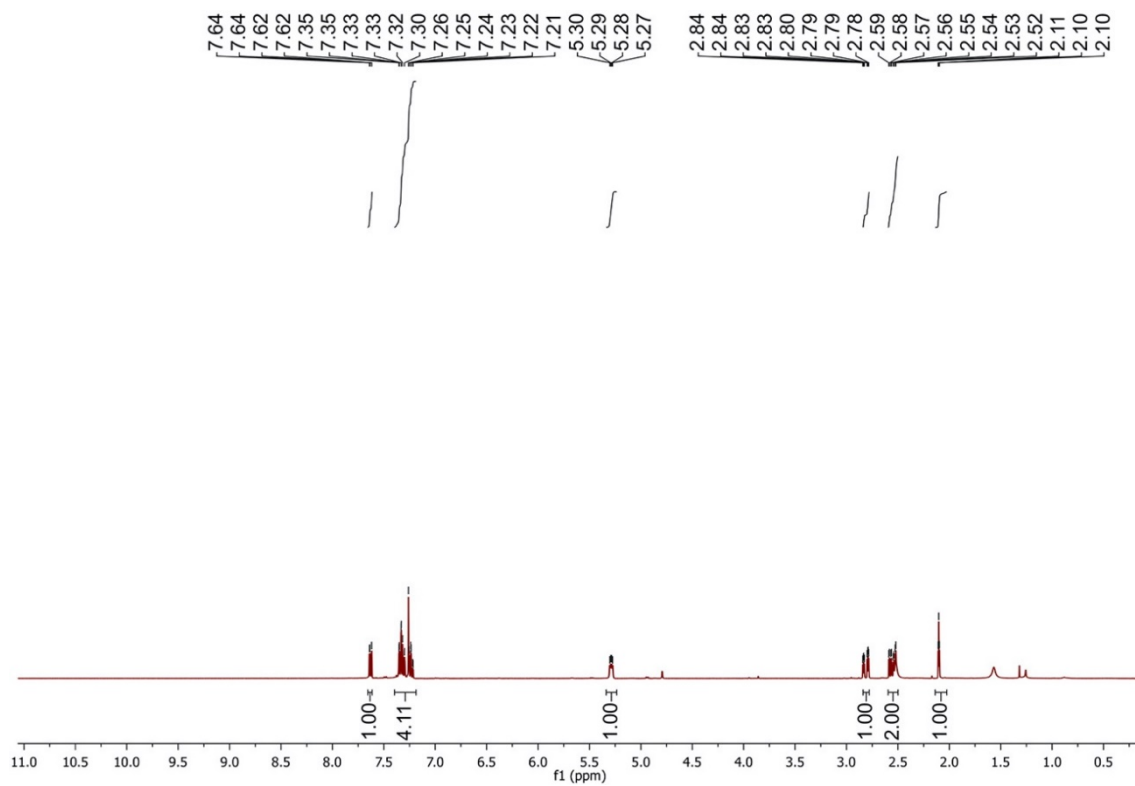


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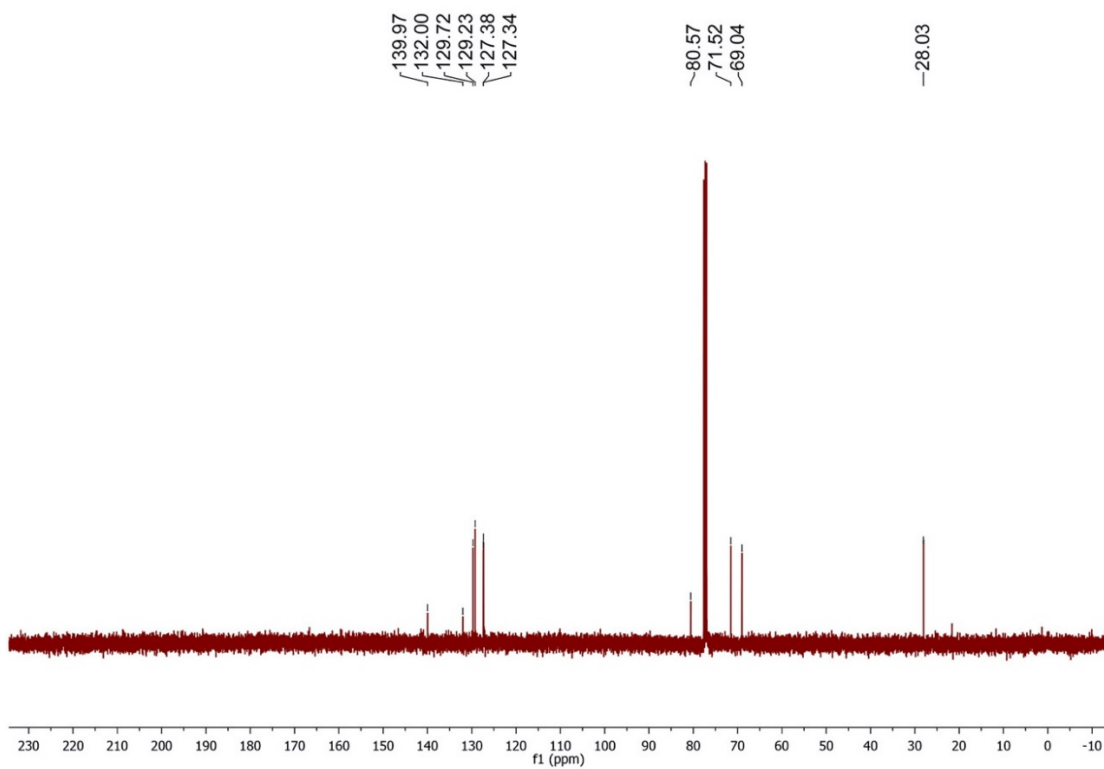


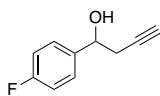


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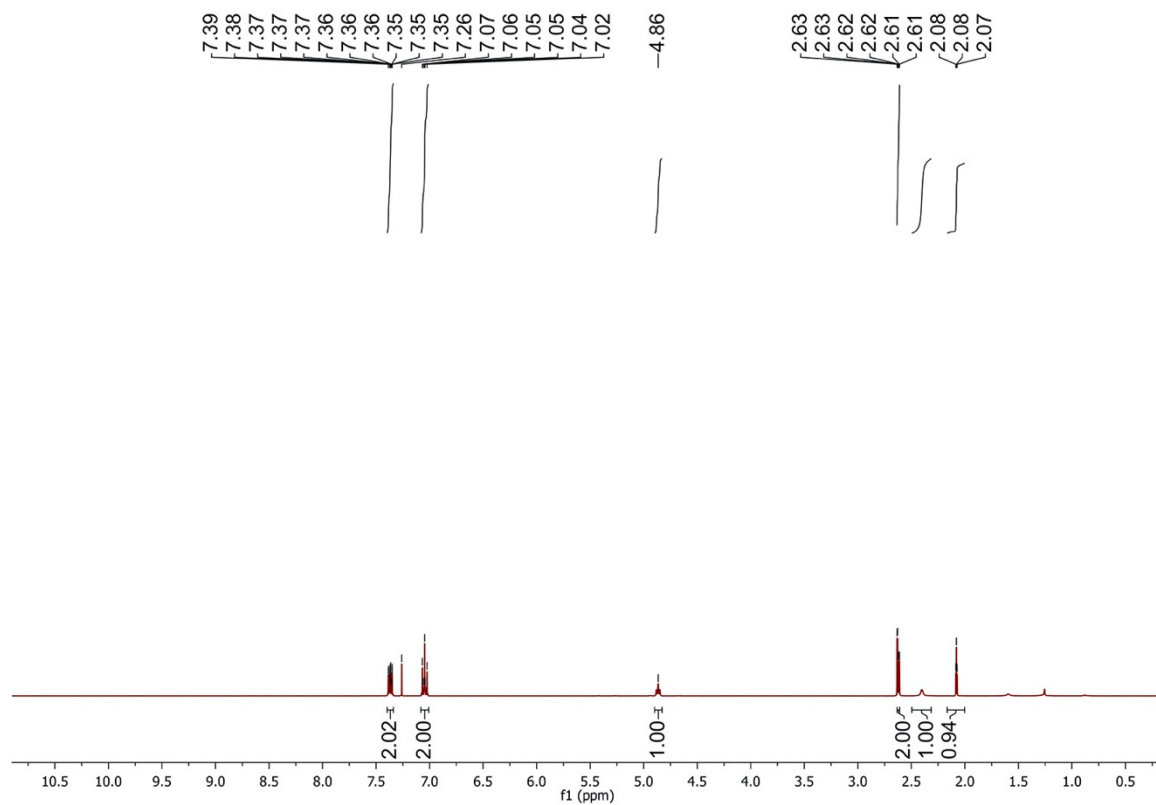


$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)

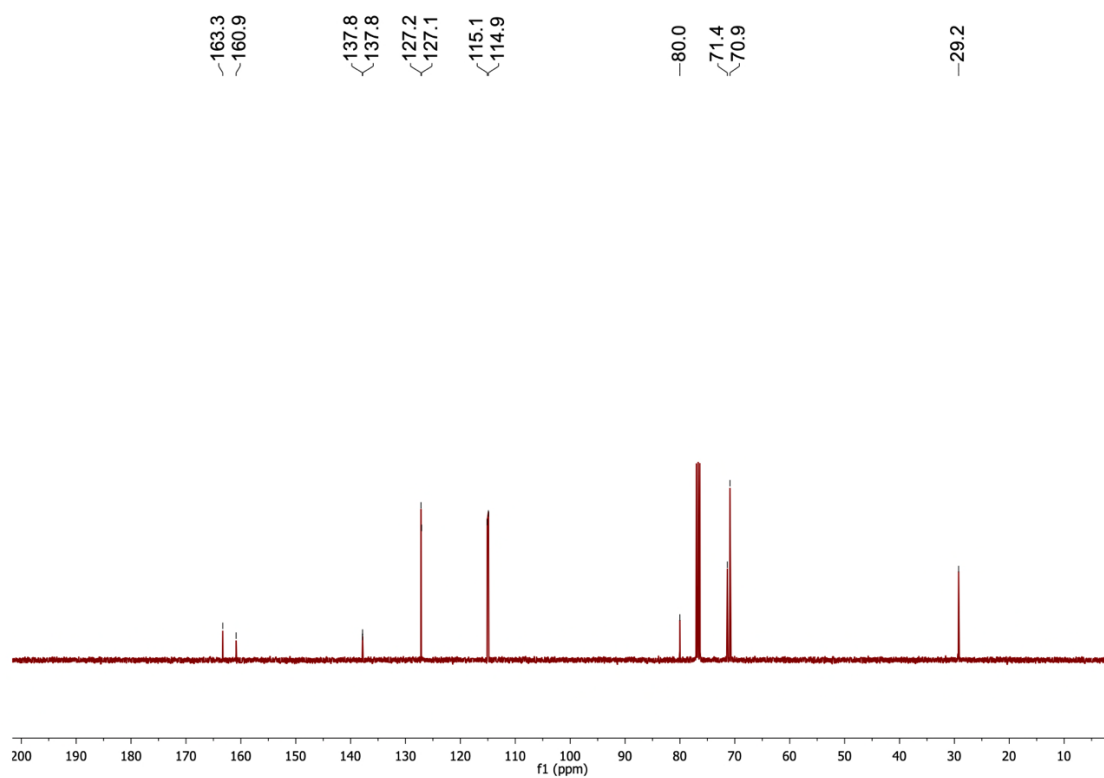




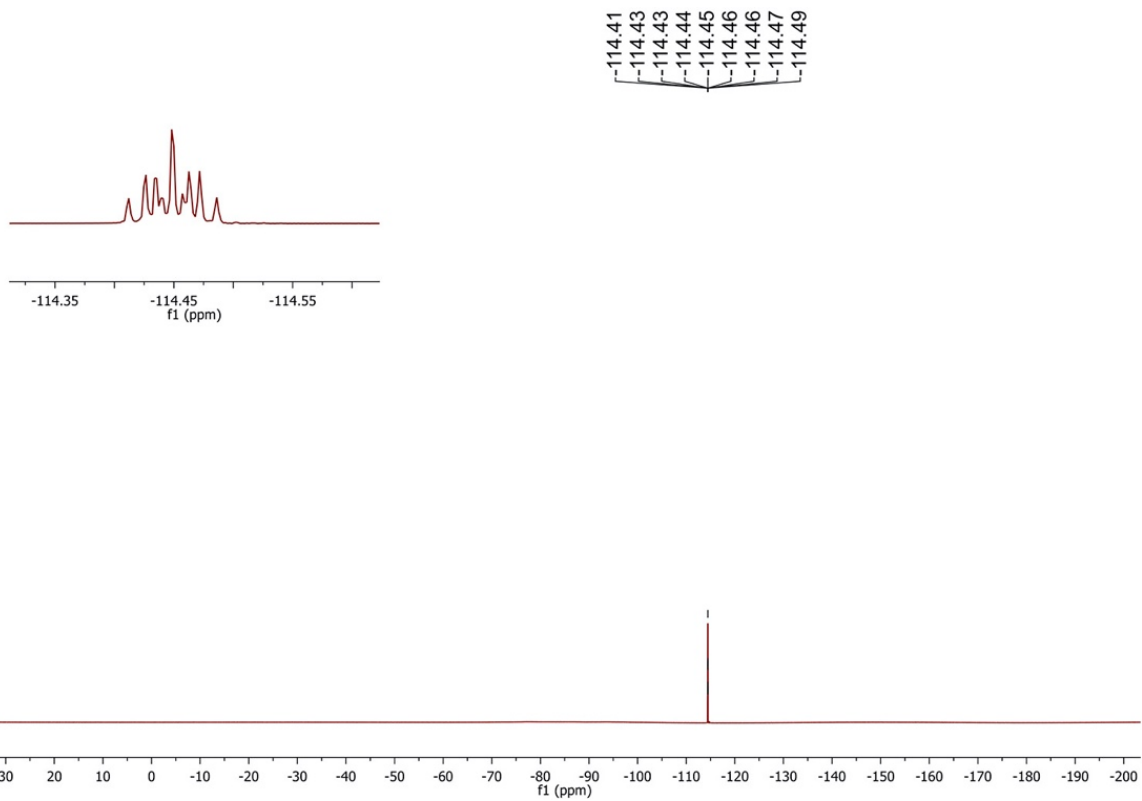
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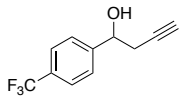


$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)

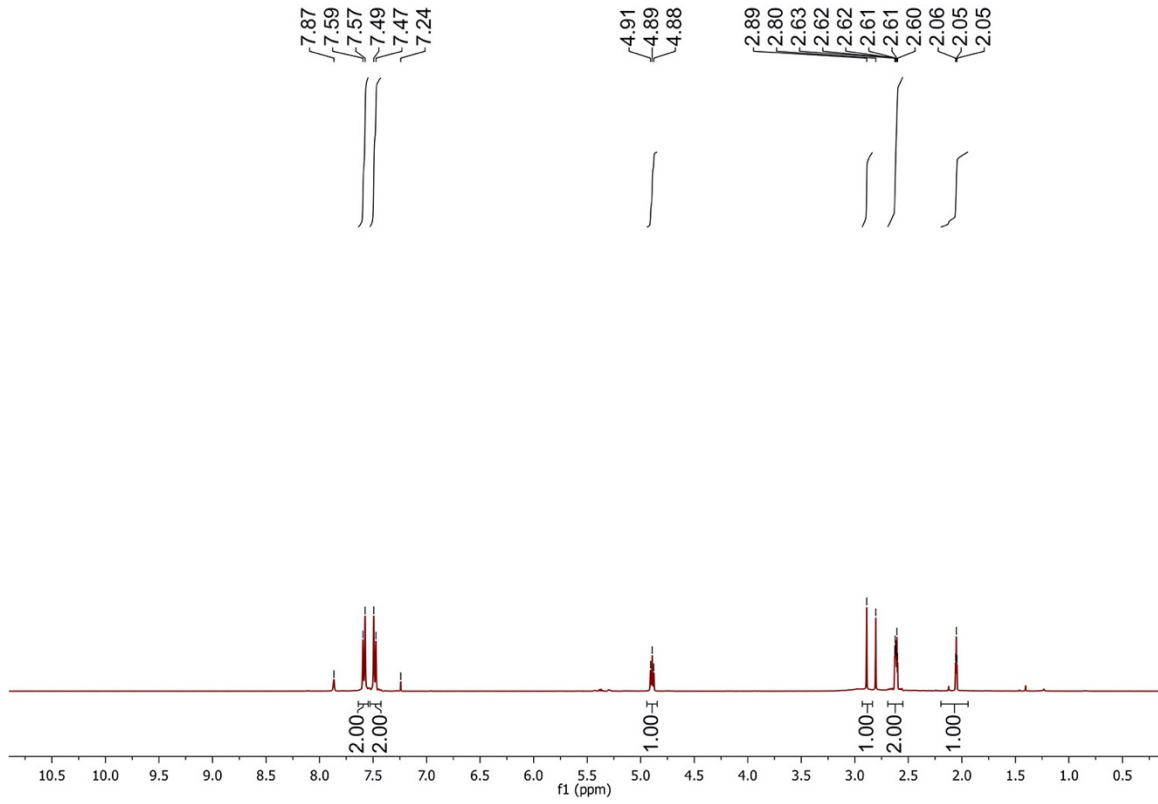


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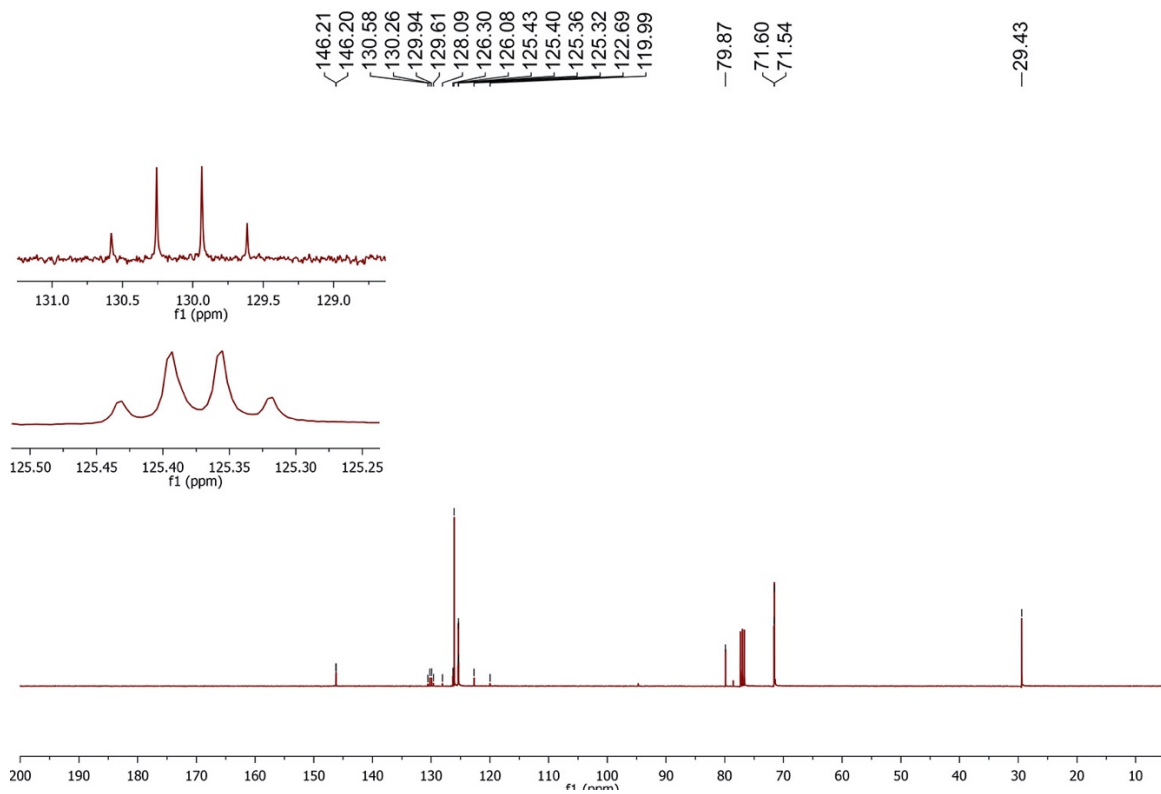




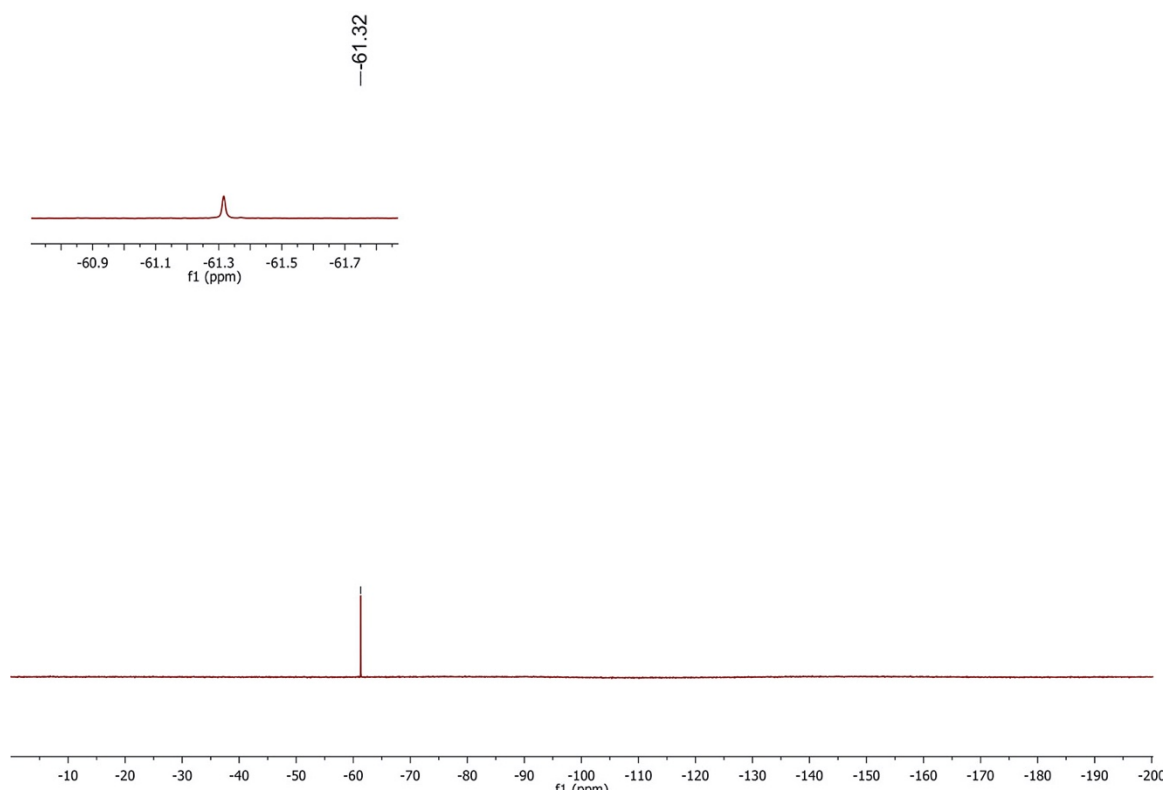
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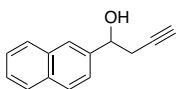


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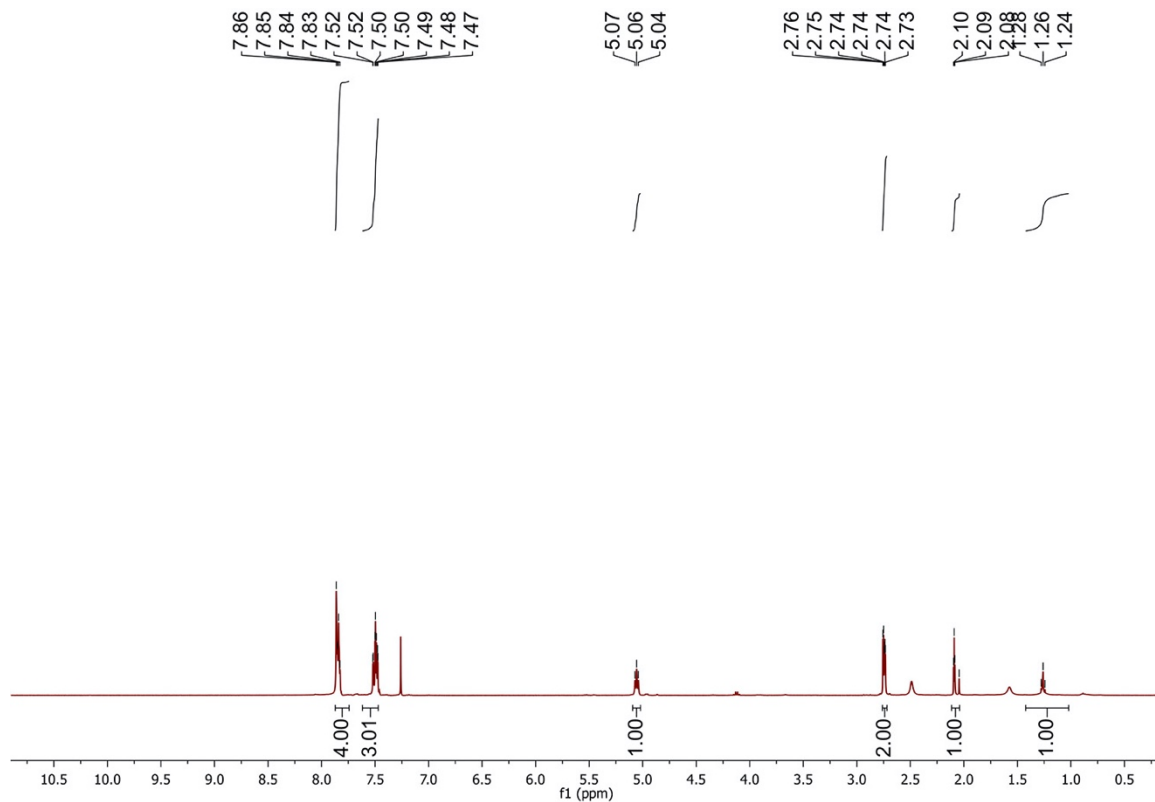


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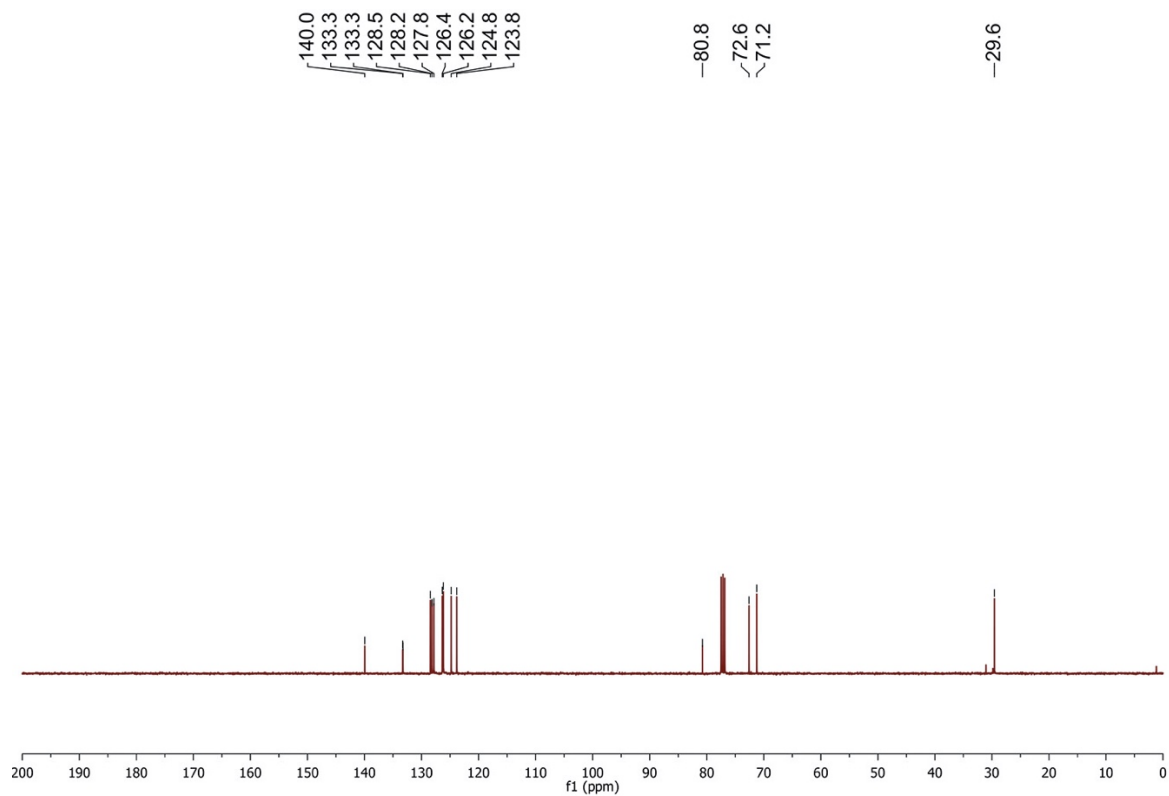


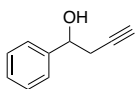


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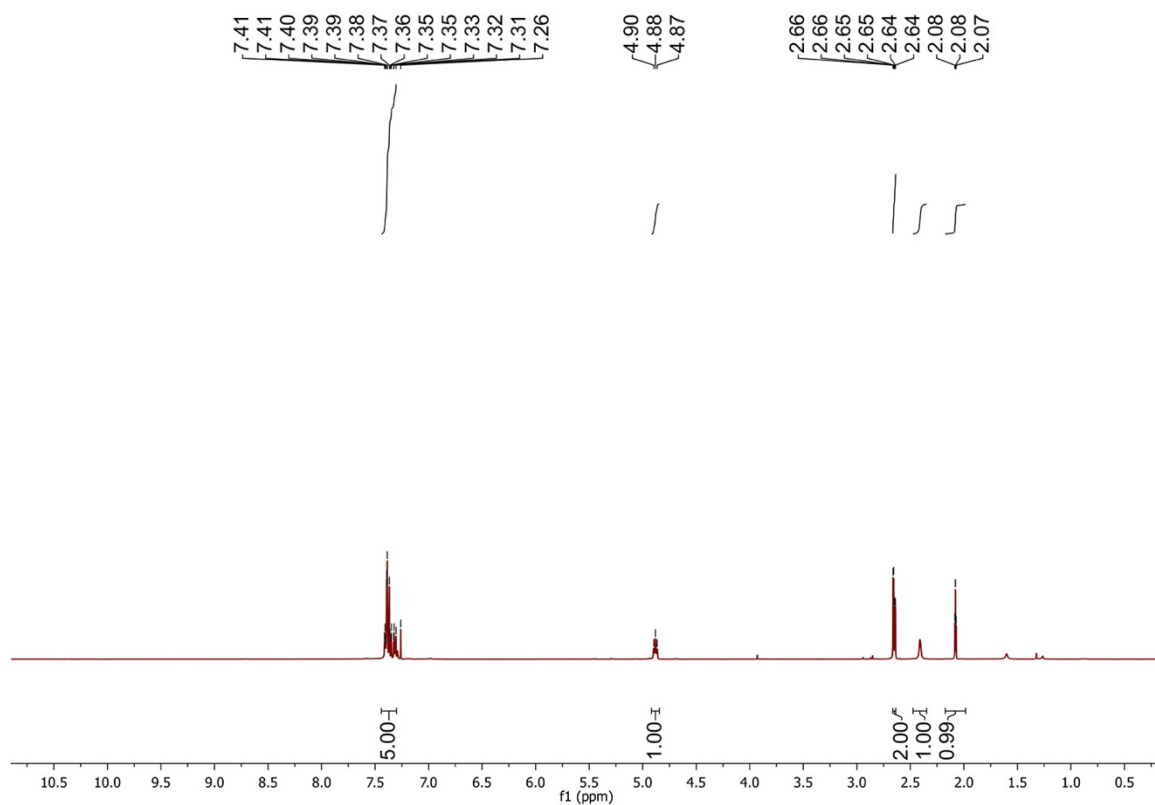


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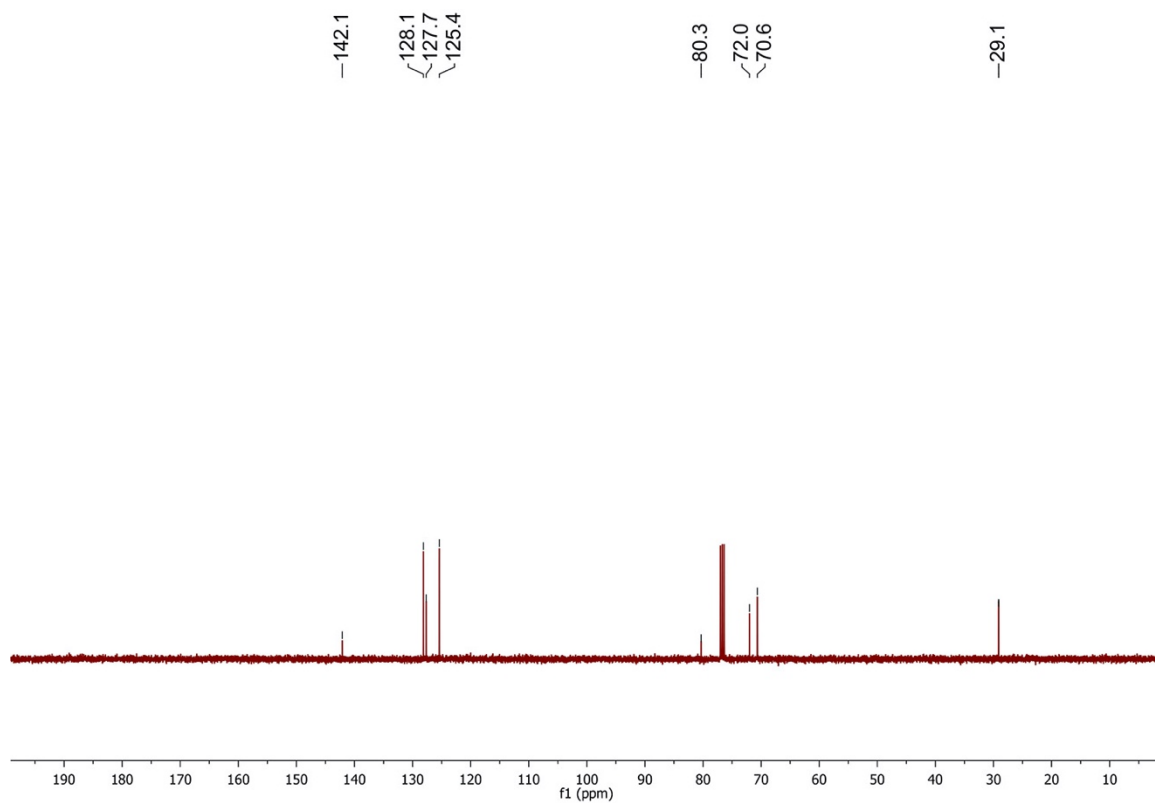


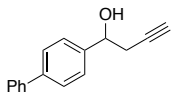


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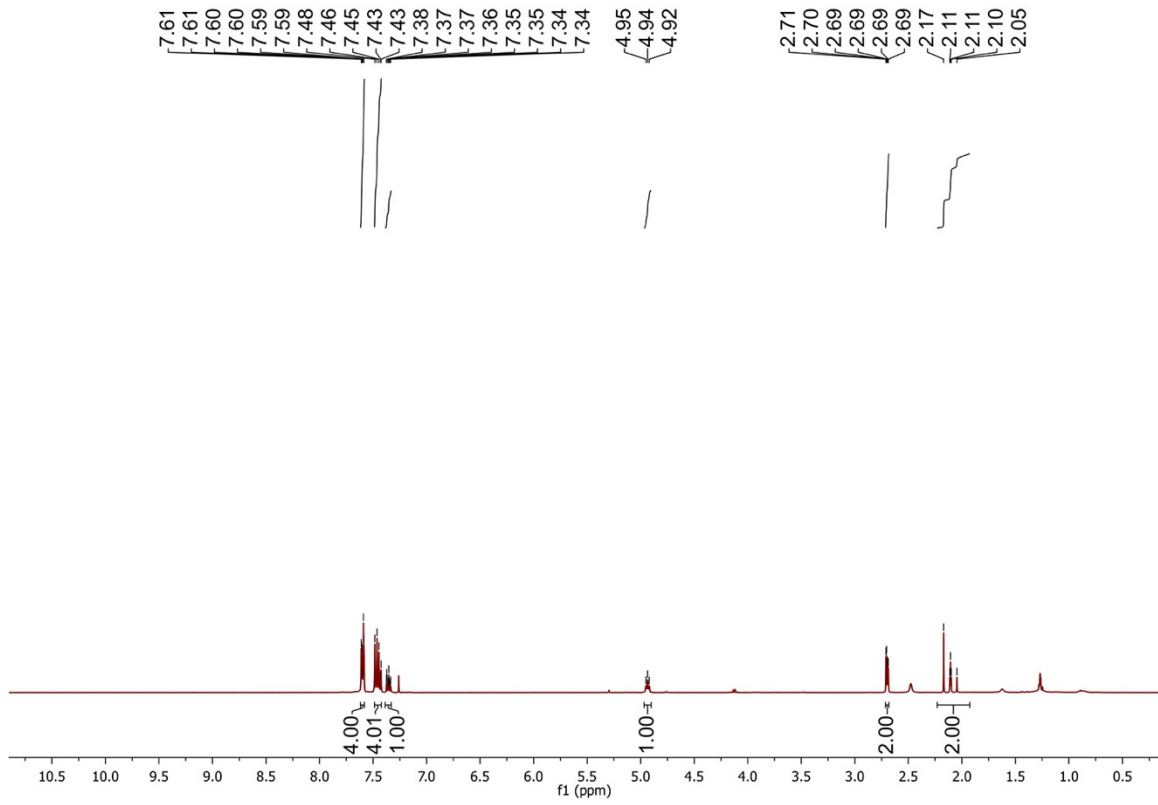


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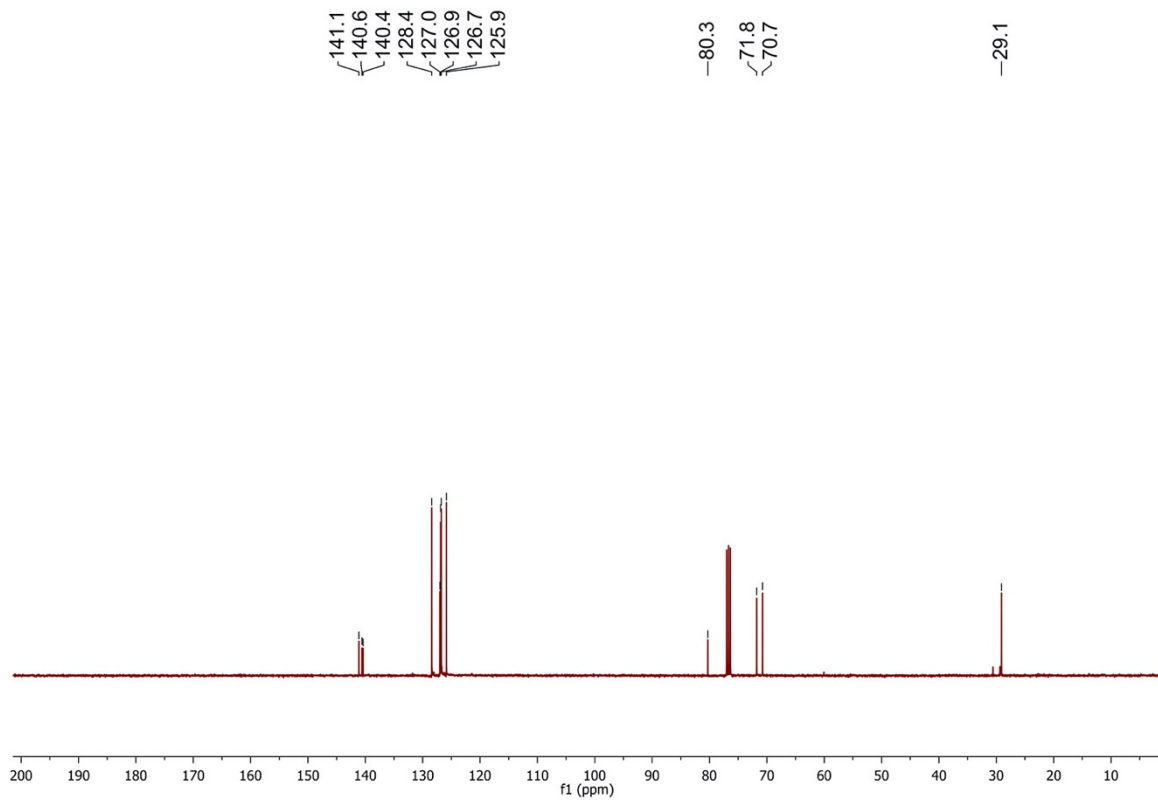


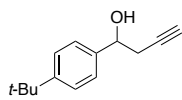


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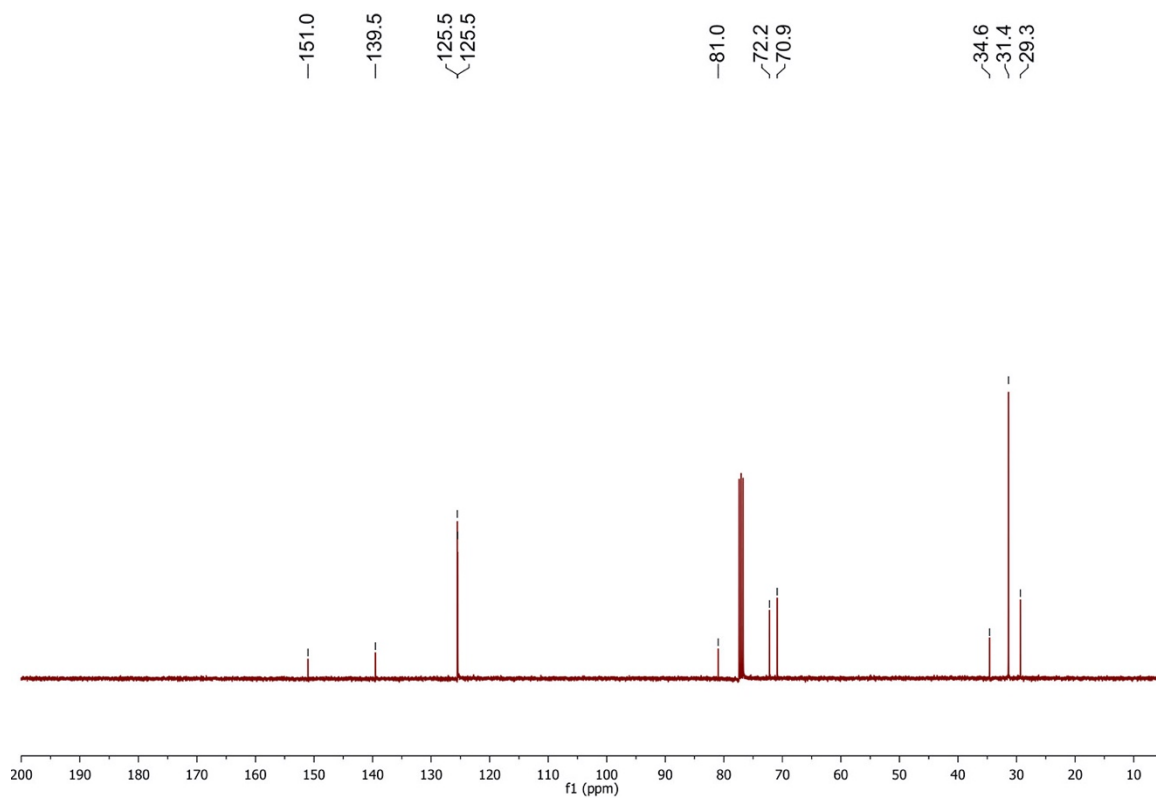


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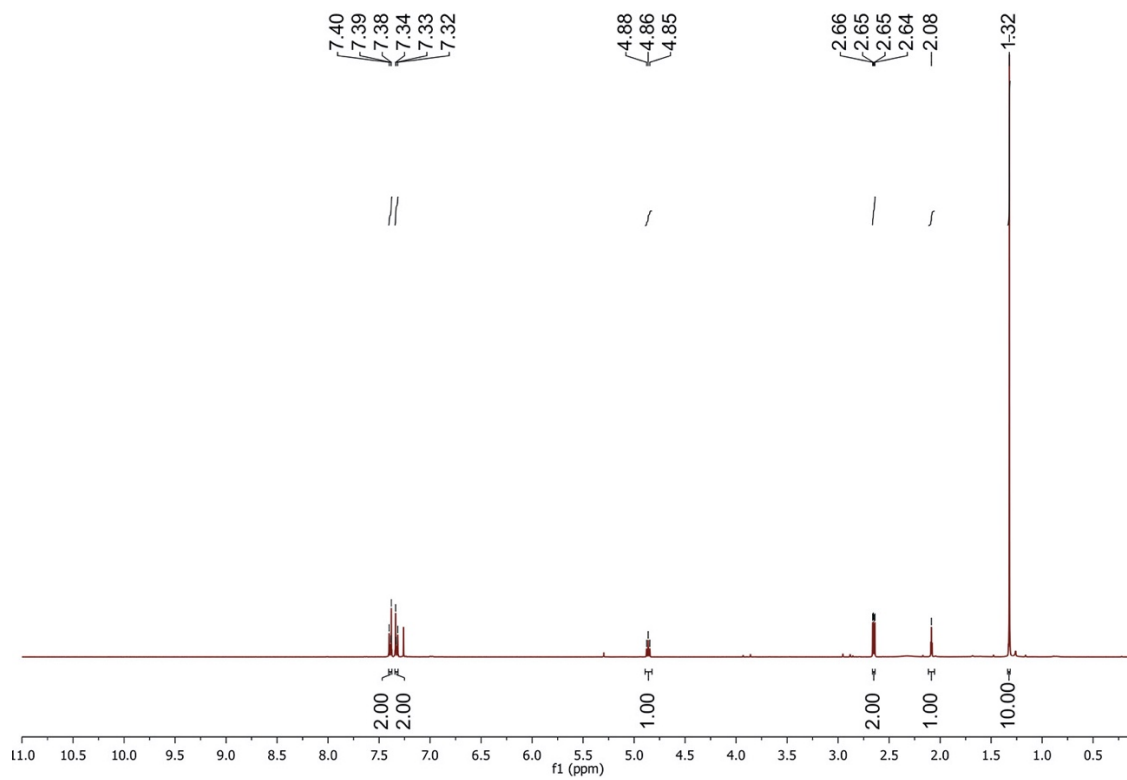


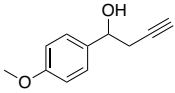


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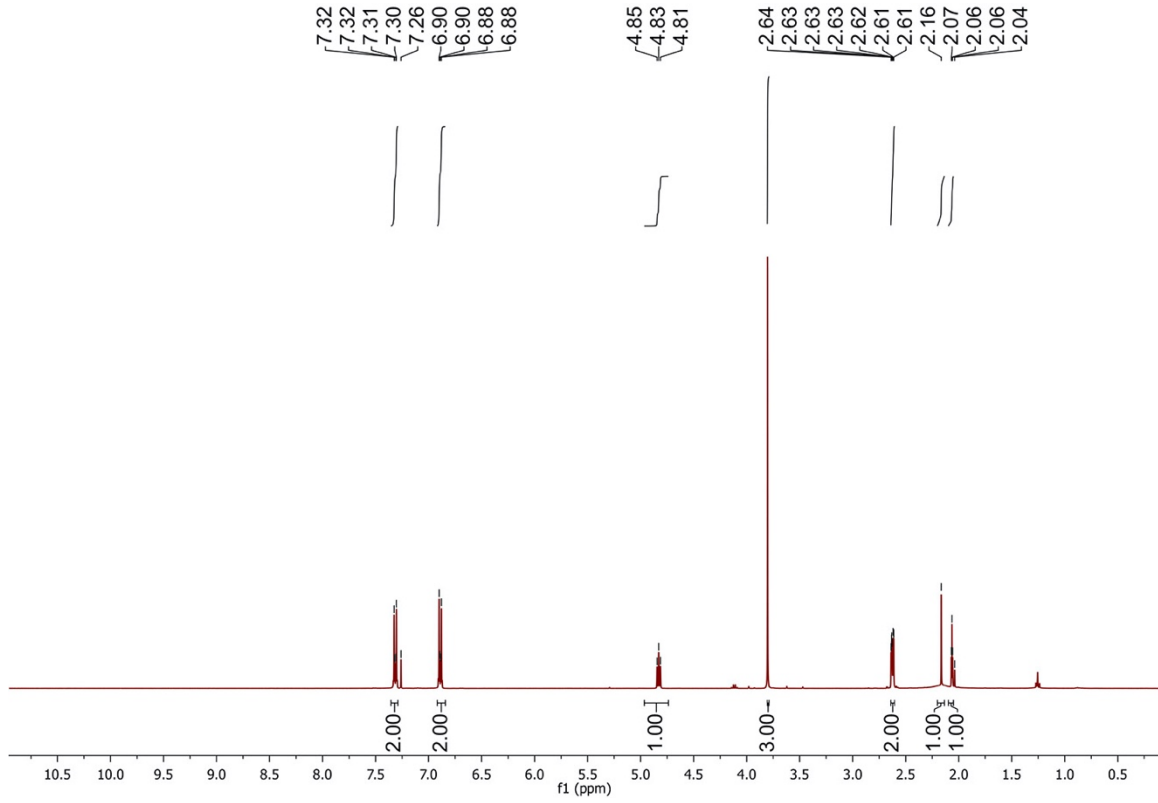


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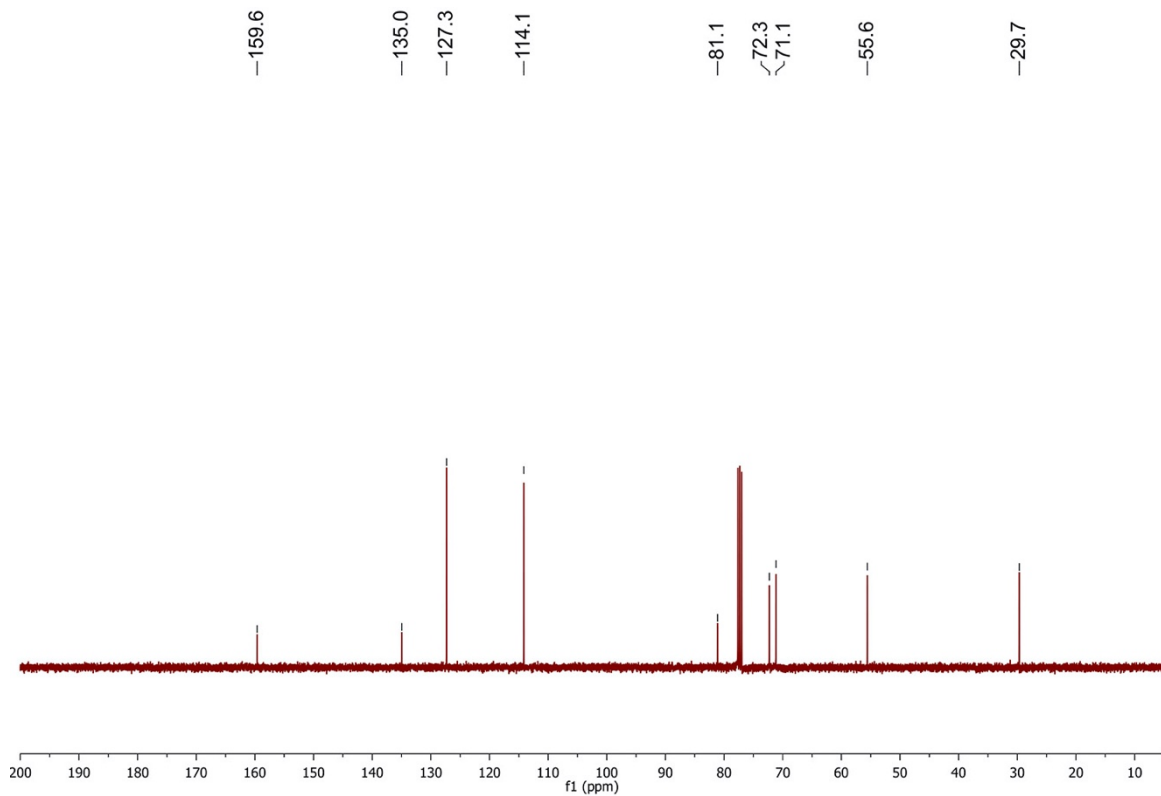


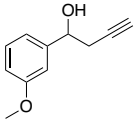


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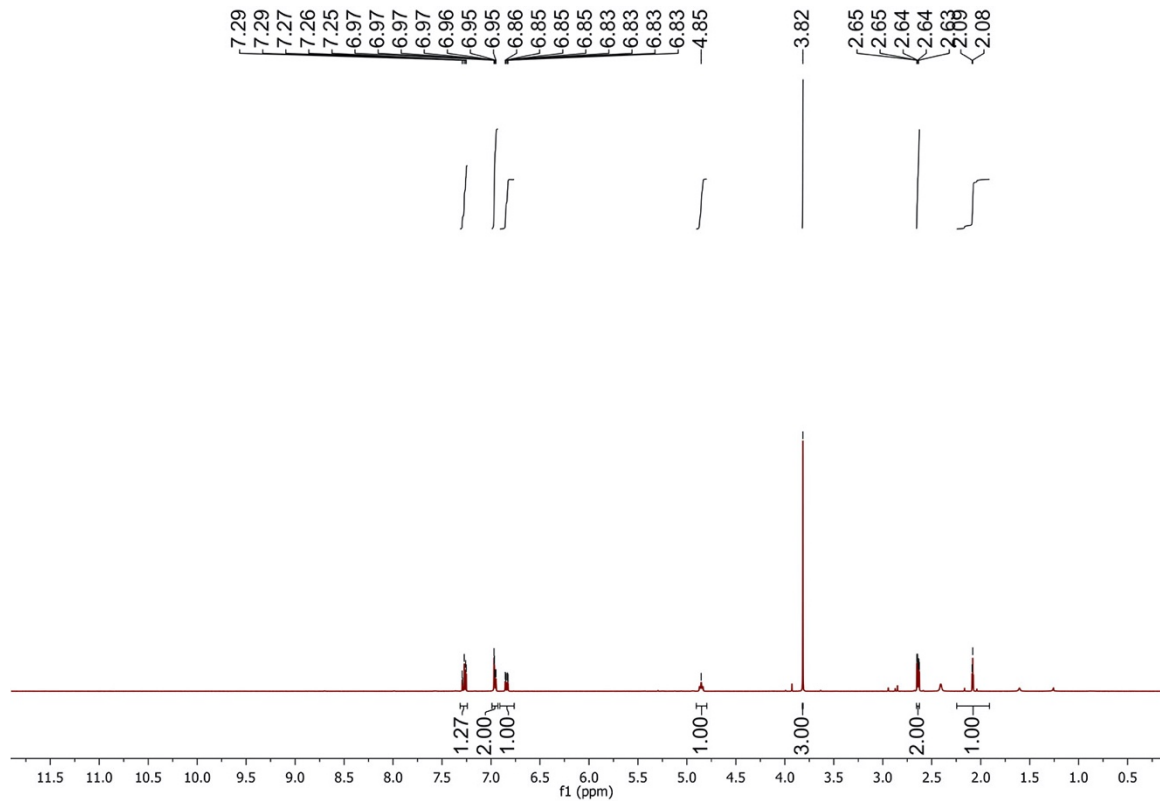


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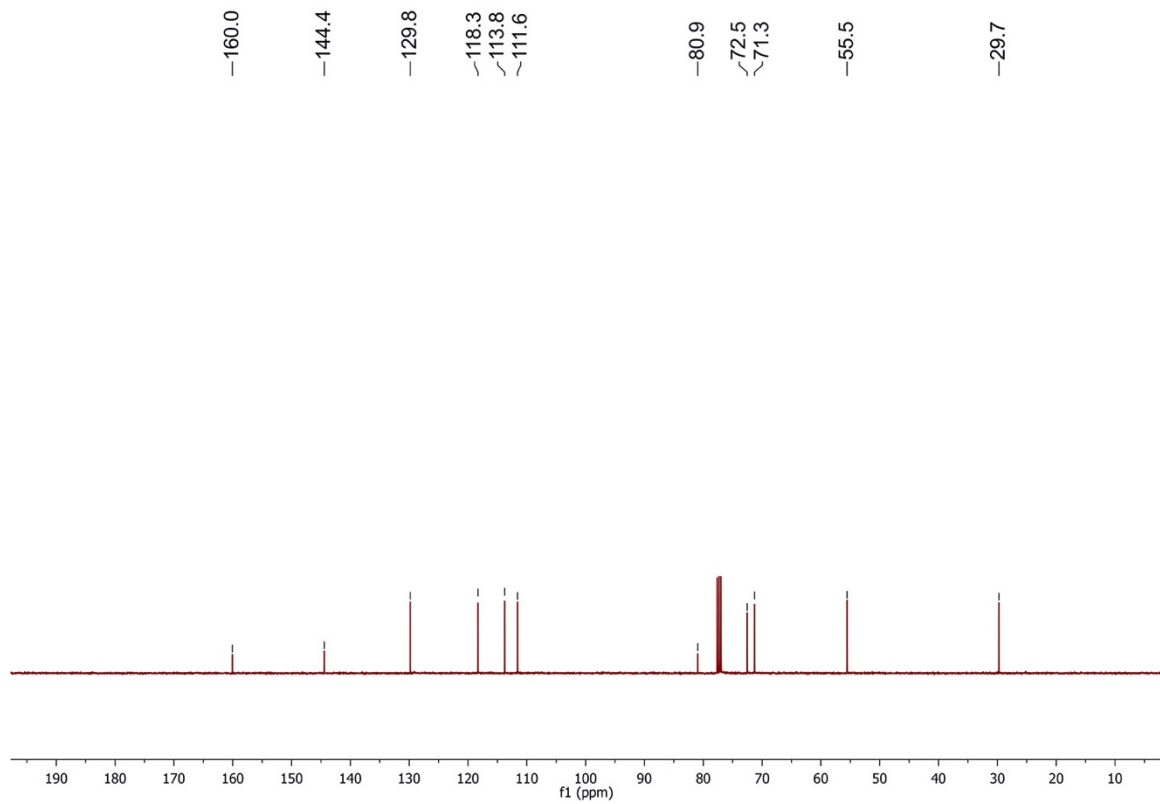


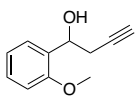


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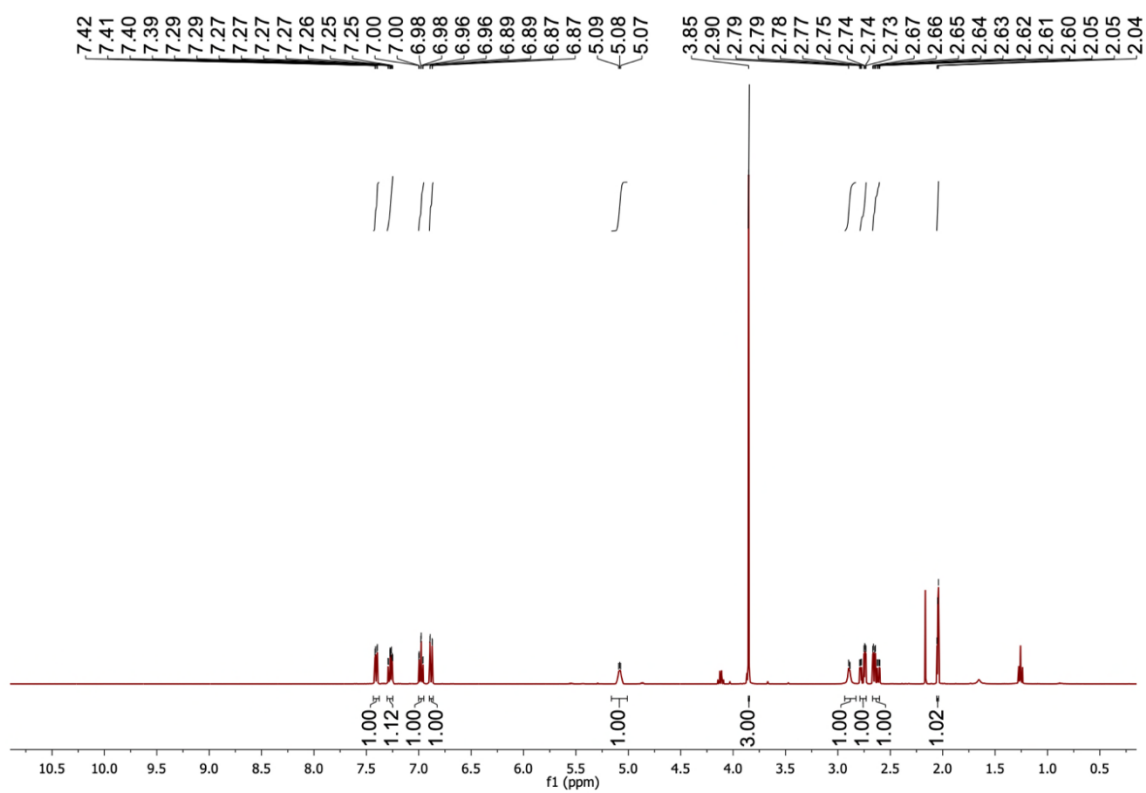


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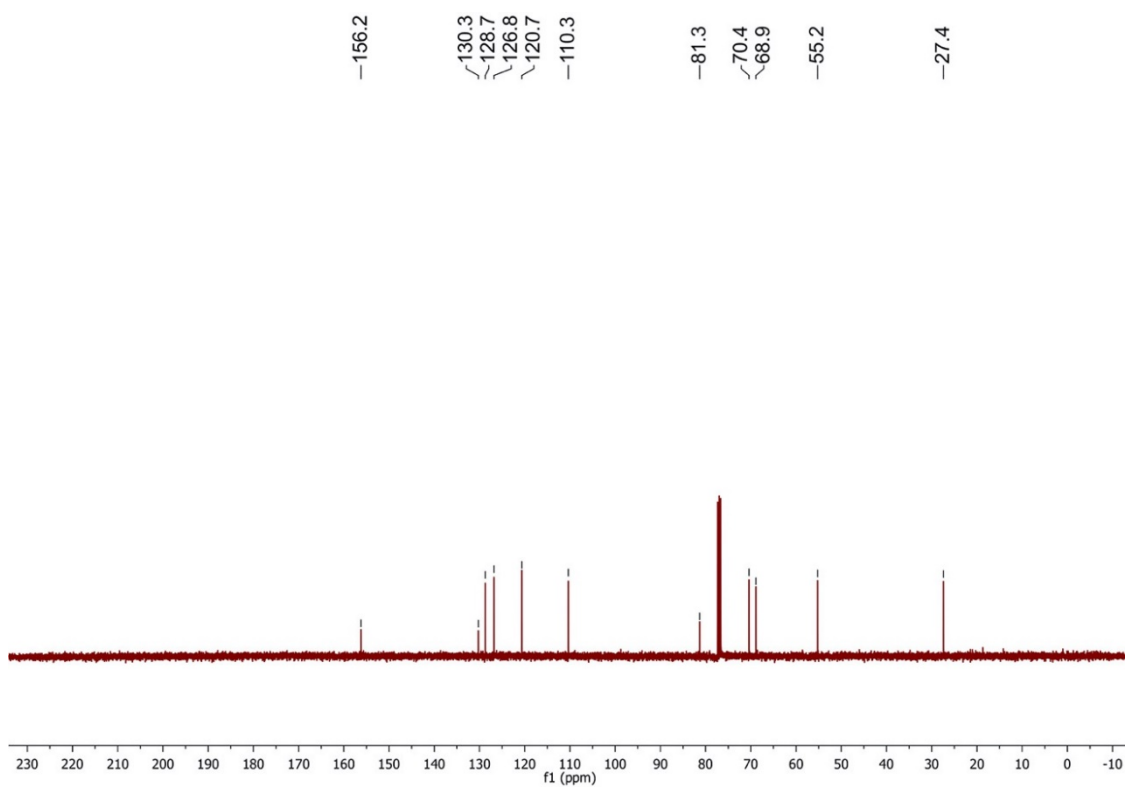


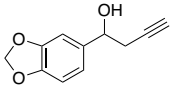


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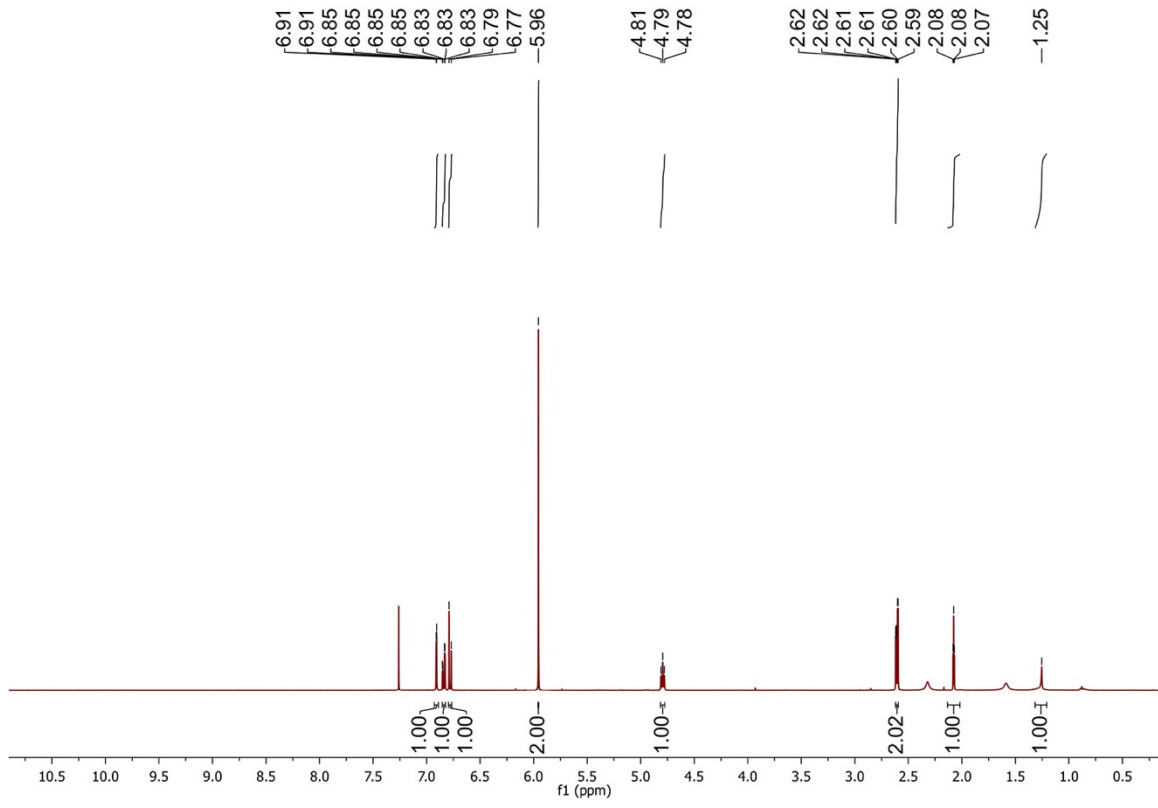


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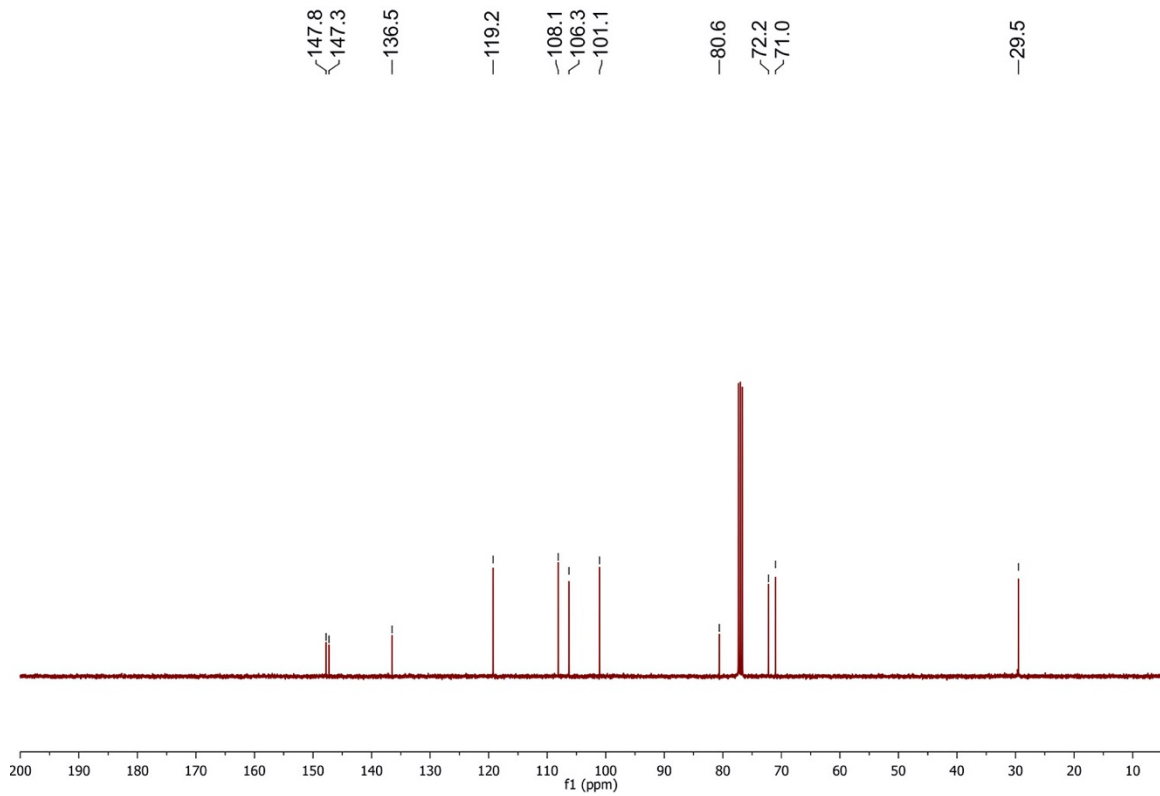


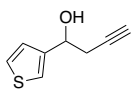


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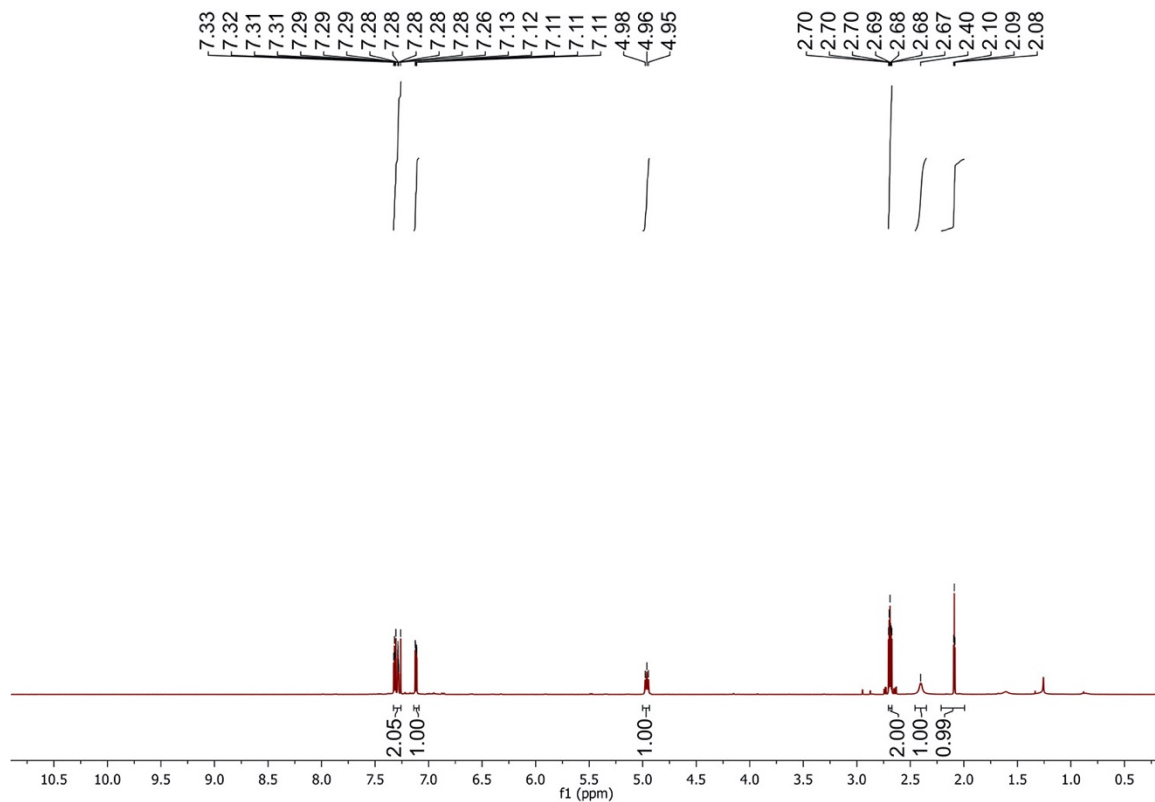


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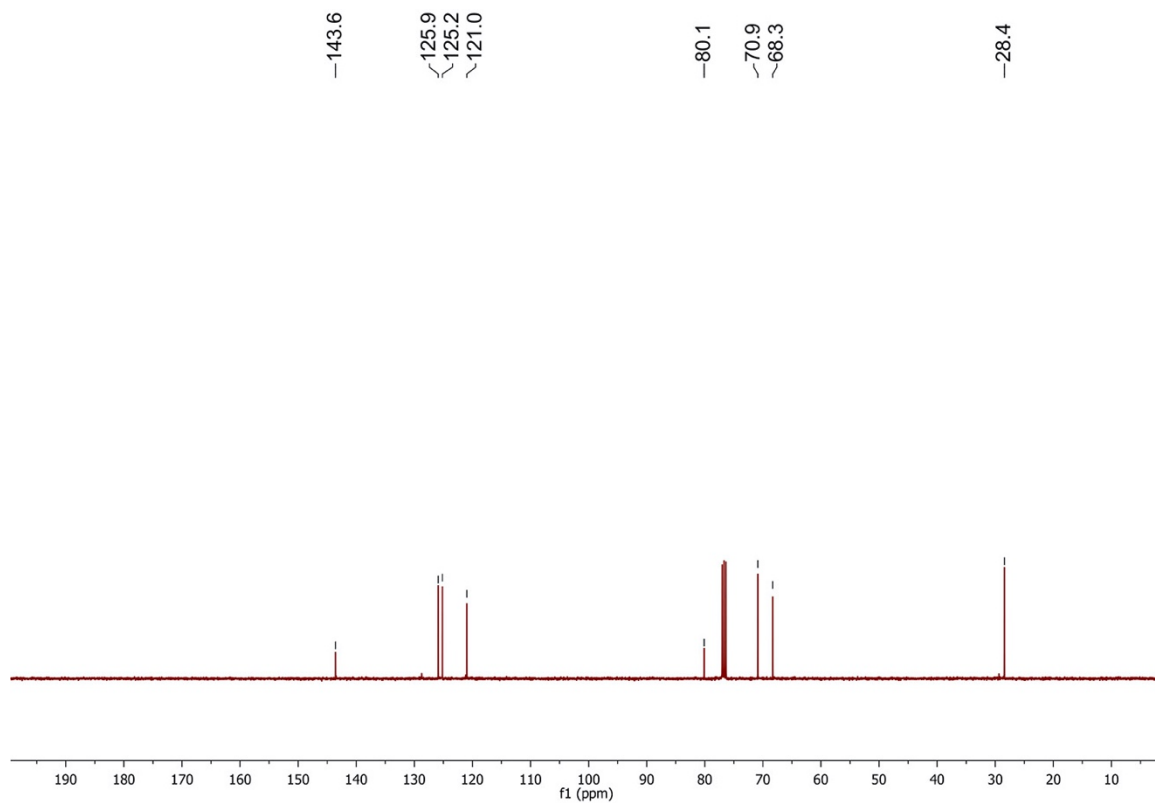


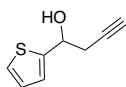


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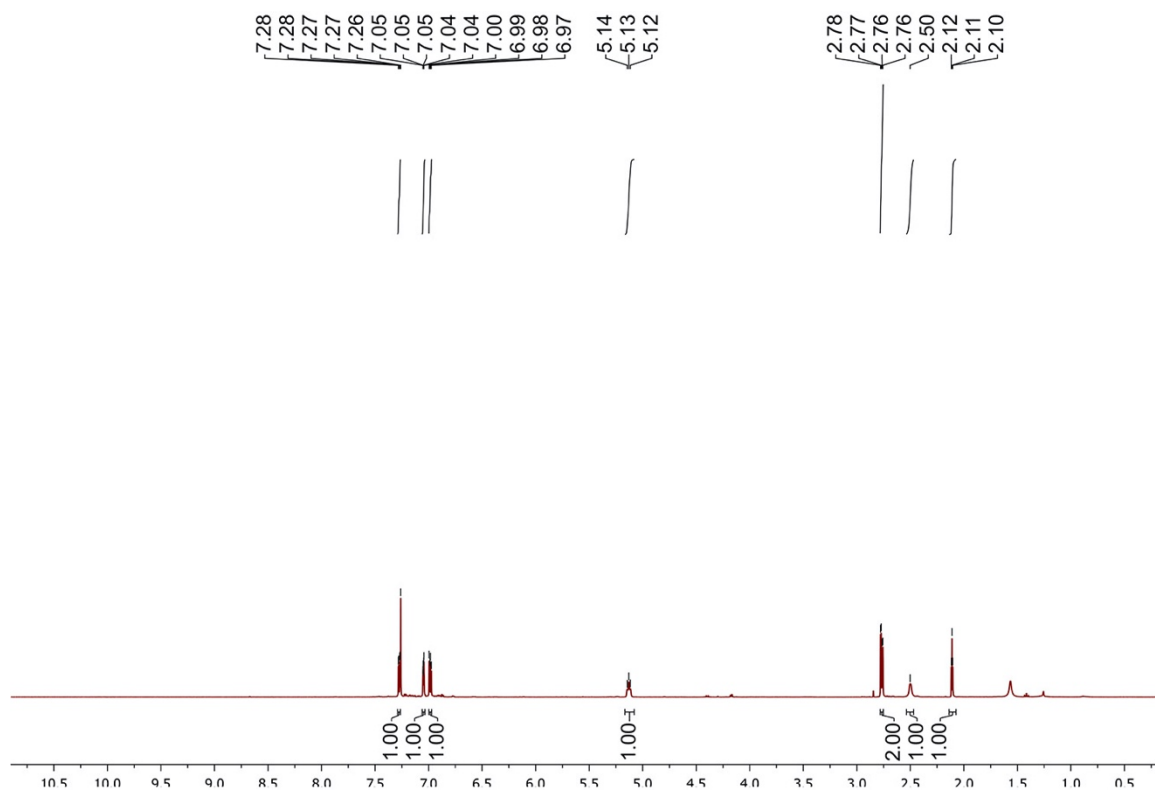


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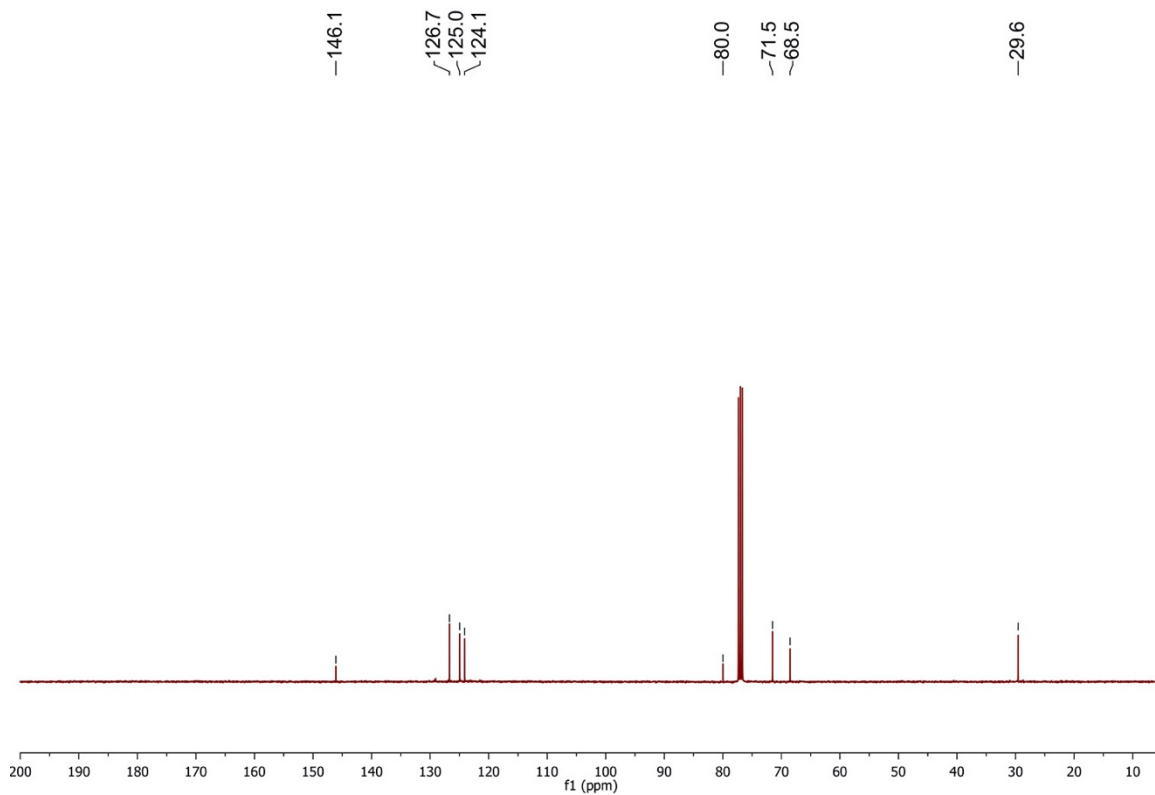


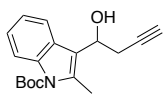


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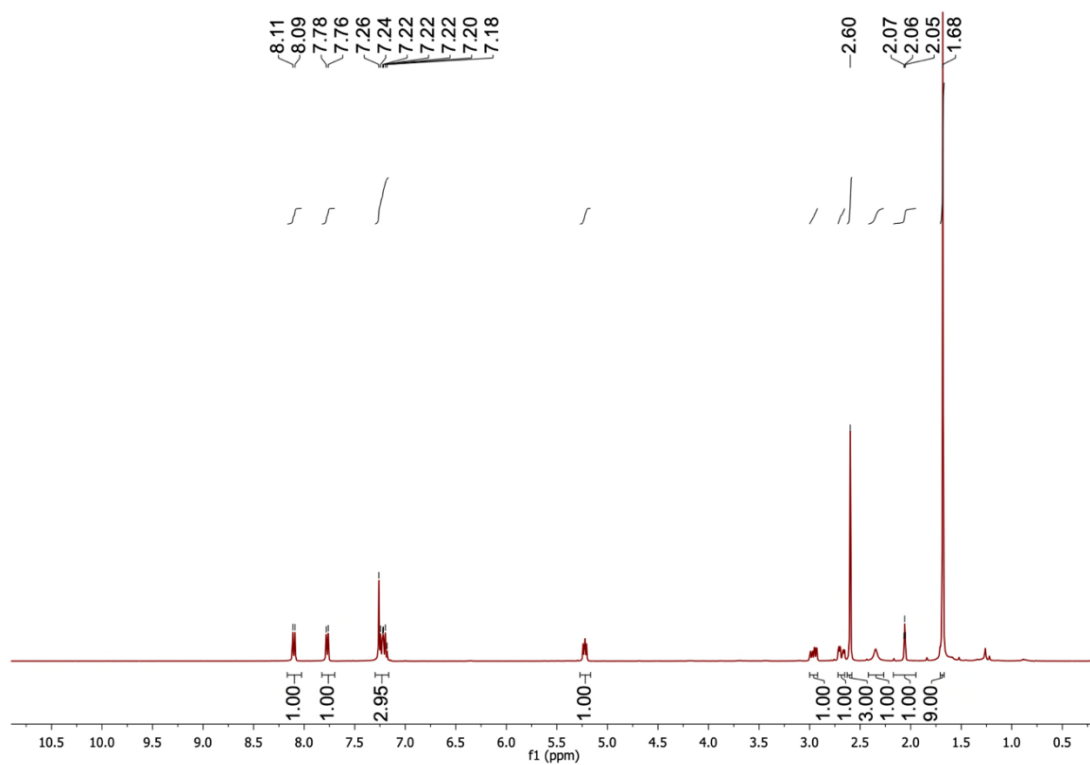


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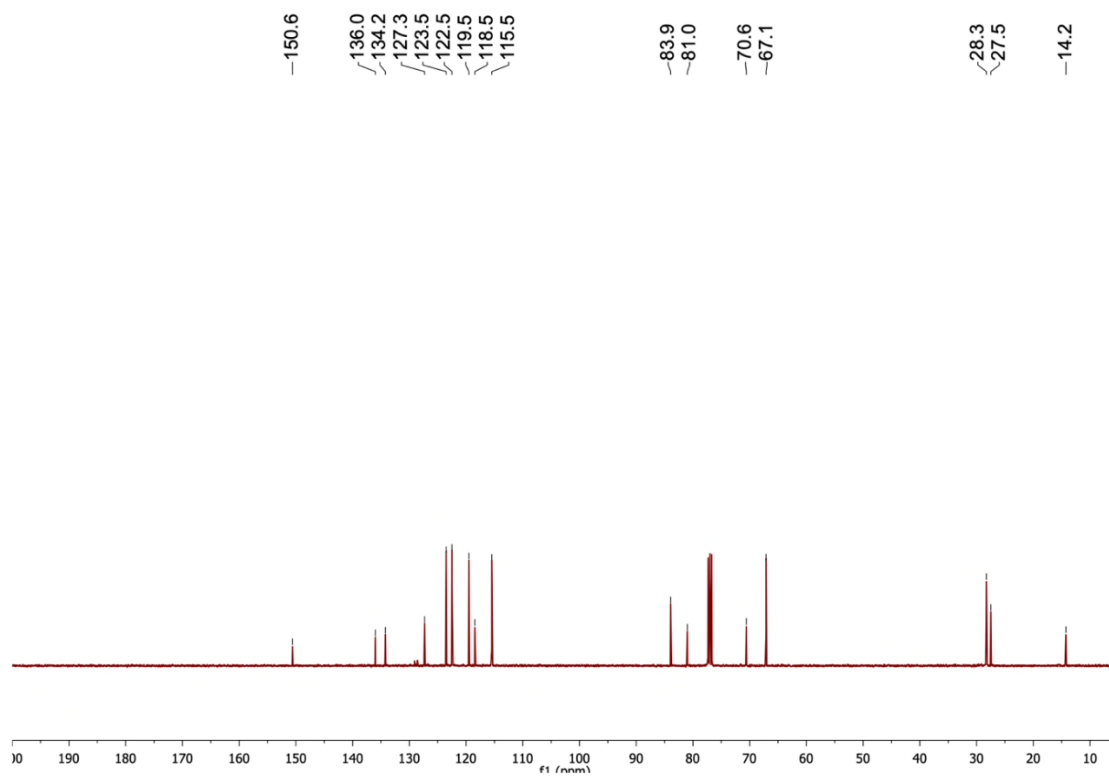


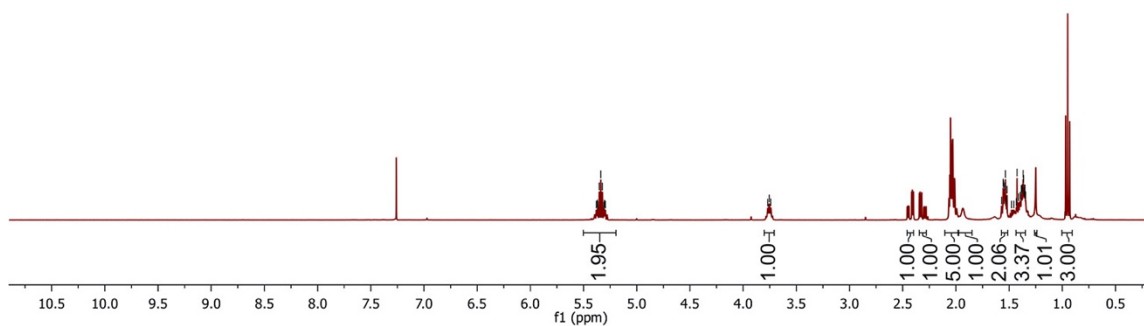
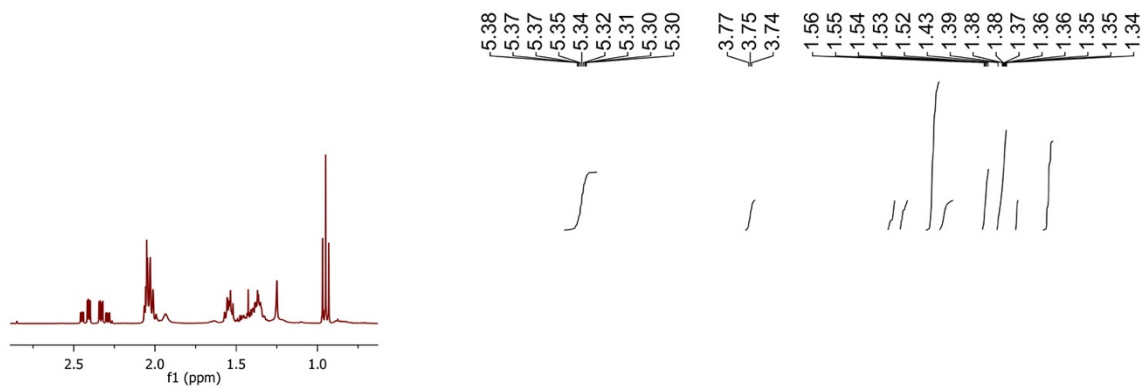
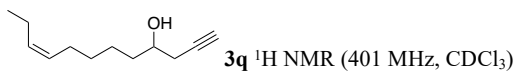


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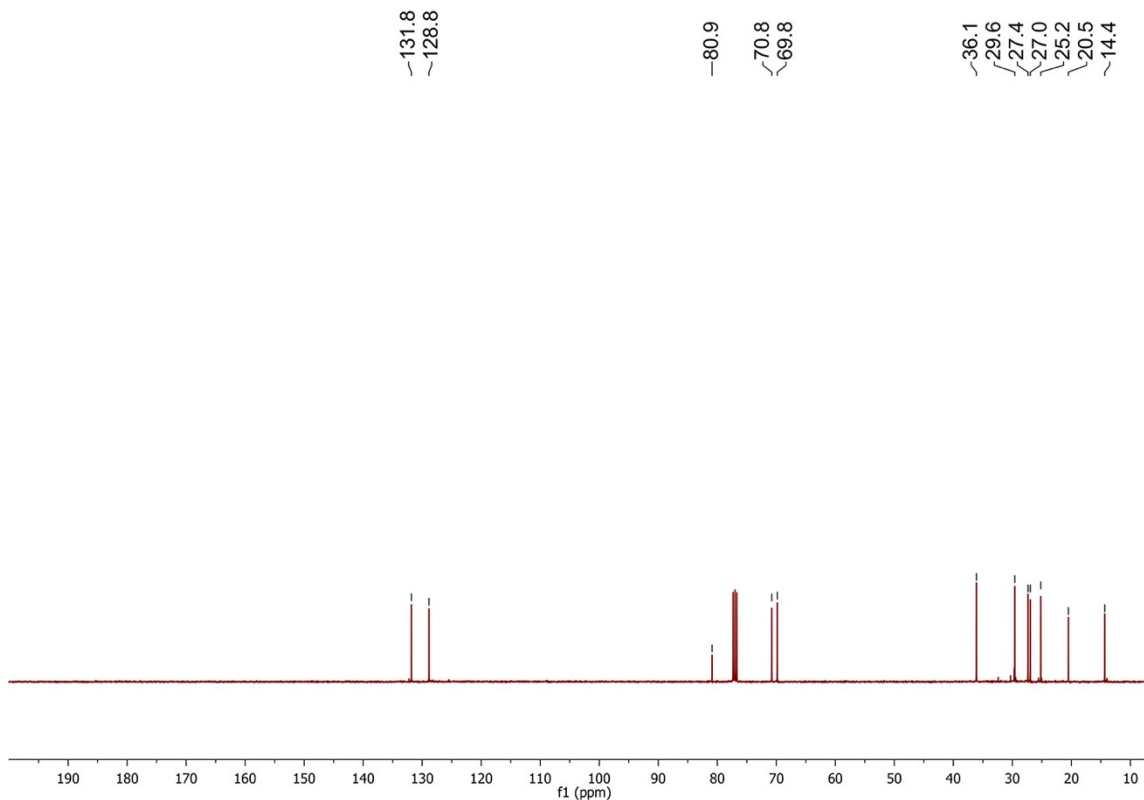


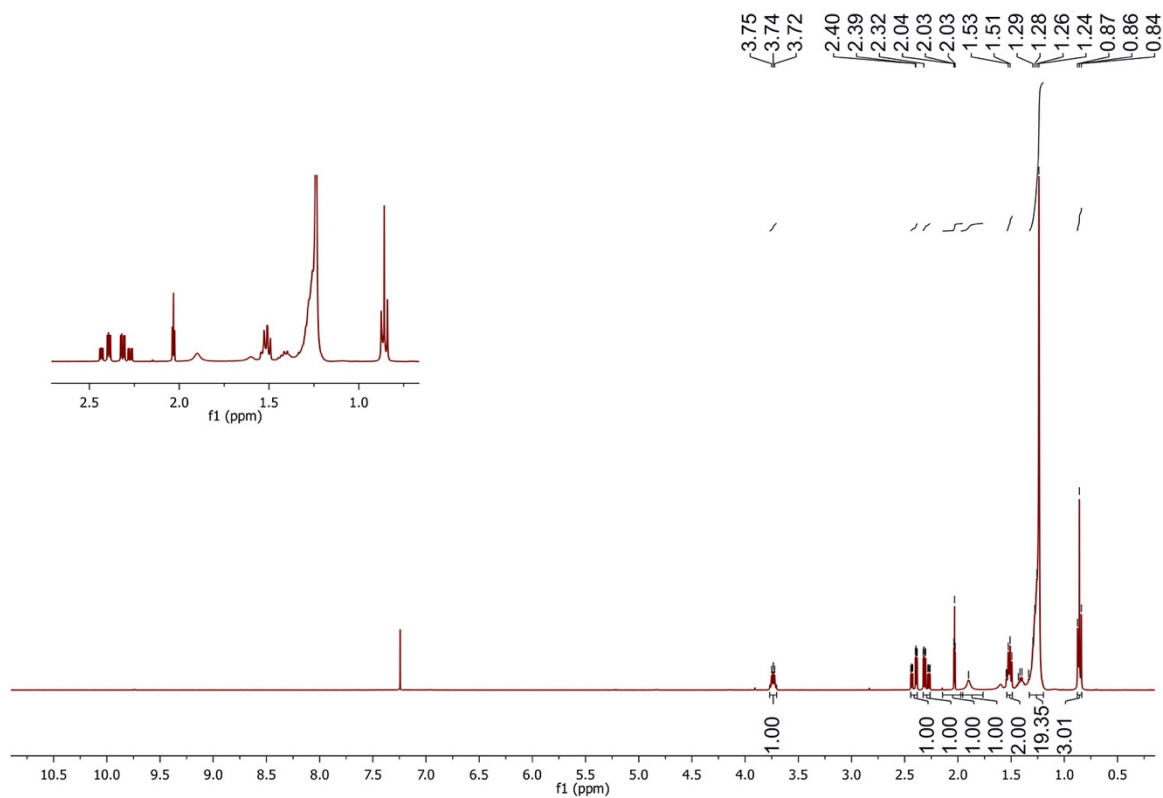
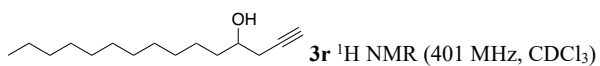
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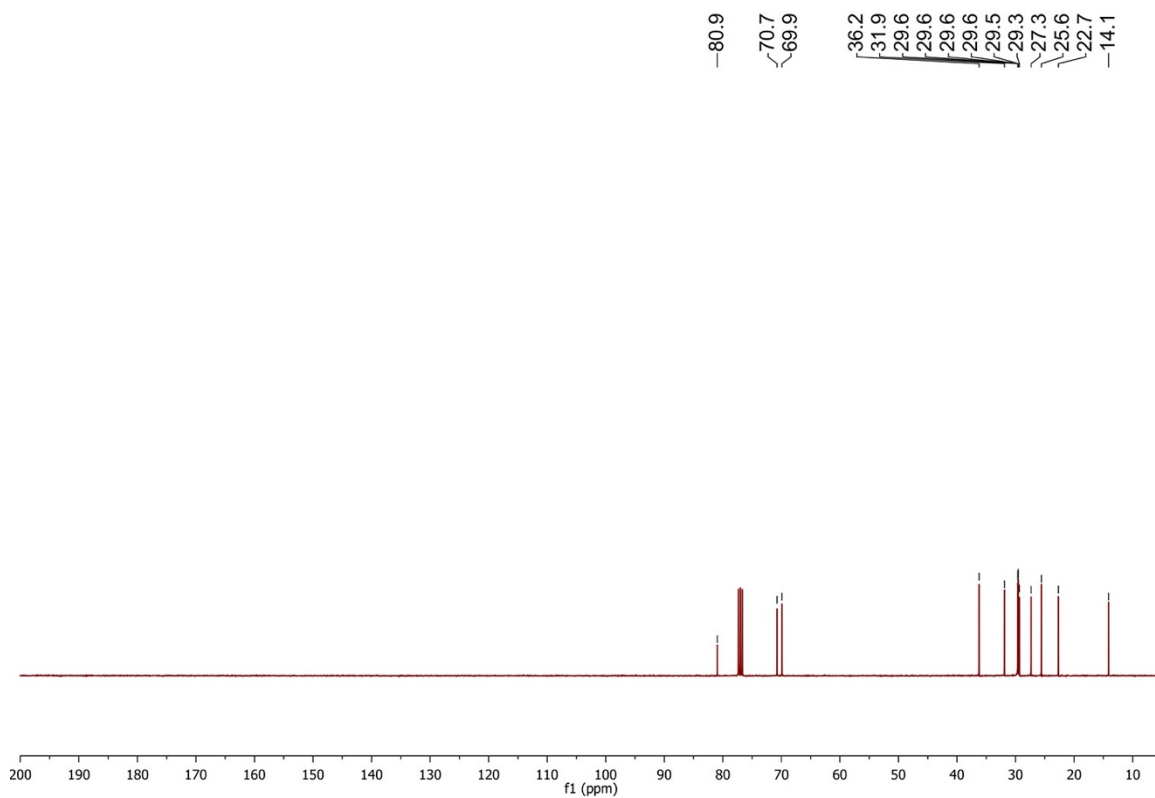


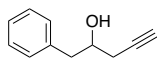
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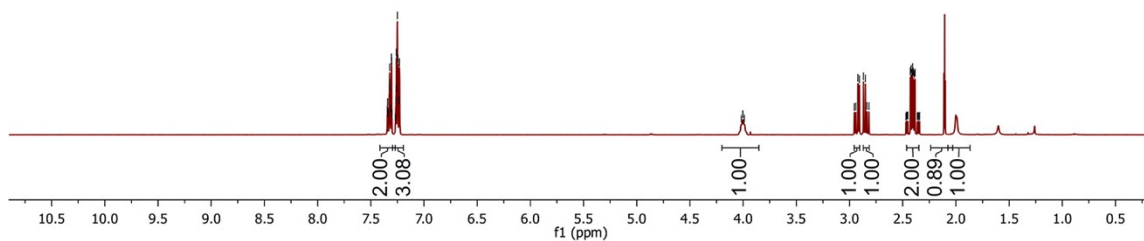
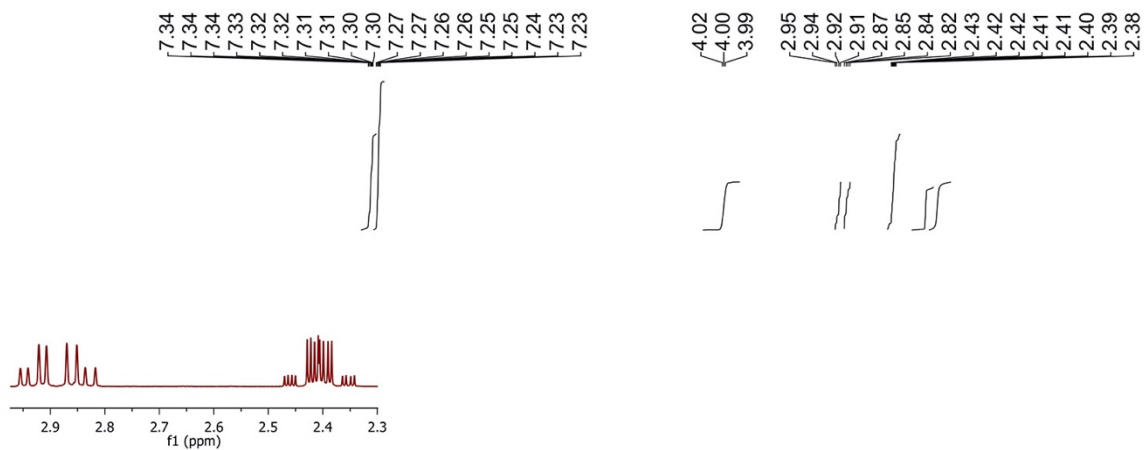


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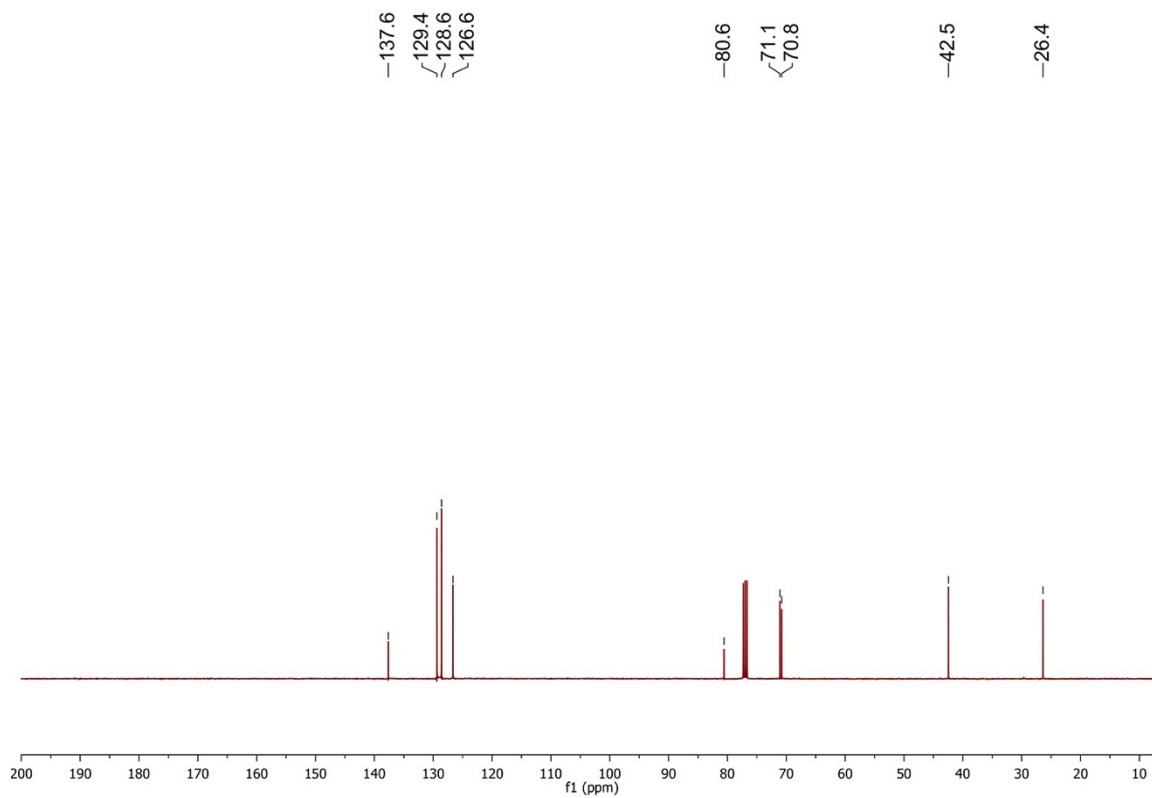


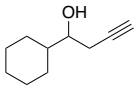


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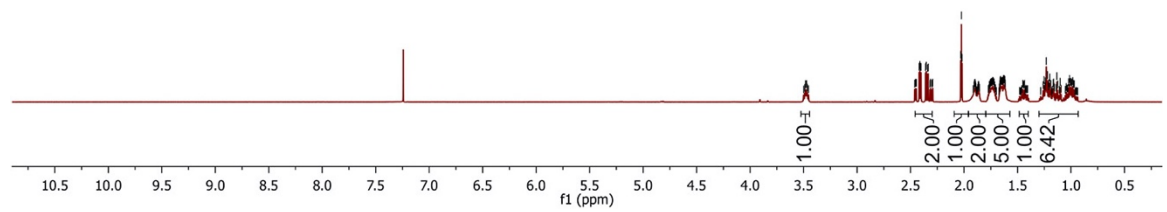
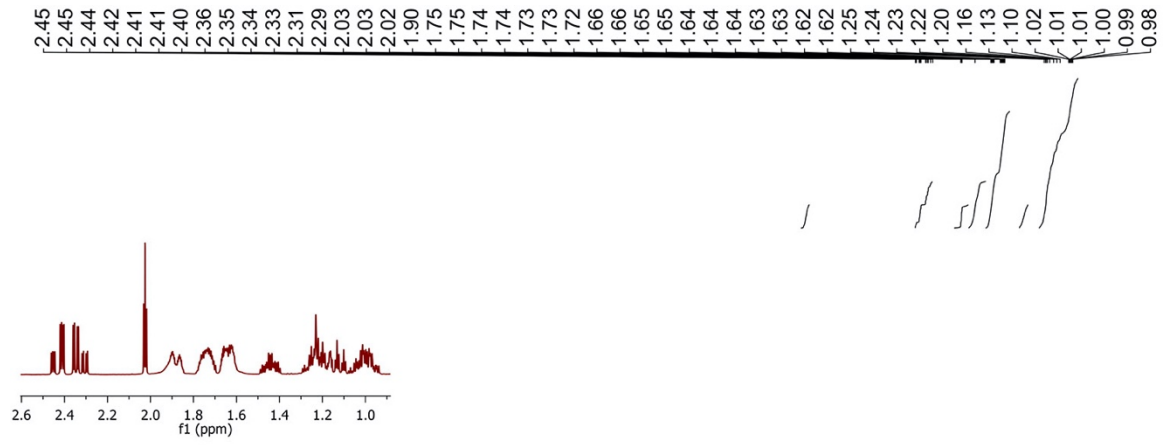


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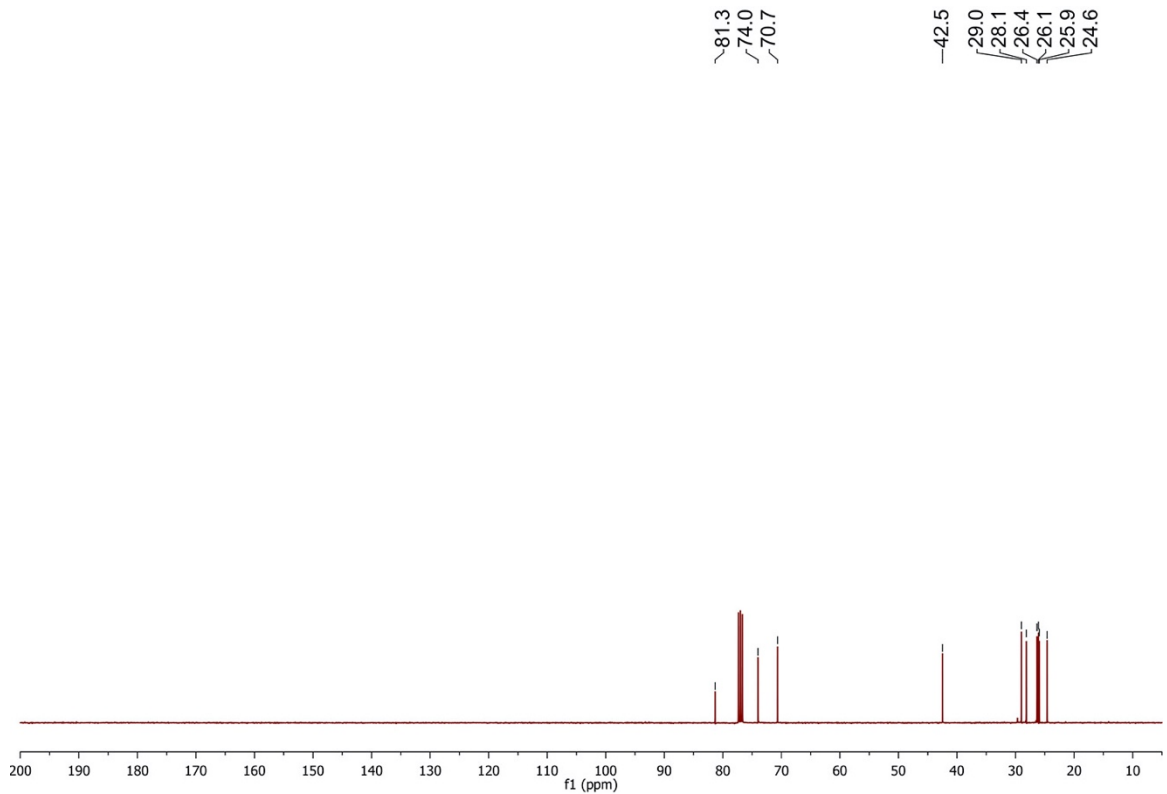


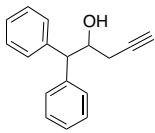


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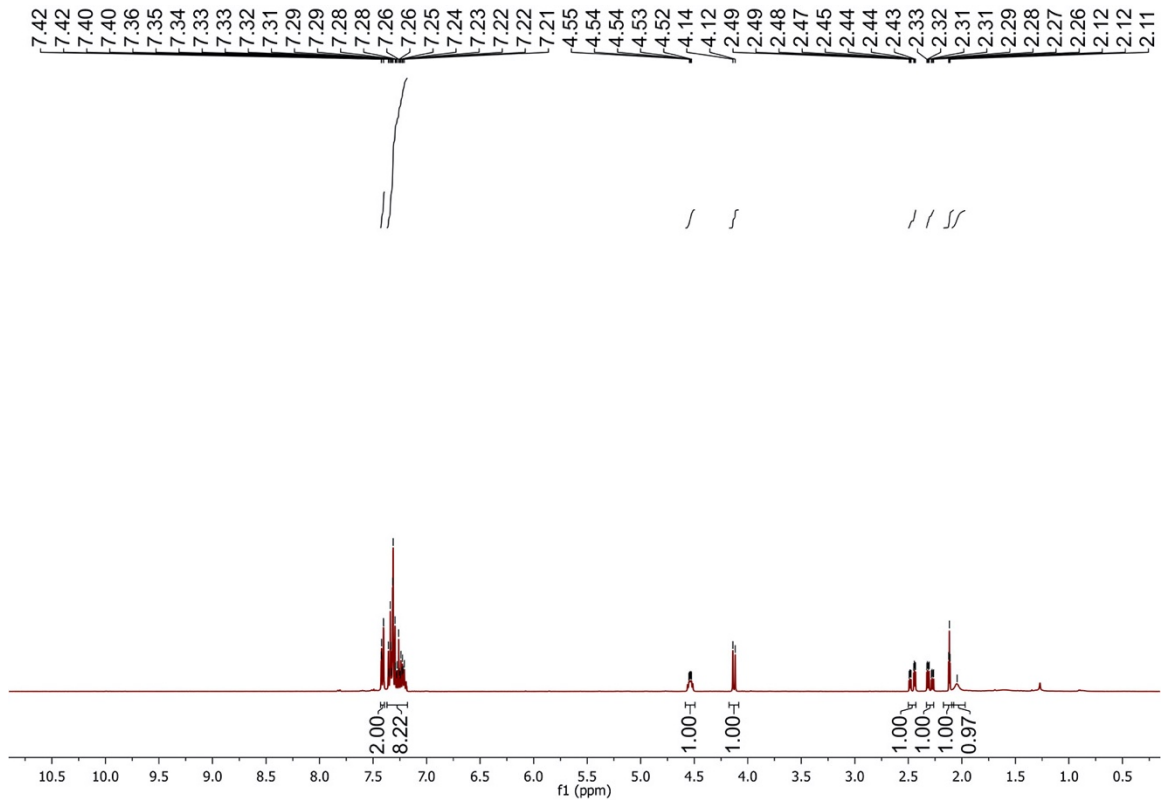


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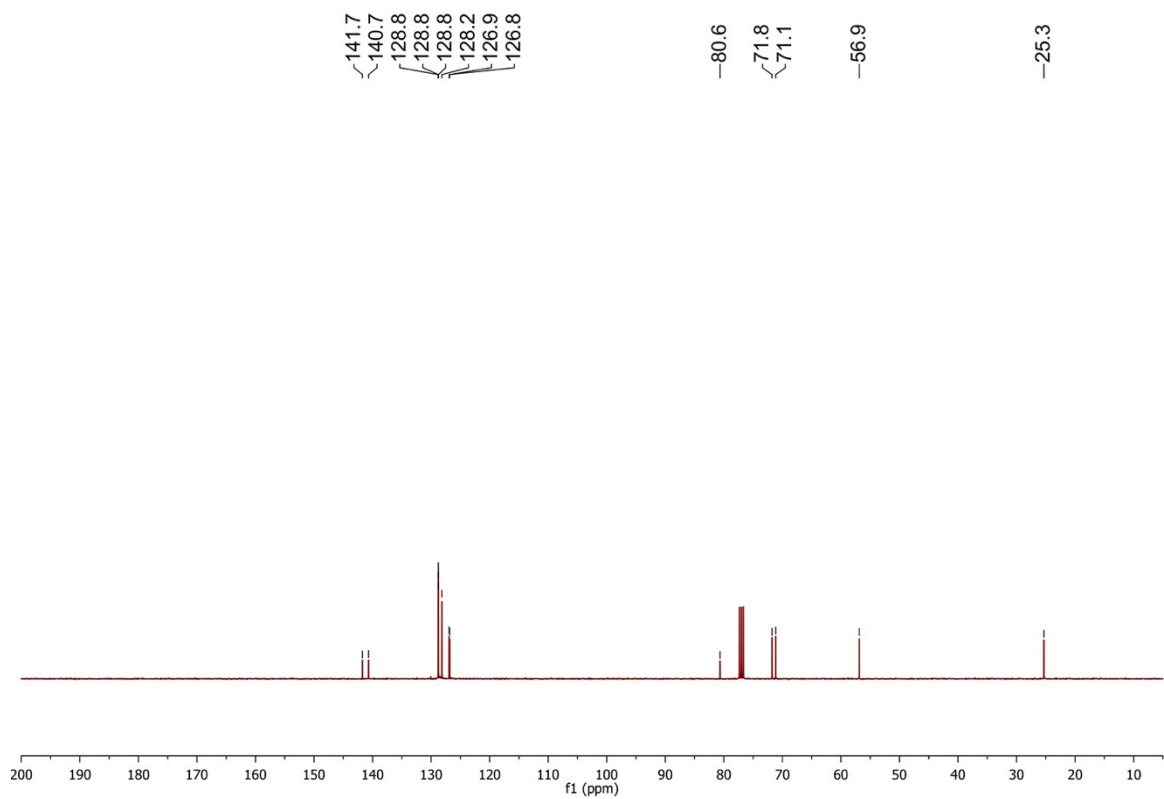


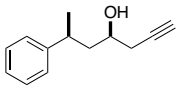


3u ^1H NMR (401 MHz, CDCl_3)

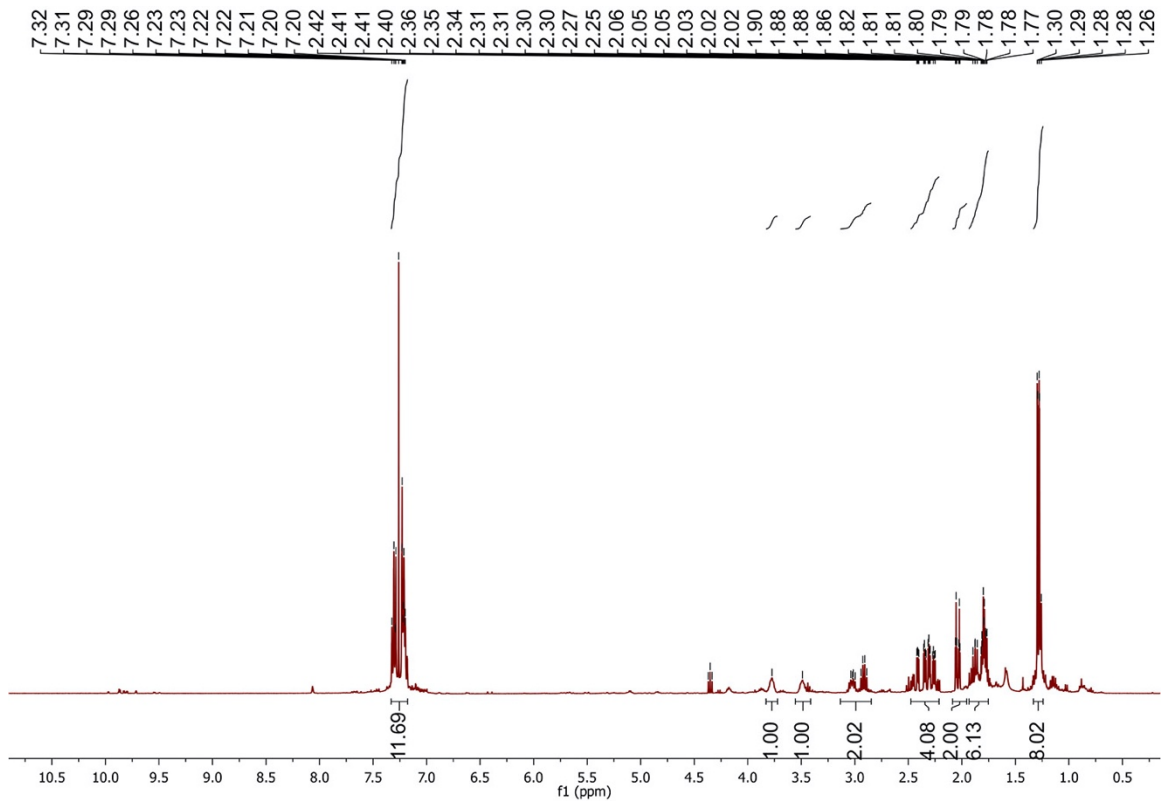


$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)

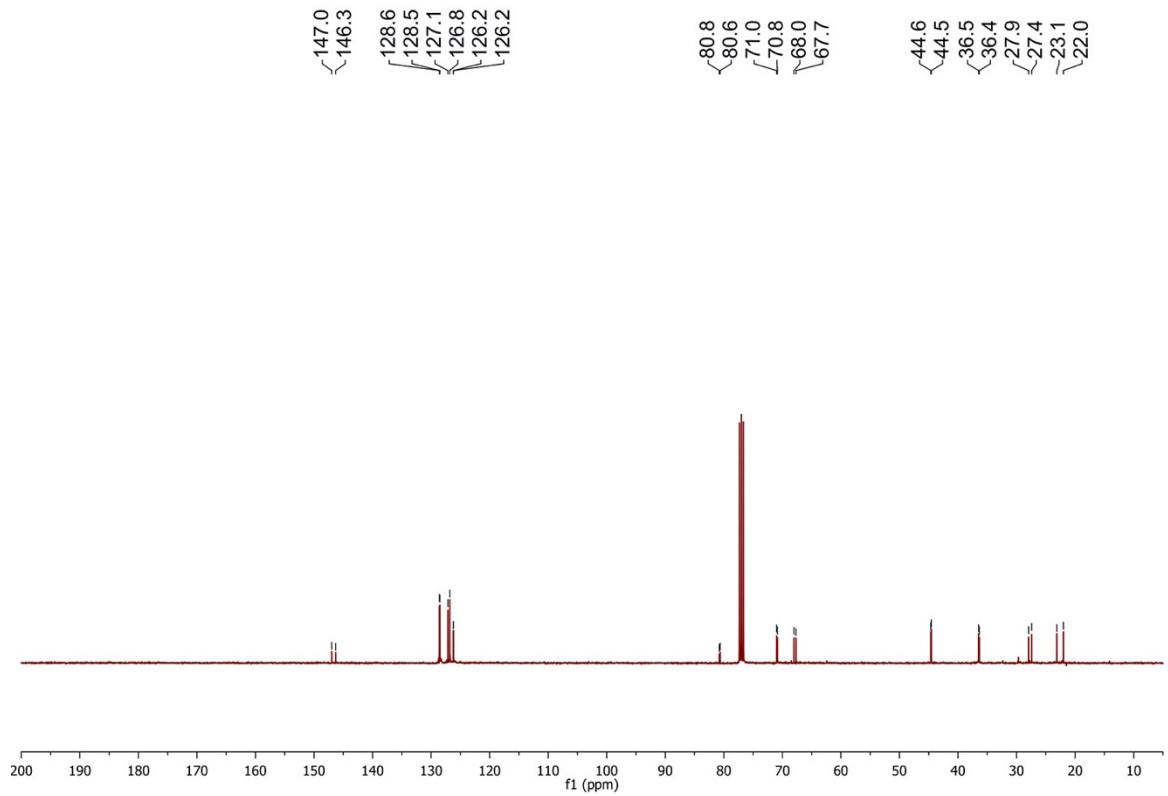


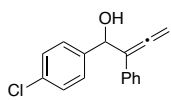


3v ^1H NMR (401 MHz, CDCl_3)

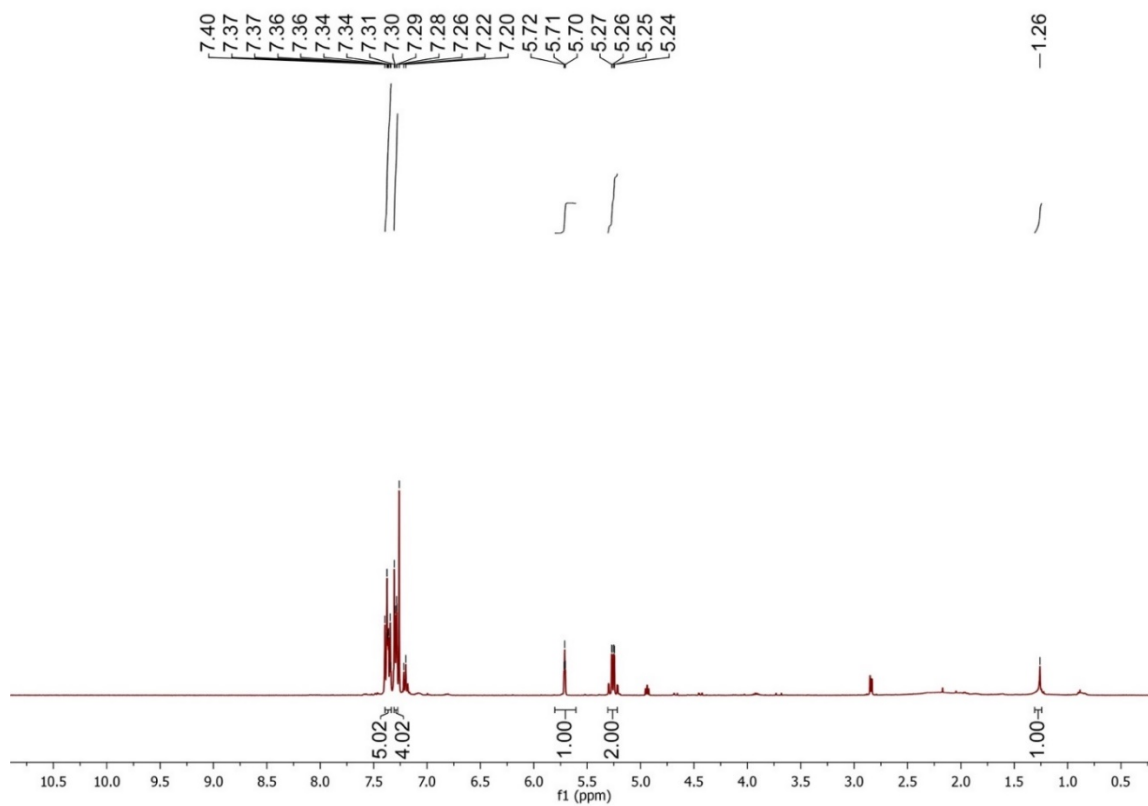


$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)

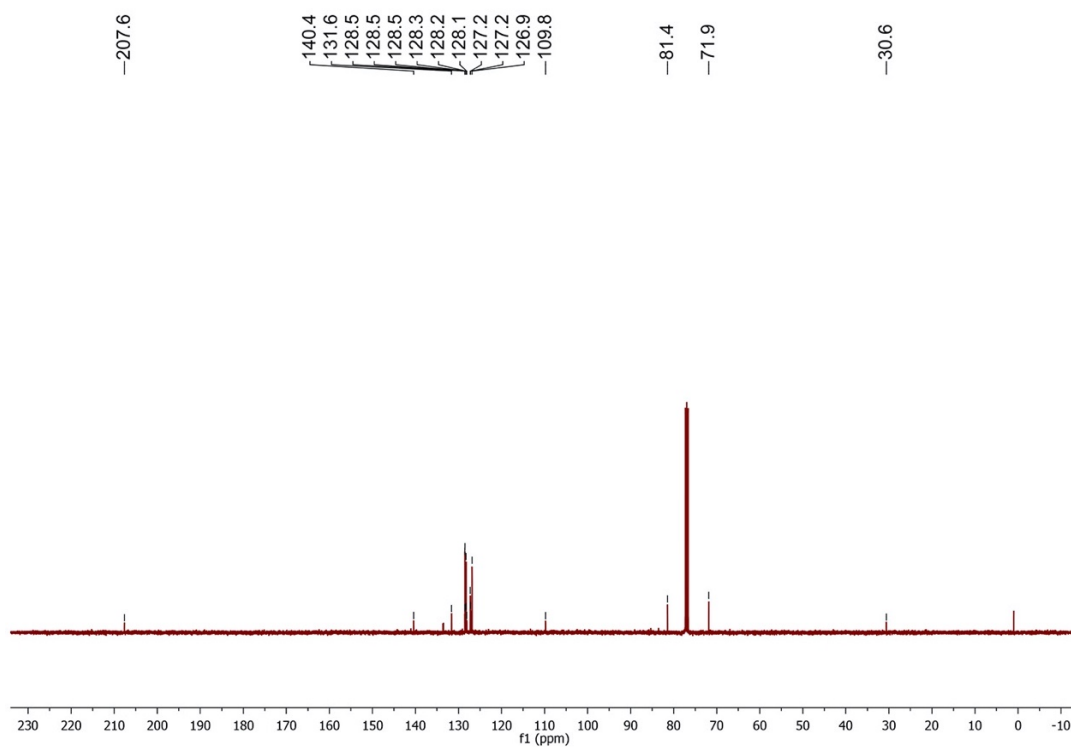


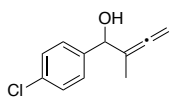


4a ^1H NMR (401 MHz, CDCl_3)

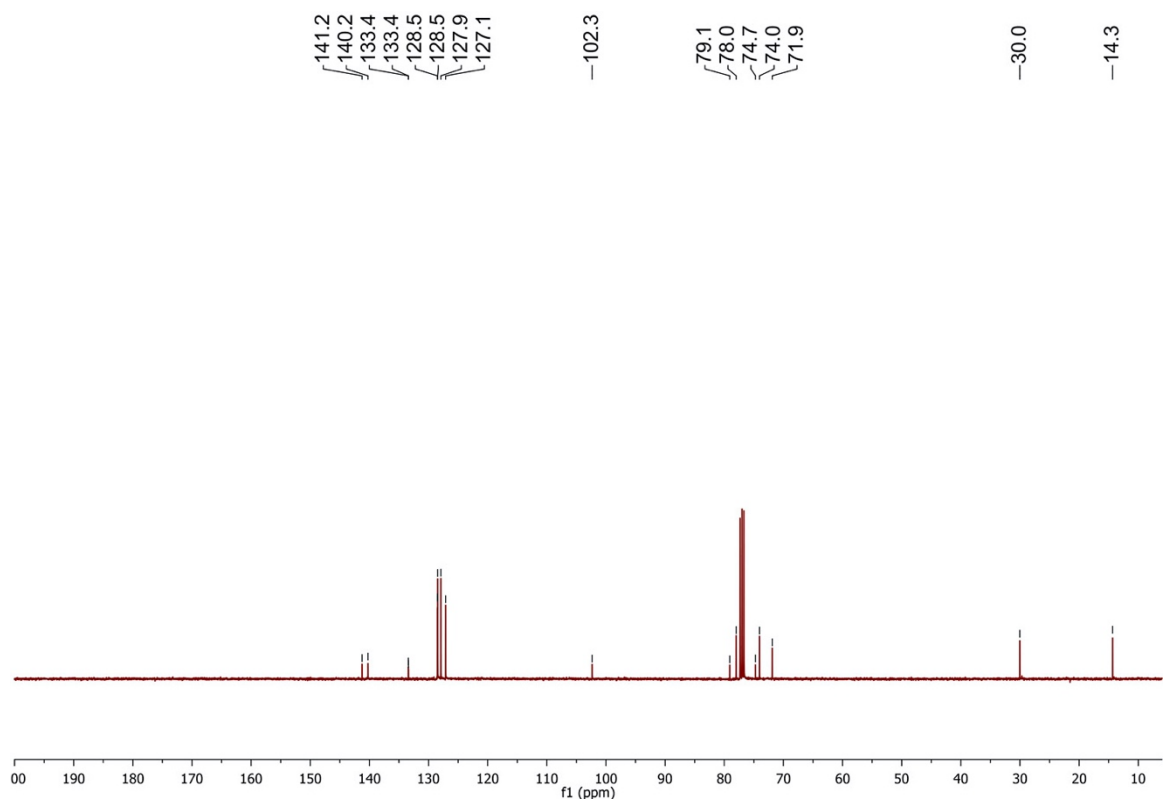
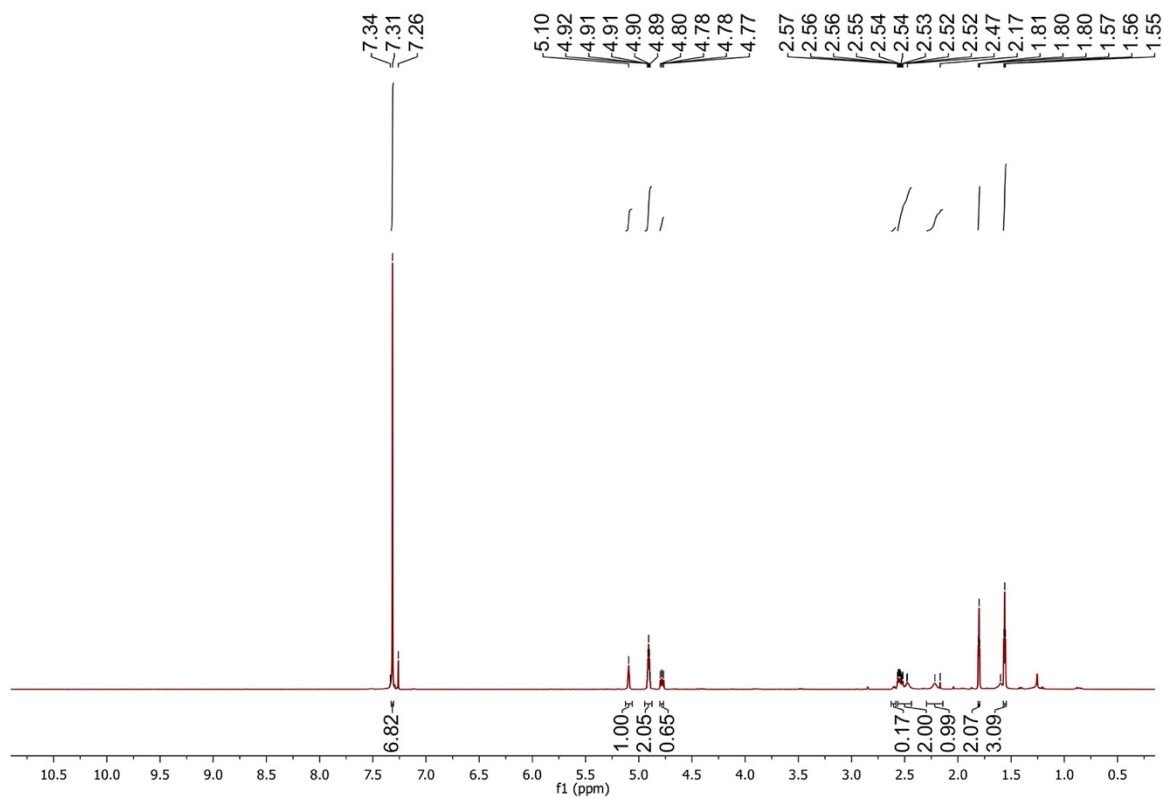


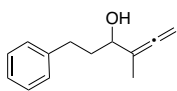
$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)



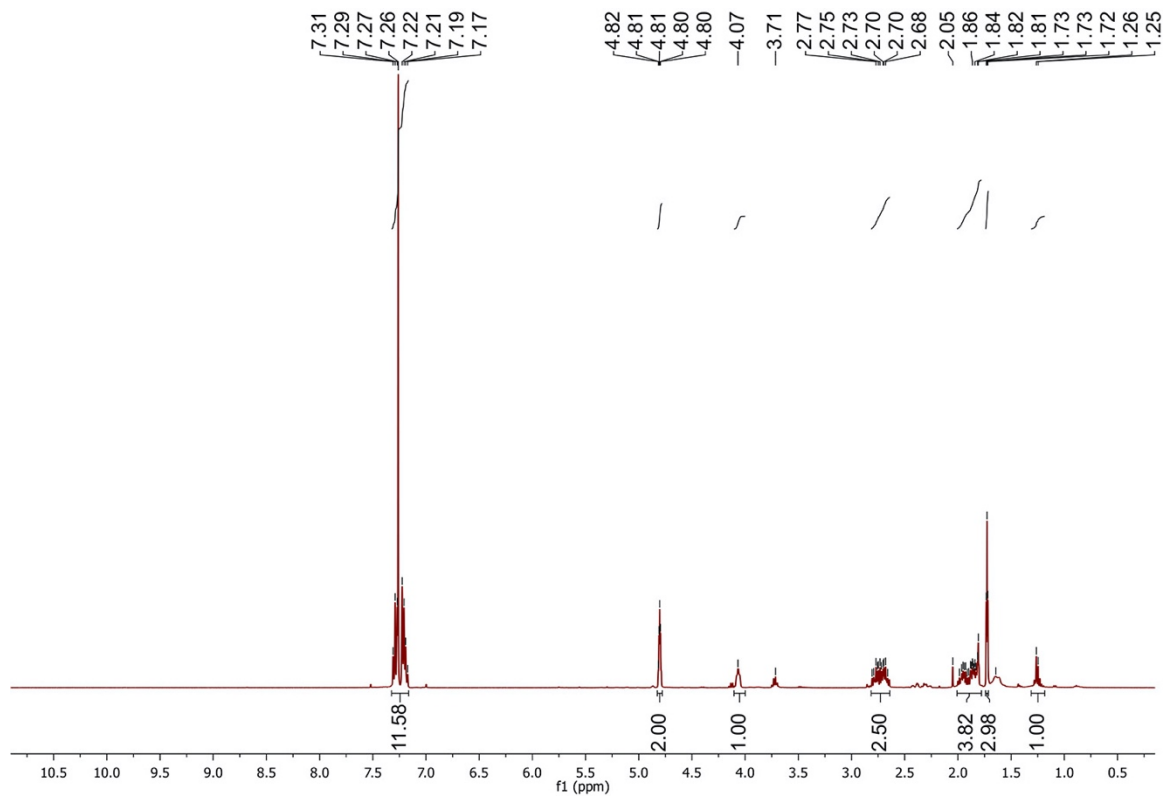


4b ^1H NMR (401 MHz, CDCl_3)

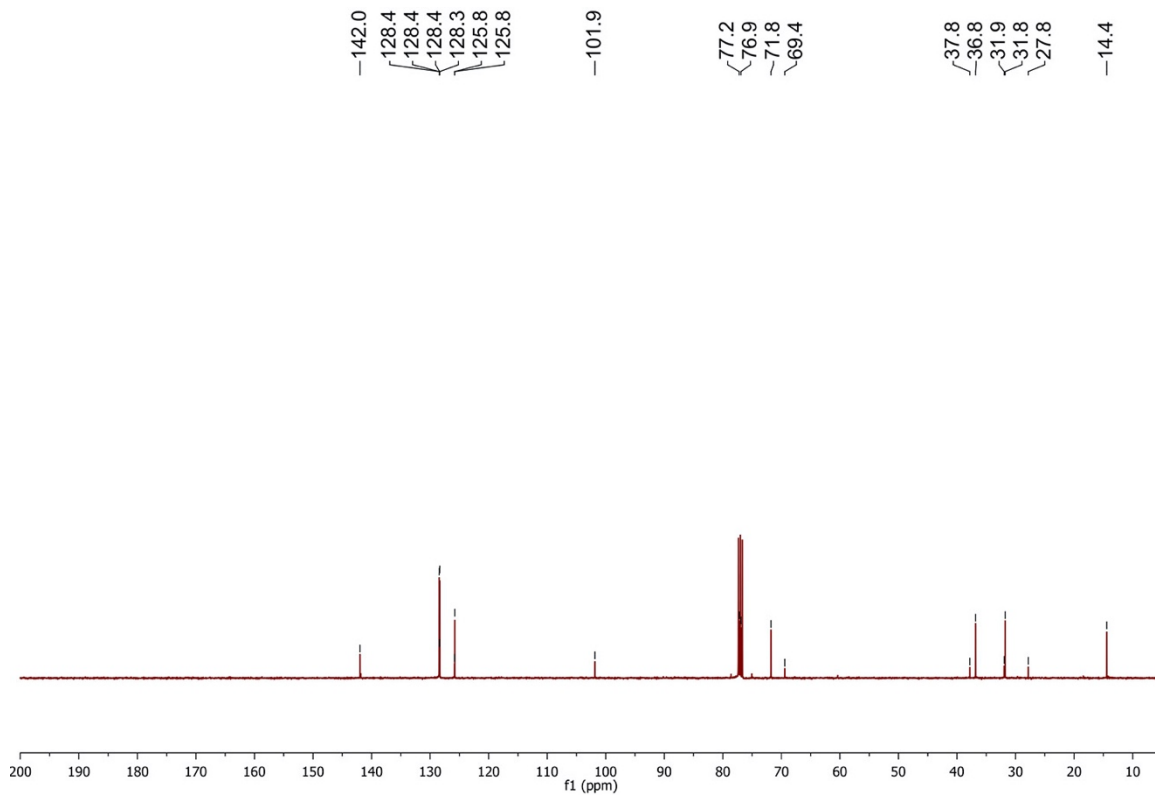


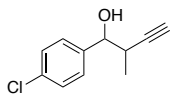


4c ^1H NMR (401 MHz, CDCl_3)

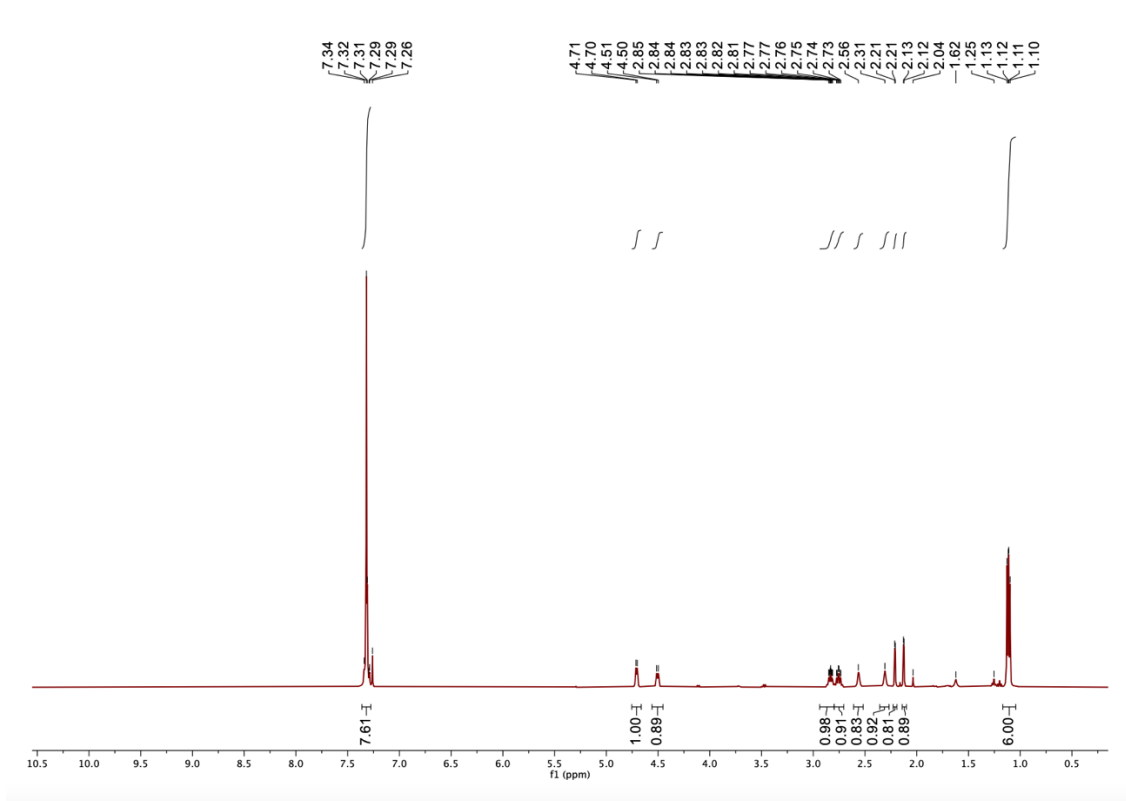


$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)

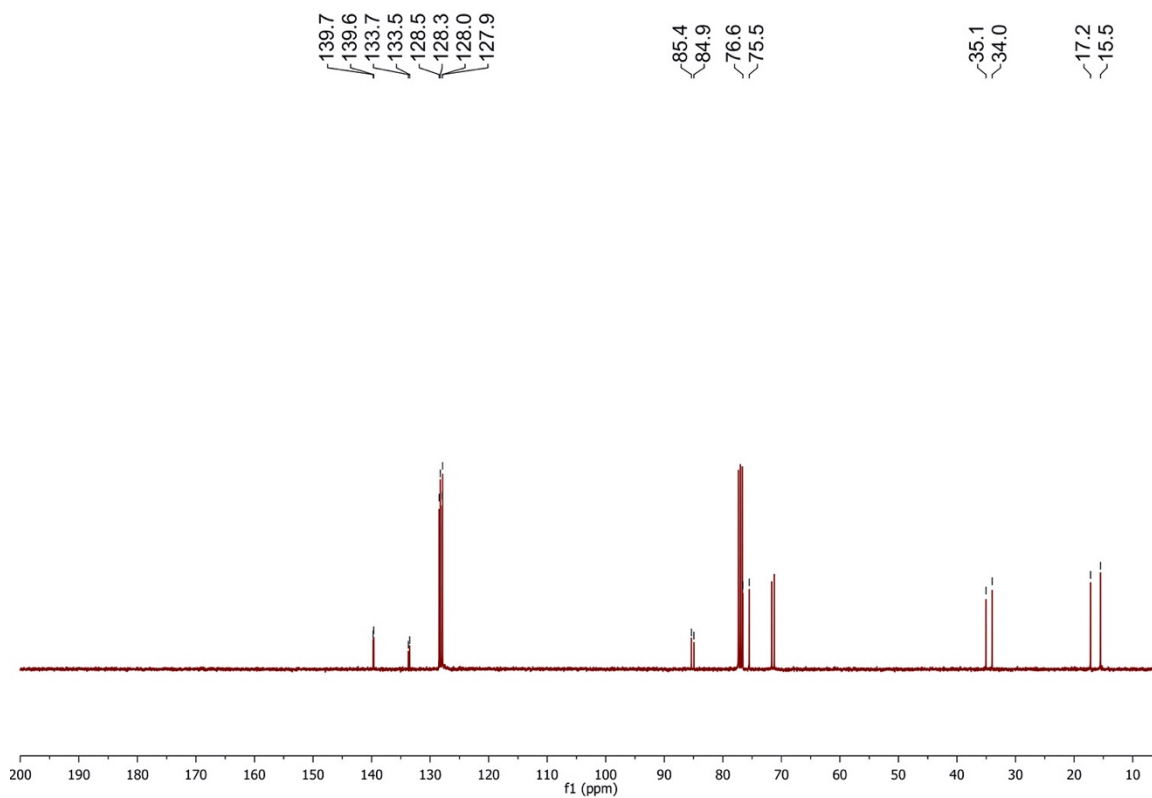


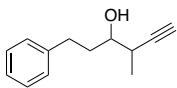


5a ^1H NMR (401 MHz, CDCl_3)

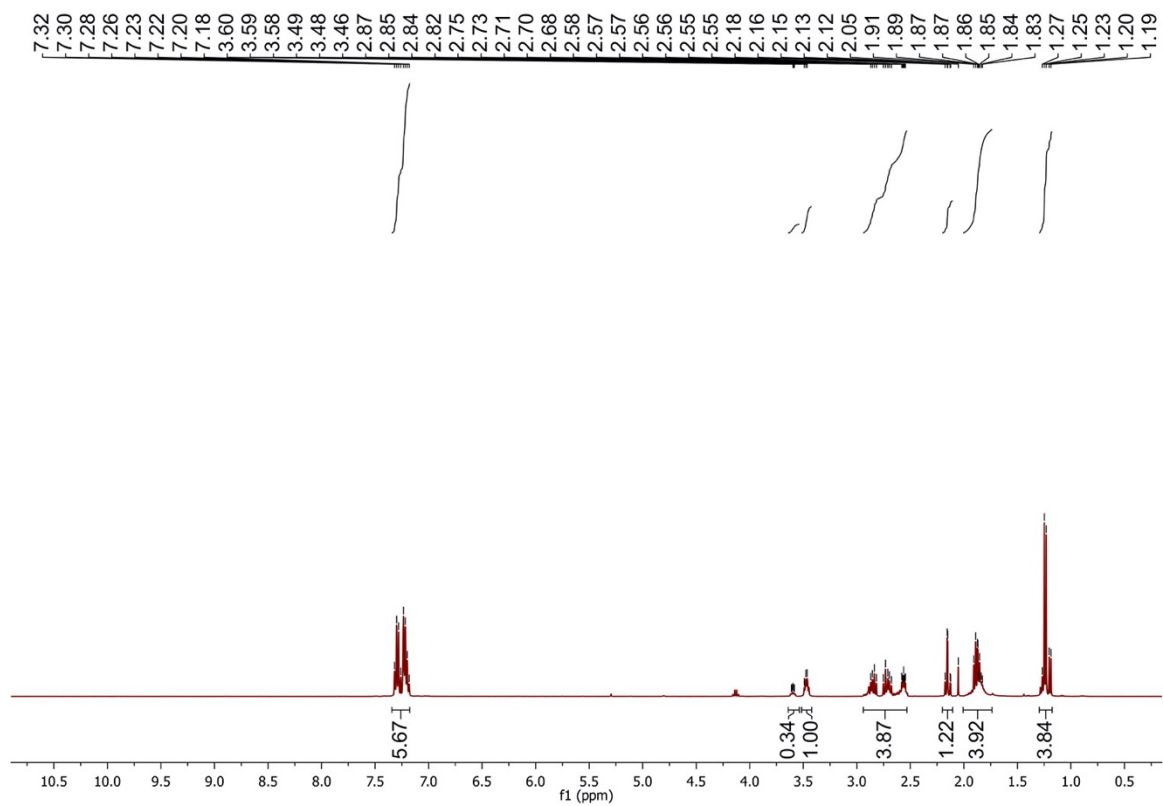


$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)





5b ^1H NMR (401 MHz, CDCl_3)



$^{13}\text{C}\{^1\text{H}\}$ NMR (101 MHz, CDCl_3)

