



ALMA MATER STUDIORUM
UNIVERSITÀ DI BOLOGNA

ARCHIVIO ISTITUZIONALE
DELLA RICERCA

Alma Mater Studiorum Università di Bologna Archivio istituzionale della ricerca

Autologous Fat Grafting for the Oral and Digital Complications of Systemic Sclerosis: Results of a Prospective Study

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Pignatti, M., Spinella, A., Cocchiara, E., Boscaini, G., Luseti, I.L., Citriniti, G., et al. (2020). Autologous Fat Grafting for the Oral and Digital Complications of Systemic Sclerosis: Results of a Prospective Study. *AESTHETIC PLASTIC SURGERY*, 2020, 1-4 [10.1007/s00266-020-01848-2].

Availability:

This version is available at: <https://hdl.handle.net/11585/764646> since: 2020-07-07

Published:

DOI: <http://doi.org/10.1007/s00266-020-01848-2>

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>).
When citing, please refer to the published version.

(Article begins on next page)

This is the accepted manuscript of:

Pignatti, M., Spinella, A., Cocchiara, E. et al. Autologous Fat Grafting for the Oral and Digital Complications of Systemic Sclerosis: Results of a Prospective Study. *Aesth Plast Surg* 44, 1820–1832 (2020).

Final version available: <https://doi.org/10.1007/s00266-020-01848-2>

Rights / License:

The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>)

When citing, please refer to the published version.

AUTOLOGOUS FAT GRAFTING FOR THE ORAL AND DIGITAL COMPLICATIONS OF SYSTEMIC SCLEROSIS. RESULTS OF A PROSPECTIVE STUDY

Running title: Fat grafting in Systemic Sclerosis

Pignatti M, Spinella A, Cocchiara E, Boscaini G, Lusetti IL, Citriniti G, Lumetti F, Setti G, Dominici M, Salvarani C, De Santis G, Giuggioli D.

Marco Pignatti: Plastic Surgery, Policlinico di Sant'Orsola. University of Bologna. DIMES. Bologna, Italy.

ORCID ID: <https://orcid.org/0000-0002-8259-496X>
mrpignatti@gmail.com

Amelia Spinella: MD PhD, Rheumatology Unit, University of Modena and Reggio Emilia, Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy. ORCID ID: <https://orcid.org/0000-0003-4941-8507>
amelia.spinella@unimore.it

Emanuele Cocchiara: MD Rheumatology Unit, University of Modena and Reggio Emilia, Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy.
ORCID ID: <https://orcid.org/0000-0003-0409-7236>
cocchiara2001@yahoo.it

Giulia Boscaini: Plastic Surgery, University of Modena and Reggio Emilia, Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy.
boscaini.giulia@gmail.com

Irene Laura Lusetti: Plastic Surgery, University of Modena and Reggio Emilia, Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy.
irenelaura.lusetti@gmail.com

Giorgia Citriniti:, Rheumatology Unit, University of Modena and Reggio Emilia, Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy.
ORCID ID: <https://orcid.org/0000-0003-1990-2891>
giorgia.citriniti@gmail.com

Federica Lumetti: Research fellow, Rheumatology Unit, University of Modena and Reggio Emilia, Medical School, Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy.
ORCID ID: <https://orcid.org/0000-0003-4671-7982>
fedelumetti@gmail.com

Giacomo Setti: DDS, Unit of Dentistry and Oral-Maxillofacial Surgery, University of Modena and Reggio Emilia, Italy.
ORCID ID: <https://orcid.org/0000-0002-2933-8174>
setti.giacomo@gmail.com

Massimo Dominici MD PhD Department of Medical and Surgical Sciences, University of Modena and Reggio Emilia, Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy.
massimo.dominici@unimore.it

Carlo Salvarani MD. Rheumatology Unit, University of Modena and Reggio Emilia, Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy.
carlo.salvarani@unimore.it

Giorgio De Santis MD. Plastic Surgery, University of Modena and Reggio Emilia, Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy.
giorgio.desantis@unimore.it

Dilia Giuggioli. MD Rheumatology Unit, University of Modena and Reggio Emilia. Azienda Ospedaliero-Universitaria, Policlinico di Modena, Modena, Italy.
ORCID ID: <https://orcid.org/0000-0002-0041-3695>

Corresponding Author:

Marco Pignatti: Plastic Surgery, Policlinico di Sant'Orsola. University of Bologna.
DIMES. Bologna, Italy.
ORCID ID: <https://orcid.org/0000-0002-8259-496X>
mrpignatti@gmail.com
[tel. 00393493774545](tel:00393493774545)

None of the Authors has any commercial interest in the subject of study and does not have received any financial or material support.

Presented in part at the EULAR meeting as Poster (THURSDAY, 13 JUNE 2019).

Spinella A, Pignatti M, Citriniti G, *et al*
THU0349 AUTOLOGOUS FAT GRAFTING IN THE TREATMENT OF
PATIENTS WITH SYSTEMIC SCLEROSIS: CURRENT EXPERIENCE
AND FUTURE PROSPECTS
Annals of the Rheumatic Diseases 2019;**78**:456.

Abstract

Background

Systemic Sclerosis (SSc) is a connective tissue disease. Skin involvement of the mouth and hand may compromise function and quality of life. Autologous fat grafting has been described as a specific treatment of these clinical features.

We report the results of our prospective study designed to treat and prevent skin complications in systemic sclerosis.

Materials and methods

We treated 25 patients with mouth and/or hand involvement (microstomia, xerostomia, skin sclerosis, Raynaud's phenomenon and long-lasting digital ulcers) with autologous fat grafting, according to the Coleman's technique, around the mouth and/or at the base of each finger. The surgical procedures were repeated in each patient every 6 months for a total of two or three times. Clinical data were collected before the first surgery and again 6 months after each surgical procedure.

Pain, skin thickness, saliva production and disability were assessed with validated tests

Results

Overall we performed 63 autologous fat grafting sessions (either on the mouth, on the hands or on both anatomical areas). Results at 6 months after the last session included

improvement of xerostomia evaluated with a sialogram, reduction of the skin tension around the mouth and, in the hands, reduction of the Raynaud phenomenon as well as skin thickness. Pain was reduced while the perception of disability improved. Digital ulcers healed completely in 8/9 patients.

Conclusions

Our results confirm the efficacy and safety of autologous fat grafting for the treatment of skin complications and digital ulcers due to systemic sclerosis. In addition, the patients' subjective well-being improved.

Level of Evidence: IV

Keywords: Systemic Sclerosis, autologous fat grafting, stem cell transplantation, xerostomia, microstomia, digital ulcers

AUTOLOGOUS FAT GRAFTING FOR THE ORAL AND DIGITAL COMPLICATIONS OF SYSTEMIC SCLEROSIS. RESULTS OF A PROSPECTIVE STUDY

INTRODUCTION

Systemic Sclerosis (SSc) is a connective tissue disease characterized by endothelial dysfunction, specific autoimmune abnormalities and accumulation of collagen and other matrix component in the skin and target internal organs [1-3]. The multifaceted Clinical features of SSc appear to be related to the variable contribution of the above mentioned pathogenetic mechanism that together with a multistep process are responsible for the heterogeneous clinical manifestations of SSc.(REF) Fibrotic and vascular manifestations such as Raynaud's phenomenon, digital ulcers (DUs), pulmonary hypertension and cutaneous and visceral fibrosis are the most frequent manifestation of scleroderma [4-8]. The extent of skin fibrosis led to the classification in two distinct clinical subset of SSc namely, limited cutaneous SSc with skin involvement distal to the elbows and knees and diffused cutaneous SSc with skin involvement extending to the proximal limbs and/or trunk [1, 17-19]. Facial involvement frequently present in both subtypes is associated with disfiguring mask-like stiffness of the face [20-22], together with xerostomia and the loss of elasticity and fibrosis of the perioral area, makes eating, drinking, and personal and dental care difficult [8]. DUs are one the most frequent and severe manifestation of SSc , they are present in up to 50% of patients [6, 9-11]. DUs are very painful, hard to heal, may be complicated by infections and gangrene, sometimes requiring amputation [12]. DUs are also related to functional

disability with negative impact on the patients' quality of life. [11-16]. The complex and multifactorial nature of SSc constitutes a great therapeutic challenge.

The discovery of adipose tissue-derived stem cells has opened new therapeutic possibilities in plastic and regenerative surgery. In the last decades autologous fat grafting (AFG) has been successfully used to treat a progressively larger number of clinical conditions characterized by skin atrophy or fibrosis such as radio-induced tissue damage, scars, post-surgical pain [23-28]. Recently, also patients affected by scleroderma have been treated by grafting of autologous fat, with different techniques [29-30].

In this prospective study, we report the results of our treatment protocol with multiple sessions of autologous fat grafting for facial and hand dysfunctions related to systemic sclerosis.

The study was approved by the local ethical committee of the University of Modena and Reggio Emilia (number 275/16) and performed according to the criteria of the Helsinki declaration. All patients gave their written consent.

MATERIALS AND METHODS

We performed repeated surgical procedures in 25 scleroderma patients affected by mouth and/or hand involvement: microstomia, xerostomia, skin sclerosis, and long-lasting digital ulcers. All patients fulfilled the American College of Rheumatology criteria for SSc [7]. The clinical features of the patients at the time of treatment and number of procedures are reported in **Table 1**. To be included in the study, the SSc patients had to have an opening of oral commissure <5 cm or/and skin involvement of

the hands, with functional limitation consisting in abnormal finger flexion and extension, an increased skin thickness as measured by the modified Rodnan Skin Score (mRSS) , presence of Raynaud phenomenon, evaluated with Raynaud Condition Score (RCS).

Digital ulcers lasting for more than 6 months and not responding to conventional systemic and local treatments were present in 9 of the 25 patients.

All the patients received standard medical therapy for scleroderma vasculopathy, (ie: calcium channel blockers, prostanoids and/or anti-endothelin receptors)y, therapy that they did not discontinue during the surgical treatment.

Exclusion criteria were: severe SSc cardio-pulmonary involvement not allowing the patients to undergo surgery; insufficient subcutaneous fat for harvesting; refusal to participate in the study and age under 18 years or over 85 years.

The treatment plan included three sessions of surgery, each one consisting in autologous fat grafting of the affected areas (either the mouth area or the hands or both) performed at intervals of 6 months from each other. **Figure 1**

When patients required both mouth and hand treatment, the two sites were operated on during the same surgical session, and that was considered as a single procedure.

Three procedures, planned every 6 months, of AFG were recommended to all patients even if improved after the first procedure, in the belief, derived from the experience with AFG in other clinical conditions (such as breast reconstruction and radiodermatitis) (31), that they were necessary to induce and maintain the beneficial effects of AFG.

Not every patient completed the three surgical sessions as planned, due to worsening of the clinical condition (not related to treatment) or to personal choice.

The patients' clinical data were collected before the first surgery and again 6 months after each surgical procedure, just before the following procedure. A final follow-up evaluation was performed 6 months after the last surgical procedure, and therefore 12 months (two sessions) or 18 months (three sessions) after the first autologous fat grafting.

The data collected included clinical and serological features of SSc and clinimetric assessment of hands and mouth, sialometry, and the evolution of the digital ulcers, when present.

Clinimetric measures included mouth opening (taken on the midline at maximum forced opening, between the upper and lower lip margin); middle finger-wrist distance (length taken from the tip of middle finger to the distal volar wrist crease with the hand fully extended and supinated); middle finger-palm distance (distance between middle finger tip and palmar skin when the patient was asked to make a fist as a measure of the flexion of fingers on the palm).

Skin thickness was measured with the modified Rodnan Skin Score (mRSS), the Raynaud phenomenon with the Raynaud Condition Score (RCS).

Sialometry, the measurement of the amount of the unstimulated whole saliva (UWS) per minute, is necessary for the diagnosis of hyposalivation. The sialometry, that was performed according to the technique reported in the literature, is considered to be positive for hyposalivation if less than 1.5 ml of saliva are collected in 15 minutes, a production of 0.1 ml/min representing the cut-off limit for diagnosis [32-35].

Furthermore, the patients were asked to fill out a questionnaire in order to express their level of satisfaction with hand and mouth function.

Pain was evaluated by the Visual Analog Scale (VAS) for pain and by the short-form McGill Pain Questionnaire,(SF-MPQ), that investigates sensory and affective descriptors. The perception of disability was measured by the Health Assessment Questionnaire-HAQ and by the Mouth Handicap in Systemic Sclerosis scale-MHISS [16].

Surgical Procedure

All the patients received one preoperative dose of 2g of Cefazolin e.v. (Clindamycin 600mg, if allergic to cephalosporins).

The procedure was performed under sedation and regional block with local anesthetic, to obtain adequate analgesia on the areas where the fat was harvested and where it was injected. Local anesthetic alone would be insufficient to control pain and maintain comfort while repeated sessions of general anesthesia would have been more invasive and unnecessary. Furthermore, orotracheal intubation is difficult in patients affected by microstomia.

We performed tumescent infiltration of the donor areas (flanks or trochanteric areas) with a modified Klein solution (50 ml of saline solution, 0.5 ml of 1:1.000 adrenalin and 10 ml of 2% Mepivacain) through an epidural 23 Gauge needle attached to a 20 ml Luer-lock syringe.

The original Coleman's technique for harvesting and purification of the fat was used [36].

An aspiration cannula (Micro Aspiration Cannulas Black & Black Surgical FAC, 12 gauge x 15 cm Luer-lock) was introduced through a 3 mm incision and a total volume of 30 ml to 50 ml of fat and fluids was aspirated in 10 ml Luer-lock syringes. The syringes were centrifuged for 3 minutes at 3000 rpm/1900 RCF (MPW-223e

centrifuge, "Mpw Medical Instruments" Spółdzielnia Pracy, Warsaw, Poland) and the infranatant fat was isolated and transferred to 2.5 ml syringes.

Mouth

For the treatment of the mouth, after infraorbital block and mental nerve block with mepivacaine 1%, a skin access was created at each naso-jugal crease at the level of the oral commissure with a 19 gauge needle to introduce a Coleman injection cannula (Tissue Injection Cannulas Type I Black & Black Surgical 19gauge x 7cm).

The cannula was connected to the 2.5 ml syringes loaded with the centrifuged, purified autologous fat that was then injected in the subcutaneous and submucosal plane with the retrograde technique, as shown in **Figure 2**.

Two ml of fat were grafted in each of the eight sites around the mouth, for a total of 16 ml (**Figure 3**).

The amount of AFG was standardized. For the perioral area previous experience showed us that due to the thin subcutaneous tissue and the stiff skin and mucosa a volume of 2 ml per each subunit, was adequate to obtain a filling effect, avoiding excessive tissue tension and hypoperfusion. A total of 16 ml was therefore used for the area.

Hands

After wrist block (median, ulnar and dorsal radial nerve) with 5ml of mepivacaine 1%, skin accesses were created with a 19 gauge needle at each finger commissure and/or dorsum of proximal phalanx depending on the tightness of the skin which also determined the maximum amount of fat injected. The fat was grafted around the neurovascular bundle at the base of the fingers in an amount ranging from 0.5 to 1 ml on each side of each finger, for a total of up to 10 ml per hand (**Figure 4**). The volume

injected was never more than 1 ml for each neurovascular bundle to prevent the potentially severe complication of finger ischemia.

Patients that required mouth and hand treatment were treated at both sites during the same session.

Statistical Analysis

All descriptive data were expressed as mean \pm standard deviation (SD). Comparison between values at the baseline and after treatment was performed by paired t-test and Wilcoxon's test for continuous and non-continuous variables, respectively. Data analysis was carried out using an SPSS statistical package (version 22.0, IBM Software, USA) and P value was considered to be significant when <0.05 .

RESULTS

Overall, we performed 63 surgical procedures.

Of the 25 patients treated, 12 interrupted the treatment after the second surgical procedure: six because of other SSc-related medical problems, four because they were satisfied with the benefits already achieved after the first two procedures, and two because no longer followed by our Rheumatology center. The number of patients who received two or three treatments and the sites treated are reported in Table 2

The results obtained are reported below.

Mouth

At the last follow up all the 17 patients affected by microstomia reported subjective improvement of perioral skin tension ($p=0.0238$).

However, the maximum “mouth opening distance” assessment did not significantly change from baseline (mean variation -0.1 , $p = ns$). Figure 5 and 6 show the results obtained in two patients.

A normalization of saliva production to more than $0.1\text{ml}/\text{min}$ was documented in all patients. This result corresponded to a subjective amelioration of xerostomia in 10/14 subjects ($p=0.0269$).

Hands

There was a complete healing of digital ulcers in 8/9 patients ($p=0.0297$) and an improvement in the remaining patient. Figure 7, 8, 9.

All the 12 patients with hand involvement reported an improvement in hand tension ($p=0.0037$), but the improvement of the clinimetric measures was not significant ($p = ns$).

The Raynaud phenomenon, measured with the Raynaud Condition Score, significantly improved from 6.0 ± 1.8 to 3.8 ± 1.6 ($p<0.0001$), while the Modified Rodnan Skin Score showed only a trend towards improvement (from 9.5 ± 6.1 to 8.9 ± 5.7 ; $p=0.083$).

Pain, evaluated by SF-MPQ, scrutinizing sensory ($p=0.3340$) and affective ($p=0.2234$) descriptors, decreased, although not significantly, in all patients, who required lower doses of analgesic drugs.

The clinical changes were mirrored by the subjective feeling of increased well being. In particular, the perception of disability decreased with a trend towards amelioration, when measured with the tests cited above (HAQ, $p=0.063$ and MHISS, $p=0.097$). The subjective assessment and clinimetric measures before and after the last treatment are reported In **Table 3**.

Side effects

Side effects of the treatment included small ecchymotic areas around the mouth, , and temporary edema and paresthesia in the hands of two patients Both problems resolved within the first month after treatment. No other complications, in particular, no vascular occlusions or infections, were recorded.

DISCUSSION

Subcutaneous fat is an excellent source of adult stem cells and it has the advantage of being easy to harvest.

Adipose-derived stem cells (ADSCs), similar to bone-marrow-derived stem cells, are able to differentiate into multiple mesodermal tissue types. In contrast to bone-marrow-derived stem cells, ADSCs can be easily harvested by liposuction, and their abundance avoids the need for expansion in culture. For these reasons the adipose tissue can be considered an innovative source of mesenchymal stem cells suitable for cell-based therapy in regenerative medicine [21, 27, 37].

ADSCin addition to function as filler, have immune-modulatory properties and are able to secrete angiogenetic factors that facilitate tissue repair [37].

Increasing evidence shows that AFG in sclerotic tissues may decrease collagen deposition and increase elasticity and vascularization [37]. In fact, the procedure has been successfully used to regenerate atrophic or fibrotic skin in a large number of

clinical conditions such as radio-dermatitis, burn scars, linear scleroderma, and various types of morphea [24-27, 38].

Several studies have reported the use of fat grafting to treat either mouth or hand SSc complications [29, 30].

We conducted a comprehensive literature search in the MEDLINE, EMBASE, PubMed, Science Citation Index, and Google Scholar databases (up to June 2019) using the keywords "systemic sclerosis", "scleroderma", "microstomia", "digital ulcers", "autologous fat grafting".

A list of the published articles on the topic is reported in Table 4.

Blezien et al. recently reported in this journal the treatment of oral complications of systemic sclerosis in seven patients by fat grafting, demonstrating satisfying results in most of the observed parameters.(39)

Of particular interest is the study by Del Papa et al. who performed a randomized controlled trial comparing AFG with a sham procedure [40], confirming their previous data [41,42] on the efficacy of AFG for the treatment of indolent digital ulcers and mouth opening in SSc patients. Furthermore, they showed an increase in the neovascularization of the treated perioral skin [42].

After collection, fat can be processed in various ways, including the Coleman's technique, as in our study [36]. There is no agreement among authors regarding the best method for processing fat transfer.

We chose the original Coleman's technique because of our previous experience with it in other conditions.

It would have been interesting to have a control group to try a different fat injection technique, but the number of our patients was not sufficient to obtain statistically

significant results. However, the other reported techniques were not shown to have a superior success rate. [21].

A few years ago, Onesti et al. used sedimentation by gravity as a method to eliminate nonviable components of the lipoaspirate in five patients with mouth functional disability while five more were treated with cell-factory prepared adipose-derived stromal cells. At the one-year follow-up they noticed that both procedures obtained significant results in mouth opening capacity and MHISS scores but neither one emerged as the first-choice technique [43].

Fat tissue provides an abundant source of stromal vascular fraction cells for immediate administration and can also give rise to a substantial number of cultured, multipotent adipose-derived stromal cells [44].

Recently Magalon et al. reported the results of a study on the molecular profile and proangiogenic activity of the adipose-derived stromal vascular fraction [45]. The autologous stromal vascular fraction from adipose tissue is an alternative to cultured adipose-derived stem cells. They concluded that the stromal vascular fraction from patients with SSc presents similar distribution of hematopoietic and regenerative subpopulations compared with healthy donors, supporting the rationale for the therapeutic use of autologous SSc-stromal vascular fraction [45].

Whether autologous adipose-derived stromal vascular fraction could turn out to be an unwanted source of profibrotic myofibroblasts in SSc [46] does not seem likely, on the basis of our results in a SSc cohort with an 18 months follow-up. Also, Capelli et al. showed that ADSCs obtained from patients with SSc exhibit phenotypic pattern, proliferation, immunosuppressive properties and differentiation potential that are similar to the ones observed in healthy controls [47].

The actual mechanism of fat graft survival is not completely understood [48], but no significant difference appears to exist in survival of grafted fat obtained from different harvest and implantation techniques [21, 23].

Improvement of safety and outcomes of autologous fat grafting procedures may come from so-called “enrichment strategies”.

A recent review of the literature found that platelet-rich plasma and adipose-derived stem cells appeared to have a beneficial effect when used to augment and improve the viability of fat grafts. However, randomized controlled clinical studies are still needed [49, 50]

On the contrary, differences in the disease process, in the number of stromal adipose stem cells obtained by lipoaspirate, and in the underlying conditions associated with poor revascularization, may significantly impact the engraftment [21, 45].

Our study including 25 SSc patients treated with two or three grafting of autologous fat and followed for up to 18 months, confirmed the role of the AFG technique in the treatment of SSc complications [51-55].

Our study, in comparison to previous published ones, has a large patient population sample, a long follow up, includes treatment of hands in addition to the perioral area, provides a detailed surgical technique description and adds the use of salivation test to obtain objective data on improvement of xerostomia.

In particular, we observed complete healing of long-lasting digital ulcers in 8 /9 patients and a significant improvement in hand tension and in the Raynaud phenomenon as evaluated by means of RCS. (Figure 7, 8, 9)

A subjective improvement in perioral skin tension and in the sensation of xerostomia was also reported. Almadori *et al.* in 2019 reported similar results when investigating mouth function following treatment with lipotransfer in 62 SSc patients (6.85 ± 5.07) ($p < 0.0001$). All of treated patients had manifestation of sicca syndrome, followed by post-operative improvement that contributed of 21,6% (1.51 ± 1.2 , $p < 0.0001$) to MHISS overall score. [56].

In addition we report data of 5 patients in which sialometry was evaluated pre- and post-operatively to provide more evidence to the subjective results.

Despite the limited number of cases, a positive trend in salivary flow rate was observed confirming the improvements reported by the patients.

Several articles report on the use of stem cells therapy in salivary glands hypofunction in animal models. [57,58].

In addition, the differentiation of ADSc in salivary gland cells in association with platelet-rich fibrin (PRF) has been showed in vitro . [59].

Therefore, we hypothesize that, with the support of well-designed clinical and histological studies, AFG could be proposed not only for scleroderma but also for the treatment of “dry mouth” in other rheumatic conditions.

However, the clinimetric measures of mouth opening and of hand flexion and extension did not improve. We believe that the improvement in mouth opening and perioral elasticity could be masked by the concomitant increase of lip volume.

Concerning hands, clinimetric assessment did not show significant enhancement, probably because joint involvement and tendon retraction, frequently observed in SSc, were responsible for the hand disability, and were therefore less influenced by AFG treatment.

Finally, our patients reported an overall subjective amelioration in their quality of life and did not complain of any significant adverse effect, demonstrating a good safety profile of the technique.

CONCLUSIONS

In conclusion, the results obtained in 25 patients treated with two or three sessions of AFG and evaluated by multiple tests, including the spit test, confirmed the efficacy of AFG to treat the perioral complications of SSc. In addition, there was a modest healing of digital ulcers, associated to a subjective improvement of pain. These clinical results were reflected by the subjective improvement in well-being of the patients.

REFERENCES

1. LeRoy EC, Black C, Fleischmajer R, Jablonska S, Krieg T, Medsger Jr TA, et al. (1988). Scleroderma (systemic sclerosis): classification, subsets and pathogenesis. *J Rheumatol*; 15:202-5.
2. Medsger Jr TA, Steen VD. (2004). Classification and prognosis. In: Clements PJ, Furst DE, editors. *Systemic sclerosis*. Philadelphia: Williams & Wilkins; p.51-64.
3. Denton CP. (2015). Systemic sclerosis: from pathogenesis to target therapy. *Clin Exp Rheumatol*; 33(4Suppl92):3-7.
4. Elhai M, Meune C, Avouac J, Kahan A, Allanore Y. (2012). Trends in mortality in patients with systemic sclerosis over 40 years: a systematic review and meta-analysis of cohort studies. *Rheumatology*; 51:1017-26.
5. Ioannidis JPA, Vlachoyiannopoulos PG, Haidich AB, Medsger TA, Lucas M, Michet CJ, et al. (2005). Mortality in systemic sclerosis: an international meta-analysis of individual patient data. *Am J Med*;118:2-10.
6. Ferri C, Valentini G, Cozzi F, Sebastiani M, Michelassi C, La Montagna G, et al. (2002). Systemic sclerosis: demographic, clinical, and serologic features and survival in 1,012 Italian patients. *Medicine*; 81:139-53.

7. Van den Hoogen F, Khanna D, Fransen J, et al. (2013) Classification criteria for systemic sclerosis: an American College of Rheumatology/European League against Rheumatism collaborative initiative. *Arthritis Rheum* 65:2737-47.
8. Hachulla E, Launay D. (2011). Diagnosis and classification of systemic sclerosis. *Clin Rev Allergy Immunol*;40:78-83.
9. Ferri C, Sebastiani M, Lo Monaco A, Iudici M, Giuggioli D, Furini F, Manfredi A, Cuomo G, Spinella A, Colaci M, Govoni M, Valentini G. (2014). Systemic sclerosis evolution of disease pathomorphosis and survival. Our experience on Italian patients' population and review of the literature. *Autoimmun Rev* 13(10):1026-34.
10. Walker UA, Tyndall A, Czirják L, Denton C, Farge-Bancel D, Kowal-Bielecka, et al. (2007) Clinical risk assessment of organ manifestations in systemic sclerosis: A report from the EULAR Scleroderma Trials and Research Group database. *Ann Rheum Dis* 66:754-763.
11. Steen V, Denton CP, Pope JE, Matucci-Cerinic M. (2009). Digital ulcers: Overt vascular disease in systemic sclerosis. *Rheumatology*; 48(Suppl 3):19-24.
12. Giuggioli D, Manfredi A, Colaci M, et al. (2012). Scleroderma digital ulcers complicated by infection with fecal pathogens. *Arthritis Care Res* 64:295-7.
13. Hachulla E, Clerson P, Launay D, et al. (2007). Natural history of ischemic digital ulcers in systemic sclerosis: single-center retrospective longitudinal study. *J Rheumatol* 34:2423-30.

14. Mouthon L, Carpentier PH, Lok C, et al (2014). ECLIPSE Study Investigators. Ischemic digital ulcers affect hand disability and pain in systemic sclerosis. *J Rheumatol* 41:1317-23.
15. Guillevin L, Hunsche E, Denton CP, et al, (2013). DUO Registry Group. Functional impairment of systemic scleroderma patients with digital ulcerations: results from the DUO Registry. *Clin Exp Rheumatol* 31:71-80.
16. Rannou F, Poiraudau S, Berezne' A, et al. (2007). Assessing disability and quality of life in systemic sclerosis: construct validities of the Cochin Hand Function Scale, Health Assessment Questionnaire (HAQ), Systemic Sclerosis HAQ, and Medical Outcomes Study 36-Item Short Form Health Survey. *Arthritis Rheum* 57:94-102.
17. Clements PJ, Lachenbruch PA, Seibold JR, et al. (1993). Skin thickness score in systemic sclerosis: an assessment of interobserver variability in 3 independent studies. *J Rheumatol* 20:1892-6.
18. Daumas A, Rossi P, Arieu-Bonnet D, et al. (2014). Generalized calcinosis in systemic sclerosis. *QJM* 107:219-21.
19. Mouthon L. (2013). Hand involvement in systemic sclerosis. *Presse Med* 42:1616-26.
20. Maddali-Bongi S, Del Rosso A, Mikhaylova S, et al. (2014). Impact of hand and face disabilities on global disability and quality of life in systemic sclerosis patients. *Clin Exp Rheumatol*. 32(6Suppl86):15-20.
21. Magalon G, Daumas A, Sautereau N, Magalon J, Sabatier F, Granel B. (2015). Regenerative Approach to Scleroderma with Fat Grafting. *Clin Plast Surg*. 42(3):353-64.

22. Arkachaisri T, Vilaiyuk S, Li S, O'Neil KM, Pope E, Higgins G, et al. (2009). Localized Scleroderma Clinical and Ultrasound Study Group. The localized scleroderma skin severity index and physician global assessment of disease activity: A work in progress toward development of localized scleroderma outcome measures. *J. Rheumatol* 36(12):2819-2829.
23. Coleman SR. (2006) Structural fat grafting: more than a permanent filler. *Plast Reconstr Surg* 118:108S-120S.
24. Rigotti G, Marchi A, Galie M, et al. (2007). Clinical treatment of radiotherapy tissue damage by lipoaspirate transplant: a healing process mediated by adipose-derived adult stem cells. *Plast Re- constr Surg* 119:1409-22 discussion 1423-4.
25. Klinger M, Caviggioli F, Klinger FM, et al. (2013). Autologous fat graft in scar treatment. *J Craniofac Surg* 24:1610-15.
26. Juhl AA, Karlsson P, Damsgaard TE. (2016) Fat grafting for alleviating persistent pain after breast cancer treatment: A randomized controlled trial. *J Plast Reconstr Aesthet Surg.* 69(9):1192-202.
27. Coleman SR, Katzel EB. (2015) Fat grafting for facial filling and regeneration. *Clin Plast Surg.* 42(3):289-300
28. Rigotti G, Charles-de-Sá L, Gontijo-de-Amorim NF, et al. (2016). Expanded stem cells, stromal-vascular fraction, and platelet-rich plasma enriched fat: comparing results of different facial rejuvenation approaches in a clinical trial. *Aesthet Surg J.* 36(3):261-270
29. Scuderi N, Ceccarelli S, Onesti MG, Fioramonti P, Guidi C, Romano F, et al. (2013). Human adipose-derived stromal cells for cell-based therapies in the treatment of systemic sclerosis. *Cell Transplant* 22(5):779-95.

30. Bene MD, Pozzi MR, Rovati L, Mazzola I, Erba G, Bonomi S. (2014). Autologous fat grafting for scleroderma-induced digital ulcers. An effective technique in patients with systemic sclerosis. *Handchir Mikrochir Plast Chir.* 46(4):242-7.
31. Rigotti G1, Marchi A, Galiè M, Baroni G, Benati D, Krampera M, Pasini A, Sbarbati A. Clinical treatment of radiotherapy tissue damage by lipoaspirate transplant: a healing process mediated by adipose-derived adult stem cells. *Plast Reconstr Surg.* 2007 Apr 15;119(5):1409-22; discussion 1423-4.
32. Navazesh M, Christensen C, Brightman V. (1992). Clinical criteria for the diagnosis of salivary gland hypofunction. *J Dent Res.* 71(7):1363-9.
33. Navazesh M. (1993). Methods for collecting saliva. *Ann N Y Acad Sci.* 694:72-7.
34. Falcão DP, da Mota LM, Pires AL, Bezerra AC. (2013) Sialometry: aspects of clinical interest. *Rev Bras Reumatol.* 53(6):525-31.
35. Varoni EM, Federighi V, Decani S, Carrassi A, Lodi G, Sardella A. (2016). The effect of clinical setting on the unstimulated salivary flow rate. *Arch Oral Biol.* 69:7-12.
36. Coleman SR. (2002). Hand rejuvenation with structural fat grafting. *Plast Reconstr Surg* 110(7):1731-44; discussion 1745-7.36.
37. Gimble JM, Katz AJ, Bunnell BA. (2007). Adipose-derived stem cells for regenerative medicine. *Cir Res.* 100 (9):1249-60.
38. Chen B, Wang X, Long X, Zhang M, Huang J, Yu N, et al. (2018). Supportive Use of Adipose-Derived Stem Cells in Cell-Assisted Lipotransfer for Localized Scleroderma. *Plast Reconstr Surg* 141(6):1395-1407.

39. Blezien O, D'Andrea F, Nicoletti GF, Ferraro GA. (2017). Effects of Fat Grafting Containing Stem Cells in Microstomia and Microcheilia Derived from Systemic Sclerosis. *Aesthetic PlastSurg* 41(4):839-844.
40. Del Papa N, Di Luca G, Andracco R, Zaccara E, Maglione W, Pignataro F, et al. (2019). Regional grafting of autologous adipose tissue is effective in inducing prompt healing of indolent digital ulcers in patients with systemic sclerosis: results of a monocentric randomized controlled study. *Arthritis Res Ther* 21(1):7.
41. Del Papa N, Di Luca G, Sambataro D, Zaccara E, Maglione W, Gabrielli A, et al. (2015). Regional implantation of autologous adipose tissue-derived cells induces a prompt healing of long-lasting indolent digital ulcers in patients with systemic sclerosis. *Cell Transplant*. 24(11):2297-305.
42. Del Papa N, Caviggioli F, Sambataro D, Zaccara E, Vinci V, Di Luca G, et al. (2015). Autologous fat grafting in the treatment of fibrotic perioral changes in patients with systemic sclerosis. *Cell Transplant* 24(1):63-72.
43. Onesti MG, Fioramonti P, Carella S, Fino P, Marchese C, Scuderi N. (2016). Improvement of Mouth Functional Disability in Systemic Sclerosis Patients over One Year in a Trial of Fat Transplantation versus Adipose-Derived Stromal Cells. *Stem Cells Int* 2016:2416192. doi: 10.1155/2016/2416192.

44. Bora P, Majumdar AS. (2017). Adipose tissue-derived stromal vascular fraction in regenerative medicine: a brief review on biology and translation. *Stem Cell Res Ther* 8(1):145.
45. Magalon J, Velier M, Simoncini S, François P, Bertrand B, Daumas A, et al. (2019). Molecular profile and proangiogenic activity of the adipose-derived stromal vascular fraction used as an autologous innovative medicinal product in patients with systemic sclerosis. *Ann Rheum Dis.* 78(3):391-398.
46. Manetti M. (2019). Could autologous adipose-derived stromal vascular fraction turn out an unwanted source of profibrotic myofibroblasts in systemic sclerosis? *Ann Rheum Dis.* 2019 Mar 13. pii: annrheumdis-2019-215288. doi: 10.1136/annrheumdis-2019-215288.
47. Capelli C, Zaccara E, Cipriani P, Di Benedetto P, Maglione W, Andracco R, et al. (2017). Phenotypical and Functional Characteristics of In Vitro-Expanded Adipose-Derived Mesenchymal Stromal Cells From Patients With Systematic Sclerosis. *Cell Transplant.* 26(5):841-854.
48. Granel B, Daumas A, Jouve E, Harle' J-R, Nguyen P-S, Chabannon C, et al. (2014). Safety, tolerability and potential efficacy of injection of autologous adipose-derived stromal vascular fraction in the fingers of patients with systemic sclerosis: an open-label phase I trial. *Ann Rheum Dis* 81:2056.
49. Vyas KS, Vasconez HC, Morrison S, Mogni B, Linton S, Hockensmith L, Kabir T, Zielins E, Najor A, Bakri K, Mardini S. Fat Graft Enrichment Strategies: A Systematic Review. *Plast Reconstr Surg.* 2020 Mar;145(3):827-841. doi: 10.1097/PRS.0000000000006557.

50. Xiong S, Yi C, Pu LLQ.
An Overview of Principles and New Techniques for Facial Fat Grafting. *Clin Plast Surg*. 2020 Jan;47(1):7-17. doi: 10.1016/j.cps.2019.08.001. Epub 2019 Oct 21. Review.
51. Guillaume-Jugnot P, Daumas A, Magalon J, Sautereau N, Veran J, Magalon G, et al. (2016). State of the art. Autologous fat graft and adipose tissue-derived stromal vascular fraction injection for hand therapy in systemic sclerosis patients. *Curr Res Transl Med* 64(1):35-42.
52. Daumas A, Magalon J, Jouve E, Truillet R, Casanova D, Giraud L, et al. (2017). Long-term follow-up after autologous adipose-derived stromal vascular fraction injection into fingers in systemic sclerosis patients. *Curr Res Transl Med* 65(1):40-43.
53. Sautereau N, Daumas A, Truillet R, Jouve E, Magalon J, Veran J, et al. (2016). Efficacy of Autologous Microfat Graft on Facial Handicap in Systemic Sclerosis Patients. *Plast Reconstr Surg Glob Open* 4(3):e660.
54. Virzì F, Bianca P, Giammona A, Apuzzo T, Di Franco S, Mangiapane LR, et al. (2017). Combined platelet-rich plasma and lipofilling treatment provides great improvement in facial skin-induced lesion regeneration for scleroderma patients. *Stem Cell Res Ther* 8(1):236.
55. Gheisari M, Ahmadzadeh A, Nobari N, Iranmanesh B, Mozafari N. (2018). Autologous Fat Grafting in the Treatment of Facial Scleroderma. *Dermatol Res Pract* 2018 Aug 1;2018:6568016. doi: 10.1155/2018/6568016. eCollection 2018.

56. Almadori A, Griffin M, Ryan CM, Hunt DF, Hansen E, Kumar R, Abraham DJ, Denton CP, Butler PEM.
Stem cell enriched lipotransfer reverses the effects of fibrosis in systemic sclerosis. *PLoS One*. 2019 Jul 17;14(7):e0218068.
doi:10.1371/journal.pone.0218068. eCollection 2019.
57. Yoo C, Vines JB, Alexander G, Murdock K, Hwang P, Jun HW. Adult stem cells and tissue engineering strategies for salivary gland regeneration: a review. *Biomater Res*. 2014 Jul 24;18:9. doi: 10.1186/2055-7124-18-9.
eCollection 2014. Review.
58. Lim JY, Ra JC, Shin IS, Jang YH, An HY, Choi JS, Kim WC, Kim YM.
Systemic transplantation of human adipose tissue-derived mesenchymal stem cells for the regeneration of irradiation-induced salivary gland damage. *PLoS One*. 2013 Aug 9;8(8):e71167. doi: 10.1371/journal.pone.0071167.
eCollection 2013.
59. Dai TQ, Zhang LL, An Y, Xu FF, An R, Xu HY, Liu YP, Liu B.
In vitro transdifferentiation of adipose tissue-derived stem cells into salivary gland acinar-like cells. *Am J Transl Res*. 2019 May 15;11(5):2908-2924. eCollection 2019.

LEGENDS to FIGURES

Fig 1: *study flowchart.*

The autologous fat was grafted in the affected sites: either in the perioral area or in the fingers or in both sites.

After 6 months, clinical data were collected and a second AFG treatment was offered.

After another 6 months, clinical data were collected again and a third AFG treatment was offered.

After another 6 months a final follow-up data collection concluded the study

Fig 2: *Injection cannula connected to a 2.5ml syringe loaded with purified fat is introduced laterally to the oral commissure*

Fig 3: *The eight schematic areas of grafting: three in the upper lip, three in the lower lip (one central and two lateral), one at each commissure. Two ml of fat were injected with the retrograde technique into each area using many radiating passages at the subcutaneous level and in the submucosal plane, for a total of 16 ml of purified fat.*

Fig 4: *After creating a skin access with a 19 gauge needle at the medial and lateral dorsal side of each proximal phalanx, an injection cannula, connected to a syringe, is introduced. One finger of the surgeon controls, by palpation, the correct position of the tip of the cannula around the neurovascular bundle at the volar side of the*

phalanx, where the fat is positioned with retrograde technique. In this image, the cannula is introduced in the ulnar side of the proximal phalanx of the middle finger

Fig 5: Before a)c) and after b)d) treatment pictures of one patient with closed and open mouth. Improvement of lip thickness (comparison a-b) and mouth opening (c-d)

Fig 6: Before a)c) and after b)d) treatment pictures of one patient with closed and open mouth. Improvement of lip thickness (comparison a-b) and mouth opening (c-d)

*Fig 7: Second finger of the right hand. Proximal inter phalangeal joint exposure
a) pre-operative image, b) 1 week after AFG, c) 1 month after AFG*

Fig 8:

Improvement of skin fibrosis of the hands and healing of the fingertip ulcers

a), Before treatment

b) 6 months after the second session of AFG

c) 2 years after the third session of AFG

Fig 9:

Gradual healing of the middle fingertip ulcer

a) Before treatment

b) 6 months after the first session of AFG

c) 6 months after the second session of AFG

d) 2 years after the third session of AFG

LEGENDS to TABLES

Table 1. *Clinical features of SSc patients before treatment. Number of procedures*

lcSSc-L/dcSSc-D: limited/diffuse cutaneous systemic sclerosis

Scl70/ACA: anti-topoisomerase I or anti-Scl 70 antibodies/anti-centromere antibodies

RCS: Raynaud Condition Score (0-10)

mRSS:modified Rodnan Skin Score

Table 2: *The number of patients who received two or three treatments and the sites treated*

Table 3: *Subjective assessment and clinimetric measures before and after treatment.*

RCS: Raynaud Condition Score

mRSS: modified Rodnan Skin Score

HAQ: Health Assessment Questionnaire. Perception of disability measure

VAS: VisualAnalog Scale

SF-MPQ: Short-form McGill Pain Questionnaire . Pain measure, scrutinizing sensory and affective descriptors

MHISS: Mouth Handicap in Systemic Sclerosis scale- Perception of disability measure.

ns: not significant

Table 4: *Published studies on AFG in cohorts of SSc patients*

Pts: patients;

DUs: digital ulcers.

w: weeks;

mo: months;

yr: years.

MHISS: Mouth Handicap in Systemic Sclerosis.

CRRT: cutaneous resonance running time.

Kind of graft: AFG: autologous fat grafting;

PRP: platelet rich plasma;

ADSVF: autologous adipose-derived stromal vascular fraction;

ADSCs: adipose-derived stem cells. ASCs: adipose- derived stromal cells.