

Alma Mater Studiorum Università di Bologna
Archivio istituzionale della ricerca

Natural Compounds as a Strategy to Optimize " In Vitro" Expansion of Stem Cells

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Natural Compounds as a Strategy to Optimize " In Vitro" Expansion of Stem Cells / Tullia Maraldi, Cecilia Prata , Pasquale Marrazzo, Silvana Hrelia , Cristina Angeloni. - In: REJUVENATION RESEARCH. - ISSN 1549-1684. - ELETTRONICO. - 23:2(2020), pp. 93-106. [10.1089/rej.2019.2187]

Availability:

This version is available at: <https://hdl.handle.net/11585/758942> since: 2020-05-15

Published:

DOI: <http://doi.org/10.1089/rej.2019.2187>

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>).
When citing, please refer to the published version.

(Article begins on next page)

This is the final peer-reviewed accepted manuscript of:

Maraldi, T., Prata, C., Marrazzo, P., Hrelia, S., & Angeloni, C. (2019). Natural Compounds as a Strategy to Optimize “In Vitro” Expansion of Stem Cells. *Rejuvenation Research*, 23(2), 93–106.

The final published version is available online at: <https://doi.org/10.1089/rej.2019.2187>.

Rights / License:

The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>)

When citing, please refer to the published version.

Natural Compounds as a Strategy to Optimize “*In Vitro*” Expansion of Stem Cells

Tullia Maraldi,¹ Cecilia Prata,² Pasquale Marrazzo,³ Silvana Hrelia,^{3,*} and Cristina Angeloni^{4,*}

Abstract

The efficient use of stem cells for transplantation is often limited by the relatively low number of stem cells collected. The *ex vivo* expansion of human stem cells for clinical use is a potentially valuable approach to increase stem cell number. Currently, most of the procedures used to expand stem cells are carried out using a 21% oxygen concentration, which is about 4- to 10-fold greater than the concentration characteristic of their natural niches. Hyperoxia might cause oxidative stress with a deleterious effect on the physiology of cultured stem cells. In this review, we investigate and critically examine the available information on the ability of natural compounds to counteract hyperoxia-induced damage in different types of stem cells *ex vivo*. In particular, we focused on proliferation and stemness maintenance in an attempt to draw up useful indications to define new culture media with a promoting activity on cell expansion *in vitro*.

Keywords: stem cell expansion, natural compounds, hyperoxia, oxidative stress, cell proliferation, stemness maintenance

Introduction

HUMAN STEM CELLS are being investigated for the treatment of many different diseases, like cardiovascular and autoimmune diseases and traumatic lesions.¹ Among stem cells, particular interest has been focused on mesenchymal stem cells (hMSC) that can be derived from many tissues but are often insufficient to reach a relevant number of cells adequate for clinical use. Moreover, patient age, gender, genetic background, and clinical history influence their quantity and quality.

At present, most procedures to expand stem cells are carried out at an ambient oxygen concentration where cells are exposed to 21% O₂, which is 4- to 10-fold greater than the concentration characteristic of their “*in vivo*” niches.^{2,3} Several studies have demonstrated the deleterious impact of a 21% oxygen tension on the physiology of stem cells, for example, neuronal stem cells,⁴ bone marrow stem cells,^{5,6} umbilical cord stem cells,⁷ or adipose tissue stem cells.^{8,9} In fact, *in vitro* culture of MSCs exposed to a 21% oxygen concentration might increase the production of reactive oxygen species (ROS) and, as a consequence, to be subjected to environmental oxidative stress. In general, ROS,

including hydrogen peroxide (H₂O₂) and superoxide anion (O₂^{•−}), plays crucial roles in stem and progenitor physiology. Nevertheless, the “redox window” hypothesis must be considered: suitable ROS level is vital for physiological cellular functions,¹⁰ whereas excess ROS contribute to adverse effects on MSCs, including premature senescence, increase of the population doubling time, damages to DNA,^{11,12} and reduced engraftment after transplantation.¹³ Human MSCs are particularly susceptible to early senescence and typically halt proliferation at approximately passage 8, which limits their expansion potential and therapeutic value.¹⁴ Another issue to be considered is the need of long-term cryopreservation of stem cells, a routine practice that requires extremely controlled and safe techniques of specimen storage. However, more and more studies are showing contradictory results on the influence of stem cell cryopreservation and thawing, such as wide stresses at physical and biological level, mitochondrial damages, modifications to basal respiration and ATP synthesis, apoptosis and necrosis, shortening of telomeres and cellular senescence, damage to DNA, and oxidative stress.¹⁵ These results have raised serious alarms concerning the therapeutic effectiveness and safety of stem cells. Ability to

¹Department of Surgery, Medicine, Dentistry and Morphological Sciences, University of Modena and Reggio Emilia, Modena, Italy.

²Department of Pharmacy and Biotechnology, Alma Mater Studiorum—University of Bologna, Bologna, Italy.

³Department for Life Quality Studies, Alma Mater Studiorum—University of Bologna, Rimini, Italy.

⁴School of Pharmacy, University of Camerino, Camerino, Italy.

*These authors contributed equally to this work.

enhance MSC culture expansion and delay senescence could substantially improve efficacy for autologous stem cell therapy. On these bases, a promising strategy to counteract oxidative stress induced by hyperoxia or by cryopreservation and thawing could be the use of compounds with antioxidant activity. In particular, many natural antioxidant compounds have been demonstrated to counteract oxidative stress by directly scavenging free radicals or activating antioxidant defense pathways.^{16–18} Moreover, the safety of many natural antioxidant compounds has been largely demonstrated^{19–21} and this is an important issue considering that stem cell expansion has principally performed with the intent of implantation.

Another important issue to take into consideration in using natural antioxidant compounds to counteract oxidative stress in stem cells is that these compounds can induce changes in cells that should be checked before using them for treatment. A good antioxidant compound should not impair stem cell stemness, potency, and proliferation. In this review we investigate and critically examine the available information on the ability of natural compounds to counteract hyperoxia-induced damage in different type of stem cells *ex vivo*. In particular, we focused on proliferation and stemness maintenance in an attempt to draw up useful indications to define new culture media with a promoting activity on cell expansion *in vitro*.

From the Niche to the Bank

In living organisms, stem cells reside in a specific micro-environment called niche. Quiescent stem cells are located in the niche until specific signals finish the dormant state and activate the differentiation process. In particular, this dynamic microenvironment accomplishes primarily three functions, thanks to secreted or cell surface molecules: stem cell proliferation control, stem cell daughters fate determination, and protection of stem cells from exhaustion or death.²² Not only structural and soluble factors, but also hypoxia plays a crucial role for the stemness maintenance of mesenchymal, embryonic, neural stem, and hematopoietic cells, and could also impact on proliferation and differentiation.²³

In MSC niches, the local oxygen concentrations fluctuate depending on stem cell type: between 10% and 15% O₂ in the adipose tissue,²⁴ about 1.3% both in the amniotic fluid and amniotic membrane,²² and between 1% and 7% O₂ in bone marrow.²⁵

The bone marrow niche is the most studied in literature. Although bone marrow has a highly complex vascular network, the entire bone interior is relatively hypoxic, with pO₂ in the range of 1%–4%.²⁶ In general, HSCs stay quiescent in the bone marrow niche, whereas various stimuli that cause loss of quiescence induce strong entry into the cell cycle, and trigger proliferation frequently accompanied by stress, apoptosis, and damage to DNA.^{27,28} Zhou et al.²⁹ observed that a low-oxygen niche in bone marrow reduces ROS generation, therefore providing long-term protection against oxidative stress. *Ex vivo* expansion techniques should promote symmetric stem cell divisions,³⁰ and consequently stem cell self-renewal without inducing differentiation and apoptosis.

Indeed, oxygen concentration has a major role in the proliferation and differentiation of stem cells, also in the placenta. It has been demonstrated that cytotrophoblasts

proliferate in low-oxygen tension (2% O₂), whereas they differentiate in high-oxygen tension (20% O₂).³¹ The reduction of oxygen tension below a specific threshold leads to the initiation of a gene expression program by the transcription factor, hypoxia-inducible factor-1 (HIF-1). HIF-1 signaling pathway plays a fundamental role in regulating key processes such as energy metabolism, angiogenesis, cell proliferation, and viability depending on the availability of oxygen in the cell (reviewed in Ref.³²). The low-oxygen tension stabilizes HIF-1, the oxygen-sensitive monomer of the HIF-1, since its ubiquitination and proteasomal degradation are prevented.^{33,34} HIF-1 is linked to transcription of at least 70 genes holding functional hypoxia-responsive element. Most of these genes are involved in cell survival, through the expression of cell surface receptors, angiogenic growth factors, glucose transporters, and glycolytic enzymes.^{34,35} Moreover, under low-oxygen tension stem cell pluripotency is maintained by HIF-2 interaction with the promoter region of OCT4 causing increased transcriptional activity.³⁶ OCT4 is the principal regulator of pluripotency in embryonic stem cells (ESCs) and essential to obtain induced pluripotent stem cells (iPSCs) but disappears upon differentiation.³⁷

For example, bone marrow HSCs (BHSCs) are quiescent *in vivo* through conservation of a hypoxic phenotype with high levels of HIF-1 α protein and the hypoxic marker pimonidazole, even when contiguous to arterioles.³⁸ Thus, in preserving quiescence into the niche, HSCs must depend on anaerobic glycolysis for energy production.³⁹ Indeed, glycolysis is induced by HIF-1 α in response to hypoxia, expressing pyruvate dehydrogenase kinase 1 (PDK1), which inhibits pyruvate dehydrogenase and avoids mitochondrial acetyl-Co-A production and oxidation. The clear result is a metabolic shift that induces glycolysis and decreases ROS generation in hypoxia. Correspondingly, expansion of MSCs in atmospheric oxygen levels upregulates OXPHOS genes and mitochondrial respiration, and accelerates cellular senescence, while hypoxic culture conditions delay senescence.⁴⁰

It is known that the use of O₂ in several fundamental metabolic processes by living organisms has got an evolutionary price, as O₂ metabolism is the main source of ROS in the cells. The condition known as oxidative stress occurs when the endogenous production of ROS exceeds the cellular capacity of the antioxidant systems. Unfortunately, not always the cellular adaptive processes are able to counteract the accumulation of oxidative damage to key molecules, such as lipids, DNA, and proteins.²⁹ While all the cells of an organism might be subjected to the accumulation of damages triggered by oxidative damage, the effects of ROS on stem cells in most self-renewing tissues are of specific interest to the processes of aging and cancer development owing to their undifferentiated condition and longevity of replicative potential.^{41,42}

Excessive ROS production leads to the overexpression of the tumor suppressor genes, p21, p53, and p16.^{43,44} Accordingly, oxidative stress is one of the principal causes of physiological decline during cellular senescence. Forkhead box (Fox) O proteins control ROS generation in stem cells by regulating catalase and SOD expression.^{45,46} It has been observed that the increase of ROS level in FoxO-deficient mice is associated with the disruption of HSC quiescence, increase in HSC apoptosis, and impairment in hematopoietic repopulating abilities.

In animal models with damaged cellular antioxidant defense system, such as SOD2, an antioxidant enzyme that is present in mitochondria and modulated by FoxO3, a potential correlation between ROS and HSC impairment has been observed. SOD2 evoked a protective activity in mouse neural stem cells (NSCs), and its upregulation improved NSC's survival both *in vitro* and *in vivo*. In addition, the alterations related to aging in SOD2 modulation by FoxO3 and DNA damage, recognized by serine/threonine protein kinase ATM (ataxia telangiectasia mutated), contribute to HSC functionality reduction with aging.⁴⁶ HSC self-renewal capacity relies on ATM-mediated reduction of ROS production as demonstrated by studies carried out in *Atm*^{-/-}. There is a crosstalk with the ATM-p53 signaling converging on p16 to trigger stem cell senescence.⁴⁷

Other key modulators of aging, oxidative stress, and stem cell functionality are sirtuins (SIRT), a family of NAD-dependent protein deacetylases. Recently it has been suggested, a role for SIRT1 in maintaining MSC growth and differentiation that have been observed to deteriorate with age. Interestingly, its activation in aged cells protects the cells from p53-dependent apoptosis or senescence.⁴⁸ SIRT3, is crucial in the aged HSCs in the maintenance of mitochondria metabolism and in controlling oxidative stress. Moreover, ectopic upregulation of SIRT3 enhances the function of HSCs during aging by increasing SOD2 antioxidant activity.⁴⁹

On these bases, the aging of stem cells may be controlled by regulating their metabolic and redox state, which in turn might impact on the intracellular accumulation of ROS. The condition defined as “cell culture stress” might induce genetic instability and therefore constitutes a biological risk for cell therapy protocols. These pathways should be reversed or avoided to obtain an efficient stem cell expansion *in vitro*.

Recently, the increasing interest in the use of MSCs in the clinical practice has led to a high number of studies on the effects of culture conditions and expansion on pathways, behavior, and function of stem cells. As *in vivo* HSC self-renewal and differentiation are regulated by niche cells secreting cytokines and growth factors, including pleiotrophin, angiopoietin-1, -3, thrombopoietin, granulocyte colony-stimulating factor, stem cell factor, and Notch ligands,⁵⁰ cytokines were the first drugs studied for HSC *ex vivo* expansion. Overall, cytokines and their combinations can maintain HSCs and progenitors counteracting apoptosis in *in vivo* proliferation but leading to a contained human HSCs (hHSCs) expansion. For this reason, further elements are obviously essential for a positive expansion of hHSCs *ex vivo*.

The impact of oxygen concentration in adult stem cell phenotype, morphology, proliferative capacity, and functionality has been largely investigated with controversial outcomes.

For example, MSCs cultured in low oxygen tension (1%–10% oxygen) has revealed enhanced regenerative/reparative properties in animal models of heart, brain, and lung damage.^{51–53}

In particular, several studies have shown that human stem cells cultured in a range of 1%–5% of oxygen tensions showed improved cell proliferation and longer lifespan.^{5,12,54,55} Culturing in hypoxia have also been demonstrated to avoid double-strand breaks (DSB) and chromosomal abnormalities in several types of stem cells.^{56,57} In agreement with this notion, Estrada

et al.¹¹ showed that, compared with cells grown at 3% O₂, short-term growth of hMSC at 20% O₂ tension enhances oxidative stress and DNA damage markers, such as DSB, chromosomal aberrations, aneuploidy, and telomere shortening rates. In this study, they showed that expansion at 3% O₂ significantly increases the growth rate of adipose tissue-derived hMSC with a better conservation of telomere length. However, telomerase activity in hMSC was low and insensitive to O₂ concentration suggesting that the protection from oxidative stress, due to the growth at 3% O₂, avoids telomere erosion.

Moreover, hypoxia upregulated the expression of OCT4 and CXCR7 in MSCs, and deferred modifications in phenotypic characteristics such as morphology, cell size, and senescence.⁵⁸ Furthermore, besides these effects, hypoxia also increased the levels of MSC-secreted factors, such as matrix metalloprotease-(1 and 3) and secreted lysyl oxidase (LOX), which are required for migration and cell matrix adhesion.⁵⁹

However, other authors described different results. For example, Pezzi et al.⁶⁰ found that cells exposed to protracted hypoxia behaved similar to their controls in normoxia concerning viability, immunophenotyping, differentiation, and ROS levels. Cells subjected to low oxygen tensions (4%–1%) showed lower proliferation, smaller cell size, greater cellular complexity, lower mitochondrial activity, and autophagy. These data validate a study by Holzwarth et al.,⁶¹ that, by culturing MSCs in conditions of 21%, 5%, 3%, and 1% oxygen, concluded that hypoxia limits MSC's proliferation in incubation period of 7 days. This inconsistency could be due to heterogeneity of culture conditions, such as supplementation and period of exposure to hypoxia, as well as intradonor variability that may explain the contradictory effects of hypoxia on the proliferation and differentiation capacities of MSCs.^{61–64} Furthermore, donor's age directly worsens cell proliferative capacity, concurring to the variability of results.⁶⁵

Supplementation with molecule affecting ROS levels has been tested in many studies. For example, El Alami et al.⁶⁶ demonstrated that for human dental pulp stem cells (hDPSCs) cultured under 21% O₂, oxidative stress promotes the sequential activation of p38 MAPK, p21, and the nuclear factor erythroid 2-related factor 2 (Nrf-2) antioxidant defense pathway. The antioxidant properties of p21 depend on Nrf2 activity that upregulates HO-1 and NQO1. Therefore, incubation with Trolox, a water-soluble analog of vitamin E, or with a p38 inhibitor, reestablishes high proliferation rate of hDPSCs even under 21% O₂.

Source of ROS and Antioxidant Defense System in Stem Cells

ROS include nonradical species, such as hydrogen peroxide and singlet oxygen, as well as radical species (superoxide, hydroxyl, hydroperoxy, peroxy, and alkoxy radicals). Biologically, superoxide and hydrogen peroxide play a pivotal role in modulating redox signaling.⁶⁷ The effects exerted by these molecules is related to their nature, intracellular levels, and localization: ROS can be involved not only in macromolecule's oxidative damage and cell death induction, but also in signal transduction linked to cell proliferation and gene transcription activation.⁶⁸ In particular, numerous experimental evidences suggest a clear

correlation between intracellular H_2O_2 levels and features of stem and progenitor cells.^{10,41,69–71} As is well known, excess ROS contributes to pathological conditions, nevertheless, a proper and controlled ROS production is mandatory for physiological cellular functions, the so-called and previously cited “redox window” hypothesis.^{72,73}

Much remains to be learned about the identification of the main ROS source in stem cells. In mammalian cells, a number of ROS sources are known, including: mitochondrial OXPHOS,^{67,74} plasma membrane NOXs,⁷⁵ cytochrome P-450, and b5 enzymes at endoplasmic reticulum level,⁷⁶ peroxisome oxidases,⁷⁷ lipoxygenases,⁷⁸ and xanthine oxidase.⁷⁹

The fewer mitochondria content of stem cell compared with their progeny is in accordance with their utilization of glycolysis rather than OXPHOS for ATP production.^{39,80} Since it is often assumed that mitochondria are the main ROS source,⁶⁷ members of NOX family could represent major cellular sources of ROS in HSCs, as also suggested by Drehmer et al.⁸¹ in relation to the metabolic switches for adipogenic stem cell differentiation. hHSCs indeed express NOX1, 2, and 4 and various regulatory subunits of NOXs.^{80,82} and Piccoli et al. estimated that NOX-mediated extramitochondrial O_2 consumption accounts for about half of the endogenous cell respiration in hHSCs.⁸⁰ Interestingly, mouse BHSC-enriched LSK-cells express NOX1, 2, and 4, whereas NOX4 is not present in HPCs, Lin[−] cells, and mononuclear cells from mouse BM, suggesting that NOX4 expression is downregulated during HSC differentiation and that NOX4 may play an important role in the regulation of HSC function.⁸³ However, the increase in ROS production in HSCs is associated with upregulation of NOX4 expression,⁸³ but is not related to mitochondrial dysfunction nor elevated mitochondrial superoxide production.⁸⁴

Nevertheless, it has been shown that cells, including HSCs, from Bmi1^{−/−} mice exhibit abnormal mitochondrial function with consequently an increased ROS production.⁸⁵ In addition, an elevation of mitochondrial biogenesis and oxidative activities seems to be related to an increased production of ROS in HSCs from Tsc1^{−/−} mice.⁸⁶

ROS derived from various cellular sources and compartments could be used by different stem cells to perform unique functions. For example, Udx1 (a dual NOX), allows the conversion of $\text{O}_2^{\bullet-}$ to H_2O_2 to prevent polyspermy and regulate cleavage in the zygote of the sea urchin.⁸⁷ Cardiovascular differentiation of ESCs depends on increased ROS generation derived from various NOXs.⁸⁸ However, an increase in mitochondrial mass and production of ATP and ROS is linked to human ESC (hESC) differentiation.⁸⁹

To regulate redox signaling and to counteract excessive amounts of ROS derived from aerobic metabolism, a complex antioxidant system plays an important role. In particular, MnSOD plays a primary role in the cell protection from mitochondrial ROS. Nevertheless, although MnSOD has been extensively studied in relation to aging and tumor biology, the crucial role of MnSOD in stem cell physiology still remains unclear.⁹⁰ The cytosolic Cu/Zn SOD isoform not only is present in stem cell but also represent a factor secreted by MSCs. Importantly, MSCs have been reported to restore the radiation-induced reduction of Cu/Zn SOD levels after whole thorax irradiation, suggesting a paracrine action exerted by MSC-derived SOD1 in the protective function exerted by MSC.⁹¹

Outside mitochondria, the glutathione-based systems (GSH/GSSG ratio regulated by glutathione S-transferase, glutathione peroxidase/reductase) usually represent the major redox-regulatory mechanisms leading to H_2O_2 -scavenging mechanism involving its reduction to water and a similar reaction is catalyzed by CAT inside the peroxisome.

Thioredoxin is mainly localized in the endoplasmic reticulum and reduces disulfide bonds of oxidatively modified proteins. Sundaramoorthy et al.⁹² demonstrated that the treatment with TXN, 24 hours following irradiation, mitigates radiation-induced cell injury of hematopoietic stem in mice.

Natural Compounds to Optimize Expansion of Stem Cells *In Vitro*

Many studies have explored the effect of natural antioxidant compounds against hyperoxia-induced oxidative stress in *ex vivo* cultures of different stem cells to increase cell proliferation, maintain potency, counteract senescence and DNA damages (Table 1). In particular, purified phytochemicals and their potential *in vivo* metabolites, plant extracts, combinations of purified compounds, and/or extracts have been investigated.

Purified Phytochemicals

Many purified phytochemicals have been studied for their ability to enhance proliferation and maintain stem cell potency, and the majority of them are polyphenolic compounds, in particular flavonoids (Fig. 1).

Epigallocatechin gallate [(−)-cis-3,3',4',5,5',7-hexahydroxyflavane-3-gallate] is the most abundant and biologically active catechin in green tea. This compound is a polyphenol belonging to the flavonoid family whose members are known for their *in vitro* and *in vivo* antioxidant activity.⁹³ Wang et al.⁹⁴ showed that epigallocatechin gallate treatment (10–40 μM) significantly increased cell proliferation of adult hippocampal neural progenitor cell cultures. Another study demonstrated that low concentrations (0.1 μM) of epigallocatechin gallate increased the number of dividing cells and stimulated neurosphere formation of cochlea NSCs, confirming the positive effect of this flavonoid on stem cell proliferation.⁹⁵

Quercetin (3,3',4',5,7-pentahydroxyflavone) is a natural flavonoid present in many edible plants, including grapes, apples, onions, and berries.⁹⁶ The effect of this flavonoid, along with its major *in vivo* metabolite, quercetin-3-O-glucuronide, has been considered in cultures of human embryonic NSCs.⁹⁷ The results showed that quercetin and quercetin-3-O-glucuronide have totally opposed effects on cell viability. Quercetin (20–60 μM) decreased NSC viability, decreased Akt phosphorylation, and increased apoptosis. In contrast, quercetin-3-O-glucuronide (20–60 μM) increased proliferation through the Akt/cyclin D1 and BDNF signaling pathway. Quercetin's negative effect on cell viability was also observed in hESCs. In particular, quercetin (25–200 μM) induced mitochondrial ROS production that were responsible for quercetin-mediated hESC cell death.⁹⁸ From our point of view, as quercetin is a very active compound, the range of concentrations tested in these studies are too high to evoke a proliferative effect. Further studies should be carried out to assess lower quercetin

TABLE 1. PHYTOCHEMICALS WITH A POSITIVE EFFECT ON STEM CELL PROLIFERATION AND STEMNESS MAINTENANCE IN VITRO

Compounds	Effective concentration	Type of stem cell	Molecular targets	References
Epigallocatechin-3-gallate	10–40 μ M	Mice hippocampal neuronal progenitor cells	Not investigated	88
Quercetin-3-O-glucuronide	0.1 μ M	Mouse cochlear NSCs	Not investigated	89
Kaempferol	20–60 μ M	Human embryonic NSCs	\uparrow Akt/cyclin D1 and \uparrow BDNF signaling pathway	91
Naringenin	20 μ M	mESCs	\uparrow Nanog	94
	1–100 μ g/mL	hBMSCs	Not investigated	96
	1–100 μ g/mL	hAFSCs	Not investigated	97
Icariin	10 nM–1 μ M	hBMSCs		99
	100 nM	mNSCs	\uparrow ERK	100
	10 μ M	hNSCs	\uparrow FGFR1, \uparrow Wnt family, \downarrow GSK-3 β	101
	100 μ M	rNSCs	\uparrow cyclin D1, \uparrow p21	102
	20–320 μ M	rBMSCs	\uparrow pERK, \uparrow pp38, \uparrow Elk-1, \uparrow c-Myc	103
	100 nM	rASCs	\uparrow RhoA/ROCK signaling pathway	104
Icaritin	5–10 nM	mESCs	\downarrow CDX2 and p130, \uparrow cyclin E/CDK2, \uparrow Oct4, \uparrow Nanog, \uparrow Klf4, \uparrow Sox2	99,106
	100 nM	hMSCs	\uparrow STAT3 signaling pathway	107
Chrysin	Loaded on 3D nanofibers (15% wt)	hASCs	\uparrow Oct4, \uparrow Nanog, \uparrow Rex-1, \uparrow Sox2	108
	Loaded on 3D nanofibers (5 μ M)	mMSCs	\downarrow cyclin C1, \uparrow cyclin B1, \uparrow cyclin E1	109
3,2'-dihydroxyflavone	5 μ M	mESCs and iPSCs	\uparrow STA3, \uparrow Akt, \downarrow GSK-3 β , \downarrow ERK, \uparrow Oct4, \uparrow Nanog, \uparrow Fgf4, \uparrow Sox2, \uparrow Cripto	110
Morin hydrate	1–2 mg/L	mNSCs		112
Resveratrol	10 nM–10 μ M	hMSCs	\uparrow NO/cGMP signal	114
	10 nM–10 μ M	hMSCs	\uparrow Estrogen receptor dependent MAPK pathway	115
	0.1 μ M	hMSCs	\uparrow Sirtuin 1, \uparrow Sirtuin 2, \uparrow Birc4, \uparrow Birc5	116
	50–500 nM	mESCs	\uparrow PcnA, \uparrow cyclin D, \uparrow cyclin A, \uparrow Oct4, \uparrow Sox2,	117
	50 μ M	hESCs	\uparrow SIRT1-ERK signaling pathway	118
Di-(2-ethylhexyl) phthalate	100–600 μ M	rNSCs	\uparrow Sox2	132
Silymarin	50–100 μ gM	hBMSCs	\uparrow GSH	139
Daucosterol	5 μ M	rNSCs	\uparrow IGF1, \uparrow pAkt	119
Salvianolic acid B	5–20 μ M	rNSPCs	\uparrow pAkt, \uparrow Nestin, \uparrow Notch-1	121
Oleuropein	10–100 μ M	hASCs		123
Curcumin	0.1–0.5 μ M	C17.2 cells	\uparrow pERK, \uparrow pp38	125
Astaxanthin	5–10 ng/mL	Mouse neuronal progenitor cells	\uparrow PI3K and MEK signaling pathway, \uparrow Rex1, \uparrow CDK1, \uparrow CDK2, \uparrow Oct4, \uparrow Sox2, \uparrow Nanog, \uparrow Klf4	128
Essential oil of <i>Lippia origanoides</i>	1 μ g/mL	hASCs		130
Hydroethanolic extract <i>Cirsium vulgare</i>	200–1000 μ g/mL	rNSCs		132
Water extract of <i>Hibiscus sabdariffa</i>	0.5–1.0 μ g/mL	mBMSCs	\uparrow GSH, \uparrow SOD	135
<i>Drynaria fortune</i>	0.1 g/L	DPSCs		136
<i>Epimedium</i>	200–400 μ g/mL	Rat hippocampal NSCs		137
Blueberry extract, green tea extract, carnosine, vitamin D3	500 ng/mL, 500 ng/mL, 20 μ M, 5 μ M	Human hematopoietic stem cells		140
Epigallocatechin-3-gallate, sulforaphane	10 μ M, 1 μ M	hAFSCs	\uparrow GSH, \uparrow NQO1, \uparrow TR, \uparrow Oct4, \uparrow Nanog, \downarrow p16, \downarrow β -gal	141

3D, three-dimensional; ASCs, adipose-derived stem cells; ERK, extracellular signal-regulated kinase; hAFSCs, human amniotic fluid-derived stem cells; hBMSCs, human bone mesenchymal stem cells; hESCs, human embryonic stem cells; iPSCs, induced pluripotent stem cells; mBMSC; mESCs, murine embryonic stem cells; mMSCs, mouse mesenchymal stem cells; NSCs, neural stem cells; ROCK, rho-associated protein kinase; STAT3, signal transduction activator transcription factor 3; rASCs; rBMSCs; rNSCs; rNSPCs; mNSCs; hNSCs; hASCs; TR, thioredoxin reductase.

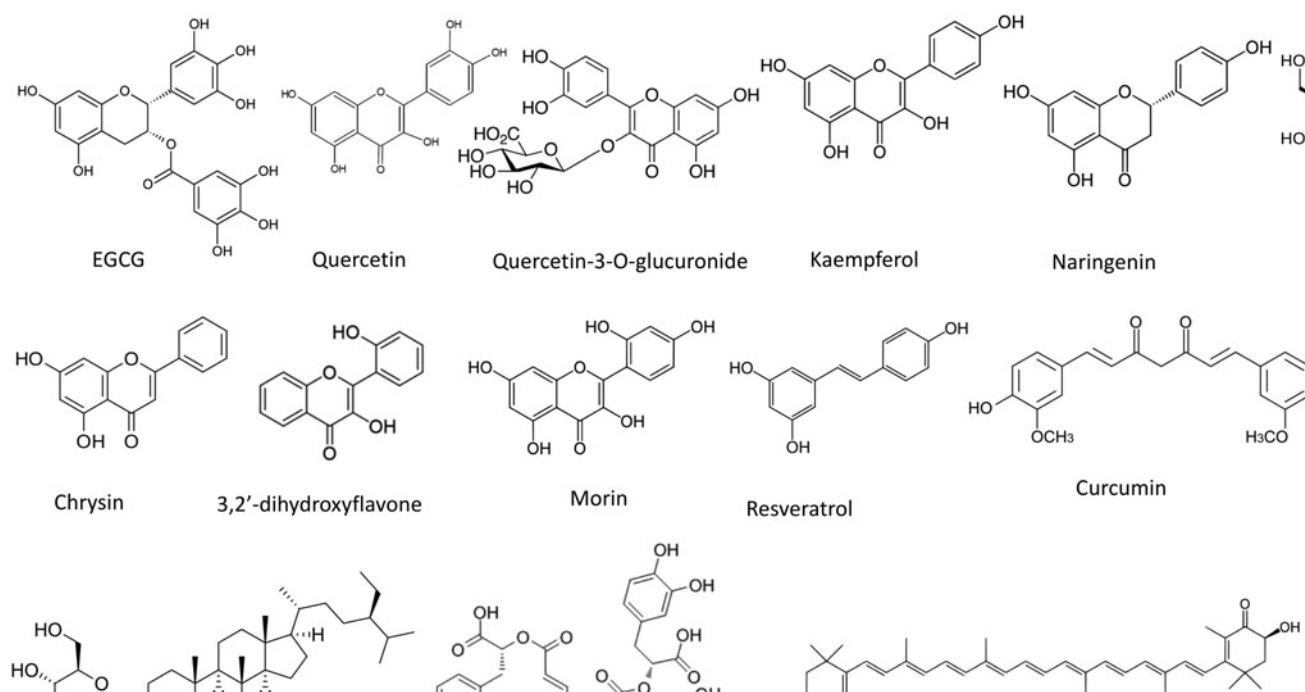


FIG. 1. Chemical structure of phytochemicals used to optimize expansion of stem cell *in vitro*.

concentrations on NSCs viability. Our hypothesis is reinforced by the study of Nichols et al.,⁹⁹ which showed that 0.3–3 μM quercetin, but not 10 μM quercetin, increased cell viability of primary cortical neuron subjected to oxygen-glucose deprivation.

Kaempferol (3,4',5,7-tetrahydroxyflavone) is a flavonoid structurally similar to quercetin apart from the lack of the hydroxyl group in 3' position. The treatment with high concentrations (200 μM) of kaempferol negatively affected murine ESCs (mESCs) viability and growth and increased mitochondrial ROS, although the resulting cell colonies were smaller but remain pluripotent.¹⁰⁰ Nevertheless, lower concentrations (20 μM) of this flavonoid increased the pluripotency marker expression in mESCs but, at the same time, inhibited proper differentiation of mESCs.

Naringenin (4',5,7-trihydroxyflavanone) is a flavonoid compound belonging to the flavanone class mostly present in citrus fruits, including lemon, orange, tangerine, and grapefruit.¹⁰¹ The supplementation of naringenin (1–100 $\mu\text{g/mL}$) to human bone MSCs (hBMSCs) resulted in enhanced cell proliferation.¹⁰² The activity of naringenin was also studied in human amniotic fluid-derived stem cells (hAFSCs) by Liu et al.¹⁰³ In agreement with the previous study, in the presence of naringenin, hAFSC proliferation increased dose dependently between 1 and 100 $\mu\text{g/mL}$. Unfortunately, no mechanisms have been suggested to explain the observed effect on cell proliferation.

Icariin (4'-O-methyl-8- γ,γ -dimethylallylkaempferol-3-rhamnoside-7-glucoside) and its derivatives are flavonoids obtained from traditional Chinese medicinal herbs of the *Epimedium* genus.¹⁰⁴ The first study that observed a promoting activity of icariin on proliferation of stem cells was carried out by Fan et al.¹⁰⁵ Icariin in the range 10 nM–1 μM induced the *in vitro* cell proliferation of hBMSCs. The same effect was observed on NSCs derived from forebrains of mice embryos.¹⁰⁶ Icariin promoted neurosphere formation

and proliferation of NSCs in a dose-dependent manner, achieving the maximum effects at 100 nM. The authors suggested that these effects were partially mediated by the extracellular signal-regulated kinase (ERK)/MAPK. The regulatory roles of icariin in increasing cell proliferation of NSCs *in vitro* was also investigated in human NSCs derived from fetuses.¹⁰⁷ The treatment of human NSCs with 10 μM icariin led to a higher proliferation rate compared with control cells. The microarray analysis carried out in this study showed that icariin modulates the expression of different genes in the Wnt and bFGF pathways, in particular, icariin significantly enhanced the expression of FGFR1 and three important Wnt family members: CTNNB1, FZD7, DVL3, and, at the same time, reduced the expression of GSK-3 β . These results were further defined and broadened by Fu et al.¹⁰⁸ who observed that icariin promoted the growth and proliferation of rat hippocampus NSCs in a dose-dependent manner through the regulation of the cell cycle genes, cyclin D1 and p21. Higher concentrations of icariin (20–320 μM) increased proliferation of rat BMSCs.¹⁰⁹ The increased rBMSC proliferation appeared to be mediated by the phosphorylation of ERK and p38 and by the overexpression of MAPK targets, Elk-1 and c-Myc. Another study evidenced that 100 nM icariin promotes *in vitro* proliferation of rat adipose-derived stem cells (ASCs) through the RhoA and Rho-associated protein kinase (ROCK) signaling pathways.¹¹⁰ Icariin has also been investigated in relation to the cellular damages caused by oxidative stress experienced during cryopreservation and thawing.¹¹¹ The addition of icariin to the freezing solution containing DMSO significantly increased the postthawed cell viability, decreased the apoptosis rate, improved cell adherence, and maintained the mitochondrial functions, as compared with the freezing solution containing DMSO alone. Moreover, icariin inhibited oxidative stress and upregulated heat shock proteins 70 and 90.

Icaritin [3,5,7-trihydroxy-2-(4-methoxyphenyl)-8-(3-methylbut-2-enyl)chromen-4-one], an icariin derivative, has been shown to increase proliferation and enhance self-renewal of mouse ESCs.^{105,112} Icaritin cooperates and stimulates ER α , functioning as a effective inhibitor of CDX2 and p130, which successively activates cyclin E/CDK2 signaling to trigger G1/S phase progression and upregulates pluripotency markers like Klf4, Oct4, Nanog, and Sox2. The proliferative effect of different concentrations of icaritin for different times has also been investigated in MSCs.¹¹³ Icaritin treatment increased MSC proliferation after 2 and 3 weeks of exposure with the highest MSC proliferation measured with 100 nM icaritin treatment. The study suggests that icaritin promotes the proliferation of human adult MSCs by the activation of the signal transduction activator transcription factor 3 (STAT-3), with a resulting increase of cysteine (C)-X-C motif chemokine receptor 4 (CXCR4) expression and activity.

Another highly investigated flavonoid in relation to the optimization of stem cell cultures *in vitro* is chrysin (5,7-dihydroxyflavone), a flavone predominantly present in honey, propolis, and passion fruit.¹¹⁴ In a recent article, ASCs were cultured on three-dimensional nanofibrous mats loaded with ASCs.¹¹⁴ Interestingly, ASCs, cultured in the presence of chrysin, exhibited improved adhesion and increased viability compared with cultures on nanofibers without chrysin or tissue culture polystyrene. Moreover, ASCs on chrysin-based nanofibers showed enhanced stemness maintenance capability evidenced by the overexpression of four stemness markers: Nanog, Sox-2, Oct-4, and Rex-1. In another study, chrysin was incorporated into biocomposite scaffolds encompassing chitosan, carboxymethyl cellulose, and nanohydroxyapatite, and mouse MSCs (mMSCs) were seeded on these scaffolds.¹¹⁵ Chrysin released from scaffolds enhanced cell proliferation down-regulating cyclin C1 mRNA, thus inducing the transition of cells from G0 phase to G1 phase, upregulating Cyclin B1 and E1 and promoting the progression of mMSCs from G1 phase to S and G2/M phases.

A study explored the effect of different flavonoids characterized by different hydroxylation pattern in the B ring of skeleton of diphenylpropane (C6 C3 C6) on cell growth and pluripotency marker expression in mouse embryonic stem cells and iPSCs.¹¹⁶ Among the tested compounds only 3,2'-dihydroxyflavone increased cell growth and the expression of pluripotency marker Sox2, Oct4, Nanog, Cripto, and Fgf4. The mechanisms behind these effects were a strong activation (AKT and STAT3) or inhibition (ERK and GSK3 β) of self-renewal-related kinase of both ESCs and iPSCs. These data suggest that 3,2'-dihydroxyflavone could be useful for the maintenance of high-quality pluripotent ESCs and iPSCs.

Morin hydrate (3,5,7,2',4' pentahydroxyflavone) is a polyphenol compound extracted from *Prunus dulcis*, *Castanea sativa*, *Morus alba*, and other fruits.¹¹⁷ It has been observed that morin hydrate (1–2 mg/L) promotes growth and proliferation of isolated mouse inner ear NSCs as measure by MTT assay and BrdU incorporation.¹¹⁸

Resveratrol (3, 5, 4'-trihydroxystilbene), a nonflavonoid polyphenol with stilbene structure, is found in different plants such as cranberries, blueberries, grapes, peanuts, and eucalyptus.¹¹⁹

The first studies that evidenced a stimulatory activity of resveratrol on stem cell proliferation were conducted on human bone marrow-derived mesenchymal stem cells (hMSCs).^{120,121} In particular, 10 nM to 10 μ M resveratrol induced mBMSCs cell proliferation measured by [³H]-thymidine incorporation through the NO/cGMP signal¹²⁰ and stimulates hBMSCs proliferation through an estrogen receptor-dependent mechanism and coupling to ERK1/2 phosphorylation.¹²¹ To gain better understanding of the effect of this polyphenols on MSCs, Peltz et al.¹²² investigated the effect of short-term (14 days) versus long-term (28–70 days) exposure to resveratrol at different concentrations (0.1–10 μ M) in hMSCs. Resveratrol showed a dose-dependent activity on hMSCs self-renewal as a result of its combinatorial effect on cell doubling time, and on cell senescence and cell proliferation rate. At 0.1 μ M, resveratrol showed the highest benefit, enhancing stem cell self-renewal over both short- and long-term exposure, meanwhile 5 or 10 μ M treatment inhibited cell self-renewal. In accordance with these observations, the expression of genes involved in cell survival (Sirtuin 1, Sirtuin 2, Birc4, and Birc5) was increased by a lower resveratrol concentration but inhibited by a higher resveratrol concentration. A study conducted on mouse ESCs showed that nM concentrations of resveratrol (50 and 500 nM) restored the pluripotency of ESCs differentiated with retinoic acid by modulating the JAK-STAT3 signaling pathway which, in turn, stimulates the transcription of the stemness key factors, OCT4 and SOX2.¹²³ In addition, the same resveratrol concentrations enhanced ESC proliferation through the induction of genes related to proliferation, such as Cyclin A, Cyclin D, and PcnA. Of note, the authors observed that higher concentrations of resveratrol played a nonsignificant role in the regulation of ESC self-renewal suggesting that the timing and concentration of resveratrol are fundamental in eliciting its final activity on self-renewal of ESCs. These aspects were further investigated by Safaeinejad et al.¹²⁴ who indicated that 50 μ M resveratrol might induce cell proliferation of hESCs by the modulation of the cell cycle. Interestingly, this resveratrol concentration did not negatively affect pluripotent markers and upregulated antiapoptotic markers. The authors suggested that resveratrol maintained self-renewal of hESCs at least in part through “SIRT1-MEK/ERK” axis.

Daucosterol (β -sitosterol β -D-glucoside) is a phytosterol present in many higher plants, especially in plant oils, nuts, and seeds.¹²⁵ It has been observed that 5 μ M daucosterol induced rat NSC proliferation and increased the percentage of daughter cells reentering the cell cycle by the modulation of numerous genes, especially increasing the expression of IGF1 and phosphorylation of Akt.¹²⁵

Salvianolic acid B is a hydrophilic compound present in *Salvia miltiorrhiza Bunge*, commonly known as Danshen and widely used for the treatment of various diseases, such as cardiovascular disease, Alzheimer's, hyperlipidemia, and acute cerebrovascular disease.¹²⁶ Salvianolic acid B significantly increased the proliferation of rat neural stem/pre-cursor cells (NSPCs) by activating the PI3K/Akt signal pathway and maintained NSPC's self-renewal by increasing the expression of two self-renewal markers, Nestin and Notch-1.¹²⁷

Oleuropein is a phenolic compound present in olive oil with high antioxidant capacity.¹²⁸ Human adipose-derived

MSCs showed a higher proliferative rate in the presence of 10–100 μ M oleuropein as measured by WST assay.¹²⁹

Curcumin is a polyphenol isolated from the rhizomes of the plant *Curcuma longa* (turmeric) and is a commonly used spice throughout the world.¹³⁰ This natural antioxidant has been investigated in relation to the proliferation of multipotent neural progenitor cells (C17.2 cells).¹³¹ Low doses (0.1 and 0.5 μ M) of curcumin induced C17.2 cell proliferation, whereas high doses (≥ 10 μ M) triggered a cytotoxic effect. The authors demonstrated that the mechanisms by which curcumin elicits this effect is by activating ERK and p38 MAP kinases.

Astaxanthin is a xanthophyll carotenoid ubiquitous in nature, especially in the marine environment, and is found in high amounts in algae, and in aquatic animals such as salmon, trout, krill, and lobster.¹³² It has been shown that astaxanthin possesses a particularly strong antioxidant capacity compared with other carotenoids.¹³³ Treatment with astaxanthin (5–10 ng/mL) increased proliferation and colony formation of mouse neural progenitor cells in a dose/time-dependent manner through the activation of the PI3K and MEK signaling pathways.¹³⁴ Moreover, astaxanthin upregulated transcription factors associated to proliferation, such as CDK1, CDK2, and Rex1, and increased the expression of stemness genes (OCT4, Nanog, SOX2, and KLF4) suggesting that astaxanthin can also improve stem cell potency.

In conclusion, different phytochemicals showed a promising activity in increasing stem cell proliferation and promoting stemness maintenance suggesting their use in *ex vivo* expansion of stem cells.

Plant Extracts

Not only purified phytochemicals, but also extracts from officinal and not officinal plants have been demonstrated to influence stem cell expansion and stemness maintenance.

Lippia origanoides is an aromatic plant widely found throughout the Brazilian Amazon. Its essential oil has antioxidant activity, thanks to the presence of the monoterpenoids, thymol, p-cymene, and myrcene, and the sesquiterpene, caryophyllene.¹³⁵ The supplementation of the essential oil from *L. origanoides* (1 μ g/mL) in the culture medium of stem cells isolated from human adipose tissue induced a strong increase of cell proliferation especially at passage 3.¹³⁶ The authors suggested that this effect could be attributed to thymol, the principal constituent of the oil of *L. origanoides* used in the study (88.2%).

Cirsium vulgare is a common plant belonging to Asteraceae family whose main components are flavonoids and phenolic acids.¹³⁷ The effects of a *C. vulgare* hydroethanolic extract (200–1000 μ g/mL) and its main component, di-(2-ethylhexyl) phthalate (100–600 μ M) have been investigated on the proliferation and pluripotency of neonatal rat hippocampus-derived NSCs. Both the extract and di-(2-ethylhexyl) phthalate at the higher concentrations were able to increase proliferation and the phthalate upregulated the expression of the stemness marker Sox2.¹³⁸

Hibiscus sabdariffa is a plant rich in flavonoids whose biological effects seem to be mediated by its antioxidant activity.^{139,140} The supplementation of a water extract of *H. sabdariffa* to murine bone marrow-derived hematopoietic stem cells showed a cell-genoprotective potential and modulated proliferation and intracellular antioxidant status,

without causing a remarkable loss of surface marker expression for HSC.¹⁴¹

Huang et al.¹⁴² studied the proliferative effect of a commercial flavonoid extract of *Drynaria fortunei*, a common type of traditional Chinese herb, on stem cells isolated from rat dental pulp. The extract (0.01–0.1 g/L) induced dental pulp stem cell proliferation in a dose-dependent manner by increasing the number of stem cells entering into S phase dose dependently.

Epimedium flavonoids, extracted from a traditional Chinese *Epimedium* herb, promoted the proliferation of rat NSCs.¹⁴³ Unfortunately, the authors did not characterize the extract they used.

Silymarin, a combination of flavonolignans obtained from *Silybum marianum*, has been shown to have a strong antioxidant activity.¹⁴⁴ Silymarin (50 and 100 μ M) increased cell viability of MSCs from human bone marrow by elevating intracellular GSH.¹⁴⁵

Combination of Purified Phytochemicals and/or Plant Extracts

One of the first study that took into account the possibility to use a combination of natural antioxidant compounds to stimulate the proliferation of stem cells was conducted by Bickford et al.¹⁴⁶ Human hematopoietic stem cells derived from bone marrow were supplemented with different concentrations of catechin, carnosine, blueberry extract, green tea extract, and vitamin D₃ alone or in combination. All the tested compounds exhibited a positive effect on cell proliferation, although the combined treatments showed a stronger effect with respect to the single treatments. In particular, the combination of blueberry extract, green tea extract, carnosine, and vitamin D₃ led to a higher proliferation rate with respect to the positive controls treated with human granulocyte colony-stimulating factor. Of all the tested compounds, blueberry extract significantly increased cell proliferation when combined with the other compounds.

Very recently, we demonstrated that a combination of epigallocatechin-3-gallate and sulforaphane exhibited a higher efficacy against oxidative stress induced by hyperoxia with respect to the single antioxidant treatments in hAFSCs.¹⁴⁷ In particular, the combined treatment was more effective in reducing endogenous ROS generation, enhancing GSH levels and increasing the endogenous antioxidant enzymes glutathione reductase, NAD(P)H:quinone oxidoreductase-1 (NQO1), and thioredoxin reductase (TR). Moreover, the combined treatment upregulated the pluripotency markers, Oct4 and Nanog, and downregulated the senescence-associated gene markers, p16 and β -gal.

Luo et al.¹⁴⁸ investigated the effect of two different combinations of antioxidants, a proprietary antioxidant supplement from Sigma-Aldrich (AOS, Catalogue Number: Sigma A1345) at 10,000-, 50,000-, and 200,000-fold dilution, and a homemade cocktail of L-ascorbate, L-glutathione (20 mM), and α -tocopherol acetate at the concentrations of 20, 4, and 1 μ M, respectively, on genomic stability of iPSCs during long-term *ex vivo* expansion. Both the antioxidant treatment did not reduce cell growth and maintained stemness up to 2 months. Furthermore, the expression of 53BP1 and ATM, two critical molecules involved in DNA damage and repair,^{149,150} were not modulated by the antioxidant

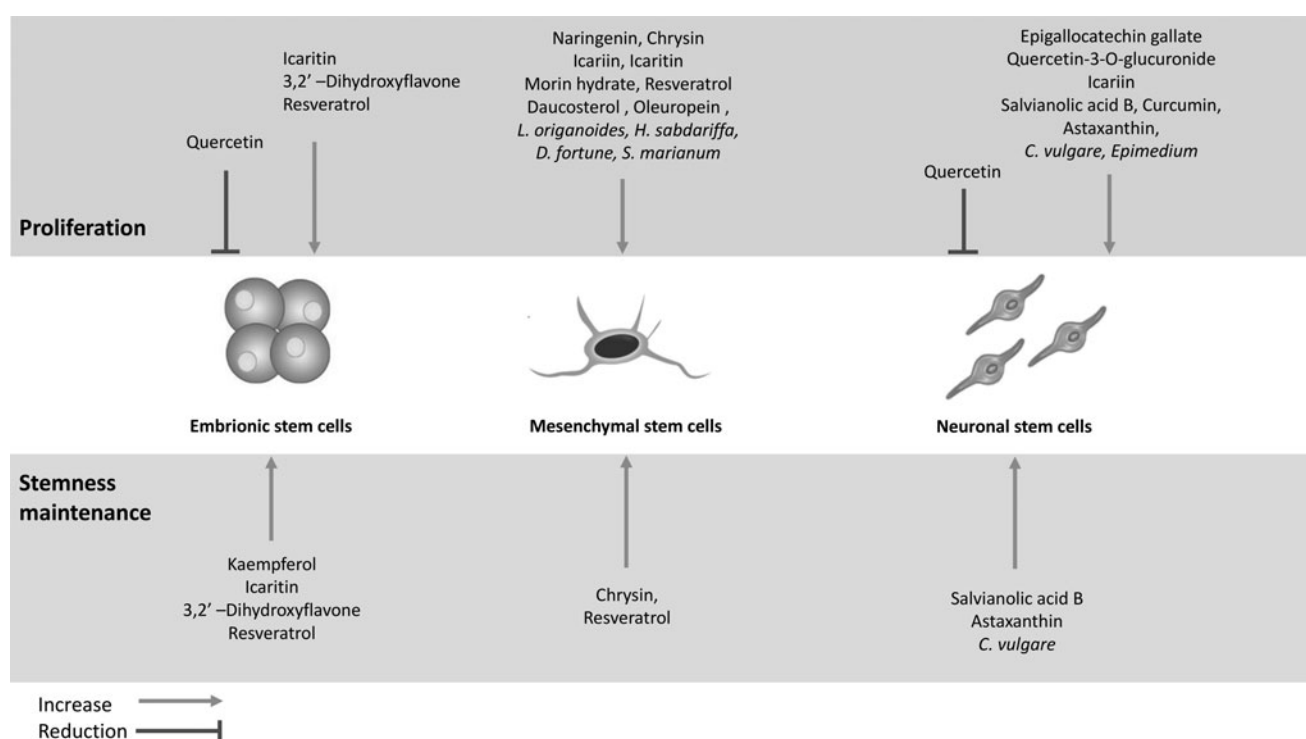


FIG. 2. Effect of phytochemicals and plant extracts on proliferation and stemness maintenance of different stem cell types.

supplementation, and array CGH analysis demonstrated a reduction of the genetic aberration events only in iPSC line supplemented with the homemade antioxidant cocktail. Of course, further experiments are needed to corroborate the benefit of antioxidants for long-term *in vitro* expansion of iPSCs.

In conclusion, the use of different phytochemicals or plant extracts demonstrated to be effective in reducing hyperoxia-induced damage *in vitro*. Of note, any compounds elicited its effect on cell proliferation or stemness maintenance targeting specific molecular targets and at different concentrations in the different cell cultures (Table 1). On these bases, the use of a combination of different phytochemicals and plant extracts seems to be the most promising approach to maximize the effect.

Conclusions

Today, the potential application of stem cells is dramatically increasing, and stem cell therapy will probably be applied in numerous chronic degenerative disease therapies in the future.

So, the maintenance of high-quality stem cells in culture without any side effect in proliferation, potency, or differentiation potential will be critical for all present and future biomedical applications. The best way to isolate, culture, and maintain stem cells from primary cultures should be under their physiological conditions, that is, low oxygen concentration and low glucose. Nevertheless, this often is not possible due to economical and/or practical reasons and therefore, the possibility to use natural antioxidants compounds represents a potentially very effective alternative. The use of natural products, and above all of natural phy-

tochemicals, opens new frontiers. Many of these compounds, generally recognized as safe by the American Food and Drug Administration, can act at different molecular and cellular levels to modulate proliferation and stemness maintenance in several stem cell types (Fig. 2). Further studies are necessary to deepen the knowledge on the wide world of natural compounds that could be used in stem cell cultures, on their mechanisms of action, and on the optimal dose and association of different phytochemicals to give an important contribution to stem cell application in regenerative medicine.

Acknowledgment

Data reported in this review were partly obtained, thanks to the support of Fondazione Cassa di Risparmio di Bologna (Italy).

Author Disclosure Statement

No competing financial interests exist.

Funding Information

References

1. Siegel G, Schäfer R, Dazzi F. The immunosuppressive properties of mesenchymal stem cells. *Transplantation* 2009;87:S45–S49.
2. Chow DC, Wenning LA, Miller WM, Papoutsakis ET. Modeling pO(2) distributions in the bone marrow hematopoietic compartment. I. Krogh's model. *Biophys J* 2001; 81:675–684.

3. Antoniou ES, Sund S, Homsy EN, Challenger LF, Rameshwar P. A theoretical simulation of hematopoietic stem cells during oxygen fluctuations: Prediction of bone marrow responses during hemorrhagic shock. *Shock* 2004; 22:415–422.
4. Rodrigues CA, Diogo MM, da Silva CL, Cabral JM. Hypoxia enhances proliferation of mouse embryonic stem cell-derived neural stem cells. *Biotechnol Bioeng* 2010; 106:260–270.
5. Dos Santos F, Andrade PZ, Boura JS, Abecasis MM, da Silva CL, Cabral JM. Ex vivo expansion of human mesenchymal stem cells: A more effective cell proliferation kinetics and metabolism under hypoxia. *J Cell Physiol* 2010; 223:27–35.
6. Hung SP, Ho JH, Shih YR, Lo T, Lee OK. Hypoxia promotes proliferation and osteogenic differentiation potentials of human mesenchymal stem cells. *J Orthop Res* 2012; 30:260–266.
7. Lavrentieva A, Majore I, Kasper C, Hass R. Effects of hypoxic culture conditions on umbilical cord-derived human mesenchymal stem cells. *Cell Commun Signal* 2010; 8:18.
8. Efimenko A, Starostina E, Kalinina N, Stolzing A. Angiogenic properties of aged adipose derived mesenchymal stem cells after hypoxic conditioning. *J Transl Med* 2011; 9:10.
9. Kim JH, Song SY, Park SG, Song SU, Xia Y, Sung JH. Primary involvement of NADPH oxidase 4 in hypoxia-induced generation of reactive oxygen species in adipose-derived stem cells. *Stem Cells Dev* 2012; 21:2212–2221.
10. Urao N, Ushio-Fukai M. Redox regulation of stem/progenitor cells and bone marrow niche. *Free Radic Biol Med* 2013; 54:26–39.
11. Estrada JC, Albo C, Benguría A, et al. Culture of human mesenchymal stem cells at low oxygen tension improves growth and genetic stability by activating glycolysis. *Cell Death Differ* 2012; 19:743–755.
12. Fehrer C, Brunauer R, Laschober G, et al. Reduced oxygen tension attenuates differentiation capacity of human mesenchymal stem cells and prolongs their lifespan. *Aging Cell* 2007; 6:745–757.
13. Mohamadnejad M, Pournasr B, Bagheri M, et al. Transplantation of allogeneic bone marrow mesenchymal stromal cell-derived hepatocyte-like cells in homozygous familial hypercholesterolemia. *Cytotherapy* 2010; 12:566–568.
14. Casciaro F, Beretti F, Zavatti M, et al. Nuclear Nox4 interaction with prelamin A is associated with nuclear redox control of stem cell aging. *Aging (Albany NY)* 2018; 10: 2911–2934.
15. Karimi-Busheri F, Rasouli-Nia A, Weinfeld M. Key issues related to cryopreservation and storage of stem cells and cancer stem cells: Protecting biological integrity. *Adv Exp Med Biol* 2016; 951:1–12.
16. Angeloni C, Leoncini E, Malaguti M, Angelini S, Hrelia P, Hrelia S. Modulation of phase II enzymes by sulforaphane: Implications for its cardioprotective potential. *J Agric Food Chem* 2009; 57:5615–5622.
17. Lu MC, Ji JA, Jiang ZY, You QD. The Keap1-Nrf2-ARE pathway as a potential preventive and therapeutic target: An update. *Med Res Rev* 2016; 36:924–963.
18. Islam MS, Nagasaka R, Ohara K, et al. Biological abilities of rice bran-derived antioxidant phytochemicals for medical therapy. *Curr Top Med Chem* 2011; 11:1847–1853.
19. Ma Y, Zeng M, Sun R, Hu M. Disposition of flavonoids impacts their efficacy and safety. *Curr Drug Metab* 2014; 15:841–864.
20. Minich DM, Bland JS. A review of the clinical efficacy and safety of cruciferous vegetable phytochemicals. *Nutr Rev* 2007; 65:259–267.
21. Shipkowski KA, Betz JM, Birnbaum LS, et al. Naturally complex: Perspectives and challenges associated with Botanical Dietary Supplement Safety assessment. *Food Chem Toxicol* 2018; 118:963–971.
22. Bertin E, Piccoli M, Franzin C, et al. First steps to define murine amniotic fluid stem cell microenvironment. *Sci Rep* 2016; 6:37080.
23. Mohyeldin A, Garzón-Muvdi T, Quiñones-Hinojosa A. Oxygen in stem cell biology: A critical component of the stem cell niche. *Cell Stem Cell* 2010; 7:150–161.
24. Bizzarri A, Koehler H, Cajlakovic M, et al. Continuous oxygen monitoring in subcutaneous adipose tissue using microdialysis. *Anal Chim Acta* 2006; 573–574:48–56.
25. Harrison JS, Rameshwar P, Chang V, Bandari P. Oxygen saturation in the bone marrow of healthy volunteers. *Blood* 2002; 99:394.
26. Spencer JA, Ferraro F, Roussakis E, et al. Direct measurement of local oxygen concentration in the bone marrow of live animals. *Nature* 2014; 508:269–273.
27. Walter D, Lier A, Geiselhart A, et al. Exit from dormancy provokes DNA-damage-induced attrition in haematopoietic stem cells. *Nature* 2015; 520:549–552.
28. Wilson A, Laurenti E, Oser G, et al. Hematopoietic stem cells reversibly switch from dormancy to self-renewal during homeostasis and repair. *Cell* 2008; 135:1118–1129.
29. Zhou D, Shao L, Spitz DR. Reactive oxygen species in normal and tumor stem cells. *Adv Cancer Res* 2014; 122: 1–67.
30. Morrison SJ, Kimble J. Asymmetric and symmetric stem-cell divisions in development and cancer. *Nature* 2006; 441:1068–1074.
31. Genbacev O, Zhou Y, Ludlow JW, Fisher SJ. Regulation of human placental development by oxygen tension. *Science* 1997; 277:1669–1672.
32. Schofield CJ, Ratcliffe PJ. Signalling hypoxia by HIF hydroxylases. *Biochem Biophys Res Commun* 2005; 338: 617–626.
33. Berra E, Benizri E, Ginouvès A, Volmat V, Roux D, Pouyssegur J. HIF prolyl-hydroxylase 2 is the key oxygen sensor setting low steady-state levels of HIF-1α in normoxia. *EMBO J* 2003; 22:4082–4090.
34. Berra E, Ginouvès A, Pouyssegur J. The hypoxia-inducible-factor hydroxylases bring fresh air into hypoxia signalling. *EMBO Rep* 2006; 7:41–45.
35. Semenza GL. Hydroxylation of HIF-1: Oxygen sensing at the molecular level. *Physiology (Bethesda)* 2004; 19:176–182.
36. Covellos KL, Kehler J, Yu H, et al. HIF-2α regulates Oct-4: Effects of hypoxia on stem cell function, embryonic development, and tumor growth. *Genes Dev* 2006; 20: 557–570.
37. Sterneckert J, Höing S, Schöler HR. Concise review: Oct4 and more: The reprogramming expressway. *Stem Cells* 2012; 30:15–21.
38. Boulais PE, Frenette PS. Making sense of hematopoietic stem cell niches. *Blood* 2015; 125:2621–2629.
39. Simsek T, Kocabas F, Zheng J, et al. The distinct metabolic profile of hematopoietic stem cells reflects their location in a hypoxic niche. *Cell Stem Cell* 2010; 7:380–390.

40. Pattappa G, Thorpe SD, Jegard NC, Heywood HK, de Bruijn JD, Lee DA. Continuous and uninterrupted oxygen tension influences the colony formation and oxidative metabolism of human mesenchymal stem cells. *Tissue Eng Part C Methods* 2013;19:68–79.
41. Kobayashi CI, Suda T. Regulation of reactive oxygen species in stem cells and cancer stem cells. *J Cell Physiol* 2012;227:421–430.
42. Shyh-Chang N, Daley GQ, Cantley LC. Stem cell metabolism in tissue development and aging. *Development* 2013;140:2535–2547.
43. Jung MS, Jin DH, Chae HD, et al. Bcl-xL and E1B-19K proteins inhibit p53-induced irreversible growth arrest and senescence by preventing reactive oxygen species-dependent p38 activation. *J Biol Chem* 2004;279:17765–17771.
44. Macip S, Igarashi M, Fang L, et al. Inhibition of p21-mediated ROS accumulation can rescue p21-induced senescence. *EMBO J* 2002;21:2180–2188.
45. Tothova Z, Kollipara R, Huntly BJ, et al. FoxOs are critical mediators of hematopoietic stem cell resistance to physiologic oxidative stress. *Cell* 2007;128:325–339.
46. Oh J, Lee YD, Wagers AJ. Stem cell aging: Mechanisms, regulators and therapeutic opportunities. *Nat Med* 2014;20:870–880.
47. Shao L, Li H, Pazhanisamy SK, Meng A, Wang Y, Zhou D. Reactive oxygen species and hematopoietic stem cell senescence. *Int J Hematol* 2011;94:24–32.
48. Ong ALC, Ramasamy TS. Role of Sirtuin1-p53 regulatory axis in aging, cancer and cellular reprogramming. *Ageing Res Rev* 2018;43:64–80.
49. Brown K, Xie S, Qiu X, et al. SIRT3 reverses aging-associated degeneration. *Cell Rep* 2013;3:319–327.
50. Kumar S, Geiger H. HSC niche biology and HSC expansion ex vivo. *Trends Mol Med* 2017;23:799–819.
51. Cruz FF, Rocco PR. Hypoxic preconditioning enhances mesenchymal stromal cell lung repair capacity. *Stem Cell Res Ther* 2015;6:130.
52. Lan YW, Choo KB, Chen CM, et al. Hypoxia-preconditioned mesenchymal stem cells attenuate bleomycin-induced pulmonary fibrosis. *Stem Cell Res Ther* 2015;6:97.
53. Wakai T, Narasimhan P, Sakata H, et al. Hypoxic preconditioning enhances neural stem cell transplantation therapy after intracerebral hemorrhage in mice. *J Cereb Blood Flow Metab* 2016;36:2134–2145.
54. Basciano L, Nemos C, Foliguet B, et al. Long term culture of mesenchymal stem cells in hypoxia promotes a genetic program maintaining their undifferentiated and multipotent status. *BMC Cell Biol* 2011;12:12.
55. Grayson WL, Zhao F, Bunnell B, Ma T. Hypoxia enhances proliferation and tissue formation of human mesenchymal stem cells. *Biochem Biophys Res Commun* 2007;358:948–953.
56. Li TS, Marbán E. Physiological levels of reactive oxygen species are required to maintain genomic stability in stem cells. *Stem Cells* 2010;28:1178–1185.
57. Forsyth NR, Musio A, Vezzoni P, Simpson AH, Noble BS, McWhir J. Physiologic oxygen enhances human embryonic stem cell clonal recovery and reduces chromosomal abnormalities. *Cloning Stem Cells* 2006;8:16–23.
58. Kim DS, Ko YJ, Lee MW, et al. Effect of low oxygen tension on the biological characteristics of human bone marrow mesenchymal stem cells. *Cell Stress Chaperones* 2016;21:1089–1099.
59. Ejtehadifar M, Shamsasenjan K, Movassaghpour A, et al. The effect of hypoxia on mesenchymal stem cell biology. *Adv Pharm Bull* 2015;5:141–149.
60. Pezzi A, Amarin B, Laureano Á, et al. Effects of hypoxia in long-term in vitro expansion of human bone marrow derived mesenchymal stem cells. *J Cell Biochem* 2017;118:3072–3079.
61. Holzwarth C, Vaegler M, Gieseke F, et al. Low physiologic oxygen tensions reduce proliferation and differentiation of human multipotent mesenchymal stromal cells. *BMC Cell Biol* 2010;11:11.
62. Salim A, Nacamuli RP, Morgan EF, Giaccia AJ, Longaker MT. Transient changes in oxygen tension inhibit osteogenic differentiation and Runx2 expression in osteoblasts. *J Biol Chem* 2004;279:40007–40016.
63. Lennon DP, Edmison JM, Caplan AI. Cultivation of rat marrow-derived mesenchymal stem cells in reduced oxygen tension: Effects on in vitro and in vivo osteochondrogenesis. *J Cell Physiol* 2001;187:345–355.
64. Malladi P, Xu Y, Chiou M, Giaccia AJ, Longaker MT. Effect of reduced oxygen tension on chondrogenesis and osteogenesis in adipose-derived mesenchymal cells. *Am J Physiol Cell Physiol* 2006;290:C1139–C1146.
65. Choudhery MS, Badowski M, Muise A, Pierce J, Harris DT. Donor age negatively impacts adipose tissue-derived mesenchymal stem cell expansion and differentiation. *J Transl Med* 2014;12:8.
66. El Alami M, Viña-Almunia J, Gambini J, et al. Activation of p38, p21, and NRF-2 mediates decreased proliferation of human dental pulp stem cells cultured under 21% O₂. *Stem Cell Rep* 2014;3:566–573.
67. Brown GC, Borutaite V. There is no evidence that mitochondria are the main source of reactive oxygen species in mammalian cells. *Mitochondrion* 2012;12:1–4.
68. Dröge W. Free radicals in the physiological control of cell function. *Physiol Rev* 2002;82:47–95.
69. Case J, Ingram DA, Haneline LS. Oxidative stress impairs endothelial progenitor cell function. *Antioxid Redox Signal* 2008;10:1895–1907.
70. Ito K, Hirao A, Arai F, et al. Reactive oxygen species act through p38 MAPK to limit the lifespan of hematopoietic stem cells. *Nat Med* 2006;12:446–451.
71. Ushio-Fukai M, Rehman J. Redox and metabolic regulation of stem/progenitor cells and their niche. *Antioxid Redox Signal* 2014;21:1587–1590.
72. Naka K, Muraguchi T, Hoshii T, Hirao A. Regulation of reactive oxygen species and genomic stability in hematopoietic stem cells. *Antioxid Redox Signal* 2008;10:1883–1894.
73. Jang YY, Sharkis SJ. A low level of reactive oxygen species selects for primitive hematopoietic stem cells that may reside in the low-oxygenic niche. *Blood* 2007;110:3056–3063.
74. Starkov AA. The role of mitochondria in reactive oxygen species metabolism and signaling. *Ann N Y Acad Sci* 2008;1147:37–52.
75. Nordzike DE, Medraño-Fernandez I. The plasma membrane: A platform for intra- and intercellular redox signaling. *Antioxidants (Basel)* 2018;7.
76. Jezek P, Hlavatá L. Mitochondria in homeostasis of reactive oxygen species in cell, tissues, and organism. *Int J Biochem Cell Biol* 2005;37:2478–2503.

77. Del Río LA, López-Huertas E. ROS Generation in peroxisomes and its role in cell signaling. *Plant Cell Physiol* 2016;57:1364–1376.
78. Ivanov I, Heydeck D, Hofheinz K, et al. Molecular enzymology of lipoxygenases. *Arch Biochem Biophys* 2010; 503:161–174.
79. Ben-Mahdi MH, Dang PM, Gougerot-Pocidalo MA, O'Dowd Y, El-Benna J, Pasquier C. Xanthine oxidase-derived ros display a biphasic effect on endothelial cells adhesion and FAK phosphorylation. *Oxid Med Cell Longev* 2016;2016:9346242.
80. Piccoli C, Ria R, Scrima R, et al. Characterization of mitochondrial and extra-mitochondrial oxygen consuming reactions in human hematopoietic stem cells. Novel evidence of the occurrence of NAD(P)H oxidase activity. *J Biol Chem* 2005;280:26467–26476.
81. Drehmer DL, de Aguiar AM, Brandt AP, et al. Metabolic switches during the first steps of adipogenic stem cells differentiation. *Stem Cell Res* 2016;17:413–421.
82. Piccoli C, D'Aprile A, Scrima R, et al. Role of reactive oxygen species as signal molecules in the pre-commitment phase of adult stem cells. *Ital J Biochem* 2007;56:7.
83. Wang Y, Liu L, Pazhanisamy SK, Li H, Meng A, Zhou D. Total body irradiation causes residual bone marrow injury by induction of persistent oxidative stress in murine hematopoietic stem cells. *Free Radic Biol Med* 2010;48: 348–356.
84. Yamaguchi M, Kashiwakura I. Role of reactive oxygen species in the radiation response of human hematopoietic stem/progenitor cells. *PLoS One* 2013;8:e70503.
85. Liu J, Cao L, Chen J, et al. Bmi1 regulates mitochondrial function and the DNA damage response pathway. *Nature* 2009;459:387–392.
86. Chen C, Liu Y, Liu R, Ikenoue T, Guan KL, Zheng P. TSC-mTOR maintains quiescence and function of hematopoietic stem cells by repressing mitochondrial biogenesis and reactive oxygen species. *J Exp Med* 2008;205: 2397–2408.
87. Wong JL, Wessel GM. Reactive oxygen species and Udx1 during early sea urchin development. *Dev Biol* 2005;288: 317–333.
88. Schmelter M, Ateghang B, Helmig S, Wartenberg M, Sauer H. Embryonic stem cells utilize reactive oxygen species as transducers of mechanical strain-induced cardiovascular differentiation. *FASEB J* 2006;20:1182–1184.
89. Cho YM, Kwon S, Pak YK, et al. Dynamic changes in mitochondrial biogenesis and antioxidant enzymes during the spontaneous differentiation of human embryonic stem cells. *Biochem Biophys Res Commun* 2006;348:1472–1478.
90. Sheshadri P, Kumar A. Managing odds in stem cells: Insights into the role of mitochondrial antioxidant enzyme MnSOD. *Free Radic Res* 2016;50:570–584.
91. Klein D, Steens J, Wiesemann A, et al. Mesenchymal stem cell therapy protects lungs from radiation-induced endothelial cell loss by restoring superoxide dismutase 1 expression. *Antioxid Redox Signal* 2017;26:563–582.
92. Sundaramoorthy P, Wang Q, Zheng Z, et al. Thioredoxin mitigates radiation-induced hematopoietic stem cell injury in mice. *Stem Cell Res Ther* 2017;8:263.
93. Olivares-Vicente M, Barrajon-Catalan E, Herranz-Lopez M, et al. Plant-derived polyphenols in human health: Biological activity, metabolites and putative molecular targets. *Curr Drug Metab* 2018;19:351–369.
94. Wang Y, Li M, Xu X, Song M, Tao H, Bai Y. Green tea epigallocatechin-3-gallate (EGCG) promotes neural progenitor cell proliferation and sonic hedgehog pathway activation during adult hippocampal neurogenesis. *Mol Nutr Food Res* 2012;56:1292–1303.
95. Zhang Y, He Q, Dong J, Jia Z, Hao F, Shan C. Effects of epigallocatechin-3-gallate on proliferation and differentiation of mouse cochlear neural stem cells: Involvement of PI3K/Akt signaling pathway. *Eur J Pharm Sci* 2016;88: 267–273.
96. Angeloni C, Hrelia S. Quercetin reduces inflammatory responses in LPS-stimulated cardiomyoblasts. *Oxid Med Cell Longev* 2012;2012:837104.
97. Baral S, Pariyar R, Kim J, Lee HS, Seo J. Quercetin-3-O-glucuronide promotes the proliferation and migration of neural stem cells. *Neurobiol Aging* 2017;52:39–52.
98. Kim SY, Jeong HC, Hong SK, Lee MO, Cho SJ, Cha HJ. Quercetin induced ROS production triggers mitochondrial cell death of human embryonic stem cells. *Oncotarget* 2017;8:64964–64973.
99. Nichols M, Zhang J, Polster BM, et al. Synergistic neuroprotection by epicatechin and quercetin: Activation of convergent mitochondrial signaling pathways. *Neuroscience* 2015;308:75–94.
100. Correia M, Rodrigues AS, Perestrelo T, et al. Different concentrations of kaempferol distinctly modulate murine embryonic stem cell function. *Food Chem Toxicol* 2016; 87:148–156.
101. Lee CH, Jeong TS, Choi YK, et al. Anti-atherogenic effect of citrus flavonoids, naringin and naringenin, associated with hepatic ACAT and aortic VCAM-1 and MCP-1 in high cholesterol-fed rabbits. *Biochem Biophys Res Commun* 2001;284:681–688.
102. Zhang P, Dai KR, Yan SG, et al. Effects of naringin on the proliferation and osteogenic differentiation of human bone mesenchymal stem cell. *Eur J Pharmacol* 2009;607:1–5.
103. Liu M, Li Y, Yang ST. Effects of naringin on the proliferation and osteogenic differentiation of human amniotic fluid-derived stem cells. *J Tissue Eng Regen Med* 2017; 11:276–284.
104. Zhong S, Ge J, Yu JY. Icariin prevents cytokine-induced β -cell death by inhibiting NF- κ B signaling. *Exp Ther Med* 2018;16:2756–2762.
105. Fan JJ, Cao LG, Wu T, et al. The dose-effect of icariin on the proliferation and osteogenic differentiation of human bone mesenchymal stem cells. *Molecules* 2011;16:10123–10133.
106. Huang JH, Cai WJ, Zhang XM, Shen ZY. Icariin promotes self-renewal of neural stem cells: An involvement of extracellular regulated kinase signaling pathway. *Chin J Integr Med* 2014;20:107–115.
107. Yang P, Guan YQ, Li YL, Zhang L, Li L. Icariin promotes cell proliferation and regulates gene expression in human neural stem cells in vitro. *Mol Med Rep* 2016;14:1316–1322.
108. Fu X, Li S, Zhou S, Wu Q, Jin F, Shi J. Stimulatory effect of icariin on the proliferation of neural stem cells from rat hippocampus. *BMC Complement Altern Med* 2018;18:34.
109. Qin S, Zhou W, Liu S, Chen P, Wu H. Icariin stimulates the proliferation of rat bone mesenchymal stem cells via ERK and p38 MAPK signaling. *Int J Clin Exp Med* 2015; 8:7125–7133.

110. Ye Y, Jing X, Li N, Wu Y, Li B, Xu T. Icaritin promotes proliferation and osteogenic differentiation of rat adipose-derived stem cells by activating the RhoA-TAZ signaling pathway. *Biomed Pharmacother* 2017;88:384–394.
111. Chen S, Deng X, Ma K, et al. Icaritin improves the viability and function of cryopreserved human nucleus pulposus-derived mesenchymal stem cells. *Oxid Med Cell Longev* 2018;2018:3459612.
112. Tsang WP, Zhang F, He Q, et al. Icaritin enhances mESC self-renewal through upregulating core pluripotency transcription factors mediated by ER α . *Sci Rep* 2017;7:40894.
113. Lim RZL, Li L, Yong EL, Chew N. STAT-3 regulation of CXCR4 is necessary for the prenylflavonoid Icaritin to enhance mesenchymal stem cell proliferation, migration and osteogenic differentiation. *Biochim Biophys Acta Gen Subj* 2018;1862:1680–1692.
114. Deldar Y, Zarghami F, Pilehvar-Soltanahmadi Y, Dashedpour M, Zarghami N. Antioxidant effects of chrysin-loaded electrospun nanofibrous mats on proliferation and stemness preservation of human adipose-derived stem cells. *Cell Tissue Bank* 2017;18:475–487.
115. Menon AH, Soundarya SP, Sanjay V, Chandran SV, Balagangadharan K, Selvamurugan N. Sustained release of chrysin from chitosan-based scaffolds promotes mesenchymal stem cell proliferation and osteoblast differentiation. *Carbohydr Polym* 2018;195:356–367.
116. Han D, Kim HJ, Choi HY, et al. 3,2'-Dihydroxyflavone-treated pluripotent stem cells show enhanced proliferation, pluripotency marker expression, and neuroprotective properties. *Cell Transplant* 2015;24:1511–1532.
117. Kuzu M, Kandemir FM, Yildirim S, Kucukler S, Caglayan C, Turk E. Morin attenuates doxorubicin-induced heart and brain damage by reducing oxidative stress, inflammation and apoptosis. *Biomed Pharmacother* 2018;106:443–453.
118. He Q, Jia Z, Zhang Y, Ren X. Morin hydrate promotes inner ear neural stem cell survival and differentiation and protects cochlea against neuronal hearing loss. *J Cell Mol Med* 2017;21:600–608.
119. Burns J, Yokota T, Ashihara H, Lean ME, Crozier A. Plant foods and herbal sources of resveratrol. *J Agric Food Chem* 2002;50:3337–3340.
120. Song LH, Pan W, Yu YH, Quarles LD, Zhou HH, Xiao ZS. Resveratrol prevents CsA inhibition of proliferation and osteoblastic differentiation of mouse bone marrow-derived mesenchymal stem cells through an ER/NO/cGMP pathway. *Toxicol In Vitro* 2006;20:915–922.
121. Dai Z, Li Y, Quarles LD, et al. Resveratrol enhances proliferation and osteoblastic differentiation in human mesenchymal stem cells via ER-dependent ERK1/2 activation. *Phytomedicine* 2007;14:806–814.
122. Peltz L, Gomez J, Marquez M, et al. Resveratrol exerts dosage and duration dependent effect on human mesenchymal stem cell development. *PLoS One* 2012;7:e37162.
123. Li N, Du Z, Shen Q, et al. Resveratrol enhances self-renewal of mouse embryonic stem cells. *J Cell Biochem* 2017;118:1928–1935.
124. Safaiejad Z, Nabuini M, Peymani M, Ghaedi K, Nasr-Esfahani MH, Baharvand H. Resveratrol promotes human embryonic stem cells self-renewal by targeting SIRT1-ERK signaling pathway. *Eur J Cell Biol* 2017;96:665–672.
125. Jiang LH, Yang NY, Yuan XL, et al. Daucosterol promotes the proliferation of neural stem cells. *J Steroid Biochem Mol Biol* 2014;140:90–99.
126. Kim R, Lee S, Lee CY, et al. *Salvia miltiorrhiza* enhances the survival of mesenchymal stem cells under ischemic conditions. *J Pharm Pharmacol* 2018;70:1228–1241.
127. Zhuang P, Zhang Y, Cui G, et al. Direct stimulation of adult neural stem/progenitor cells in vitro and neurogenesis in vivo by salvianolic acid B. *PLoS One* 2012;7:e35636.
128. Angeloni C, Malaguti M, Barbalace MC, Hrelia S. Bioactivity of olive oil phenols in neuroprotection. *Int J Mol Sci* 2017;18.
129. Ji ST, Kim YJ, Jung SY, et al. Oleuropein attenuates hydrogen peroxide-induced autophagic cell death in human adipose-derived stem cells. *Biochem Biophys Res Commun* 2018;499:675–680.
130. Typek R, Dawidowicz AL, Wianowska D, Bernacik K, Stankevič M, Gil M. Formation of aqueous and alcoholic adducts of curcumin during its extraction. *Food Chem* 2019;276:101–109.
131. Kim SJ, Son TG, Park HR, et al. Curcumin stimulates proliferation of embryonic neural progenitor cells and neurogenesis in the adult hippocampus. *J Biol Chem* 2008;283:14497–14505.
132. Galasso C, Orefice I, Pellone P, et al. On the neuroprotective role of astaxanthin: New perspectives? *Mar Drugs* 2018;16.
133. Kidd P. Astaxanthin, cell membrane nutrient with diverse clinical benefits and anti-aging potential. *Altern Med Rev* 2011;16:355–364.
134. Kim JH, Nam SW, Kim BW, et al. Astaxanthin improves stem cell potency via an increase in the proliferation of neural progenitor cells. *Int J Mol Sci* 2010;11:5109–5119.
135. Fontenelle RO, Morais SM, Brito EH, et al. Chemical composition, toxicological aspects and antifungal activity of essential oil from *Lippia sidoides* Cham. *J Antimicrob Chemother* 2007;59:934–940.
136. Brito FN, Vendramin FS, Lopes CTA, et al. Proliferation of human adipose tissue-derived stem cells stimulated by oil rich in thymol of *Lippia organoides*. *Acta Cir Bras* 2018;33:431–438.
137. Nalewajko-Sieliwoniuk E, Malejko J, Twarowska P, Timoszuk M, Nazaruk J. Postcolumn determination of polyphenolic antioxidants in *Cirsium vulgare* (Savi) Ten. extracts. *J Sep Sci* 2017;40:3830–3838.
138. Abdanipour A, Noori-Zadeh A, Mesbah-Namin SA, Bakhtiyari S, Nejatbakhsh R, Anarkooli IJ. Di-(2-ethylhexyl) phthalate-induced hippocampus-derived neural stem cells proliferation. *Cell J* 2017;19:166–172.
139. Hirunpanich V, Utaipat A, Morales NP, et al. Hypocholesterolemic and antioxidant effects of aqueous extracts from the dried calyx of *Hibiscus sabdariffa* L. in hypercholesterolemic rats. *J Ethnopharmacol* 2006;103:252–260.
140. Micucci M, Malaguti M, Toschi TG, et al. Cardiac and vascular synergic protective effect of *Olea europea* L. *Leaves* and *Hibiscus sabdariffa* L. *Flower Extracts*. *Oxid Med Cell Longev* 2015;2015:318125.
141. Abdul Hamid Z, Lin Lin WH, Abdalla BJ, et al. The role of *Hibiscus sabdariffa* L. (Roselle) in maintenance of ex vivo murine bone marrow-derived hematopoietic stem cells. *ScientificWorldJournal* 2014;2014:258192.

142. Huang XF, Yuan SJ, Yang C. Effects of total flavonoids from *Drynaria fortunei* on the proliferation and osteogenic differentiation of rat dental pulp stem cells. *Mol Med Rep* 2012;6:547–552.
143. Yao R, Zhang L, Li X, Li L. Effects of Epimedium flavonoids on proliferation and differentiation of neural stem cells in vitro. *Neurol Res* 2010;32:736–742.
144. Zaidi SNF, Mahboob T. Prevention of liver cirrhosis by Silymarin. *Pak J Pharm Sci* 2017;30:1203–1211.
145. Ahmadi-Ashtiani H, Allameh A, Rastegar H, Soleimani M, Barkhordari E. Inhibition of cyclooxygenase-2 and inducible nitric oxide synthase by silymarin in proliferating mesenchymal stem cells: Comparison with glutathione modifiers. *J Nat Med* 2012;66:85–94.
146. Bickford PC, Tan J, Shytle RD, Sanberg CD, El-Badri N, Sanberg PR. Nutraceuticals synergistically promote proliferation of human stem cells. *Stem Cells Dev* 2006;15:118–123.
147. Marrazzo P, Angeloni C, Freschi M, et al. Combination of epigallocatechin gallate and sulforaphane counteracts in vitro oxidative stress and delays stemness loss of amniotic fluid stem cells. *Oxid Med Cell Longev* 2018: 5263985.
148. Luo L, Kawakatsu M, Guo CW, et al. Effects of antioxidants on the quality and genomic stability of induced pluripotent stem cells. *Sci Rep* 2014;4:3779.
149. Noon AT, Goodarzi AA. 53BP1-mediated DNA double strand break repair: Insert bad pun here. *DNA Repair (Amst)* 2011;10:1071–1076.
150. Kitagawa R, Kastan MB. The ATM-dependent DNA damage signaling pathway. *Cold Spring Harb Symp Quant Biol* 2005;70:99–109.

Address correspondence to:
 Cristina Angeloni
 School of Pharmacy
 University of Camerino
 Via Gentile III da Varano
 Camerino 62032
 Italy

E-mail: cristina.angeloni@unicam.it