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**Biodegradation of polyvinyl chloride plastic films by enriched anaerobic
marine consortia**

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Abstract

Plastics remarkably contribute to marine litter, which is raising serious concerns. Currently, little is known about the fate of most plastics entering the marine environment and their potential biodegradation rate and extent under anoxic conditions.

In this work, biodegradation of polyvinyl chloride (PVC) films by consortia enriched from marine samples (litter and water) was evaluated in anaerobic microcosms. After 7 months, three microcosms showed dense biofilms on plastic surfaces, gravimetric weight losses up to $11.7 \pm 0.6\%$, marked decreases in thermal stability and average molecular weight of the polymer, suggesting microbial attack towards polymer chains. After 24 months, further three consortia showed the same abilities. Microbial communities analyzed at month 24 included taxa closely related to those previously reported as halogenated organic compounds degraders. The study is the first report on PVC biodegradation by marine anaerobic microbes and provides insights on potential biodegradation of the plastic film introduced into the sea by native microbes.

Keywords: polyvinyl chloride, plastics colonization, anaerobic marine environment, biodegradation, microbial consortia, microcosms, community composition, dehalogenation.

1. Introduction

In 2018, global plastic production was 359 million tons, 61.8 of which were produced in Europe (PlasticsEurope, 2019). Over one third of plastics is used to produce disposable products, such as packaging, disposable bags, etc., with a lifespan of 3 years (Gewert *et al.*, 2015; PlasticsEurope, 2019; Shah *et al.*, 2008; Wang *et al.*, 2016). From 2006, the amount of plastics wastes that is disposed of in landfill decreased of about 44%, and in 2016 for the first time, recycling overcome landfilling in the EU countries (PlasticsEurope, 2019). Plastic wastes are also widespread in the marine environment where they were found to be the largest debris contaminating marine habitats, with an estimation of more than 250,000 tons (Avio *et al.*, 2017; Oberbeckmann *et al.*, 2016; Wang *et al.*, 2016; Zettler *et al.*, 2013); and between 1.000 and 3.000 tons of floating plastics have been reported in Mediterranean Sea (Avio *et al.*, 2017). This plastic contamination is generated by direct plastic (macro and micro) transfer into aquatic systems or derived from physical/mechanical degradation of macroplastics in landfill that produces microplastics, which are transferred through leachate to rivers and finally to seas and oceans (Avio *et al.*, 2017; Galloway *et al.*, 2017). Further, additives present in plastics formulations are normally not covalently bonded to the polymer chains and could therefore leach out from plastics and enter the marine environment (Avio *et al.*, 2017; Gewert *et al.*, 2015; Harrison *et al.*, 2014). Plastics entering the marine environment could also absorb persistent organic pollutants (POPs) due to the hydrophobic characteristics of these compounds or be a vector agent for the spreading of harmful organisms inhabiting plastic surfaces (Avio *et al.*, 2017; Harrison *et al.*, 2014; Oberbeckmann *et al.*, 2016; Wang *et al.*, 2016; Zettler *et al.*, 2013).

Among petroleum-based plastics, the PVC is the third one in terms of European plastic demand (5 million tons in 2018) (PlasticsEurope, 2019). PVC plastics have a wide range of applications, such as food packaging, electronics, coatings, medical devices etc., due to low cost, long term stability and mechanical properties depending on quantity and quality of plasticizers added (Bueno-

69 Ferrer *et al.*, 2010; Glas *et al.*, 2014; Reddy *et al.*, 2010). The environmental concern about PVC
70 plastics is due to their high contents of chlorine and additives required for their processing.
71 Additives and stabilizers, e.g. heavy metals or phthalates, provide the necessary physical/chemical
72 and stability properties to the final products but have hazardous characteristics against humans
73 (Glas *et al.*, 2014). Chlorine atoms could produce environmentally harmful chlorinated compounds,
74 HCl or chlorinated dioxins, when disposed of in landfills or incinerated for energy recovery (Glas *et*
75 *al.*, 2014; Reddy *et al.*, 2010).

76 In the last years, there has been an increasing interest in the evaluation of biodegradation of
77 petroleum-based plastics (Ali *et al.*, 2014; Shah *et al.*, 2014; Yang *et al.*, 2015; Tribedi and Dey,
78 2017; Peixoto *et al.*, 2017; Giacomucci *et al.*, 2019; Raddadi and Fava, 2019) with the perspective
79 to develop innovative strategies for mitigating their environmental impact. Considering their
80 hydrophobic nature and the very low bioavailability, the formation of biofilm on the polymer/plastic
81 surface is a critical step necessary for the biodegradation (Das *et al.*, 2012; Reisser *et al.*, 2014). To
82 date, few PVC polymer and plastic biodegradation studies have been reported and were performed
83 mainly under aerobic conditions and in terrestrial systems (Ali *et al.*, 2014; Anwar *et al.*, 2016;
84 Giacomucci *et al.*, 2019; Shah *et al.* 2008; Webb *et al.*, 2000). Marine anaerobic microorganisms
85 have not been studied yet in depth as plastic biodegraders even if they should be the first actors
86 involved in mitigating the marine litter effects associated with the plastics already in the marine
87 ecosystems.

88 The Mediterranean Sea has been identified as a plastic accumulating ecosystem, especially for
89 floating polymeric items (Ruiz-Orejón *et al.*, 2017; Suaria and Aliani, 2014). Oxygen availability in
90 marine water is decreasing due to climate change with the result of modifying by the expansion of
91 low-oxygen zones, especially in coastal systems (Breitburg *et al.* 2018; Dang and Lovell, 2016).
92 Although most waters in the ocean are oxic and contain aerobic microbes, biofilms on submerged
93 surfaces and marine particles in oxic waters may additionally harbor hypoxic and anoxic
94 microenvironments suitable for microbial anaerobic metabolisms. Hence, anaerobic microbes living

in sessile form could be found even in aerobic condition as biofilm structure preserves them from oxygen present in the surrounding environment and thus some bacteria associated to marine fouling need suboxic or anoxic microniches within particles to support microaerophilic or anaerobic metabolism (Celikkol-Aydin *et al.*, 2016; Dang and Lovell, 2016).

The aim of the present study was to evaluate the biodegradation of a virgin PVC plastic film under marine anaerobic conditions. For this purpose, 16 anaerobic consortia were enriched from marine samples (water with litter fraction) in the presence of PVC films in laboratory microcosms and their biodegradation abilities were assessed after 7 and 24 months incubation. At the end of the experiment, the PVC film-associated microbial communities were characterized.

2. Materials and Methods

2.1. Sampling and pre-enrichment of marine anaerobic consortia

Marine samples were collected from four sampling stations (S1 to S4) in Elefsis Bay (Greece) at a depth of 0 to 1.2 m of the water column. The samples, named S1 to S4 and consisting of marine water and a litter/debris fraction, were used as source of microbial communities. To increase the anaerobic microbial biomass to be used as inoculum in biodegradation assays, a pre enrichment step of the microbes adhered to the collected marine litter was performed. Specifically, four anaerobic microcosms from each sampling station were set up, using four different growth media, in 100 ml serum bottles with working volume of 40 ml. For this purpose 4 ml of litter fraction, 16 ml of marine water and further 20 ml of the anaerobic specific medium containing 24 g/L of sodium chloride were mixed. The four culture media (M1, M2, M3 and M4; composition reported in Supplementary material S1) were used in order to try to favour the growth of different anaerobic consortia, i.e. acidogenic/methanogenic (M1 and M2), nitrate reducing (M3) and sulfate reducing (M4) microorganisms. The sixteen different microcosms were flushed with pure sterile nitrogen (in the case of M1, M3 and M4 media) or a mix of nitrogen and carbon dioxide (N₂:CO₂=80:20, for the M2 medium) and incubated at 20°C. Microbial growth was evaluated after 3 months incubation

120 based on gas composition analysis of the headspace of each microcosm (H₂, CO₂, CH₄) by gas
121 chromatography (Agilent 3000 MicroGC, Agilent technologies, Santa Clara, CA, USA) equipped
122 with thermal conductivity detector (TCD). The analytical conditions were as follows: injector
123 temperature 90 °C; column temperature 60 °C; sampling time 20 s; injection time 50 ms; column
124 pressure 25 psi; run time 44 s; carrier gas N₂. When microbial growth was detected, fresh medium
125 was added to a final volume of 70 ml and microcosms were incubated under the same conditions
126 until a significant growth was detected, before using as inocula in the biodegradation experiment.

127 A set of four negative controls with the corresponding non inoculated media were also prepared.

128

129 **2.2. PVC film**

130 PVC film containing a total of 30%_{w/w} of additives was provided by an Italian plastic producing
131 company. In order to better monitor the gravimetric weight loss, thick PVC plastic films (300 µm),
132 obtained after pressing several PVC thin layers (20 µm each) at 17.44 bar and 160°C for 4-5 min,
133 were used. Thick films were then cut into small pieces (1.5 cm × 1.5 cm and 1.5 cm × 3.2 cm),
134 degreased and sterilized by 30 min immersion in 70%_{v/v} ethanol solution and washed with sterile
135 distilled water before use. The films used for the determination of gravimetric weight loss were
136 dried under vacuum to constant weight before being used as substrate for microbial growth.

137

138 **2.3. PVC biodegradation assays**

139 The sixteen microbial communities from the pre-enrichment step were used as inocula for PVC
140 biodegradation experiments (Figure S1). Specifically, sixteen microcosms were setup in 100 ml
141 serum bottles with 50 ml of the corresponding medium containing 10 g/l of PVC films as major
142 carbon and energy source and inoculated with 20%_{v/v} of pre-enriched cultures. Inoculated
143 microcosms and abiotic controls were incubated under static conditions at 20°C. Samples of plastic

144 films and culture broth were withdrawn after 7 and 24 months incubation and subjected to the
145 following analyses in order to detect putative microbial growth and PVC films biodegradation.

146 2.3.1. *Evaluation of the microbial growth and colonization of the PVC film surface*

147 Viability and growth of anaerobic consortia were evaluated by measuring headspace gas
148 production and composition (H₂, CO₂, CH₄) using MicroGC-TCD.

149 Biofilm formation was assessed by quantifying the proteins available onto the PVC film
150 surfaces. For this purpose, sampled PVC films were incubated overnight (4°C, 150 rpm) in a 6 M
151 urea solution. Protein quantification according to Lowry method (Lowry *et al.*, 1951) was then
152 performed on urea solutions, and urea 6 M was used as blank.

153 2.3.2. *Evaluation of PVC films biodegradation*

154 *Gravimetric weight loss.* Measurement of the dry weight of PVC film was determined after biofilm
155 removal by incubation with a 6 M urea solution overnight at 4°C and further washing with distilled
156 water. The washed PVC films were dried under vacuum at room temperature to constant weight (at
157 least 24 h) before weighing using a balance with a 6 digit accuracy. The weight loss percentage was
158 calculated as follows:

$$159 \text{ Weight loss (\%)} = (\text{Initial weight} - \text{Final weight}) / \text{Initial weight} * 100.$$

160 *Thermogravimetric analysis (TGA).* Dried plastic samples of 5 to 10 mg were subjected to TGA
161 using a Perkin Elmer TGA7 thermal analyzer under nitrogen atmosphere (gas flow: 40 ml/min).
162 The thermograms were recorded from 40°C to 800°C at a heating rate of 10°C/min. The onset
163 degradation temperature (T_{onset}) and the maximum degradation temperature (T_{max}) were noted. Non
164 incubated PVC film and PVC resin were used as reference.

165 *Gel permeation chromatography (GPC).* The molecular weight of the PVC films were determined
166 using GPC. Measurements were carried out on a HPLC Lab Flow 2000 equipped with Phenomenex
167 Phenogel Mixed 5µ MXM/MXL columns and a Linear Instrument UVIS-200 detector operating at
168 254 nm. Tetrahydrofuran was used as mobile phase (1 ml/min) after calibration with polystyrene

standards of known molecular mass. A sample concentration of 3 mg/ml was applied. Non incubated PVC film and PVC films incubated under abiotic conditions were used as references.

2.3.3. Statistical analyses

Results of gravimetric weight losses and GPC analyses were statistically evaluated using one-way ANOVA. Post-hoc Tukey test was applied to determine whether gravimetric or molecular weight changes of PVC films showed significantly different extents. Statistically significant results were depicted by *p-values* < 0.05. All the analyses were performed using GraphPad Prism version 8.0.0 for Windows (GraphPad Software, San Diego, California USA, www.graphpad.com).

2.4. Composition of planktonic and biofilm communities

After 24 months incubation, total DNA was extracted using a Powersoil[®] DNA isolation kit (MOBIO, Carlsbad, CA, USA) from planktonic (from 2 ml of culture broth) and biofilm (PVC adhered microbial cells) microbial communities. Bacterial partial 16S rRNA gene was amplified using 357F-GC and 907R primers and PCR conditions reported by Polo *et al.* (2010) except for MgCl₂ and dNTP mix concentration of 1.8-2 mM and 0.2 μM, respectively and annealing temperature of 55°C. Archaeal PCRs were performed using Arch344F-GC and Arch918R (Giovannoni *et al.*, 1988; Stahl and Amann, 1991) primer pair. The archaeal thermal protocol was a modification of the bacterial protocol by elongating each step of 15 sec. DGGE was according to Polo *et al.* (2010), using denaturant gradients of 40–60% and 30–70% for bacterial and archaeal communities, respectively. Principal bands were excised, eluted in 50 μl milliQ water (30°C for 3 h), amplified and identified by sequencing (Macrogen Inc., Republic of Korea). The sequences were checked for chimeras using DECIPHER software (Wright *et al.*, 2012), analyzed using the BLASTN software and submitted to the Genbank database (accession numbers MH520751 to MH520766).

3. Results and discussion

3.1. PVC films biodegradability after 7 months

3.1.1. Microbial growth monitoring

Before performing the biodegradation assays, a pre-enrichment step (growth in the absence of PVC film) was performed (please refer to the scheme of the experimental design in supplementary Figure S1) and microbial growth was confirmed by headspace gas analysis. The pre-enriched consortia were used as inocula in the biodegradation assays. Microbial growth in the presence of PVC films as major carbon and energy source was monitored by measuring the volume and composition of the gas produced in the headspace of each consortium. After 7 months incubation, significant gas volumes were measured in 10 microcosms (Figure 1A) reaching a maximum of 19.0 ± 1.0 ml, while no gas production was detected in the four abiotic controls. Headspace gas analysis showed that CO₂ was produced in all the microcosms while hydrogen was recorded only in the case of S2-M1 consortium (5.8 ± 0.0 ml) (Figure 1A). Methane (max volume = 5.7 ± 0.1 ml) was produced by the four consortia enriched in M2 medium, while very low/no methane production was recorded in microcosms enriched in M1, M3 and M4 media. These results showed that growth of the native marine microbial communities occurred in the presence of PVC plastic film as major C and energy source. Plasticizers probably sustained the observed growth, in line with previous studies carried out with PVC plastic under sulfate reducing conditions with bacteria enriched from landfill (Tsuchida *et al.*, 2011).

Based on these results, the 10 microcosms displaying clear microbial growth on PVC films (S2-M1, S2-M2, S3-M2, S4-M2, S1-M3, S3-M3, S4-M3, S2-M4, S3-M4 and S4-M4) were selected for further analysis.

3.1.2. Evaluation of microbial colonization of PVC film surface.

Among PVC films incubated in the 10 microcosms, four showed significantly higher (>10 times) amount of adhered proteins compared to their corresponding abiotic controls (Figure 1B),

219 suggesting an extensive microbial colonization of plastic surface. These include S2-M2, S3-M2, S4-
220 M3 and S2-M4, with a protein content (mean \pm SD) of 57.3 \pm 2.9, 45.6 \pm 2.3, 88.9 \pm 4.4 and 76.3 \pm 3.8 μ g
221 of BSA equivalent/mg of PVC film, respectively. Microbial colonization of PVC films was reported
222 under aerobic conditions. Specifically, unpretreated plasticized PVC films exposed to the
223 atmosphere (Blackley, Manchester, United Kingdom) for 95 weeks were found to be colonized by
224 fungi while no bacterial colonization was recorded (Webb *et al.*, 2000). Dang *et al.* (2008)
225 evaluated biofilm formation on different polymer surfaces including PVC plates, submerged in
226 seawater (1 m below the water surface) near the Qingdao coast for 72 h. Biofilm formation was
227 shown to occur on the different surfaces based on detection of 126 operational taxonomic units with
228 prokaryotes being predominant.

229 To the best of our knowledge, to date there is only one report on microbial colonization of PVC
230 plastic surface under anaerobic conditions. The Archaeal strain *Methanosarcina barkei* was found
231 to be able to adhere on PVC films after 2 h of exposure and to putatively produce exopolymetric
232 substances (EPS), as revealed by epifluorescence and scanning electron microscope observations
233 (Nguyen *et al.*, 2016).

234 Biofilm formation is considered as the first step of biodegradation activity against water
235 insoluble polymeric materials (Das *et al.*, 2012; Reisser *et al.*, 2014; Wang *et al.*, 2016; Zettler *et*
236 *al.*, 2013). Hence, only microbial consortia that showed significantly high adhered protein contents
237 on PVC films, i.e. S2-M2, S3-M2, S4-M3 and S2-M4 (Figure 1B), were selected for further
238 analyses. Indeed, for the PVC films incubated in these microcosms, a 10-fold protein content was
239 recorded compared to the corresponding abiotic controls. This threshold was fixed in order to
240 overcome the limitations of the Lowry assay related to the interference of molecules other than
241 proteins (like compounds that derive from the additives components of the plastic film), in the
242 colorimetric measurements (Everette *et al.*, 2010).

3.1.3. Evaluation of PVC biodegradation

Biodegradation activity was evaluated on film samples withdrawn from the four (S2-M2, S3-M2, S4-M3 and S2-M4) selected microcosms. Gravimetric measurements revealed a statistically higher decrease in their weights ($p < 0.05$; Figure 2 A-D) compared to those recovered from the corresponding abiotic controls. Specifically, weight reductions (mean \pm SD%) of $11.67\pm0.58\%$, $10.71\pm0.54\%$, $6.26\pm0.31\%$ and $5.77\pm0.29\%$ were recorded for the films incubated in S4-M3, S2-M4, S2-M2 and S3-M2 microcosms, respectively. Among the biological effect of the four selected consortia, the highest gravimetric weight decrease was recorded by films incubated with S4-M3 and S2-M4 consortia, which showed the same statistical film weight reduction followed by the action of S2-M2 and S3-M2 consortia ($p < 0.05$; Figure 2 D). The corresponding abiotic controls incubated in the growth media M3, M4 and M2 showed statistically equal weight losses of $3.71\pm0.28\%$, $3.91\pm0.2\%$ and $3.51\pm0.18\%$ respectively ($p < 0.05$; Figure 2 A-D) due to additives leaching from the PVC films (Kastner *et al.*, 2012; Suhrhoff and Scholz-Bottcher, 2016; Wang *et al.*, 2016). These evidences are the first proof of plastic biodegradation; indeed, the reduction of gravimetric weight, which is a direct evidence of the loss of material, is widely used in biodegradation tests (Shah *et al.* 2008; Wang *et al.*, 2016). Thus some components of the PVC films were degraded by the enriched microbial communities. Since the films used in this study had about 30%_{w/w} of additives, further analyses were performed in order to elucidate if the biodegradation regarded the sole additives or also the PVC polymer.

TGA showed a lower thermal stability of the films recovered from the four active microcosms (Figure 2A-C) compared to those incubated under abiotic conditions. Specifically, a T_{\max} ranging from 287°C to 277°C was recorded for the former compared to a T_{\max} of 292°C for those of abiotic controls or 294°C for the non incubated PVC film. Considering that the presence of additives (citrates, polyadipates, epoxidized soybean oil, Zn etc; data supplied by the production company) is lowering the thermal stability of PVC resins once in PVC plastic film, the (bio)degradation of the sole additives would lead to an increase of the film thermal stability i.e. a shift of the TGA curve

269 toward that of the resin. On the opposite, here a reduction of such parameter was observed and
270 would suggest that a microbial attack towards PVC polymer chains occurred.

271 Also GPC analyses exhibited a decrease of the average molecular weight ($M_n\%$; mean \pm SD) from
272 100% to 91.1 \pm 3.2%, 94.3 \pm 3.3%, 95.5 \pm 3.3%, and 98.1 \pm 3.4% for PVC films incubated with S4-M3,
273 S3-M2, S2-M2, and S2-M4 consortia, respectively (Figure 2E). According to statistical analysis,
274 PVC film incubated with S4-M3 exhibited a statistically lower $M_n(\%)$ compared to the film
275 incubated with its abiotic control ($p < 0.05$, Figure 2E), suggesting that this consortium was able to
276 statistically reduce PVC molecular weight. On the other hand, no statistically significant differences
277 were recorded between molecular weights of films incubated with all consortia and pristine PVC
278 film ($p > 0.05$; Figure 2E). However, the fact that the differences between biotically treated and
279 pristine PVC films are not statistically significant does not imply that no biological action has
280 occurred and the interpretation of the results should focus on the scientific importance rather than
281 on the statistical significance. Indeed, considering the nonbiodegradable nature of the polymer even
282 small changes that are not statistically significant are important from a biodegradation view point.
283 Hence, the GPC results are in line with those of TGA and suggest that some polymer chains were
284 biodegraded with the production of low M_w PVC chains in the case of S4-M3, S3-M2 and S2-M2
285 microcosms (Figure 2E). On the other hand, GPC analyses revealed an increase of M_n of abiotic
286 controls up to 105.5 \pm 3.7% (mean \pm SD; Figure 2E) although no appreciable changes in their thermal
287 stability were recorded. Such results could be explained by a loss of water-soluble low molecular
288 weight additives other than thermal stabilizers, which has led to M_n increase without change in the
289 thermal stability.

290

291 **3.2. PVC films biodegradation after 24 months**

292 Figure 3 shows the gravimetric weight reductions of the PVC films incubated in the 16
293 biologically active microcosms or in the corresponding abiotic controls after 24 months incubation.
294 According to statistical analysis, eleven consortia (three in M2, 4 in M3 and 4 in M4 medium) were

295 shown to significantly reduce film weight compared to their abiotic controls ($p < 0.05$; Figure 3).
296 Four and three consortia enriched in M4 and M3 media reduced the gravimetric weight (mean \pm SD)
297 of the PVC films by up to $12.97\pm0.65\%$ ($p < 0.05$; showed by d letter in Figure 3). A statistically
298 lower weight loss of up to $9.23\pm0.46\%$ ($p < 0.05$; showed by c and e letters in Figure 3) was
299 recorded for three microcosms set up in M2 and for S2-M3 consortium, while a maximum weight
300 loss of $4.21\pm0.21\%$, statistically comparable to that recorded under abiotic conditions
301 ($4.32\pm0.22\%$), was observed for the four consortia enriched in M1 and in S4-M2 consortium ($p <$
302 0.05 ; Figure 3).

303 Based on these results, only 11 consortia (S1-M2, S2-M2, S3-M2, S1-M3, S2-M3, S3-M3, S4-
304 M3, S1-M4, S2-M4, S3-M4 and S4-M4), exhibiting statistically significant gravimetric weight
305 losses ($p < 0.05$), were selected for further analyses.

306 Compared to abiotic control and pristine (non-incubated) PVC films, the films withdrawn from
307 S2 and S3 consortia enriched in M2, the S4 consortium enriched in M3 as well as S1, S2 and S4
308 consortia enriched in M4 displayed a decrease of TGA-assessed thermal stability (Table 1),
309 confirming the evidences observed after 7 months incubation.

310 On the other hand, the GPC analyses showed an increase of the M_n (%) of the films incubated
311 with 8/11 consortia (S2-M2, S3-M2, S2-M3, S4-M3, S1-M4, S2-M4, S3-M4 and S4-M4) including
312 those that exhibited M_n (%) lower than 100% at month 7, even if no statistically significant
313 differences were found among M_n (%) after the analysis of variance (ANOVA: $p = 0.19$; $F = 1.44$;
314 $df = 14$) (Figure 2E, Figure 4). Once again, the fact that no statistically significant differences in M_n
315 (%) were observed does not imply no modifications of the M_n (%) has occurred. Indeed, it is
316 important to notice that M_n values for polymeric substances are very high numbers and therefore
317 differences in such number should be very high to be considered statistically different.
318 Consequently, in the case of data obtained from a biodegradation of materials known as “non
319 biodegradable”, small differences in M_n (%) are often observed and should be acknowledged as of
320 scientific importance rather than of statistical significance. Hence, these results suggest that the

three consortia found to be the most active at the 7th month together with consortia S1, S2 and S4 enriched in M4 that displayed activity after longer incubation period, are apparently able to degrade PVC plasticizers and polymer chains. Indeed, considering that biodegradation of low molecular weight compounds affects prevalently the M_n (%) rather than the gravimetric weight, the M_n increase observed after 24 months incubation could be explained by the full biodegradation of the short polymer chains putatively produced within the first months of incubation and/or the release of the broken chains and of the lower molecular weight additives into the growth medium.

3.3. Characterization of microbial communities

Characterization of the anaerobic communities enriched in the presence of PVC films was performed on the 11 different consortia exhibiting a mass reduction significantly higher than the corresponding abiotic controls after 24 months incubation.

From the DGGE profiles (Figure 5), 13 (band #1 to #13) and 3 (band #A to #C) bands were cut from the bacterial and archaeal communities, respectively.

The bacterial biofilm communities enriched in M2, M3 and M4 media were characterized by the presence of two bands that were detected in most consortia and which include band #8 and band #10 detected in 9 and 8 out of the 11 consortia, respectively (Figure 5, Table 2). The band #8 has highest sequence similarity with different Clostridia including uncultured *Clostridium* clone reported from an Alaskan mesothermic petroleum reservoir (Pham *et al.*, 2009), Clostridiales bacteria able to degrade the explosive UXO5-23 (Zhao *et al.*, 2007) or *Clostridium* sp. AN-D reported from high pressure prokaryotic enrichments from deep marine gas hydrate sediments (Parkes *et al.*, 2009). The band #10 has 98% similarity with *Acetobacterium* sp. Other two bands (band #9 and band #7) which have, respectively, 100% and 99% of similarity with *Dethiosulfovibrio* sp. and *Sporobacter* sp. were detected in 3/11 consortia (Figure 5, Table 2). Different other bands were detected in some of the communities enriched in specific media and having homologies with: i) *Erysipelothrix* sp. (band #1) mainly in M4 communities; ii) *Fusibacter*

347 (band #13) and *Psychromonas* (band #4) in some M3 communities and iii) *Desulfovibrio* (band
348 #12), *Cupriavidus* (band #5), *Cohaesibacter* sp. (band #11) and *Pleomorphochaeta* (band #6)
349 detected mainly in M2 consortia (Figure 5, Table 2).

350 For all the consortia, almost no differences in the composition of the bacterial biofilm and its
351 corresponding planktonic community were observed. Similarly, composition of the planktonic and
352 biofilm archaeal communities for each consortium were very similar. They were composed of bands
353 having high similarities with i) *Methanosaeta* sp. (band A) present in all M4 and M3 communities,
354 ii) *Methanococcoides* sp. (band B) detected in the three M2 communities and iii) *Methanogenium*
355 sp. (band C) found only in some M2 communities (Figure 5, Table 2).

356 Although the chemical analyses indicate the biodegradation of PVC polymer by the consortia
357 S2-M2, S3-M2, S4-M3, S1-M4, S2-M4 and S4-M4, none common microbial phylotype was
358 observed in all such consortia and this seems to suggest that more than a microbial specie were
359 involved in the biodegradation. The occurrence of high NaCl contents in the microcosm media did
360 not allow to determine changes in chlorine ion concentration in the culture supernatants and
361 therefore to assess if the degradation was accompanied by the dehalogenation of PVC polymer.
362 However, the following observations support PVC dechlorination. Among the microbial
363 components of two communities (S1-M2; S1-M3), bands having a high similarity with the
364 facultative organohalide respiring bacterium *Desulfovibrio dechloracetivorans* (band #12, S1-M2)
365 (Sun *et al.*, 2000) and with *Fusibacter* sp. (band #13, S1-M3) were detected. *Fusibacter* sp. was
366 reported as dominant component of tidal flat communities where *Dehalococcoides*-like bacteria
367 were not detected although dechlorination of PCE-to-cis-DCE has occurred (Lee *et al.*, 2011).
368 Further, bands having highest sequence similarity with Clostridia were detected in almost all
369 consortia. The class of Clostridia includes bacterial species (bacteria from the genus *Clostridium*)
370 reported to play an important role in dechlorinating consortia, producing H₂ that is used as electron
371 donor by the dechlorinating bacteria (Bowman *et al.*, 2010).

372 Bacteria of the genus *Acetobacterium* were detected in 8/11 consortia. Among these
373 homoacetogens, some species have been reported to be able to mineralize tetrachloromethane (Egli
374 *et al.*, 1988). *Acetobacterium* spp. was also detected among the components of consortia able to
375 dechlorinate 1,1,2,2-tetrachloroethane (Manchester *et al.*, 2012). Patil *et al.* (2014) reported
376 successful dechlorination of tetrachloroethene by a microbial community that did not include
377 classical dechlorinators due to the synergistic work of the community i.e. *Enterobacter* and
378 *Desulfovibrio* spp. as putative dechlorinators and *Bacteroidetes* and *Firmicutes* as acetate
379 fermenters (Patil *et al.*, 2014).

380 Other bands having high similarity with Bacteroidales bacterium CF (band#3, 93% of identity),
381 were also detected. This bacterium was found to be the most abundant non dechlorinating bacterium
382 in a community that reductively dechlorinates multichlorine molecules; it is able to ferment lactate,
383 ethanol and glucose in the presence of concentrations of chloroform or trichloroethane that usually
384 inhibit reductive dechlorination and methanogenesis (Tang *et al.*, 2013).

385 The archaeal communities present in 8/11 consortia enriched in M3 and M4 media were
386 composed mainly by members of the Phylum Euryarchaeota, with a unique member of the
387 Methanosarcinales (Genus *Methanosaeta*, species *pelagica*, Figure 5, Table 2) known for its
388 acetoclastic methanogenic activities. Oh *et al.* (2008) showed that acetoclastic methanogenic
389 Archaea may play an indirect role in dechlorination of halogenated organic compounds such as
390 PCBs by synthesizing the corrinoid cofactor cobalamin required for vitamin B12 synthesis, which is
391 necessary for assembling functional reductive dehalogenase systems (Patil *et al.*, 2014; Rupakula *et*
392 *al.*, 2014; Yan *et al.*, 2013). Cobalamine and other precursors are typically produced by
393 methanogenic Archaea as cofactors for methyltransferases involved in methane production (Yan *et*
394 *al.*, 2013).

395 Specialized dehalorespiring bacteria among the components of the anaerobic consortia studied
396 were, however, not detected. This could be due to PCR bias i.e. the necessity to use more specific

397 amplification primers in order to reveal the presence of such bacteria (Aulenta *et al.*, 2004;
 398 Dowideit *et al.*, 2010; Duhamel *et al.*, 2006; Manchester *et al.*, 2012).
 399 Besides of the bacterial and archaeal phyla potentially playing an important role in dechlorinating
 400 PVC, the other bacterial and archaeal clades were detected and could be involved in additives
 401 biodegradation. These include *Dethiosulfovibrio* sp.; *Erysipelothrix* sp.; *Sporobacter* sp.;
 402 *Psychromonas* sp.; *Cupriavidus* sp. and *Cohaesibacter* sp. (Bonin *et al.*, 2002; Canion *et al.*, 2013;
 403 Diels *et al.*, 2009; GrechMora *et al.*, 1996; Hwang *et al.*, 2008; Pantazidou *et al.*, 2007; Stackebrandt
 404 *et al.*, 2006; Surkov *et al.*, 2001) as well as *Methanococcoides* sp. (99% of identity, Figure 5, Table
 405 2) detected in three M2 consortia or *Methanogenium* sp. detected only in S1-M2 consortium (Figure
 406 5, Table 2) (Miller *et al.*, 2016).

407

408 **4. Conclusions**

409 A significant biodegradation of plasticized untreated PVC films by marine anaerobic
 410 microbial consortia was demonstrated in this study. After 7 months incubation, three of 16 enriched
 411 consortia were able to degrade the films apparently acting against both the additives and the
 412 polymer chains. For these consortia, GPC showed a decrease in M_n suggesting that some polymer
 413 chain scission took place leading to the formation of smaller fragments, while TGA and gravimetric
 414 weight loss (%) revealed a decrease in thermal stability and a mass loss. Prolongation of the
 415 incubation to 24 months allowed the selection of other three consortia able to display the same
 416 potential.

417 To our knowledge, this is the first study reporting on biodegradation of plasticized PVC films by
 418 marine microbial communities enriched under anoxic conditions and providing information about
 419 the composition of such communities potentially responsible for it. The PVC film-associated
 420 communities of the biodegrading microcosms encompasses microbial phyla closely related to those

421 previously reported from laboratory/field enrichments for their ability to degrade halogenated
422 organic compounds and hydrocarbons.

423 The enriched consortia originate from marine samples and the activity recorded was obtained
424 under lab conditions mimicking the sulfate reducing, nitrate reducing and methanogenic conditions
425 occurring in marine environment. Thus the present study provides relevant insights on the potential
426 fate of PVC plastic films occurring in anoxic marine impacted ecosystems.

427

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References

- Ali M.I., Ahmed S., Robson G., Javed I., Ali N., Atiq N., Hameed A., 2014. Isolation and molecular characterization of polyvinyl chloride (PVC) plastic degrading fungal isolates, J. Basic. Microbiol. 54, 18-27. DOI: 10.1002/jobm.201200496.
- Anwar M.S., Kapri A., Chaudhry V., Mishra A., Ansari M.W., Souche Y., Nautiyal C.S., Zaidi M.G., Goel R., 2016. Response of indigenously developed bacterial consortia in progressive degradation of polyvinyl chloride, Protoplasma. 253, 1023-1032. DOI: 10.1007/s00709-015-0855-9.
- Aulenta F., Rossetti S., Majone M., Tandoi V., 2004. Detection and quantitative estimation of *Dehalococcoides* spp. in a dechlorinating bioreactor by a combination of fluorescent in situ hybridisation (FISH) and kinetic analysis, Appl. Microbiol. Biotechnol. 64, 206-212. DOI: 10.1007/s00253-003-1503-4.
- Avio C.G., Gorbi S., Regoli F., 2017. Plastics and microplastics in the oceans: From emerging pollutants to emerged threat, Mar. Environ. Res. 128, 2-11. DOI: 10.1016/j.marenvres.2016.05.012.
- Bonin P.C., Michotey V.D., Mouzdahir A., Rontani J.F., 2002. Anaerobic biodegradation of squalene: Using DGGE to monitor the isolation of denitrifying Bacteria taken from enrichment cultures, FEMS Microbiol. Ecol. 42, 37-49. DOI: 10.1111/j.1574-6941.2002.tb00993.x.
- Bowman K.S., Dupre R.E., Rainey F.A., Moe W.M., 2010. *Clostridium hydrogeniformans* sp. nov. and *Clostridium cavendishii* sp. nov., hydrogen-producing bacteria from chlorinated solvent-contaminated groundwater, Int. J. Syst. Evol. Microbiol. 60, 358-363. DOI: 10.1099/ijs.0.013169-0.
- Breitburg D., Levin L.A., Oschlies A., Gregoire M., Chavez F.P., Conley D.J., Garcon V., Gilbert D., Gutierrez D., Isensee K., Jacinto G.S., Limburg K.E., Montes I., Naqvi S.W.A., Pitcher G.C., Rabalais N.N., Roman M.R., Rose K.A., Seibel B.A., Telszewski M., Yasuhara M.,

459 Zhang J., 2018. Declining oxygen in the global ocean and coastal waters. *Science*. 359, 6371-
 460 6382. DOI: 10.1126/science.aam7240.

461 Bueno-Ferrer C., Garrigos M.C., Jimenez A., 2010. Characterization and thermal stability of
 462 poly(vinyl chloride) plasticized with epoxidized soybean oil for food packaging, *Polym.*
 463 *Degrad. Stabil.* 95, 2207-2212. DOI: 10.1016/j.polymdegradstab.2010.01.027.

464 Canion A., Prakash O., Green S.J., Jahnke L., Kuypers M.M., Kostka J.E., 2013. Isolation and
 465 physiological characterization of psychrophilic denitrifying bacteria from permanently cold
 466 Arctic fjord sediments (Svalbard, Norway), *Environ. Microbiol.* 15, 1606-1618. DOI:
 467 10.1111/1462-2920.12110.

468 Celikkol-Aydin S., Gaylarde C. C., Lee T., Melchers R. E., Witt D. L., Beech I. B., 2016. 16S
 469 rRNA gene profiling of planktonic and biofilm microbial populations in the Gulf of Guinea
 470 using Illumina NGS. *Mar Environ Res* 122, 105-112. DOI: 10.1016/j.marenvres.2016.10.001.

471 Dang H., Li T., Chen M., Huang G., 2008. Cross-ocean distribution of *Rhodobacterales* bacteria as
 472 primary surface colonizers in temperate coastal marine waters, *Appl. Environ. Microbiol.* 74,
 473 52-60. DOI: 10.1128/aem.01400-07.

474 Dang H. and Lovell C. R., 2016. Microbial surface colonization and biofilm development in marine
 475 environments. *Microbiol. Mol. Biol. Rev.* 80, 91-138. DOI: 10.1128/MMBR.00037-15.

476 Das G., Bordoloi N.K., Rai S.K., Mukherjee A.K., Karak N., 2012. Biodegradable and
 477 biocompatible epoxidized vegetable oil modified thermostable poly(vinyl chloride): thermal
 478 and performance characteristics post biodegradation with *Pseudomonas aeruginosa* and
 479 *Achromobacter* sp., *J. Hazard. Mater.* 209-210, 434-442. DOI: 10.1016/j.jhazmat.2012.01.043.

480 Diels L., Van Roy S., Taghavi S., Van Houdt R., 2009. From industrial sites to environmental
 481 applications with *Cupriavidus metallidurans*, *Antonie van Leeuwenhoek.* 96, 247-258. DOI:
 482 10.1007/s10482-009-9361-4.

483 Dowideit K., Scholz-Muramatsu H., Miethling-Graff R., Vigelahn L., Freygang M., Dohrmann
 484 A.B., Tebbe C.C., 2010. Spatial heterogeneity of dechlorinating bacteria and limiting factors

485 for in situ trichloroethene dechlorination revealed by analyses of sediment cores from a
 486 polluted field site, FEMS Microbiol. Ecol. 71, 444-459. DOI: 10.1111/j.1574-
 487 6941.2009.00820.x.

488 Duhamel M., Edwards E.A., 2006. Microbial composition of chlorinated ethene-degrading cultures
 489 dominated by *Dehalococcoides*, FEMS Microbiol. Ecol. 58, 538-549. DOI: 10.1111/j.1574-
 490 6941.2006.00191.x.

491 Egli C., Tschan T., Scholtz R., Cook A.M., Leisinger T., 1988. Transformation of
 492 tetrachloromethane to dichloromethane and carbon dioxide by *Acetobacterium woodii*, App.
 493 and environ. microbiol. 54, 2819-2824. PMCID: PMC204379; PMID: 3145712

494 Everette J.D., Bryant Q.M., Green A.M., Abbey Y.A., Wangila G.W., Walker R.B., 2010.
 495 Thorough study of reactivity of various compound classes toward the Folin-Ciocalteu reagent,
 496 J. Agric. Food Chem. 58, 8139-8144. DOI: 10.1021/jf1005935.

497 Galloway T.S., Cole M., Lewis C., 2017. Interactions of microplastic debris throughout the marine
 498 ecosystem, Nat. Ecol. Evol. 1, 116-123. DOI: 10.1038/s41559-017-0116.

499 Gewert B., Plassmann M.M., MacLeod M., 2015. Pathways for degradation of plastic polymers
 500 floating in the marine environment, Environ. Sci.-Proc. Imp. 17, 1513-1521. DOI:
 501 10.1039/c5em00207a

502 Giacomucci L., Raddadi N., Soccio M., Lotti N., Fava F., 2019. Polyvinylchloride biodegradation
 503 by *Pseudomonas citronellolis* and *Bacillus flexus*. N. Biotechnol. 52, 35-41. DOI:
 504 10.1016/j.nbt.2019.04.005.

505 Giovannoni S.J., Delong E.F., Olsen G.J., Pace N.R., 1988. Phylogenetic Group-Specific
 506 Oligodeoxynucleotide Probes for Identification of Single Microbial-Cells, J. Bacteriol. 170,
 507 720-726. DOI: 10.1128/jb.170.2.720-726.1988.

508 Glas D., Hulsbosch J., Dubois P., Binnemans K., De Vos D.E., 2014. End-of-life treatment of
 509 poly(vinyl chloride) and chlorinated polyethylene by dehydrochlorination in ionic liquids,
 510 Chem. Sus. Chem. 7, 610-617. DOI: 10.1002/cssc.201300970.

511 GrechMora I., Fardeau M.L., Patel B.K.C., Ollivier B., Rimbault A., Prensier G., Garcia J.L.,
 512 GarnierSillam E., 1996. Isolation and characterization of *Sporobacter termitidis* gen nov sp
 513 nov, from the digestive tract of the wood-feeding termite *Nasutitermes lujae*, Int. J. Syst.
 514 Bacteriol. 46, 512-518. DOI: 10.1099/00207713-46-2-512.

515 Harrison J.P., Schratzberger M., Sapp M., Osborn A.M., 2014. Rapid bacterial colonization of low-
 516 density polyethylene microplastics in coastal sediment microcosms, BMC Microbiol. 14, 232-
 517 246. DOI: 10.1186/s12866-014-0232-4.

518 Hwang C.Y., Cho B.C., 2008. *Cohaesibacter gelatinilyticus* gen. nov., sp. nov., a marine bacterium
 519 that forms a distinct branch in the order Rhizobiales, and proposal of *Cohaesibacteraceae* fam.
 520 nov, Int. J. Syst. Evol. Micr. 58, 267-277. DOI: doi:10.1099/ijs.0.65016-0.

521 Kastner J., Cooper D. G., Maric M., Dodd P., Yargeau V., 2012. Aqueous leaching of di-2-
 522 ethylhexyl phthalate and "green" plasticizers from poly(vinyl chloride). Sci. Total Environ.
 523 432, 357-364. DOI: 10.1016/j.scitotenv.2012.06.014.

524 Lee J., Lee T.K., Löffler F.E., Park J., 2011. Characterization of microbial community structure and
 525 population dynamics of tetrachloroethene-dechlorinating tidal mudflat communities,
 526 Biodegradation. 22, 687-698. DOI: 10.1007/s10532-010-9429-x.

527 Lowry O.H., Rosebrough N.J., Farr A.L., Randall R.J., 1951. Protein measurement with the Folin
 528 phenol reagent, The Journal of biological chemistry. 193, 265-275. PMID: 14907713.

529 Manchester M.J., Hug L.A., Zarek M., Zila A., Edwards E.A., 2012. Discovery of a trans-
 530 dichloroethene-respiring Dehalogenimonas species in the 1,1,2,2-tetrachloroethane-
 531 dechlorinating WBC-2 consortium, Appl. Environ. Microbiol. 78, 5280-5287. DOI:
 532 10.1128/AEM.00384-12.

533 Miller H.M., Matter J.M., Kelemen P., Ellison E.T., Conrad M.E., Fierer N., Ruchala T., Tominaga
 534 M., Templeton A.S., 2016. Modern water/rock reactions in Oman hyperalkaline peridotite
 535 aquifers and implications for microbial habitability, Geochim. Cosmochim. Ac. 179, 217-241.
 536 DOI: 10.1186/1471-2180-10-149.

537 Nguyen V., Karunakaran E., Collins G., Biggs C.A., 2016. Physicochemical analysis of initial
 538 adhesion and biofilm formation of *Methanosarcina barkeri* on polymer support material,
 539 Colloids Surf. B Biointerfaces. 143, 518-525. DOI: 10.1016/j.colsurfb.2016.03.042.
 540 Oberbeckmann S., Osborn A.M., Duhaime M.B., 2016. Microbes on a bottle: substrate, season and
 541 geography influence community composition of microbes colonizing marine plastic debris,
 542 PloS one. 11, e0159289. DOI: 10.1371/journal.pone.0159289.
 543 Oh K.H., Ostrofsky E.B., Cho Y.C., 2008. Molecular characterization of polychlorinated biphenyl-
 544 dechlorinating populations in contaminated sediments, J. Microbiol. 46, 165-173. DOI:
 545 10.1007/s12275-007-0214-4.
 546 Pantazidou M., Kapniaris S., Katsiri A., Christidis A., 2007. Pollutant trends and hazard ranking in
 547 Elefsis Bay, Greece, Desalination. 210, 69-82. DOI: 10.1016/j.desal.2006.05.034.
 548 Parkes R.J., Seltek G., Webster G., Martin D., Anders E., Weightman A.J., Sass H., 2009.
 549 Culturable prokaryotic diversity of deep, gas hydrate sediments: first use of a continuous high-
 550 pressure, anaerobic, enrichment and isolation system for subseafloor sediments (DeepIsoBUG),
 551 Environ. Microbiol. 11, 3140-3153. DOI: 10.1111/j.1462-2920.2009.02018.x
 552 Patil S.S., Adetutu E.M., Aburto-Medina A., Menz I.R., Ball A.S., 2014. Biostimulation of
 553 indigenous communities for the successful dechlorination of tetrachloroethene
 554 (perchloroethylene)-contaminated groundwater, Biotechnol. Lett. 36, 75-83. DOI:
 555 10.1007/s10529-013-1369-1.
 556 Peixoto J., Silva L.P., Kruger R.H., 2017. Brazilian Cerrado soil reveals an untapped microbial
 557 potential for untreated polyethylene biodegradation, J. Hazard. Mater. 324, 634-644. DOI:
 558 10.1016/j.jhazmat.2016.11.037.
 559 Pham V.D., Hnatow L.L., Zhang S., Fallon R.D., Jackson S.C., Tomb J.F., DeLong E.F., Keeler
 560 S.J., 2009. Characterizing microbial diversity in production water from an Alaskan
 561 mesothermic petroleum reservoir with two independent molecular methods, Environ.
 562 Microbiol. 11, 176-187. DOI: 10.1111/j.1462-2920.2008.01751.x.

563 PlasticsEurope, 2019. Plastics – the Facts 2019, in PlasticsEurope 2019.

564 Polo A., Cappitelli F., Brusetti L., Principi P., Villa F., Giacomucci L., Ranalli G., Sorlini C., 2010.

565 Feasibility of removing surface deposits on stone using biological and chemical remediation

566 methods, *Microb. Ecol.* 60, 1-14. DOI: 10.1007/s00248-009-9633-6.

567 Raddadi, N. and F. Fava, 2019. Biodegradation of oil-based plastics in the environment: existing

568 knowledge and needs of research and innovation. *Sci Total Environ* 679, 148-158. DOI:

569 10.1016/j.scitotenv.2019.04.419.

570 Reddy M.S., Yamaguchi T., Okuda T., Tsai T.Y., Nakai S., Nishijima W., Okada M., 2010.

571 Feasibility study of the separation of chlorinated films from plastic packaging wastes, *Waste*

572 *Manag.* 30, 597-601. DOI: 10.1016/j.wasman.2009.11.028.

573 Reisser J., Shaw J., Hallegraeff G., Proietti M., Barnes D.K., Thums M., Wilcox C., Hardesty B.D.,

574 Pattiaratchi C., 2014. Millimeter-sized marine plastics: a new pelagic habitat for

575 microorganisms and invertebrates, *PloS one.* 9, e100289. DOI: 10.1371/journal.pone.0100289.

576 Ruiz-Orejón L.F., Sardá R., Ramis-Pujol J., 2016. Floating plastic debris in the Central and Western

577 Mediterranean Sea, *Mar. Environ. Res.*, 120, 136-144. DOI:10.1016/j.marenvres.2016.08.001.

578 Rupakula A., Lu Y., Kruse T., Boeren S., Holliger C., Smidt H., Maillard J., 2014. Functional

579 genomics of corrinoid starvation in the organohalide-respiring bacterium *Dehalobacter*

580 *restrictus* strain PER-K23, *Front. Microbiol.* 5, 751-760. DOI: 10.3389/fmicb.2014.00751.

581 Shah A.A., Hasan F., Hameed A., Ahmed S., 2008. Biological degradation of plastics: a

582 comprehensive review, *Biotechnol. Adv.* 26, 246-265. DOI: 10.1016/j.biotechadv.2007.12.005.

583 Shah A.A., Kato S., Shintani N., Kamini N.R., Nakajima-Kambe T., 2014. Microbial degradation of

584 aliphatic and aliphatic-aromatic co-polyesters, *Appl. Microbiol. Biotechnol.* 98, 3437-3447.

585 DOI: 10.1007/s00253-014-5558-1.

586 Stackebrandt E., Rebolí A.C., Farrar W.E., 2006. The genus *Erysipelothrix*, in: Dworkin M.,

587 Falkow S., Rosenberg E., Schleifer K.-H., Stackebrandt E. (Eds.) *The prokaryotes: volume 4.*

588 Bacteria: firmicutes, cyanobacteria, Springer US, New York, NY, pp. 492-510. DOI:
589 10.1007/0-387-30744-3.

590 Stahl D.A., Amann R.I., 1991. Development and application of nucleic acid probes in bacterial
591 systematics, in: Stackebrandt E., Goodfellow M. (Eds.) Nucleic acid techniques in bacterial
592 systematics, John Wiley and sons, Chichester, England. pp. 205-248. DOI:
593 10.1002/jobm.3620310616.

594 Suaria G., Aliani S., 2014. Floating debris in the Mediterranean Sea, Mar. Pollut. Bull. 86, 494-504.
595 DOI:10.1016/j.marpolbul.2014.06.025.

596 Suhrhoff T. J. and Scholz-Bottcher B. M., 2016. Qualitative impact of salinity, UV radiation and
597 turbulence on leaching of organic plastic additives from four common plastics - A lab
598 experiment. Mar. Pollut. Bull. 102, 84-94. DOI: 10.1016/j.marpolbul.2015.11.054.

599 Sun B., Cole J.R., Sanford R.A., Tiedje J.M., 2000. Isolation and characterization of *Desulfovibrio*
600 *dechloracetivorans* sp. nov., a marine dechlorinating bacterium growing by coupling the
601 oxidation of acetate to the reductive dechlorination of 2-chlorophenol, Appl. Environ.
602 Microbiol. 66, 2408-2413. DOI: 10.1128/aem.66.6.2408-2413.2000.

603 Surkov A.V., Dubinina G.A., Lysenko A.M., Glöckner F.O., Kuever J., 2001. *Dethiosulfovibrio*
604 *russensis* sp. nov., *Dethosulfovibrio marinus* sp. nov. and *Dethosulfovibrio acidaminovorans*
605 sp. nov., novel anaerobic, thiosulfate- and sulfur-reducing bacteria isolated from 'Thiodendron'
606 sulfur mats in different saline environments, Int. J. Syst. Evol. Micr. 51, 327-337. DOI:
607 10.1099/00207713-51-2-327.

608 Tang S., Edwards E.A., 2013. Complete genome sequence of bacteroidales strain CF from a
609 chloroform-dechlorinating enrichment culture, Genome Announc. 1, e01066-01013. DOI:
610 10.1128/genomeA.01066-13.

611 Tribedi, P., Dey, S., 2017. Pre-oxidation of low-density polyethylene (LDPE) by ultraviolet light
612 (UV) promotes enhanced degradation of LDPE in soil, Environ. Monit. Assess. 189, 624. DOI:
613 10.1007/s10661-017-6351-2.

614 Tsuchida D., Kajihara Y., Shimidzu N., Hamamura K., Nagase M., 2011. Hydrogen sulfide
615 production by sulfate-reducing bacteria utilizing additives eluted from plastic resins, Waste
616 Manag. Res. 29, 594-601. DOI: 10.1177/0734242X10388556.

617 Wang J., Tan Z., Peng J., Qiu Q., Li M., 2016. The behaviors of microplastics in the marine
618 environment, Mar. Environ. Res. 113, 7-17. DOI: 10.1016/j.marenvres.2015.10.014.

619 Webb J.S., Nixon M., Eastwood I.M., Greenhalgh M., Robson G.D., Handley P.S., 2000. Fungal
620 colonization and biodeterioration of plasticized polyvinyl chloride, Appl. Environ. Microb. 66,
621 3194-3200. DOI: 10.1128/Aem.66.8.3194-3200.2000.

622 Wright E.S., Yilmaz L.S., Noguera D.R., 2012. DECIPHER, a search-based approach to chimera
623 identification for 16S rRNA sequences, Appl. Environ. Microbiol. 78, 717-725. DOI:
624 10.1128/AEM.06516-11.

625 Yan J., Im J., Yang Y., Löffler F.E., 2013. Guided cobalamin biosynthesis supports
626 *Dehalococcoides mccartyi* reductive dechlorination activity, Philos. Trans. R. Soc. Lond. B.
627 Biol. Sci. 368, 20120320. DOI: 10.1098/rstb.2012.0320.

628 Yang Y., Yang J., Wu W.M., Zhao J., Song Y., Gao L., Yang R., Jiang L., 2015. Biodegradation
629 and mineralization of polystyrene by plastic-eating mealworms: part 2. Role of gut
630 microorganisms, Environ. Sci. Technol. 49, 12087-12093. DOI: 10.1021/acs.est.5b02663.

631 Zettler E.R., Mincer T.J., Amaral-Zettler L.A., 2013. Life in the “Plastisphere”: microbial
632 communities on plastic marine debris, Environ. Sci. Technol. 47, 7137-7146. DOI:
633 10.1021/es401288x.

634 Zhao J.S., Manno D., Hawari J., 2007. Abundance and diversity of octahydro-1,3,5,7-tetranitro-
635 1,3,5,7-tetrazocine (HMX)-metabolizing bacteria in UXO-contaminated marine sediments,
636 FEMS Microbiol. Ecol. 59, 706-717. DOI: 10.1111/j.1574-6941.2006.00248.x

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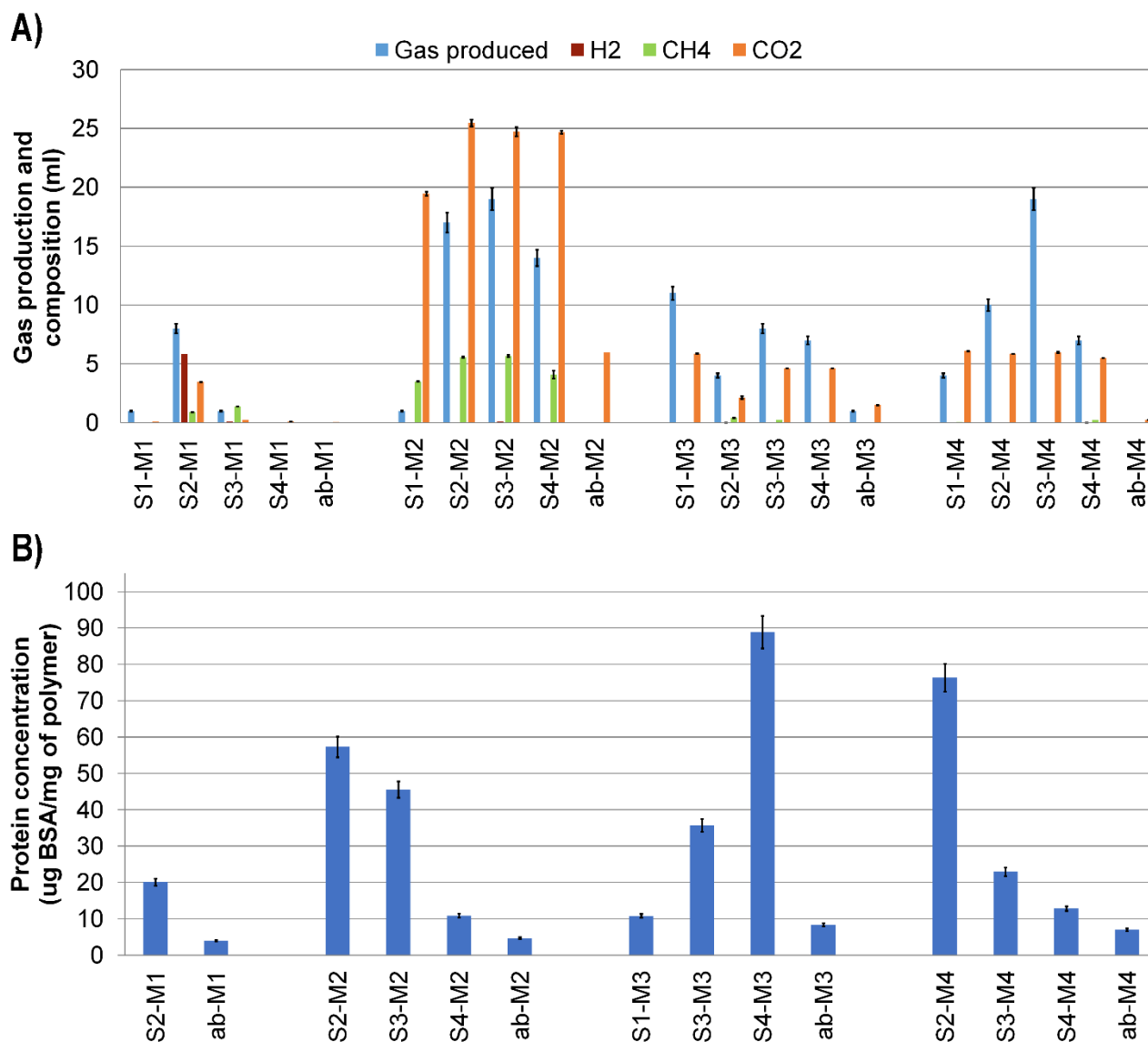


Figure 1. Evaluation of microbial growth and biofilm formation after 7 months incubation.

Volume and composition of gases produced by the sixteen different enriched marine consortia (the combination of 4 native marine/litter communities, i.e. S1 to S4, grown in the presence of 4 different media, i.e. M1 to M4) (a); Concentration of proteins adhered on surfaces of PVC films incubated under abiotic conditions (ab-M1 to ab-M4) or with the ten different enriched marine consortia showing highest microbial growth by gas analysis (b). Bars indicate standard deviation.

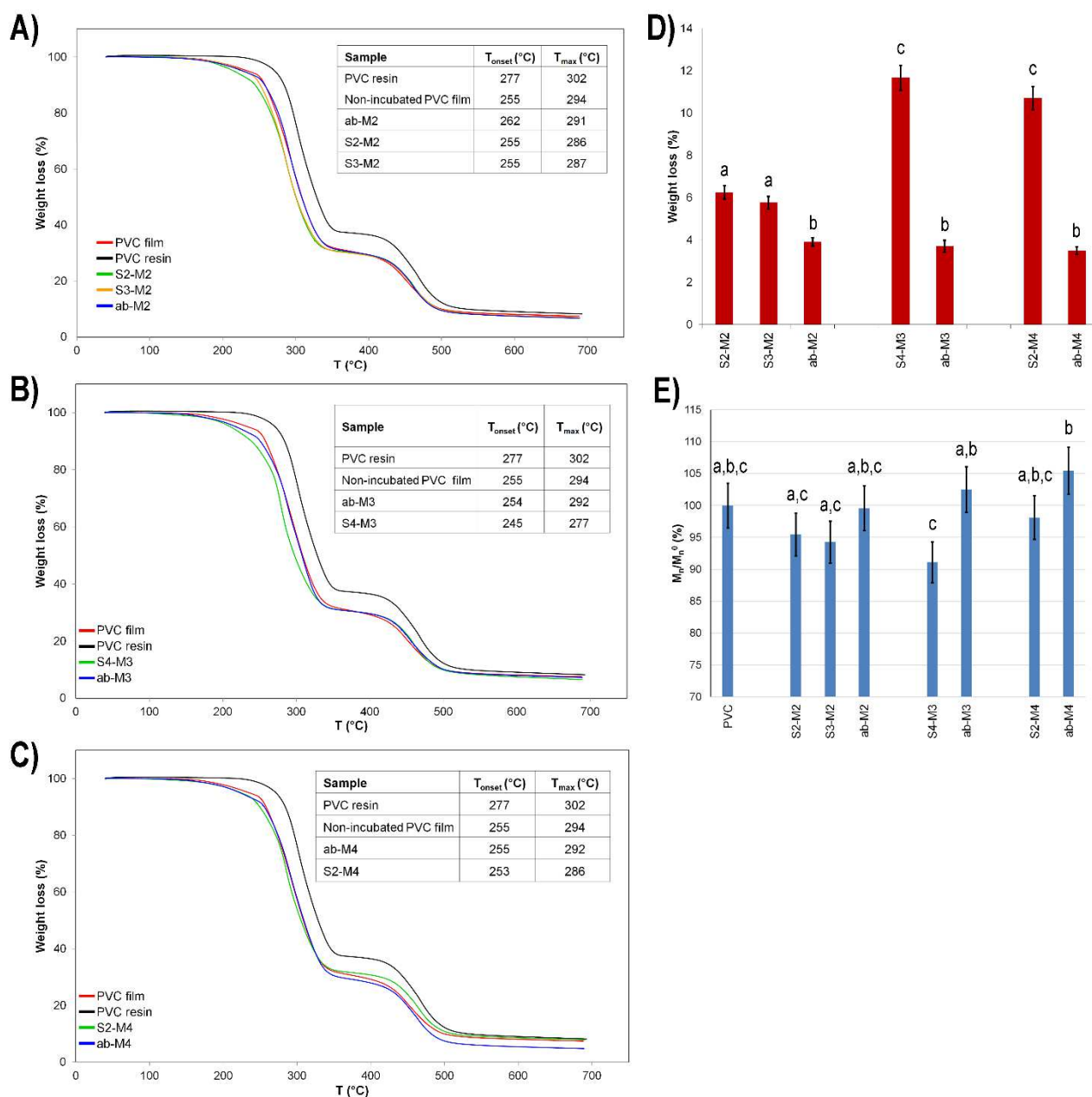
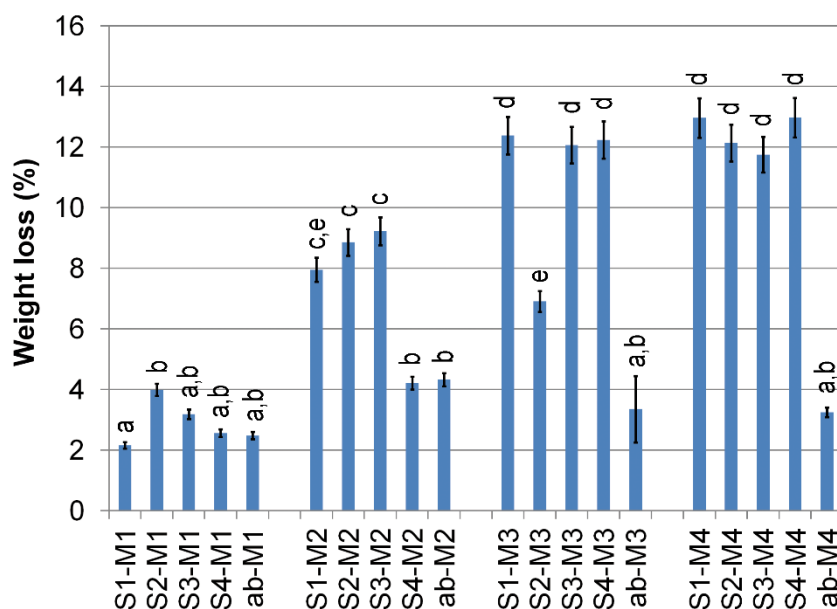


Figure 2. Chemical-physical analyses on PVC films withdrawn at month 7. TGA curves of PVC films incubated with the four different consortia enriched in M2 (A), M3 (B) and M4 (C) media or in the four corresponding abiotic controls. Gravimetric weight loss percentage of PVC films incubated with the four different consortia or in the four corresponding abiotic controls (D) (ANOVA: $p < 0.0001$; $F = 249.9$; $df = 6$). Number average molecular weight (Mn) of selected PVC films (ANOVA: $p = 0.0025$; $F = 5.42$; $df = 7$) (E). Mn (%) is defined as $(Mn/Mn_0) \times 100$, where Mn is the molecular weight at sampling time t and Mn₀ is the molecular weight of the non-incubated PVC film. Bars indicate standard deviation. Letters a, b, c show results of post-hoc analyses for film

657 gravimetric or molecular weight losses ($p < 0.05$), different letters indicate a significant statistical
 658 difference in gravimetric or molecular weight reduction values.

659



660

661 **Figure 3.** Gravimetric measurements at month 24. Weight loss (%) of PVC films incubated with
 662 the sixteen enriched consortia and the corresponding four abiotic controls. Bars indicate standard
 663 deviation (ANOVA: $p < 0.0001$; $F = 222.3$; $df = 19$). Letters a, b, c, d, e show results of post-hoc
 664 analyses for film gravimetric or molecular weight losses ($p < 0.05$), different letters indicate a
 665 significant statistical difference in gravimetric weight reduction values.

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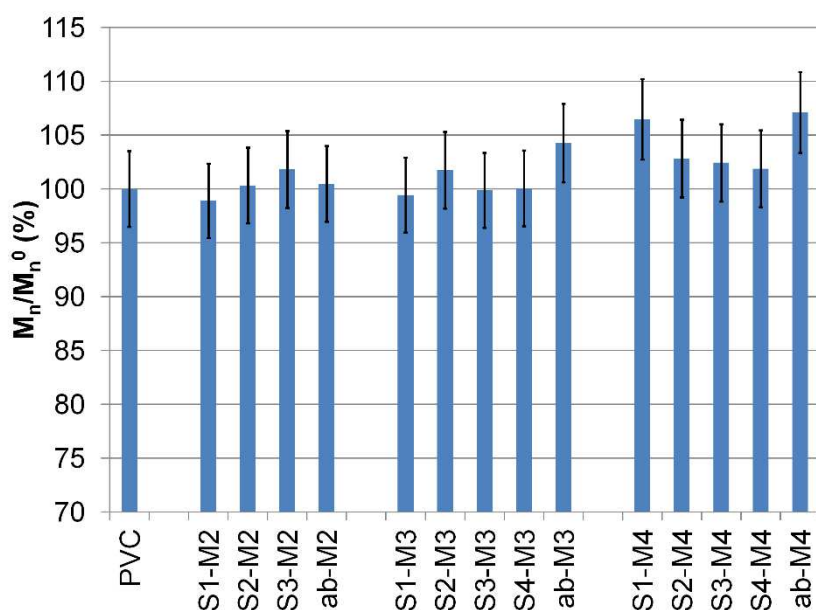


Figure 4. PVC mean molecular weight at month 24. GPC results for PVC films incubated with eleven different consortia enriched in M2, M3 and M4 media and their corresponding abiotic controls (ab-M1; ab-M2; ab-M3 and ab-M4) (ANOVA: $p = 0.19$; $F = 1.44$; $df = 14$).

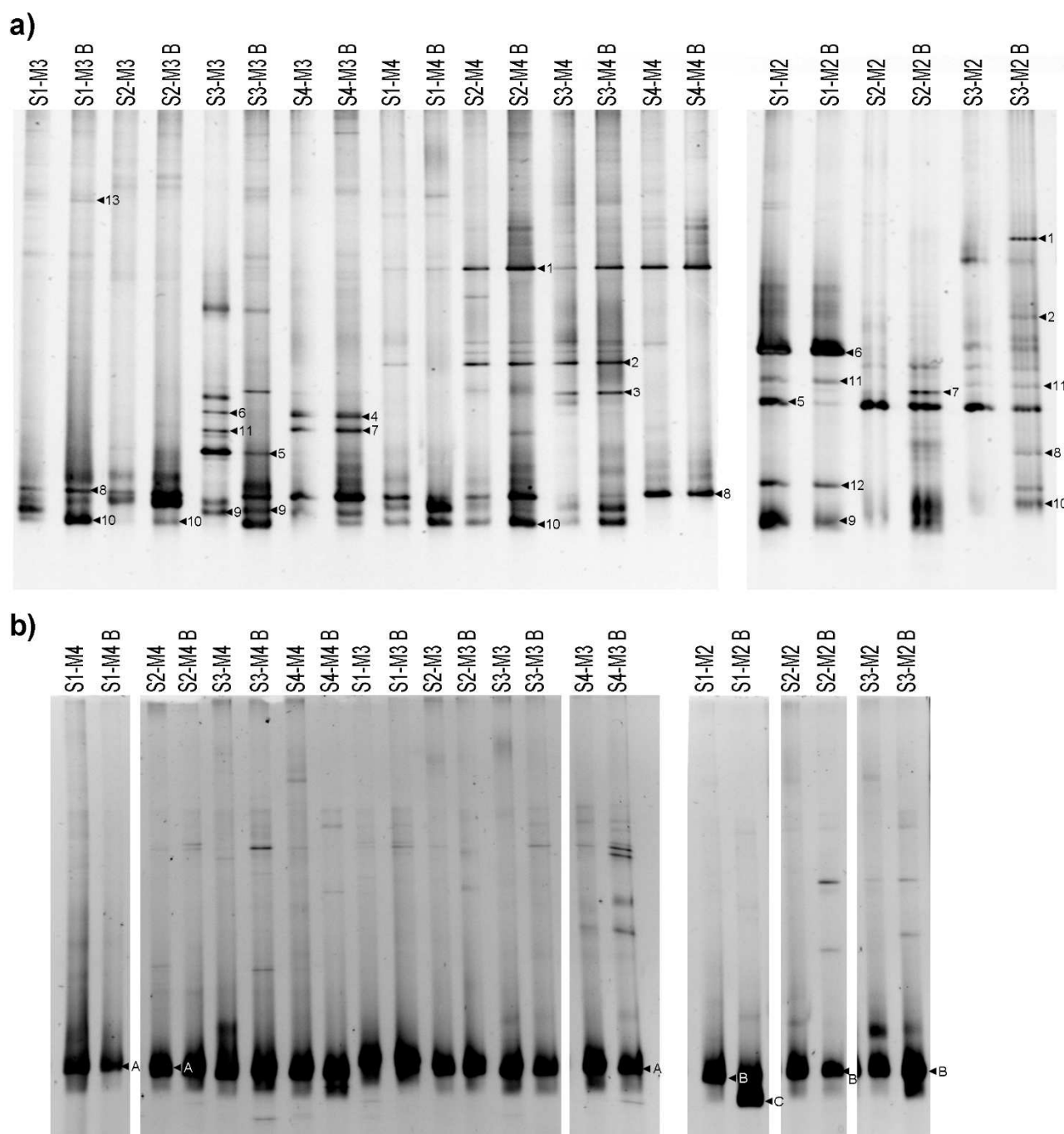
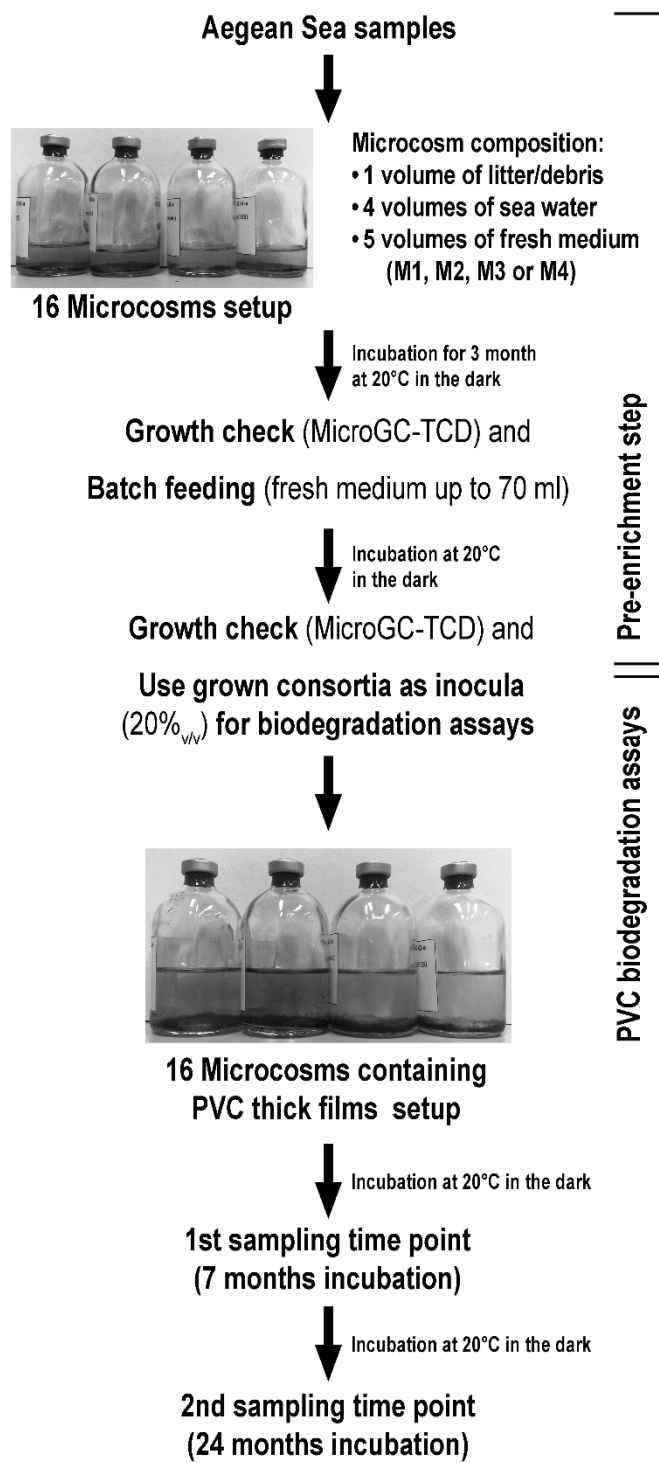


Figure 5. Bacterial (a) and archaeal (b) diversities of the 24 months PVC film-enriched anaerobic communities. DGGE profiles of 16S rRNA genes amplified from planktonic and biofilm microbial communities from the eleven different consortia showing high weight losses compared to the corresponding abiotic controls. Names over the lanes refer to the ID of the consortium. The letter B after consortia names stands for “biofilm community”. The identity of sequences of bands marked with arrows is given in Table 2 according to the band ID (1 to 13 and A to C).



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680 **Figure S1.** Experimental design. Schematic presentation of the experimental sequence for
 681 assessment of PVC film biodegradation.

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Supplementary material – S1

Supplementary material S1 contains the composition of the four media used to enrich the different marine anaerobic consortia and used to perform PVC biodegradation study.

Medium M1 was composed of (g/l): sodium formate, 2; Na₂CO₃, 0.1; (NH₄)₂SO₄, 0.5; mineral solution (per litre: KH₂PO₄, 18 g; NaCl, 18 g; CaCl₂•2H₂O, 0.53 g; MgCl₂•6H₂O, 0.4 g; MnCl₂•4H₂O, 0.2 g; CoCl₂•6H₂O, 0.02 g), 50 ml; acetic acid, 1.53 ml; vitamin solution (per litre: thiamine-HCl, 20 mg; Ca-D-pantothenate, 20 mg; nicotinamide, 20 mg; riboflavin, 20 mg; pyridoxine-HCl, 20 mg; biotin, 0.5 mg; folic acid, 0.25 mg; vitamin B₁₂, 0.2 mg), 5 ml; hemin solution (10 mg in 100 ml of 50% ethanolic), 5 ml; cysteine-sulphide solution (L-cysteine-HCl•H₂O, 2.5 g/l; Na₂S•9H₂O, 2.5 g/l; pH 11), 5 ml; pH 6.7 (Cacciari et al. 1993).

Medium M2 was composed of (g/l): glucose, 2; yeast extract, 1 g; tryptone, 1 g; beef extract, 0.5 g; K₂HPO₄, 1.5 g; MgCl₂•6H₂O, 0.21 g; FeSO₄•7H₂O, 0.1 g; trace element solution (per litre: MnSO₄•7H₂O, 0.01 g; ZnSO₄•7H₂O, 0.05 g; H₃BO₃, 0.01 g; CaCl₂•2H₂O, 0.01 g; Na₂MoO₄, 0.01 g; CoCl₂•6H₂O, 0.2 g), 10 ml; vitamin solution (per litre: L-ascorbic acid, 0.025 g; citric acid, 0.02 g; pyridoxine-HCl, 0.05 g; para-aminobenzoic acid, 0.01 g; D-biotin, 0.01 g; vitamin B₁, 0.02 g; riboflavin, 0.025 g) 1 ml; 5 %_{w/v} L-cysteine solution, 10 ml; pH 7.2-7.5.

Medium M3 was composed of (g/l): glucose, 2; KH₂PO₄, 0.5; NH₄Cl, 0.3; MgSO₄•7H₂O, 0.5; CaCl₂•2H₂O, 0.1; NaNO₃, 3; NaHCO₃ solution, 30 ml; vitamins solution, 1 ml; EDTA-chelated mixture of trace elements, 1 ml; selenite-tungstate solution, 1 ml; pH 7.2. All solutions composition were from Widdel and Bak (1992).

Medium M4 was composed per litre of distilled water: sodium lactate, 4.5 g; Na₂SO₄, 6 g; KH₂PO₄, 0.2 g; NH₄Cl, 0.25 g; MgCl₂•6H₂O, 0.4 g; KCl, 0.5 g; CaCl₂•2H₂O, 0.15 g; DSMZ SL-10 trace elements solution, 1 ml; selenite-tungstate solution (Widdel and Bak, 1992), 1 ml; mixed vitamin solution (per 100 ml of 10 mM sodium phosphate pH 7.1: p-aminobenzoic acid, 4 mg; biotin, 1 mg; nicotinic acid, 10 mg; D,L-Ca pantothenate, 5 mg; pyridoxine-HCl, 15 mg; thiamine-HCl, 10 mg), 1 ml; 0.005%_{w/v} vitamin B12 solution, 1 ml; 84 mg/l CaHCO₃ solution, 30 ml; 4%_{w/v} sulfide solution, 7.5 ml; pH 7.0-7.5.

711 **Tables**

712 **Table 1. TGA at month 24.** Results of TGA of PVC resin, PVC film (non-incubated) and PVC films incubated under abiotic conditions (ab-M2
713 to ab-M4) or with the consortia enriched in three media that showed lower thermal stability compared to abiotic controls.

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Sample	T _{onset} (°C)	T _{max} (°C)
PVC resin	277	302
PVC film	255	294
ab-M2	261	290
S2-M2	260	284
S3-M2	259	283
ab-M3	260	289
S4-M3	263	285
ab-M4	260	289
S1-M4	263	283
S2-M4	262	285
S4-M4	261	284

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Table 2. Identification of microorganisms characterizing the different PVC film associated communities according to DGGE profiles.

Band ID	Closest cultured relative (BLAST)	Accession No.	Identity (nt%)	Putative classification	No. positives ^a
<i>Bacteria</i>					
1	<i>Erysipelothrix</i> sp. (KX156777)	MH520751	95	<i>Erysipelotrichaceae</i>	5/11
2	Uncultured <i>CFB</i> group bacterium (FJ024711) *	MH520752	98	<i>CFB</i> group	3/11
3	<i>Bacteroidales</i> bacterium CF (CP006772)	MH520753	93	<i>Bacteroidetes</i>	2/11
4	<i>Psychromonas</i> sp. (NR116830)	MH520754	99	<i>Psychromonas</i>	1/11
5	<i>Cupriavidus</i> sp. (MG948149)	MH520755	100	<i>Cupriavidus</i>	4/11
6	<i>Pleomorphochaeta</i> sp. (NR134177)	MH520756	98	<i>Pleomorphochaeta</i>	4/11
7	<i>Sporobacter</i> sp. (KT183425)	MH520757	99	<i>Ruminococcaceae</i>	3/11
8	<i>Clostridium</i> sp. a-nd (FN397991)	MH520758	98	<i>Clostridiaceae</i>	9/11
9	<i>Dethiosulfovibrio</i> sp. (NR029034)	MH520759	100	<i>Dethiosulfovibrio</i>	3/11
10	<i>Acetobacterium</i> sp. (NR074548)	MH520760	98	<i>Acetobacterium</i>	8/11
11	<i>Cohaesibacter</i> sp. (KT324976)	MH520761	99	<i>Cohaesibacter</i>	2/11
12	<i>Desulfovibrio</i> sp. (KU892724)	MH520762	99	<i>Desulfovibrio</i>	1/11
13	<i>Fusibacter</i> sp. (KJ420408)	MH520763	96	<i>Fusibacter</i>	1/11
<i>Archaea</i>					
A	<i>Methanosaeta pelagica</i> (NR113571)	MH520764	99	<i>Methanosaeta</i>	8/11
B	<i>Methanococcoides methylutens</i> (CP009518)	MH520765	99	<i>Methanococcoides</i>	3/11
C	<i>Methanogenium</i> sp. (NR104730)	MH520766	99	<i>Methanogenium</i>	1/11

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725 ^a Number of samples positive for the presence of the specific band in DGGE analysis compared to total number of PVC plastic associated biofilm
726 communities analyzed.

727 * The sequence related to band #2 demonstrated a very low homology with cultivable bacteria, it was analyzed including the
728 uncultured/environmental sample sequences present in nucleotide databases.

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