

Prognostic Utility of the New Definition of Difficult-to-Treat Resistance Among Patients With Gram-Negative Bloodstream Infections

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Background. To compare the prognostic utility of the new definition of difficult-to-treat resistance (DTR) vs established definitions in a cohort of patients with Gram-negative bloodstream infections (GNBSIs).

Methods. This was a retrospective single-center study of adult patients with monomicrobial GNBSI, hospitalized from 2013 to 2016. DTR was defined as isolate demonstrating intermediate or resistant phenotype to all reported agents in the carbapenem, beta-lactam, and fluoroquinolone classes. Carbapenem resistance (CR) was defined according to 2015 Centers for Disease Control and Prevention criteria. Each isolate was further classified according to the Magiorakos et al. criteria as non-multidrug-resistant (non-MDR), MDR, extensively drug-resistant (XDR), or pan-drug-resistant (PDR). The primary outcome was all-cause 30-day mortality.

Results. Overall, 1576 patients were analyzed. Enterobacteriaceae accounted for 88.7% of BSIs, with *Escherichia coli* (n = 941) and *Klebsiella pneumoniae* (n = 326) being the most common pathogens. *Pseudomonas aeruginosa* was the most common nonfermentative bacteria (n = 130, 8.2%). Overall, 11% of strains were defined as DTR and 13% as CR. Episodes were further classified as non-MDR (68.8%), MDR (21.9%), XDR (8.8%), and PDR (0.4%). The prevalence rates of DTR, CR, and XDR were similar among Enterobacteriaceae and *Acinetobacter baumannii*, whereas they differed in *P. aeruginosa*. All the analyzed resistance definitions significantly improved prediction of 30-day mortality when introduced into a baseline multivariate model, to a similar degree: 9%, 10%, and 11% for DTR, Magiorakos, and CR definitions, respectively.

Conclusions. DTR seems a promising tool to identify challenging GNBSIs, mainly those due to *P. aeruginosa*. With the availability of new agents for CR infections, further multicenter assessments of DTR are needed.

Keywords. bloodstream infection; Gram-negative; carbapenem resistance; difficult-to-treat resistance; all-cause 30-day mortality.

Increasing rates of antibiotic resistance among Gram-negative bacteria have prompted investigators to analyze several issues concerning patients with severe infections due to these microorganisms.

In a majority of studies, the definition adopted for multidrug resistance was that proposed in 2008 by the US Centers for Disease Control and Prevention (CDC) and the European Centre for Disease Prevention and Control [1]. The authors defined 3 resistance phenotypes: multidrug resistance (MDR) as nonsusceptibility to ≥ 1 agent in ≥ 3 antimicrobial categories;

extensive drug resistance (XDR) as susceptibility limited to ≤ 2 categories; and pan-drug resistance (PDR) as nonsusceptibility to all agents in all antimicrobial categories [1]. Although epidemiologically useful, this definition has the limitation of weighing all antibiotics equally and only considering their in vitro activity, regardless of their “real-life” effectiveness and toxicity, limiting the bedside applicability of MDR and XDR categories. Indeed, MDR and XDR infections were not consistently associated with poorer patient outcomes in some studies [2, 3]; initial appropriate therapy has failed to improve the outcomes of patients with such drug resistance categories [4]. These issues are pivotal in designing and evaluating clinical trials on the therapeutic management of MDR Gram-negative infections [5].

Indeed, in a white paper from the Infectious Diseases Society of America (IDSA) on the conduct of clinical trials for the treatment of drug-resistant bacteria, the authors proposed a new concept of “extreme drug resistance” as an alternative to the Magiorakos XDR class. They defined XDR organisms as those resistant to all Food and Drug Administration (FDA)-approved,

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systematically active antibacterial agents except for those known to be substantially more toxic than or less efficacious than alternative agents [6]. For Gram-negative bacteria, they reported the example of strains resistant to all FDA-approved agents except for aminoglycosides, tigecycline, or colistin. In accordance with this concept, a new definition of resistance for Gram-negative infections has been recently proposed by Kadri et al. [7]. The authors defined difficult-to-treat resistance (DTR) as a treatment-limiting resistance to all firstline agents including all beta-lactams and fluoroquinolones [7]. This definition should reflect the use of second-line agents, such as those mentioned in the IDSA white paper, which are characterized by poorer pharmacokinetic properties and increased risk of toxicity, resulting in a better prediction of poor outcome. The 5-year prevalence of DTR, the associated risk factors, and the impact on in-hospital mortality rates were analyzed using a very large US cohort of patients with Gram-negative bloodstream infections (GNBSIs) [7]. However, in this study, the prevalence of DTR was very low and administrative data were used.

The aim of our study was to compare the prognostic utility of DTR vs established resistance definitions in a cohort of patients with GNBSIs hospitalized in a tertiary teaching hospital from an area with a high prevalence of antibiotic resistance.

METHODS

Study Design and Setting

We performed a retrospective cohort study of patients hospitalized at S. Orsola-Malpighi Hospital, a 1450-bed tertiary care university institute in Bologna, in the region of Emilia-Romagna (Northern Italy), from January 1, 2013, to December 31, 2016.

Patients were identified through microbiology databases. Clinical charts and hospital records were reviewed to gather study variables using a case report form (CRF) for up to 90 days after the index blood cultures (BCs). The accuracy of the data was systematically reviewed by a senior investigator before inclusion in the database.

Our ethics committee approved the study; informed consent was waived due to the retrospective noninterventive study design. Data were collected anonymously.

Participants

We included all adult (≥ 18 years) patients diagnosed with Gram-negative BSI, defined as ≥ 1 positive BC obtained from a patient suspected of having infection. Patients were considered only once at the time of the first episode (index BCs).

Patients were excluded from the analysis if they were found to have (i) a polymicrobial BSI, defined as growth of >1 micro-organism, excluding potential contaminants (ie, coagulase-negative staphylococci, *Corynebacterium* spp., *Propionibacterium* spp.); (ii) died within 72 hours of drawing the index BCs; or (iii) no clinical data available.

Variables and Definitions

For each GNBSI, we determined if the infection met the proposed definition for DTR according to Kadri et al. [7]. Specifically, DTR was defined as any GNBSI isolate demonstrating an intermediate or resistant phenotype to all reported agents in the carbapenem, beta-lactam, and fluoroquinolone categories (including additional agents when results were available). *Stenotrophomonas maltophilia*, which showed resistance to all tested antimicrobials (including TMP/SMX, levofloxacin, and minocycline), was also considered DTR.

Isolates were further classified according to the Magiorakos et al. criteria [1] as non-MDR, MDR, XDR, or PDR.

In addition, we categorized antibiotic class resistance as carbapenem resistance (CR), extended-spectrum cephalosporin resistance (ECR), and fluoroquinolone resistance (FQR) based on Centers for Disease Control and Prevention (CDC) surveillance definitions (<https://gis.cdc.gov/grasp/PSA/Downloads/AR-PhenotypeDefinitions.pdf>). Finally, beta-lactam/betalactamase inhibitor resistance (BL/BLI-R) was assessed according to European Committee for Antimicrobial Susceptibility Testing (EUCAST) criteria.

The primary outcome used to assess the prognostic significance of each resistance definition was 30-day mortality, defined as all-cause mortality within 30 days of the index BC [8].

We analyzed patient risk factors for their association with 30-day mortality, including age, sex, and underlying disease severity according to the Charlson comorbidity index [9]. Immunosuppression included neutropenia (neutrophil count $< 500/\text{mm}^3$), solid organ transplantation, hematopoietic stem cell transplantation, corticosteroid therapy at a dosage higher than or equivalent to prednisone 16 mg/day ≥ 15 days, and uncontrolled HIV infection ($< 200 \text{ CD4}/\text{mm}^3$).

BSI was classified according to the site of acquisition into nosocomial, health care-associated, and community-acquired using Friedman's criteria [10]. Clinical severity at infection onset was assessed according to updated sepsis definitions [11]. BSI sources were established according to CDC criteria [12]. In the absence of a recognized source, BSI was considered primary. BSI was defined as complicated when the infection source was not fully removable.

According to the causative species and susceptibility to carbapenems, etiologies were classified into 3 groups: (i) carbapenem-susceptible Enterobacteriaceae (CSE), (ii) carbapenem-resistant Enterobacteriaceae (CRE), and (iii) nonfermentative Gram-negative bacteria (NF-GNB).

Empirical therapy was defined as antibiotics administered before the susceptibility report was available. It was considered appropriate when at least 1 in vitro active drug (according to the susceptibility pattern of the isolate) was administered within 24 hours of drawing the index BC. Delayed or no active antibiotic administration within this period was considered inappropriate empirical therapy. Definitive antibiotic therapy was defined as

antibiotic treatment administered according to susceptibility results. Combination therapy was defined as a regimen including >1 anti-Gram-negative drug irrespective of relative in vitro activity. Duration of antibiotic treatment was defined as the number of consecutive days during which the patient received an appropriate antibiotic regimen. Source control was defined as the removal of the infection source within 7 days of index BC, including the performance of nonsurgical or surgical procedures to treat an obstructive focus, collection, or abscess at any site, including, among others, the urinary tract, biliary tract, and surgical site, and the removal of any device deemed the source of the BSI.

Microbiology

BCs were incubated using the BACTEC FX Automated Blood Culture System (Becton Dickinson, Franklin Lakes, NJ, USA). All positive BCs were processed with the Maldi Biotyper MALDI-TOF system (Bruker Daltonics, Bremen, Germany) for rapid and reliable species identification of microorganisms. Antimicrobial susceptibility testing of strains was performed using the Vitek 2 automated system (bioMérieux, Marcy l'Etoile, France). Enterobacteriaceae minimal inhibitory concentrations (MICs) were interpreted using EUCAST clinical breakpoints for all tested antibiotics.

Statistical Analysis

For the descriptive analysis, categorical variables were presented as absolute numbers and their relative frequencies. Continuous variables were presented as mean and standard deviation if normally distributed or as median and interquartile range (IQR) if non-normally distributed.

Univariate and multivariate analysis were performed to assess the relationship of study variables with 30-day all-cause mortality. First, categorical variables were compared using the χ^2 or Fisher exact test when appropriate, and continuous variables were compared using the Mann-Whitney *U* test. Then, significant and clinically relevant covariates identified in univariate analysis were introduced by a backward selection approach into a multivariable Cox regression survival model to ensure that all

correlations between predictors were considered, using a *P* cutoff of .05. Patients were considered from the day of BSI onset (index BCs) until death or day 30. The discrimination and calibration of the Cox regression model were then analyzed without any variable defining a resistance category of the bloodstream isolate (baseline mortality model), vs the addition of 1 of the following categories: (i) Magiorakos et al. classifications (non-MDR, MDR, XDR, and PDR); (ii) DTR; and (iii) CR definitions, described previously. All analysis was performed with STATA IC 13.1 (Stata Corp., College Station, TX, USA) using the STCOXCAL package to compare model calibration. Model discrimination was assessed by the Harrel C statistic and Net Reclassification Index (NRI) of each resistance definition with the baseline survival model [13].

RESULTS

According to the study criteria (Supplementary Figure 1), 1576 patients with a first episode of monomicrobial GN-BSI during the study period were analyzed. The median age (IQR) was 72 (59–82) years, and 55.7% were male. The general characteristics of the study population are reported in Supplementary Table 1.

Enterobacteriaceae accounted for 88.7% of BSIs, with 1259 carbapenem-susceptible (CSE) and 140 carbapenem-resistant (CRE) pathogens. *Escherichia coli* was the most common causative microorganism (59.7%), followed by *Klebsiella pneumoniae* (20.7%). *Pseudomonas aeruginosa* was the most common nonfermentative bacteria (8.2%).

Overall, 11% of strains were defined as DTR. Distribution of resistance categories was as follows: non-MDR 68.8%, MDR 21.9%, XDR 8.8%, and PDR 0.4%. The distribution of antibiotic class resistance was: FQR 46.6%, BL/BLIR 44.7%, ESCR 36.1%, and CR 13.1%. The prevalence of resistance categories and classes of antibiotic resistance varied across pathogens, as shown in Table 1.

Source control and appropriate empirical therapy were performed in 27.3% and 68% of cases, respectively. In both

Table 1. Prevalence of Resistance Among the Main Gram-Negative Species According to the Analyzed Definitions

	<i>E. coli</i> (n = 941), No. (%)	<i>K. pneumoniae</i> (n = 326), No. (%)	<i>P. aeruginosa</i> (n = 130), No. (%)	<i>A. baumannii</i> (n = 33), No. (%)
Resistance categories				
Non-MDR	742 (78.9)	133 (40.8)	111 (85.4)	11 (33.3)
MDR	197 (20.9)	69 (21.2)	19 (14.6)	3 (9.1)
XDR	2 (0.2)	117 (35.9)	0	19 (57.6)
PDR	0	7 (2.1)	0	0
Antibiotic class resistance				
BL/BLIR	371 (39.5)	215 (66)	37 (38.5)	NA
ECR	296 (31.5)	207 (63.5)	26 (20)	NA
CR	1 (0.1)	140 (42.9)	36 (27.7)	22 (66.7)
FQR	438 (46.5)	198 (60.7)	33 (25.4)	22 (66.7)
New definition				
DTR	1 (0.1)	138 (42.3)	10 (7.7)	22 (66.7)

Abbreviations: BL/BLIR, betalactam/betalactamase inhibitor resistance; CR, carbapenem resistance; DTR, difficult-to-treat resistance; ECR, extended-spectrum cephalosporin resistance; FQR, fluoroquinolone resistance; MDR, multidrug resistance; PDR, pandrug resistance; XDR, extensive drug resistance.

Table 2. Univariate Analysis of Risk Factors for All-Cause 30-Day Mortality

	Survivors (n = 1412), No. (%)	Nonsurvivors (n = 164), No. (%)	p
Demographics			
Age, median (IQR), y	72 (59–82)	72 (62–83)	.20
Male sex	782 (55.4)	96 (58.5)	.46
Comorbidities			
Charlson index, median (IQR)	6 (4–8)	6.6 (4.5–8.8)	.003
Immunosuppression	292 (20.7)	40 (24.4)	.31
Ward of admission			
Medical	1136 (80.5)	110 (67.1)	<.001
Surgical	181 (12.8)	23 (14)	
ICU	95 (6.7)	31 (18.9)	
Site of BSI acquisition			
Community-acquired	415 (29.4)	22 (13.4)	<.001
Health care-associated	249 (17.6)	32 (19.5)	
Hospital-acquired	748 (53)	110 (67.1)	
CRE carrier at BSI onset			
	150 (10.6)	33 (20.1)	.001
Clinical severity at BSI onset			
SOFA, median (IQR)	3 (1–5)	5 (3–7)	<.001
Septic shock	98 (6.9)	45 (27.4)	<.001
Source of BSI			
Undefined	265 (18.8)	33 (20.1)	.75
Urinary tract	560 (39.7)	33 (20.1)	<.001
Biliary tract	205 (14.5)	19 (11.6)	.35
Intra-abdominal	170 (12)	25 (15.2)	.26
Lower respiratory tract	99 (7)	24 (14.6)	.001
CVC-related	71 (5)	19 (11.6)	.001
Complicated BSI	358 (25.4)	53 (32.3)	.06
Etiology			
<i>Escherichia coli</i>	881 (62.4)	60 (36.6)	<.001
<i>Klebsiella pneumoniae</i>	280 (19.8)	46 (28)	.01
<i>Enterobacter</i> spp.	70 (5)	7 (4.3)	.71
<i>Proteus</i> spp.	41 (2.9)	14 (8.5)	.001
<i>Pseudomonas aeruginosa</i>	107 (7.6)	23 (14)	.007
<i>Acinetobacter baumannii</i>	21 (1.6)	10 (6.1)	.001
<i>Stenotrophomonas maltophilia</i>	10 (0.7)	4 (2.4)	.05
Etiology category			
CSE	1163 (82.4)	96 (58.5)	
CRE	109 (7.7)	31 (18.9)	
NF-GNB	140 (9.9)	37 (22.6)	
Resistance categories^a			
Non-MDR	1005 (71.2)	80 (48.8)	<.001
MDR	295 (21)	50 (30.5)	
XDR	107 (7.6)	32 (19.5)	
PDR	5 (0.4)	2 (1.2)	
Antibiotic class resistance^a			
ECR	487 (34.5)	82 (50)	<.001
BL/BLIR	612 (43.3)	93 (56.7)	<.001
CR	154 (10.9)	53 (32.3)	<.001
FQR	626 (44.3)	109 (66.5)	<.001
New definition			
DTR	129 (9.1)	45 (27.4)	<.001
Therapeutic management			
Source control	386 (27.3)	45 (27.4)	1
Appropriate empirical therapy	990 (70.1)	85 (51.8)	<.001

Abbreviations: BL/BLIR, betalactam/betalactamase inhibitor resistance; BSI, bloodstream infection; CR, carbapenem resistance; CRE, carbapenem-resistant Enterobacteriaceae; CVC, central venous catheter; DTR, difficult-to-treat resistance; ECR, extended-spectrum cephalosporin resistance; FQR, fluoroquinolone resistance; ICU, intensive care unit; IQR, interquartile range; MDR, multidrug resistance; PDR, pandrug resistance; SOFA, sequential organ failure assessment; XDR, extensive drug resistance.

^aResistance categories were mutually exclusive, whereas antibiotic class resistances were not.

the empirical and definitive treatment cohorts, the antibiotic classes most commonly used were BL/BLI and carbapenems. Combination therapy was administered in 16.2% and 21.6% of empirical and definitive regimens, respectively (data shown in [Supplementary Table 1](#)).

All-cause 30-day mortality was 10.4%, with 5% of patients later presenting with a BSI relapse within 90 days after index BCs.

Compared with patients who were alive at day 30 ([Table 2](#)), nonsurviving patients exhibited higher Charlson index scores and were more likely to be in the ICU at BSI onset or have a hospital-acquired BSI. Nonsurviving patients also exhibited higher SOFA scores and higher rates of septic shock, nonurinary infection sources, and etiologies other than *E. coli*. In terms of therapeutic management, only appropriate empiric therapy was significantly associated with a lower mortality rate. All resistance definitions were associated with significantly higher 30-day mortality rates by univariate analysis ([Supplementary Figure 2](#)).

At multivariate analysis, the independent risk factors for all-cause 30-day mortality were Charlson index, SOFA score, septic shock, CVC-related BSI, BSI due to CRE or NF-GNB, and complicated BSI, whereas urinary source, source control, and active empiric therapy were protective factors ([Table 3](#)). The predicted impact of Magiorakos (non-MDR, MDR, XDR, and PDR), CR, and DTR definitions on 30-day survival adjusted for significant survival covariates is shown in [Figure 1](#). The impact of DTR was also analyzed, including the 19 patients who died within 72 hours of index BC for whom clinical data were available, without observing different results ([Supplementary Figure 3](#)).

Incorporation of the resistance definitions into the baseline mortality model significantly improved discrimination of the multivariate model for predicting 30-day mortality; the net reclassification improvement was 9%, 10%, and 11% for DTR, Magiorakos et al., and CR definitions, respectively

([Supplementary Table 2](#)). Similarly, calibration of the baseline mortality risk model was improved with inclusion of each resistance definition, particularly for predicted 30-day mortality risk >20%, as shown in [Figure 2](#).

DISCUSSION

We analyzed the prevalence of the new proposed definition of antibiotic resistance for Gram-negative bacteria and difficult-to-treat resistance in a cohort of 1576 patients with monomicrobial GN-BSI. In addition, we compared DTR with the previously proposed definitions of Magiorakos et al. and CR according to the 2015 CDC criteria. In our study, the prevalence of DTR was 11%. It varied across species and was highest among *K. pneumoniae* and *A. baumannii* BSIs. In these pathogens, CR and DTR rates were comparable, whereas they differed in *P. aeruginosa*. Specifically, DTR seemed to identify better than CR and XDR categories the cases of *P. aeruginosa* with limited treatment options. All the analyzed definitions significantly improved the prediction of 30-day mortality to a similar degree when introduced into a baseline mortality prediction model.

In the daily practice, DTR and CR definitions offer some important advantages over Magiorakos criteria as (i) being easier to establish; (ii) providing more descriptive information that enhances pathogen-directed treatment; and (iii) capturing excess mortality attributable to both discordant empirical regimens and subsequent reliance on less effective and/or more toxic compounds (eg, colistin, tigecycline, and aminoglycosides).

Some authors have observed that CR, when appropriately applied, encompasses most DTR Gram-negative infections, providing useful information for guiding therapy [14]. This was confirmed in our study for Enterobacteriaceae and *A. baumannii* but not for *P. aeruginosa*. Unfortunately, the low number of *P. aeruginosa* BSIs limited our ability to analyze the prognostic significance of DTR in this subgroup. In addition, our epidemiology and the therapeutic approach to CR infections during the study period could have influenced our results. Indeed, with the introduction of new drugs for CR infections, the predictive value of CR has been changing [15]. This fact underlines a strength of a new definition: “DTR is not a fixed phenotype but rather a flexible framework” [16]. Indeed, the authors who proposed this definition recognized the need to periodically revise the rubric of firstline, high-efficacy, and low-toxicity agents in order to continue to capture, with the DTR definition, how resistance is perceived and confronted at the bedside.

The extreme drug resistance and DTR concepts were primarily developed to design clinical trials on new antibacterial agents for drug-resistant infections [6]. Resistance to all firstline drugs should reflect excess mortality attributable not only to initial inappropriate therapy, but also to the use of alternative drugs with suboptimal pharmacokinetic/pharmacodynamic (PK/PD) profiles and greater toxicity [17]. Indeed, in

Table 3. Multivariate Analysis of Risk Factors for All-Cause 30-Day Mortality

Covariate	aHR (95% CI)	P
Charlson comorbidity score	1.12 (1.06–1.18)	<.001
Septic shock	2.91 (1.81–4.70)	<.001
SOFA score	1.12 (1.07–1.18)	<.001
Urinary tract source	0.64 (0.42–0.96)	<.03
CVC-associated infection	2.19 (1.27–3.79)	.005
Etiology category		
CSE	Reference	
CRE	1.95 (1.26–3.02)	.003
Nonfermentative	2.43 (1.59–3.73)	<.001
Complicated BSI	2.02 (1.26–3.24)	.003
Source control	0.38 (0.23–0.65)	<.001
Active empiric therapy	0.68 (0.49–0.95)	.02

Abbreviations: aHR, adjusted hazard ratio; BSI, bloodstream infection; CI, confidence interval; CSE, carbapenem-susceptible Enterobacteriaceae; CRE, carbapenem-resistant Enterobacteriaceae; CVC, central venous catheter; NFGN, nonfermentative Gram-negative; SOFA, sequential organ failure assessment.

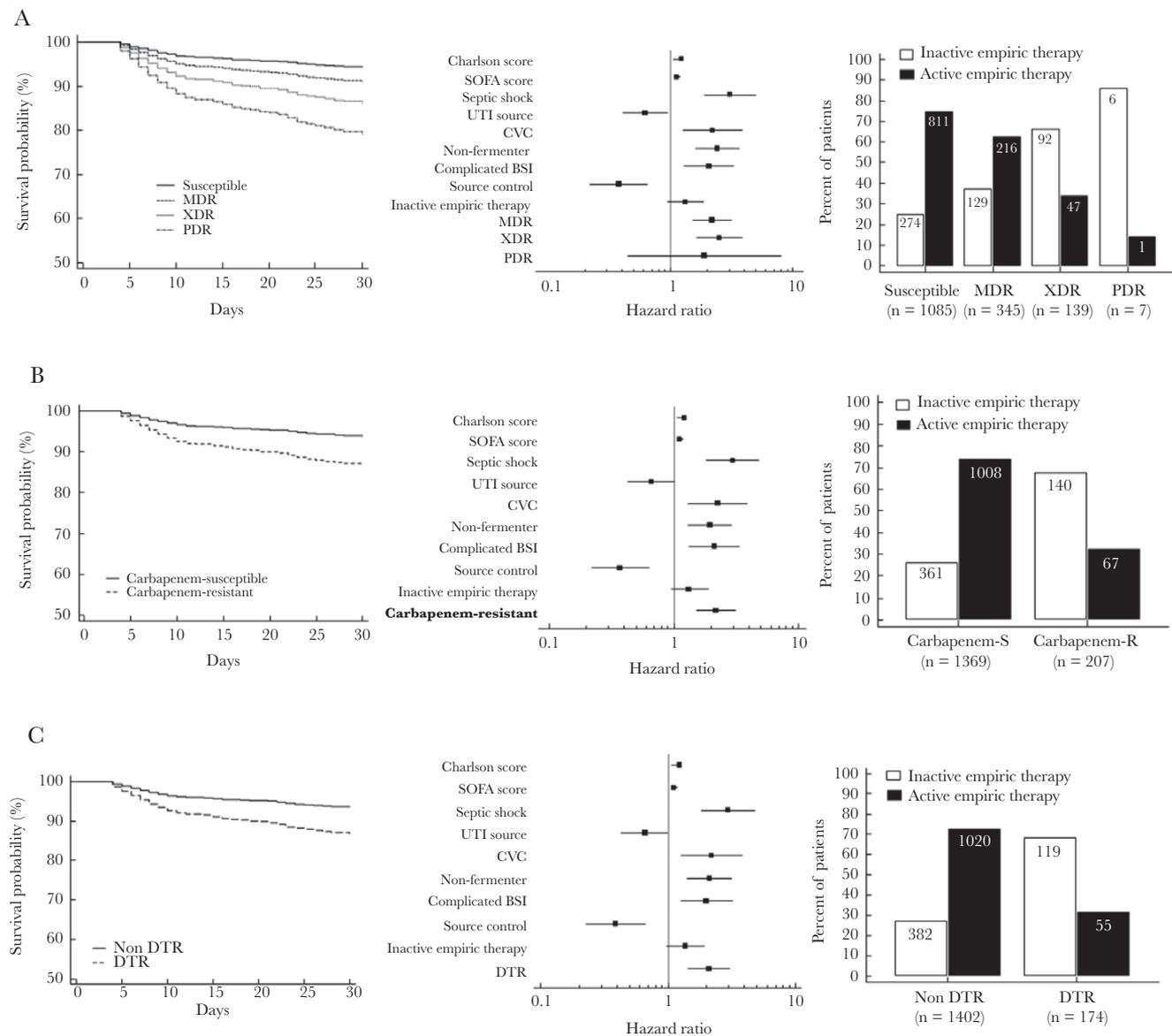


Figure 1. Survival curves for different resistance categories and forest plots according to Cox multivariate analysis of risk factors for all-cause 30-day mortality. The baseline model included the following resistance definitions: Magiorakos criteria (A); carbapenem resistance (B); difficult-to-treat resistance (C). Numbers and rates of active and inactive therapy for each category according to each resistance definition are shown. Abbreviations: BSI, bloodstream infection; CVC, central venous catheter; DTR, difficult-to-treat resistance; MDR, multidrug resistance; PDR, pandrug resistance; SOFA, sequential organ failure assessment; UTI, urinary tract infection; XDR, extensive drug resistance.

our basic model for mortality prediction, active empiric therapy was an independent protective factor, along with source control. However, when drug resistance categories were added to the model, the association between initial appropriate therapy and mortality was no longer significant, whereas source control remained a strong protective factor. It is worth noting that our multivariate analysis focused solely on appropriate empiric therapy; that is, it is still possible to receive inappropriate empiric therapy even in patients infected with susceptible isolates. Indeed, in our analysis, we found that nearly 25% of patients with “susceptible” Gram-negative pathogens did not receive

appropriate empirical therapy, thus providing one explanation of how MDR or DTR resistance definitions could be retained simultaneously in a multivariate model adjusted for inappropriate therapy. Another possible explanation is that both resistance categories and in vitro active therapy do not take into account eventual drug exposure in real life. These considerations underline the need to determine local microbiology and optimize dosing schedules to improve the rates of appropriate empiric therapy and patient survival.

Our study has several limitations. The single-center design could limit the generalizability of our results. However, this is

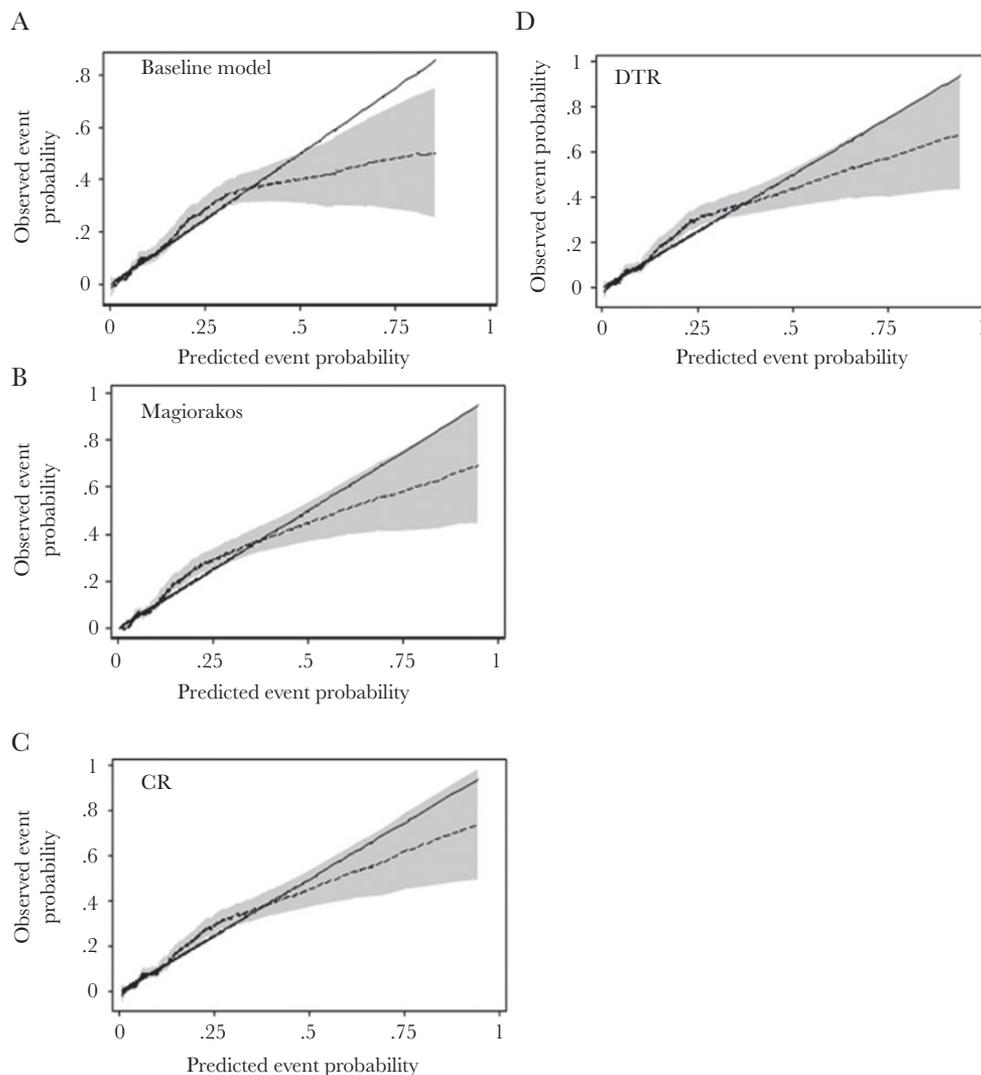


Figure 2. Calibration of 30-day mortality risk model by resistance definition. Smoothed pseudo-values (dashed lines) with pointwise 95% confidence intervals (shaded area) are plotted against predicted 30-day mortality probabilities. The solid line is the line of identity, denoting perfect calibration. Some miscalibration is evident with all models at predicted probabilities >0.5. A, Baseline mortality model without susceptibility categories. B, Magiorakos et al. definitions. C, Carbapenem resistance. D, Difficult-to-treat resistance. Abbreviations: CR, carbapenem resistance; DTR, difficult-to-treat resistance;

the first validation of the DTR definition in a Southern European country, where the prevalence and impact on mortality of antibiotic resistance are much higher than in the population used to develop the definition [18]. In addition, in US studies, both urban and rural hospitals were included, diluting the prevalence and impact of DTR. Our cohort is from a large tertiary teaching hospital, reflecting the complexity and epidemiology of patients managed in similar institutions from our area. The retrospective collection of patient and microbiological data could have limited integrity and accuracy. However, a senior investigator revised all CRFs and reconciled data reports and missing data with medical records before including information in the database. This approach ensured that patient-level data were accurate and of high quality, whereas in prior studies clinical information was mainly obtained from administrative data [7, 19]. Most

antibiotic susceptibility data were generated by an automated system (Vitek 2) that could have over- or underestimated MICs for some antibiotics in some episodes. However, this reflects real life, as in most hospitals physicians establish treatment on the basis of laboratory results generated using similar methods. Finally, clinical competency can contribute to the outcomes of patients with drug-resistant GN-BSI. However, this was not systematically assessed in our retrospective analysis.

To conclude, this is the first validation of DTR in a large non-US cohort of patients with GN-BSI. DTR seems a promising tool to identify challenging cases, mainly among patients with *P. aeruginosa* BSI. However, due to the high prevalence of CR in our study, mainly among *K. pneumoniae* and *A. baumannii*, the CR category was associated with the highest net reclassification improvement in predicting mortality. Further studies assessing

the value of DTR in multinational European cohorts, considering also newly available drugs for CR infections, are encouraged.

Supplementary Data

Supplementary materials are available at *Open Forum Infectious Diseases* online. Consisting of data provided by the authors to benefit the reader, the posted materials are not copyedited and are the sole responsibility of the authors, so questions or comments should be addressed to the corresponding author.

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Author contributions. M.G.: study design, data analysis, and drafting the manuscript; L.B.: data collection and support in data analysis and drafting the manuscript; R.P.: revision of collected data; M.B.: revision of collected data; M.M.: data collection; L.P.: data collection; A.T.: data collection; G.F.: data collection; L.M.: support in data collection and analysis; S.A.: support in collection and revision of microbiological data; R.L.: data analysis and drafting the manuscript; P.V.: study design and manuscript revision.

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