

SUPPLEMENTARY DATA

NSUN2 introduces 5-methylcytosines in mammalian mitochondrial tRNAs

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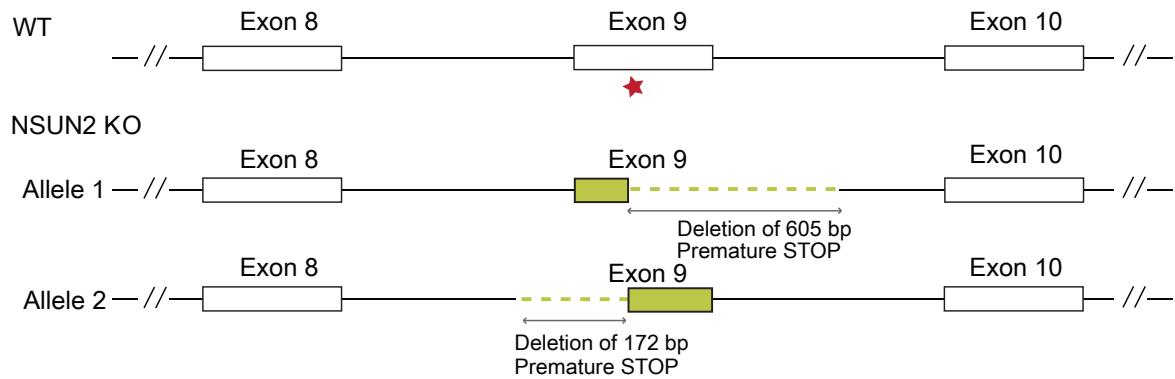
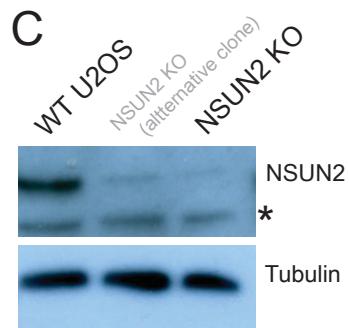
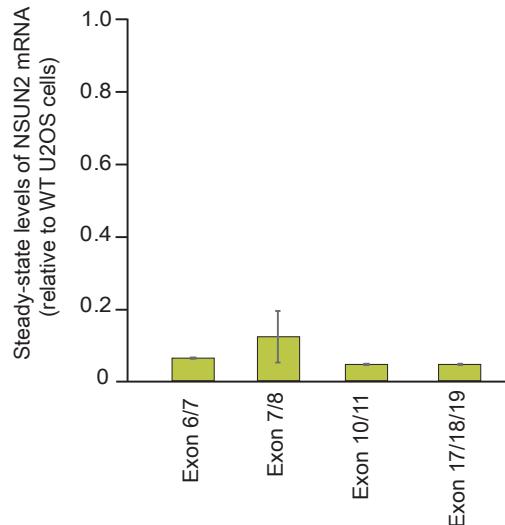
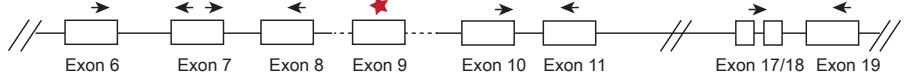
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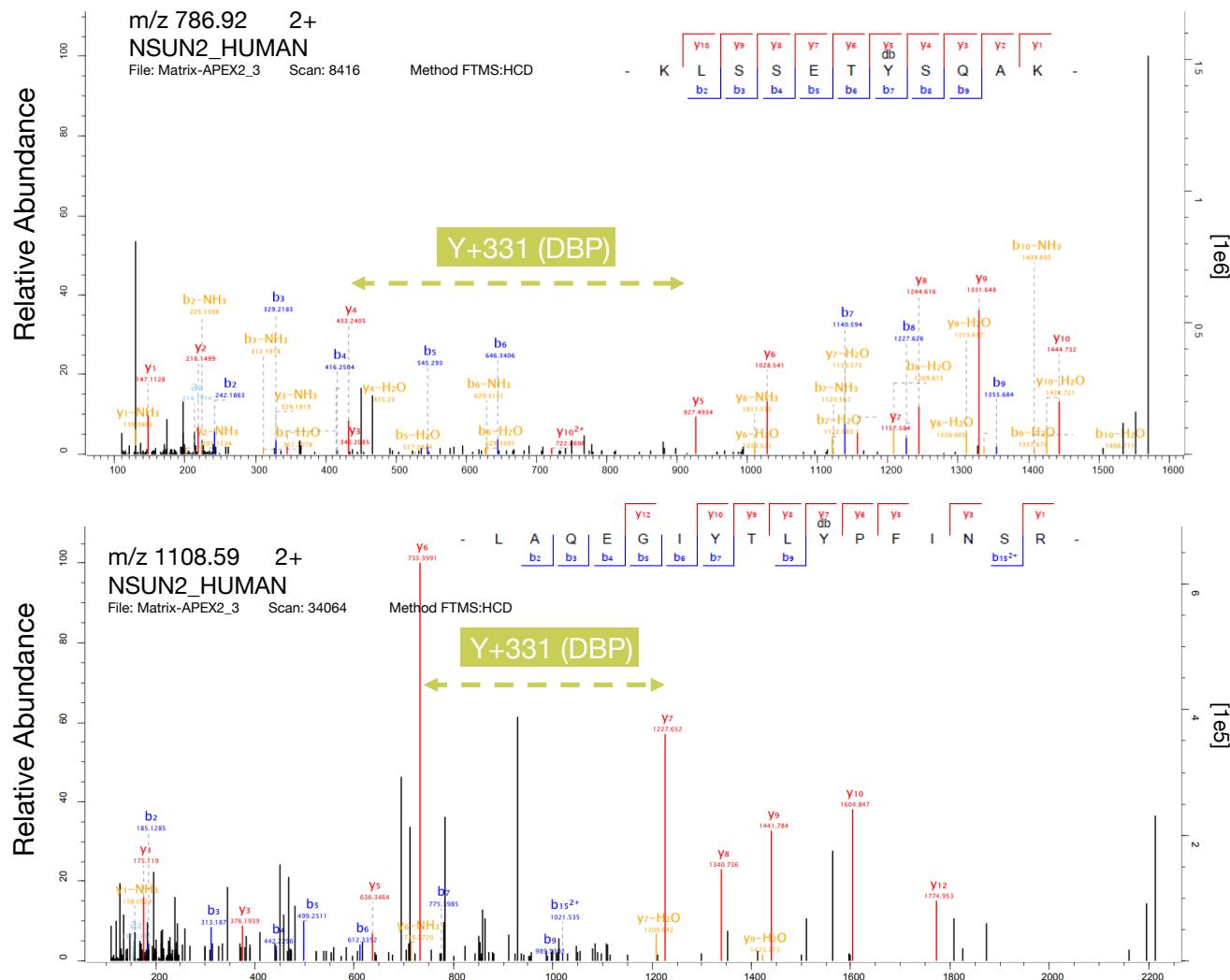
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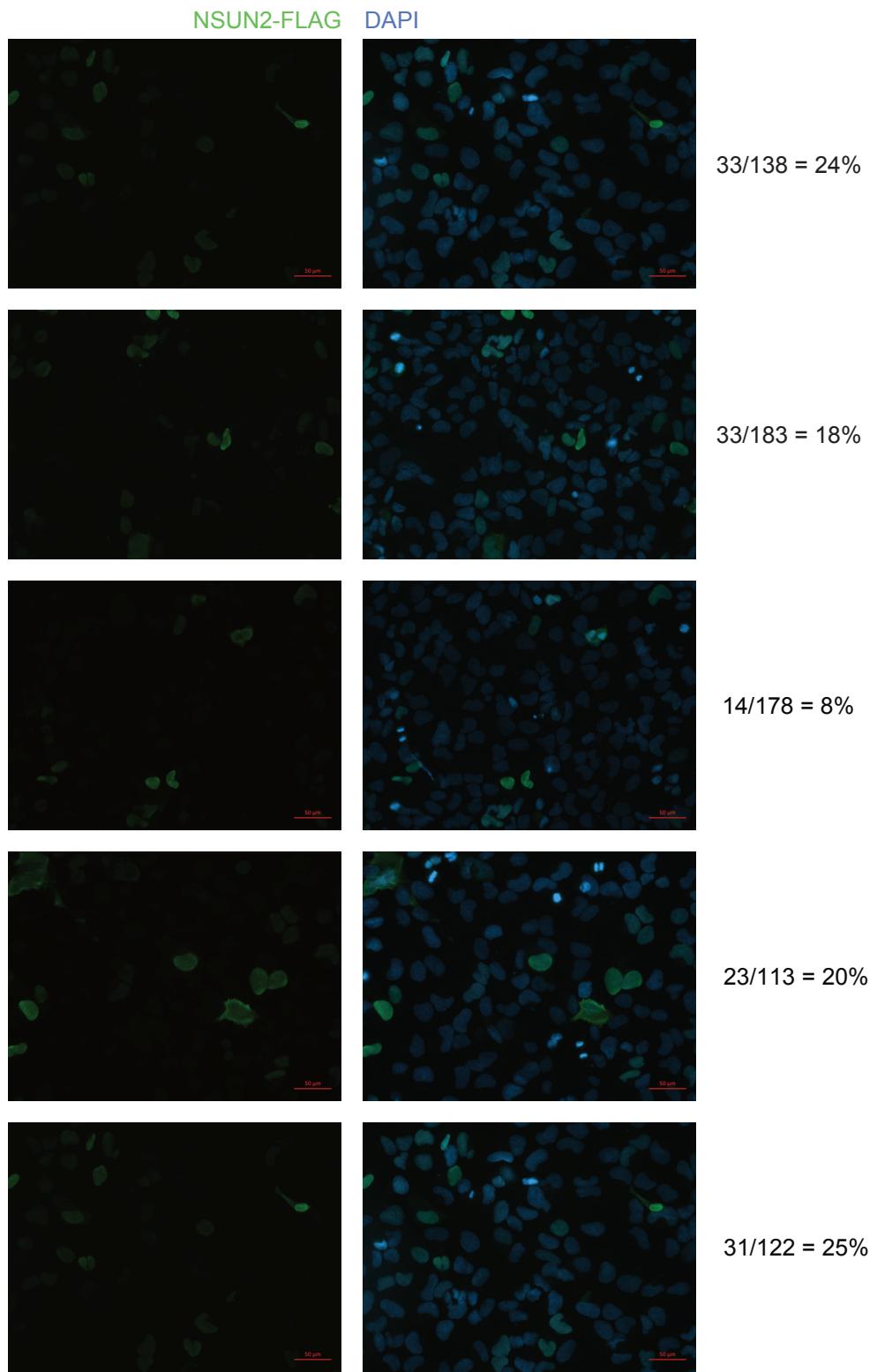
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Supplementary Figure S1 | Generation and characterisation of NSUN2 CRISPR/Cas9 KO cells

(A) Schematic representation of a fragment of the NSUN2 gene structure spanning exons 8 to 10. Red star indicates the site in exon 9, which was targeted by the CRISPR/Cas9 gRNAs (**Supplementary Table S1**). Genomic changes in the U2OS NSUN2 KO cell line as detected by PCR and Sanger sequencing are indicated in green and described. **(B)** *Top*: the NSUN2 gene structure spanning exons 6 to 19 with indicated RT-qPCR primer binding sites. *Bottom*: RT-qPCR analysis of NSUN2 mRNA expression levels in the NSUN2 KO cells for 4 pairs of primers. The expression was normalised to GAPDH. n=2. **(C)** Western blot analysis of the steady-state levels of NSUN2 in U2OS WT and NSUN3 KO cell line. Note: alternative CRISPR/Cas9 KO line is shown, which was not used in the present study.



Supplementary Figure S2 | Detailed MS/MS spectra of NSUN2 DBP-peptides.
Spectra for KLSSETYSQAK and LAQEGIYTLYPFINSR (DBP-modified Y underlined)
labelled by mitochondrial matrix-targeted APEX2.



Supplementary Figure S3 | Transient expression of NSUN2 in human NSUN2 KO cells

Representative images of immunofluorescence labelling after transient expression of a NSUN2.FLAG.STREP2 construct in human NSUN2 KO cells. Cells were stained for FLAG (green) and DAPI (blue). Scale bar: 50 μ M. Average percentage of transfected cells was 18 %.

Supplementary Table S1 | List and sequences of gRNA used to generate human NSUN2 KO line

gRNA name	gRNA sequence
gRNA_C321A_1_fwd	pACCGCAATCCGCAGCTGTAAGCTA
gRNA_C321A_1_rev	pAAACTAGCTTACAGCTGCGGATTG
gRNA_C321A_2_fwd	pACCGGTGTTCACTAAACCCTATTG
gRNA_C321A_2_rev	pAACCCAATAGGGTTAGTGAACAC

Supplementary Table S2 | List of oligonucleotides used in this study

Purpose	5' to 3' sequence	
Primers used for cloning into pcDNA5-FST2		
NSUN2 start	forward	GACGGTACCATGGGCGGGCGGTGCGGGG
NSUN2 stop	reverse	GTCCTCGAGCCGGGTGGATGGACCCC
qPCR primers for DNA analysis		
mt-CO1	forward	TGCTAGCCGCAGGCATTACT
	reverse	CGGGATCAAAGAAAGTTGTGTTT
RNaseP	forward	GCCTACACTGGAGTCCGTGCTACT
	reverse	CTGACCACACACGAGCTGGTAGAA
qPCR primers for RT-qPCR		
NSUN2 ex17-18	forward	AAGCAAAGGACCTGGCAAAG
	reverse	CAGCCCCATCATCCTGAGAT
NSUN2 ex10	forward	AGTGGATGCCTGGAATCACA
	reverse	GGGAACATGGTAGGTCGGAT
NSUN2 ex11	forward	CTGGCTCAAAGACCACACAG
	reverse	GTTGACCACCATGATGCAGG
NSUN2 ex6	forward	CCTGCATCATGGTGGTCAAC
	reverse	TCTCATAGTGCCGTCTCCAC
NSUN2 ex7	forward	
	reverse	
NSUN2 ex7	forward	
	reverse	
NSUN2 ex8	forward	
	reverse	

Supplementary Table S2 - continued**Targeted RNA BS-seq primers human**

hMT-TS2 RT primer		TAAAAAAACCATTATTAAACA
hMT-TS2 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGGTTATAAGAATTGTTAATTATG
hMT-TS2 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGTAAAAAAACCATTATTAAACA
hMT-TM RT primer		TTAGTTAATAAGTTATTGGGT
hMT-TM 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGTTAGTTAAATAAGTTATTGGGT
hMT-TM 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGAACCAACATTTCAAATA
hMT-TH RT primer		TAAAAATCATAAACCTC
hMT-TH 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGTTAAATATTAGATTGTGAATTG
hMT-TH 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGAACAAAAATCATAAACCTC
hMT-TL1 RT primer		TATTAAAAAAAAAAATTAAACCTC
hMT-TL1 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGGTAATTGTATAAAATTAAAAT
hMT-TL1 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGTATTAAAAAAAATTAAACCTC
hMT-TE RT primer		TATTCTCACACAAACTACAACCA
hMT-TE 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGAAATATAATGATGGTTTTATA
hMT-TE 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGTATTCTCACACAAACTACAACCA
hMT-TF RT primer		TATTATAAAATAATATAAACC
hMT-TF 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGTAAAGTAATATATTGAAATGTT
hMT-TF 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGTATTATAAAATAATATAAACC
hMT-TY RT primer		ATAATAAAAAAAAAACCTAACCC
hMT-TY 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGGTTGAGTGAAGTATTGGATTGAA
hMT-TY 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGATAAAAAAAACCTAACCC

Supplementary Table S2 - continued**Targeted RNA BS-seq primers mouse**

mMT-TH RT primer		AATAAATAAAAAAATTATTC
mMT-TH 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGAAAATTAGATTGTGAATTG
mMT-TH 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGAATAAATAAAAATTATTC
mMT-TL1 RT primer		TATTAAGAAAATTAAACCTC
mMT-TL1 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGTGTGAAGATTAAAATTGT
mMT-TL1 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGTATTAAGAAAATTAAACCTC
mMT-TL2 RT primer		TACTTTATTAAATTACACCA
mMT-TL2 1 stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGATAATAGTAATTATTGGTTTAGGA
mMT-TL2 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGTACTTTATTAAATTACACCA
mMT-TY RT primer		TAATAAAAAAAATTAAACCTC
mMT-TY 1 stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGGAGTAAGTATTAGATTGTAAT
mMT-TY 1 stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGTAATAAAAAAAATTAAACCTC
mMT-TN RT primer		CTAAATTAAACAAAATTAAACCTA
mMT-TN 1 stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGTAATAGGGTATTAGTTGTTAA
mMT-TN 1 stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGCTAAATTAAACAAAATTAAACCTA
mMT-TS2 RT primer		TAAAAAACCATATTAAACA
mMT-TS2 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGTTGTAAAGATTGTTATTATG
mMT-TS2 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGTAAAAAACCATATTAAACA
mMT-TE RT primer		TATTTCTACACAAACATTCAACTA
mMT-TE 1stage PCR primer with overhang	forward	TCGTCGGCAGCGTCAGATGTGTATAAGAGACAGTGTGATTTTTATGTTATTGG
mMT-TE 1stage PCR primer with overhang	reverse	GTCTCGTGGGCTCGGAGATGTGTATAAGAGACAGTATTCTACACAAACATTCAACTA