

Review

Marine Sponge Natural Products with Anticancer Potential: An Updated Review

Cinzia Calcabrini ^{1,†}, Elena Catanzaro ^{1,†}, Anupam Bishayee ^{2,*}, Eleonora Turrini ¹ 
and Carmela Fimognari ^{1,*}

¹ Department for Life Quality Studies, Alma Mater Studiorum-University of Bologna, Corso d'Augusto 237, 47921 Rimini, Italy; cinzia.calcabrini@unibo.it (C.C.); elena.catanzaro2@unibo.it (E.C.); eleonora.turrini@unibo.it (E.T.); carmela.fimognari@unibo.it (C.F.)

² Department of Pharmaceutical Sciences, College of Pharmacy, Larkin University, Miami, FL 33169, USA

* Correspondence: abishayee@ularkin.org or abishayee@gmail.com (A.B.); carmela.fimognari@unibo.it (C.F.); Tel.: +1-305-760-7511 (A.B.); +39-0541-434658 (C.F.)

† These authors contributed equally to this article.

Received: 25 August 2017; Accepted: 9 October 2017; Published: 13 October 2017

Abstract: Despite the huge investment into research and the significant effort and advances made in the search for new anticancer drugs in recent decades, cancer cure and treatment continue to be a formidable challenge. Many sources, including plants, animals, and minerals, have been explored in the oncological field because of the possibility of identifying novel molecular therapeutics. Marine sponges are a prolific source of secondary metabolites, a number of which showed intriguing tumor chemopreventive and chemotherapeutic properties. Recently, Food and Drug Administration-approved drugs derived from marine sponges have been shown to reduce metastatic breast cancer, malignant lymphoma, and Hodgkin's disease. The chemopreventive and potential anticancer activity of marine sponge-derived compounds could be explained by multiple cellular and molecular mechanisms, including DNA protection, cell-cycle modulation, apoptosis, and anti-inflammatory activities as well as their ability to chemosensitize cancer cells to traditional antitumor chemotherapy. The present article aims to depict the multiple mechanisms involved in the chemopreventive and therapeutic effects of marine sponges and critically explore the limitations and challenges associated with the development of marine sponge-based anticancer strategy.

Keywords: marine sponges; cancer; apoptosis; cell cycle; cancer chemoprevention; cancer chemotherapy

1. Introduction

Natural compounds represent a useful source of active molecules and play a noteworthy role in the drug discovery process. Inter alia, for several years, cancer therapy and prevention witnessed an exponential use of natural compounds as effective alternatives or supporting elements to traditional antitumor therapy. Nowadays, about 60% of all newly discovered anticancer drugs derive from natural compounds [1].

Cancer is a world burden. Malignant tumors still claim about 8.8 million victims per year and are the cause of 1 out of 6 deaths, globally [2]. Two major issues of current anticancer therapy are low efficacy and safety; as a result, the identification of new anticancer strategies, endowed with a better pharmaco-toxicological profile, to be used alone or in association with conventional chemotherapy, is essential. In this context, natural compounds could play a pivotal role since they are usually less toxic than traditional chemotherapy agents, they are effective, inexpensive and easily available [3]. In particular, natural compounds are often characterized by a better safety profile than traditional anticancer agents and the ability to inhibit cancer formation and development through the interaction with multiple cell signaling pathways [3]. This property makes it possible to affect multiple hallmarks

of cancer at the same time, thus to effectively fight a complex disease, such as neoplasms. Furthermore, adapting Hippocrates' precept "Primum non nocere" and following the World Health Organization's (WHO) policy of prevention as a long-term strategy for the control of cancer, many natural compounds could represent potent chemopreventive agents.

If we ask ourselves what are the most renowned natural compounds currently used to cure major illnesses, we are sure that most readers would mention the cardiac glycoside digoxin or the antitumor taxanes or vinca alkaloids, all compounds obtained from the land. However, 70% of our planet is covered by water and marine biodiversity is unrivaled. This is the reason why medical research has focused on the marine world in the last six decades, starting to exploit its huge potential. Besides, modern resources and technological advances have made it possible to reach the unexplored sea depths, making marine biota accessible to researchers. Thus, in the 1950s Bergmann and Feeney [4] extracted for the first time an unusual nucleoside from the sponge *Tethya crypta*, which would later be renowned as the anticancer drug vidarabine [4]. Around 28,000 new compounds of marine origin were discovered [5]. Among all marine organisms, sponges, which belong to the phylum of *Porifera*, represent the richest source of natural marine compounds, contributing to 30% of all natural marine products identified so far [6]. This fact should not be surprising. Sponges are sessile invertebrates; they do not have an innate immune system or mechanic defense structures like shells or spines, therefore the only method they have of preserving themselves is to produce metabolites that act as a self-defense device and let them adapt to the environment [7]. Moreover, the biodiversity of sponges' intraphylum translates into a chemical variety of the molecules produced. So far, particular nucleosides, sterols, alkaloids, peroxides, terpenes, fatty acids, amino acid derivatives and cyclic peptides have been discovered as active compounds and at least 60 of them have chemopreventive and/or anticancer potential [8]. These remarkable properties could be justified by the sponge's ability to affect multiple cellular and molecular events, including DNA protection, cell cycle, apoptosis and inflammation as well as by their ability to chemosensitize cancer cells to traditional antitumor chemotherapy.

The aim of this review is to illustrate and provide an update on the potential chemopreventive and antitumor effects of marine sponges, with particular focus on their ability to affect different pathways involved in cancer formation and development, and critically examine the limitations and challenges associated with the development of marine sponge-based anticancer strategy.

2. Proapoptotic and Anti-Inflammatory Effects

Apoptosis, or programmed cell death, is a physiological mechanism that occurs to preserve cellular homeostasis. It is essential for many biological activities, such as embryonic development, cell turnover, immune system maturation, and regulation of tissue homeostasis; accordingly, resistance to this process is itself a hallmark of cancer [9]. Furthermore, specific alterations in the apoptotic pathway are tumor distinctive, hence they may turn out to be a selective target for antitumor therapy [9].

Two main pathways can activate apoptosis: the mitochondrial or intrinsic pathway, and the receptor-mediated or extrinsic pathway. The intrinsic pathway refers to apoptotic cell death triggered by an enormous variety of intracellular damage or stress signals such as DNA damage, oxidative stress, endoplasmic reticulum (ER) stress, and cytokine withdrawal [10]. This pathway is regulated by the Bcl-2 family and involves mitochondrial outer membrane permeabilization (MOMP). Bcl-2 family counts pro- and anti-apoptotic proteins, and the balance between these elements decides the fate of the cell. Basically, when proapoptotic stress occurs, BH3-only proteins are recruited and activate Bax and Bak, which in turn trigger MOMP. As a result, cytochrome *c*, the second mitochondria-derived activator of caspases (SMAC) and other mitochondrial intermembrane space proteins are liberated into the cytosol. In this way, the apoptotic protease-activating factor 1 (APAF1) is able to reach cytochrome *c*, assembles the apoptosome, and activates caspase 9. Then, caspase 9 triggers the effector caspase-3 and caspase-7 that lead to apoptosis. At the same time, SMAC blocks the caspase inhibitor X-linked inhibitor of apoptosis protein (XIAP) leading to apoptosis. On their part, Bcl-2 homology (BH) 3-only proteins, and in particular Bid, can be activated by caspase-8 and lead directly

to MOMP activation, thus representing the junction between the intrinsic and the extrinsic apoptotic pathway. Apoptosis induced by the extrinsic pathway occurs in response to extracellular stress signals like the pathogen- or damage-associated molecular patterns (PAMP or DAMP) or through the propagation of cytokines [10]. The extrinsic pathway requires the activation of specific receptors, such as Fas (CD95/TNFRSF6/Apo-1) or tumor necrosis factor (TNF)-family receptors, such as TNF-related apoptosis-inducing ligand (TRAIL) receptor (TRAILR). Upon binding with the designated ligands, a caspase cascade is triggered. At that point, adaptor proteins, such as Fas-associated death domain protein (FADD), make caspase-8 and caspase-10 dimerize, which in turn activate the effector caspase-3 and caspase-7, leading to apoptosis [10].

Intriguingly, inflammation, by now considered the seventh fully-fledged hallmark of cancer, has several converging pathways with apoptosis [9]. In fact, all TNF-receptor family members trigger both apoptosis and the activation of transcription factors leading to cytokine and chemokine production, but in a different chronological order [11]. While Fas receptor and TRAILR trigger first apoptosis and then inflammation, the TNF receptor 1 (TNFR1) first allows the translation of inflammatory genes and then apoptosis [11]. TNF-TNFR1 binding activates the nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B), c-Jun amino-terminal kinase (JNK), the mitogen-activated protein kinases (MAPKs), and p38 [11]. NF- κ B is considered the most important link between cancer and inflammation. It is a transcription factor that regulates many biological processes, such as cytokine production, immunity modulation, cell growth, both cell survival and apoptosis, and tumorigenesis. Its constitutive activation has been described in several cancer types [12] and in all cancer phases (initiation, promotion and progression) [13]. NF- κ B consists of heterodimers or homodimers of the NF- κ B/Rel protein family, which mediate NF- κ B dimerization, nuclear import, and specific DNA binding. Depending on the stimulus, NF- κ B can be activated through different pathways. Microbial infections or inflammatory signals, such as TNF α and interleukin-1 (IL-1), act via I κ B/I κ B kinase (IKK). Briefly, in basal conditions NF- κ B is coupled to its inhibitor I κ B. When IKK is triggered, it phosphorylates I κ B that in turn makes the p50-RELA-dimer free to reach the nucleus and allows the modulation of gene transcription. Otherwise, TNF cytokine family can induce the non-canonical pathway. This pathway requires the activation of IKK α by the NF- κ B-inducing kinase (NIK). The formation of the complex NIK-IKK α -p100 (NF- κ B subunit) leads to the phosphorylation of the p100 subunit and the nuclear translocation of p52-RELB dimer.

So far, more than 60 compounds obtained from sea sponges have shown their potential anticancer activity through the induction of apoptosis. For the majority of them, the proapoptotic mechanism is not completely elucidated and only their ability to induce one or more hallmarks of apoptosis has been demonstrated (Table 1). Some of them are at the same time involved in the inflammatory pathway. Selected compounds for which cellular and molecular mechanisms were explored in depth are presented and discussed in more detail in the text.

Table 1. Apoptosis-inducing compounds isolated from marine sponges.

Compounds	Sponge	Cell Line	Concentration Range (μM ^a)	Chromatin Condensation/DNA Fragmentation	Phosphatidylserin Externalization	Caspase Activation	PARP Cleavage	Reference
Aaptamine	<i>Aaptos</i> sp.	THP-1	50–200		✓			[14]
Demethyl(oxy)aaptamine	<i>Aaptos</i> sp.	THP-1	10–25		✓			[14]
Isoaaptamine	<i>Aaptos</i> sp.	THP-1	10–25		✓			[14]
Microsclerodermin A	<i>Amphibleptula</i> sp.	AsPC-1	2.4	✓		3 and 7		[15]
		BxPC-3	2.4	✓		3 and 7		
		PANC-1	2.4	✓		3 and 7		
Isofistularin-3	<i>Aplysina aerophoba</i>	Raji	50	✓		3 and 7	✓	[16]
		U937	50	✓		3 and 7	✓	
Spongiatriol	<i>Australian spongia</i> sp.	AsPC-1	6.8	✓		3 and 7		[17]
		PANC-1	6.8	✓		3 and 7		
		MIA PaCa-2	6.8			3 and 7		
		BxPC-3	6.8			3 and 7		
Laulimalide	<i>Cacospongia mycofijiensis</i>	MDA-MB-435	0.1			3	✓	[18]
Scalaradial	<i>Cacospongia scalaris</i>	HeLa	10 $\mu\text{g}/\text{mL}$			3		[19]
		T47D	10 $\mu\text{g}/\text{mL}$	✓				
Candidaspongiolide	<i>Candidaspongia</i> sp.	U251 HCT116	0.05–0.10		✓	3 and 12	✓	[20]
Callyspongiolol	<i>Callyspongia</i> sp.	HL-60	31.0–77.5	✓				[21]
Crambescidin 800	<i>Crambe crambe</i>	HepG2	0.5–2.5		✓	3		[22]
Crambescidin 816	<i>Crambe crambe</i>	HepG2	0.5–2.5		✓	3		[22]
Crambescidin 830	<i>Crambe crambe</i>	HepG2	0.5–2.5		✓	3		[22]
Smenospongine	<i>Dactylospongia elegans</i>	U937	5–15	✓				[23]
		HL-60						
Pectenotoxin-2	<i>Dinophysis fortii</i> and <i>Dinophysis acuminata</i>	Hep3B	0.01 $\mu\text{g}/\text{mL}$		✓	3, 8 and 9		[24]
		U937	0.008–0.010 $\mu\text{g}/\text{mL}$	✓	✓	3	✓	[25,26]
Salarin C	<i>Fascaplysinopsis</i> sp.	K562 normoxic and hypoxic conditions	0.01–0.2	✓	✓	3 and 9	✓	[27,28]
Cacospongiolide	<i>Fasciospongia cavernosa</i>	HeLa T47D	10 $\mu\text{g}/\text{mL}$ 10 $\mu\text{g}/\text{mL}$	✓		3		[19]
Lasonolide A	<i>Forcepia</i> sp.	CA46, Ramos, Daudi, HL-60, MDA-MD-231, MCF-7, HCT-116, HT-29	0.1	✓				[29]

Table 1. Cont.

Compounds	Sponge	Cell Line	Concentration Range (μM ^a)	Chromatin Condensation/DNA Fragmentation	Phosphatidylserin Externalization	Caspase Activation	PARP Cleavage	Reference
Geoditin A	<i>Geodia japonica</i>	HL-60 HT-29	1.6 to 25 $\mu\text{g}/\text{mL}$ 5–30	✓ ✓	✓	3		[30,31]
Stelletin A	<i>Geodia japonica</i>	HL-60	4 $\mu\text{g}/\text{mL}$			3		[32]
Ilimaquinone	<i>Hippospongia metachromia</i>	HCT116	2.5–10	✓	✓	3 and 8	✓	[33,34]
		PC-3	2–10	✓				[35]
Spongistatin 1	<i>Spirastrella spinispirulifera</i> , <i>Hyrtios erecta</i>	MCF-7	0.0002–0.0005	✓		slight activation of 2, 3, 6, 7, 8 and 9		[36]
		Jurkat	0.0002	✓		2, 3, 7, 8 and 9	✓	[37]
		L3.6pl	0.00001–0.01	✓				[38]
Heteronemin	<i>Hyrtios</i> sp.	K562	1.4–5.6	✓	✓	3, 8 and 9	✓	[39]
		DU145	0.01–1 $\mu\text{g}/\text{mL}$	✓		3, 8 and 9		
		PC-3	0.01–1 $\mu\text{g}/\text{mL}$	✓		3, 8 and 9		[40]
		LNCaP	0.01 $\mu\text{g}/\text{mL}$	✓				
		T24 A498	0.1–0.8 $\mu\text{g}/\text{mL}$ 0.5–3	✓ ✓	✓	3 and 9 3, 8 and 9	✓ ✓	[41] [42]
Bastadin 6	<i>lanthella</i> sp.	HUVEC	0.01–1	✓		3 and 7		[43]
Irciniastatin A	<i>Ircinia ramose</i> , <i>Psammocinia</i> sp.	Jurkat	0.01	✓		3, 8 and 9		[44]
Jaspolid B	<i>Jaspis</i> sp.	Bel-7402	0.5	✓				
		HepG2	10–20	✓				[45]
Stelletin B	<i>Jaspis stellifera</i>	K562	0.012–0.054		✓	3 and 9	✓	[46]
		A549	0.02–1		✓	3 and 7	✓	[47]
		SF295	0.04–1		✓		✓	[48]
Jaspine B or Pachastrissamine	<i>Jaspis</i> sp. <i>Pachastrissa</i> sp.	B16 HaCaT	5 5 $\mu\text{g}/\text{mL}$	✓	✓	3 and 9 3	✓	[49,50]
Petrosterol-3,6-dione	<i>Lanthella</i> sp.	HL-60	19.9	✓				[51]
5 α ,6 α -epoxy-petrosterol	<i>Lanthella</i> sp.	HL-60	21.3	✓				[51]
petrosterol	<i>Lanthella</i> sp.	HL-60	21.5	✓				[51]
Leiodermatolide	<i>Leiodermatium</i> sp.	AsPC-1	0.01	✓				
		BxPC-3	0.01	✓		3		[52]
		MIA PaCa-2	0.01	✓		3		
Naamidine A	<i>Leucetta chagosensis</i>				✓	3, 8 and 9	✓	[53]
Monanchocidin A	<i>Monanchora pulchra</i>	HeLa	1.39	✓	✓	3 and 7		
			2.01					[54]

Table 1. Cont.

Compounds	Sponge	Cell Line	Concentration Range (μM ^a)	Chromatin Condensation/DNA Fragmentation	Phosphatidylserin Externalization	Caspase Activation	PARP Cleavage	Reference
Monanchocidin B	<i>Monanchora pulchra</i>	HeLa	0.58 1.36	✓		3 and 7		[54]
Monanchocidin C	<i>Monanchora pulchra</i>	HeLa	1.84 1.31	✓		3 and 7		[54]
Ptilomycalin A	<i>Monanchora pulchra</i>	HeLa	1.1 0.5	✓		3 and 7		[54]
Monanchomycalin B	<i>Monanchora pulchra</i>	HeLa	1.5 1.72	✓		3 and 7		[54]
Normonanchocidin D	<i>Monanchora pulchra</i>	HeLa	2.1 5.2	✓		3 and 7		[54]
Urupocidin A	<i>Monanchora pulchra</i>	HeLa	28.7 27	✓				[54]
Pulchrarin A	<i>Monanchora pulchra</i>	HeLa	51 58	✓		3 and 7		[54]
Pateamine	<i>Mycale</i> sp.	32D	0.1		✓			[55]
Mycalamide A	<i>Mycale</i> sp.	32D	0.1		✓			[55]
Latrunculin A	<i>Negombata magnifica</i>	MKN45 NUGC-4	–10 0.01–10			3 and 7		[56]
Kuanoniamines A	<i>Oceanapia sagittaria</i>	MCF-7	0.5–2.5	✓				[57]
Kuanoniamines C	<i>Oceanapia sagittaria</i>	MCF-7	1.0–2.5	✓				[57]
Dideoxypetrosynol A	<i>Petrosia</i> sp.	SK-MEL-2	0.1–0.3 $\mu\text{g}/\text{mL}$	✓		3 and 9	✓	[58]
Psammaplin A	<i>Psammaplysilla</i> sp.	Human endometrial Ishikawa		✓				[59]
Psammaplysene A	<i>Psammaplysilla</i>	Ishikawa ECC1	1 1				✓	[60]
(1' <i>R</i> ,5' <i>S</i> ,6' <i>S</i>)-2-(3',5'-dibromo-1',6'-dihydroxy-4'-oxocyclohex-2'-enyl)acetonitrile	<i>Pseudoceratina</i> sp.	K562	7.7–30.8		✓	3 and 9	✓	[61]
13 <i>E</i> ,17 <i>E</i> -globostellatic acid X methyl ester	<i>Rhabdastrella globostellata</i>	HUVEC	1–10			3 and 7		[51]
Rhabdastrellic acid-A	<i>Rhabdastrella globostellata</i>	HL-60		✓		3	✓	[62]

Table 1. Cont.

Compounds	Sponge	Cell Line	Concentration Range (μM ^a)	Chromatin Condensation/DNA Fragmentation	Phosphatidylserin Externalization	Caspase Activation	PARP Cleavage	Reference	
Rhizochalin or Rhizocalinin	<i>Rhizochalina incrustata</i>	HL-60	10–25		✓	3, 8 and 9		[63]	
		HT-29	1–6	✓	✓	3	✓	[64]	
		THP-1	1–10		✓	✓		✓	[65]
		PC-3	0.5–4		✓	✓	8	✓	
		DU-145	0.5–4		✓	✓	8	✓	
		22Rv1	0.5–4		✓	✓	8	✓	[66]
VCaP	0.5–4		✓	✓	8	✓			
Ircinin-1	<i>Sarcotragus</i>	SK-MEL-2	25–50	✓		3 and 9	✓	[67]	
Sipholenol A	<i>Siphonochalina</i> sp.	HepG2	17.18	✓		3		[68]	
		HCT-116	14.8	✓		3		[69]	
Sipholenol L	<i>Siphonochalina</i> sp.	HepG2	24	✓		3		[68]	
		HCT-116	19.8	✓		3		[69]	
Smenamides A and B	<i>Smenospongia aurea</i>	Calu-1	0.05–0.1		✓			[70]	
(Z)-stelletic acid C	<i>Stelletta</i> sp.	U937	17.2–103.3		✓	3, 8 and 9		[71]	
Renieramycin M	<i>Xestospongia</i> sp.	H460	5–40	✓					
		U373MG	0.0031			3	✓	[72,73]	

^a If not specified.

Renieramycin M is a tetrahydroisoquinoline extracted from the blue sponge *Xestospongia* sp. belonging to the same chemical family of ecteinascidin 743 (ET-743, YondelisTM, or Trabectedin), the first marine anticancer agent approved by the European Union and by the Food and Drug Administration (FDA) for the treatment of advanced soft tissue sarcomas [74,75]. At nanomolar concentrations, it is cytotoxic for human colon, lung, prostate, brain, and breast cancer cells [73,76,77]. It conveys human non-small cell lung cancer H460 cells to p53-dependent apoptosis through the activation of the intrinsic pathway [72]. It is also able to stimulate anoikis. Anoikis is a particular type of apoptosis known as “detachment-induced apoptosis” and it is involved in the inhibition of cancer metastasization. Indeed, avoiding anoikis means avoiding apoptosis in cells that have lost adhesion to the extracellular matrix, allowing them to survive and develop a tumor [78]. Renieramycin M overcomes resistance to anoikis in H460 cells and in anoikis-resistant H460 cells [72,79]. Resistance to anoikis is associated with the up-regulation of phosphorylated extracellular signal-regulated kinase (p-ERK), phosphorylated ATP-dependent tyrosine kinase (p-AKT), Bcl-2, and Mcl-1. At subtoxic concentrations (0.5–2.5 μ M for 24 h) [72], renieramycin M reduced all these protein levels [79]. Furthermore, renieramycin M decreased the expression of CD133, CD44 and ALDH1A1, which are markers of cancer stem cells [80].

Aptamines are alkaloids extracted from different species of *Aaptos* sp. showing important potential anticancer activity. Dyshlovoy and coll. studied in particular three compounds (aaptamine, demethyl(oxy)aaptamine and isoaptamine) and demonstrated their ability to trigger apoptosis in a monocytic leukemia cell line (THP-1). Among the three molecules, demethyl(oxy)aaptamine and isoaptamine exhibited the strongest activity (Table 1). In a murine epidermal cell line (JB6 Cl41), all of them activated NF- κ B and the transcription factor activator protein-1 (AP-1). Demethyl(oxy)aaptamine and isoaptamine deregulated p53 transcriptional activity, while aaptamine did not affect it [14]. NF- κ B and AP-1 partake in several biological processes, such as cell proliferation and migration, inflammation and apoptosis. AP-1 is a protein complex constituted by JUN, FOS, activating transcription factor (ATF), and musculoaponeurotic fibrosarcoma (MAF) family members that bind a shared DNA site, the AP-1-binding site [81]. NF- κ B and AP-1 both have a dual role as tumor initiators and suppressors, depending on their composition, cell lines, microenvironment and stimuli [82]. In this respect, for example, a concurrent activation of these complexes is crucial to trigger apoptosis through the Fas ligand expression in leukemia cells (Jurkat) by DNA damaging agents [83]. Activation of AP-1 and NF- κ B, and decrease in p53 levels are observed in the JB6 Cl41 response to cisplatin. For this reason, Dyshlovoy and coll. concluded that modulation of AP-1, NF- κ B, and p53 is involved in aaptamine-mediated apoptosis [14].

Psammaplysene A (PsA) is a bromotyrosine derivative extracted from the marine sponge *Psammaplysilla* sp. Its potential anticancer activity has been explored in two endometrial cancer cell lines, Ishikawa and ECC-1. In both cell lines, PsA induced apoptosis and increased the expression of nuclear Forkhead box O1 (FOXO1) protein [60] FOXO1 is a transcription factor that plays an important role in the apoptotic process. Among others, FOXO1 activates the proapoptotic Bim protein or induces TRAIL-mediated apoptosis [84]. FOXO1 moves back and forth between nucleus and cytoplasm, creating a homeostatic equilibrium. Protein kinase B (Akt) can phosphorylate, inactivate and extrude FOXO1 from the nucleus, an event that inhibits apoptosis [85,86]. This event becomes pathologic when Akt is constitutively activated or when the tumor suppressor phosphatase and tensin homolog (PTEN) gene is mutated. Almost 50% of type I endometrial cancers are characterized by this mutation [87], and FOXO1 levels are significantly lower or absent in endometrial tumors [88,89]. To demonstrate that the induction of apoptosis and the increased expression of FOXO1 are related to each other, Berry et al. [60] evaluated the proapoptotic potential of PsA in silenced FOXO1 ECC-1 cells and in overexpressing FOXO-1 Iwasawa cells. Compared to the respective wild-type cells, PsA induced PARP cleavage to a lesser extent in the FOXO-1 silenced cell line, and to a greater extent in the overexpressing ones, demonstrating FOXO1 involvement in its modus operandi.

Stellettin B, a triterpene isolated from the marine sponge *Jaspis stellifera*, showed potential anticancer activity in various cell lines, including human glioblastoma (SF295), human non-small cell lung cancer (A549), and human chronic myeloid leukemia cells (K562 and KU812). In all four cell lines, Stellettin B induced an increase in the reactive oxygen species (ROS) levels that was linked to apoptosis. In leukemia cells, it triggered the mitochondrial pathway through upregulation of Bak and Bax protein levels, downregulation of Bcl-2, and induction of MOMP [46]. In SF295 and A549 cell lines, the pathway was not investigated [48].

Tang et al. [49] demonstrated that Stellettin B blocked the phosphorylation of Akt in SF295, with no effect on phospholipid kinase phosphatidylinositol 3-kinase (PI3K), its main activator. Thus, even if Stellettin B targets some Akt upstream proteins different from PI3K, the PI3K/Akt pathway is involved in its proapoptotic potential. Since their upstream effectors p38 and ERK were not modulated by Stellettin B, the involvement of the Ras/MAPK pathway was excluded. More recently, Chen et al. [47] and Wang et al. [48] confirmed Stellettin B's ability to trigger apoptosis via Akt inhibition in A549, K562 and KU812 cells. However, as opposed to the evidence on glioblastoma cells, the authors showed the inhibition of the expression of p110, the catalytic subunit of PI3K, and the phosphorylation of phosphoinositide-dependent kinase-1 (PDK1), another Akt upstream protein [46,47]. An interesting experiment would be to evaluate the PDK1 levels in Stellettin B-treated SF295 cells. In the leukemic cell lines, Stellettin B inhibited Stat5 phosphorylation, a PI3K upstreaming protein, increasing the evidence that PI3K/Akt is involved in its cytotoxic mechanism [46]. In both leukemia and lung cancer cells, the expression of Akt downstream proteins were investigated after Stellettin B treatment, and a decrease in mTOR, a negative modulator of autophagy, was recorded, suggesting a role of this process in the potential antitumor activity of this sponge derivative [46,47].

The dibromotyrosine derivative (1'*R*,5'*S*,6'*S*)-2-(3',5'-dibromo-1',6'-dihydroxy-4'-oxocyclohex-2'-enyl) acetonitrile (DT), isolated from the sponge *Pseudoceratina* sp., showed proapoptotic activity on leukemia K562 cells. Relying on ROS production, it activated the intrinsic pathway, as demonstrated by the activation of caspase-9, and induced the disruption of MOMP. In addition, different mechanisms intertwine in DT activity. Certainly, oxidative stress plays a pivotal role in its potential antitumor activity. Indeed, oxidative stress seems to drive the modulation of the other two pathways involved in its cytotoxicity, i.e., PI3K/Akt and IKK/NF- κ B [61].

Su et al. [61] showed that, unlike PsA and Stellettin B, DT induced apoptosis through the inhibition of FOXO genes and the activation of PI3K/Akt. DT represses PTEN activation and stimulates Akt phosphorylation that would lead to FOXO deactivation.

The IKK/NF- κ B pathway is involved in different cellular events, such as inflammation or cell survival [90] and it has been demonstrated that ROS-mediated apoptosis is linked to the repression of this pathway [91,92]. DT reduced IKK/NF- κ B downstream protein expression via ROS stimulation, suggesting its implication in DT-induced K562 apoptosis [61].

Another cellular mechanism involved in sponge-mediated apoptosis is ER-stress. ER-stress is triggered when an imbalance between protein folding capacity and functional demand occurs. The most common pathway involved in ER homeostasis restoration involves the unfolded protein response (UPR). UPR compensates for the accumulation of misfolded or unfolded proteins, but if stress is prolonged or the adaptive response is unsuccessful, apoptosis ensues. Protein kinase RNA (PKR)-like 67 ER kinase (PERK) is one of the ER transmembrane receptors that govern UPR. Among other kinases, both PERK and PKR can phosphorylate and inhibit eukaryotic translation initiation factor 2 subunit 1 (eIF2 α). When eIF2 α is phosphorylated, it is unable to efficiently initiate translation, leading to the inhibition of global protein synthesis and to apoptosis mediated by different pathways [93].

Candidaspongolide (CAN) causes apoptosis in human U251 glioma and HCT116 colon carcinoma cells in a singular way. It triggers caspase-12 and, to a lesser extent, caspase-3 activation. Caspase-12 is involved in ER stress-induced apoptosis in rodents [94], while its role in humans is not clear yet [95,96]. Nonetheless, ER-stress was demonstrated to play a crucial role in CAN-induced apoptosis in the human cancer cells tested. In fact, CAN phosphorylates eIF2 α through PKR activation [20]. Of note,

UPR was not involved in CAN-mediated ER stress [20]. Since eIF2 α phosphorylation is essential for CAN-mediated apoptosis and other UPR effectors, such as C/EBP homologous protein (CHOP) were not modulated, Trisciuglio and coll. presumed that the PKR/eIF2 α pathway activates NF- κ B or the FADD/caspase-8 and Apaf-1/caspase-9 pathways, leading to apoptosis [20].

The methanol and hexane extracts of the marine sponge *Geodia cydonium* showed potential anticancer activity on different rabbit colon carcinoma and myosarcoma cell lines, even if the mechanism of action has not been investigated [97]. On the contrary, the methanol extract did not exert any cytotoxic or proapoptotic effect on breast cancer cells (MCF-7) [98]. However, in the same cell line, the extract exhibited an anti-inflammatory activity [98]. It decreased vascular endothelial growth factor (VEGF) levels and other proinflammatory cytokines such as interferon- α (IFN- α) and TNF- α , mutually interconnected with NF- κ B. Moreover, it directly downregulated the gene expression of two NF- κ B subunits, NF- κ B1 and c-Rel [98]. All this information suggests that *Geodia cydonium* is endowed with an anti-inflammatory potential mediated by NF- κ B inactivation.

Heteronemin is a sesterterpene isolated from the sponge *Hyrtios* sp. that modulates the transcriptional level of several pathways related to cell survival, such as cell cycle and apoptosis, and inflammation. Heteronemin triggers both the intrinsic and the extrinsic apoptotic pathways in chronic myeloid leukemia K562 and human renal cell carcinoma A498 cells [39,42] and negatively modulates NF- κ B at subtoxic concentrations in K562 [39]. Precisely, it inhibited the PI3K/AKT pathway and the MAPK family member ERK in A498 cells [39,42]. In K652 cells, heteronemin targets NF- κ B upstream protein MAPK, sensitized K652 cells to TNF- α -mediated apoptosis, and inhibited the phosphorylation of p38, another MAPK protein that turned out to be crucial for heteronemin-mediated apoptosis, and c-Jun NH2-terminal kinase (JNK) [42]. JNK belongs to the MAPK family and could trigger apoptosis or autophagy. In the heteronemin-treated A498 cells, it led to autophagy [42]. One interesting fact is that the inhibition of autophagy significantly increased heteronemin-induced apoptosis [42]. Thus, the combination of heteronemin with autophagy inhibitors could be an intriguing therapeutic strategy.

The precise mechanism of the potential anticancer activity of heteronemin has not been completely elucidated and, since MAPK is involved in both anti-inflammatory and proapoptotic pathways, further studies are required to understand the role of its modulation in the proapoptotic and anti-inflammatory potential of heteronemin [39].

Dideoxypetrosynol A is a polyacetylene isolated from *Petrosia* sp. known for its proapoptotic activity on human skin melanoma SK-MEL-2 and human leukemia U937 cells mediated by the mitochondrial pathway [58,99]. In U937 cells, it exhibited very interesting anti-inflammatory properties through the inhibition of the arachidonic acid (AC) pathway. Cyclooxygenase-1 (COX-1) and cyclooxygenase-2 (COX-2) are two crucial enzymes in AC metabolism. COX-1 is the constitutive form, while COX-2 is inducible [99]. These two enzymes catalyze the conversion of AC to the inflammatory mediator prostaglandins (PG). An over-induction of COX-2 could bring to an overproduction of PGE₂ and facilitates the proliferation of neoplastic cells [100,101]. COX-2 up-regulation has been described in several kinds of cancer and is involved in carcinogenesis, metastatic potential, and angiogenesis [102]. Dideoxypetrosynol A markedly downregulated COX-2 mRNA and protein expression in U937 cells, while it did not affect COX-1 levels. It also induced a dose-dependent decrease in PGE₂ amount [99].

Other crucial promoters and mediators of inflammation are the proinflammatory cytokines, such as the interleukin family. For instance, interleukin 8 (IL-8) is involved in many inflammatory disorders, from arthritis to cancer. It activates neutrophils and, at the same time, has chemo-attractant properties. It promotes angiogenesis in physiological situations, such as wound healing, but in case of existent tumors this property could be linked to cancer ability to create new blood vessels, activate survival-signaling pathways, and promote metastasis [103]. Thus, the inhibition of IL-8 can provide a strategy to contrast cancer-related inflammation. Theopederins K and L from the marine sponge *Discodermia* sp. and mycalamide A from *Mycala* sp. are non-specific protein synthesis inhibitors able to

inhibit IL-8 secretion in different pancreatic cancer cell lines [104]. Their ability to block IL-8 deserves more in-depth study to fully understand their impact on the tumor process.

3. Antiproliferative Effects

Cancer cells are characterized by sustaining proliferative signaling that could result from the insensitivity to proliferation-inhibitory signals arising from the stroma or specific gene expression patterns [105]. Many marine compounds display antiproliferative activity targeting the modulators of the cell-cycle progression. A list of marine spongean compounds with antiproliferative activity is presented in Table 2. Selected compounds for which cellular and molecular mechanisms were explored in depth are presented and discussed in more detail in the text.

Table 2. Antiproliferative compounds isolated from marine sponges.

Compounds	Sponges	Cell Lines	Concentrations (μM *)	Phase of Cell-Cycle Arrest	Molecular Targets	Reference
Aaptamine	<i>Aaptos aaptos</i>	NT2	1–50	G2/M	/	[106]
		HepG2	50–100	G2/M	↓ cyclins D and E, CDK2	[107]
		HCC-LM3		S	↑ p21	
	<i>Aaptos suberitoides</i>	MG63	30 $\mu\text{g}/\text{mL}$	G2/M	↑ p21	[108]
		K562	20–100	G2/M	↑ p21	[109]
Aphrocallistin	<i>Aphrocallistes beatrix beatrix</i>	Panc-1	≤ 46.5	G0/G1	/	[110]
Aragusterol A	<i>Xestospongia</i> sp.	A549	1–10	Late G1	↓ CDK2, CDK4 ↓ cyclins D1, A, E ↓ pRb	[111]
Batzelline A and B Isobatzelline A, C, D Secobatzelline A and B Isobatzelline E	<i>Batzella</i> sp.	AsPC-1	5 or 25 $\mu\text{g}/\text{mL}$	S G2/M	Intercalate into DNA and/or inhibit Topoisomerase II activity	[112]
Calyculin A	<i>Discodermia calyx</i>	MDA-MB-468 MCF-7 MDA-MB-231	0.01	G0/G1	↓ cyclin D1	[113]
Crambescidin 800	<i>Monanchora unguiculata</i>	K562	0.15–1.5	S	↑ p21	[114]
Crambescidin 800, 816 and 830	<i>Crambe crambe</i>	HepG2	2.5	G0/G1	↓ cyclins A, D ↓ CDK2, 6, 1 ↑ CDKN2A, 2D, 1A	[22]
Dictyostatin-1	<i>Corallistidae</i> sp.	A549	0.01–1	G2/M	↑ micronuclei, asters and abnormal mitotic spindles formation	[115]
Dideoxypetrosynol A	<i>Petrosia</i> sp.	U937	0.2–1 $\mu\text{g}/\text{mL}$	G0/G1	↑ cyclin D1 ↓ cyclin E ↑ pRB-E2F1 complex and p16	[116]
(+)–Discodermolide	<i>Discodermia dissoluta</i>	MCF-7, CA46	0.01–1	G2/M	Stabilize microtubules	[117]
		A549	0.07–0.166	G2/M	abnormal mitotic spindles ↓ microtubules dynamicity	[118]
Geodiamolide A, B, H and I	<i>Geodia corticostylifera</i>	T47D, MCF7	50 ng/mL	Not investigated	↑ disorganization of actin filaments	[119]
(19Z)–Halichondramide	<i>Chondrosia corticata</i>	A549	0.025–0.1	G2/M	↑ p53, GADD45 ↓ CDC2, CDC25C, cyclin B1, cyclin A	[120]
Hemiasterlin, Hemiasterlin A and B	<i>Hemiasterella minor</i>	MCF-7	0.0005–0.01	G2/M	↑ abnormal mitotic spindles formation	[121]
Jaspolid B	<i>Jaspis</i> sp.	Bel-7402 HepG2	20	G0/G1	↑ microtubule disassembly	[45]
Laulimalide	<i>Cacospongia mycofijiensis</i>	MDA-MB-435	0.02	G2/M	Microtubule stabilization	[18]
		A-10 SK-OV-3	0.02–2		↑ micronuclei and abnormal mitotic spindles formation	

Table 2. Cont.

Compounds	Sponges	Cell Lines	Concentrations (μM ^a)	Phase of Cell-Cycle Arrest	Molecular Targets	Reference
Leiodermatolide	<i>Leiodermatium</i> sp.	PANC-1	0.01–0.1	G2/M	↓ mitotic spindles formation and microtubule elongation	[52]
		A549	0.01–1	G2/M	↓ mitotic spindles formation	[122]
		U2OS	0.018–0.23	G2/M	Tubulin disruption Centrosome amplification Micronuclei formation	[123]
Pachymatismin	<i>Pachymatisma johnstonii</i>	DU145	4–16		Microtubules depolymerization	[124]
		NSCLC-N6	2–20 $\mu\text{g}/\text{mL}$	G0/G1		[125]
		NSCLC-N6 subcutaneous xenografts	0.5–5 mg/kg		↓ tumor growth	[126]
Peloruside A	<i>Mycale hentscheli</i>	H441	0.01–1	G2/M	Microtubule stabilization, ↑ micronuclei and abnormal mitotic spindles formation	[127]
		MCF-7	0.025–0.1	G2/M	↓ microtubule dynamicity (growth rate, growth length, time spent growing)	[128]
PHC-1	<i>Phyllospongia chondrodes</i>	K562	0.1–5 $\mu\text{g}/\text{mL}$	G0/G1	↑ haemoglobin, glycophorin A and enucleation	[129]
PM050489, PM060184	<i>Lithoplocamia lithistoides</i>	A549	$0.25-1 \times 10^{-3}$	G2/M [130]	↓ microtubules formation binding $\alpha\beta$ tubulin dimers	[131]
PM060184					0.001	↑ abnormal mitotic spindles formation, ↓ CDK1, ↑ p21
		HCT116	0.01			
		MDA-MB-231 subcutaneous xenografts	16 mg/kg		↑ formation of multinucleated cells	
Siphonol-A	<i>Siphonochalina siphonella</i>	PC-3	7.9	G0/G1	/	[133]
Smenospongine	<i>Dactylospongia elegans</i>	K562	3–15	G0/G1	↑ p21, ↓ p57, ↓ pRb; ↑ haemoglobin, glycophorin A	[23,134]
(8E,13Z,20Z)-strobilinin/ (7E,13Z,20Z)-felixinin 1:1	<i>Psammocinia</i> sp.	HeLa	10–50	S	↓ topoisomerase I and polymerase alpha-primase activities	[130]

^a If not specified.

Aragusterol A is a steroid isolated from *Xestospongia* sp. that exhibits antiproliferative activity on many human cancer cell lines (breast, ovarian and cervical cancer, lung, oral epidermoid, gastric, colon cancer and leukemia) [111]. It inhibits protein and nucleic acid synthesis and induces cell-cycle arrest at late G1 phase in human non-small cell lung cancer. The cell-cycle block was dependent on the reduced expression of CDKs and cyclins involved in the G1-S transition, such as CDK2, CDK4, cyclin D1, A, and E. In particular, the decrease in the complex cyclin E-CDK2 activity led to a reduced pRb phosphorylation and, consequently, to an antiproliferative effect [135].

(19Z)-Halichondramide is a macrolide produced by *Chondrosia corticata*. It showed antiproliferative activity against non-small cell lung cancer cells (A549) through the block in the G2/M phase. The arrest in the G2/M phase was related to an increase in p53 and GADD45 (growth arrest and DNA damage), and to the downregulation of their downstream target genes including cyclins B1 and A, CDC2 and its phosphatase activator CDC25C [120]. Additionally, (19Z)-Halichondramide suppressed the constitutively activated Akt/mTOR pathway, responsible for cell proliferation and survival.

In particular, this natural compound induced the downregulation of mTOR and in the same way reduced the expression of the mTOR precursor Akt, its survival effectors p70 S6 kinase and the eukaryotic initiation factor 4E (eIF4E)-binding protein 1 (4EBP1). Furthermore, a downregulation was noticed of the AMP-activated protein kinases (AMPKs), ERK, and p38, which regulate cell proliferation [120].

Smenospongine is a sesquiterpene founded in *Dactylosporgia elegans* [23]. It has antiproliferative effects in a human chronic myelogenous leukemia cell line (K562) through the cell-cycle arrest in G0/G1 phase mediated by p21 increase and p57 decrease [134]. Concurrently with the G0/G1 block, an increase in peroxidase activity, a marker of hemoglobin production, was noticed. K562 differentiation was also confirmed by the increase in glycophorine A, a surface sialoglycoprotein expressed during erythroid differentiation [136], and the inhibition of Crkl phosphorylation by Bcr–Abl tyrosine kinase, a marker of chronic myelogenous leukemia pathogenesis [134,137]. A more recent study showed the antiproliferative effect of smenospongine also in other leukemia cells (HL60 and U937), characterized by a G0/G1 arrest. Furthermore, the study reported the involvement of the p21-pRb pathway in the smenospongine-mediated G0/G1 arrest, but no effect of the marine compound on p21 promoter was recorded, leaving how smenospongine increases p21 unclear [23]. Likewise, the spongean sesterterpene PHC-1–PHC-7 induced K562 differentiation as demonstrated by G0/G1 arrest, and increased hemoglobin production, glycophorine A expression, and enucleation of cells [129].

Another compound inducing K562 differentiation is Crambescidin 800, an alkaloid isolated from *Monanchora unguiculata*. Like smenospongine, crambescidin 800 increased hemoglobin peroxidase activity and p21 expression; no alterations in p27 expression were noticed. Unlike smenospongine, it caused an accumulation of cells in the S-phase [114]. Furthermore, morphological changes with development of neurite formation were observed in murine neuroblastoma cells treated with crambescidin 800 [114]. A G0/G1 block was recorded in a human hepatocellular carcinoma cell line treated with crambescidin 800 or, more powerfully, with crambescidin 830 and 816. The cell-cycle block was accompanied by the downregulation of gene expression of CDK1 and 2, cyclins A and D, and by the upregulation of some cyclin-dependent kinase inhibitors (CDKN2A, 2D and 1A) at mRNA levels. Crambescidins also activated p53-downstream target genes, such as GADD45 and 14-3-3- σ , which in turn inactive CDK1 and lead to cell-cycle arrest. At the same time, PUMA, another p53-target gene, was activated, leading to apoptosis via the intrinsic pathway (see also Table 1) [22].

The cell-cycle arrest could be the consequence of the alterations in microtubule-tubulin kinetics since they play a pivotal role in mitosis and cell division: the microtubule network reorganizes, in a highly coordinated manner, into a mitotic spindle that attaches and separates chromosomes. Therefore, over the years, several anticancer agents targeting microtubule have been developed. These compounds bind tubulin and induce microtubule stabilization or destabilization, which leads to disruption of the mitotic spindle, cell-cycle block in M phase and, eventually, cell death [138]. The best-known antitubulin drugs are taxoids and vinca alkaloids. Microtubules are made up of 13 parallel protofilaments arranged in a circle. The polymerization occurs by the addition of α , β -tubulin heterodimers and GTP hydrolysis at the plus ends; the depolymerization process, instead, occurs with the release of GDP at the minus ends. Stabilizing agents, such as vinca alkaloids, induce assembly of heterodimers by increasing lateral protofilament interactions. Destabilizing agents, such as taxanes, reduce polymerization by inducing an altered conformation [139].

Marine sponge-derived compounds, such as avarols, arenastatins, halichondrin, jaspolid, milnamide, and spongistatin are microtubules-destabilizing agents; instead, dactyloides, dictyostatin, discodermolide, hemiasterlin, laulimalide, peloruside A, and zampanolide act as stabilizing microtubules [140]. However, clinical studies for many of them (avarols, arenastatins, spongistatins, dactyloides, dictyostatins, laulimalides, and zampanolides) are lacking because of their low availability in nature or long synthesis processes, rapid inactivation, or toxicity [141].

Laulimalide and the less potent isolaulimalide are poor substrates of the P-glycoprotein (Pgp) efflux pump, as demonstrated in human ovarian cancer cells overexpressing Pgp and

vinblastine-resistant [18]. Likewise, discodermolide is effective in human colon and ovarian carcinoma cell lines overexpressing Pgp and resistant to paclitaxel [142]. Peloruside A is also not efficiently extruded by Pgp since its cytotoxic activity was similar in multidrug resistance (MDR)-overexpressing Chinese hamster ovary cells and Pgp-overexpressing human ovarian carcinoma cells resistant to paclitaxel, compared to parental cell lines [143].

Peloruside A, isolated from *Mycale hentscheli*, is a microtubule stabilizer that binds β -tubulin at the laulimalide/peloruside domain, promotes G2/M arrest, and induces the formation of micronuclei, multiple bundles of microtubules and asters in mitosis, as shown in lung adenocarcinoma cells [127,143]. The patent for peloruside A belongs to the University of Victoria [144]. A study by the same University recently demonstrated that peloruside A forms a hydrogen bond with aspartic acid 297 of human β -tubulin. This evidence suggests a unique mechanism for microtubule stabilization of peloruside A [145].

A clinically important observation is that peloruside A proved to be more effective in reducing tumor volume compared to docetaxel and paclitaxel in two lines of human non-small cell lung cancer implanted in athymic *nu/nu* mice: 74–99% of tumor growth inhibition depending on the cell line implanted against 40–50% induced by taxanes. Of note, peloruside A was better tolerated than taxanes. However, the combination therapy of peloruside A plus docetaxel or paclitaxel resulted in high toxicity and mortality [146]. In a further study performed on Pgp-overexpressing ovarian xenografted-bearing mice, peloruside A was less effective in tumor growth inhibition than doxorubicin and paclitaxel; it was, however, better tolerated, with 14% of animal death against 43% recorded for the reference drugs [146]. Currently, clinical studies are on standby due to the limited quantities of peloruside A obtained from natural matrix. To overcome this problem, much effort is therefore being made towards upscaling its synthesis [147]. Another issue to be considered is the resistance to peloruside A recorded in *in vitro* experimental settings. Resistance seems to be linked to mutations in β -tubulin and downregulation of vimentin [148,149]. Moreover, a point mutation at D297 residue resulted in resistance to peloruside A [145].

PM060184, isolated from the marine sponge *Lithoplocamia lithistoides*, is a microtubule-destabilizing agent that inhibits the addition of further tubulin subunits at the plus ends of microtubules or creates a complex with α/β -tubulin dimers [150]. Through the binding with the maytansine site, it impedes the formation of longitudinal tubulin–tubulin interactions [150], leading to disruption of microtubules and cell-cycle arrest in G2/M phase [131]. It exhibited high antiproliferative activity in 24 cancer cell lines at very low concentrations [average IC_{50} (the drug concentration that inhibits cell proliferation by 50% compared with untreated cells) = 0.7 nM] [131]. The antiproliferative activity was also confirmed in six xenograft models of human cancer. The administration of 16 mg/kg of PM060184 on days 0, 7 and 14 did not induce systemic or local toxicity and determined an overall tumor regression in four models [132]. Moreover, the antiproliferative activity was maintained in *in vitro* and *in vivo* models with Pgp-mediated chemoresistance [131].

4. Chemosensitizing Properties

The resistance to anticancer drugs is still one of the major problems that determines the failure of antineoplastic chemotherapy. Some types of tumor have an innate ability to resist antineoplastic drugs, but many others develop chemoresistance during or after therapy, resulting in refractory cancers. When resistance against drugs with different mechanisms or structures occurs, it is defined as MDR. One of the most frequent mechanisms of resistance is due to an increased efflux of drugs out of the cells, mediated by the ATP-binding cassette (ABC) transporters. Forty-nine members belong to this protein family and the three most common transporters related to MDR are: Pgp (also known as multidrug resistance protein 1, MDR1), MDR-associated protein 1 (MRP1), and breast cancer resistance protein (BCRP) [151]. Normally present in almost all tissues at low levels, these efflux pumps are expressed at higher levels in epithelial cells with secretory functions (intestine, kidney, liver, placenta, and blood-brain barrier). Their overexpression in many types of cancer leads to intrinsic drug

resistance against anticancer drugs, such as vinca alkaloids, anthracyclines, taxanes, camptothecines, and epipodophyllotoxins [152]. However, these proteins are also highly inducible in cancer tissues. For example, a single doxorubicin perfusion increased 3–15-fold MDR1 gene expression in less than 1 h in biopsies of sarcoma pulmonary metastases, whereas no alteration was noticed in normal lung tissues of the same patients [153]. All three proteins have similar substrate specificity and promote the elimination of hydrophobic compounds. Furthermore, MRP1 and BCRP extrude metabolic conjugates, differently from Pgp [154]. To counteract MDR, three classes of ABC transporter inhibitors have been synthesized [155]. *In vitro*, inhibitors of these efflux pumps are able to sensitize resistant cancer cells; however, they proved less efficient in *in vivo* models. Research is therefore still moving towards the identification of new compounds acting as chemosensitizing agents and to be administered in association with antitumoral therapy.

In the following sections, we present some examples of associations of marine spongean compounds and anticancer drugs, focusing on the mechanisms of the interaction. Experimental details and pharmacological effects of these compounds are summarized in Table 3.

Table 3. Biological effects of compounds isolated from marine sponges in association with radiotherapy or anticancer chemotherapy.

Drug Associations	Sponge	Cell Line	Concentrations (μM ^a)	CI	Biological Effect	Reference
Debromohymenialdisine (DBH) + Radiotherapy	<i>Stylissa flabeliformis</i>	MCF-7	3 (DBH) + 2–5 Gy		↓ pChk1/2, survival rate and cancer stem cell subpopulation	[156]
(+)–Discodermolide (D) + Taxol (T)		A549	0.1–5 (T) + 0.5–25 (D) (1:5 molar ratio)	0.396 ^b	↑ antiproliferative effect and aneuploidy	[157]
		MCF-7		0.273 ^b		
		SKOV-3		0.476 ^b		
(+)–Discodermolide + Paclitaxel (PT)	<i>Discodermia dissoluta</i>	A549	0.07 (D) + 0.02 (PT)	0.59 ± 0.04	Microtubules stabilization G2/M arrest apoptosis	[158]
		SKOV-3	0.001 (D) + 0.02 (T) or 0.02 (D) + 0.001 (T)	≤0.7	↑ antiproliferative effect and aneuploidy	[159]
(+)–Discodermolide + Taxol		SKOV-3 xenograft-bearing athymic (<i>nu/nu</i>) female mice	5 mg/kg (D) + 20 mg/kg (T)		↓ tumor volume and vascularization	
Fascaplysin (F) + Camptothecin (C) 10-hydroxycamptothecin (HC)	<i>Fascaplysinopsis Bergquist</i> sp.	NCI-H417	0.5 (F) + 0.5 (C)	0.53		[160]
			1 (F) + 2 (HC)	0.82		
Panicein A (PA) + Doxorubicin (Doxo)	<i>Haliclona (Soestella) mucosa</i>	MEWO	10 (PA) + 2 (Doxo)		↓ IC ₅₀ and Doxorubicin cellular efflux ↑ apoptosis	[161]
		A375	25 (PA) + 1.5 (Doxo)			
Peloruside A (P) + Paclitaxel (PT)		1A9	0.005–0.02 (P) + 0.005–0.015 (PT)	0.48–0.96	↑ G2/M arrest	[162]
		HL-60	0.015–0.03 (P) + 0.02–0.04 (PT)	0.16–0.87		
Peloruside A (P) + Etoposide (E)	<i>Mycale hentscheli</i>	1A9	0.005–0.025 (P) + 0.005–0.01 (E)	0.41–0.96	↑ G2/M arrest	[162]
		HL-60	0.02–0.125 (P) + 0.01–0.02 (E)	0.08–1.04		

CI = combination index. CI ≤ 0.7: synergy; 0.7 < CI < 1.2: additivity; CI ≥ 1.2 antagonism; ^a If not specified; ^b CI as a mean.

Lamellarin O is an alkaloid isolated from the Australian marine sponge *Ianthella* sp. It was studied as an inhibitor of ABC transporters known to induce MDR [163]. Cytotoxicity, accumulation and efflux studies were performed on human cancer cell lines (both wild-type or overexpressing one of the ABC transporters). Lamellarin O inhibited (1) Pgp in human Pgp-overexpressing colon cancer cells reversing P-gp mediated doxorubicin resistance; (2) MRP1 in human MRP1-overexpressing ovarian carcinoma cells; and (3) BCRP in human mitoxantrone-resistant non-small cell lung cancer cell lines increasing intracellular mitoxantrone [163]. Western blot and in silico docking analyses identified the BCRP inhibitory pharmacophore of lamellarin O [163].

Aragusterol A is a steroid isolated from *Xestospongia* sp. It was found to inhibit the growth of cisplatin- and doxorubicin-resistant cancer cells. For example, the cisplatin IC₅₀ in cisplatin-resistant non-small lung cancer cells was 30.3 μM (versus 0.18 μM in their parental drug-sensitive cells); the IC₅₀ of aragusterol A was 0.18 μM (versus 0.42 μM in the drug-sensitive cells). Instead, a partial cross-resistance was found in two human doxorubicin-resistant cell lines. Indeed, the doxorubicin IC₅₀ in doxorubicin-resistant leukemia cells was 5.05 μM (versus 0.10 μM in the drug-sensitive cells); the IC₅₀ of aragusterol A was 0.73 μM (versus 0.12 μM in their parental cells). Even if the resistance to doxorubicin is usually mediated by Pgp, the possible interaction between the marine steroid and Pgp was not investigated [111].

Agosterol A, a spongean sterol isolated from *Spongia* sp., was able to reverse multidrug chemoresistance to vincristine, colchicine, doxorubicin, and etoposide in human epidermoid carcinoma cells overexpressing Pgp or MRP1 [164,165]. The natural compound increased the intracellular accumulation of vincristine and reduced its efflux, restoring intracellular drug concentrations similar to those in the parental drug-sensitive cell line. The ability of agosterol A to inhibit efflux transporters was due to its direct interaction with both Pgp and MRP1 [164]. Moreover, a study revealed its ability to competitively inhibit the transport of amphipathic substrates by MRP1. Agosterol A was also able to decrease glutathione intracellular levels, lowering the formation of drug-glutathione conjugates, which are substrates of MRP1. The glutathione depletion was independent of its activity on MRP1, since a similar decrease was noticed also in the non-MRP1-overexpressing cells [165].

The sipholane triterpenoids (sipholenol A, sipholenone E, sipholenol L, and siphonellinol D), isolated from the sponge *Callyspongia siphonella*, were studied as Pgp inhibitors [166,167]. They were able to enhance the cytotoxicity of paclitaxel, colchicine and vinblastine in drug-resistant human epidermoid carcinoma cells overexpressing Pgp. No altered IC₅₀ was observed in the original clone. The triterpenoids were specific inhibitors for Pgp. In fact, in the same cell lines, they had no effect on the IC₅₀ of cisplatin, which is not a substrate of this glycoprotein. Furthermore, they did not modify the cytotoxic profile of the above reported anticancer drugs when administered to cell lines knockout for Pgp or expressing other types of resistance proteins (i.e., MDR1, MDR7, or BCRP) [166,167]. The sipholanes' ability to reverse chemoresistance and increase the intracellular accumulation of drugs is due to a decreased efflux activity of Pgp by a direct interaction with the substrate binding-site of Pgp. The compounds lack effects on Pgp expression [166,167].

Panicein A hydroquinone, isolated from the Mediterranean sponge *Haliclona* (*Soestella*) *mucosa*, was found to increase the cytotoxic and proapoptotic activity of doxorubicin in two human melanoma cell lines (MEWO metastatic cells and malignant melanoma A375 cells) [161]. Both cell lines express the protein Patched, which is the receptor for the Hedgehog signaling pathway. This pathway regulates many processes during embryonic development. The aberrant activation of this pathway is linked to tumorigenesis and metastatization as well as resistance to antitubercular drugs [168]. Patched is also a multidrug efflux pump that extrudes many drugs out of cells. The synergistic effect observed for the doxorubicin plus panicein A hydroquinone association was due to a reduced doxorubicin efflux by Patched receptor [169]. The analysis performed on a structural model of Patched identified a panicein-binding site near to that for doxorubicin, suggesting a possible interference of the marine compound with Patched drug efflux activity [161]. Since Patched is expressed prevalently in cancerous cells in adults, its inhibitors could have a higher selectivity than compounds acting on ABC

transporters, which are expressed ubiquitously. Furthermore, the cytotoxic effect against cancerous cells overexpressing a gene target of the Hedgehog pathway was not observed on non-transformed cells [170].

Fascaplysin, an alkaloid isolated from the *Fascaplysinopsis bergquist* sp. [171], induces ROS generation, cell-cycle arrest, and apoptosis [160]. It showed a synergistic cytotoxicity with the topoisomerase I inhibitors camptothecin, 10-hydroxycamptothecin and topotecan on chemoresistant NCI-H417 small-cell lung cancer. The mechanism of action is still unclear. However, fascaplysin could interfere with the topoisomerase I activity since no interaction or minor antagonism was found with other classes of antineoplastic drugs, such as DNA crosslinkers (cisplatin, carboplatin, oxaliplatin and mitomycin), topoisomerase II inhibitors (etoposide), DNA intercalating agents (doxorubicin), inhibitors of microtubules (vinblastine, and docetaxel), nucleoside analogs (gemcitabine and cytarabine) [160].

Manzamine A is an alkaloid found in the sponges *Haliclona* sp., *Pachypellina* sp., *Pellina* sp. and *Xestospongia* sp. It is endowed with various pharmacological properties, such as potential anticancer, antimicrobial, and anti-inflammatory activities [172]. This marine compound increased the sensitivity to TRAIL-apoptosis inducers in human pancreatic adenocarcinoma cells [173].

A useful strategy to improve the clinical application of an anticancer drug might be to widen its therapeutic window. Adverse effects are the limiting factors of many anticancer therapies, such as nephrotoxicity and ototoxicity for cisplatin [174,175], cardiotoxicity for doxorubicin [176] or pulmonary toxicity for bleomycin [177]. Natural compounds with selective cytoprotective abilities in non-transformed cells could therefore allow the use of higher concentrations of anticancer drugs.

Aaptamine and aeroplysinin-1 are two alkaloids derived from *Aaptos suberitoides* [178] and *Aplysina aerophoba* [179], which were studied as cytoprotective agents in rat tubular kidney cells and rat glomerular endothelial cells. Reduced cytotoxicity and genotoxicity were observed with 1–2 μ M of alkaloids only in cisplatin-treated cells; instead, no protection was noticed against oxaliplatin or doxorubicin [180]. Aaptamine showed greater cytoprotective effects compared to aeroplysinin-1 in normal cells, and was more selective than aeroplysinin-1 in human bladder carcinoma cells. Aaptamine did not alter cisplatin or oxaliplatin cytotoxicity in transformed cells, whereas aeroplysinin-1 induced a slight resistance to platinum compounds [180]. However, the cytoprotective mechanism of these spongean alkaloids is still unclear. They seem not to influence cisplatin influx/efflux because cisplatin-DNA adducts formation weren't reduced. Of note, they activated the checkpoint kinase-1 and induced a reduction of Bax, suggesting that these compounds modulate the DNA-damage response mechanisms [180].

The inhibition of G2 phase DNA-damage checkpoint (Chk1 and 2) could be a useful strategy to sensitize cancer cells to DNA-damaging anticancer drugs. DNA damage activates the phosphoinositide kinases ataxia-telangectasia muted (ATM) and ATM- and Rad3-related (ATR), which transmit the signals to Chk1 and Chk2. Chk1 and Chk2 in turn phosphorylate Wee1 kinase and Cdc25 phosphatases. Both actions lead to the inactivation of the cyclin B-CDK1 complex and G2 arrest, which favors DNA repair [181].

Debromohymenialdisine (DBH), isolated from *Stylissa flabeliformis*, is a potent inhibitor against Chk1 and Chk2, without effect on ATM or ATR in human breast cancer cells with a dominant negative mutant 53-defective. Of note, it exhibited this inhibitory activity at concentrations 8-fold lower than its IC_{50} [182]. DBH used at a non-cytotoxic concentration (3 μ M) in association with radiotherapy (2 or 5 Gy) was able to reduce Chk1/2 phosphorylation in human breast cancer cells (MCF7). The combined treatment induced a lower MCF7 survival rate compared to only radiotherapy. The effect was stable over time (24–72 h). Furthermore, DBH reduced the rate of cancer stem cell subpopulation in a time-dependent manner, reaching 65% of inhibition 8 days after irradiation [156]. However, an antagonistic effect was observed in combination with oxaliplatin in human colon cancer cells: cytotoxicity and apoptotic events as well as DNA cross-link formation induced by oxaliplatin were antagonized by DBH [183].

Usually, the strategy adopted in planning combination therapy is to select two drugs with different mechanisms of action, better if they modulate two different pathways. In the recent past, however, the association between compounds acting on the same target achieved resounding success. One example of the latter-mentioned strategy is the association of two microtubule stabilizer agents, i.e., (+)-discodermolide, extracted from *Discodermia dissoluta*, and paclitaxel. This combination therapy led in non-small cell lung cancer cells to a synergistic suppression of microtubule dynamic parameters, such as the shortening rate and length or the length-based rescue frequency (the period of growth or shortening excursions or the period of pause). In the cellular context, these events signified a synergistic increase in G2/M arrest and apoptotic events [158]. The synergism between discodermolide and taxol was also observed in human ovarian carcinoma cells. The antiproliferative effect was synergic at not equimolar ratios and at low concentrations (≤ 20 nM), which induced aneuploidy and caspase-independent cell death. Only at higher concentrations, was a mitotic arrest observed, as previously reported by Honore et al. [158,159]. Likewise, a combination therapy exploiting low doses of both drugs (discodermolide at 5 mg/kg and taxol at 20 mg/kg) showed a synergistic tumor suppression and antiangiogenic activity in ovarian cancer xenograft-bearing mice, without development of side effects [159]. It is worth highlighting that the ability of taxol to inhibit endothelial cell chemotaxins and invasiveness occurred at concentrations lower than its endothelial antiproliferative activity [184]. Of note, the association between low doses of two stabilizer agents instead of high doses of a single drug most likely means lower treatment toxicity and improvement of therapeutic compliance. The license of discodermolide was bought by Novartis, but the clinical development of the compound was halted because of its toxicity [185]. The research is now focusing on synthetic analogues or hybrid molecules [186].

5. Chemoprevention

The high systemic toxicity and the high failure rates of traditional cancer therapies have enhanced the search for new agents, which could prevent and/or slow down cancer growth and, at the same time, improve patients' compliance [187]. Indeed, carcinogenesis is a long and complex process that presents many opportunities for interventions. For many types of cancer, such as breast malignancies, the WHO specifically suggested chemoprevention as an effective therapy.

Somatic cell mutations are pivotal in tumor initiation and in the other phases of carcinogenesis. By definition, chemopreventive agents trigger protective mechanisms either inside cells or in the extracellular environment in order to prevent or inhibit DNA mutations and cancer initiation. Since 1995, it has been known that marine organisms and, in particular, marine sponges do not develop cancer. This is due to their efficient bio-transformation and detoxification systems that counteract the generation of DNA lesions induced by carcinogens [188,189]. This evidence stimulated the idea that sponges and their metabolites could exert chemopreventive activity in humans. So far, only a few compounds isolated from marine sponges have been tested for their ability to inhibit cancer initiation.

Arenosclera brasiliensis ethanolic extract is one example of a potential chemopreventive agent obtained from marine sponges. The crude extract was not genotoxic per se and prevented DNA damage [190]. Indeed, *Salmonella typhimurium* reverse mutation tests (Ames test) or direct plasmid treatments showed that the extract prevented mitomycin C, sodium azide, and 2-aminofluorene mutagenicity [190], by acting through intracellular and extracellular mechanisms and favoring DNA repair.

The antioxidant verongiaquinol and manzamine A, isolated from the red sea sponge *Aplysina* sp. and *Acanthostrongylophora* sp., respectively, are other two antimutagenic compounds. They reduced base-pair substitution, frame-shift and transitional mutagenicity induced by sodium azide and methyl methanesulfonate, thus probably acting through different mechanisms [191]. The proposed mechanisms of action involved the adsorption of mutagens [192,193] or the induction of DNA glycosylase enzymes, which repair alkylated DNA bases [194].

The ethanolic extract of *Haliclona koremella* has antioxidant and free-radical scavenger characteristics, which are responsible for its antimutagenic activity. In fact, the extract reduced the number of micronucleated polychromatic erythrocytes induced by metronidazole, a drug whose genotoxic potential is related to the induction of oxidative stress [195].

6. Clinical Studies

It is worth noting the fact that, despite being successful in preclinical models, around 95% of new anticancer drugs fail in clinical trials [196]. Furthermore, many compounds showing very promising properties in in vitro models suffer pharmacokinetic problems in animal models [197].

To date, only one anticancer drug derived from marine sponges is commercially available: eribulin mesylate (brand name Halaven). Eribulin mesylate (EM) is a synthetic analog of helicondrin B, extracted from the *Porifera Halichondria okadai* [198]. It was approved by the FDA in 2010 [199] and by the European Medicines Agency (EMA) in 2011 for the treatment of metastatic breast cancer or locally advanced breast cancer as third-line therapy after two chemotherapeutic regimens with an anthracycline and a taxane [200]. In 2014, the American Society of Clinical Oncology recommended EM as a second- and later-line chemotherapy in women with human epidermal growth factor receptor 2 (HER2)-negative advanced breast cancer [201].

EM is a microtubule destabilizing agent that increases the formation of aberrant mitotic spindles leading to an irreversible mitotic arrest [202]. It exhibited a selective activity for proliferating cells since no effect was recorded in human quiescent immortalized fibroblasts [203]. Differently from what was observed for other microtubule-depolymerizing compounds [204], EM acts on the abnormal tumor vasculature, where it increases the formation of microvessels and, therefore, tumor perfusion [205,206]. The enhanced vascularization could lead to a duplex benefit: the improved delivery of drugs into the hypoxic core of the tumor and the reduction of chemoresistance driven by hypoxia [207]. Another characteristic of EM, which could positively affect the outcome, is to reverse the epithelial-to-mesenchymal transition. During this process, epithelial cells adopt the mesenchymal phenotype, which is related to drug resistance, invasion, metastasis, and changes in stem cell phenotype [208]. EM was able to reverse this transition in three triple-negative breast cancer cell lines, decreasing their mesenchymal gene and protein profiles in favor of the epithelial one [209]. In preclinical studies, EM showed antitumor activity against many types of cancer, such as breast, ovarian, colon, pancreatic, head and neck, non-small-cell and small-cell lung cancer, leiomyosarcoma, fibrosarcoma, glioblastoma, melanoma, and acute lymphocytic leukemia [203,210,211]. In clinical trials performed on patients with locally advanced or metastatic cancer, EM increased the progression-free and overall survival compared to the reference drug, with manageable side effects, such as neutropenia, fatigue, and peripheral neuropathy [212,213].

In 2016, the FDA approved EM for the treatment of metastatic or unresectable liposarcoma as second-line chemotherapy after a prior anthracycline-containing regimen. Moreover, EM was designed as an orphan drug, encouraging its development for future applications [214]. In the phase III clinical trial on previously treated patients with advanced liposarcoma or leiomyosarcoma, EM increased overall survival compared to dacarbazine (13.5 vs. 11.5 months). However, patients developed intermediate or serious adverse effects more frequently than when treated with the reference drug (67% vs. 56%); mortality was higher than dacarbazine (4% vs. 1%) [215]. Adverse effects include neutropenia, numbness and peripheral neuropathy, and cardiomyopathy with risk of death [215].

To date, there are 131 clinical trials on EM, among them 84 regarding breast cancer (53 are ongoing) [216]. In 28 clinical trials (8 have been completed or terminated), EM was studied as a combination therapy with other antineoplastic drugs, such as carboplatin (NCT03032614), gemcitabine (NCT00410553), or irinotecan (NCT02318589), but also with new compounds like PQR309 (NCT02723877) or POL6326 (NCT01837095) [217].

E7974 is a synthetic analog of hemiasterlin, isolated from the marine sponge *Hemiasterella minor* [218]. E7974 inhibits microtubule polymerization binding mostly to the

α -subunit into the α/β -tubulin heterodimer interface. It alters the architecture of the mitotic spindles, induces G2/M arrest, and apoptosis [219]. It showed a strong antiproliferative activity against many cancer cell lines [219]. It was also effective in paclitaxel-resistant cell lines and in human xenograft-cancer models sensitive or not to paclitaxel. It does not bind to β -tubulin, the major site of mutations in taxane-resistant cell lines, and is a poor substrate of Pgp efflux pump [220,221]. So far, three clinical phase I studies with different schemes of drug administration have been completed (NCT00165802, NCT00130169, NCT00121732) [217]. They were conducted in patients with refractory solid tumors, and the disease was stabilized in 35%. The most relevant toxicity was neutropenia. However, all toxic effects were reversible and manageable [221].

Up to now, there are two phase I clinical studies on PM060184; one has been completed (NCT01299636) and one is in the recruiting status (NCT02533674) [217]. The completed clinical trial enrolled 20 patients with advanced solid tumor such as breast cancer, non-small cell lung cancer, and urothelial cancer. One patient showed partial response and three patients achieved disease stability. Severe developed toxicities were bowel obstruction, vomiting, tumor pain, peripheral motor neuropathy, neutropenia, and thrombocytopenia [222]. The ongoing clinical study will deal with the association between PM060184 and gemcitabine in patients with advanced solid tumors [217].

Considering the high social impact of cancer pathology, finding new therapeutic strategies for the treatment of late-stage or refractory tumors is essential. Taking into account the fact that such drugs should improve the quality of life of those patients that are already physically and psychologically devastated by the disease and the chemotherapy, the authorization by the FDA for these drugs has been simplified. In particular, for this category of drug products a deferral of genotoxicity studies during the initial phase of clinical drug development is authorized [223]. Usually, these compounds are characterized by a slightly better overall- or progression-free-survival than the reference-listed drug, e.g., EM versus decarbazine. Moreover, although adverse reactions are common, they are reversible and manageable [221]. However, for clinical trials enrolling patients at early stages of cancer such as EM in NCT01439282 and NCT01593020 studies, the exclusion of genotoxic and mutagenic studies should not be recommended. However, genotoxic studies have been reported for only very few natural products originated from sponges. Ingenamine G from *Pachychalina alcaloidifera* proved genotoxic in human proliferating lymphocytes [224].

7. Conclusions

The increasing knowledge of the ability of compounds isolated from marine sponges to target various events of the carcinogenetic process suggests that they could serve as new tools for both preventive and therapeutic interventions. The evidence reported in this review highlights the great value of marine sponge derivatives as a promising source of anticancer compounds. The compounds presented here evoked their potential antitumor activity by different mechanisms, such as cell-cycle arrest, anti-inflammatory activity, apoptosis, induction of ER stress, and interaction with many different targets involved in cancer development, such as mitochondrial membrane, PARP, cytochrome *c*, Akt, and caspases (Figure 1).

More than one pathway is usually affected by each sponge derivative. Aaptamines, for example, are capable of triggering apoptosis, inhibiting cell proliferation and acting as cytoprotective elements against traditional chemotherapeutic agents. This highlights their multi-target nature enabling them to effectively counteract the biological complexity of cancer. One aspect worthy of note is that some marine sponge derivatives exhibit potential anticancer effects through particular and partially novel mechanisms. An example is eribulin, which modulates microtubule dynamics through a mechanism different from other antimicrotubular chemotherapy drugs.

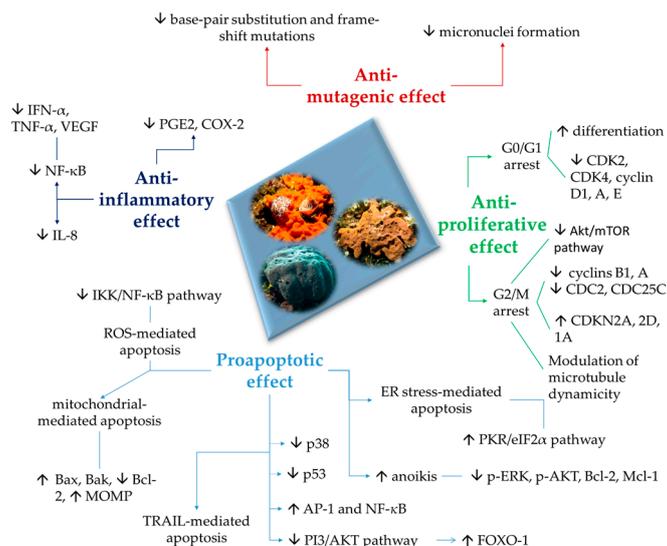


Figure 1. Pathways involved in anti-mutagenic, anti-inflammatory, anti-proliferative and proapoptotic effects of the marine compounds.

Moreover, many sponge compounds showed additive or synergistic effects when tested with traditional antitumor drugs. This strategy is very interesting since it makes it possible to lower the dose of traditional antitumor drugs, thus decreasing their toxicity and improving their therapeutic index. In addition, the combined therapy can circumvent the insurgence or overcome multidrug resistance, which still represents a significant limitation for traditional anticancer treatments. The synergy they show indicates these molecules as very interesting leads for drug development. Thus, future research should focus on the assessment and exploitation of the clinical anticancer potential of the most promising candidates isolated from marine sponges, balancing both their broad-spectrum effectiveness and their toxicity in order to carefully define their risk–benefit ratio.

Author Contributions: Carmela Fimognari and Anupam Bishayee conceived the manuscript. Cinzia Calcabrini, Elena Catanzaro, Anupam Bishayee, Eleonora Turrini and Carmela Fimognari wrote the manuscript. Carmela Fimognari and Anupam Bishayee critically revised the article for intellectual content.

Conflicts of Interest: The authors declare no conflict of interest.

References

- Newman, D.J.; Cragg, G.M.; Snader, K.M. Natural products as sources of new drugs over the period 1981–2002. *J. Nat. Prod.* **2003**, *66*, 1022–1037. [CrossRef] [PubMed]
- World Health Organization. Available online: <http://www.who.int/mediacentre/factsheets/fs297/en/> (accessed on 6 July 2017).
- Amin, A.R.; Kucuk, O.; Khuri, F.R.; Shin, D.M. Perspectives for cancer prevention with natural compounds. *J. Clin. Oncol.* **2009**, *27*, 2712–2725. [CrossRef] [PubMed]
- Sagar, S.; Kaur, M.; Minneman, K.P. Antiviral lead compounds from marine sponges. *Mar. Drugs* **2010**, *8*, 2619–2638. [CrossRef] [PubMed]
- Blunt, J.W.; Copp, B.R.; Keyzers, R.A.; Munro, M.H.; Prinsep, M.R. Marine natural products. *Nat. Prod. Rep.* **2015**, *32*, 116–211. [CrossRef] [PubMed]
- Mehbub, M.F.; Lei, J.; Franco, C.; Zhang, W. Marine sponge derived natural products between 2001 and 2010: Trends and opportunities for discovery of bioactives. *Mar. Drugs* **2014**, *12*, 4539–4577. [CrossRef] [PubMed]
- Da Rocha, A.B.; Lopes, R.M.; Schwartzmann, G. Natural products in anticancer therapy. *Curr. Opin. Pharmacol.* **2001**, *1*, 364–369. [CrossRef]

8. Essack, M.; Bajic, V.B.; Archer, J.A. Recently confirmed apoptosis-inducing lead compounds isolated from marine sponge of potential relevance in cancer treatment. *Mar. Drugs* **2011**, *9*, 1580–1606. [[CrossRef](#)] [[PubMed](#)]
9. Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. *Cell* **2011**, *144*, 646–674. [[CrossRef](#)] [[PubMed](#)]
10. Vanden Berghe, T.; Kaiser, W.J.; Bertrand, M.J.; Vandenabeele, P. Molecular crosstalk between apoptosis, necroptosis, and survival signaling. *Mol. Cell. Oncol.* **2015**, *2*, e975093. [[CrossRef](#)] [[PubMed](#)]
11. Walczak, H. Death receptor-ligand systems in cancer, cell death, and inflammation. *Cold Spring Harb. Perspect. Biol.* **2013**, *5*, a008698. [[CrossRef](#)] [[PubMed](#)]
12. Prasad, S.; Ravindran, J.; Aggarwal, B.B. Nf- κ B and cancer: How intimate is this relationship. *Mol. Cell. Biochem.* **2010**, *336*, 25–37. [[CrossRef](#)] [[PubMed](#)]
13. Naugler, W.E.; Karin, M. Nf- κ B and cancer-identifying targets and mechanisms. *Curr. Opin. Genet. Dev.* **2008**, *18*, 19–26. [[CrossRef](#)] [[PubMed](#)]
14. Dyshlovoy, S.A.; Fedorov, S.N.; Shubina, L.K.; Kuzmich, A.S.; Bokemeyer, C.; Keller-von Amsberg, G.; Honecker, F. Aaptamines from the marine sponge *Aaptos* sp. Display anticancer activities in human cancer cell lines and modulate ap-1-, Nf- κ B-, and p53-dependent transcriptional activity in mouse j6 cl41 cells. *Biomed. Res. Int.* **2014**, *2014*, 469309. [[CrossRef](#)] [[PubMed](#)]
15. Guzman, E.A.; Maers, K.; Roberts, J.; Kemami-Wangun, H.V.; Harmody, D.; Wright, A.E. The marine natural product microsclerodermin a is a novel inhibitor of the nuclear factor kappa B and induces apoptosis in pancreatic cancer cells. *Investig. New Drugs* **2015**, *33*, 86–94. [[CrossRef](#)] [[PubMed](#)]
16. Florean, C.; Schnekenburger, M.; Lee, J.Y.; Kim, K.R.; Mazumder, A.; Song, S.; Kim, J.M.; Grandjenette, C.; Kim, J.G.; Yoon, A.Y.; et al. Discovery and characterization of isofistularin-3, a marine brominated alkaloid, as a new DNA demethylating agent inducing cell cycle arrest and sensitization to trail in cancer cells. *Oncotarget* **2016**, *7*, 24027–24049. [[CrossRef](#)] [[PubMed](#)]
17. Guzman, E.; Maher, M.; Temkin, A.; Pitts, T.; Wright, A. Spongiatriol inhibits nuclear factor kappa B activation and induces apoptosis in pancreatic cancer cells. *Mar. Drugs* **2013**, *11*, 1140–1151. [[CrossRef](#)] [[PubMed](#)]
18. Mooberry, S.L.; Tien, G.; Hernandez, A.H.; Plubrukarn, A.; Davidson, B.S. Laulimalide and isolaulimalide, new paclitaxel-like microtubule-stabilizing agents. *Cancer Res.* **1999**, *59*, 653–660. [[PubMed](#)]
19. De Stefano, D.; Tommonaro, G.; Malik, S.A.; Iodice, C.; De Rosa, S.; Maiuri, M.C.; Carnuccio, R. Cacospongionolide and scalaradial, two marine sesterterpenoids as potent apoptosis-inducing factors in human carcinoma cell lines. *PLoS ONE* **2012**, *7*, e33031. [[CrossRef](#)] [[PubMed](#)]
20. Trisciuglio, D.; Uranchimeg, B.; Cardellina, J.H.; Meragelman, T.L.; Matsunaga, S.; Fusetani, N.; Del Bufalo, D.; Shoemaker, R.H.; Melillo, G. Induction of apoptosis in human cancer cells by candidaspongiolide, a novel sponge polyketide. *J. Natl. Cancer Inst.* **2008**, *100*, 1233–1246. [[CrossRef](#)] [[PubMed](#)]
21. Umeyama, A.; Matsuoka, N.; Mine, R.; Nakata, A.; Arimoto, E.; Matsui, M.; Shoji, N.; Arihara, S.; Takei, M.; Hashimoto, T. Polyacetylene diols with antiproliferative and driving Th1 polarization effects from the marine sponge *Callyspongia* sp. *J. Nat. Med.* **2010**, *64*, 93–97. [[CrossRef](#)] [[PubMed](#)]
22. Roel, M.; Rubiolo, J.A.; Guerra-Varela, J.; Silva, S.B.; Thomas, O.P.; Cabezas-Sainz, P.; Sanchez, L.; Lopez, R.; Botana, L.M. Marine guanidine alkaloids crambescidins inhibit tumor growth and activate intrinsic apoptotic signaling inducing tumor regression in a colorectal carcinoma zebrafish xenograft model. *Oncotarget* **2016**, *7*, 83071–83087. [[CrossRef](#)] [[PubMed](#)]
23. Kong, D.; Aoki, S.; Sowa, Y.; Sakai, T.; Kobayashi, M. Smenospongine, a sesquiterpene aminoquinone from a marine sponge, induces g1 arrest or apoptosis in different leukemia cells. *Mar. Drugs* **2008**, *6*, 480–488. [[PubMed](#)]
24. Shin, D.Y.; Kim, G.Y.; Kim, N.D.; Jung, J.H.; Kim, S.K.; Kang, H.S.; Choi, Y.H. Induction of apoptosis by pectenotoxin-2 is mediated with the induction of DR4/DR5, EGR-1 and NAG-1, activation of caspases and modulation of the Bcl-2 family in p53-deficient Hep3B hepatocellular carcinoma cells. *Oncol. Rep.* **2008**, *19*, 517–526. [[CrossRef](#)] [[PubMed](#)]
25. Kim, M.O.; Moon, D.O.; Heo, M.S.; Lee, J.D.; Jung, J.H.; Kim, S.K.; Choi, Y.H.; Kim, G.Y. Pectenotoxin-2 abolishes constitutively activated Nf- κ B, leading to suppression of Nf- κ B related gene products and potentiation of apoptosis. *Cancer Lett.* **2008**, *271*, 25–33. [[CrossRef](#)] [[PubMed](#)]

26. Moon, D.O.; Kim, M.O.; Kang, S.H.; Lee, K.J.; Heo, M.S.; Choi, K.S.; Choi, Y.H.; Kim, G.Y. Induction of G2/M arrest, endoreduplication, and apoptosis by actin depolymerization agent pectenotoxin-2 in human leukemia cells, involving activation of erk and jnk. *Biochem. Pharmacol.* **2008**, *76*, 312–321. [[CrossRef](#)] [[PubMed](#)]
27. Ben-Califa, N.; Bishara, A.; Kashman, Y.; Neumann, D. Salarin c, a member of the salarin superfamily of marine compounds, is a potent inducer of apoptosis. *Investig. New Drugs* **2012**, *30*, 98–104. [[CrossRef](#)] [[PubMed](#)]
28. Del Poggetto, E.; Tanturli, M.; Ben-Califa, N.; Gozzini, A.; Tusa, I.; Cheloni, G.; Marzi, I.; Cipolleschi, M.G.; Kashman, Y.; Neumann, D.; et al. Salarin C inhibits the maintenance of chronic myeloid leukemia progenitor cells. *Cell Cycle* **2015**, *14*, 3146–3154. [[CrossRef](#)] [[PubMed](#)]
29. Zhang, Y.W.; Ghosh, A.K.; Pommier, Y. Lasonolide a, a potent and reversible inducer of chromosome condensation. *Cell Cycle* **2012**, *11*, 4424–4435. [[CrossRef](#)] [[PubMed](#)]
30. Cheung, F.W.; Li, C.; Che, C.T.; Liu, B.P.; Wang, L.; Liu, W.K. Geoditin a induces oxidative stress and apoptosis on human colon HT29 cells. *Mar. Drugs* **2010**, *8*, 80–90. [[CrossRef](#)] [[PubMed](#)]
31. Liu, W.K.; Ho, J.C.; Che, C.T. Apoptotic activity of isomalabaricane triterpenes on human promyelocytic leukemia hl60 cells. *Cancer Lett.* **2005**, *230*, 102–110. [[CrossRef](#)] [[PubMed](#)]
32. Liu, W.K.; Cheung, F.W.; Che, C.T. Stelletin a induces oxidative stress and apoptosis in hl-60 human leukemia and Incap prostate cancer cell lines. *J. Nat. Prod.* **2006**, *69*, 934–937. [[CrossRef](#)] [[PubMed](#)]
33. Do, M.T.; Na, M.; Kim, H.G.; Khanal, T.; Choi, J.H.; Jin, S.W.; Oh, S.H.; Hwang, I.H.; Chung, Y.C.; Kim, H.S.; et al. Ilimaquinone induces death receptor expression and sensitizes human colon cancer cells to trail-induced apoptosis through activation of ros-erk/p38 mapk-chop signaling pathways. *Food Chem. Toxicol.* **2014**, *71*, 51–59. [[CrossRef](#)] [[PubMed](#)]
34. Lee, H.Y.; Chung, K.J.; Hwang, I.H.; Gwak, J.; Park, S.; Ju, B.G.; Yun, E.; Kim, D.E.; Chung, Y.H.; Na, M.; et al. Activation of p53 with ilimaquinone and ethylsmenoquinone, marine sponge metabolites, induces apoptosis and autophagy in colon cancer cells. *Mar. Drugs* **2015**, *13*, 543–557. [[CrossRef](#)] [[PubMed](#)]
35. Lu, P.H.; Chueh, S.C.; Kung, F.L.; Pan, S.L.; Shen, Y.C.; Guh, J.H. Ilimaquinone, a marine sponge metabolite, displays anticancer activity via gadd153-mediated pathway. *Eur. J. Pharmacol.* **2007**, *556*, 45–54. [[CrossRef](#)] [[PubMed](#)]
36. Schneiders, U.M.; Schyschka, L.; Rudy, A.; Vollmar, A.M. Bh3-only proteins mcl-1 and bim as well as endonuclease g are targeted in spongistatin 1-induced apoptosis in breast cancer cells. *Mol. Cancer Ther.* **2009**, *8*, 2914–2925. [[CrossRef](#)] [[PubMed](#)]
37. Schyschka, L.; Rudy, A.; Jeremias, I.; Barth, N.; Pettit, G.R.; Vollmar, A.M. Spongistatin 1: A new chemosensitizing marine compound that degrades xiap. *Leukemia* **2008**, *22*, 1737–1745. [[CrossRef](#)] [[PubMed](#)]
38. Rothmeier, A.S.; Schneiders, U.M.; Wiedmann, R.M.; Ischenko, I.; Bruns, C.J.; Rudy, A.; Zahler, S.; Vollmar, A.M. The marine compound spongistatin 1 targets pancreatic tumor progression and metastasis. *Int. J. Cancer* **2010**, *127*, 1096–1105. [[CrossRef](#)] [[PubMed](#)]
39. Schumacher, M.; Cerella, C.; Eifes, S.; Chateauvieux, S.; Morceau, F.; Jaspars, M.; Dicato, M.; Diederich, M. Heteronemin, a spongean sesterterpene, inhibits tnf alpha-induced NF-κB activation through proteasome inhibition and induces apoptotic cell death. *Biochem. Pharmacol.* **2010**, *79*, 610–622. [[CrossRef](#)] [[PubMed](#)]
40. Wu, J.C.; Wang, C.T.; Hung, H.C.; Wu, W.J.; Wu, D.C.; Chang, M.C.; Sung, P.J.; Chou, Y.W.; Wen, Z.H.; Tai, M.H. Heteronemin is a novel c-Met/STAT3 inhibitor against advanced prostate cancer cells. *Prostate* **2016**, *76*, 1469–1483. [[CrossRef](#)] [[PubMed](#)]
41. Huang, H.H.; Kuo, S.M.; Wu, Y.J.; Su, J.H. Improvement and enhancement of antibladder carcinoma cell effects of heteronemin by the nanosized hyaluronan aggregation. *Int. J. Nanomed.* **2016**, *11*, 1237–1251. [[CrossRef](#)] [[PubMed](#)]
42. Wu, S.Y.; Sung, P.J.; Chang, Y.L.; Pan, S.L.; Teng, C.M. Heteronemin, a spongean sesterterpene, induces cell apoptosis and autophagy in human renal carcinoma cells. *Biomed. Res. Int.* **2015**, *2015*, 738241. [[CrossRef](#)] [[PubMed](#)]
43. Aoki, S.; Cho, S.H.; Ono, M.; Kuwano, T.; Nakao, S.; Kuwano, M.; Nakagawa, S.; Gao, J.Q.; Mayumi, T.; Shibuya, M.; et al. Bastadin 6, a spongean brominated tyrosine derivative, inhibits tumor angiogenesis by inducing selective apoptosis to endothelial cells. *Anticancer Drugs* **2006**, *17*, 269–278. [[CrossRef](#)] [[PubMed](#)]
44. Chinen, T.; Nagumo, Y.; Watanabe, T.; Imaizumi, T.; Shibuya, M.; Kataoka, T.; Kanoh, N.; Iwabuchi, Y.; Usui, T. Irciniastatin a induces jnk activation that is involved in caspase-8-dependent apoptosis via the mitochondrial pathway. *Toxicol. Lett.* **2010**, *199*, 341–346. [[CrossRef](#)] [[PubMed](#)]

45. Wei, S.Y.; Li, M.; Tang, S.A.; Sun, W.; Xu, B.; Cui, J.R.; Lin, W.H. Induction of apoptosis accompanying with g(1) phase arrest and microtubule disassembly in human hepatoma cells by jaspolide b, a new isomalabaricane-type triterpene. *Cancer Lett.* **2008**, *262*, 114–122. [[CrossRef](#)] [[PubMed](#)]
46. Chen, Y.; Zhou, Q.; Zhang, L.; Zhong, Y.; Fan, G.; Zhang, Z.; Wang, R.; Jin, M.; Qiu, Y.; Kong, D. Stelletin b induces apoptosis in human chronic myeloid leukemia cells via targeting pi3k and stat5. *Oncotarget* **2017**, *8*, 28906–28921. [[CrossRef](#)] [[PubMed](#)]
47. Wang, R.; Zhang, Q.; Peng, X.; Zhou, C.; Zhong, Y.; Chen, X.; Qiu, Y.; Jin, M.; Gong, M.; Kong, D. Stelletin b induces g1 arrest, apoptosis and autophagy in human non-small cell lung cancer A549 cells via blocking PI3K/Akt/mTOR pathway. *Sci. Rep.* **2016**, *6*, 27071. [[CrossRef](#)] [[PubMed](#)]
48. Tang, S.A.; Zhou, Q.; Guo, W.Z.; Qiu, Y.; Wang, R.; Jin, M.; Zhang, W.; Li, K.; Yamori, T.; Dan, S.; et al. In vitro antitumor activity of stelletin b, a triterpene from marine sponge jaspis stellifera, on human glioblastoma cancer sf295 cells. *Mar. Drugs* **2014**, *12*, 4200–4213. [[CrossRef](#)] [[PubMed](#)]
49. Salma, Y.; Lafont, E.; Therville, N.; Carpentier, S.; Bonnafant, M.J.; Levade, T.; Genisson, Y.; Andrieu-Abadie, N. The natural marine anhydrophytosphingosine, jaspine b, induces apoptosis in melanoma cells by interfering with ceramide metabolism. *Biochem. Pharmacol.* **2009**, *78*, 477–485. [[CrossRef](#)] [[PubMed](#)]
50. Yoo, H.; Lee, Y.S.; Lee, S.; Kim, S.; Kim, T.Y. Pachastrissamine from *Pachastrissa* sp. Inhibits melanoma cell growth by dual inhibition of Cdk2 and erk-mediated foxo3 downregulation. *Phytother. Res.* **2012**, *26*, 1927–1933. [[CrossRef](#)] [[PubMed](#)]
51. Nguyen, H.T.; Chau, V.M.; Tran, T.H.; Phan, V.K.; Hoang, T.H.; Nguyen, T.D.; Nguyen, X.N.; Tai, B.H.; Hyun, J.H.; Kang, H.K.; et al. C29 sterols with a cyclopropane ring at C-25 and 26 from the vietnamese marine sponge *ianthella* sp. In addition, their anticancer properties. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 4584–4588. [[PubMed](#)]
52. Guzman, E.A.; Xu, Q.; Pitts, T.P.; Mitsushashi, K.O.; Baker, C.; Linley, P.A.; Oestreicher, J.; Tendyke, K.; Winder, P.L.; Suh, E.M.; et al. Leiodermatolide, a novel marine natural product, has potent cytotoxic and antimetastatic activity against cancer cells, appears to affect microtubule dynamics, and exhibits antitumor activity. *Int. J. Cancer* **2016**, *139*, 2116–2126. [[CrossRef](#)] [[PubMed](#)]
53. LaBarbera, D.V.; Modzelewska, K.; Glazar, A.I.; Gray, P.D.; Kaur, M.; Liu, T.; Grossman, D.; Harper, M.K.; Kuwada, S.K.; Moghal, N.; et al. The marine alkaloid naamidine a promotes caspase-dependent apoptosis in tumor cells. *Anticancer Drugs* **2009**, *20*, 425–436. [[CrossRef](#)] [[PubMed](#)]
54. Dyshlovoy, S.A.; Tabakmakher, K.M.; Hauschild, J.; Shchekaleva, R.K.; Otte, K.; Guzii, A.G.; Makarieva, T.N.; Kudryashova, E.K.; Fedorov, S.N.; Shubina, L.K.; et al. Guanidine alkaloids from the marine sponge monanchora pulchra show cytotoxic properties and prevent egf-induced neoplastic transformation in vitro. *Mar. Drugs* **2016**, *14*, 133. [[CrossRef](#)] [[PubMed](#)]
55. Hood, K.A.; West, L.M.; Northcote, P.T.; Berridge, M.V.; Miller, J.H. Induction of apoptosis by the marine sponge (mycale) metabolites, mycalamide a and pateamine. *Apoptosis* **2001**, *6*, 207–219. [[CrossRef](#)] [[PubMed](#)]
56. Konishi, H.; Kikuchi, S.; Ochiai, T.; Ikoma, H.; Kubota, T.; Ichikawa, D.; Fujiwara, H.; Okamoto, K.; Sakakura, C.; Sonoyama, T.; et al. Latrunculin a has a strong anticancer effect in a peritoneal dissemination model of human gastric cancer in mice. *Anticancer Res.* **2009**, *29*, 2091–2097. [[PubMed](#)]
57. Kijjoa, A.; Wattanadilok, R.; Campos, N.; Nascimento, M.S.; Pinto, M.; Herz, W. Anticancer activity evaluation of kuanoniamines a and c isolated from the marine sponge oceanapia sagittaria, collected from the gulf of thailand. *Mar. Drugs* **2007**, *5*, 6–22. [[CrossRef](#)] [[PubMed](#)]
58. Choi, H.J.; Bae, S.J.; Kim, N.D.; Jung, J.H.; Choi, Y.H. Induction of apoptosis by dideoxypetrosynol a, a polyacetylene from the sponge *petrosia* sp., in human skin melanoma cells. *Int. J. Mol. Med.* **2004**, *14*, 1091–1096. [[CrossRef](#)] [[PubMed](#)]
59. Ahn, M.Y.; Jung, J.H.; Na, Y.J.; Kim, H.S. A natural histone deacetylase inhibitor, psammaphin a, induces cell cycle arrest and apoptosis in human endometrial cancer cells. *Gynecol. Oncol.* **2008**, *108*, 27–33. [[CrossRef](#)] [[PubMed](#)]
60. Berry, E.; Hardt, J.L.; Clardy, J.; Lurain, J.R.; Kim, J.J. Induction of apoptosis in endometrial cancer cells by psammaphysene a involves foxo1. *Gynecol. Oncol.* **2009**, *112*, 331–336. [[CrossRef](#)] [[PubMed](#)]
61. Su, J.H.; Chen, Y.C.; El-Shazly, M.; Du, Y.C.; Su, C.W.; Tsao, C.W.; Liu, L.L.; Chou, Y.; Chang, W.B.; Su, Y.D.; et al. Towards the small and the beautiful: A small dibromotyrosine derivative from *Pseudoceratina* sp. sponge exhibits potent apoptotic effect through targeting IKK/NFκB signaling pathway. *Mar. Drugs* **2013**, *11*, 3168–3185. [[CrossRef](#)] [[PubMed](#)]

62. Guo, J.F.; Zhou, J.M.; Zhang, Y.; Deng, R.; Liu, J.N.; Feng, G.K.; Liu, Z.C.; Xiao, D.J.; Deng, S.Z.; Zhu, X.F. Rhabdastrellin acid-a inhibited pi3k/akt pathway and induced apoptosis in human leukemia hl-60 cells. *Cell. Biol. Int.* **2008**, *32*, 48–54. [[CrossRef](#)] [[PubMed](#)]
63. Jin, J.O.; Shastina, V.; Park, J.I.; Han, J.Y.; Makarieva, T.; Fedorov, S.; Rasskazov, V.; Stonik, V.; Kwak, J.Y. Differential induction of apoptosis of leukemic cells by rhizochalin, two headed sphingolipids from sponge and its derivatives. *Biol. Pharm. Bull.* **2009**, *32*, 955–962. [[CrossRef](#)] [[PubMed](#)]
64. Khanal, P.; Kang, B.S.; Yun, H.J.; Cho, H.G.; Makarieva, T.N.; Choi, H.S. Aglycon of rhizochalin from the rhizochalina incrustata induces apoptosis via activation of amp-activated protein kinase in ht-29 colon cancer cells. *Biol. Pharm. Bull.* **2011**, *34*, 1553–1558. [[CrossRef](#)] [[PubMed](#)]
65. Fedorov, S.N.; Makarieva, T.N.; Guzii, A.G.; Shubina, L.K.; Kwak, J.Y.; Stonik, V.A. Marine two-headed sphingolipid-like compound rhizochalin inhibits egf-induced transformation of jb6 p+ cl41 cells. *Lipids* **2009**, *44*, 777–785. [[CrossRef](#)] [[PubMed](#)]
66. Dyshlovoy, S.A.; Otte, K.; Alsdorf, W.H.; Hauschild, J.; Lange, T.; Venz, S.; Bauer, C.K.; Bähring, R.; Amann, K.; Mandanchi, R.; et al. Marine compound rhizochalinin shows high in vitro and in vivo efficacy in castration resistant prostate cancer. *Oncotarget* **2016**, *7*, 69703–69717. [[CrossRef](#)] [[PubMed](#)]
67. Choi, H.J.; Choi, Y.H.; Yee, S.B.; Im, E.; Jung, J.H.; Kim, N.D. Ircinin-1 induces cell cycle arrest and apoptosis in sk-mel-2 human melanoma cells. *Mol. Carcinog.* **2005**, *44*, 162–173. [[CrossRef](#)] [[PubMed](#)]
68. Abdel-Lateff, A.; Al-Abd, A.M.; Alahdal, A.M.; Alarif, W.M.; Ayyad, S.E.; Al-Lihaibi, S.S.; Hegazy, M.E.; Al Mohammadi, A.; Abdelghany, T.M.; Abdel-Naim, A.B.; et al. Antiproliferative effects of triterpenoidal derivatives, obtained from the marine sponge *Siphonochalina* sp., on human hepatic and colorectal cancer cells. *Z. Naturforsch. C* **2016**, *71*, 29–35. [[CrossRef](#)] [[PubMed](#)]
69. Sobahi, T.R.A.; Ayyad, S.N.; Abdel-Lateff, A.; Algandaby, M.M.; Alorfi, H.S.; Abdel-Naim, A.B. Cytotoxic metabolites from callispongia siphonella display antiproliferative activity by inducing apoptosis in hct-116 cells. *Pharmacogn. Mag.* **2017**, *13*, S37–S40. [[PubMed](#)]
70. Teta, R.; Irollo, E.; Della Sala, G.; Pirozzi, G.; Mangoni, A.; Costantino, V. Smenamides a and b, chlorinated peptide/polyketide hybrids containing a dolapyrrolidinone unit from the caribbean sponge *Smenospongia aurea*. Evaluation of their role as leads in antitumor drug research. *Mar. Drugs* **2013**, *11*, 4451–4463. [[CrossRef](#)] [[PubMed](#)]
71. Park, C.; Kim, G.Y.; Kim, W.I.; Hong, S.H.; Park, D.I.; Kim, N.D.; Bae, S.J.; Jung, J.H.; Choi, Y.H. Induction of apoptosis by (z)-stellettin acid c, an acetylenic acid from the sponge *Stelletta* sp., is associated with inhibition of telomerase activity in human leukemic u937 cells. *Chemotherapy* **2007**, *53*, 160–168. [[CrossRef](#)] [[PubMed](#)]
72. Halim, H.; Chunchacha, P.; Suwanborirux, K.; Chanvorachote, P. Anticancer and antimetastatic activities of renieramycin m, a marine tetrahydroisoquinoline alkaloid, in human non-small cell lung cancer cells. *Anticancer Res.* **2011**, *31*, 193–201. [[PubMed](#)]
73. Tabunoki, H.; Saito, N.; Suwanborirux, K.; Charupant, K.; Satoh, J. Molecular network profiling of u373mg human glioblastoma cells following induction of apoptosis by novel marine-derived anti-cancer 1,2,3,4-tetrahydroisoquinoline alkaloids. *Cancer Cell. Int.* **2012**, *12*, 14. [[CrossRef](#)] [[PubMed](#)]
74. Gordon, E.M.; Sankhala, K.K.; Chawla, N.; Chawla, S.P. Trabectedin for soft tissue sarcoma: Current status and future perspectives. *Adv. Ther.* **2016**, *33*, 1055–1071. [[CrossRef](#)] [[PubMed](#)]
75. FDA. Fda Approves New Therapy for Certain Types of Advanced Soft Tissue Sarcoma. Available online: <https://www.fda.gov/newsevents/newsroom/pressannouncements/ucm468832.htm> (accessed on 18 September 2017).
76. Charupant, K.; Daikuhara, N.; Saito, E.; Amnuoyopol, S.; Suwanborirux, K.; Owa, T.; Saito, N. Chemistry of renieramycins. Part 8: Synthesis and cytotoxicity evaluation of renieramycin m-jorunnamycin a analogues. *Bioorg. Med. Chem.* **2009**, *17*, 4548–4558. [[CrossRef](#)] [[PubMed](#)]
77. Saito, N.; Tanaka, C.; Koizumi, Y.-I.; Suwanborirux, K.; Amnuoyopol, S.; Pummangura, S.; Kubo, A. Chemistry of renieramycins. Part 6: Transformation of renieramycin m into jorumycin and renieramycin j including oxidative degradation products, mimosamycin, renierone, and renierol acetate. *Tetrahedron* **2004**, *60*, 3873–3881. [[CrossRef](#)]
78. Frisch, S.M.; Francis, H. Disruption of epithelial cell-matrix interactions induces apoptosis. *J. Cell. Biol.* **1994**, *124*, 619–626. [[CrossRef](#)] [[PubMed](#)]
79. Sirimangkalakitti, N.; Chamni, S.; Suwanborirux, K.; Chanvorachote, P. Renieramycin m sensitizes anoikis-resistant h460 lung cancer cells to anoikis. *Anticancer Res.* **2016**, *36*, 1665–1671. [[PubMed](#)]

80. Sirimangkalakitti, N.; Chamni, S.; Suwanborirux, K.; Chanvorachote, P. Renieramycin m attenuates cancer stem cell-like phenotypes in h460 lung cancer cells. *Anticancer Res.* **2017**, *37*, 615–621. [[CrossRef](#)] [[PubMed](#)]
81. Eferl, R.; Wagner, E.F. Ap-1: A double-edged sword in tumorigenesis. *Nat. Rev. Cancer* **2003**, *3*, 859–868. [[CrossRef](#)] [[PubMed](#)]
82. Gopalakrishnan, A.; Tony Kong, A.N. Anticarcinogenesis by dietary phytochemicals: Cytoprotection by nrf2 in normal cells and cytotoxicity by modulation of transcription factors NF- κ B and ap-1 in abnormal cancer cells. *Food. Chem. Toxicol.* **2008**, *46*, 1257–1270. [[CrossRef](#)] [[PubMed](#)]
83. Kasibhatla, S.; Brunner, T.; Genestier, L.; Echeverri, F.; Mahboubi, A.; Green, D.R. DNA damaging agents induce expression of fas ligand and subsequent apoptosis in t lymphocytes via the activation of NF- κ B and ap-1. *Mol. Cell.* **1998**, *1*, 543–551. [[CrossRef](#)]
84. Lu, H.; Huang, H. Foxo1: A potential target for human diseases. *Curr. Drug Targets* **2011**, *12*, 1235–1244. [[CrossRef](#)] [[PubMed](#)]
85. Biggs, W.H., 3rd; Meisenhelder, J.; Hunter, T.; Cavenee, W.K.; Arden, K.C. Protein kinase b/akt-mediated phosphorylation promotes nuclear exclusion of the winged helix transcription factor fkhrl. *Proc. Natl. Acad. Sci. USA* **1999**, *96*, 7421–7426. [[CrossRef](#)] [[PubMed](#)]
86. Brunet, A.; Bonni, A.; Zigmond, M.J.; Lin, M.Z.; Juo, P.; Hu, L.S.; Anderson, M.J.; Arden, K.C.; Blenis, J.; Greenberg, M.E. Akt promotes cell survival by phosphorylating and inhibiting a forkhead transcription factor. *Cell* **1999**, *96*, 857–868. [[CrossRef](#)]
87. Tashiro, H.; Blazes, M.S.; Wu, R.; Cho, K.R.; Bose, S.; Wang, S.I.; Li, J.; Parsons, R.; Ellenson, L.H. Mutations in pten are frequent in endometrial carcinoma but rare in other common gynecological malignancies. *Cancer Res.* **1997**, *57*, 3935–3940. [[PubMed](#)]
88. Goto, T.; Takano, M.; Albergaria, A.; Briese, J.; Pomeranz, K.M.; Cloke, B.; Fusi, L.; Feroze-Zaidi, F.; Maywald, N.; Sajin, M.; et al. Mechanism and functional consequences of loss of foxo1 expression in endometrioid endometrial cancer cells. *Oncogene* **2008**, *27*, 9–19. [[CrossRef](#)] [[PubMed](#)]
89. Ward, E.C.; Hoekstra, A.V.; Blok, L.J.; Hanifi-Moghaddam, P.; Lurain, J.R.; Singh, D.K.; Buttin, B.M.; Schink, J.C.; Kim, J.J. The regulation and function of the forkhead transcription factor, forkhead box O1, is dependent on the progesterone receptor in endometrial carcinoma. *Endocrinology* **2008**, *149*, 1942–1950. [[CrossRef](#)] [[PubMed](#)]
90. Hutti, J.E.; Pfefferle, A.D.; Russell, S.C.; Sircar, M.; Perou, C.M.; Baldwin, A.S. Oncogenic pi3k mutations lead to Nf- κ B-dependent cytokine expression following growth factor deprivation. *Cancer Res.* **2012**, *72*, 3260–3269. [[CrossRef](#)] [[PubMed](#)]
91. Lampiasi, N.; Azzolina, A.; Umezawa, K.; Montalto, G.; McCubrey, J.A.; Cervello, M. The novel Nf- κ B inhibitor dhmeq synergizes with celecoxib to exert antitumor effects on human liver cancer cells by a ros-dependent mechanism. *Cancer Lett.* **2012**, *322*, 35–44. [[CrossRef](#)] [[PubMed](#)]
92. Madonna, G.; Ullman, C.D.; Gentilcore, G.; Palmieri, G.; Ascierto, P.A. Nf- κ B as potential target in the treatment of melanoma. *J. Transl. Med.* **2012**, *10*, 53. [[CrossRef](#)] [[PubMed](#)]
93. Rasheva, V.I.; Domingos, P.M. Cellular responses to endoplasmic reticulum stress and apoptosis. *Apoptosis* **2009**, *14*, 996–1007. [[CrossRef](#)] [[PubMed](#)]
94. Szegezdi, E.; Fitzgerald, U.; Samali, A. Caspase-12 and er-stress-mediated apoptosis: The story so far. *Ann. N. Y. Acad. Sci.* **2003**, *1010*, 186–194. [[CrossRef](#)] [[PubMed](#)]
95. Nakagawa, T.; Zhu, H.; Morishima, N.; Li, E.; Xu, J.; Yankner, B.A.; Yuan, J. Caspase-12 mediates endoplasmic-reticulum-specific apoptosis and cytotoxicity by amyloid-beta. *Nature* **2000**, *403*, 98–103. [[CrossRef](#)] [[PubMed](#)]
96. Smith, W.W.; Jiang, H.; Pei, Z.; Tanaka, Y.; Morita, H.; Sawa, A.; Dawson, V.L.; Dawson, T.M.; Ross, C.A. Endoplasmic reticulum stress and mitochondrial cell death pathways mediate A53T mutant alpha-synuclein-induced toxicity. *Hum. Mol. Genet.* **2005**, *14*, 3801–3811. [[CrossRef](#)] [[PubMed](#)]
97. Bayazit, V. Cytotoxic effects of some animal and vegetable extracts and some chemicals on liver and colon carcinoma and myosarcoma. *Saudi Med. J.* **2004**, *25*, 156–163. [[PubMed](#)]
98. Costantini, S.; Romano, G.; Rusolo, F.; Capone, F.; Guerriero, E.; Colonna, G.; Ianora, A.; Ciliberto, G.; Costantini, M. Anti-inflammatory effects of a methanol extract from the marine sponge *Geodia cydonium* on the human breast cancer MCF-7 cell line. *Mediat. Inflamm.* **2015**, *2015*, 204975. [[CrossRef](#)] [[PubMed](#)]

99. Park, C.; Jung, J.H.; Kim, N.D.; Choi, Y.H. Inhibition of cyclooxygenase-2 and telomerase activities in human leukemia cells by dideoxypetrosynol a, a polyacetylene from the marine sponge *Petrosia* sp. *Int. J. Oncol.* **2007**, *30*, 291–298. [[CrossRef](#)] [[PubMed](#)]
100. Gately, S.; Kerbel, R. Therapeutic potential of selective cyclooxygenase-2 inhibitors in the management of tumor angiogenesis. *Prog. Exp. Tumor Res.* **2003**, *37*, 179–192. [[PubMed](#)]
101. Smith, W.L.; Garavito, R.M.; DeWitt, D.L. Prostaglandin endoperoxide synthases (cyclooxygenases)-1 and -2. *J. Biol. Chem.* **1996**, *271*, 33157–33160. [[CrossRef](#)] [[PubMed](#)]
102. Cao, Y.; Prescott, S.M. Many actions of cyclooxygenase-2 in cellular dynamics and in cancer. *J. Cell. Physiol.* **2002**, *190*, 279–286. [[CrossRef](#)] [[PubMed](#)]
103. Waugh, D.J.; Wilson, C. The interleukin-8 pathway in cancer. *Clin. Cancer Res.* **2008**, *14*, 6735–6741. [[CrossRef](#)] [[PubMed](#)]
104. Guzman, E.A.; Harmody, D.; Pitts, T.P.; Vera-Diaz, B.; Winder, P.L.; Yu, Y.; Wright, A.E. Inhibition of il-8 secretion on bxp-3 and mia paca-2 cells and induction of cytotoxicity in pancreatic cancer cells with marine natural products. *Anticancer Drugs* **2017**, *28*, 153–160. [[CrossRef](#)] [[PubMed](#)]
105. Deshpande, A.; Sicinski, P.; Hinds, P.W. Cyclins and cdk in development and cancer: A perspective. *Oncogene* **2005**, *24*, 2909–2915. [[CrossRef](#)] [[PubMed](#)]
106. Dyshlovoy, S.A.; Naeth, I.; Venz, S.; Preukschas, M.; Sievert, H.; Jacobsen, C.; Shubina, L.K.; Gesell Salazar, M.; Scharf, C.; Walther, R.; et al. Proteomic profiling of germ cell cancer cells treated with aaptamine, a marine alkaloid with antiproliferative activity. *J. Proteome Res.* **2012**, *11*, 2316–2330. [[CrossRef](#)] [[PubMed](#)]
107. Li, Q.L.; Zhang, P.P.; Wang, P.Q.; Yu, H.B.; Sun, F.; Hu, W.Z.; Wu, W.H.; Zhang, X.; Chen, F.; Chu, Z.Y.; et al. The cytotoxic and mechanistic effects of aaptamine on hepatocellular carcinoma. *Anticancer Agents Med. Chem.* **2015**, *15*, 291–297. [[CrossRef](#)] [[PubMed](#)]
108. Aoki, S.; Kong, D.; Suna, H.; Sowa, Y.; Sakai, T.; Setiawan, A.; Kobayashi, M. Aaptamine, a spongean alkaloid, activates p21 promoter in a p53-independent manner. *Biochem. Biophys. Res. Commun.* **2006**, *342*, 101–106. [[CrossRef](#)] [[PubMed](#)]
109. Jin, M.; Zhao, W.; Zhang, Y.; Kobayashi, M.; Duan, H.; Kong, D. Antiproliferative effect of aaptamine on human chronic myeloid leukemia k562 cells. *Int. J. Mol. Sci.* **2011**, *12*, 7352–7359. [[CrossRef](#)] [[PubMed](#)]
110. Wright, A.E.; Roth, G.P.; Hoffman, J.K.; Divlianska, D.B.; Pechter, D.; Sennett, S.H.; Guzman, E.A.; Linley, P.; McCarthy, P.J.; Pitts, T.P.; et al. Isolation, synthesis, and biological activity of aphrocallistin, an adenine-substituted bromotyramine metabolite from the hexactinellida sponge *Aphrocallistes beatrix*. *J. Nat. Prod.* **2009**, *72*, 1178–1183. [[CrossRef](#)] [[PubMed](#)]
111. Fukuoka, K.; Yamagishi, T.; Ichihara, T.; Nakaike, S.; Iguchi, K.; Yamada, Y.; Fukumoto, H.; Yoneda, T.; Samata, K.; Ikeya, H.; et al. Mechanism of action of aragusterol a (yta0040), a potent anti-tumor marine steroid targeting the g(1) phase of the cell cycle. *Int. J. Cancer* **2000**, *88*, 810–819. [[CrossRef](#)]
112. Guzman, E.A.; Johnson, J.D.; Carrier, M.K.; Meyer, C.I.; Pitts, T.P.; Gunasekera, S.P.; Wright, A.E. Selective cytotoxic activity of the marine-derived batzelline compounds against pancreatic cancer cell lines. *Anticancer Drugs* **2009**, *20*, 149–155. [[CrossRef](#)] [[PubMed](#)]
113. Edelson, J.R.; Brautigam, D.L. The discodermia calyx toxin calyculin a enhances cyclin d1 phosphorylation and degradation, and arrests cell cycle progression in human breast cancer cells. *Toxins* **2011**, *3*, 105–119. [[CrossRef](#)] [[PubMed](#)]
114. Aoki, S.; Kong, D.; Matsui, K.; Kobayashi, M. Erythroid differentiation in k562 chronic myelogenous cells induced by crambescidin 800, a pentacyclic guanidine alkaloid. *Anticancer Res.* **2004**, *24*, 2325–2330. [[PubMed](#)]
115. Isbrucker, R.A.; Cummins, J.; Pomponi, S.A.; Longley, R.E.; Wright, A.E. Tubulin polymerizing activity of dictyostatin-1, a polyketide of marine sponge origin. *Biochem. Pharmacol.* **2003**, *66*, 75–82. [[CrossRef](#)]
116. Park, C.; Kim, G.Y.; Kim, G.D.; Lee, W.H.; Cheong, J.H.; Kim, N.D.; Bae, S.J.; Jung, J.H.; Choi, Y.H. Suppression of u937 human monocytic leukemia cell growth by dideoxypetrosynol a, a polyacetylene from the sponge *Petrosia* sp., via induction of cdk inhibitor p16 and down-regulation of prb phosphorylation. *Oncol. Rep.* **2006**, *16*, 171–176. [[CrossRef](#)] [[PubMed](#)]
117. Ter Haar, E.; Kowalski, R.J.; Hamel, E.; Lin, C.M.; Longley, R.E.; Gunasekera, S.P.; Rosenkranz, H.S.; Day, B.W. Discodermolide, a cytotoxic marine agent that stabilizes microtubules more potently than taxol. *Biochemistry* **1996**, *35*, 243–250. [[CrossRef](#)] [[PubMed](#)]

118. Honore, S.; Kamath, K.; Braguer, D.; Wilson, L.; Briand, C.; Jordan, M.A. Suppression of microtubule dynamics by discodermolide by a novel mechanism is associated with mitotic arrest and inhibition of tumor cell proliferation. *Mol. Cancer Ther.* **2003**, *2*, 1303–1311. [[PubMed](#)]
119. Rangel, M.; Prado, M.P.; Konno, K.; Naoki, H.; Freitas, J.C.; Machado-Santelli, G.M. Cytoskeleton alterations induced by geodia corticostylifera depsipeptides in breast cancer cells. *Peptides* **2006**, *27*, 2047–2057. [[CrossRef](#)] [[PubMed](#)]
120. Bae, S.Y.; Kim, G.D.; Jeon, J.E.; Shin, J.; Lee, S.K. Anti-proliferative effect of (19Z)-halichondramide, a novel marine macrolide isolated from the sponge *Chondrosia corticata*, is associated with G2/M cell cycle arrest and suppression of mtor signaling in human lung cancer cells. *Toxicol In Vitro* **2013**, *27*, 694–699. [[CrossRef](#)] [[PubMed](#)]
121. Anderson, H.J.; Coleman, J.E.; Andersen, R.J.; Roberge, M. Cytotoxic peptides hemiasterlin, hemiasterlin a and hemiasterlin b induce mitotic arrest and abnormal spindle formation. *Cancer Chemother. Pharmacol.* **1997**, *39*, 223–226. [[CrossRef](#)] [[PubMed](#)]
122. Paterson, I.; Dalby, S.M.; Roberts, J.C.; Naylor, G.J.; Guzman, E.A.; Isbrucker, R.; Pitts, T.P.; Linley, P.; Divlianska, D.; Reed, J.K.; et al. Leiodermatolide, a potent antimitotic macrolide from the marine sponge *leiodermatium* sp. *Angew. Chem. Int. Ed. Engl.* **2011**, *50*, 3219–3223. [[CrossRef](#)] [[PubMed](#)]
123. Maillhol, D.; Willwacher, J.; Kausch-Busies, N.; Rubitski, E.E.; Sobol, Z.; Schuler, M.; Lam, M.H.; Musto, S.; Loganzo, F.; Maderna, A.; et al. Synthesis, molecular editing, and biological assessment of the potent cytotoxin leiodermatolide. *J. Am. Chem. Soc.* **2014**, *136*, 15719–15729. [[CrossRef](#)] [[PubMed](#)]
124. Sangrajang, S.; Zidane, M.; Berda, P.; More, M.T.; Calvo, F.; Fellous, A. Different microtubule network alterations induced by pachymatemin, a new marine glycoprotein, on two prostatic cell lines. *Cancer Chemother. Pharmacol.* **2000**, *45*, 120–126. [[CrossRef](#)] [[PubMed](#)]
125. Zidane, M.; Pondaven, P.; Roussakis, C.; More, M.T. Effects in vitro of pachymatemin, a glycoprotein from the marine sponge *Pachymatisma johnstonii*, on a non-small-cell bronchopulmonary carcinoma line (nslc-n6). *Anticancer Res.* **1996**, *16*, 2805–2812. [[PubMed](#)]
126. Zidane, M.; More, M.T.; Pondaven, P.; Jaquot, C.; Riou, D.; Roussakis, C. In vivo effect of pachymatemin, a new marine glycoprotein, on a human non-small-cell lung carcinoma. *In Vivo* **1997**, *11*, 185–188. [[PubMed](#)]
127. Hood, K.A.; West, L.M.; Rouwe, B.; Northcote, P.T.; Berridge, M.V.; Wakefield, S.J.; Miller, J.H. Peloruside a, a novel antimitotic agent with paclitaxel-like microtubule-stabilizing activity. *Cancer Res.* **2002**, *62*, 3356–3360. [[PubMed](#)]
128. Chan, A.; Andraea, P.M.; Northcote, P.T.; Miller, J.H. Peloruside a inhibits microtubule dynamics in a breast cancer cell line MCF7. *Investig. New Drugs* **2011**, *29*, 615–626. [[CrossRef](#)] [[PubMed](#)]
129. Aoki, S.; Higuchi, K.; Isozumi, N.; Matsui, K.; Miyamoto, Y.; Itoh, N.; Tanaka, K.; Kobayashi, M. Differentiation in chronic myelogenous leukemia cell k562 by spongean sesterterpene. *Biochem. Biophys. Res. Commun.* **2001**, *282*, 426–431. [[CrossRef](#)] [[PubMed](#)]
130. Jiang, Y.; Ahn, E.Y.; Ryu, S.H.; Kim, D.K.; Park, J.S.; Kang, S.W.; You, S.; Lee, B.J.; Jung, J.H. Mechanism of cell cycle arrest by (8e, 13z, 20z)-strobilinin/(7e, 13z, 20z)-felixinin from a marine sponge *Psammocinia* sp. *Oncol. Rep.* **2005**, *14*, 957–962. [[CrossRef](#)] [[PubMed](#)]
131. Pera, B.; Barasoain, I.; Pantazopoulou, A.; Canales, A.; Matesanz, R.; Rodriguez-Salarichs, J.; Garcia-Fernandez, L.F.; Moneo, V.; Jimenez-Barbero, J.; Galmarini, C.M.; et al. New interfacial microtubule inhibitors of marine origin, pm050489/pm060184, with potent antitumor activity and a distinct mechanism. *ACS Chem. Biol.* **2013**, *8*, 2084–2094. [[CrossRef](#)] [[PubMed](#)]
132. Martinez-Diez, M.; Guillen-Navarro, M.J.; Pera, B.; Bouchet, B.P.; Martinez-Leal, J.F.; Barasoain, I.; Cuevas, C.; Andreu, J.M.; Garcia-Fernandez, L.F.; Diaz, J.F.; et al. Pm060184, a new tubulin binding agent with potent antitumor activity including p-glycoprotein over-expressing tumors. *Biochem. Pharmacol.* **2014**, *88*, 291–302. [[CrossRef](#)] [[PubMed](#)]
133. Angawi, R.F.; Saqer, E.; Abdel-Lateff, A.; Badria, F.A.; Ayyad, S.E. Cytotoxic neviotane triterpene-type from the red sea sponge *Siphonochalina siphonella*. *Pharmacogn. Mag.* **2014**, *10*, S334–341. [[PubMed](#)]
134. Aoki, S.; Kong, D.; Matsui, K.; Kobayashi, M. Smenospongine, a spongean sesquiterpene aminoquinone, induces erythroid differentiation in k562 cells. *Anticancer Drugs* **2004**, *15*, 363–369. [[CrossRef](#)] [[PubMed](#)]
135. Rubin, E.; Tamrakar, S.; Ludlow, J.W. Protein phosphatase type 1, the product of the retinoblastoma susceptibility gene, and cell cycle control. *Front. Biosci.* **1998**, *3*, D1209–1219. [[PubMed](#)]

136. Robinson, J.; Sieff, C.; Delia, D.; Edwards, P.A.; Greaves, M. Expression of cell-surface HLA-DR, HLA-ABC and glycophorin during erythroid differentiation. *Nature* **1981**, *289*, 68–71. [[CrossRef](#)] [[PubMed](#)]
137. Nichols, G.L.; Raines, M.; Vera, J.; Lacomis, L.; Tempst, P.A.; Golde, D. Identification of crkl as the constitutively phosphorylated 39-kd tyrosine phosphoprotein in chronic myelogenous leukemia cells. *Blood* **1994**, *84*, 2912–2918. [[PubMed](#)]
138. Fanale, D.; Bronte, G.; Passiglia, F.; Calo, V.; Castiglia, M.; Di Piazza, F.; Barraco, N.; Cangemi, A.; Catarella, M.T.; Insalaco, L.; et al. Stabilizing versus destabilizing the microtubules: A double-edge sword for an effective cancer treatment option? *Anal. Cell. Pathol.* **2015**, *2015*, 690916. [[CrossRef](#)] [[PubMed](#)]
139. Field, J.J.; Waight, A.B.; Senter, P.D. A previously undescribed tubulin binder. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 13684–13685. [[CrossRef](#)] [[PubMed](#)]
140. Miller, J.H.; Singh, A.J.; Northcote, P.T. Microtubule-stabilizing drugs from marine sponges: Focus on peloruside a and zampanolide. *Mar. Drugs* **2010**, *8*, 1059–1079. [[CrossRef](#)] [[PubMed](#)]
141. Gnanambal, K.M.; Lakshminpathy, S.V. Dictyoceratidan poisons: Defined mark on microtubule-tubulin dynamics. *Life Sci.* **2016**, *148*, 229–240. [[CrossRef](#)] [[PubMed](#)]
142. Kowalski, R.J.; Giannakakou, P.; Gunasekera, S.P.; Longley, R.E.; Day, B.W.; Hamel, E. The microtubule-stabilizing agent discodermolide competitively inhibits the binding of paclitaxel (taxol) to tubulin polymers, enhances tubulin nucleation reactions more potently than paclitaxel, and inhibits the growth of paclitaxel-resistant cells. *Mol. Pharmacol.* **1997**, *52*, 613–622. [[PubMed](#)]
143. Gaitanos, T.N.; Buey, R.M.; Diaz, J.F.; Northcote, P.T.; Teesdale-Spittle, P.; Andreu, J.M.; Miller, J.H. Peloruside a does not bind to the taxoid site on beta-tubulin and retains its activity in multidrug-resistant cell lines. *Cancer Res.* **2004**, *64*, 5063–5067. [[CrossRef](#)] [[PubMed](#)]
144. Northcote, P.T.; Miller, J.H.; Hood, K.A.; West, L.M. Bioactive Compound. U.S. Patent 6,790,862 B, 14 September 2004.
145. Rowe, M.R. Resistance to Microtubule-Stabilising Agents Following Point Mutation of Human β i-Tubulin. Master's Thesis, Victoria University of Wellington, Wellington, New Zealand, June 2015.
146. Meyer, C.J.; Krauth, M.; Wick, M.J.; Shay, J.W.; Gellert, G.; De Brabander, J.K.; Northcote, P.T.; Miller, J.H. Peloruside a inhibits growth of human lung and breast tumor xenografts in an athymic nu/nu mouse model. *Mol. Cancer Ther.* **2015**, *14*, 1816–1823. [[CrossRef](#)] [[PubMed](#)]
147. Kanakkanthara, A.; Northcote, P.T.; Miller, J.H. Peloruside a: A lead non-taxoid-site microtubule-stabilizing agent with potential activity against cancer, neurodegeneration, and autoimmune disease. *Nat. Prod. Rep.* **2016**, *33*, 549–561. [[CrossRef](#)] [[PubMed](#)]
148. Kanakkanthara, A.; Eras, J.; Northcote, P.T.; Cabral, F.; Miller, J.H. Resistance to peloruside a and laulimalide: Functional significance of acquired betai-tubulin mutations at sites important for drug-tubulin binding. *Curr. Cancer Drug Targets* **2014**, *14*, 79–90. [[CrossRef](#)] [[PubMed](#)]
149. Kanakkanthara, A.; Rawson, P.; Northcote, P.T.; Miller, J.H. Acquired resistance to peloruside a and laulimalide is associated with downregulation of vimentin in human ovarian carcinoma cells. *Pharm. Res.* **2012**, *29*, 3022–3032. [[CrossRef](#)] [[PubMed](#)]
150. Prota, A.E.; Bargsten, K.; Diaz, J.F.; Marsh, M.; Cuevas, C.; Liniger, M.; Neuhaus, C.; Andreu, J.M.; Altmann, K.H.; Steinmetz, M.O. A new tubulin-binding site and pharmacophore for microtubule-destabilizing anticancer drugs. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 13817–13821. [[CrossRef](#)] [[PubMed](#)]
151. Gottesman, M.M.; Fojo, T.; Bates, S.E. Multidrug resistance in cancer: Role of atp-dependent transporters. *Nat. Rev. Cancer* **2002**, *2*, 48–58. [[CrossRef](#)] [[PubMed](#)]
152. Holohan, C.; Van Schaeybroeck, S.; Longley, D.B.; Johnston, P.G. Cancer drug resistance: An evolving paradigm. *Nat. Rev. Cancer* **2013**, *13*, 714–726. [[CrossRef](#)] [[PubMed](#)]
153. Abolhoda, A.; Wilson, A.E.; Ross, H.; Danenberg, P.V.; Burt, M.; Scotto, K.W. Rapid activation of mdr1 gene expression in human metastatic sarcoma after in vivo exposure to doxorubicin. *Clin. Cancer Res.* **1999**, *5*, 3352–3356. [[PubMed](#)]
154. Gameiro, M.; Silva, R.; Rocha-Pereira, C.; Carmo, H.; Carvalho, F.; Bastos, M.L.; Remiao, F. Cellular models and in vitro assays for the screening of modulators of p-gp, mrp1 and bcrp. *Molecules* **2017**, *22*, 600. [[CrossRef](#)] [[PubMed](#)]

155. Li, W.; Zhang, H.; Assaraf, Y.G.; Zhao, K.; Xu, X.; Xie, J.; Yang, D.H.; Chen, Z.S. Overcoming abc transporter-mediated multidrug resistance: Molecular mechanisms and novel therapeutic drug strategies. *Drug Resist. Updates* **2016**, *27*, 14–29. [[CrossRef](#)] [[PubMed](#)]
156. Yang, Z.X.; Sun, Y.H.; He, J.G.; Cao, H.; Jiang, G.Q. Increased activity of chk enhances the radioresistance of MCF-7 breast cancer stem cells. *Oncol. Lett.* **2015**, *10*, 3443–3449. [[CrossRef](#)] [[PubMed](#)]
157. Martello, L.A.; McDaid, H.M.; Regl, D.L.; Yang, C.P.; Meng, D.; Pettus, T.R.; Kaufman, M.D.; Arimoto, H.; Danishefsky, S.J.; Smith, A.B.; et al. Taxol and discodermolide represent a synergistic drug combination in human carcinoma cell lines. *Clin. Cancer Res.* **2000**, *6*, 1978–1987. [[PubMed](#)]
158. Honore, S.; Kamath, K.; Braguer, D.; Horwitz, S.B.; Wilson, L.; Briand, C.; Jordan, M.A. Synergistic suppression of microtubule dynamics by discodermolide and paclitaxel in non-small cell lung carcinoma cells. *Cancer Res.* **2004**, *64*, 4957–4964. [[CrossRef](#)] [[PubMed](#)]
159. Huang, G.S.; Lopez-Barcons, L.; Freeze, B.S.; Smith, A.B., 3rd; Goldberg, G.L.; Horwitz, S.B.; McDaid, H.M. Potentiation of taxol efficacy and by discodermolide in ovarian carcinoma xenograft-bearing mice. *Clin. Cancer Res.* **2006**, *12*, 298–304. [[CrossRef](#)] [[PubMed](#)]
160. Hamilton, G. Cytotoxic effects of faspaplysin against small cell lung cancer cell lines. *Mar. Drugs* **2014**, *12*, 1377–1389. [[CrossRef](#)] [[PubMed](#)]
161. Fiorini, L.; Tribalat, M.A.; Sauvard, L.; Cazareth, J.; Lalli, E.; Broutin, I.; Thomas, O.P.; Mus-Veteau, I. Natural paniceins from mediterranean sponge inhibit the multidrug resistance activity of patched and increase chemotherapy efficiency on melanoma cells. *Oncotarget* **2015**, *6*, 22282–22297. [[CrossRef](#)] [[PubMed](#)]
162. Wilmes, A.; Bargh, K.; Kelly, C.; Northcote, P.T.; Miller, J.H. Peloruside a synergizes with other microtubule stabilizing agents in cultured cancer cell lines. *Mol. Pharm.* **2007**, *4*, 269–280. [[CrossRef](#)] [[PubMed](#)]
163. Huang, X.C.; Xiao, X.; Zhang, Y.K.; Talele, T.T.; Salim, A.A.; Chen, Z.S.; Capon, R.J. Lamellarin o, a pyrrole alkaloid from an australian marine sponge, *Ianthella* sp., reverses bcrp mediated drug resistance in cancer cells. *Mar Drugs* **2014**, *12*, 3818–3837. [[CrossRef](#)] [[PubMed](#)]
164. Aoki, S.; Chen, Z.S.; Higasiyama, K.; Setiawan, A.; Akiyama, S.; Kobayashi, M. Reversing effect of agosterol a, a spongean sterol acetate, on multidrug resistance in human carcinoma cells. *Jpn. J. Cancer Res.* **2001**, *92*, 886–895. [[CrossRef](#)] [[PubMed](#)]
165. Chen, Z.S.; Aoki, S.; Komatsu, M.; Ueda, K.; Sumizawa, T.; Furukawa, T.; Okumura, H.; Ren, X.Q.; Belinsky, M.G.; Lee, K.; et al. Reversal of drug resistance mediated by multidrug resistance protein (mrp) 1 by dual effects of agosterol a on mrp1 function. *Int. J. Cancer* **2001**, *93*, 107–113. [[CrossRef](#)] [[PubMed](#)]
166. Abraham, I.; Jain, S.; Wu, C.P.; Khanfar, M.A.; Kuang, Y.; Dai, C.L.; Shi, Z.; Chen, X.; Fu, L.; Ambudkar, S.V.; et al. Marine sponge-derived sipholane triterpenoids reverse p-glycoprotein (abcb1)-mediated multidrug resistance in cancer cells. *Biochem. Pharmacol.* **2010**, *80*, 1497–1506. [[CrossRef](#)] [[PubMed](#)]
167. Shi, Z.; Jain, S.; Kim, I.W.; Peng, X.X.; Abraham, I.; Youssef, D.T.; Fu, L.W.; El Sayed, K.; Ambudkar, S.V.; Chen, Z.S. Sipholenol a, a marine-derived sipholane triterpene, potently reverses p-glycoprotein (abcb1)-mediated multidrug resistance in cancer cells. *Cancer Sci.* **2007**, *98*, 1373–1380. [[CrossRef](#)] [[PubMed](#)]
168. Gupta, S.; Takebe, N.; Lorusso, P. Targeting the hedgehog pathway in cancer. *Ther. Adv. Med. Oncol.* **2010**, *2*, 237–250. [[CrossRef](#)] [[PubMed](#)]
169. Bidet, M.; Tomico, A.; Martin, P.; Guizouarn, H.; Mollat, P.; Mus-Veteau, I. The hedgehog receptor patched functions in multidrug transport and chemotherapy resistance. *Mol. Cancer Res.* **2012**, *10*, 1496–1508. [[CrossRef](#)] [[PubMed](#)]
170. Rifai, Y.; Arai, M.A.; Koyano, T.; Kowithayakorn, T.; Ishibashi, M. Terpenoids and a flavonoid glycoside from acacia pennata leaves as hedgehog/gli-mediated transcriptional inhibitors. *J. Nat. Prod.* **2010**, *73*, 995–997. [[CrossRef](#)] [[PubMed](#)]
171. Segraves, N.L.; Lopez, S.; Johnson, T.A.; Said, S.A.; Fu, X.; Schmitz, F.J.; Pietraszkiewicz, H.; Valeriote, F.A.; Crews, P. Structures and cytotoxicities of faspaplysin and related alkaloids from two marine phyla—*Faspaplysinopsis* sponges and *didemnum* tunicates. *Tetrahedron Lett.* **2003**, *44*, 3471–3475. [[CrossRef](#)]
172. Radwan, M.; Hanora, A.; Khalifa, S.; Abou-El-Ela, S.H. Manzamines: A potential for novel cures. *Cell Cycle* **2012**, *11*, 1765–1772. [[CrossRef](#)] [[PubMed](#)]
173. Guzman, E.A.; Johnson, J.D.; Linley, P.A.; Gunasekera, S.E.; Wright, A.E. A novel activity from an old compound: Manzamine a reduces the metastatic potential of aspc-1 pancreatic cancer cells and sensitizes them to trail-induced apoptosis. *Investig. New Drugs* **2011**, *29*, 777–785. [[CrossRef](#)] [[PubMed](#)]

174. Manohar, S.; Leung, N. Cisplatin nephrotoxicity: A review of the literature. *J. Nephrol.* **2017**. [[CrossRef](#)] [[PubMed](#)]
175. Waissbluth, S.; Peleva, E.; Daniel, S.J. Platinum-induced ototoxicity: A review of prevailing ototoxicity criteria. *Eur. Arch. Otorhinolaryngol.* **2017**, *274*, 1187–1196. [[CrossRef](#)] [[PubMed](#)]
176. Koleini, N.; Kardami, E. Autophagy and mitophagy in the context of doxorubicin-induced cardiotoxicity. *Oncotarget* **2017**, *8*, 46663–46680. [[CrossRef](#)] [[PubMed](#)]
177. Shippee, B.M.; Bates, J.S.; Richards, K.L. The role of screening and monitoring for bleomycin pulmonary toxicity. *J. Oncol. Pharm. Pract.* **2016**, *22*, 308–312. [[CrossRef](#)] [[PubMed](#)]
178. Pham, C.D.; Hartmann, R.; Muller, W.E.; de Voogd, N.; Lai, D.; Proksch, P. Aaptamine derivatives from the Indonesian sponge *Aaptos suberitoides*. *J. Nat. Prod.* **2013**, *76*, 103–106. [[CrossRef](#)] [[PubMed](#)]
179. Ebel, R.; Brenzinger, M.; Kunze, A.; Gross, H.J.; Proksch, P. Wound activation of protoxins in marine sponge *Aplysina aerophoba*. *J. Chem. Ecol.* **1997**, *23*, 1451–1462. [[CrossRef](#)]
180. Funk, F.; Kruger, K.; Henninger, C.; Watjen, W.; Proksch, P.; Thomale, J.; Fritz, G. Spongean alkaloids protect rat kidney cells against cisplatin-induced cytotoxicity. *Anticancer Drugs* **2014**, *25*, 917–929. [[CrossRef](#)] [[PubMed](#)]
181. Khalil, H.S.; Tummala, H.; Chakarov, S.; Zhelev, N.; Lane, D.P. Targeting atm pathway for therapeutic intervention in cancer. *Biodiscovery* **2012**. [[CrossRef](#)]
182. Curman, D.; Cinel, B.; Williams, D.E.; Rundle, N.; Block, W.D.; Goodarzi, A.A.; Hutchins, J.R.; Clarke, P.R.; Zhou, B.B.; Lees-Miller, S.P.; et al. Inhibition of the g2 DNA damage checkpoint and of protein kinases chk1 and chk2 by the marine sponge alkaloid debromohymenialdisine. *J. Biol. Chem.* **2001**, *276*, 17914–17919. [[CrossRef](#)] [[PubMed](#)]
183. Pires, I.M.; Ward, T.H.; Dive, C. Oxaliplatin responses in colorectal cancer cells are modulated by chk2 kinase inhibitors. *Br. J. Pharmacol.* **2010**, *159*, 1326–1338. [[CrossRef](#)] [[PubMed](#)]
184. Belotti, D.; Vergani, V.; Drudis, T.; Borsotti, P.; Pitelli, M.R.; Viale, G.; Giavazzi, R.; Taraboletti, G. The microtubule-affecting drug paclitaxel has antiangiogenic activity. *Clin. Cancer Res.* **1996**, *2*, 1843–1849. [[PubMed](#)]
185. Shaw, S.J. The structure activity relationship of discodermolide analogues. *Mini Rev. Med. Chem.* **2008**, *8*, 276–284. [[CrossRef](#)] [[PubMed](#)]
186. Tangutur, A.D.; Kumar, D.; Krishna, K.V.; Kantevari, S. Microtubule targeting agents as cancer chemotherapeutics: An overview of molecular hybrids as stabilising and destabilising agents. *Curr. Top. Med. Chem.* **2017**. [[CrossRef](#)] [[PubMed](#)]
187. Fimognari, C.; Lenzi, M.; Hrelia, P. Chemoprevention of cancer by isothiocyanates and anthocyanins: Mechanisms of action and structure-activity relationship. *Curr. Med. Chem.* **2008**, *15*, 440–447. [[CrossRef](#)] [[PubMed](#)]
188. Aiub, C.; Giannerini, A.; Ferreira, F.; Mazzei, J.; Stankevics, L.; Lobo-Hajdu, G.; Guimaraes, P.; Hajdu, E.; Felzenszwalb, I. Genotoxic evaluation of extracts from *Aplysina fulva*, a Brazilian marine sponge. *Mutat. Res.* **2006**, *611*, 34–41. [[CrossRef](#)] [[PubMed](#)]
189. De Flora, S.; Bagnasco, M.; Bennicelli, C.; Camoirano, A.; Bojnemirski, A.; Kurelec, B. Biotransformation of genotoxic agents in marine sponges. Mechanisms and modulation. *Mutagenesis* **1995**, *10*, 357–364. [[CrossRef](#)] [[PubMed](#)]
190. Stankevics, L.; Aiub, C.; Maria, L.C.; Lobo-Hajdu, G.; Felzenszwalb, I. Genotoxic and antigenotoxic evaluation of extracts from *Arenosclera brasiliensis*, a Brazilian marine sponge. *Toxicol. In Vitro* **2008**, *22*, 1869–1877. [[CrossRef](#)] [[PubMed](#)]
191. Aqil, F.; Zahin, M.; El Sayed, K.A.; Ahmad, I.; Orabi, K.Y.; Arif, J.M. Antimicrobial, antioxidant, and antimutagenic activities of selected marine natural products and tobacco cembranoids. *Drug Chem. Toxicol.* **2011**, *34*, 167–179. [[CrossRef](#)] [[PubMed](#)]
192. Ferguson, L.R. Antimutagens as cancer chemopreventive agents in the diet. *Mutat. Res.* **1994**, *307*, 395–410. [[CrossRef](#)]
193. Nogueira, M.E.; Passoni, M.H.; Biso, F.I.; Longo Mdo, C.; Cardoso, C.R.; Santos, L.C.; Varanda, E.A. Investigation of genotoxic and antigenotoxic activities of *Melampodium divaricatum* in *Salmonella typhimurium*. *Toxicol. In Vitro* **2006**, *20*, 361–366. [[CrossRef](#)] [[PubMed](#)]
194. Steele, V.E.; Kelloff, G.J. Development of cancer chemopreventive drugs based on mechanistic approaches. *Mutat. Res.* **2005**, *591*, 16–23. [[CrossRef](#)] [[PubMed](#)]

195. Dobias, L.; Cerna, M.; Rossner, P.; Sram, R. Genotoxicity and carcinogenicity of metronidazole. *Mutat. Res.* **1994**, *317*, 177–194. [[CrossRef](#)]
196. Hickman, J.A.; Graeser, R.; de Hoogt, R.; Vidic, S.; Brito, C.; Gutekunst, M.; van der Kuip, H.; Consortium, I.P. Three-dimensional models of cancer for pharmacology and cancer cell biology: Capturing tumor complexity in vitro/ex vivo. *Biotechnol. J.* **2014**, *9*, 1115–1128. [[CrossRef](#)] [[PubMed](#)]
197. Lipinski, C.A. Drug-like properties and the causes of poor solubility and poor permeability. *J. Pharmacol. Toxicol. Methods* **2000**, *44*, 235–249. [[CrossRef](#)]
198. Hirata, Y.; Uemura, D. Halichondrins-antitumor polyether macrolides from a marine sponge. *Pure Appl. Chem.* **1986**, *58*, 701–710. [[CrossRef](#)]
199. Drugs.com. Fda Approves Halaven. Available online: <https://www.drugs.com/newdrugs/fda-approves-halaven-late-stage-breast-cancer-2406.html> (accessed on 20 July 2017).
200. Ro, J.; Cheng, F.T.; Sriuranpong, V.; Villalon, A.; Smruti, B.K.; Tsang, J.; Yap, Y.S.; Asian Working Group for Eribulin Clinical. Patient management with eribulin in metastatic breast cancer: A clinical practice guide. *J. Breast Cancer* **2016**, *19*, 8–17. [[CrossRef](#)] [[PubMed](#)]
201. Partridge, A.H.; Rumble, R.B.; Carey, L.A.; Come, S.E.; Davidson, N.E.; Di Leo, A.; Gralow, J.; Hortobagyi, G.N.; Moy, B.; Yee, D.; et al. Chemotherapy and targeted therapy for women with human epidermal growth factor receptor 2-negative (or unknown) advanced breast cancer: American society of clinical oncology clinical practice guideline. *J. Clin. Oncol.* **2014**, *32*, 3307–3329. [[CrossRef](#)] [[PubMed](#)]
202. Towle, M.J.; Salvato, K.A.; Wels, B.F.; Aalfs, K.K.; Zheng, W.; Seletsky, B.M.; Zhu, X.; Lewis, B.M.; Kishi, Y.; Yu, M.J.; et al. Eribulin induces irreversible mitotic blockade: Implications of cell-based pharmacodynamics for in vivo efficacy under intermittent dosing conditions. *Cancer Res.* **2011**, *71*, 496–505. [[CrossRef](#)] [[PubMed](#)]
203. Towle, M.J.; Salvato, K.A.; Budrow, J.; Wels, B.F.; Kuznetsov, G.; Aalfs, K.K.; Welsh, S.; Zheng, W.; Seletsky, B.M.; Palme, M.H.; et al. In vitro and in vivo anticancer activities of synthetic macrocyclic ketone analogues of halichondrin b. *Cancer Res.* **2001**, *61*, 1013–1021. [[PubMed](#)]
204. Kanthou, C.; Tozer, G.M. Microtubule depolymerizing vascular disrupting agents: Novel therapeutic agents for oncology and other pathologies. *Int. J. Exp. Pathol.* **2009**, *90*, 284–294. [[CrossRef](#)] [[PubMed](#)]
205. Agoulnik, S.I.; Kawano, S.; Taylor, N.; Oestreicher, J.; Matsui, J.; Chow, J.; Oda, Y.; Funahashi, Y. Eribulin mesylate exerts specific gene expression changes in pericytes and shortens pericyte-driven capillary network in vitro. *Vasc. Cell* **2014**, *6*, 3. [[CrossRef](#)] [[PubMed](#)]
206. Funahashi, Y.; Okamoto, K.; Adachi, Y.; Semba, T.; Uesugi, M.; Ozawa, Y.; Tohyama, O.; Uehara, T.; Kimura, T.; Watanabe, H.; et al. Eribulin mesylate reduces tumor microenvironment abnormality by vascular remodeling in preclinical human breast cancer models. *Cancer Sci.* **2014**, *105*, 1334–1342. [[CrossRef](#)] [[PubMed](#)]
207. Dybdal-Hargreaves, N.F.; Risinger, A.L.; Mooberry, S.L. Eribulin mesylate: Mechanism of action of a unique microtubule-targeting agent. *Clin. Cancer Res.* **2015**, *21*, 2445–2452. [[CrossRef](#)] [[PubMed](#)]
208. Steinestel, K.; Eder, S.; Schrader, A.J.; Steinestel, J. Clinical significance of epithelial-mesenchymal transition. *Clin. Transl. Med.* **2014**, *3*, 17. [[CrossRef](#)] [[PubMed](#)]
209. Yoshida, T.; Ozawa, Y.; Kimura, T.; Sato, Y.; Kuznetsov, G.; Xu, S.; Uesugi, M.; Agoulnik, S.; Taylor, N.; Funahashi, Y.; et al. Eribulin mesilate suppresses experimental metastasis of breast cancer cells by reversing phenotype from epithelial-mesenchymal transition (emt) to mesenchymal-epithelial transition (met) states. *Br. J. Cancer* **2014**, *110*, 1497–1505. [[CrossRef](#)] [[PubMed](#)]
210. Kolb, E.A.; Gorlick, R.; Reynolds, C.P.; Kang, M.H.; Carol, H.; Lock, R.; Keir, S.T.; Maris, J.M.; Billups, C.A.; Desjardins, C.; et al. Initial testing (stage 1) of eribulin, a novel tubulin binding agent, by the pediatric preclinical testing program. *Pediatr. Blood Cancer* **2013**, *60*, 1325–1332. [[CrossRef](#)] [[PubMed](#)]
211. Towle, M.J.; Nomoto, K.; Asano, M.; Kishi, Y.; Yu, M.J.; Littlefield, B.A. Broad spectrum preclinical antitumor activity of eribulin (halaven(r)): Optimal effectiveness under intermittent dosing conditions. *Anticancer Res.* **2012**, *32*, 1611–1619. [[PubMed](#)]
212. Cortes, J.; O’Shaughnessy, J.; Loesch, D.; Blum, J.L.; Vahdat, L.T.; Petrakova, K.; Chollet, P.; Manikas, A.; Dieras, V.; Delozier, T.; et al. Eribulin monotherapy versus treatment of physician’s choice in patients with metastatic breast cancer (embrace): A phase 3 open-label randomised study. *Lancet* **2011**, *377*, 914–923. [[CrossRef](#)]

213. Kaufman, P.A.; Awada, A.; Twelves, C.; Yelle, L.; Perez, E.A.; Velikova, G.; Olivo, M.S.; He, Y.; Dutcus, C.E.; Cortes, J. Phase iii open-label randomized study of eribulin mesylate versus capecitabine in patients with locally advanced or metastatic breast cancer previously treated with an anthracycline and a taxane. *J. Clin. Oncol.* **2015**, *33*, 594–601. [CrossRef] [PubMed]
214. FDA. Fda Approves First Drug to Show Survival Benefit in Liposarcoma. Available online: <https://www.fda.gov/newsevents/newsroom/pressannouncements/ucm483714.htm> (accessed on 17 August 2017).
215. Schoffski, P.; Chawla, S.; Maki, R.G.; Italiano, A.; Gelderblom, H.; Choy, E.; Grignani, G.; Camargo, V.; Bauer, S.; Rha, S.Y.; et al. Eribulin versus dacarbazine in previously treated patients with advanced liposarcoma or leiomyosarcoma: A randomised, open-label, multicentre, phase 3 trial. *Lancet* **2016**, *387*, 1629–1637. [CrossRef]
216. ClinicalTrials.gov. Available online: <https://clinicaltrials.gov/ct2/results?cond=breast+cancer&term=eribulin+mesylate&cntry1=&state1=&Search=Search> (accessed on 20 July 2017).
217. ClinicalTrials.gov. Available online: https://clinicaltrials.gov/ct2/results?term=eribulin+mesylate&type=&rslt=&age_v=&gndr=&cond=Cancer&intr=&titles=combination&outc=&spons=&lead=&id=&cntry1=&state1=&cntry2=&state2=&cntry3=&state3=&locn=&rcv_s=&rcv_e=&lup_s=&lup_e= (accessed on 23 July 2017).
218. Talpir, R.; Benayahu, Y.; Kashman, Y.; Pannell, L.; Schleyer, M. Hemiasterlin and geodiamolide ta; two new cytotoxic peptides from the marine sponge *Hemiasterella* minor (kirkpatrick). *Tetrahedron Lett.* **1994**, *35*, 4453–4456. [CrossRef]
219. Kuznetsov, G.; TenDyke, K.; Towle, M.J.; Cheng, H.; Liu, J.; Marsh, J.P.; Schiller, S.E.; Spyvee, M.R.; Yang, H.; Seletsky, B.M.; et al. Tubulin-based antimetabolic mechanism of e7974, a novel analogue of the marine sponge natural product hemiasterlin. *Mol. Cancer Ther.* **2009**, *8*, 2852–2860. [CrossRef] [PubMed]
220. Kuznetsov, G.; Towle, M.J.; Liu, J.; Cheng, H.; Tendyke, K.; Kowalczyk, J.J.; Campagna, S.; Littlefield, B.A. Tubulin-based antimetabolic mechanism of novel hemiasterlin analog E7974. In *Proceedings of the 96th Annual Meeting 2005; AACR: Anaheim, CA, USA, 2005; Volume 65*, p. 810.
221. Rocha-Lima, C.M.; Bayraktar, S.; Macintyre, J.; Raez, L.; Flores, A.M.; Ferrell, A.; Rubin, E.H.; Poplin, E.A.; Tan, A.R.; Lucarelli, A.; et al. A phase 1 trial of e7974 administered on day 1 of a 21-day cycle in patients with advanced solid tumors. *Cancer* **2012**, *118*, 4262–4270. [CrossRef] [PubMed]
222. Hidalgo, M.; Boni, V.; Tolcher, A.; Smith, L.; Cubillo, A.; Rasco, D.; Calvo, E.; Amaya, A.; Ordoñez, E.; Patnaik, A.; et al. 361 phase i, open-label, dose-escalating clinical and pharmacokinetic study of the novel antimicrotubulin agent pm060184 administered over 10 minutes on days 1–3 and 15–17 every 28 days to patients with advanced malignant solid tumors. *Eur. J. Cancer* **2015**, *51*, S74. [CrossRef]
223. FDA. Guidance for Industry. Available online: <https://www.fda.gov/downloads/Drugs/Guidances/ucm085389.pdf> (accessed on 25 July 2017).
224. Cavalcanti, B.C.; Sombra, C.M.; de Oliveira, J.H.; Berlinck, R.G.; de Moraes, M.O.; Pessoa, C. Cytotoxicity and genotoxicity of ingenamine g isolated from the brazilian marine sponge *Pachychalina alcaloidifera*. *Comp. Biochem. Physiol. C Toxicol. Pharmacol.* **2008**, *147*, 409–415. [CrossRef] [PubMed]



© 2017 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).