IMPACT OF ENGINEERED NANOPARTICLES ON VIRULENCE OF XANTHOMONAS ORYZAE PV ORYZAE AND ON RICE SENSITIVITY AT ITS INFECTION

Giuliano Degrassi ⁽¹⁾, Livia Vittori Antisari ⁽²⁾, Vittorio Venturi ⁽¹⁾, Serena Carbone ^{(2)*}, Antonietta M. Gatti ⁽³⁾, Chiara Gambardella ⁽⁴⁾, Carla Falugi ⁽⁴⁾, Gilmo Vianello ⁽²⁾

(1) International Centre for Genetic Engineering and Biotechnology, Trieste, Italy
(2) Department of Agricultural Sciences, University of Bologna, Italy
(3) Nanodiagnostics srl. Modena, Italy

Abstract

The present work of nanotoxicity wants to propose a new plant model starting from the rice plant. The model takes into consideration the impact of engineered nanoparticles (Ag, Co, Ni, CeO₂, Fe₃O₄, TiO₂) on rice plants that were weakened by infections of *Xanthomonas oryzae* pv *oryzae* bacteria. The results indicate that some NPs increase the rice sensitivity to the pathogen while others decrease the virulence of the pathogen towards rice. No-enrichment in component metal concentration is detected in above organs of rice, with exception of Ni-NPs treatment. An imbalance of major elements in infected rice crops treated with NPs was investigated.

Keywords: engineered nanoparticles, Ag, Co, Ni, CeO₂, rice, Xanthomonas oryzae

Introduction

Nanoparticles (NPs) are entities that can be either of natural origin, or generated by engineering processes or unintentionally produced by combustion and NPs will find their way into aquatic, terrestrial and atmospheric environment (Dinish et al., 2010). Nanoparticle research is currently an area of intense scientific interest due to a wide variety of potential applications in biomedical, optical and electronic fields, but could also present possible negative effects from the medical and environmental point of view. Most of these effects are due to the high surface to volume ratio, which can make the particles very reactive or catalytic due to their size, namely from 1 to100 nm. They can pass through cell membranes in organisms, and their interactions with biological systems are relatively unknown. The importance of understanding the NPs impact on the environment is growing due to the increased release into the environment and also because they are present in several products already on the market. The plants are essential base of environment and NPs will inevitably interact with plants and these interactions such as uptake and accumulation in plant biomass will greatly affect their fate and

⁽⁴⁾ Department of Earth, Environmental and Life Sciences, University of Genova, Italy

^{*} Corresponding author: e.mail serena.carbone2@unibo.it

transport in the environment. NPs could exert physical and/or chemical toxicity on plants (Ma et al, 2011). The interaction between NPs and plants were object of studies (Lin et al., 2007), focusing the potential toxicity and both positive and negative or negligible effects have been described (Menard et al., 2006). Evidence that NPs penetrate into plant cell were also reported with or without showing adverse effects (Khodakovskaya et al., 2009; Birbaum et al., 2010; Cifuentes et al., 2010).

In this work, we propose a complex biological systems represented by rice crop and Xanthomonas oryzae pv oryzae, its pathogen to study the effects of NPs on crop, pathogen and the interaction between crop-pathogen. The rice represents the staple food for more than two billion people every day. There are many bacteria associated to rice, some of these are pathogenic and cause severe damages to the crop, such as Xanthomonas oryzae pv oryzae (Shen and Ronald, 2002), Burkholderia glumae and B. plantarii (Cottyn et al., 1996; Coenye et al., 1999) and Pseudomonas fuscovaginae (Mattiuzzo et al., 2011). In particular, Xanthomonas oryzae py oryzae (Xoo) cause bacterial blight one of the most destructive rice disease, resulting in 10% to 80% yield losses, endangering worldwide food security (Zhu et al., 2000). Protection of crop plants from bacterial disease can substantially improve agricultural production. Synthetic or fumigant chemicals including inorganic, organic and metallic compounds are agents controlling the crop diseases with advise impact on the environment.. The alternative is the resistance of rice by means of new transgenic technique. Recently, engineered NPs metal-based were tested as antimicrobial agents; they entry in cells of many bacteria (e.g. Escherichia coli, Pseudomonas). The graphene oxide exhibits a high efficiency to inactive the bacteria pathogen of rise (Chen et al., 2013).

With these considerations, the aim of this study was to evaluate 1) the efficiency of *Xanthomonas oryzae* pv *oryzae* (*Xoo*) exposed to metal (Ag, Co, Ni) and metal oxides (CeO₂, Fe₃O₄, TiO₂) nanoparticles infecting rice crop; 2) the effects of NPs on rice crop infected by Xoo after their addition to soil, mimicking the NPs contamination due to polluted biosolid or wastewater.

Images using electron microscopy (ESEM-EDX) were carried out to evaluate the pathogens culture, while the assessment of the uptake and translocation of NPs in crops was determined using inductive coupled plasma spectrometry (ICP-OES) after acidic mineralization of different organs (root, stem, panicles/seeds).

Materials and methods

Bacterial strain and growth conditions

The bacterial strain used in this study is *Xanthomonas oryzae* pv. *oryzae* XKK.12 (*Xoo*), isolated in Kerala State (India) and previously reported as being highly virulent to rice. (Ferluga *et al.*, 2007). *Xoo* was grown at 28°C on peptone-yeast extract liquid medium (PY; 0,8% peptone, 0.2% yeast extract, 0,2% K₂HPO₄, 0,05% KH₂PO₄, 0,025% MgSO₄ 7H₂O, 0.5% glucose w/v) or on peptone-sucrose agar plates (peptone 1%, sucrose 1%, agar 1,5% w/v). *Xoo* was grown in PY medium with and without NPs at concentration of 100 μg ml⁻¹.

Rice growth

The plant used for the study of host-bacteria interaction is *Oryza sativa* Italian cultivar Baldo. Rice was grown either in a green house (without control of humidity and light and temperature kept below 32°C) for CeO₂, Co, TiO₂, Ni -NPs treatments and in a phytotron (with temperature of 27°C, relative humidity of 50% and a light-dark photoperiod of 16:8) for Ag-NP treatment. Rice was fertilized with 10 g/m² of iron sulfate and 5 g/m² of ammonium sulphate every two weeks, and daily watered. For this reason we present the data separated for the two different growth conditions.

Nanoparticles

The plants were exposed to: magnetite (Fe₃O₄; 15-20 nm), passivated cobalt (Co, 28 nm), titanium oxide (TiO₂; 20-160 nm), nickel (Ni, 62 nm), silicon oxide (SiO₂, 4-40 nm), cerium oxide (CeO₂, 50-105 nm), tin oxide (SnO₂, 61 nm). All the chemicals and reagents were purchased from Sigma Aldrich (Italy), unless otherwise stated. SnO₂, CeO₂ and Fe₃O₄ NPs were provided by Nanostructured & Amorphous Materials, Inc (Houston, USA) with at least 98% purity; SiO₂ was provided by Tal Materials Inc (Michigan, USA) with at least 98% purity. By manufacturer's information the specific surface area was 14.2, 8-15, > 40 m and 400 m²/g and the particle size, calculated from the specific surface area and by Transmission Electron Microscopy was 61, 50-105, 20-30, 4-40 nm for SnO₂, CeO₂ Fe₃O₄ and SiO₂, respectively. Stocks of NPs at concentration of 10 mg ml⁻¹ were used and always sonicated before the appropriate dilution.

Experimental design

To assess the influence of NPs on the virulence of *Xanthomonas oryzae* pv *oryzae* (*Xoo*), rice leaves were infected by means of the clipping method previously reported by Ferluga *et al.*, (2009). Briefly, 45 to 60 days old rice leaves, grown in green house and phytotron, were infected with overnight cultures of *Xoo* following the schema in which the control was the rice infected by *Xoo* (R/*Xoo*) and the treatments were: i) rice+NP infected by *Xoo* (R-NP/*Xoo*), ii) rice infected by *Xoo*-NP treated (R/*Xoo*-NP) and iii) rice+NP infected by *Xoo*+NP (R-NP/*Xoo*-NP). The NPs were added to soil at 500 μg kg⁻¹ of dry weight, while the pathogens were grown in pure culture with NPs at concentration of 100 μg ml⁻¹.

Optical density OD₆₀₀ of the overnight cultures of *Xoo* grown with NPs was measured and the cell number diluted to $1x10^9$ ml⁻¹. These diluted cultures were used to inoculate rice leaves, 3 to 4 leaves per plant, ten plants for each treatment combination (R/*Xoo*-NP and R-NP/*Xoo*-NP) were tested. Scissors were dipped into the cultures and then used to cut the tip of rice leaves. Plants were then incubated for 24 hours in a humidity chamber (27°C and more than 95% of humidity), then moved to the green house. Virulence and symptoms were measured after two weeks by measuring the lesion lengths caused by the pathogens along the central vein (Figure 1). A preliminary experiment compared all the treatments, while a

second experiment was performed limited to Co-, CeO₂-, and TiO₂-NPs, which had confirmed the most virulent.

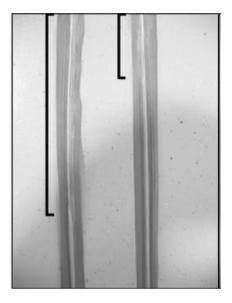


Figure 1
Lesion along the central vein on leaves inoculated with Xanthomonas oryzae pv oryzae by the clipping methods. The measure of the lesions in cm is the parameter of the bacteria virulence

Plants were grown in trays (50x40x9cm) as twenty plants per tray, one tray for each NP tested. Rice plants were grown up to the completion of the flowering stage and seeds production and they were harvested and analyzed for: i) average number of panicles per plant, ii) number of seeds containing starch ("full seeds" vs "empty seeds"); iii) percentage of germination of "full seeds" (seeds were germinated in microtitre plates, one seed in one well with the addition of $100 \, \Box 1$ of sterile water. In addition seeds were tested for the germination ability under standard germination condition (in Petri dishes, onto a water-soaked filter paper, at $28^{\circ}C$ for four days).

At the end of the rice growth cycle ten rice plants were sampled and divided in root, stem and inflorescences. The rice tissues were dried in a forced air oven (T<40°C) and homogenized (in a blender with blades made of pure titanium, carefully checked to not introduce any further metal contamination to samples). Unwashed plant samples were used. The mineralization of rice tissues was carried out according to Vittori Antisari et al., (2011). Briefly, 0.25 g sub-sample of plant tissues was dissolved in 8 mL of concentrated HNO₃ (suprapure, Meck) plus 2 mL of H₂O₂ (Carlo Erba for electronic use). The mineralization was carried out in Teflon bombs in a microwave oven (Milestone, 1200). After cooling, solutions were made up to 20 mL with milli-Q water and then filtered with Whatmann 42. The accuracy of the instrumental method and analytical procedures used was checked using reference material, which was run after every 10 samples to check for drift in the sensitivity. The analytical quality of the results was checked against the following reference materials, which certify values of the studied elements close to the measured ones: CRM 060 (aquatic plants), CRM 062 (Olive leaves)

provided by the European Commission Institute for Reference Materials and Measurements. The solution as analyzed by Inductive Coupled Plasma Optical Emission Spectrometry (ICP-OES) (Spectro Ametek, Arcos).

Statistical analysis of data was performed by one-way ANOVAs analysis of variance with post-hoc Turkey's multiple comparison test.

Results

Greenhouse experiments: Influence of NPs treatments on rice-Xoo system.

After 14 days from infection with *Xanthomonas oryzae* pv *oryzae* (*Xoo*) the length of lesions (in cm) was analysed and the treatments with Co-, CeO₂-, and TiO₂-NPs were more virulent than the control (R/Xoo), as well as the preventive treatment of Xoo with Co-NPs (R/Xoo-Co) showed a lower lesion length than the control. The rice plants contaminated with Ni- and Fe3O4- NPs showed the same behaviour and no significant differences than the control (data no shown).

The exposure of the rice plants at Co-, CeO₂- and TiO₂-NPs, added into soil through irrigation water, showed more virulent disease and the crop seemed to be more sensitive to *Xoo* infection. These results induced to plan new experiments testing again Co-, CeO₂ and TiO₂-NPs. One-way ANOVA test was performed to underline statistically differences among the different treatments and the results for CeO₂-, Co-, and TiO₂-NPs treatments were reported (Table 1).

Table 1. Using One-way Anova analysis of variance with post-hoc Turkey's (p<0.05) multiple comparison test of Figure 2 (Ti, Ce and Co only)

Rice+TiO2-NP	Mean Diff.	q	P < 0.05	95% CI of diff
Rice+Ti/Xoo+Ti vs Rice+Ti/Xoo	-2.1	2.5	ns	-5.0 to 0.75
ice+Ti/Xoo+Ti vs Rice/Xoo	6.6	9.0	***	4.1to 9.1
Rice+Ti/Xoo vs Rice/Xoo	8.8	11.7	***	6.2 to 11.3
Rice +CeO2-NP	Mean Diff.	q	P < 0.05	95% CI of diff
Rice+Ce/Xoo+Si vs Rice+Ce/Xoo	1.0	1.3	ns	-1.5 to 3.5
Rice+Ce/Xoo+Si vs Rice/Xoo	6.3	9.7	***	4.1 to 8.4
Rice+Ce/Xoo vs Rice/Xoo	5.3	8.2	***	3.1 to 7.4
Rice +Co-NP	Mean Diff.	q	P < 0.05	95% CI of diff
Rice+Co/ Xoo+Co vs Rice+Co/Xoo	-0.4596	0.5605	ns	-3.5 to 2.6
Rice+Co/Xoo+Co vs Rice/Xoo	2.9	4.3	*	0.4 to 5.4
Rice+Co/Xoo+Co vs Rice/Xoo+Co	6.1	9.0	***	3.6 to 8.6
Rice+Co/Xoo vs Rice/Xoo	3.4	4.8	**	0.8 to 5.9
Rice+Co/Xoo vs Rice/Xoo+Co	6.6	9.3	***	3.9 to 9.2
Rice/Xoovs Rice/Xoo+Co	3.2	6.1	***	1.3 to 5.2

The results obtained confirmed that *Xoo* grown in laboratory media in presence of Co-NPs was less virulent in the rice plant. In addition the rice plant displays

slightly higher sensitivity to Xoo infection when they are grown after addition of Co-NPs. CeO₂- and TiO₂-NPs into soil inducing significantly (p<0.05) higher pathogen virulence than the control one.

Interaction between Xanthomonas oryzae pv oryzae and NPs using ESEM-EDX.

The interaction between *Xoo* and NPs was detected by means of electron microscopy (ESEM-EDX), in Figure 1 the pathogen was grown in PY medium without NPs, while in figure 2 and 4 two examples of *Xoo* grown in the presence of Fe₃O₄- and Co-NP, respectively, were reported.

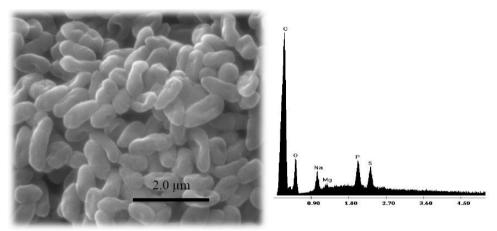


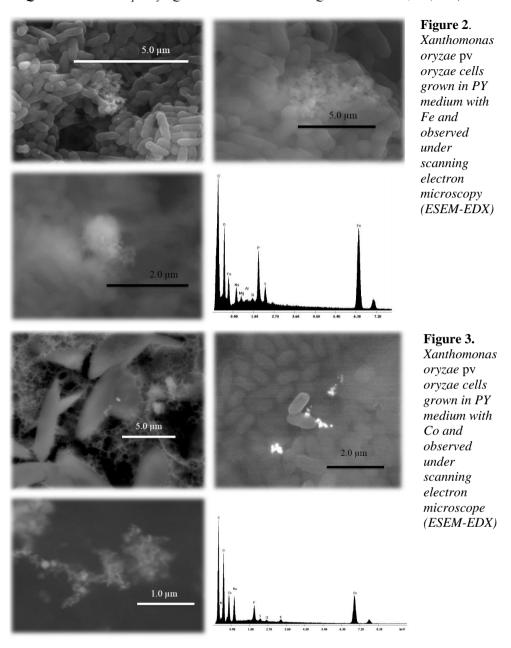
Figure 1. Xanthomonas oryzae pv oryzae cells grown in PY medium without NPs and observed by electron microscopy (ESEM-EDX)

The electron microscope images showed that the magnetite (Figure 2) seemed to form a cluster and aggregates of nanoparticles on the cells of Xoo. The Co-NP showed little aggregates of smaller sizes, isolated particles (Figure 3), also the presence of Co-NP seemed to released organic substances by cells (figure 3).

Panicle and seed formation and fertility.

The rice plants overcome the pathogen infection were grown up to the seeds formation; the number of both panicles and seeds were counted (Table 2). No significantly differences were found between the treatments and the control, even if, as expected, the rice no-exposed at Xoo and at NPs showed high parameters, the contrary for the crops infected by the pathogens. The calculation of germination percentage was performed with "full seeds" and no significant differences in germination rate were observed in among the different seed batches/samples (Table 2).

Although the statistic elaboration of data (ANOVA) did not show significant results, nevertheless a strong trend was seen for the effects exerted by the exposure to the different NPs.



Thus, we calculated the relative risk (RR) showing the probability of the effect occurring in the exposed group versus a non-exposed group. (Sistrom and Garvan, 2004)

$$RR = \frac{P \text{ event when exposed}}{P \text{ event when non-exposed}}$$

Generally, on the basis of this statistical approach the probability of negative effect on seed germination arising from NPs exposure increased than the control ones; about 2.5 times greater in the plants exposed to Ni-NP than the control. The incidence of exposure to NPs was relevant mainly for the exposure of rice plants at NPs. In fact the presence of Ni-, Co- and Fe3O4- NPs could decrease the germination of the seed on rice plants contaminated. At the same time the presence of Co-NP (R+Co/Xoo and R+Co/Xoo-Co) in infected plants could ameliorate the germination phenomenon.

Table2. Number panicles/rice plants, full seed and germination percentage are reported.

	Number panicles/plant		Full seed		Germination	
		SD	%	SD	%	SD
R	2.7	0.7	88.1	2.1	89.0	6.5
R/Xoo	1.8	1.1	76.4	0.5	81.2	3.5
R/Xoo-Co	2.6	0.5	86.6	0.6	83.7	4.0
R+Ce	1.8	0.8	79.4	0.3	84.2	7.4
R+Ce/Xoo	1.8	1.0	79.4	0.2	82.0	9.5
R+Ce/Xoo-Ce	1.8	0.7	87.1	0.2	86.3	5.5
R+Co	1.4	0.6	72.1	0.4	87.3	3.4
R+Co/Xoo	1.7	0.7	72.1	0.1	88.3	4.5
R+Co/Xoo-Co	1.2	0.4	85.1	0.8	86.3	2.5
R+Ni	1.4	0.6	84.7	1.0	78.0	8.0
R+Ni/Xoo	1.3	0.7	85.4	0.2	75.7	8.4
R+Ni/Xoo-Ni	1.5	0.5	84.0	0.4	80.3	8.6
R+Fe3O4	1.7	0.7	81.7	0.8	81.2	6.9
R+Fe/Xoo	1.8	0.7	85.8	0.6	80.3	9.0
R+Fe/Xoo-Fe	1.6	0.7	77.6	0.5	82.0	6.1
R+TiO ₂	1.4	0.6	79.4	0.9	83.0	4.2
R+Ti/Xoo	1.4	0.7	78.1	0.6	83.7	4.0
R+Ti/Xoo-Ti	1.4	0.5	79.4	0.7	82.3	5.1

Total concentration of elements arising NPs in rice crops

Component metal concentration (e.g. Ce, Co, Ni, Fe, Ti) arising from NPs in rice crops organs is detected (Table 3). The ANOVA test showed that the concentration of Co, Ce and Ni in roots was statistically higher (p<0.05) than the respective controls (R/Xoo and R/Xoo-Co), while no differences were found for Fe_3O_4 - and TiO_2 -NPs treatments (Table 3).

In both control and NP-treatment, Ce and Co concentration in epigeous organs was always lower than detection limits (DL) The storage of Fe in rice plants treated with magnetite and infected by both Xoo and Xoo-Fe₃O₄ was lower in roots and stem (p<0.01) than those of respective controls, while no significant difference was found in seeds. In TiO₂-NP treatments, the Ti concentration in stems was significant higher (p<0.01) than the controls, while no difference was found for the other organs. The Ni amount was detected in stem in rice plant exposed at Ni-NPs, the Ni concentration was also significant higher in roots (p<0.05 and 0.01, respectively) in both rice treatments with Ni-NP added to soil.

	Ce	Co	Fe	Ni	Ti
			μg g-1		
Panicles					
R-NP/Xoo	DL	DL	33.7	DL	2.1
R/Xoo	DL	DL	31.2	DL	2.6
ANOVA			ns		ns
R-NP/Xoo-NP	DL	DL	21.9	DL	1.7
R/Xoo-Co	DL	DL	25.9	DL	1.6
ANOVA			ns		ns
Stem					
R-NP/Xoo	DL	DL	47.8	DL	2.7
R/Xoo	DL	DL	62.1	DL	0.9
ANOVA			**		**
R-NP/Xoo-NP	DL	DL	42.7	0.9	2.7
R/Xoo-Co	DL	DL	75.5	DL	0.9
ANOVA			**	**	**
Root					
R-NP/Xoo	2.3	5.9	1757	3.4	31.3
R/Xoo	DL	0.5	2307	1.2	29.3
ANOVA	**	**	ns	*	ns
R-NP/Xoo-NP	3.1	5.2	1403	11.2	24.3
R/Xoo-Co	DL	0.6	1787	1.6	20.9
ANOVA	**	**	*	**	ns

Table 3
Total concentration elements arising from nanoparticles in rice organs.
ANOVA one-way is performed using Tukey test (p<0.05)

Distribution of major elements in rice plant organs

The rice plants harvested at the end of the crop cycle were analysed for major elements; in Table 4 the data obtained by rice treated with NPs and inoculated with Xoo.

Generally, the Ca and Na concentration in panicles of rice plants treated with NPs were lower (p<0.05) than those of control. The major elements concentration in stem varied as a function of NPs treatments without a visible trend. We report a very low concentration of K and P in the stem of R-Ni/Xoo treatment, as well as a very high concentration of both of these nutrients in roots. Low P concentration was detected in roots treated with Co- and Fe_3O_4 -NPs/Xoo.

The major nutrients concentration was not different between the rice plants treated with NPs and the pathogens incubated with NPs and the control in panicles (Table 4), with some exception, the values in stems were lower in rice treated than the control. In particular, the treatments with CeO₂ and Ni-NPs showed the lack of K accumulation in rice stems (Table 4). Ni-NP treatment showed high nutrients concentration in roots, for the other treatments no trend were observed.

Discussion

The interaction between NPs and *Xanthomonas oryzae* pv *oryzae* is tested measuring the length of infection in the leaves (Niňo-Liu et al., 2006) and Co-NP

Table 4. Major elements in rice plant organs. The bold number are significantly different by control.

	Ca	K	Mg	Na	P	S
	mg kg-1					
Panicles						
R-Co/Xoo	664	4579	977	299	2322	874
R-CeO ₂ /Xoo	826	3907	1113	246	2243	898
R-Fe ₃ O ₄ /Xoo	594	4495	945	343	1971	937
R-Ni/Xoo	594	4495	945	343	1971	937
R-TiO ₂ /Xoo	589	4309	909	322	1973	913
Control (R/Xoo)	725	4410	1038	482	1896	1142
Stem						
R-Co/Xoo	4829	3626	1827	1823	435	1579
R-CeO ₂ /Xoo	2974	2983	1721	2168	262	1573
R-Fe ₃ O ₄ /Xoo	3401	2744	1995	4206	710	2011
R-Ni/Xoo	3939	169	1724	5444	146	1489
R-TiO ₂ /Xoo	4772	2761	1995	2454	207	2023
Control (R/Xoo)	3661	3338	2808	3270	371	1966
Roots						
R-Co/Xoo	5750	8117	1486	2886	582	1630
R-CeO ₂ /Xoo	4868	7460	1166	4728	1250	2528
R-Fe ₃ O ₄ /Xoo	6454	7074	1430	3716	821	1928
R-Ni/Xoo	10941	10415	3147	5451	3041	2857
R-TiO ₂ /Xoo	8526	7179	1490	3809	1458	2749
Control (R/Xoo)	9402	6741	1785	3052	1068	2861
	Ca	K	Mg	Na	P	S
D	mg kg-1					
Panicles	520	4200	000	250	2217	924
R-Co/Xoo-Co	539 609	4299 4346	900 1043	259 291	2217 2213	824 995
R-CeO ₂ /Xoo-CeO ₂ R-Fe ₃ O ₄ /Xoo-Fe ₃ O ₄	540	3941	1043	249	2100	995 881
R-Ni/X00-Ni	340 447	4411	849	288	1833	976
R-TiO ₂ /Xoo-TiO ₂	618	4835	1000	344	2190	980
Control (R/X00-Co)	715	4320	1025	478	1883	1005
Stem	/13	4320	1023	470	1003	1003
R-Co/Xoo-Co	4495	3458	1771	1444	286	1532
R-CeO ₂ /Xoo-CeO ₂	2717	nd	1506	2749	315	1725
R-Fe ₃ O ₄ /Xoo-Fe ₃ O ₄	3067	5236	2209	2300	297	1355
R-Ni/Xoo-Ni	3763	nd	1644	5786	175	1472
R-TiO ₂ /Xoo- TiO ₂	2506	2290	1468	2833	548	1616
Control (R/Xoo-Co)	4350	4074	2877	3683	297	1804
Roots		,.		2000		100,
R-Co/Xoo-Co	6905	6770	1542	2837	1041	2090
R-CeO ₂ /Xoo-CeO ₂	7754	7758	1454	3780	1808	2935
R-Fe ₃ O ₄ /Xoo-Fe ₃ O ₄	4937	7783	1349	4698	1153	2363
R-Ni/Xoo-Ni	9267	9003	2335	4488	2338	2740
R-TiO ₂ /Xoo- TiO ₂	7569	7585	1536	3510	1624	2610

treatment shows the more efficiency in inhibiting the rice disease. For this reason, in all the experiments, the symptoms of control without NPs exposure (Xoo) is compared with Xoo grown in the presence of Co-NP. The culturing pathogens treated with Co-NP shows an interaction between the single nanoparticles and pathogens cells, while the interaction between the other NPs and the pathogen cells showed cluster and agglomerations of nanoparticles. These later treatments do not alter the Xoo virulence and pathogenicity. The exposure of rice crops at some NPs (Co-, CeO2 and TiO2) magnify the virulence of pathogen attack and the hypothesis can be due to a more vulnerability of crops exposed to NPs. Many discordant results were found in literature: about the interaction between nanoparticles and crops for examples nano-TiO2 showed no evidence of phytotoxicity in terms of germination and root elongation of tomatoes (Song et al., 2013) on the other hands, genotoxicity of TiO2-NP was assessed in vivo in plants and in vitro in human lymphocytes (Gosh et al., 2010).

Low translocation of nanoparticles in rice organs is found in this experiments, the infection with Xoo do not affect the NPs translocation. ICP-OES data shows that the component metal concentration in above rice organs (e.g. leaves, stem) is low for both infection treatments. Different translocation behaviour for NPs is detected for other crops (Wang et al., 2012; Servin et al., 2012; 2013; Vittori Antisari et al., 2015).

Generally, little is known about the influence of NPs on plant mineral composition or on the macro- and microelement accumulation in different plant organs. In our case a decrease of Ca and Na concentration in panicles is not always balanced by an accumulation in the roots. In all cases, with exception for R-Ni/Xoo and R-Ni/Xoo-Ni, it appears that the plant will absorb a smaller amount from the soil. This phenomenon is associated with an increase of P in above organs of rice. This evidence suggests a competitive inhibition of major nutrients due to NPs added into soils and also Xoo infection.

Conclusions

The question of the uptake, bioaccumulation, biotransformation and risk of NPs in food crops is still not well understood.

The metal oxides and metallic engineered NPs were found to have differing effects on *Xanthomonas oryzae* pv *oryzae* and Co-NP inhibits the virulence of this important disease of rice.

The presence of nanoparticle in soil not influence the disease of rice plants, exacerbating the symptoms. An imbalance of major elements in infected rice crops treated with NPs was investigated.

Acknowledgments This study was supported by the INESE project funded by the Italian Institute of Technology (IIT, Genoa, Italy).

References

BIRBAUM K., BROGIOLI R., SCHELLENBERG M., MARTINOIA E., STARK W.J., GÜNTHER D., *ET AL.* (2010) No evidence for cerium dioxide nanoparticle translocation in maize plants. Environmental Science & Technology, 44:8718-8723.

CIFUENTES Z., CUSTARDOY L., DE LA FUENTE J., MARQUINA C., IBARRA M.R., RUBIALES D. *ET AL.* (2010) Absorption and translocation to the aerial part of magnetic carbon-coated nanoparticles through the root of different crop plants. Journal of Nanobiotechnology, 8:26-34.

CHEN J., WANG X., HAN H. (2013) A new function of graphene oxide emerges: Inactivating phytopathogenic bacterium Xanthomonas oryzae pv. Oryzae. J. Nanopart. Res., 15:1658-1672. DOI: 10.1007/s11051-013-1658-6

COENYE T., HOLMES B., KERSTERS K., GOVAN J.R., VANDAMME P. (1999) Burkholderia cocovenenans (van Damme et al. 1960) Gillis et al. 1995 and Burkholderia vandii Urakami et al. 1994 are junior synonyms of Burkholderia gladioli (Severini 1913) Yabuuchi et al. 1993 and Burkholderia plantarii (Azegami et al. 1987) Urakami et al. 1994, respectively. Int J Syst Bacteriol., 1:37-42.

COTTYN B., VAN OUTRYVE M.F., CEREZ M.T., DE CLEENE M., SWINGS J., MEW T.W. (1996) Bacterial diseases of rice. II. Characterization of pathogenic bacteria associated with sheath rot complex and grain discoloration of rice in the Philippines. Plant Dis., 80:438-445.

DINESH R., ANANDARAJ M., SRINIVASAN V., HAMZA S. (2012) Engineered nanoparticles in the soil and their potential implications to microbial activity. Geoderma, 173–174:19–27. DOI:10.1016/j.geoderma.2011. 12.018

FALUGI C., ALUIGI M.G., CHIANTORE M.C., PRIVITERA D., RAMOINO P., GATTI A.M., FABRIZI A., PINSINO A., MATRANGA V. (2012) Toxicity of metal oxide nanoparticles in immune cells of the sea urchin. Mar Environ. Res., 76:114-121.

FERLUGA S, BIGIRIMANA J, HÖFTE M, VENTURI V. (2007) A LuxR homologue of Xanthomonas oryzae pv. oryzae is required for optimal rice virulence. Mol Plant Pathol. 8:529-538. DOI: 10.1111/j.1364-3703.2007.00415.x.

GATTI A.M., BOSCO P., RIVASI F., BIANCA S., ETTORE G., GAETTI L., MONTANARI S., BARTOLONI G., GAZZOLO D. (2011) Heavy metals nanoparticles in fetal kidney and liver tissues. Front Biosci. (Elite Ed) 3:221-226.

GHOSH M., BANDYOPADHYAY M., MUKHERJEE A. (2010) Genotoxicity of titanium dioxide (TiO₂) nanoparticles at two trophic levels: Plant and human lymphocytes. Chemosphere, 81:1253–1262.

IANNITTI T., CAPONE S., GATTI A., CAPITANI F., CETTA F., PALMIERI B. (2010) Intracellular heavy metal nanoparticle storage: progressive accumulation within lymph nodes with transformation from chronic inflammation to malignancy. Int J Nanomedicine, 5:955-960.

LIN D., XING B. (2007) Phytotoxicity of nanoparticles: Inhibition of seed germination and root growth. Environ Pollut. 150:243–250.

KHODAKOVSKAYA M., DERVISHI E., MAHMOOD M., XU Y., LI Z., WATANABE F. *ET AL.* (2009) Carbon nanotubes are able to penetrate plant seed coat and dramatically affect seed germination and plant growth. ACS Nano, 3:3221-3227.

NIŇO-LIU D.O., RONALD P.C., BOGDANOVE A.J. (2006) *Xantomonas oryzae* pathovars: model pathogens of a model crop. Molecular Plant Pathology, 7:303-324.

MA X., GEISER-LEE J., DENG Y., KOLMAKOV A. (2010) Interactions between engineered nanoparticles (ENPs) and plants: phytotoxicity, uptake and accumulation. Science of The Total Environment, 408:3053-3061.

MATTIUZZO M, BERTANI I, FERLUGA S, CABRIO L, BIGIRIMANA J, GUARNACCIA C, PONGOR S, MARAITE H, VENTURI V. (2011) The plant pathogen Pseudomonas fuscovaginae contains two conserved quorum sensing systems involved in virulence and negatively regulated by RsaL and the novel regulator RsaM. Environ. Microbiol., 13:145-62.

MNYUSIWALLA A., DAAR A.S., SINGER P.A. (2003) 'Mind the gap': science and ethics in nanotechnology. Nanotechnology, 14(3):R9. DOI:10.1088/0957-4484/14/3/201.

SERVIN A.D., CASTILLO-MICHEL H., HERNANDEZ-VIEZCAS J.A., DIAZ B.C., PERALTAVIDEA J.R., GARDEA-TORRESDEY J.L. (2012) Synchrotron micro-XRF and micro-XANES confirmation of the uptake and translocation of TiO₂ nanoparticles in cucumber (Cucumis sativus) plants. Environ. Sci. Technol., 46:7637–7643. DOI:10.1021/es300955b.

SERVIN A.D., MORALES M.I., CASTILLO-MICHEL H., HERNANDEZ-VIEZCAS J.A., MUNOZ B., ZHAO L., NUNEZ J.E., PERALTA-VIDEA J.R., GARDEA-TORRESDEY J.L. (2013) Synchrotron verification of TiO2 accumulation in cucumber fruit: a possible pathway of TiO2 nanoparticle transfer from soil into the food chain. Environ. Sci. Technol., 47:11592–11598. DOI:10.1021/es403368j

SHEN Y., RONALD P. (2002) Molecular determinants of disease and resistance in interactions of *Xanthomonas oryzae* pv. *oryzae* and rice. Microbes Infect., 4:1361-1367.

SISTROM C.L., GARVAN C.W. (2004) Proportions, Odds, And Risk. Radiology, 230 (1):12–9.

SONG U., JUN H., WALDMAN B., ROH J., KIM Y., YI J., LEE E.J. (2013) Functional analyses of nanoparticle toxicity: a comparative study of the effects of TiO2 and Ag on tomatoes (*Lycopersicon esculentum*). Ecotoxicol. Environ. Saf., 93:60–67. DOI:10.1016/j. ecoenv.2013.03.033

VITTORI ANTISARI L., CARBONE S., FABRIZI A., GATTI A., VIANELLO G. (2011). Response of soil microbial biomass to CeO₂ nanoparticles. EQA, 7:135-150. DOI: 10.6092/issn.2281-4485/3829.

VITTORI ANTISARI L., CARBONE S., GATTI A., VIANELLO G., NANNIPIERI P. (2014) Uptake and translocation of metals and nutrients in tomato grown in soil polluted with metal oxide (CeO_2 , Fe_3O_4 , SnO_2 , TiO_2) or metallic (Ag, Co, Ni) engineered nanoparticles. Environmental Science and Pollution. Online First 1-13. DOI 10.1007/s11356-014-3509-0

YING J. (2001) Nanostructured Materials. New York: Academic Press. ISBN 0-12-744451-3.

ZHU Y., CHEN H., FAN J., WANG Y., LI Y., *ET AL*. (2000) Genetic diversity and disease control in rice. Nature, 406:718–722

WANG Q., MA X., ZHANG W., PEI H., CHEN Y. (2012) The impact of cerium oxide nanoparticles on tomato (*Solanum lycopersicum* L.) and its implications for food safety. Metallomics, 4:1105–1112. DOI:10.1039/c2mt20149f.

IMPACT DE NANOPARTICULES DE SYNTHÈSE SUR LA VIRULENCE DE XANTHOMONAS ORYZAE PV ORYZAE ET SUR LA SENSIBILITÉ DU RIZ A SA INFECTION

Résumé

Il propose un nouveau procès sur la nanotoxicité contre les plantes de riz. Le modèle expérimental prend en compte l'impact des nanoparticules de synthèse (Ag, Co, Ni, CeO₂, Fe₃O₄, TiO₂) sur des plants de riz qui ont été affaiblis par *Xanthomonas oryzae pv oryzae* infections bactériennes. Les résultats indiquent que certains NP améliorer la sensibilité de l'agent pathogène de riz tandis que d'autres diminuent la virulence de l'agent pathogène contre rice. Aucun enrichissement significatives dans les concentrations de métaux dans les organes supérieurs de riz, à l'exception d'un traitement avec des nanoparticules de Ni. Il a été constaté un déséquilibre des macro éléments dans des plants de riz infectés traités avec des nanoparticules.

Mots-clés: nanoparticules de synthese, Ag, Co, Ni, CeO₂, riz, Xanthomonas oryzae

IMPATTO DELLE NANOPARTICELLE INGEGNERIZZATE SULLA VIRULENZA DI XANTHOMONAS ORYZAE PV ORYZAE E SULLA SENSIBILITÀ DEL RISO ALLA SUA INFEZIONE

Riassunto

Il presente lavoro propone una nuova sperimentazione relativa alla nanotossicità nei confronti di piante di riso. Il modello sperimentale prende in considerazione l'impatto delle nanoparticelle ingegnerizzate (Ag, Co, Ni, CeO₂, Fe₃O₄, TiO₂) su piante di riso che sono state indebolite da infezioni batteriche di *Xanthomonas oryzae pv oryzae*. I risultati indicano che alcune NP aumentano la sensibilità del riso al patogeno mentre altri diminuiscono la virulenza del patogeno nei confronti del riso. Non si rilevano arricchimenti nelle concentrazioni di metalli negli organi superiori del riso, ad eccezione del trattamento con nanoparticelle di Ni. E' stato riscontrato uno squilibrio degli elementi maggiori nelle piante di riso infette trattate con nanoparticelle.

Parole chiave: nanoparticelle ingegnerizzate, Ag, Co, Ni, CeO₂, riso, Xanthomonas oryzae