



Original Investigation | Oncology

Histopathological Response After Neoadjuvant Chemotherapy for High-Risk Soft-Tissue Sarcomas

A Secondary Analysis of a Randomized Clinical Trial

Sandro Pasquali, MD, PhD; Paola Collini, MD; Cleofe Romagosa, MD; Jean-Michel Coindre, MD; Sara Pizzamiglio, PhD; Paolo Verderio, PhD; Valeria Duroni, MSc; Marta Barisella, MD; Emanuela Palmerini, MD; Vittorio Quagliuolo, MD; Javier Martin Broto, MD; Antonio Lopez Pousa, MD; Giovanni Grignani, MD; Antonella Brunello, MD; Jean-Yves Blay, MD; Iwona Lugowska, MD; Valeria Fontana, MSc; Giuseppe Bianchi, MD; Elena Palassini, MD; Salvatore Lorenzo Renne, MD; Paolo Giovanni Casali, MD; Rosalba Miceli, PhD; Marta Sbaraglia, MD; Marco Gambarotti, MD; Silvia Bagué, MD; Angelo Paolo Dei Tos, MD; Silvia Stacchiotti, MD; Alessandro Gronchi, MD

Abstract

IMPORTANCE Treatment of high-risk soft-tissue sarcoma (STS) of extremity or trunk wall involves neoadjuvant chemotherapy (NACT) followed by surgery. Histopathological response could estimate patient outcomes.

OBJECTIVE To characterize morphological changes in surgical specimens of patients treated with NACT with or without radiotherapy (RT) to identify histopathological features that stratify risk of recurrence and ultimately estimate the benefit from neoadjuvant treatments.

DESIGN, SETTING, AND PARTICIPANTS This was a preplanned prospective secondary analysis of the ISG-ST5 1001 clinical trial, a study with both a randomized clinical trial (conducted between 2011 and 2016) and a nonrandomized patient cohort (included between 2016 and 2020) at 32 centers across Italy, Spain, France, and Poland. Participants were patients with STS randomly assigned to receive either anthracycline plus ifosfamide or histotype-tailored (also termed histology tailored) NACT. Data analyses were performed from January to June 2023.

INTERVENTION Participants received 3 cycles of anthracycline plus ifosfamide or histotype-tailored NACT with or without RT followed by surgery.

MAIN OUTCOMES AND MEASURES The primary outcome was disease-free survival (DFS). Histopathological features considered included the proportion of stainable tumor cells, tumor necrosis, hemorrhage, fibrohistiocytic reaction with hemosiderin, sclerosis or fibrosis, and sclerohyalinosis. The proportion of stainable tumor cells was classified according to the European Organization for Research and Treatment of Cancer–Soft Tissue and Bone Sarcoma Group categories or as absent or present. The continuous variable of sclerohyalinosis, expressed as a percentage, was categorized based on the second tertile of its distribution (20%). Tumor necrosis, hemorrhage, fibrohistiocytic reaction with hemosiderin, sclerosis or fibrosis, which were also expressed as a percentage, were classified as absent or present.

RESULTS A total of 388 patients (201 in randomized cohort, 187 in nonrandomized cohort; median [IQR] age, 50 [41-60] years; 245 males [63.1%]) were evaluable for histopathological response. In the randomized cohort, after a median (IQR) follow-up of 86 (70-99) months, 115 of 201 patients (57.2%) developed a disease recurrence. The proportion of stainable tumor cells (>1%) was not associated with DFS (hazard ratio [HR], 1.47; 95% CI, 0.36-5.98; $P = .59$). Necrosis (>1%) was associated with shorter DFS (HR, 3.11; 95% CI, 1.36-7.14; $P = .007$), while sclerohyalinosis greater than 20% was

(continued)

Key Points

Question Does histopathological response after neoadjuvant chemotherapy (NACT) estimate outcomes in patients with high-risk soft-tissue sarcoma (STS) of extremity or trunk wall?

Findings In this preplanned secondary analysis of a clinical trial involving 388 patients, the proportion of stainable tumor cells was not associated with disease-free survival (DFS). Necrosis was associated with worse DFS, and presence of sclerohyalinosis greater than 20% estimated improved DFS.

Meaning These findings suggest that the presence of sclerohyalinosis may serve as a novel histopathological marker of favorable response to NACT in STS, challenging the current emphasis on the proportion of stainable tumor cells and informing patient risk stratification and treatment evaluation.

+ Supplemental content

Author affiliations and article information are listed at the end of this article.

Open Access. This is an open access article distributed under the terms of the CC-BY License.

Abstract (continued)

associated with longer DFS (HR, 0.51; 95% CI, 0.28-0.94; $P = .03$). Exclusion of patients who received preoperative RT did not alter these associations. In patients randomly assigned to anthracycline plus ifosfamide ($n = 98$), sclerohyalinosis greater than 20% remained associated with longer DFS (HR, 0.24; 95% CI, 0.09-0.67; $P = .007$). These findings were confirmed when a broader cohort ($n = 187$) was included.

CONCLUSIONS AND RELEVANCE In this secondary analysis of a randomized clinical trial, the proportion of stainable tumor cells, currently considered as the most relevant posttreatment change, did not stratify patient risk. The findings support consideration of the presence of sclerohyalinosis (>20%) to identify patients with the best outcome after NACT.

JAMA Network Open. 2025;8(11):e2540177. doi:10.1001/jamanetworkopen.2025.40177

Introduction

Histopathological response to neoadjuvant therapies, usually defined as presence of residual stainable tumor cells and other posttreatment changes, is considered a surrogate end point for identifying patients who benefit from these treatment strategies.¹⁻⁴ Soft-tissue sarcomas (STS) are a rare group of malignant neoplasms in which surgery is the mainstay of treatment.⁵ Neoadjuvant chemotherapy (NACT) is considered when the risk of tumor metastasis is high to increase the chance of cure for patients, particularly in selected STS histotypes emerging in the extremity or trunk wall.⁵

The value of quantification of residual stainable tumor cells in estimating patient outcomes is well established for Ewing sarcoma of bone and osteosarcoma.^{6,7} Despite challenges in interpreting pathological responses in these tumors (eg, potential misclassification due to cutoff use), histopathological changes after neoadjuvant treatment remain among the most relevant factors in estimating patient outcomes. In STS, tumor necrosis represents an intrinsic feature of STS characterized by high malignant neoplasm grade,^{8,9} which is one of the major determinants of the indication for NACT.^{10,11} Morphological analysis cannot differentiate between tumor-intrinsic and treatment-induced necrosis, thus lowering the utility of this tumor characteristic for assessing treatment response in STS. Retrospective analysis from a clinical trial on NACT associated tumor response at magnetic resonance imaging (MRI) with stainable tumor cells, which defined a very good response when they were represented in less than 10% of the tumor. The European Organization for Research and Treatment of Cancer–Soft Tissue and Bone Sarcoma Group (EORTC-STBSG) included this information in a classification of histopathological tumor response after NACT. Consensus on the role of quantification of residual stainable tumor cells in posttreatment changes, including their cutoffs, is lacking. We hypothesized that NACT with or without radiotherapy (RT) induces histopathological changes to tumor tissue that could estimate patient outcomes. In this secondary analysis, we aimed to characterize morphological changes in surgical specimens of patients treated with NACT with or without RT to identify histopathological features that stratify risk of recurrence and ultimately estimate the benefit from neoadjuvant treatments.

Methods

This preplanned prospective secondary analysis of the ISG-STG 1001 clinical trial (ClinicalTrials.gov Identifier: [NCT01710176](https://clinicaltrials.gov/ct2/show/study/NCT01710176)) was conducted from 2011 to 2020. Information about this trial, along with the study protocol, is described elsewhere¹²⁻¹⁵ and is available in [Supplement 1](#). In brief, the ISG-STG 1001 trial, conducted at 32 centers across Italy, Spain, France, and Poland, included both a randomized clinical trial (between 2011 and 2016) and a nonrandomized patient cohort (between 2016 and 2020). The randomized trial was an open-label, phase 3 study that randomly assigned

patients to receive either 3 cycles of anthracycline plus ifosfamide or histotype-tailored (also termed *histology tailored*) NACT followed by surgery. The appropriate independent ethics committee at each participating center approved the protocol and all amendments. The ISG-ST5 1001 trial was conducted in accordance with the Declaration of Helsinki.¹⁶ All patients provided written informed consent before enrollment. We followed the Consolidated Standards of Reporting Trials (CONSORT) reporting guideline.

Assessment of Histopathological Response

Histopathological analysis of posttreatment surgical specimens was performed, assessing all available slides. For each enrolling center, 1 local pathologist was responsible for sampling the surgical specimen, following the study protocol. This process was conducted in collaboration with the operating surgeon and, where possible, the radiologist, leading to a histopathological diagnosis. Subsequently, blinded national central reviewers (1 per participating country) were engaged to independently review all national cases and complete the corresponding forms. Whenever possible, cases underwent histological analysis during meetings that included both pathologists and radiologists, occurring concurrently with the radiological review. A lead investigator (P.C.) meticulously reviewed all submitted forms. In the event of discrepancies, cases were requested for reevaluation until a consensus was achieved.

The following histopathological features were considered and scored a percentage relative to the whole tumor bed: proportion of stainable tumor cells in the tumor bed and each posttreatment change in the tumor bed, including tumor necrosis, hemorrhage, fibrohistiocytic reaction with hemosiderin, sclerosis or fibrosis, and sclerohyalinosis (**Figure 1**). The sum of the percentages was 100%. The occurrence of a cystic component was considered, especially for synovial sarcomas where posttreatment macroscopic diameter was shorter than that measured at preoperative radiological imaging. Stainable tumor cells were considered to be neoplastic cells that retained their structural integrity and properly absorbed hematoxylin and eosin staining, allowing for clear visualization of nuclear and cytoplasmic features. Stainable cells are characterized by distinctly visible nuclei under hematoxylin. The disappearance of this staining is a key diagnostic marker, serving as a histological sign of cell necrosis. Stainable tumor cells were quantified as a percentage that represents the proportion between the areas of stainable tumor cells and areas of the tumor.

Sclerosis or fibrosis was defined by the proliferation of fibroblasts and increased deposition of collagen fibers in the absence of a hyaline matrix. Histologically, it was presented as an expansion of the stromal component with interwoven or compact collagen bundles, often accompanied by spindle-shaped fibroblasts.

Sclerohyalinosis (eFigure 1 in [Supplement 2](#)) was defined by the presence of a dense, homogenized collagen with a paucicellular deposition of an eosinophilic hyaline matrix. Histologically, it exhibited a glassy, acellular appearance with sparse, atrophic or spindle-shaped fibroblasts embedded within the matrix. Scattered inflammatory cells—predominantly lymphocytes and macrophages—may be present, along with rare stainable tumor cells. The hyalinized stroma may show focal dystrophic calcifications, and in some cases, small, obliterated vascular structures can be observed.

Statistical Analysis

Statistical analysis investigated the association between each characteristic of histopathological response and disease-free survival (DFS). DFS, which was the primary outcome of the ISG-ST5 1001 trial, was defined as the time between surgery and the first recurrence or the last follow-up for nonrecurrence.

First, analyses were conducted in patients included in the randomized cohort. Second, analyses were restricted to patients who received NACT without preoperative RT and to patients who were randomly assigned to receive anthracycline plus ifosfamide, to reduce possible confounding effect of

RT and histotype-tailored NACT, respectively. Third, the randomized and nonrandomized cohorts of the ISG-1001 trial were merged.

The proportion of stainable tumor cells was classified according to EORTC-STBSG categories or as absent or present. The continuous variable of sclerohyalinosis, expressed as a percentage, was categorized based on its distribution, and the second tertile was used as the cutoff point. Tumor necrosis, hemorrhage, fibrohistiocytic reaction with hemosiderin, and sclerosis or fibrosis, also expressed as a percentage, were classified as absent or present.

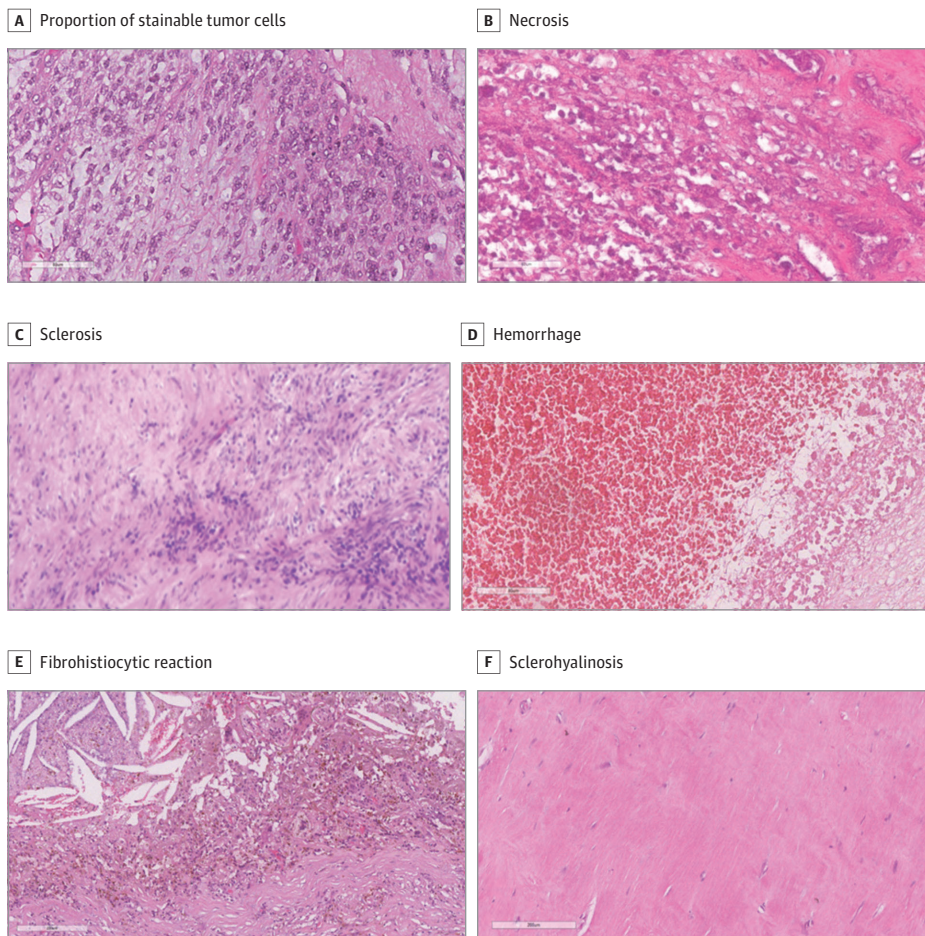
Detailed information about statistical tests used in this analysis are provided in the eMethods in Supplement 2. Data analyses were performed from January to June 2023 using SAS Studio, version 5.2 (SAS Institute Inc). Statistical significance was set at $P = .05$.

Results

Three hundred eighty-eight patients (median [IQR] age, 50 [41-60] years; 143 females [36.9%], 245 males [63.1%]), from a total of 548 enrolled in the ISG-ST5 1001 clinical trial, had surgical specimens including a residual tumor evaluable for histopathological response (Figure 2, Table 1).

Histopathological response was assessed in 201 (70.0%) of 287 patients in the randomized cohort. Among excluded patients, 47 had re-excision without macroscopic disease, 7 did not have surgery, and 32 had missing information on pathological response. Primary outcome of this trial is detailed in previous reports.^{12,13} In addition, the present analysis included 24 patients with myxoid liposarcoma

Figure 1. Features Considered to Characterize Histopathological Tumor Response in Trial Participants Treated With Neoadjuvant Chemotherapy With or Without Radiotherapy



Histopathological images show residual stainable tumor cells (A) and posttreatment changes (B-F) in a surgical specimen of pretreated sarcoma (formalin-fixed paraffin-embedded tissue with hematoxylin-eosin stain magnification x200).

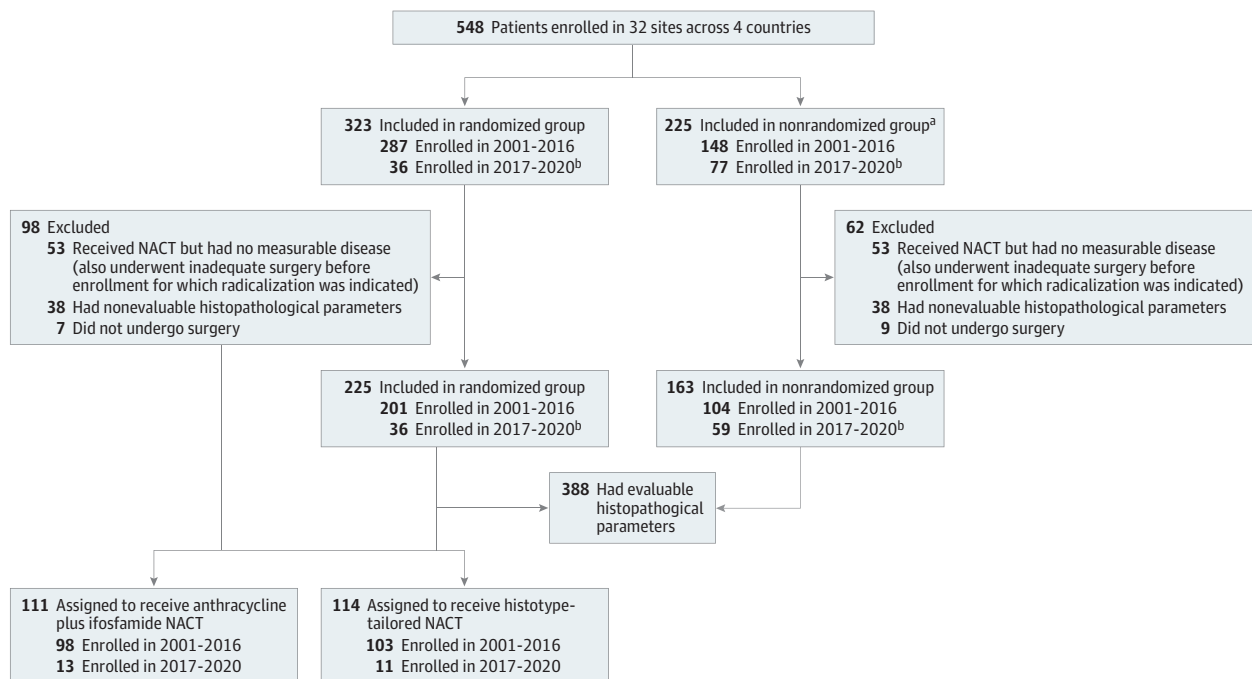
who were enrolled in the expansion cohort of the randomized trial.¹⁴ In this analysis, we also evaluated data from 163 patients in the nonrandomized cohort. Patient and tumor characteristics are reported in Table 1.

After a median (IQR) follow-up of 72 (43-93) months, 156 of 388 patients (40.2%) developed a disease recurrence. In the randomized cohort (n = 201), the median (IQR) follow-up was 86 (70-99) months, and 115 patients (57.2%) developed a disease recurrence.

The distribution of characteristics of histopathological response to neoadjuvant treatment suggested a relatively low prevalence for each of the considered morphological features (eTable and eFigure 2 in Supplement 2). In the randomized cohort, a proportion of 1% or more stainable tumor cells were detected in most patients (194 [96.5%]), underlying the rare occurrence of a pathological complete response in high-risk STS treated with 3 cycles of NACT. Presence ($\geq 1\%$) of necrosis in 169 patients (84.1%) was associated with shorter DFS (hazard ratio [HR], 3.11; 95% CI, 1.36-7.14; $P = .007$) (Figure 3A), and greater than 20% of sclerohyalinosis in 42 patients (20.9%) was associated with a lower risk of recurrence (HR, 0.51; 95% CI, 0.28-0.94; $P = .03$) (Figure 3B). The other evaluated characteristics of histopathological response did not show any association with DFS (Table 2).

The proportion of stainable tumor cells was not associated with DFS either when dichotomized as present or absent (HR, 1.47; 95% CI, 0.36-5.98; $P = .59$) or when categorized according to the EORTC-STBSG classification (Table 2, Figure 3C). Only 7 patients (3.5%) had absence of stainable tumor cells or a proportion of stainable tumor cells lower than 1%. To further investigate these findings, patients who had neoadjuvant RT together with NACT (32 [15.9%]) were excluded from this analysis, leaving patients treated only with NACT. The findings confirmed that the presence of necrosis in 142 (84.1%) was associated with shorter DFS (HR, 3.06; 95% CI, 1.23-7.60; $P = .02$), while greater than 20% of sclerohyalinosis detected in 31 patients (18.3%) was associated with a reduced risk of recurrence (HR, 0.45; 95% CI, 0.21-0.93; $P = .03$). The main outcome of the ISG-ST5 1001 trial suggested that anthracycline plus ifosfamide was the regimen preferred for high-risk patients

Figure 2. CONSORT Diagram of the ISG-1001 Clinical Trial



NACT indicates neoadjuvant chemotherapy.

^a Affected by histologies not randomized after central pathological review and treated with anthracycline plus ifosfamide.

^b Includes classification of patients after protocol amendment 3.0 in January 2017.

compared with histotype-tailored NACT. Consequently, we considered only patients treated with anthracycline plus ifosfamide in the randomized cohort and observed the association between presence of sclerohyalinosis higher than 20% and lower risk of recurrence (24 of 98 [24.5%]; HR, 0.24 [95% CI, 0.09-0.67], $P = .007$). Tumor necrosis remained associated with shorter DFS in patients who received histotype-tailored NACT (84 of 103 [85.4%]; HR, 9.78 [95% CI, 1.35-71.18], $P = .02$), supporting the hypothesis of necrosis being tumor-intrinsic rather treatment-related.

To increase the robustness of these results, both cohorts of randomized and nonrandomized patients were analyzed together ($N = 388$). The analysis showed that the presence of necrosis (313 [80.6%]; HR, 2.54 [95% CI, 1.51-4.26], $P < .01$) and sclerohyalinosis greater than 20% (111 [28.6%]; HR, 0.64 [95% CI, 0.44-0.93], $P = .02$) could be used to estimate DFS. The analysis of the overall patient series also confirmed the lack of association when the proportion of stainable tumor cells was dichotomized as present or absent and when categorized according to the EORTC-STBSG classification (Figure 3D). To address potential confounding from the pooled data from both randomized and nonrandomized patients, we adjusted for baseline variables and largely found consistency with the main analysis. No association between DFS and sclerohyalinosis was found.

Table 1. Clinical-Pathological Features of Patients With High-Risk Soft-Tissue Sarcomas of Extremity or Trunk Wall Treated With NACT With or Without Radiotherapy and Surgery in the ISG-1001 Clinical Trial

Characteristic	Patients, No. (%)				
	Anthracycline plus ifosfamide NACT (n = 274)			Histotype-tailored NACT (n = 114)	
	All (n = 274)	Randomized (n = 111)	Nonrandomized (n = 163)	Randomized (n = 114)	All (N = 388)
Age, y					
Mean (SD)	50.5 (12.0)	47.6 (11.2)	52.5 (12.2)	48.6 (13.4)	49.9 (12.5)
Median (IQR)	51 (41-60)	47 (39-57)	53 (44-62)	50 (40-60)	50.5 (41-60)
Sex					
Male	173 (63.1)	70 (63.1)	103 (63.2)	72 (63.2)	245 (63.1)
Female	101 (36.9)	41 (36.9)	60 (36.8)	42 (36.8)	143 (36.9)
Tumor size, mm					
Mean (SD)	119.1 (54.9)	114.1 (45.9)	122.5 (60.2)	111 (66.3)	116.7 (58.6)
Range	10-350	32-270	10-350	46-680	10-680
Median (IQR)	110 (80-149)	107 (83-135)	115 (80-152)	99.5 (75-140)	107 (80-142.5)
Histologic type					
High-grade myxoid liposarcoma	70 (25.6)	35 (31.5)	35 (21.5)	31 (27.2)	101 (26.0)
Synovial sarcoma	40 (14.6)	25 (22.5)	15 (9.2)	25 (21.9)	65 (16.8)
Malignant peripheral nerve sheath tumor	10 (3.7)	10 (9.0)	0	7 (6.1)	17 (4.4)
Leiomyosarcoma	22 (8.0)	11 (9.9)	11 (6.8)	11 (9.7)	33 (8.5)
Undifferentiated pleomorphic sarcoma	67 (24.5)	28 (25.2)	39 (23.9)	40 (35.1)	107 (27.6)
Myxofibrosarcoma ^a	37 (13.5)	0	37 (22.7)	0	37 (9.5)
Unclassified spindle cell ^a	8 (2.9)	1 (0.9)	7 (4.3)	0	8 (2.1)
Pleomorphic liposarcoma ^a	14 (5.1)	1 (0.9)	13 (8.0)	0	14 (3.6)
Pleomorphic rhabdomyosarcoma	6 (2.2)	0	6 (3.7)	0	6 (1.6)
Radiotherapy					
Preoperative	117 (42.7)	18 (16.2)	99 (60.7)	17 (14.9)	134 (34.5)
Postoperative	122 (44.5)	76 (68.5)	46 (28.2)	81 (71.1)	203 (52.3)
Preoperative and postoperative	9 (3.3)	2 (1.8)	7 (4.3)	1 (0.9)	10 (2.6)
None	26 (9.5)	15 (13.5)	11 (6.8)	15 (13.2)	41 (10.6)

Abbreviation: NACT, neoadjuvant chemotherapy.

^a Histologies allowed only in the nonrandomized cohort of the study.

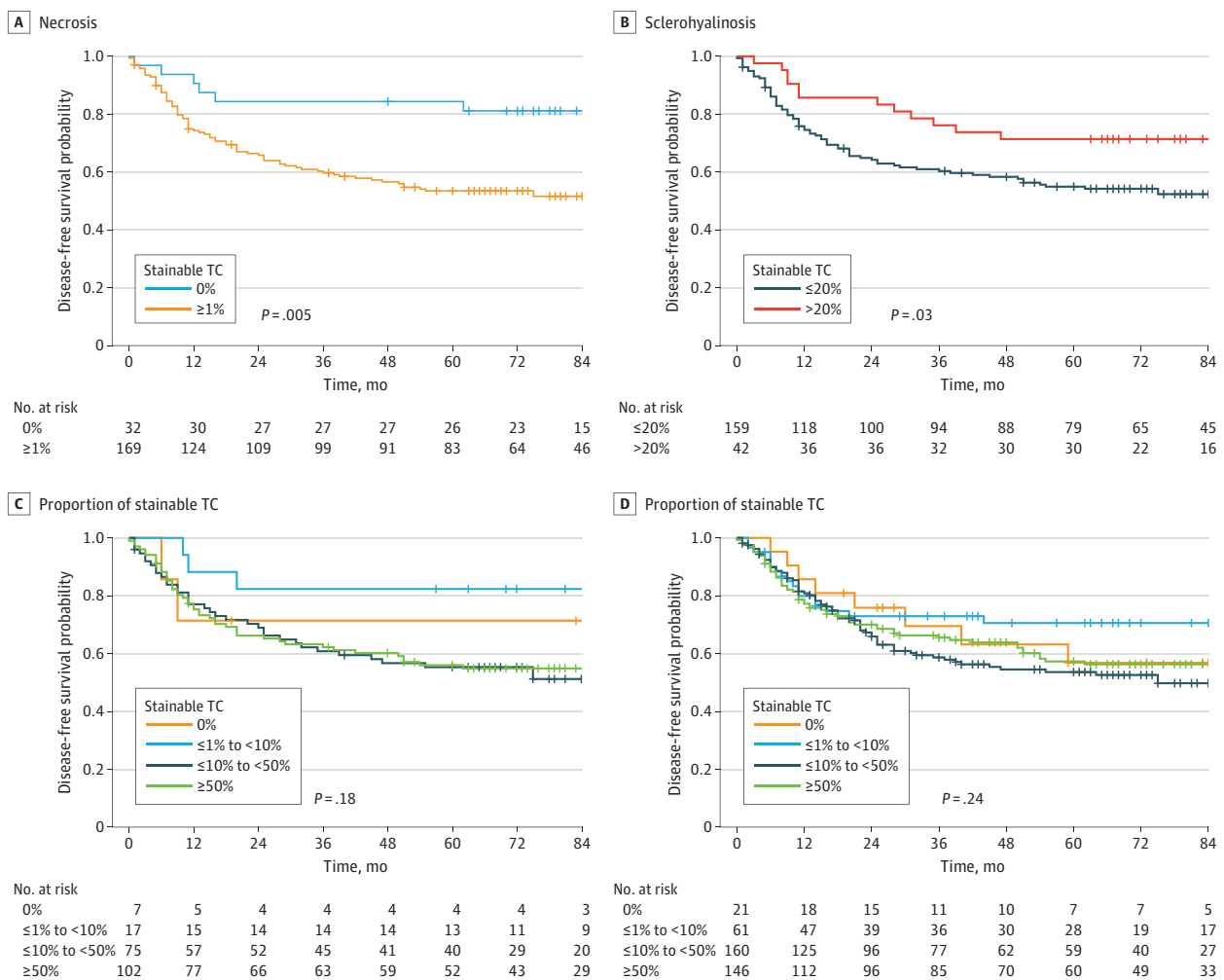
Discussion

To our knowledge, this secondary analysis is the first prospective study to provide an in-depth characterization of the histopathological response to NACT in patients with high-risk STS of extremities or trunk wall. The proportion of stainable tumor cells, considered as the most relevant posttreatment change in STS,¹⁷ did not stratify patient risk. This study supported consideration of the occurrence of more than 20% of sclerohyalinosis to identify patients with the best outcome after anthracycline-based NACT.

Criteria for Histopathological Response

In sarcomas, criteria for stratification of histopathological response have been established for osteosarcoma^{6,7} and Ewing sarcoma.^{7,18,19} In osteosarcoma, the percentage of histological response, including necrosis, fibrosis, and calcification, was ultimately considered to be a proxy of resistance to treatment and presence of minimal residual disease.²⁰ As a consequence, histological response is routinely documented in postoperative pathology report.²⁰ In osteosarcoma, a response is considered good when the proportion of stainable tumor cells is 10% or less or when the histological response is 90% or greater.⁶ For Ewing sarcoma of bone, the optimal threshold is less clearly

Figure 3. Kaplan-Meier Curves for Disease-Free Survival by Necrosis, Sclerohyalinosis, and Pathologic Tumor Response in the Randomized Cohort and by All Evaluable Patients



TC indicates tumor cell.

defined. Previous studies suggest that a complete (100%) response is the best indicator of a favorable outcome,²¹ whereas earlier reports classified a good response as achieving between 90% and 100% posttreatment changes.^{19,22} The same level of evidence has not been reached to date for STS, where studies have been conducted on different regimens, including systemic NACT mainly based on regimens containing an anthracycline, locoregional NACT administered through isolated limb perfusion, RT, or a combination of RT and chemotherapy.²³⁻³⁵ Histopathological response was retrospectively analyzed in the phase 3 clinical trial that compared 3 cycles with 5 cycles of perioperative chemotherapy.^{12,23,24,36} Histopathological posttreatment modifications after NACT correlated with tumor response detected at preoperative magnetic resonance imaging.²³ Radiologic changes were associated with better outcomes for patients who showed a histopathological response.²⁴ A substantial histopathological response (considered as a very good response) was defined as the presence of 10% or less stainable tumor cells after neoadjuvant treatments and surgery.²³ This evidence enables the EORTC-STBSG to generate their scoring system for histopathological response given the rate of stainable tumor cells,¹⁷ and this scoring system has been prospectively tested in the ISG-ST5 1001 study. Further analysis of the consistency between histopathological and radiologic characteristics of tumor response in a subgroup of patients warrants additional research.

In other retrospective studies, large variations exist, including but not limited to heterogeneity in STS histotypes, chemotherapy drugs and schedules, the combination of neoadjuvant RT, time interval between NACT surgery, and characteristics of histopathological response.²³⁻³⁵ These studies included a relatively high prevalence of patients with low-risk STS who were treated primarily with neoadjuvant RT and achieved a pathologic complete response, generally defined as the presence of only posttreatment changes and absence of stainable tumor cells in approximately 25% of patients.²⁶ This result was consistent with a recent retrospective study of patients with localized STS treated with preoperative RT alone or combined with chemotherapy in the NRG Oncology/Radiation Therapy Oncology 9514 and O630 nonrandomized clinical trials.³⁷ In the present study,

Table 2. Association Between Characteristics of Histopathological Response and Patient Disease-Free Survival in Patients in the Randomized Cohort of the ISG-1001

Variable and category, %	Patients, No. (%)		HR (95% CI)	P value
	Total (n = 201)	No association (n = 86)		
Proportion of stainable tumor cells				
0, Reference group	7 (3.5)	2 (2.3)	NA	NA
>0 to <1	0	NA	NA	NA
1 to <10	17 (8.5)	3 (3.5)	0.50 (0.08-2.98)	.44
10 to <50	75 (37.3)	36 (41.9)	1.66 (0.40-6.90)	.49
≥50	102 (50.7)	45 (52.3)	1.53 (0.37-6.30)	.56
Tumor necrosis				
0, Reference group	32 (15.9)	6 (7.0)	NA	.007
≥1	169 (84.1)	80 (93.0)	3.11 (1.36-7.14)	
Hemorrhage				
0, Reference group	134 (66.7)	56 (65.1)	NA	.44
≥1	67 (33.3)	30 (34.9)	1.19 (0.76-1.86)	
Fibrohistiocytic reaction with hemosiderin				
0, Reference group	165 (82.1)	69 (80.2)	NA	.28
≥1	36 (17.9)	17 (19.8)	1.34 (0.79-2.28)	
Fibrosis or sclerosis				
0, Reference group	140 (69.7)	53 (61.6)	NA	.05
≥1	61 (30.3)	33 (38.4)	1.54 (1.00-2.38)	
Sclerohyalinosis ^a				
≤20, Reference group	159 (79.1)	74 (86.0)	NA	.03
>20	42 (20.9)	12 (14.0)	0.51 (0.28-0.94)	

Abbreviations: HR, hazard ratio; NA, not applicable.

^a Sclerohyalinosis was categorized on the second tertile.

only a minority of patients (7 of 201 [3.5%]) had a proportion of stainable tumor cells less than 1%, and this low proportion likely explains the lack of association with patient outcomes. We cannot ascertain whether the low response rate was associated with the study design, particularly the 3 cycles of NACT, which may not lead to clinically significant histopathological response, as previously discussed.

Sclerohyalinosis

In this study, the presence of greater than 20% of sclerohyalinosis estimated patient DFS, which is consistent with the complete substitution of tumor and tumor microenvironment with acellular sclerohyaline matrix. This feature was highlighted in a previous EORTC study of neoadjuvant RT in STS.³⁸ In contrast to the proportion of stainable tumor cells, the presence of at least 20% hyalinization and fibrosis was associated with improved estimation of patient outcome and, if confirmed in future studies, could serve as a meaningful end point in neoadjuvant clinical trials. However, it remains impossible to assess the magnitude of the increased proportion of sclerohyalinosis after NACT compared with the baseline proportion.

Several STS histotypes are characterized by the transcription of gene sets related to extracellular matrix and exhibit a spectrum of proteins involved in its remodeling.^{39,40} Sclerohyalinosis was associated with outcomes in patients with melanoma treated with neoadjuvant immune checkpoint inhibitors or *BRAF/MEK*-targeted therapies for locally advanced disease.^{41,42} The association between sclerohyalinosis and DFS observed in our study suggests possible mechanisms of extracellular matrix remodeling through chemotherapy-induced changes in the tumor cells and tumor microenvironment. In the ISG-STS 1001 trial, we showed the association between anthracycline-based NACT and changes in tumor immune microenvironment, a condition that further supports the hypothesis that sclerohyalinosis is treatment-induced. In undifferentiated pleomorphic sarcoma, the NEOSARCOMICS trial also confirmed increased survival for patients with immune infiltration, although it suggested a lack of correlation between immune infiltrate and histopathological response.⁴³ It remains unclear whether the identified threshold of sclerosis-hyalinosis is dependent on tumor histologic type and treatment regimen—in terms of both type of drugs used and duration of therapy—or if it can be considered as a more generalizable parameter.

Limitations

This analysis is limited in several ways. First, it lacked validation in independent series of high-risk STS. This limitation extends to the data-driven cutoff used for sclerohyalinosis (>20%), which might restrict its generalizability and hence necessitate future studies to validate and refine this cutoff. However, surgical specimens were prospectively and systematically sampled and evaluated by pathologists with expertise in soft-tissue tumors, which resulted in an unprecedented collection of data. Second, our evaluations were limited by the inability to assess histopathological response in the tumor mass before neoadjuvant therapy. Therefore, this study cannot determine the extent to which the observed tumor characteristics after NACT are true posttreatment changes or intrinsic features of the tumor. Third, although we selected the most prevalent sarcoma histotypes in the extremities and trunk wall, it was not powered to evaluate differences in the characteristics of histopathological response for each sarcoma histotype. Therefore, we cannot rule out possible differences in response to NACT containing anthracycline plus ifosfamide with or without RT across different sarcomas, as suggested by histologic type-specific retrospective analysis in undifferentiated pleomorphic sarcoma and myxofibrosarcoma. To minimize this issue, we accounted for some differences, such as the need to adapt tumor measurements in case of cystic synovial sarcoma. The addition of neoadjuvant RT to chemotherapy may represent a substantial confounder; thus, as a mitigation strategy, we performed subgroup analyses.

Finally, it may be possible that 3 cycles of NACT may not result in a meaningful histopathological response. This study tested 3 cycles of full-dose anthracyclines plus ifosfamide, which was consistently tested following the previous randomized clinical trial performed by the Italian and

Spanish sarcoma groups (ISG and GEIS) and demonstrated the noninferiority of 3 cycles over 5 cycles of anthracycline plus ifosfamide for DFS and overall survival.^{36,44} Three cycles were not meant to maximize the histopathological tumor response, which may be higher with the implementation of 5 cycles and a more extensive use of preoperative RT. Conversely, the aim of 3 cycles was to lower systemic risk and minimize toxic effect. When the intent of the preoperative treatment is local, increasing NACT and combining it with RT may be considered.

Conclusions

The findings of this secondary analysis of a clinical trial of patients with high-risk STS treated with NACT questioned the EORTC-STBSG criteria for assessment of histopathological response to chemotherapy, making the assessment of response to radiologic imaging likely more effective at this stage.^{23,24,45} Specifically, the proportion of stainable tumor cells, considered as the most relevant posttreatment change, did not stratify patient risk and supported the consideration of presence of sclerohyalinosis (>20%) in identifying patients with the best outcome after NACT. Additionally, this study suggests caution when a clinical trial in patients with primary STS considers histopathological response as the primary end point.

ARTICLE INFORMATION

Accepted for Publication: September 3, 2025.

Published: November 6, 2025. doi:10.1001/jamanetworkopen.2025.40177

Open Access: This is an open access article distributed under the terms of the [CC-BY License](#). © 2025 Pasquali S et al. *JAMA Network Open*.

Corresponding Authors: Paola Collini, MD, Soft Tissue Tumor Pathology Unit, Department of Advanced Diagnostics (paola.collini@istitutotumori.mi.it); Alessandro Gronchi, MD, Sarcoma Service, Department of Surgery (alessandro.gronchi@istitutotumori.mi.it), Fondazione IRCCS Istituto Nazionale dei Tumori, Milano, Italy.

Author Affiliations: Molecular Pharmacology, Department of Experimental Oncology, Fondazione Istituto di Ricovero e Cura a Carattere Scientifico (IRCCS) Istituto Nazionale dei Tumori di Milano, Milano, Italy (Pasquali); Soft Tissue Tumor Pathology Unit, Department of Advanced Diagnostics, Fondazione IRCCS Istituto Nazionale dei Tumori di Milano, Milano, Italy (Collini); Pathology Department, Vall d'Hebron University Hospital, Barcelona, Spain (Romagosa); Department of Pathology, Institut Bergonié, Bordeaux, France (Coindre); Institut National de la Santé et de la Recherche Médicale (INSERM) U1218 ACTION, Institut Bergonié, Bordeaux, France (Coindre); Unit of Bioinformatics and Biostatistics, Fondazione IRCCS Istituto Nazionale dei Tumori, Milan, Italy (Pizzamiglio, Verderio, Duroni); Pathology Unit, Azienda SocioSanitaria Territoriale (ASST) Fatebenefratelli Sacco, Milan, Italy (Barisella); Osteoncology, Bone and Soft Tissue Sarcomas and Innovative Therapies Unit IRCCS Istituto Ortopedico Rizzoli, Bologna, Italy (Palmerini); Miller School of Medicine, University of Miami, Miami, Florida (Palmerini); Surgery Department, IRCCS Humanitas Research Hospital, Rozzano, Italy (Quagliuolo); Oncology Department, Fundación Jiménez Díaz University Hospital, Madrid, Spain (Martin Broto); Medical Oncology Department, Hospital de la Santa Creu i Sant Pau, Carrer de Sant Quintí, Barcelona, Spain (Pousa); Medical Oncology Unit, Città della Salute e della Scienza Hospital, Turin, Italy (Grignani); Department of Oncology, Medical Oncology 1 Unit, Istituto Oncologico Veneto IOV IRCCS, Padova, Italy (Brunello); Centre Léon Bérard & Université Claude Bernard Lyon 1, Lyon, France (Blay); Department of Soft Tissue/Bone Sarcoma and Melanoma, Centrum Onkologii, Instytut, Marii Skłodowskiej-Curie, Warsaw, Poland (Lugowska); Clinical Trial Center and Department of Epidemiology, IRCCS Ospedale Policlinico San Martino, IST Istituto Nazionale per la Ricerca sul Cancro, Genoa, Italy (Fontana); Orthopedic Oncology Unit, IRCCS Istituto Ortopedico Rizzoli, Bologna, Italy (Bianchi); Department of Cancer Medicine, Fondazione IRCCS Istituto Nazionale dei Tumori di Milano, Milano, Italy (Palassini, Casali, Stacchiotti); Department of Pathology, IRCCS Humanitas Research Hospital, Rozzano, Milan, Italy (Renne); Surgical Pathology and Cytopathology Unit, Department of Medicine-DIMED, University of Padua, Padua, Italy (Sbaraglia, Dei Tos); Department of Data Science, Fondazione IRCCS Istituto Nazionale dei Tumori di Milano, Milano, Italy (Miceli); Department of Pathology, IRCCS Istituto Ortopedico Rizzoli, Bologna, Italy (Gambarotti); Pathology Department, Hospital de la Santa Creu i Sant Pau, Universitat Autònoma de Barcelona, Barcelona, Spain (Bagué); Sarcoma Service, Department of Surgery, Fondazione IRCCS Istituto Nazionale dei Tumori di Milano, Milano, Italy (Gronchi).

Author Contributions: Drs Gronchi and Collini had full access to all of the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis.

Concept and design: Pasquali, Collini, Verderio, Duroni, Fontana, Renne, Casali, Dei Tos, Stacchiotti, Gronchi.

Acquisition, analysis, or interpretation of data: Pasquali, Collini, Romagosa, Coindre, Pizzamiglio, Verderio, Duroni, Barisella, Palmerini, Quagliuolo, Martín Broto, Lopez Pousa, Grignani, Brunello, Blay, Lugowska, Bianchi, Palassini, Renne, Casali, Miceli, Sbaraglia, Gambarotti, Bague, Dei Tos, Stacchiotti, Gronchi.

Drafting of the manuscript: Pasquali, Collini, Pizzamiglio, Verderio, Duroni, Blay, Fontana, Miceli, Gronchi.

Critical review of the manuscript for important intellectual content: Pasquali, Collini, Romagosa, Coindre, Verderio, Barisella, Palmerini, Quagliuolo, Martín Broto, Lopez Pousa, Grignani, Brunello, Blay, Lugowska, Bianchi, Palassini, Renne, Casali, Sbaraglia, Gambarotti, Bague, Dei Tos, Stacchiotti, Gronchi.

Statistical analysis: Pizzamiglio, Verderio, Duroni, Blay, Fontana, Miceli, Gronchi.

Obtained funding: Pasquali, Blay.

Administrative, technical, or material support: Pasquali, Romagosa, Barisella, Palmerini, Grignani, Brunello, Blay.

Supervision: Collini, Verderio, Duroni, Palmerini, Quagliuolo, Martín Broto, Lopez Pousa, Casali, Sbaraglia, Dei Tos, Stacchiotti, Gronchi.

Conflict of Interest Disclosures: Dr Romagosa reported receiving grants from the SELNET Consortium during the conduct of the study. Dr Palmerini reported receiving personal fees from Daiichi Sankyo, Deciphera, Eusa Pharma, SynOx Therapeutics, Ipsen Biopharmaceuticals, and Servier outside the submitted work. Dr Martín Broto reported receiving personal fees from PharmaMar, Bayer, GSK, Deciphera, Boehringer, Cogent, Roche, and Adium; receiving grants from Deciphera, PharmaMar, Eli Lilly, BMS, Pfizer, Boehringer, Synox, Abbisko, Biosplice, Lixte, Karyopharm, Rain, Inhibrx, Immunome, Phillogen, Cebiotex, PTC, and Springworks; and being CEO of Sarcoma Research Solutions outside the submitted work. Dr Grignani reported receiving personal fees and grants from PharmaMar during the conduct of the study and personal fees from Novartis, Incyte, Deciphera, Lilly, Merck, and Gentilini outside the submitted work. Dr Brunello reported receiving personal fees from Boehringer Ingelheim, Deciphera, and GSK and nonfinancial support from PharmaMar outside the submitted work. Dr Lugowska reported receiving personal fees from Roche, Pfizer, Incyte, Immunocore, Astra, Sairopa, MSD, Amgen, Ascendis, and Clininote and nonfinancial support from Agenus outside the submitted work. Dr Casali reported receiving grants from Abbisko Therapeutics, Advenchen, Ayala Pharm, Blueprint, Boehringer Ingelheim, Cogent Bio, Daiichi Sankyo, Deciphera, Eisai, Eli Lilly, Epizyme Inc, Foghorn, Glaxo Smith Kline, Hutchmed, Immunome, IDRX, Inhibrix, Karyopharm, Novartis, PharmaMar, Rain Therapeutics, SpringWorks, and Syneos Health outside the submitted work. Dr Stacchiotti reported receiving personal fees from Agenus, Bayer, Boehringer, Deciphera, Daiichi Sankyo, Gentili, Glaxo Smith Kline, Ipsen, Merck Serono, NEC Oncoimmunity, Parabilis, PharmaMar, and Servier; and being President of the Italian Sarcoma Group and President of the Connective Tissue Oncology Society outside the submitted work. Dr Gronchi reported receiving personal fees and grants from PharmaMar during the conduct of the study; personal fees from Novartis, Pfizer, Bayer, Lilly, Deciphera, SpringWorks, and Boehringer Ingelheim; and grants from Nanobiotix outside the submitted work. No other disclosures were reported.

Funding/Support: This work was supported by grants from PharmaMar, which provided trabectedin for high-grade myxoid liposarcoma to Italian Sarcoma Group (Dr Gronchi); grant EUROSARC FP7 278472 from the European Union; grants from NetSarc, LYRICAN (INCA-DGOS-INSERM 12563), and DEPGYN (RHU4) for the French sites; grant RF-2019-12370923 from the Italian Ministry of Health (Dr Gronchi); and AIRC Individual Grant-Next Gen Clinician Scientist "Fondazione 13 Marzo" (Dr Pasquali).

Role of the Funder/Sponsor: The funders had no role in the design and conduct of the study; collection, management, analysis, and interpretation of the data; preparation, review, or approval of the manuscript; and decision to submit the manuscript for publication.

Data Sharing Statement: See [Supplement 3](#).

REFERENCES

1. Lerner SP, McConkey DJ, Tangen CM, et al. Association of molecular subtypes with pathologic response, PFS, and OS in a phase II study of COXEN with neoadjuvant chemotherapy for muscle-invasive bladder cancer. *Clin Cancer Res*. 2024;30(2):444-449. doi:10.1158/1078-0432.CCR-23-0602
2. Deutsch JS, Cimino-Mathews A, Thompson E, et al. Association between pathologic response and survival after neoadjuvant therapy in lung cancer. *Nat Med*. 2024;30(1):218-228. doi:10.1038/s41591-023-02660-6
3. Yam C, Abuhadra N, Sun R, et al. Molecular characterization and prospective evaluation of pathologic response and outcomes with neoadjuvant therapy in metaplastic triple-negative breast cancer. *Clin Cancer Res*. 2022;28(13):2878-2889. doi:10.1158/1078-0432.CCR-21-3100

4. D'Alessio A, Stefanini B, Blanter J, et al. Pathological response following neoadjuvant immune checkpoint inhibitors in patients with hepatocellular carcinoma: a cross-trial, patient-level analysis. *Lancet Oncol*. 2024;25(11):1465-1475. doi:10.1016/S1470-2045(24)00457-1
5. Gronchi A, Miah AB, Dei Tos AP, et al; ESMO Guidelines Committee, EURACAN and GENTURIS. Electronic address: clinicalguidelines@esmo.org. Soft tissue and visceral sarcomas: ESMO-EURACAN-GENTURIS Clinical Practice Guidelines for diagnosis, treatment and follow-up[☆]. *Ann Oncol*. 2021;32(11):1348-1365. doi:10.1016/j.annonc.2021.07.006
6. Picci P, Bacci G, Campanacci M, et al. Histologic evaluation of necrosis in osteosarcoma induced by chemotherapy: regional mapping of viable and nonviable tumor. *Cancer*. 1985;56(7):1515-1521. doi:10.1002/1097-0142(19851001)56:7<1515::AID-CNCR2820560707>3.0.CO;2-6
7. Huvos AG, Rosen G, Marcove RC. Primary osteogenic sarcoma: pathologic aspects in 20 patients after treatment with chemotherapy en bloc resection, and prosthetic bone replacement. *Arch Pathol Lab Med*. 1977;101(1):14-18.
8. Coindre JM, Trojani M, Contesso G, et al. Reproducibility of a histopathologic grading system for adult soft tissue sarcoma. *Cancer*. 1986;58(2):306-309. doi:10.1002/1097-0142(19860715)58:2<306::AID-CNCR2820580216>3.0.CO;2-7
9. Trojani M, Contesso G, Coindre JM, et al. Soft-tissue sarcomas of adults; study of pathological prognostic variables and definition of a histopathological grading system. *Int J Cancer*. 1984;33(1):37-42. doi:10.1002/ijc.2910330108
10. Woll PJ, Reichardt P, Le Cesne A, et al; EORTC Soft Tissue and Bone Sarcoma Group and the NCIC Clinical Trials Group Sarcoma Disease Site Committee. Adjuvant chemotherapy with doxorubicin, ifosfamide, and lenograstim for resected soft-tissue sarcoma (EORTC 62931): a multicentre randomised controlled trial. *Lancet Oncol*. 2012;13(10):1045-1054. doi:10.1016/S1470-2045(12)70346-7
11. Pasquali S, Pizzamiglio S, Touati N, et al; EORTC – Soft Tissue and Bone Sarcoma Group. The impact of chemotherapy on survival of patients with extremity and trunk wall soft tissue sarcoma: revisiting the results of the EORTC-STBSG 62931 randomised trial. *Eur J Cancer*. 2019;109:51-60. doi:10.1016/j.ejca.2018.12.009
12. Gronchi A, Ferrari S, Quagliuolo V, et al. Neoadjuvant chemotherapy in high-risk soft tissue sarcomas: a randomised clinical trial from the Italian Sarcoma Group, the Spanish Sarcoma Group (GEIS), the Italian French Group (FSG) and the the Polish Sarcoma Group (PSG). *Lancet Oncol*. 2017;18(6):812-822. doi:10.1016/S1470-2045(17)30334-0
13. Gronchi A, Palmerini E, Quagliuolo V, et al. Neoadjuvant chemotherapy in high-risk soft tissue sarcomas: final results of a randomized trial from Italian (ISG), Spanish (GEIS), French (FSG), and Polish (PSG) Sarcoma Groups. *J Clin Oncol*. 2020;38(19):2178-2186. doi:10.1200/JCO.19.03289
14. Gronchi A, Palmerini E, Quagliuolo V, et al. Neoadjuvant chemotherapy in high-grade myxoid liposarcoma: results of the expanded cohort of a randomized trial from Italian (ISG), Spanish (GEIS), French (FSG), and Polish Sarcoma Groups (PSG). *J Clin Oncol*. 2024;42(8):898-906. doi:10.1200/JCO.23.00908
15. Pasquali S, Vallacchi V, Lalli L, et al. Spatial distribution of tumour immune infiltrate predicts outcomes of patients with high-risk soft tissue sarcomas after neoadjuvant chemotherapy. *EBioMedicine*. 2024;106:105220. doi:10.1016/j.ebiom.2024.105220
16. World Medical Association. World Medical Association Declaration of Helsinki: ethical principles for medical research involving human subjects. *JAMA*. 2013;310(20):2191-2194. doi:10.1001/jama.2013.281053
17. Wardelmann E, Haas RL, Bovée JV, et al. Evaluation of response after neoadjuvant treatment in soft tissue sarcomas: the European Organization for Research and Treatment of Cancer-Soft Tissue and Bone Sarcoma Group (EORTC-STBSG) recommendations for pathological examination and reporting. *Eur J Cancer*. 2016;53:84-95. doi:10.1016/j.ejca.2015.09.021
18. van der Woude HJ, Bloem JL, Taminiau AH, Nooy MA, Hogendoorn PC. Classification of histopathologic changes following chemotherapy in Ewing's sarcoma of bone. *Skeletal Radiol*. 1994;23(7):501-507. doi:10.1007/BF00223077
19. Akerman M. Tumour necrosis and prognosis in Ewing's sarcoma. *Acta Orthop Scand Suppl*. 1997;273:130-132. doi:10.1080/17453674.1997.11744718
20. Strauss SJ, Frezza AM, Abecassis N, et al; ESMO Guidelines Committee, EURACAN, GENTURIS and ERN PaedCan. Electronic address: clinicalguidelines@esmo.org. Bone sarcomas: ESMO-EURACAN-GENTURIS-ERN PaedCan Clinical Practice Guideline for diagnosis, treatment and follow-up. *Ann Oncol*. 2021;32(12):1520-1536. doi:10.1016/j.annonc.2021.08.1995
21. Albergo JI, Gaston CL, Laitinen M, et al. Ewing's sarcoma: only patients with 100% of necrosis after chemotherapy should be classified as having a good response. *Bone Joint J*. 2016;98-B(8):1138-1144. doi:10.1302/0301-620X.98B8.37346

22. Righi A, Pacheco M, Palmerini E, et al. Histological response to neoadjuvant chemotherapy in localized Ewing sarcoma of the bone: a retrospective analysis of available scoring tools. *Eur J Surg Oncol*. 2021;47(7):1778-1783. doi:10.1016/j.ejso.2021.02.009
23. Stacchiotti S, Collini P, Messina A, et al. High-grade soft-tissue sarcomas: tumor response assessment-pilot study to assess the correlation between radiologic and pathologic response by using RECIST and Choi criteria. *Radiology*. 2009;251(2):447-456. doi:10.1148/radiol.2512081403
24. Stacchiotti S, Verderio P, Messina A, et al. Tumor response assessment by modified Choi criteria in localized high-risk soft tissue sarcoma treated with chemotherapy. *Cancer*. 2012;118(23):5857-5866. doi:10.1002/cncr.27624
25. Danieli M, Barretta F, Radaelli S, et al. Pathological and radiological response following neoadjuvant treatments in primary localized resectable myxofibrosarcoma and undifferentiated pleomorphic sarcoma of the extremities and trunk wall. *Cancer*. 2023;129(21):3417-3429. doi:10.1002/cncr.34945
26. Bonvalot S, Wunder J, Gronchi A, et al. Complete pathological response to neoadjuvant treatment is associated with better survival outcomes in patients with soft tissue sarcoma: results of a retrospective multicenter study. *Eur J Surg Oncol*. 2021;47(8):2166-2172. doi:10.1016/j.ejso.2021.02.024
27. Salah S, Lewin J, Amir E, Abdul Razak A. Tumor necrosis and clinical outcomes following neoadjuvant therapy in soft tissue sarcoma: a systematic review and meta-analysis. *Cancer Treat Rev*. 2018;69:1-10. doi:10.1016/j.ctrv.2018.05.007
28. Andreou D, Werner M, Pink D, et al. Histological response assessment following neoadjuvant isolated limb perfusion in patients with primary, localised, high-grade soft tissue sarcoma. *Int J Hyperthermia*. 2016;32(2):159-164. doi:10.3109/02656736.2015.1109146
29. Andreou D, Werner M, Pink D, et al. Prognostic relevance of the mitotic count and the amount of viable tumour after neoadjuvant chemotherapy for primary, localised, high-grade soft tissue sarcoma. *Br J Cancer*. 2015;112(3):455-460. doi:10.1038/bjc.2014.635
30. Mullen JT, Hornicek FJ, Harmon DC, et al. Prognostic significance of treatment-induced pathologic necrosis in extremity and truncal soft tissue sarcoma after neoadjuvant chemoradiotherapy. *Cancer*. 2014;120(23):3676-3682. doi:10.1002/cncr.28945
31. Menendez LR, Ahlmann ER, Savage K, Cluck M, Fedenko AN. Tumor necrosis has no prognostic value in neoadjuvant chemotherapy for soft tissue sarcoma. *Clin Orthop Relat Res*. 2007;455(455):219-224. doi:10.1097/01.blo.0000238864.69486.59
32. Eilber FC, Rosen G, Eckardt J, et al. Treatment-induced pathologic necrosis: a predictor of local recurrence and survival in patients receiving neoadjuvant therapy for high-grade extremity soft tissue sarcomas. *J Clin Oncol*. 2001;19(13):3203-3209. doi:10.1200/JCO.2001.19.13.3203
33. Vaynrub M, Taheri N, Ahlmann ER, et al. Prognostic value of necrosis after neoadjuvant therapy for soft tissue sarcoma. *J Surg Oncol*. 2015;111(2):152-157. doi:10.1002/jso.23775
34. Gannon NP, Stemm MH, King DM, Bedi M. Pathologic necrosis following neoadjuvant radiotherapy or chemoradiotherapy is prognostic of poor survival in soft tissue sarcoma. *J Cancer Res Clin Oncol*. 2019;145(5):1321-1330. doi:10.1007/s00432-019-02885-4
35. Chang HY, Dermawan JK, Kuba MG, et al. Clinicopathologic and molecular correlates to neoadjuvant chemotherapy-induced pathologic response in breast angiosarcoma. *Genes Chromosomes Cancer*. 2024;63(5):e23240. doi:10.1002/gcc.23240
36. Gronchi A, Frustaci S, Mercuri M, et al. Short, full-dose adjuvant chemotherapy in high-risk adult soft tissue sarcomas: a randomized clinical trial from the Italian Sarcoma Group and the Spanish Sarcoma Group. *J Clin Oncol*. 2012;30(8):850-856. doi:10.1200/JCO.2011.37.7218
37. Wang D, Harris J, Kraybill WG, et al. Pathologic complete response and clinical outcomes in patients with localized soft tissue sarcoma treated with neoadjuvant chemoradiotherapy or radiotherapy: the NRG/RTOG 9514 and O630 nonrandomized clinical trials. *JAMA Oncol*. 2023;9(5):646-655. doi:10.1001/jamaoncol.2023.0042
38. Schaefer IM, Hornick JL, Barysaukas CM, et al. Histologic appearance after preoperative radiation therapy for soft tissue sarcoma: assessment of the European Organization for Research and Treatment of Cancer-Soft Tissue and Bone Sarcoma Group Response Score. *Int J Radiat Oncol Biol Phys*. 2017;98(2):375-383. doi:10.1016/j.ijrobp.2017.02.087
39. Pankova V, Krasny L, Kerrison W, et al. Clinical implications and molecular features of extracellular matrix networks in soft tissue sarcomas. *Clin Cancer Res*. 2024;30(15):3229-3242. doi:10.1158/1078-0432.CCR-23-3960
40. Eisinger-Mathason TS, Zhang M, Qiu Q, et al. Hypoxia-dependent modification of collagen networks promotes sarcoma metastasis. *Cancer Discov*. 2013;3(10):1190-1205. doi:10.1158/2159-8290.CD-13-0118

41. Rawson RV, Adhikari C, Bierman C, et al. Pathological response and tumour bed histopathological features correlate with survival following neoadjuvant immunotherapy in stage III melanoma. *Ann Oncol*. 2021;32(6):766-777. doi:10.1016/j.annonc.2021.03.006
42. Menzies AM, Amaria RN, Rozeman EA, et al. Pathological response and survival with neoadjuvant therapy in melanoma: a pooled analysis from the International Neoadjuvant Melanoma Consortium (INMC). *Nat Med*. 2021;27(2):301-309. doi:10.1038/s41591-020-01188-3
43. Guegan JP, El Ghazzi N, Vibert J, et al. Predictive value of tumor microenvironment on pathologic response to neoadjuvant chemotherapy in patients with undifferentiated pleomorphic sarcomas. *J Hematol Oncol*. 2024;17(1):100. doi:10.1186/s13045-024-01614-w
44. Gronchi A, Stacchiotti S, Verderio P, et al. Short, full-dose adjuvant chemotherapy (CT) in high-risk adult soft tissue sarcomas (STS): long-term follow-up of a randomized clinical trial from the Italian Sarcoma Group and the Spanish Sarcoma Group. *Ann Oncol*. 2016;27(12):2283-2288. doi:10.1093/annonc/mdw430
45. Pasquali S, Colombo C, Pizzamiglio S, et al. High-risk soft tissue sarcomas treated with perioperative chemotherapy: Improving prognostic classification in a randomised clinical trial. *Eur J Cancer*. 2018;93:28-36. doi:10.1016/j.ejca.2018.01.071

SUPPLEMENT 1.

Trial Protocol

SUPPLEMENT 2.

eMethods.

eFigure 1. Representation of Schelorohyalinosis

eFigure 2. Distribution of the Characteristics of Histopathological Response Considered in This Study in All Evaluable Patients n=388

eTable. Distribution of Characteristics of Histopathological Response

eReferences

SUPPLEMENT 3.

Data Sharing Statement