SUPPORTING INFORMATION

Chemiluminescence "add-and-measure" sensing paper based on Prussian Blue/Metal-Organic Framework MIL-101 nanozyme for rapid hydrogen peroxide detection

Héctor Martínez-Pérez-Cejuela[‡], Maria Maddalena Calabretta[†], Elisa Michelini^{†,¥,*}

[‡] Department of Analytical Chemistry, University of Valencia, Dr. Moliner 50, 46100, Burjassot, Valencia, Spain; [†]Department of Chemistry "Giacomo Ciamician", University of Bologna, Via P. Gobetti 85, 40129, Bologna, Italy; [¥] IRCCS Azienda Ospedaliero-Universitaria di Bologna, 40138 Bologna, Italy

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EXPERIMENTAL SECTION

Synthesis of MIL-101(Cr): 5 mmol of metal precursor (Cr(NO₃)₃·9H₂O) was weighed and mixed with 2 mmol of terephthalic acid (TA) in 15 mL of MQ-water. The molar ratio was set at 1:2.5:417 for TA:Cr(III):H₂O. The mixtures were homogenized and introduced in a Teflon-lined stainless steel autoclave. After its sealing, the mixture was heated up to 150 °C for 12 h. Then, the resulting green powder was collected by centrifugation at 14,000 g for 10 min and washed with DMF several times (3 x 10 mL). Finally, further purification was performed as indicated for MIL-101(Fe) in Section 2.4. with methanol at 60 °C and drying process.

Synthesis of MIL-101(Al): 2 mmol of AlCl₃·6H₂O and 3 mmol of terephthalic acid were solved in 40 mL of DMF. The molar ratio was set at 1:1.5:258 for Al(III):TA:DMF. The mixture was homogenized in the ultrasonic bath for 30 min and placed in a Teflon-lined stainless steel autoclave. The synthesis was performed for 72 h at 130 °C. The resulting palish yellow was centrifuged (14,000 g for 10 min) and washed with DMF several times (3 x 20 mL). Finally, further purification was performed as indicated for MIL-101(Fe) in the Section 2.4. with methanol at 60 °C and drying process.

Synthesis of bare PB-NPs: 1.1 mmol of citric acid was added to a previously prepared solution of 1 mM FeCl₃·6H₂O (40 mL). The mixture was homogenized and heated up to 60 °C. Then, 40 mL of 1 mM K₄Fe(CN)₆·3H₂O were added dropwise for 30 min at 60 °C under vigorous stirring. After the addition of this latter reagent, an intensive blue color appeared with a perfect dispersion of NPs. The resulting PB-NPs were naturally cooled to room temperature under mild stirring for approximately 1 h.



Figure S1. Preliminary studies in batch using 384-well plate comparing MOF(Fe), composite, and pristine PB-NPs. Experimental conditions: 5.0 μ L PB-NPs@MIL-101 dispersions (500 mg L⁻¹), 5.0 μ L of luminol 0.025 M (pH 12.0) and 10.0 μ L of 0.1 mM H₂O₂.



Figure S2. Preliminary chemiluminescence studies in paper-sensor comparing MOF(Fe), composite, and pristine PB-NPs. Experimental conditions: dried 10 μ L commercial luminol and 10 μ L of 2000 mg L⁻¹ PB-NPs@MIL-101(Fe) (or bare MIL-101(Fe) or PB-NPs) and 10 μ L of H₂O₂ 0.1 mM.



Figure S3. p-XRD spectra from 5 to 60 degree (2 Θ) for MIL-101(Cr, Al), bare PB-NPs, and their respective composites.



Figure S4. FT-IR spectra from 4000 to 400 cm⁻¹ for MIL-101(Cr, Al), bare PB-NPs, and their respective composites.



Figure S5. FT-IR spectra from 4000 to 400 cm⁻¹ for different sensors and bulk composite.



Figure S6. A) Mapping and B) EDX analysis of the PB-NPs@MIL-101(Fe) and C) EDX analysis of MIL-101(Fe).



Figure S7. SEM micrographs of A) MIL-101(Cr); B) PB-NPs@MIL-101(Cr); C) MIL-101(Al); D) PB-NPs@MIL-101(Al). Magnification and scale-bars are included in their respective images.



Figure S8. TEM micrographs of A) MIL-101(Cr); B) PB-NPs@MIL-101(Cr); C) MIL-101(Al); D) PB-NPs@MIL-101(Al). Scale-bars are included in their respective images.



Figure S9. N_2 (77 K) adsorption/desorption isotherms before and after PB-NPs impregnation. Black and red lines indicate the adsorption and desorption isotherms, respectively (A: Relative pressure (P/P₀); and B: Vol. Adsorbed (cm³ g⁻¹)



Figure S10. Different dispersions of 500 μ g of PB-NPs@MIL-101(Fe) using 1% (w/v) of PVP and SA in Tris-HCl 50 mM (pH 7.8) after their incubation without stirring for 2 h.



Figure S11. pH stability study. Stability studies at different pH were performed by analyzing H_2O_2 (0.1 mM) with the PB-NPs@MIL-101(Fe) sensing paper in 50 mM Tris-HCl by adjusting the pH to 5.0, 7.4, and 10.0, respectively



Figure S12. SEM micrographs of PB-NPs@MIL-101(Fe) at days A) 0 and B) 14. Magnification (1-2) and scale-bars are included in their respective images.

Material	Km H ₂ O ₂ (mol)	(Co)Substrate	Reaction volume	V _{max} (Mol s ⁻¹)	LDR (mol)	LOD _{H2O2} (mol)	Ref.
C0 ₃ O ₄	4.7.10-5	TMB/H ₂ O ₂	3000 µL	3.63.10-10	1.67.10-8-8.33.10-6	3.33.10-8	1
Fe ₃ O ₄	2.03.10-4	TMB/H ₂ O ₂	1316 µL	1.29.10-10	-	-	2
MWCNT	2.66.10-7	TMB/H ₂ O ₂	200 µL	2.22.10-11	$2 \cdot 10^{-10} - 3 \cdot 10^{-7}$	2.10-11	3
PB	1.54.10-6	TMB/H ₂ O ₂	200 µL	8.38.10-13	-	-	3
PB	7.82.10-6	TMB/H ₂ O ₂	1000 µL	2.4.10-11	-	-	4
HRP	4.87.10-6	TMB/H ₂ O ₂	1316 µL	1.15.10-10	-	-	2
Hemin@MO F	1.64.10-5	TMB/H ₂ O ₂	1500 μL	1.35.10-10	7.5.10-9 - 3.10-7	-	5
MIL-53(Fe)	4.2.10-8	TMB/H ₂ O ₂	1050 µL	1.95.10-11	9.98·10 ⁻¹⁰ – 2.00·10 ⁻⁸	1.37.10-10	6
MIL-101(Cr)	1.06.10-6	TMB/H ₂ O ₂	1000 µL	9.7.10-11	-	-	4
MIL-101(Fe)	5.80·10 ⁻⁹	TMB/H ₂ O ₂	100 µL	2.22.10-12	$2.40 \cdot 10^{-6} - 1 \cdot 10^{-8}$	1.5.10-11	7
MOF(Ce)	-	TMB/H ₂ O ₂	300 µL	-	1.2.10-6 - 4.8.10-6	3.10-9	8
Fe ₃ O ₄	-	ABTS/H ₂ O ₂	1000 µL	-	$5 \cdot 10^{-9} - 1 \cdot 10^{-7}$	3.00.10-9	9
MIL-101(Cr)	-	Luciferin/H ₂ O ₂	200 µL	-	$2 \cdot 10^{-6} - 2 \cdot 10^{-14}$	5.6.10-15	10
ABEI/CuFe ₂ O ₄	-	ABEI/H ₂ O ₂	100 µL	-	$1 \cdot 10^{-12} - 1 \cdot 10^{-8}$	2.5.10-13	11
POMs	-	Luminol/H ₂ O ₂	1600 µL	-	$2.67 \cdot 10^{-11} - 8 \cdot 10^{-9}$	8.10-12	12
MOF(Co) ^a	1.55.10-5	Luminol/H ₂ O ₂	4000 µL	1.29.10-5	$6 \cdot 10^{-12} - 5 \cdot 10^{-9}$	-	13
MIL-101(Cr)	-	Luminol/H ₂ O ₂	310 µL	-	$9.3 \cdot 10^{-10} - 3.1 \cdot 10^{-8}$	1.55.10-10	14
MIL-101(Fe)	3.82.10-9	Luminol/H ₂ O ₂	10 μL (45 μL)	3.65.10-5	$1 \cdot 10^{-11} - 5 \cdot 10^{-10}$	(8.2 μM) 8.2·10 ⁻¹¹	This work

Table S1. Comparison of the nanozyme sensing paper with other reported methods for H_2O_2 detection.

Abbreviations: MWCNT: Multi-Walled Carbon Nanotube ; TMB: 3,3',5,5'-Tetramethylbenzidine; ABTS: 2,2'-azino-bis(3-ethylbenzo-thiazoline-6-sulfonic acid) diammonium salt; ABEI: N-(4-aminobutyl)-N-ethylisoluminol; POM: olyoxometalates; LDR: linear dynamic range; LOD: limit of detection.

 $^a\mbox{The Km}$ and the V_{max} was calculated with TMB/H_2O_2 system

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