



OPEN COVID-19 clinical phenotypes in vaccinated and nonvaccinated solid organ transplant recipients: a multicenter validation study

Carmen Infante-Domínguez^{1,2,4,3}, Sonsoles Salto-Alejandro^{1,2,4,3}, Rocío Álvarez-Marín^{1,2}, Nuria Sabé^{2,3}, Antonio Ramos-Martínez⁴, Asunción Moreno⁵, Kamilla Ferreira de Moraes⁶, Zaira R. Palacios-Baena^{2,7}, Patricia Muñoz⁸, Mario Fernández-Ruiz^{2,9}, Marino Blanes¹⁰, Carmen Fariñas^{2,11}, Elisa Vidal^{2,12}, Esperanza Merino de Lucas^{13,14}, Mária Halpern¹⁵, Román Hernández-Gallego¹⁶, Matteo Bassetti¹⁷, Alessandra Mularoni¹⁸, Alex Gutiérrez-Dalmau¹⁹, Matteo Rinaldi^{20,21}, Silvia Jiménez-Jorge²², Marta Bodro⁵, Luis Fernando Aranha-Camargo⁶, Maricela Valerio⁸, Javier Sánchez-Céspedes^{1,2}, Belén Gutiérrez-Gutiérrez^{2,7,23}, Maddalena Giannella^{20,21}, Jesús Rodríguez-Baño^{2,7,23}, Jerónimo Pachón^{23,24}✉, Elisa Cordero^{1,2,23} & The COVIDSOT, ORCHESTRA Working Teams*

Clinical phenotypes of COVID-19, associated with mortality risk, have been identified in the general population. The present study assesses their applicability in solid organ transplant recipients (SOTR) hospital-admitted by COVID-19. In a cohort of 488 SOTR, nonvaccinated ($n = 394$) and vaccinated ($n = 94$) against SARS-CoV-2, we evaluated 16 demographic, clinical, analytical, and radiological variables to identify the clinical phenotypes A, B, and C. The median age was 61.0 (51–69) years, 330 (67.6%) and 158 (32.4%) were men and women, respectively, 415 (85%) had pneumonia, and 161 (33%) had $\text{SpO}_2 < 95\%$ at admission. All-cause mortality occurred in 105 (21.5%) cases. It was higher in nonvaccinated versus vaccinated SOTR (23.4% vs 13.8%, $P = 0.04$). Patients in the entire cohort were classified into phenotypes A ($n = 149$, 30.5%), B ($n = 187$, 38.3%), and C ($n = 152$, 31.1%), with mortality rates of 8.7%, 16.6%, and 40.1%, respectively, which were similar to those of nonvaccinated SOTR (9.5%, 16.7%, and 52.0%) and lower in vaccinated SOTR (4.4%, 15.8%, and 17.3%, respectively), with difference between nonvaccinated and vaccinated in the phenotype C ($P < 0.001$). In conclusion, COVID-19 clinical phenotypes are useful in SOTR, and all-cause mortality decreases in vaccinated patients.

Keywords Solid organ transplant recipients, COVID-19, Clinical phenotypes, Mortality, Multicenter cohort study

¹Clinical Unit of Infectious Diseases, Microbiology and Parasitology, Instituto de Biomedicina de Sevilla (IBiS), Virgen del Rocío University Hospital /CSIC/University of Seville, Seville, Spain. ²CIBERINFEC, ISCIII-CIBER de Enfermedades Infecciosas, Instituto de Salud Carlos III, Madrid, Spain. ³Service of Infectious Diseases, Bellvitge University Hospital, Bellvitge Biomedical Research Institute (IDIBELL), University of Barcelona, Hospitalet de Llobregat, Barcelona, Spain. ⁴Unit of Infectious Diseases, Microbiology, and Preventive Medicine. Hospital, Universitario Puerta de Hierro-Majadahonda-IDIPHISA, Madrid, Spain. ⁵Service of Infectious Diseases, Hospital Clinic-IDIBAPS. University of Barcelona, Barcelona, Spain. ⁶Hospital Israelita Albert Einstein, São Paulo, Brazil. ⁷Clinical Unit of Infectious Diseases and Microbiology, Virgen Macarena University Hospital, Instituto de Biomedicina de Sevilla (IBiS)/CSIC/University of Seville, Seville, Spain. ⁸Clinical Microbiology and Infectious Diseases Department, Hospital General Universitario Gregorio Marañón, Instituto de Investigación Sanitaria Gregorio Marañón (IiSGM), CIBERES, ISCIII-CIBER de Enfermedades Respiratorias. Medicine Department, School of Medicine, Universidad Complutense de Madrid, Madrid, Spain. ⁹Unit of Infectious Diseases, “12 de Octubre” University Hospital, Research Institute Hospital “12 de Octubre” (I+12), Madrid, Spain. ¹⁰Unit of Infectious Diseases, La Fe University Hospital, Valencia, Spain. ¹¹Service of Infectious Diseases, Marqués de Valdecilla University Hospital, Marqués de Valdecilla-IDIVAL, University of Cantabria, Santander, Spain. ¹²Service of Infectious Diseases, Reina Sofía University Hospital, Maimónides Biomedical Research Institute of Córdoba (IMIBIC), Córdoba, Spain. ¹³Unit of Infectious Diseases, Alicante General University Hospital, Alicante Institute of Health and Biomedical Research (ISABIAL), Alicante,

Spain.¹⁴Clinical Medicine Department, Miguel Hernández University, Elche, Spain.¹⁵Liver Transplantation Program, Quinta D'Or Hospital, Rio de Janeiro, Brazil.¹⁶Unit of Kidney Transplant, Service of Nephrology, Badajoz University Hospital, Badajoz, Spain.¹⁷Infectious Diseases Clinic, Policlinico San Martino Hospital-IRCCS, Department of Health Science, University of Genoa, Genoa, Italy.¹⁸Unit of Infectious Diseases and Infection Control, ISMETT-IRCCS-Istituto Mediterraneo per i Trapianti e Terapie ad Alta Specializzazione-Istituto di Ricovero e Cura a Carattere Scientifico, Palermo, Italy.¹⁹Renal Transplant Unit, Nephrology Service, Miguel Servet University Hospital, Aragón Institute for Health Research IIS-Aragón, Zaragoza, Spain.²⁰Department of Medical and Surgical Sciences, Alma Mater Studiorum University of Bologna, Bologna, Italy.²¹Infectious Diseases Unit, IRCCS Azienda Ospedaliero Universitaria di Bologna, Bologna, Italy.²²Clinical Research and Clinical Trials Unit (CTU), Instituto de Biomedicina de Sevilla (IBiS), Virgen del Rocío University Hospital/CSIC/University of Seville, Seville, Spain.²³Department of Medicine, School of Medicine, University of Seville, Seville, Spain.²⁴Instituto de Biomedicina de Sevilla (IBiS), Virgen del Rocío University Hospital/CSIC/University of Seville, Av. Manuel Siurot s/n, 41013 Seville, Spain.⁴³Carmen Infante-Domínguez and Sonsoles Salto-Alejandro contributed equally to this work. *A list of authors and their affiliations appears at the end of the paper. ✉email: pachon@us.es

The coronavirus disease (COVID-19) pandemic has led to significant morbidity and mortality, leading to the global collapse of the healthcare system. As of May 12, 2024, there were 775,481,326 confirmed cases and 7,049,376 deaths worldwide¹. Throughout this global health crisis, a broad spectrum of disease severity has been observed in affected individuals. Understanding the heterogeneity of clinical presentation and underlying pathogenic mechanisms is crucial for optimizing patient management and resource allocation. Extensive epidemiological, clinical, and therapeutic studies have contributed to our understanding of pathogens and diseases. This wealth of information includes insights into the risk factors for unfavorable outcomes in the general population, focusing primarily on demographics, comorbidities, symptom severity, and laboratory findings^{2–4}. In addition, factors related to immunology, virology^{5–8}, and treatment⁹ have been explored.

The global number of vaccinated individuals has been steadily increasing, with 13.5 billion vaccine doses administered as of November 26, 2023. In Western countries, particularly Andalusia and southern Spain, a remarkably high percentage of people have completed vaccination, reaching 100% in those over 60 years of age and 90% in the 40–60 age group¹⁰. Within the vaccinated population, COVID-19 predominantly affects vulnerable individuals, including those with either pluripathology or frailty, or both, and those with compromised immune responses. Furthermore, individuals receiving solid organ transplantation (SOT) or hematopoietic stem cell transplantation (HSCT) are at an elevated risk of developing severe forms of COVID-19, as seroconversion after immunization remains low^{11,12}. Therefore, understanding the factors influencing clinical outcomes, especially in patients who have received a transplant, is crucial for optimizing care. Age, underlying comorbidities, immunosuppressive regimens, and time since transplantation have been identified as risk factors associated with unfavorable outcomes, as analyzed in a prospective multicenter study involving patients who have received a solid organ transplant⁴ conducted before widespread vaccination. Moreover, pre-transplant prognostic profiles have been identified in patients with hematologic profiles¹³.

The clinical phenotypes of COVID-19 were initially defined in adult patients on hospital admission at the onset of the pandemic¹⁴. In a multicenter cohort study, three clinical phenotypes with different mortality rates were identified and validated in patients admitted to hospitals with COVID-19. To identify the phenotypes, this study developed a multinomial logistic regression model with 16 variables including age, sex, chronic lung disease, obesity, diastolic blood pressure, oxygen saturation, white blood cell and neutrophils counts, hematocrit, coagulation international normalized ratio (INR), C-reactive protein (CRP), glucose, creatinine, sodium, potassium, and type of lung infiltrate on chest radiograph. The 30-day mortality rates for phenotypes A, B, and C were 2.6%, 31.0%, and 53.4%, respectively¹⁴. This study provides valuable insights into the prognoses associated with these phenotypes and reveals potential disease severity and mortality risk indicators. It also guides personalized treatment strategies to enhance patient outcomes. However, it is essential to note that the utility of these clinical phenotypes has not yet been validated for SOT or HSCT recipients.

Data analysis from kidney transplant recipients has revealed distinct respiratory and gastrointestinal phenotypes associated with COVID-19¹⁵. Certain patients primarily display respiratory symptoms similar to the typical manifestations of COVID-19 observed in the general population. In contrast, the other patients manifested prominent gastrointestinal symptoms, a more prevalent pattern in the immunocompromised cohort. Disease severity differs based on these phenotypes, generally with patients exhibiting gastrointestinal symptoms facing a higher risk of severe outcomes.

The present study aimed to assess and validate the applicability of the clinical phenotypes identified in the general population at the onset of the pandemic¹⁴ in SOT recipients (SOTR) admitted to the hospital with COVID-19. This evaluation was conducted in two distinct cohorts of SOTR: nonvaccinated and vaccinated against SARS-CoV-2. This study aimed to identify the most accurate predictors and current clinical phenotypes within this population, serving as a valuable tool for optimal management of outpatients and inpatients.

Methods

We conducted an international retrospective and observational cohort study across 18 participating centers in Spain, Italy, and Brazil (COVIDSOT Study, clinicaltrials.gov ID: NCT04319172, first posted on March 24, 2020; EU PAS Register Number: EUPAS34349, first posted on March 31, 2020). These centers were asked to include all consecutive adult SOT recipients (SOTR) who were hospitalized for confirmed COVID-19, by a positive SARS-CoV-2 reverse transcription-PCR (RT-PCR) test or rapid antigen detection test (RADT) targeting the SARS-CoV-2 nucleocapsid protein from nasopharyngeal (NP) swabs, between January 2020 and October 2022 (nonvaccinated patients from January 2020 to March 2022, and vaccinated patients from May 2021 to October

2022), and with the availability of data in the 16 variables required to assign patients with a specific COVID-19 phenotype¹⁴.

Data collection

Anonymized data were collected through an electronic Case Report Form and entered into a database constructed explicitly for this study using the Research Electronic Data Capture (REDCap) tools¹⁶. The recorded variables included demographics, comorbidities, transplant type, and date, signs and symptoms upon admission, baseline laboratory tests, chest radiography findings, complications during hospitalization, immunosuppression management, therapeutics with purported activity against COVID-19, adjunctive strategies to modulate the host inflammatory response, and clinical outcomes. Vaccinated SOTR against SARS-CoV-2 were considered if they had received at least two vaccine doses. The clinical severity at hospital admission (basal WHO scale) and at the end of follow-up (final WHO scale) was evaluated using the WHO clinical progression scale¹⁷.

Event of interest

The clinical outcomes of the patients after a 30-day follow-up were categorized as survival or death. In cases where patients were discharged and subsequently readmitted to the hospital during the study period, only the initial hospital admission episode was considered for the analysis.

The primary stratification strategy was based on phenotypes A, B, and C, utilizing 16 demographic, clinical, analytical, and radiological variables (age, sex, chronic lung disease, obesity, diastolic blood pressure, oxygen saturation, white blood cell and neutrophils counts, hematocrit, coagulation INR, CRP, glucose, creatinine, sodium, potassium, and type of lung infiltrate on chest radiograph) of the FEN-COVID probability calculator to assign patients with a specific COVID-19 phenotype (<http://fen-covid.com/index.html>)¹⁴. The secondary stratification strategy was vaccination status (yes or no).

Statistical analysis

Two analyses were conducted. First, a descriptive analysis encompassing all patients and separate assessments of nonvaccinated and vaccinated individuals was performed using the probabilistic phenotype method. Second, the bivariate analysis focused on describing and examining the characteristics associated with 30-day mortality. Categorical variables are presented as *n* (%), and continuous variables as mean (standard deviation [SD]) or median (interquartile range [IQR]) based on the normality of the distribution. Appropriate statistical tests, such as the χ^2 -test, the Yates Correction for Continuity, Student's *t* test, or Mann–Whitney U test, were employed to assess between-group differences. Interactions, confusion, and collinearity were thoroughly explored. Variables associated with the outcome and those considered clinically relevant were incorporated into a multivariate regression analysis to assess the real impact of every exposure factor in the 30-day mortality. Survival analysis was performed using the Kaplan–Meier method. All analyses were performed using the software package SPSS (26.0. Armonk, NY: IBM Corp.). The *p*-values were derived from two-tailed tests, and those <0.05 were considered significant.

Ethics approval

The Ethics Committee of Virgen del Rocío and Virgen Macarena University Hospitals approved the study protocol (C.I. 0842-N-20), as well as by the appropriate institutional review board of each participating center (Supplementary Information), and adhered to the Helsinki Declaration. An informed consent was established as a mandatory requirement for all patients.

Results

Clinical characteristics and outcomes of all SOTR, and according to vaccination

Four hundred and eighty-eight SOTR with COVID-19, confirmed by RT-PCR and RADT in 467 and 21 cases, respectively, were included in the study from January 2020 to October 2022, with a median age of 61.0 (51–69) years, 330 (67.6%) were men and 158 (32.4%) were women (Table 1). The most frequently transplanted organs were the kidneys (*n* = 309, 63.3%) and the liver (*n* = 84, 17.2%). At the time of COVID-19 diagnosis, 186 (38.1%) patients experienced dyspnea, 415 (85.0%) had pneumonia, and 161 (33.0%) had peripheral oxygen saturation (SpO₂) <95%. The median number of days from symptom onset to diagnosis was 4 (2–9).

Regarding the analytical parameters, 47 (9.6%) patients had white blood cell counts >11,000/ μ L, 285 (58.3%) had lymphocytes <1000/ μ L, 72 (14.8%) had C-reactive protein (CRP) values \geq 100 mg/L, and 212 (43.4%) had D-dimer levels \geq 600 ng/mL. Only 88 (18%) and 80 (16.4%) patients received tocilizumab and dexamethasone, respectively. The clinical outcome, as assessed using the WHO clinical progression scale, was severe (final WHO scale 7–10) in 131 (26.8%) patients, and all-cause mortality (final WHO scale 10) occurred in 105 (21.5%). Table 1 summarizes the clinical characteristics and outcomes of all patients, including the differences between the nonvaccinated and vaccinated populations.

The nonvaccinated group (*n* = 394, 80.7%) compared to the vaccinated group (*n* = 94, 19.3%) of SOTR were younger (60 [50–68] vs. 64 [57–71], *p* < 0.05) and more frequently exhibited a temperature >37.5 °C (25.4% vs. 10.6%, *P* = 0.002), systolic pressure <90 mmHg (9.1% vs. 0%, *p* = 0.04), and diastolic pressure <60 mmHg (8.1% vs. 2.1%, *p* = 0.03), with no differences in the frequency of SpO₂ <95% (Table 1). Creatinine >1.3 mg/dL (83.0% vs. 69.5%, *p* = 0.013) and C-reactive protein \geq 100 mg/L (30.9% vs. 10.9%, *p* < 0.05) were more common in vaccinated SOTR. Vaccinated patients received dexamethasone upon hospital admission more frequently than nonvaccinated patients (61.7% vs. 5.6%, *p* < 0.001). Severe clinical outcomes (WHO scale 7–10) and all-cause mortality were higher in nonvaccinated patients than in vaccinated patients (29.7% vs. 14.9%, *p* = 0.003 and 23.4% vs. 13.8%, *p* = 0.04, respectively) (Table 1).

Variables (n, %)	All SOTR N = 488	Nonvaccinated N = 394	Vaccinated N = 94	p value
Age years (median, IQR)	61 (51–69)	60 (50–68)	64 (57–71)	0.009
Age ≥ 70 years	114 (23.4)	86 (21.8)	28 (29.8)	0.101
Male	330 (67.6)	266 (67.5)	64 (68.1)	0.915
Organ transplant				
Kidney	309 (63.3)	243 (61.7)	66 (70.2)	0.123
Liver	84 (17.2)	65 (16.5)	19 (20.2)	0.099
Heart	47 (9.6)	44 (11.2)	3 (3.2)	0.019
Lung	35 (7.2)	32 (8.1)	3 (3.2)	0.096
Pancreas	2 (0.4)	2 (0.5)	0 (0)	0.488
Combined transplant	11 (2.3)	8 (2.0)	3 (3.2)	0.496
Baseline immunosuppression				
Mycophenolate mofetil	339 (69.5)	283 (71.8)	56 (59.6)	0.410
Azathioprine	10 (2.0)	8 (2)	2 (2.1)	0.730
Cyclosporine	31 (6.4)	28 (7.1)	3 (3.2)	0.245
Tacrolimus	370 (75.8)	311 (78.9)	59 (62.8)	0.002
Sirolimus/everolimus	75 (15.4)	67 (17)	8 (8.5)	0.058
Prednisone	358 (73.4)	300 (76.1)	58 (61.7)	0.006
Comorbidities				
Diabetes mellitus	181 (37.1)	137 (34.8)	44 (46.8)	0.030
Chronic lung disease	84 (17.2)	69 (17.5)	15 (16.0)	0.751
Chronic heart disease	88 (18.0)	60 (15.2)	28 (29.8)	0.001
Chronic kidney disease	216 (44.3)	161 (40.9)	55 (58.5)	0.002
Chronic liver disease	61 (12.5)	45 (11.4)	16 (17.0)	0.140
Cancer	38 (7.8)	33 (8.4)	5 (5.3)	0.320
Morbid obesity	36 (7.4)	30 (7.6)	6 (6.4)	0.682
Symptoms and signs at diagnosis				
Pneumonia	415 (85.0)	338 (85.8)	77 (81.9)	0.344
Temperature > 37.5 °C	110 (22.5)	100 (25.4)	10 (10.6)	0.002
Dyspnea	186 (38.1)	143 (36.3)	43 (45.7)	0.170
SpO ₂ < 95%	161 (33.0)	131 (32.2)	30 (31.9)	0.805
Systolic blood pressure < 90 mmHg	16 (3.3)	16 (9.1)	0 (0)	0.040
Diastolic blood pressure < 60 mmHg	34 (7)	32 (8.1)	2 (2.1)	0.030
Impaired consciousness	21 (4.3)	18 (4.6)	3 (3.2)	0.486
Days from symptoms onset, median (IQR) (n = 423)	4 (2–9)	4 (2–8)	5 (1–12)	0.231
Time from transplant < 6 months	47 (9.6)	45 (11.4)	2 (2.1)	0.006
Analytical data				
White blood cells × 1000/μL (median, IQR)	5.8 (4.3–7.9)	5.8 (4.4–7.8)	6.0 (4.4–8.3)	0.483
White blood cells > 11,000/μL	47 (9.6)	36 (9.1)	11 (11.7)	0.449
Neutrophils × 1000/μL (median, IQR)	4.3 (3.1–6.3)	4.2 (3.2–6.2)	4.4 (3.0–6.5)	0.685
Neutrophils > 7500/μL	75 (15.3)	56 (14.2)	19 (20.2)	0.147
Lymphocytes × 1000/μL (median, IQR)	0.8 (0.5–1.1)	0.8 (0.5–1.1)	0.8 (0.5–1.2)	0.995
Lymphocytes < 1000/μL	285 (58.3)	226 (57.4)	59 (62.8)	0.539
Platelets × 1000/μL (median, IQR)	156 (122–218)	154 (120–218)	175 (125.5–219.8)	0.311
Platelets < 130,000/μL	131 (26.8)	106 (26.9)	25 (26.6)	0.743
Hematocrit (%)	38.0 (34.2–42.0)	38.6 (34.0–42.0)	37.2 (35.0–42.0)	0.542
Hematocrit < 36% (females) and < 41% (males)	262 (53.7)	209 (53.0)	53 (56.4)	0.560
Glucose (mg/dL)	116 (96–163)	115 (94–162)	132 (101–174)	0.041
Glucose > 125 mg/dL	206 (42.2)	156 (39.6)	50 (53.2)	0.016
Creatinine mg/dL (median, IQR)	1.7 (1.2–2.4)	1.6 (1.2–2.3)	1.92 (1.4–2.8)	0.017
Creatinine > 1.3 mg/dL	352 (71.8)	274 (69.5)	78 (83.0)	0.013
Na (mEq/L)	136 (133–139)	137.0 (134.0–139.0)	134 (130.8–137.0)	< 0.001
Na < 135 mEq/L	174 (35.7)	123 (31.2)	51 (54.3)	< 0.001
K (mEq/L)	4.4 (4.0–4.8)	4.4 (3.9–4.7)	4.5 (4.1–4.9)	0.047
K > 5.2 mEq/L	433 (88.7)	351 (89.0)	82 (87.2)	0.610
Continued				

Variables (n, %)	All SOTR N = 488	Nonvaccinated N = 394	Vaccinated N = 94	p value
Aspartate aminotransferase (AST) U/L (median, IQR)	29 (20–43)	30 (20–45)	27 (20–35)	0.062
AST > 30 U/L	159 (31.7)	136 (34.5)	23 (24.5)	0.627
Alanine aminotransferase (ALT) U/L (median, IQR)	21 (14–34)	23 (15–36)	12 (11–22)	<0.001
ALT > 40 U/L	78 (15.5)	68 (17.3)	10 (10.6)	0.156
Lactate dehydrogenase (LDH) U/L (median, IQR)	275 (224–349)	270 (221–352)	298 (233–336)	0.276
LDH > 300 U/L	163 (32.1)	125 (31.7)	38 (40.4)	0.137
C-reactive protein mg/L (median, IQR)	14.6 (4.8–63.7)	10.9 (3.6–38.1)	66.8 (37.1–117.7)	<0.001
C-reactive protein > 100 mg/L	72 (14.8)	43 (10.9)	29 (30.9)	<0.001
D-dimer ng/mL (median, IQR)	660 (368–1505)	614 (341–1390)	1020 (540–1710)	0.015
D-dimer > 600 ng/mL	212 (43.4)	163 (41.4)	49 (52.1)	0.059
INR (median, IQR)	1.08 (1.0–1.17)	1.08 (1.0–1.17)	1.1 (1.01–1.10)	0.369
INR > 1.86	38 (7.8)	28 (7.1)	10 (10.6)	0.251
Therapy				
Tocilizumab	88 (18)	70 (17.8)	18 (19.1)	0.870
Dexamethasone	80 (16.4)	22 (5.6)	58 (61.7)	<0.001
High flow/non-invasive mechanical ventilation	81 (16.6)	63 (16)	18 (19.1)	0.459
Mechanical ventilation	80 (16.4)	73 (18.5)	7 (7.4)	0.008
Complications during hospital stay				
Shock	4 (0.8)	3 (0.8)	1 (1.1)	0.807
Graft dysfunction	6 (1.2)	5 (1.3)	1 (1.1)	0.050
Graft lost	4 (0.8)	4 (1.0)	0 (0)	0.037
Basal WHO clinical progression scale				
Basal 2–3	15 (3.1)	11 (2.8)	4 (4.3)	0.460
Basal 4–5	447 (91.6)	367 (93.1)	80 (85.1)	0.020
Basal 6	18 (3.7)	13 (3.3)	5 (5.3)	0.351
Basal 7	8 (1.6)	3 (0.8)	5 (5.3)	0.007
Final WHO clinical progression scale				
Final 2–3	4 (0.8)	4 (1)	0 (0)	0.327
Final 4–5	340 (69.7)	260 (66.0)	80 (85.1)	<0.001
Final 6	15 (3.1)	13 (3.3)	2 (2.1)	0.554
Final 7	26 (5.3)	25 (6.3)	1 (1.1)	0.041
Final 10	105 (21.5)	92 (23.4)	13 (13.8)	0.044
Clinical phenotypes				
Phenotype A	149 (30.5)	126 (32)	23 (24.5)	0.155
Phenotype B	187 (38.3)	168 (42.6)	19 (20.2)	<0.001
Phenotype C	152 (31.1)	100 (25.4)	52 (55.3)	<0.001
Clinical phenotypes mortality				
Phenotype A mortality	13 (8.7)	12 (9.5)	1 (4.4)	0.370
Phenotype B mortality	31 (16.6)	28 (16.7)	3 (15.8)	0.611
Phenotype C mortality	61 (40.1)	52 (52.0)	9 (17.3)	<0.001

Table 1. Demographics, type of transplant, immunosuppressive therapy, clinical characteristics at hospital admission, and outcomes in all solid organ transplant recipients (SOTR) and in patients nonvaccinated and vaccinated.

Clinical phenotypes, distribution, and mortality in all SOTR and according to vaccination status

In the entire cohort of 488 SOTR, the clinical phenotypes at admission were A in 149 (30.5%) patients, B in 187 (38.3%) patients, and C in 152 (31.1%) patients, with increasing mortality rates of 8.7% ($n=13$), 16.6% ($n=31$), and 40.1% ($n=61$), respectively ($p<0.001$). The association between higher mortality rates and the phenotypes remained in the 394 nonvaccinated SOTR ($p<0.001$) from A to C: 9.5% ($n=12$), 16.7% ($n=28$), and 52% ($n=52$), respectively. However, in vaccinated SOTR, the mortality rate, although lower in phenotype A (4.4% [$n=1$]), did not differ between the three phenotypes ($p=0.11$) or between phenotypes B (15.8% [$n=3$]) and C (17.3% [$n=9$]) ($p=0.59$). Finally, mortality rates were comparable for phenotypes A and B in vaccinated and nonvaccinated patients when assessing the SOTR phenotypes according to vaccination status. However, it differed between the nonvaccinated and vaccinated SOTR belonging to phenotype C (52% vs. 17.3%, $p<0.001$) (Supplementary Fig. S1). In the Kaplan-Meier analysis, survival at day 30 was lower in SOTR with phenotype

B or C than in those with phenotype A ($p < 0.01$), regardless of vaccination. Overall survival was also lower in nonvaccinated SOTR with phenotypes B or C compared to phenotype A ($p < 0.01$), but we found no differences in survival at day 30 for vaccinated patients, irrespective of the phenotype they belonged to ($p = 0.311$) (Fig. 1).

Factors independently associated with mortality in all SOTR and according to vaccination

In the subsequent analysis, we conducted a bivariate examination of the factors associated with all-cause mortality at hospital admission on day 30 after COVID-19 diagnosis (Table 2). Deceased patients were more likely to be aged ≥ 70 years (36.2% vs. 19.8%, $p < 0.001$), have diabetes mellitus (48.6% vs. 33.9%, $p = 0.006$), and have not received the COVID-19 vaccination (87.6% vs. 78.9%, $p = 0.04$). Regarding clinical characteristics, deceased patients exhibited a higher frequency of pneumonia (92.4% vs. 83.0, $p = 0.017$), $SpO_2 < 95\%$ (48.6% vs. 28.7%, $p < 0.001$), and elevated inflammatory parameter values (neutrophils $> 7500/\mu L$ [$p = 0.016$], CRP > 100 mg/dL [$p = 0.008$], lactate dehydrogenase [LDH] > 300 U/L [$p < 0.001$], and higher median D-dimer [$p = 0.025$]).

Multivariate logistic regression analysis was performed to assess potential independent predictors of death. Age ≥ 70 years (odds ratio [OR] 2.5, $p < 0.001$), $SpO_2 < 95\%$ (OR 2.35, $p < 0.001$), creatinine > 1.3 mg/dL (OR 2.35, $p = 0.004$), and not having received COVID-19 vaccination (OR 2.21, $p = 0.018$) were identified as independent risk factors associated with all-cause mortality at day 30 (Table 3). A receiver operating characteristic (ROC) curve plot was generated for the model, with an area under the curve of 0.71 (95% CI 0.650–0.767), with a standard error of 0.03 (under the non-parametric assumption), and $p < 0.001$ (being the null hypothesis a true area = 0.50) (Fig. 2).

Concerning the nonvaccinated SOTR, all-cause mortality at day 30 occurred in 92 patients (23.4%) and was more frequent in those with age ≥ 70 years ($p < 0.001$), diabetes mellitus ($p = 0.006$), pneumonia ($p = 0.038$), dyspnea ($p < 0.001$), and $SpO_2 < 95\%$ ($p < 0.001$) at diagnosis (Supplementary Table S1). In the multivariable logistic regression analysis, age ≥ 70 years (OR 3.32, $p < 0.001$), diabetes mellitus (OR 2.01, $p = 0.037$), $SpO_2 < 95\%$ (OR 1.90, $p = 0.045$), creatinine > 1.3 mg/dL (OR 2.12, $p = 0.041$), and LDH > 300 U/L (OR 3.28, $p < 0.001$) were independently associated with all-cause mortality at day 30 (Supplementary Table S2).

Finally, in vaccinated SOTR all-cause mortality at day 30 occurred in 13 patients (13.8%) and was more frequent in those with combined transplantation (15.4% vs. 1.2%, $p = 0.007$), chronic heart disease (53.8% vs. 25.9%, $p = 0.041$), and dyspnea at hospital admission (84.6% vs. 39.5%, $p = 0.001$) (Supplementary Table S1). In the multivariable logistic regression analysis, dyspnea was the only factor independently associated with mortality (OR 15.7, $p = 0.011$) (Supplementary Table S2).

Discussion

The study findings revealed a higher all-cause mortality rate for COVID-19 in SOTR among nonvaccinated individuals than in their vaccinated counterparts. Additionally, the clinical phenotypes¹⁴ initially identified in the general population during the early stages of the COVID-19 pandemic apply to the SOTR cohort. This trend was particularly notable among nonvaccinated patients, with all-cause mortality rates progressively increasing from phenotypes A to C. In the vaccinated SOTR, phenotype A was associated with a low mortality rate, which also increased in phenotypes B and C, although with less difference between the two, owing to a three-fold decrease in patients with phenotype C in the vaccinated SOTR compared with their nonvaccinated counterparts. In a secondary analysis, independent risk factors associated with all-cause mortality at day 30 included age ≥ 70

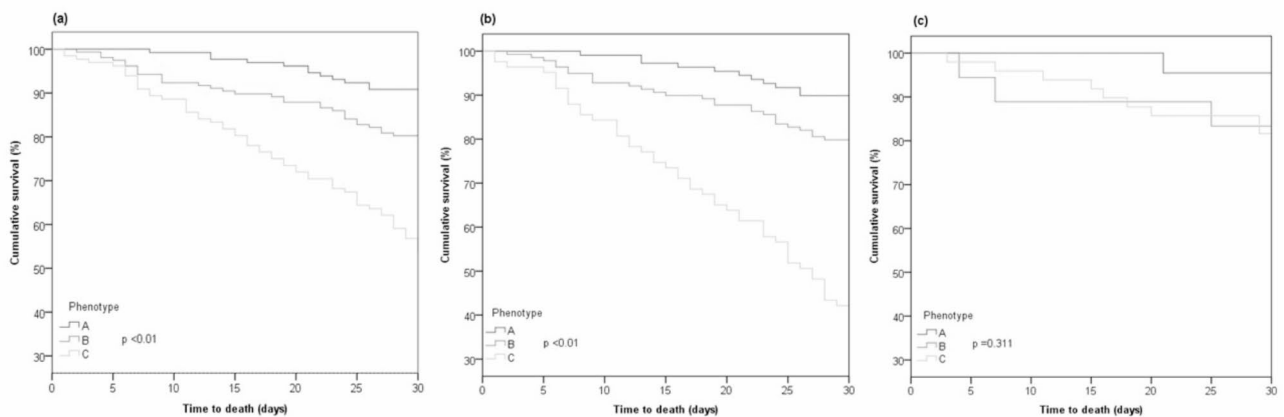


Fig. 1. Overall survival at day + 30 after COVID-19 diagnosis in patients with phenotype A, B y C in the all-solid organ transplant recipients (SOTR) cohort (a) $n = 488$, in nonvaccinated SOTR (b) $n = 394$, and in vaccinated SOTR (c) $n = 94$.

Variables (n, %)	All SOTR N = 488	Dead N = 105	Survivors N = 383	p value
Age in years (median, IQR)	61 (51–69)	64 (57–72)	59 (50–68)	<0.001
Age ≥ 70 years	114 (23.4)	38 (36.2)	76 (19.8)	<0.001
Male	330 (67.6)	71 (67.6)	259 (67.6)	0.99
Non-vaccinated	394 (80.7)	92 (87.6)	302 (78.9)	0.04
Organ transplant				
Kidney	309 (63.3)	70 (66.7)	239 (62.4)	0.422
Liver	84 (17.2)	13 (12.4)	71 (18.6)	0.311
Heart	47 (9.6)	10 (9.5)	37 (9.7)	0.966
Lung	35 (7.2)	9 (8.6)	26 (6.8)	0.530
Pancreas	2 (0.4)	0 (0)	2 (0.5)	0.458
Combined transplant	11 (2.3)	3 (2.9)	8 (2.1)	0.638
Baseline immunosuppression				
Mycophenolate mofetil	339 (69.5)	77 (73.3)	262 (68.4)	0.485
Azathioprine	10 (2.0)	1 (1)	9 (2.3)	0.586
Cyclosporine	31 (6.4)	8 (7.6)	23 (6.0)	0.718
Tacrolimus	370 (75.8)	77 (73.3)	293 (76.5)	0.824
Sirolimus/everolimus	75 (15.4)	12 (11.4)	63 (16.4)	0.378
Prednisone	358 (73.4)	79 (75.2)	279 (72.8)	0.681
Comorbidities				
Diabetes mellitus	181 (37.1)	51 (48.6)	130 (33.9)	0.006
Chronic lung disease	84 (17.2)	16 (15.2)	68 (17.8)	0.538
Chronic heart disease	88 (18.0)	22 (21.0)	66 (17.2)	0.386
Chronic kidney disease	216 (44.3)	51 (48.6)	165 (43.1)	0.316
Chronic liver disease	61 (12.5)	11 (10.5)	50 (13.1)	0.479
Cancer	38 (7.8)	11 (10.5)	27 (7.0)	0.246
Morbid obesity	36 (7.4)	8 (7.6)	28 (7.3)	0.915
Symptoms and signs at diagnosis (%)				
Pneumonia	415 (85)	97 (92.4)	318 (83.0)	0.017
Temperature > 37.5 °C	110 (22.5)	27 (25.7)	83 (21.7)	0.380
Dyspnea	186 (38.1)	63 (60.0)	123 (32.1)	<0.001
SpO ₂ < 95%	161 (33)	51 (48.6)	110 (28.7)	<0.001
Systolic blood pressure < 90 mmHg	16 (3.3)	3 (2.9)	13 (3.4)	0.874
Diastolic blood pressure < 60 mmHg	34 (7)	10 (9.5)	24 (6.3)	0.178
Impaired consciousness	21 (4.3)	10 (9.5)	11 (2.9)	<0.001
Days from symptoms onset, median (IQR)	4 (2–9)	4 (0.25–6)	5 (2–10)	0.010
Time from transplant < 6 months	47 (9.6)	15 (14.3)	32 (8.4)	0.068
Analytical data				
White blood cells × 1000/μL (median, IQR)	5.8 (4.3–7.9)	6 (4.4–9.0)	5.9 (4.3–7.6)	0.272
White blood cells > 11,000/μL	47 (9.6)	14 (13.3)	33 (8.6)	0.147
Neutrophils × 1000/μL (median, IQR)	4.3 (3.1–6.3)	4.8 (3.5–7.2)	4.2 (3.1–6.1)	0.065
Neutrophils > 7500/μL	75 (15.3)	24 (22.9)	51 (13.3)	0.016
Lymphocytes × 1000/μL (median, IQR)	0.8 (0.5–1.1)	0.7 (0.5–1.1)	0.8 (0.5–1.2)	0.110
Lymphocytes < 1000/μL	285 (58.3)	65 (61.9)	220 (57.4)	0.105
Platelets × 1000/μL (n = 445) (median, IQR)	156 (122–218)	155 (130–204)	156 (121–221.5)	0.850
Platelets < 130,000/μL	131 (26.8)	22 (21.0)	109 (28.5)	0.210
Hematocrit (%)	38.0 (34.2–42.0)	37.2 (32.0–41.8)	38.4 (34.8–42)	0.079
Hematocrit < 36% (females) or < 41% (males)	262 (53.7)	65 (61.9)	197 (51.4)	0.057
Glucose (mg/dL)	116 (96–163)	126 (105.0–167.8)	114 (94–162)	0.028
Glucose > 125 mg/dL	206 (42.2)	53 (50.4)	153 (39.9)	0.053
Creatinine mg/dL (median, IQR)	1.7 (1.2–2.4)	2.1 (1.4–3.3)	1.6 (1.2–2.3)	<0.001
Creatinine > 1.3 mg/dL	352 (71.8)	88 (83.8)	264 (68.9)	0.003
Na (mEq/L)	136 (133–139)	137 (133–139)	136 (133–139)	0.359
Na < 135 mEq/L	174 (35.7)	38 (36.19)	136 (35.5)	0.897
K (mEq/L)	4.4 (4.0–4.8)	4.6 (4.1–5.1)	4.3 (4.0–4.7)	0.005
K > 5.2 mEq/L	433 (88.7)	86 (81.9)	347 (90.6)	0.013
Continued				

Variables (n, %)	All SOTR N = 488	Dead N = 105	Survivors N = 383	p value
Aspartate aminotransferase (AST) U/L (median, IQR)	29 (20–43)	30 (20–48)	29 (20–43)	0.881
AST > 30 U/L	159 (31.7)	28 (26.7)	131 (34.2)	0.622
Alanine aminotransferase (ALT) U/L (median, IQR)	21 (14–34)	19 (13–28)	22 (14–35)	0.209
ALT > 40 U/L	78 (15.5)	12 (11.4)	66 (17.2)	0.192
Lactate dehydrogenase (LDH) U/L (median, IQR)	275 (224–349)	328 (263–412)	265 (215–333)	<0.001
LDH > 300 U/L	163 (32.1)	52 (49.5)	111 (29.0)	<0.001
C-reactive protein mg/L (median, IQR)	14.6 (4.8–63.7)	21.6 (7.5–99.1)	13 (37.13–117.68)	0.001
C-reactive protein > 100 mg/L	72 (14.8)	24 (22.9)	48 (12.5)	0.008
D-dimer ng/mL (median, IQR)	660 (368–1505)	767 (508,3–1785)	648 (340–1330)	0.025
D-dimer > 600 ng/mL	212 (43.4)	47 (44.8)	165 (43.1)	0.758
INR (median, IQR)	1.08 (1.0–1.08)	1.1 (1.03–1.19)	1.08 (1.0–1.16)	0.369
INR > 1.86	38 (7.8)	10 (9.5)	28 (7.3)	0.453
Therapy				
Tocilizumab	88 (18)	33 (31.4)	55 (14.4)	<0.001
Dexamethasone	80 (16.4)	15 (14.3)	65 (17.0)	0.984
High flow/non-invasive mechanical ventilation	81 (16.6)	47 (44.8)	34 (8.9)	<0.001
Mechanical ventilation	80 (16.4)	58 (55.2)	22 (5.7)	<0.001
Complications during hospital stay				
Shock	4 (0.8)	1 (1.0)	3 (0.8)	0.826
Graft dysfunction	6 (1.2)	2 (1.9)	4 (1.0)	0.553
Graft lost	4 (0.8)	1 (1.0)	3 (0.8)	0.839
Basal WHO clinical progression scale				
Basal 2–3	15 (3.1)	1 (1.0)	14 (3.7)	0.155
Basal 4–5	447 (91.6)	88 (83.8)	359 (93.7)	0.001
Basal 6	18 (3.7)	12 (11.4)	6 (1.6)	<0.001
Basal 7	8 (1.6)	4 (3.8)	4 (1.0)	0.048
Final WHO clinical progression scale				
Final 2–3	4 (0.8)	0 (0)	4 (1)	0.293
Final 4–5	340 (69.7)	2 (1.9)	338 (88.3)	<0.001
Final 6	15 (3.1)	0 (0)	15 (3.9)	0.039
Final 7	26 (5.3)	0 (0)	26 (6.8)	0.006
Final 10	105 (21.5)	105 (100)	0 (0)	<0.001
Clinical phenotypes				
Phenotype A	149 (30.5)	13 (12.4)	136 (35.5)	<0.001
Phenotype B	187 (38.3)	31 (29.5)	156 (40.7)	0.036
Phenotype C	152 (31.1)	61 (58.1)	91 (23.8)	<0.001

Table 2. Demographics, type of transplant, immunosuppressive therapy, clinical characteristics at hospital admission, in all solid organ transplant recipients (SOTR) and according to all-cause mortality at day + 30 after COVID-19 diagnosis.

All-cause mortality at day + 30	OR (95% CI)	p value
Variables		
Age ≥ 70 years	2.5 (1.54–4.19)	<0.001
SpO ₂ < 95%	2.35 (1.48–3.74)	<0.001
Creatinine > 1.3 mg/dL	2.35 (1.31–4.22)	0.004
Neutrophils > 7500/μL	1.78 (0.99–3.19)	0.051
Non-vaccination against COVID-19	2.21 (1.15–4.29)	0.018
Time from transplant < 6 months	1.77 (0.87–3.6)	0.113

Table 3. Multivariate logistic regression analyses of risk factors associated with all-cause 30-day mortality in the whole cohort of 488 solid organ transplant recipients.

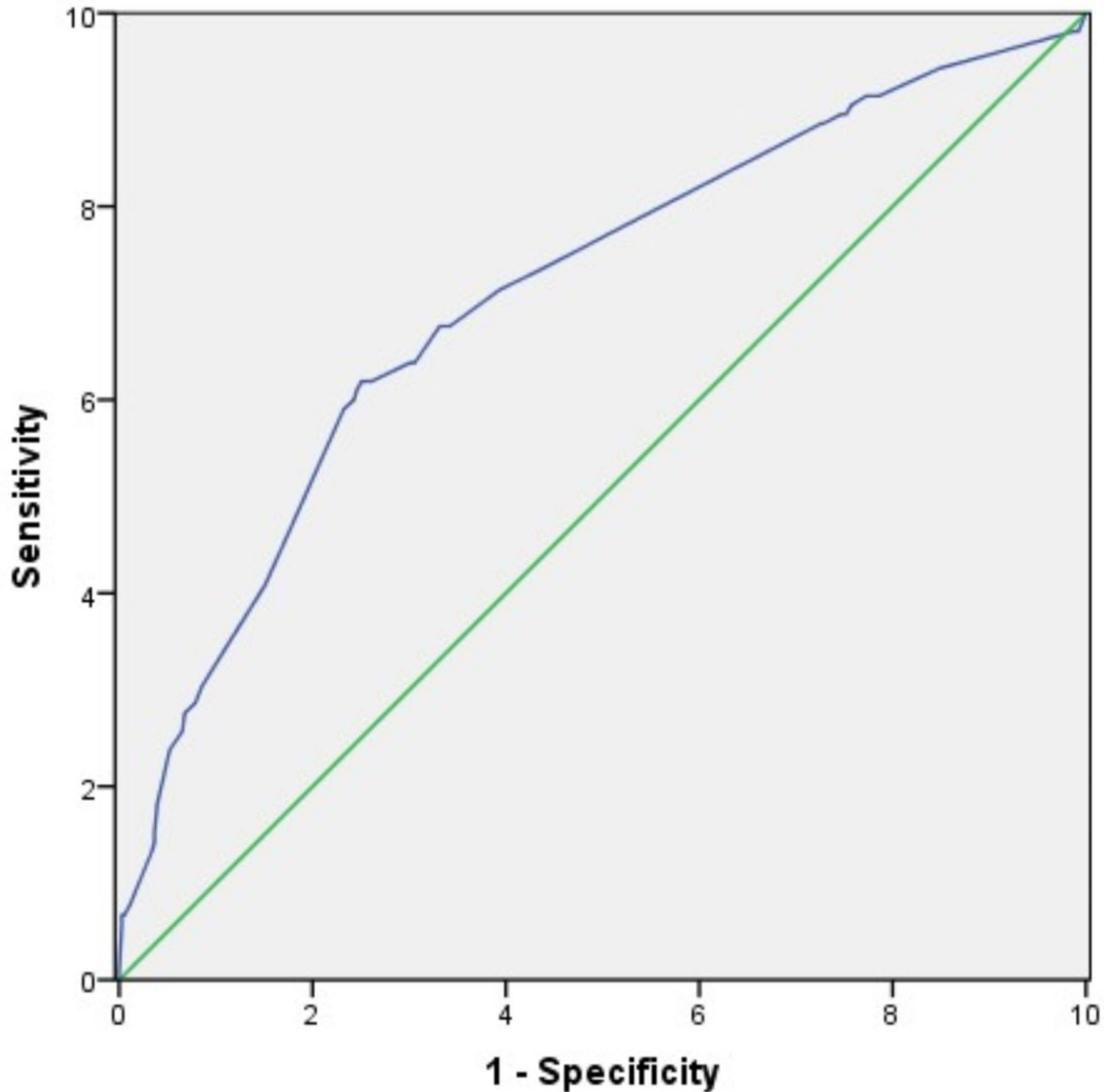


Fig. 2. Discrimination power of the final multivariable model: receiving operative curve (ROC) plot (including age ≥ 70 years, SpO₂ $< 95\%$, neutrophils $> 7500/\mu\text{L}$, non-vaccination, and time from transplant onset to COVID-19 diagnosis < 6 months), expressed by an area under the ROC of 0.71 (95% CI 0.65–0.767), SE = 0.03 (under the non-parametric assumption), and $p < 0.001$ (being the null hypothesis a true area = 0.50).

years, SpO₂ $< 95\%$, creatinine > 1.3 mg/dL, and absence of COVID-19 vaccination in the overall SOTR cohort. Among nonvaccinated SOTR, age ≥ 70 years, diabetes mellitus, SpO₂ $< 95\%$, creatinine > 1.3 mg/dL, and LDH > 300 U/L were independently associated with all-cause mortality at day 30, and in vaccinated patients, the sole factor independently linked to mortality was the presence of dyspnea at the time of COVID-19 diagnosis.

This study built on the clinical phenotypes established by analyzing immunocompetent adult patients with COVID-19 in 2020¹⁴. Sixteen demographic, clinical, analytical, and radiological variables were identified to evaluate the likelihood of death during the COVID-19 diagnosis (<http://fen-covid.com/index.html>). In the validation cohort of the general population¹⁴, a pronounced contrast in mortality rates was observed among patients with phenotypes A (3.7%), B (23.7%), and C (51.4%). The present study noted a more gradual increase in mortality rates across phenotypes, ranging from phenotype A (9.5%) to phenotype B (16.7%). This gradual increase indicates higher clinical applicability, although the mortality rate for nonvaccinated SOTR with phenotype C remained elevated (52.0%), similar to the general population. Notably, the mortality rate of phenotype C in vaccinated SOTR (17.3%) was lower than in nonvaccinated SOTR and the general population (52.0% and 51.4%, respectively).

In a previous study conducted before the availability of vaccinations, four baseline factors were identified as independent predictors of unfavorable outcomes, defined as the need for intensive care or death: advanced age, high respiratory rate, lymphopenia, and elevated LDH level⁴. In the current study, which encompassed the entire SOTR cohort, advanced age and SpO₂ < 95% potentially influenced respiratory rate, emerged as independent risk factors for mortality. The absence of COVID-19 vaccination was identified as an independent risk factor. Moreover, the mortality rate among vaccinated SOTR in this study (13.8%) was lower than that among nonvaccinated SOTR (23.4%), which, in turn, mirrored that in the general population (28.1%)¹⁴. This result underscores the need to vaccinate immunocompromised patients, including those with SOTR.

In the present study, the vaccinated status of patients was recorded depending on having received at least two vaccine doses, without considering if they might have received additional doses or the time after the last vaccine administration. A longitudinal study involving 129 kidney transplant recipients, after three doses of mRNA vaccine, revealed that only 41.1% of patients had significant neutralizing antibody titers. Moreover, analysis of the concerned variants indicated a diminished binding affinity towards the Omicron variants BA1 and BA2¹⁸. Other study carried out in 101 SOTR, found that 1-month after a third dose of mRNA vaccine 68% of patients had anti-SARS-CoV-2 antibodies, higher than the 40% after the second dose, and with increased antibody titers in those with positive antibodies after the second dose¹⁹. However, some studies in immunocompetent patients, demonstrating varied immune responses of individuals following an Omicron outbreak²⁰, and in SOTR, showing that some patients may remain at high risk for Omicron infection in spite of receiving a fourth vaccine dose²¹, underline the heterogeneity of SARS-CoV-2 immune responses between individuals, including after vaccination. Moreover, the durability of SARS-CoV-2 antibodies has been evaluated over six-month after the second dose of mRNA vaccine in 312 SOTR, using semiquantitative anti-spike antibody testing. Among the 198 patients with positive titers at one-month, 7.1% were negative at six months, and 87 (27.2%) patients of the entire cohort had negative titers at six-month²². Thus, although the vaccinated SOTR criteria in the present study was only to have received at least two vaccine doses, the inter-individual variability of the immune response and its durability decrease the risk of bias in the analysis of the 94 vaccinated SOTR in our study. Besides, stratifying the vaccinated SOTR by two vs. three or more vaccine doses, or the time from the last shot, would decrease the ability to achieve the main objective, aimed to assess and validate the applicability of the clinical phenotypes, identified in the general population, in SOT recipients with COVID-19.

The present study has several limitations. First, our cohort of SOTR was smaller than the general population cohort¹⁴, where clinical phenotypes were identified. Furthermore, our focus on hospitalized patients may restrict the generalizability of our conclusions regarding SOTR treatment in outpatient settings. Second, the period of SOTR inclusion does not exactly represent the current mortality rate of COVID-19 in these patients, which has decreased because of the COVID-19 vaccine and the antivirals availability in the clinical practice, although this is reflected in the significant lesser mortality found in the vaccinated than in the nonvaccinated patients in the present study. However, considering the current changes in the prevalence of the SARS-CoV-2 variants of interest and variants under monitoring¹, with increased vaccine dosing and availability of antivirals, future analyses to validate clinical phenotypes would be desirable. Finally, the size of the vaccinated patient cohort was smaller than that of the nonvaccinated cohort, and the limited number of deaths hampered the ability to identify independent risk factors associated with mortality robustly.

However, this study had some notable strengths. It has a robust design that involves a multicenter and international approach to enhance the generalizability and comparability of the results. Standardized and anonymous data collection contributed to the reliability of the study. The 30-day follow-up period provides a comprehensive perspective. Importantly, this study represents the first analysis of a prospective cohort of SOTR vaccinated against COVID-19 to predict mortality-related factors.

In conclusion, clinical phenotypes are valuable in assessing the risk of mortality among nonvaccinated and vaccinated SOTR, and therefore useful in guiding the level of care in patients with COVID-19. In vaccinated SOTR, dyspnea, a readily identifiable clinical symptom, emerged as an independent risk factor for increased mortality, highlighting the importance of early treatment and close monitoring of individuals exhibiting this symptom. Moreover, vaccination against SARS-CoV-2 decreases the all-cause mortality of COVID-19 by half in the SOTR, which underlies the crucial role of vaccination against SARS-CoV-2 in the SOTR and serves as a pivotal factor in reducing COVID-19 mortality.

Data availability

All relevant data are within the paper and its Supporting information files. The raw data are accessible upon reasonable request to the corresponding author.

Received: 4 June 2024; Accepted: 25 November 2024

Published online: 03 December 2024

References

1. Covid-19 cases | WHO COVID-19 Dashboard. World Health Organization. (2024). <https://data.who.int/dashboards/covid19/> (accessed 6 Nov 2024).
2. Berenguer, J. et al. Characteristics and predictors of death among 4035 consecutively hospitalized patients with COVID-19 in Spain. *Clin. Microbiol. Infect.* **26**, 1525–1536. <https://doi.org/10.1016/j.cmi.2020.07.024> (2020).
3. Lu, Q. B. et al. Comorbidities for fatal outcome among the COVID-19 patients: a hospital-based case-control study. *J. Infect.* **82**, 159–198. <https://doi.org/10.1016/j.jinf.2020.07.026> (2021).
4. Salto-Alejandre, S. et al. Risk factors for unfavorable outcome and impact of early post-transplant infection in solid organ recipients with COVID-19: a prospective multicenter cohort study. *PLoS One.* **16**, e0250796. <https://doi.org/10.1371/journal.pone.0250796> (2021).
5. Benotmane, I., Risch, S., Doderer-Lang, C., Caillard, S. & Fafi-Kremer, S. Long-term shedding of viable SARS-CoV-2 in kidney transplant recipients with COVID-19. *Am. J. Transpl.* **21**, 2871–2875. <https://doi.org/10.1111/ajt.16636> (2021).
6. Kaul, D. R. et al. Donor to recipient transmission of SARS-CoV-2 by lung transplantation despite negative donor upper respiratory tract testing. *Am. J. Transpl.* **21**, 2885–2889. <https://doi.org/10.1111/ajt.16532> (2021).
7. Kemlin, D. et al. Humoral and cellular immune correlates of protection against COVID-19 in kidney transplant recipients. *Am. J. Transpl.* **23**, 649–658. <https://doi.org/10.1016/j.ajt.2023.02.015> (2023).
8. Yang, J. et al. Augmented humoral and cellular immunity against severe acute respiratory syndrome coronavirus 2 after breakthrough infection in kidney transplant recipients who received 3 doses of coronavirus disease 2019 vaccine. *Am. J. Transpl.* **23**, 565–572. <https://doi.org/10.1016/j.ajt.2022.12.022> (2023).
9. Masotti, L. et al. Predictors of poor outcome in tocilizumab treated patients with Sars-CoV-2 related severe respiratory failure: a multicentre real-world study. *Int. Immunopharmacol.* **107**, 108709. <https://doi.org/10.1016/j.intimp.2022.108709> (2022).
10. Portal de datos estadísticos y Geoespaciales de Andalucía: Inicio. Portal de Datos Estadísticos y Geoespaciales de Andalucía | Inicio. <http://www.juntadeandalucia.es/institutoestadisticaycartografia/> (accessed 21 Mar 2024).
11. Lee, A. R. Y. B. et al. Efficacy of covid-19 vaccines in immunocompromised patients: systematic review and meta-analysis. *BMJ* **376**, e068632. <https://doi.org/10.1136/bmj-2021-068632> (2022).
12. Strasfeld, L. COVID-19 and HSCT (hematopoietic stem cell transplant). *Best Pract. Res. Clin. Haematol.* **35**, 101399. <https://doi.org/10.1016/j.beha.2022.101399> (2022).
13. Fauchoux, L., Bassolli de Oliveira Alves, L., Chevret, S. & Rocha, V. Comparison of characteristics and laboratory tests of COVID-19 hematological patients from France and Brazil during the pre-vaccination period: identification of prognostic profiles for survival. *Hematol. Transfus. Cell. Ther.* **45**, 306–316. <https://doi.org/10.1016/j.htct.2022.05.003> (2023).
14. Gutiérrez-Gutiérrez, B. et al. Identification and validation of clinical phenotypes with prognostic implications in patients admitted to hospital with COVID-19: a multicentre cohort study. *Lancet Infect. Dis.* **21**, 783–792. [https://doi.org/10.1016/S1473-3099\(21\)0019-0](https://doi.org/10.1016/S1473-3099(21)0019-0) (2021).
15. Crespo, M. et al. Respiratory and gastrointestinal COVID-19 phenotypes in kidney transplant recipients. *Transplantation* **104**, 2225–2233. <https://doi.org/10.1097/TP.0000000000003413> (2020).
16. Harris, P. A. et al. Research electronic data capture (REDCap)—A metadata-driven methodology and workflow process for providing translational research informatics support. *J. Biomed. Inf.* **42**, 377–381. <https://doi.org/10.1016/j.jbi.2008.08.010> (2009).
17. WHO Working Group on the Clinical Characterisation and Management of COVID-19 infection. A minimal common outcome measure set for COVID-19 clinical research. *Lancet Infect. Dis.* **20**, e192–e197. [https://doi.org/10.1016/S1473-3099\(20\)30483-7](https://doi.org/10.1016/S1473-3099(20)30483-7) (2020).
18. Murali, T. M. et al. Analyzing COVID-19 vaccine responses in transplant recipients. *Immunohorizons* **7**, 708–717. <https://doi.org/10.4049/immunohorizons.2300071> (2023).
19. Kamar, N. et al. Three doses of an mRNA Covid-19 vaccine in Solid-Organ transplant recipients. *N. Engl. J. Med.* **385**, 661–662. <https://doi.org/10.1056/NEJMc2108861> (2021).
20. Wu, J. et al. Heterogeneity of SARS-CoV-2 immune responses after the nationwide Omicron wave in China. *Microbiol. Spectr.* e0111724. <https://doi.org/10.1128/spectrum.01117-24> (2024).
21. Karaba, A. H. et al. A fourth dose of COVID-19 vaccine does not induce neutralization of the Omicron variant among solid organ transplant recipients with suboptimal vaccine response. *Transplantation* **106**, 1440–1444. <https://doi.org/10.1097/TP.00000000000004140> (2022).
22. Alejo, J. L., Palacios-Baena, R. & Torre-Cisneros. Six-month antibody kinetics and durability in SARS-CoV-2 mRNA vaccinated solid organ transplant recipients. *Transplantation.* **106**, e109–e110. <https://doi.org/10.1097/TP.00000000000003975> (2022).

Acknowledgements

This study was supported by Plan Nacional de I+D+i 2013-2016 and Instituto de Salud Carlos III, Subdirección General de Redes y Centros de Investigación Cooperativa, Ministerio de Ciencia, Innovación y Universidades, Spanish Network for Research in Infectious Diseases (REIPI, RD16/0016/0001 [JR-B, BG-G], RD16/0016/0005 [NS], and RD16/0016/0009 [JP, EC, JSC]), co-financed by European Development Regional Fund “A way to achieve Europe”, Operative Program Intelligence Growth 2014-2020. EC, JSC and JR-B received grants from the Instituto de Salud Carlos III, Ministerio de Ciencia e Innovación, Proyectos de Investigación sobre el SARS-CoV-2 y la enfermedad COVID-19 (COV20/00370, COV20/00580, COV20/01031). EC and JSC also were supported by the grant IM22/INF/13 from the CIBERINFEC, Instituto de Salud Carlos III, Spain. This study was also supported by CIBERINFEC - Consorcio Centro de Investigación Biomédica en Red, Instituto de Salud Carlos III, Ministerio de Ciencia e Innovación and Unión Europea – NextGenerationEU (EC and JS-C [CB21/13/00006], BG-G and JR-B [CB21/13/00012]). JSC is a researcher belonging to the program “Nicolás Monardes” (RC-0002–2022), Servicio Andaluz de Salud, Junta de Andalucía, Spain. The ORCHESTRA project has received funding from the European Union’s Horizon 2020 research and innovation programme under grant agreement No 101016167.

Author contributions

Conceptualization, E.C., J.P., J.R.-B., and B.G.-G.; methodology, E.C., J.P., J.R.-B., B.G.-G., C.I.-D., and S.S.-A.; software, S.J.-J.; validation, J.P., C.I.-D., S.S.-A., and S.J.-J.; formal analysis, J.P., C.I.-D., and S.S.-A.; data curation, R.A.-M., N.S., A.R., A.M., K.M., Z.P.-B., P.M., M.F.-R., M.B., C.F., E.V., E.M.-L., M.H., R.H.-G., M.B., A.M., A.G.-D., M.R., S.J.-J., M.B., L.F.A.-C., M.V., and J.S.-C.; writing—original draft preparation, C.I.-D. and S.S.-A.; writing—review and editing, J.P., E.C., J.R.-B., B.G.-G., and M.G.; visualization, R.A.-M., N.S., A.R., A.M., K.M., Z.P.-B., P.M., M.F.-R., M.B., C.F., E.V., E.M.-L., M.H., R.H.-G., M.B., A.M., A.G.-D., M.R., S.J.-J., M.B., L.F.A.-C., M.V., and J.S.-C.; supervision, J.P., E.C., J.R.-B., B.G.-G., and M.G.; project administration, J.P., J.S.-C., E.C.,

J.R.-B., and M.G.; funding acquisition, J.S.-C., E.C., J.R.-B., and M.G. All authors have read and agreed to the published version of the manuscript.

Declarations

Competing interests

The authors declare no competing interests.

Statement

The views expressed in this publication are the sole responsibility of the authors and the Commission is not responsible for any use that may be made of the information it contains.

Additional information

Supplementary Information The online version contains supplementary material available at <https://doi.org/10.1038/s41598-024-81099-2>.

Correspondence and requests for materials should be addressed to J.P.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by-nc-nd/4.0/>.

© The Author(s) 2024

The COVIDSOT, ORCHESTRA Working Teams

Carmen Infante-Domínguez²⁵, Sonsoles Salto-Alejandre²⁵, Rocío Álvarez-Marín²⁵, Silvia Jiménez-Jorge²⁵, Javier Sánchez-Céspedes²⁵, Jerónimo Pachón²⁵, Elisa Cordero²⁵, José Miguel Cisneros²⁵, Manuela Aguilar-Guisado²⁵, José María Álamo-Martínez²⁵, Carmen Bernal-Bellido²⁵, Gabriel Bernal-Blanco²⁵, Pedro Camacho²⁵, Marta Carretero²⁵, Carmen Cepeda-Franco²⁵, Miguel Ángel Gómez-Bravo²⁵, Antonio Grande-Trillo²⁵, Luis Miguel Marín-Gómez²⁵, Laura Merino²⁵, Gema Montilla-Cosano²⁵, María Paniagua²⁵, José Manuel Sobrino-Márquez²⁵, Diego Rangel-Sousa²⁵, Blanca Villacorta-Linaza²⁵, Nuria Sabé²⁶, José González-Costello²⁶, Laura Lladó²⁶, Eduardo Melilli²⁶, Antonio Ramos-Martínez²⁷, Jorge Calderón-Parra²⁷, Ana Arias-Milla²⁷, Asunción Moreno²⁸, Marta Bodro²⁸, Laura Linares²⁸, Sabina Herrera²⁸, María Angeles Marcos²⁸, Federico Cofán²⁸, María Angeles Castel²⁸, Jordi Colmenero²⁸, Kamilla Ferreira de Moraes²⁹, Luis Fernando Aranha Camargo²⁹, Zaira R. Palacios-Baena³⁰, Belén Gutiérrez-Gutiérrez³⁰, Jesús Rodríguez-Baño³⁰, Belén Gallego³⁰, Natalia Maldonado-Lizarazo³⁰, Patricia Muñoz³¹, Maricela Valerio³¹, Ainhoa Fernández-Yunquera³¹, Carlos Ortiz-Bautista³¹, María Luisa Rodríguez-Ferrero³¹, Mario Fernández-Ruiz³², José María Aguado³², Francisco López-Medrano³², Rafael San Juan³², Marino Blanes-Julia³³, Rosa Blanes-Hernández³³, Carmen Fariñas³⁴, Francisco Arnaiz de las Revillas-Almajano³⁴, Ignacio Fortea Ormaechea³⁴, Claudia González-Rico³⁴, Mónica Gozalo-Margüello³⁴, Aritz Gil-Ongay³⁴, Milagros Heras Vicario³⁴, Elisa Vidal³⁵, Julián Torre-Giménez³⁵, Julián Torre-Cisneros³⁵, Esperanza Merino de Lucas³⁶, Pilar González-de-la-Aleja³⁶, Silvia Otero³⁶, Héctor Pinargote-Celorio³⁶, Márcia Halpern³⁷, Elizabeth Balbi³⁷, Román Hernández-Gallego³⁸, Elena García Vinuesa-Calvo³⁸, Rocío Martínez Gallardo³⁸, Matteo Bassetti³⁹, Laura Nicolini³⁹, Antonio Vena³⁹, Alessandra Mularoni⁴⁰, Giovanna Russellì⁴⁰, Alex Gutiérrez-Dalmau⁴¹, María José Aladren-Regidor⁴¹, Javier Paul-Ramos⁴¹, Maddalena Giannella⁴², Matteo Rinaldi⁴², Cecilia Bonazzetti⁴², Natascia Caroccia⁴², Michela

**Chiara⁴², Domenico Marzolla⁴², Renato Pascale⁴², Zeno Pasquini⁴², Francesca Simone⁴²,
Beatrice Tazza⁴² & Alice Toschi⁴²**

²⁵Virgen del Rocío University Hospital-IBIS, University of Seville, Seville, Spain. ²⁶Bellvitge University Hospital-IDIBELL, University of Barcelona, Barcelona, Spain. ²⁷Hospital Universitario Puerta de Hierro-Majadahonda-IDIPHISA, Madrid, Spain. ²⁸Hospital Clinic-IDIBAPS. University of Barcelona, Barcelona, Spain. ²⁹Hospital Israelita Albert Einstein, São Paulo, Brazil. ³⁰Virgen Macarena University Hospital-IBIS, Seville, Spain. ³¹Gregorio Marañón University Hospital, Institute of Health Research Gregorio Marañón (IiSGM), Madrid, Spain. ³²12 de Octubre University Hospital/I+12, CIBERCV, Madrid, Spain. ³³La Fe University Hospital, Valencia, Spain. ³⁴Marqués de Valdecilla University Hospital-IDIVAL, University of Cantabria, Santander, Spain. ³⁵Reina Sofía University Hospital-IMIBIC, Córdoba, Spain. ³⁶Alicante General University Hospital, Alicante Institute of Health and Biomedical Research (ISABIAL), Alicante, Spain. ³⁷Quinta D'Or Hospital, Rio de Janeiro, Brasil. ³⁸Badajoz University Hospital, Extremadura, Spain. ³⁹Policlinico San Martino Hospital-IRCCS, Genoa, Italy. ⁴⁰ISMETT-IRCCS-Istituto Mediterraneo per i Trapianti e Terapie ad Alta Specializzazione-Istituto di Ricovero e Cura a Carattere Scientifico, Palermo, Italy. ⁴¹Miguel Servet University Hospital, Aragón Institute for Health Research IIS-Aragón, Zaragoza, Spain. ⁴²IRCCS Azienda Ospedaliero Universitaria di Bologna, Alma Mater Studiorum University of Bologna, Bologna, Italy.