

Alma Mater Studiorum Università di Bologna
Archivio istituzionale della ricerca

Clinical, experimental and pathophysiological effects of Yaq-001: a non-absorbable, gut-restricted adsorbent in models and patients with cirrhosis

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Liu J., MacNaughtan J., Kerbert A.J.C., Portlock T., Gonzalez J.M., Jin Y., et al. (2024). Clinical, experimental and pathophysiological effects of Yaq-001: a non-absorbable, gut-restricted adsorbent in models and patients with cirrhosis. GUT, epub before print, 1-16 [10.1136/gutjnl-2023-330699].

Availability:

This version is available at: <https://hdl.handle.net/11585/968845> since: 2024-05-05

Published:

DOI: <http://doi.org/10.1136/gutjnl-2023-330699>

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>).
When citing, please refer to the published version.

(Article begins on next page)

Clinical, Experimental and Pathophysiological effects of Yaq-001, a Non-absorbable, Gut-restricted Adsorbent in Models and Patients with Cirrhosis

Authorships and Affiliations

Author	Affiliations	email	Conflict of interest
Jinxia Liu [#]	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF. Department of Gastroenterology, Affiliated Hospital of Nantong University, Nantong, 226001, China.	liujinxia@ntu.edu.cn	
Jane Macnaughtan [#]	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	j.macnaughtan@ucl.ac.uk	
Annarein JC Kerbert	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	J.C.Kerbert@lumc.nl	
Javier Martinez	Hospital Ramón y Cajal, IRYCIS, CIBEREHD, Universidad de Alcalá, Madrid, Spain Liver Unit, Hospital Vall d'Hebron, Universitat Autònoma, CIBERehd, Barcelona, Spain	martinez.gonzalez.javier@gmail.com	
Yi Jin	Centre for Host-Microbiome Interactions, Faculty of Dentistry, Oral & Craniofacial Sciences, King's College London, SE1 9RT, UK.	yi.1.jin@kcl.ac.uk	
Frederick Clasen	Centre for Host-Microbiome Interactions, Faculty of Dentistry, Oral & Craniofacial Sciences, King's College London, SE1 9RT, UK.	frederick.clasen@crick.ac.uk	
Abeba Habtesion	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	a.habtesion@ucl.ac.uk	
Huoyan Jin	Department of Gastroenterology, Affiliated Hospital of Nantong University, Nantong, 226001, China.	jihuoyan@163.com	
Qin Jin	Department of Gastroenterology, Affiliated Hospital of Nantong University, Nantong, 226001, China.	jinqin@sina.com	

Alexandra Phillips	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	alexandra.phillips@ucl.ac.uk	
Francesco De Chiara	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	fdechiara@ibecbarcelona.eu	
Ganesh Ingavle	Centre for Regenerative Medicine and Devices, School of Applied Sciences, University of Brighton, Brighton, East Sussex, BN2 4GJ, UK. Symbiosis Centre for Stem Cell Research (SCSCR), Symbiosis School of Biological Sciences (SSBS), Symbiosis International (Deemed University), Pune 412115, India.	ganesh.ingavle@ssbs.edu.in	
Cesar Jimenez	Liver Unit, Hospital Vall d'Hebron, Universitat Autònoma, CIBERehd, Barcelona, Spain	cesar.jimenez@vhebron.net	
Giacomo Zaccherini	University of Bologna, Italy	g.zaccherini@gmail.com	
Katherine Husi	Gastroenterology, University Hospital Bern, CH	Kathrin.Husi@insel.ch	
Miguel Rodriguez-Gandia	Hospital Ramón y Cajal, IRYCIS, CIBEREHD, Universidad de Alcalá, Madrid, Spain	maldimed@gmail.com	
Paul Cordero-Sanchez	Symbiosis Centre for Stem Cell Research (SCSCR), Symbiosis School of Biological Sciences (SSBS), Symbiosis International (Deemed University), Pune 412115, India.	paul.sanchez@ucl.ac.uk	
Junpei Soeda	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	Deceased	
Lynda McConaghy	Yaqrit	lynda.mcconaghy@yaqrit.com	
Jude A Oben	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	jude.1.oben@kcl.ac.uk	
Karen Church	Yaqrit	karen.church@yaqrit.com	

Jia Li	®Department of Surgery and Cancer, Faculty of Medicine, Imperial College London, Sir Alexander Fleming Building, Imperial College Road, South Kensington, London, SW7 2AZ.	jia.li@imperial.ac.uk	
Haifeng Wu	Department of Gastroenterology, Affiliated Hospital of Nantong University, Nantong, 226001, China.	wuhaifeng1986@163.com	
Aarti Jalan	Kings College Hospital, 125 Coldharbour Lane, London SE5 9NU, UK London	aartijalan1@gmail.com	
Pere Gines	Liver Unit, Hospital Clinic of Barcelona, IDIBAPS, Faculty of Medicine and Health sciences, University of Barcelona. CIBEReHD	PGINES@clinic.ub.es	
Elsa Sola	Liver Unit, Hospital Clinic of Barcelona, IDIBAPS, Faculty of Medicine and Health sciences, University of Barcelona. CIBEReHD	ESOLA@clinic.ub.es	
Simon Eaton	ICH-UCL	s.eaton@ucl.ac.uk	
Carrie Morgan	Yaqrit	carrie.morgan@yaqrit.com	
Michal Kowalski	Yaqrit	michal.kowalski@yaqrit.com	
Daniel Green	Yaqrit	daniel.green@yaqrit.com	
Amir Gander	Tissue Access for Patient Benefit: ROYAL FREE HOSPITAL	a.gander@ucl.ac.uk	
Lindsey Ann Edwards	Institute of Liver Studies, School of Immunology and Microbial Sciences, Faculty of Life Sciences and Medicine, King's College London, 125 Coldharbour Lane, London SE5 9NU, UK	lindsey.edwards@kcl.ac.uk	
I. Jane Cox	The Roger Williams Institute of Hepatology, Foundation for Liver Research, 111 Coldharbour Lane, London SE5 9NT	j.cox@researchinliver.org.uk	
Helena Cortez-Pinto	Clínica Universitária de Gastreenterologia, Laboratório de Nutrição, Faculdade de Medicina, Universidade de Lisboa, Portugal	hlcortezpinto@gmail.com	
Reiner Wiest	Gastroenterology, University Hospital Bern, CH	reiner.wiest@insel.ch	
Francois Durand	Hepatology, Hospital Beaujon, Clichy, France	francois.durand@aphp.fr	
Paolo Caraceni	University of Bologna, Italy	paolo.caraceni@unibo.it	
Roberto Elosua	Datarus, Barcelona	relosua@datarus.eu	
Joan Vila	Datarus, Barcelona	jvila@datarus.eu	

Marco Pavesi	European Foundation for the Study of Chronic Liver Failure (EF Clif), Barcelona	mpavesi@gmail.com	
Vicente Arroyo	European Foundation for the Study of Chronic Liver Failure (EF Clif), Barcelona	vicente.arroyo@efclif.com	
Nathan Davies	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	nathan.davies@ucl.ac.uk	
Rajeshwar P Mookerjee	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	r.mookerjee@ucl.ac.uk	
Victor Vargas	Liver Unit, Hospital Vall d'Hebron, Universitat Autònoma, CIBERehd, Barcelona, Spain	vvargas@vhebron.net	
Susan Sandeman	Centre for Regenerative Medicine and Devices, School of Applied Sciences, University of Brighton, Brighton, East Sussex, BN2 4GJ, UK.	s.sandeman@brighton.ac.uk	
Gautam Mehta	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	gautam.mehta@ucl.ac.uk	
Saeed Shoaie	Centre for Host-Microbiome Interactions, Faculty of Dentistry, Oral & Craniofacial Sciences, King's College London, SE1 9RT, UK.	saeed.shoaie@kcl.ac.uk	
Julian R. Marchesi	Division of Digestive Diseases, Department of Metabolism, Digestion and Reproduction, St Mary's Hospital, Imperial College London, South Wharf Road, London, W2 1NY	j.marchesi@imperial.ac.uk	
Agustin Albillos	Hospital Ramón y Cajal, IRYCIS, CIBEREHD, Universidad de Alcalá, Madrid, Spain	agustin.albillos@uah.es	
Fausto Andreola	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF.	f.andreola@ucl.ac.uk	
Rajiv Jalan	Liver Failure Group, UCL Institute for Liver and Digestive Health, Upper third floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF. European Foundation for the Study of Chronic Liver Failure (EF Clif), Barcelona	r.jalan@ucl.ac.uk	

297

298 # Both authors share first authorship.

299

300 **Correspondence:** Rajiv Jalan, UCL Institute for Liver and Digestive Health, Upper third
301 floor, Royal Free Campus, Rowland Hill Street, Hampstead, London, NW3 2PF. Email:
302 r.jalan@ucl.ac.uk

303

304 **Keywords:** ACLF; Liver injury; organ dysfunction; LPS; endotoxemia; bacterial
305 translocation; dysbiosis; antibiotic resistance; Yaq-001

306

307 **Electronic word count:** 3991 words

308

309 **Number of figures and tables:** 8

310

311 **Conflict of Interest**

312 Rajiv Jalan is the inventor of OPA, which has been patented by UCL and licensed to
313 Mallinckrodt Pharma. He is also the founder of Yaqrit Discovery, Hepyx Limited (spin out
314 companies from University College London), and Cyberliver. He has research
315 collaborations with Yaqrit Discovery. Yaq-001 was licensed by Yaqrit Ltd. from UCL. JRM
316 has received consultancy fees from EnteroBiotix and Cultech, and speaker fees from Falk
317 Forum.

318

319 **Financial support**

320 This study was performed with support from a grant from the EU H2020, Grant Agreement
321 number: 634579 — CARBALIVE — H2020-PHC-2014-2015/H2020-PHC-2014
322 programme. JRM and the Division of Digestive Diseases at Imperial College London
323 receives financial support from the NIHR Imperial Biomedical Research Centre.

324

325 **Authors' contributions**

326 RJ, FA, JL, JM, ND - contributed to the conception and design of the study. SS and GI
327 contributed to the conception and design of the *in vitro* studies. RJ, ND, FA - provided

328 administrative, study supervision, obtained funding, material support. JL, JM, LE, YJ, FC,
329 AH, AP, FD, GI, PC, JS, JO, JL, HW, JC, SS, RM - performed experiments and
330 substantially contributed to the acquisition of data and its analysis. All authors were
331 involved in the interpretation of data. JL and JM drafted the manuscript. All authors
332 revised the manuscript critically for important intellectual content.

333

334 **Abstract**

335 **Objective:** Strategies to target bacterial translocation in cirrhosis are limited to antibiotics
336 with risk of antimicrobial resistance. This study explored the therapeutic potential of a
337 non-absorbable, gut-restricted, engineered carbon bead adsorbent, Yaq-001 in models
338 of cirrhosis and acute-on-chronic liver failure (ACLF) and, its safety and tolerability in a
339 clinical trial in cirrhosis.

340 **Design:** Performance of Yaq-001 was evaluated *in vitro*. Two-rat models of cirrhosis and
341 ACLF, [4-week, bile duct ligation (BDL)], receiving Yaq-001 for 2-weeks; and two-mouse
342 models of cirrhosis [6-week and 12-week carbon tetrachloride (CCL4)] receiving Yaq-001
343 for 6-weeks were studied. Organ and immune function, gut permeability, transcriptomics,
344 microbiome composition and metabolomics were analysed. Effect of fecal water on gut
345 permeability from animal models were evaluated on intestinal organoids. A double-blind,
346 randomized, placebo-controlled clinical trial in 28-cirrhosis patients, administered 4-
347 gram/day Yaq-001 for 3-months was performed.

348 **Results:** Yaq-001 exhibited rapid adsorption kinetics for endotoxin. *In vivo*, Yaq-001
349 reduced liver injury, progression of fibrosis, portal hypertension, renal dysfunction and
350 mortality of ACLF animals significantly. Significant impact on severity of endotoxemia,
351 hyperammonemia, liver cell death, systemic inflammation and organ transcriptomics with
352 variable modulation of inflammation, cell death and senescence in the liver, kidneys, brain
353 and colon was observed. Yaq-001 reduced gut permeability was noted in the organoids
354 and impacted positively on the microbiome composition and metabolism. Yaq-001,
355 regulated as a device met its primary end point of safety and tolerability in the clinical trial.

356 **Conclusions:** This study provides strong pre-clinical rationale and safety in patients with
357 cirrhosis to allow clinical translation.

358

359 **Significance of this study**

360 **What is already known on this topic?**

361 Current strategies to target bacterial translocation in cirrhosis are limited to antibiotics
362 with risk of resistance. Yaq-001 is an insoluble, non-absorbable, non-antibiotic,
363 engineered carbon bead of tailored porosities, which works as an adsorbent in the gut
364 and is completely excreted after oral administration.

365

366 **What this study adds?**

- 367 1. Yaq-001 rapidly adsorbs endotoxin, ammonia and bile acids without influencing
368 bacterial growth kinetics *in vitro*.
- 369 2. Yaq-001 reduces mortality of ACLF animals and impacts positively on markers of gut
370 permeability, liver injury, portal pressure, brain and kidneys in animal models of cirrhosis
371 and ACLF.
- 372 3. Yaq-001 administration was associated with positive impact on the composition of the
373 gut microbiota, reduction in severity of endotoxemia and ammonia, severity of
374 inflammation, liver cell death, signaling pathways and LPS sensitivity.
- 375 4. In animal models of liver fibrosis and cirrhosis, it reduces the severity of liver injury and
376 hepatic fibrosis.
- 377 5. Enhanced permeability of intestinal organoids following incubation with fecal water from
378 cirrhosis animals is prevented by treatment of the cirrhosis animals with Yaq-001.
- 379 6. In a Phase 2, double-blind, randomized, controlled clinical trial of Yaq-001 versus
380 placebo in patients with cirrhosis, Yaq-001 was found to be safe and well tolerated
381 providing evidence of clinical translatability.

382

383 **How this study might affect research, practice or policy?**

384 The data provide the pre-clinical rationale and clinical safety to proceed to the next phase
385 of clinical trials in patients with cirrhosis aiming to prevent the occurrence of complications.

386

387 **INTRODUCTION**

388 Gut dysbiosis and gut-derived bacterial ligands, in particular endotoxin, drive a
389 dysregulated inflammatory response, which has been implicated in the development of
390 cirrhosis and its complications such as sepsis, spontaneous bacterial peritonitis, renal
391 dysfunction and hepatic encephalopathy¹⁻³. This dysregulated inflammatory response is
392 also central in the development of acute-on-chronic liver failure (ACLF)⁴. Markers of
393 bacterial translocation such as endotoxin and bacterial DNA have been shown to be
394 associated with complications of cirrhosis and diminished survival highlighting their
395 pathogenic importance^{5,6,7}. The microbiome in cirrhosis is characterized by reduced
396 diversity and abundance of autochthonous bacteria¹. Whilst antibiotics have been shown
397 to impact positively on complications of cirrhosis, their use is associated with antibiotic
398 resistance^{8,9}. Furthermore, antibiotics reduce bacterial diversity rendering the microbiome
399 less resilient.

400
401 One of the consequences of bacterial translocation in cirrhosis is that the endotoxin-
402 sensing pathways in different organs are known to be primed resulting in heightened
403 susceptibility to organ injury^{3,10}. Adsorption of free endotoxin without exerting direct
404 effects on bacterial growth kinetics, therefore has the potential to attenuate susceptibility
405 to organ injury without producing the deleterious effects on the microbiome. Considering
406 this, we developed a synthetic non-absorbable, non-antibiotic, endotoxin sequestrant and
407 generated the hypothesis that this may be a novel therapeutic strategy to restore the
408 microbiome, prevent bacterial translocation, systemic inflammation progression of fibrosis
409 and cirrhosis complications. Yaq-001 is a non-absorbable, highly engineered, activated
410 carbon of multiple porosities tailored to the micro (<2nm) and meso-macroporous (30-200
411 nm) range and high surface area¹¹⁻¹³. These properties confer a high adsorptive capacity
412 for larger biologically relevant molecules such as bacterial toxins in addition to smaller
413 intraluminal targets. The most closely associated experimental oral adsorbent is AST-
414 120, a microporous carbon bead, which has not been shown to be efficacious in patients
415 with hepatic encephalopathy¹⁴.

416

417 In this study, we sought to determine the adsorptive capacity of Yaq-001 and its effect on
418 bacterial growth kinetics in *in vitro* studies. We then evaluated the *in vivo* biological effects
419 of Yaq-001 in four animal models representing characteristics of cirrhosis and ACLF. We
420 studied the effects of Yaq-001 on measures of multiorgan function, systemic and portal
421 hemodynamics, immune function, multiorgan transcriptomics and microbiome
422 composition. Finally, we performed a Phase 2 equivalent double-blind, multicenter,
423 randomized, placebo-controlled clinical trial to assess the safety and tolerability of Yaq-
424 001 in patients with decompensated cirrhosis.

425

426 **METHODS**

427 Methodological details are described in **Supplementary section**.

428

429 **Functional and Structural Characteristics of Yaq-001**

430 Adsorption of biomolecules of varying molecular weights (albumin, myoglobin, and
431 caffeine) was evaluated. Bacterial growth was studied for *Staphylococcus aureus* (*S.*
432 *aureus*) and *Escherichia coli* (*E. coli*). Scanning electron microscopy was performed to
433 characterise the beads and pore size distribution was assessed using mercury
434 porosimetry.

435

436 **Studies in animal models**

437 **Study design**

438 These studies aimed to characterize the therapeutic potential of Yaq-001 in rats and mice
439 to define its role in prevention of occurrence of cirrhosis, progression of cirrhosis and
440 occurrence of ACLF (**Fig.S1 and Fig.S2**).

441

442 **Animal models**

443 Four-week bile-duct ligation model of advanced fibrosis

444 a. *Cirrhosis*: Sham (n=36); Sham-Yaq-001 (n=30); BDL (n=37); BDL-Yaq-001
445 (n=44).

446 b. *Prevention of ACLF*: Sham-LPS (n=9); Sham-LPS-Yaq-001 (n=10); BDL-LPS
447 (n=16); BDL-LPS-Yaq-001 (n=12).

448

449 Yaq-001 (0.4 g/100g body weight per day) was administered for 2-weeks prior to sacrifice.

450 At the time of sacrifice, mean arterial pressure (MAP) and portal pressure were measured.

451

452 Carbon tetrachloride treated model of cirrhosis

453 c. *Advanced fibrosis and early cirrhosis (CCl₄ for 6-weeks)*: Control (n=6); Control-
454 Yaq-001 (n=6); CCl₄ (n=12); CCl₄-Yaq-001 (n=12).

455 d. *Advanced cirrhosis (CCl₄ for 12-weeks)*: Control (n=6); Control-Yaq-001 (n=6);
456 CCl₄ (n=12); CCl₄-Yaq-001 (n=12).

457
458 Yaq-001 (0.4 g/100 g body weight per day) was administered from 0-6 weeks in the 6-
459 week model and from 6-12 weeks in the 12-week model.

460

461 ***Collection and analysis of bio-samples***

462 Blood, stool and tissues samples were collected for later analysis. Portal venous blood
463 was collected where possible. Peripheral blood cells and Kupffer cell reactive oxidant
464 species (ROS) were measured. Hematoxylin-Eosin (H&E), Picrosirius Red (PSR) staining
465 and TUNEL stains were performed in liver tissues. The mRNA in different organs was
466 analyzed by using nSolver4.0 software (NanoString Technologies). To define effect on
467 the microbiome, 16s microbiome study was performed. To determine the effect of Yaq-
468 001 on modulating metabolism, urinary ¹H-NMR analysis was performed.

469

470 **Assessment of gut permeability in Intestinal organoids**

471 Permeability of mouse intestinal organoids were detected using established protocols¹⁵.
472 Fecal water generated from the stools obtained from the four groups of 6-week CCl₄ mice
473 were incubated with the organoids as described previously. Permeability of the organoids
474 were assessed.

475

476 **Clinical trial of Yaq-001 versus placebo, CARBALIVE-SAFETY study**

477 ***Study Design***

478 The CARBALIVE-SAFETY clinical trial was a first in man, multicenter, double blind
479 randomized, placebo-controlled trial of oral Yaq-001 in stable decompensated cirrhosis.
480 Details of the study protocol is available in the Supplementary section (**Fig.S3**). As Yaq-
481 001 is regulated as a device, it followed both ISO standards and ICH-GCP guidance.
482 Informed consent was obtained from each patient. The study was closely monitored and
483 overseen by an independent data safety monitoring board (NCT03202498).

484

485 Study design is described as **Fig.S4**. The primary end point was assessed at 12-weeks.
486 Blood and stool samples were taken at the time of randomization, 4-weeks and 12-weeks
487 for assessment of some of the secondary and exploratory end points. Safety

488 assessments were performed on weeks 1, 4, 8 and 12 and comprised a physical
489 examination, clinical laboratory tests, urinalysis, 12-lead ECG and an assessment of
490 reported and observed adverse events. ECGs were analyzed independently. Nutritional
491 status was assessed by the Royal Free Hospital Global Assessment tool at each safety
492 assessment together with micronutrient analysis at baseline, week 4 and 12. Vitamin B12,
493 A, D, E, folate, and K1 and, trace elements Copper, Zinc and Selenium were analyzed.

494

495 ***Main Inclusion and Exclusion Criteria***

496 The main inclusion criteria were participants aged 18-years or above, clinical diagnosis
497 of diuretic-responsive cirrhotic ascites (Child-Pugh score = 7-11 inclusive), abstinence
498 from alcohol for at least 4-weeks prior to screening and written informed consent. The
499 main exclusions were lack of informed consent, use of oral antibiotics,
500 immunosuppressants or antiviral medication within 4-weeks prior to recruitment, change
501 in dose of proton pump inhibitor therapy within 4-weeks before the start of the study
502 treatment, hospital admission for liver-related indication for at least 4-weeks (except
503 paracentesis), BMI > 35 or BMI < 18, presence of a transjugular intrahepatic
504 portosystemic shunt (See protocol in supplementary for details).

505

506 ***Randomization, Dosing and Compliance***

507 Patients were randomized 1:1 to receive 4g of oral Yaq-001 or equivalent placebo nocte
508 for 12-weeks. Treatment compliance was assessed by the number of used or unopened
509 sachets returned to the clinical site at each visit. Patients taking $\geq 70\%$ of study
510 medication were considered compliant.

511

512 ***Endpoints and Assessments***

513 Primary Endpoints

514 The main objective of this clinical investigation is to assess the safety and tolerability of
515 Yaq-001 throughout the three months' treatment period.

516

517 Secondary and Exploratory Endpoints

518 Blood and stool samples were collected for later analysis for markers of endotoxemia,
519 systemic inflammation, bile acids, short chain fatty acids, gut permeability and the
520 microbiome (results not reported in this paper).

521

522 **Statistical Analysis**

523 ***Animal Studies***

524 Based on the *in vitro* studies, we anticipated a 50% decrease in circulating endotoxin in
525 the treatment groups with an alpha error of 0.05 and power of 80%, resulting in a minimum
526 sample size of 5 animals/group. As this study included several pathophysiological end
527 points, multiple experimental groups were included. All the data accrued from these
528 studies are described in this paper. All the rats in eight groups from three independent
529 batches were included in the analysis as shown in **Fig.S1**. All the mice studied in eight
530 groups were included in **Fig.S2**.

531

532 Group comparisons for continuous variables were performed using Man-Whitney U test
533 (no-normal distribution) or unpaired t-test (normal distribution) and for categorical
534 variables by using Chi-squared test. The data were analyzed using R package (R version
535 4.4.4). 16s microbiome study and circos correlation were analyzed by using Wilcoxon
536 rank sum test and spearman correlation. Software used included Graphpad Prism 9.0
537 (GraphPad software, Inc., San Diego, CA).

538

539 ***CARBALIVE-SAFETY Clinical Trial***

540 This first-in-man clinical investigation was not powered to demonstrate statistical
541 significance for any endpoint. All statistical analyses of study data were carried out using
542 SAS v 9.3 or a later version. For categorical variables, summary tabulations of the number
543 and percentage of patients within each category (with a category for missing data) of the
544 parameter are presented. Percentage calculations are based on non-missing data unless
545 otherwise specified. Please also see statistical analysis plan (Supplementary).

546

547

548 **RESULTS**

549 **Functional and Structural Characteristics of Yaq-001**

550 Yaq-001 beads exhibited a consistent pre-defined structure with a bead diameter within
551 the 250-500 μm range and the prescribed porosity (**Fig.S5A**). Yaq-001 rapidly adsorbed
552 albumin (66.5kDa), myoglobin (16.7kDa) and caffeine (0.194kDa) representing different
553 sized biomolecules (**Fig.S5B**). Yaq-001 adsorbed LPS (18kDa) reducing the
554 concentrations from 2.5 to 1.5 EU mL^{-1} (60%) within 30 minutes. No endotoxin was
555 detected in the control solution (0 EU mL^{-1}) (**Fig.S5B**). Yaq-001 also adsorbed a range of
556 bile acids (**Fig.S5C**). Direct co-incubation of Yaq-001 with bacterial suspensions of either
557 *E. coli* or *S. aureus* indicated that Yaq-001 did not affect bacterial growth kinetics for either
558 species following direct contact in comparison to the antibiotic controls (**Fig.S5D**).
559 Mercury porosimetry showed that Yaq-001 used in the clinical trial had a consistent pore
560 size distribution plot in the meso-macroporous range from 30-200 nm (**Fig.S5E**).

561
562 Yaq-001 exhibited better performance in adsorptive capacity and effect on endotoxin
563 kinetics than AST-120(**Fig.S5**).

564

565 **Studies in animal models**

566 ***Studies in BDL rat model of advanced fibrosis***

567 Effect of Yaq-001 on liver injury and portal pressure

568 BDL rat model was used to assess the effect of Yaq-001 in cirrhosis (**Fig.1A**). Significant
569 reduction in 4-week body weight was observed in BDL rats ($p<0.0001$), which was
570 prevented by administration of Yaq-001 ($p=0.045$) (**Fig.1A**). Yaq-001 was associated with
571 a significantly lower plasma ALT ($p=0.007$). ALP, TBIL and albumin were not impacted
572 by Yaq-001 (**Fig.S6A, B, C**). Total bile acid concentrations were not different between the
573 BDL and Sham groups and there was no significant impact of Yaq-001 (**Fig.S6E**). MAP
574 was lower in BDL animals and no effect of Yaq-001 was observed (**Fig.S6F**). Yaq-001
575 resulted in a significant reduction in portal pressure compared to untreated BDL rats
576 [(median (IQR) 11.1 mm Hg (10.3-11.7) vs 12.4 mm Hg (10.8-13.3), ($p=0.025$)] (**Fig.1A**).
577 TUNEL assay showed significantly more intense staining in the liver tissue of BDL
578 compared to Sham rats (**Fig.1A**) ($p<0.0001$), which was significantly reduced in Yaq-001-

579 treated BDL rats compared to untreated-BDL rats ($p=0.025$). Collagen proportionate area
580 (CPA) was significantly higher in BDL rats ($p=0.0007$), which was unchanged with Yaq-
581 001 ($p=0.122$) (**Fig.S6D**).

582

583 Effect of Yaq-001 on ammonia, organ dysfunction, endotoxemia and bacterial
584 translocation

585 *Ammonia*: Arterial and portal venous ammonia concentrations were significantly
586 increased in BDL rats ($p<0.0001$), which was significantly reduced by Yaq-001 [($p=0.003$)
587 and ($p=0.004$) respectively] (**Fig.1A**). None of the animals showed signs of overt hepatic
588 encephalopathy.

589

590 *Kidneys*: BDL animals had significantly higher plasma creatinine ($p=0.049$), which was
591 significantly reduced with Yaq-001 ($p=0.025$) (**Fig.1A**). Urea was higher in BDL group
592 ($p=0.092$), which was reduced with Yaq-001 treatment ($p=0.095$) (**Fig.1A**).

593

594 *Gut permeability, Endotoxemia, Bacterial DNA and Cytokines*: The microbial metabolite,
595 D-lactate, a marker of gut-specific intestinal barrier damage and translocation¹⁶ was
596 significantly increased in BDL rats ($p=0.032$) and was significantly reduced by Yaq-001
597 ($p=0.035$) (**Fig.1A**). BDL rats exhibited marked endotoxemia in the portal vein and the
598 artery ($p<0.0001$ for each), which was significantly reduced with Yaq-001 [($p<0.0001$)
599 ($p=0.003$) respectively] (**Fig.1A**). Portal venous bacterial DNA was detectable in
600 significantly higher number of BDL rats ($p<0.05$), which was markedly reduced in Yaq-
601 001 administered BDL rats ($p=0.08$) (**Fig.1A**). Plasma IL- β concentration were higher in
602 the BDL rats but no significant differences were observed in TNF- α , IL-6 and IL-10. No
603 significant changes were seen with Yaq-001 (**Table S1**).

604

605 **Studies in the BDL model of ACLF**

606 This experiment was performed to determine whether Yaq-001 treatment for 2-weeks
607 prevents the occurrence of ACLF when BDL animals are administered LPS (**Fig.S1**,
608 **Fig.1B**).

609

610 *Survival:* Animals were sacrificed either at coma stages (considered as a surrogate for
611 mortality) or at 6-hours post LPS. Yaq-001 significantly reduced time to coma of BDL-LPS
612 rats compared to untreated controls ($p < 0.01$) (**Fig.1B**). All animals in the two Sham
613 groups were alive at 6-hours following LPS (data not shown).

614

615 *Liver:* Yaq-001 was associated with significantly lower ALT in BDL-LPS rats compared to
616 untreated rats ($p = 0.004$) (**Fig.1B**). No significant effect of Yaq-001 was observed on ALP,
617 TBIL and albumin (**Fig.S7 A, B, C**). The severity of fibrosis measured using CPA and the
618 body weight were unchanged (**Fig.S7D, E**).

619

620 *Systemic and Portal hemodynamics:* No significant difference in MAP was observed
621 between the groups treated with or without Yaq-001 (**Fig.S7F**) but Yaq-001 produced a
622 significant reduction in portal pressure in BDL-LPS animals compared with the untreated
623 group ($p = 0.003$), (**Fig.1B**).

624

625 *Brain:* Yaq-001 significantly reduced brain water in BDL-LPS compared with the untreated
626 group ($p = 0.017$) (**Fig.1B**). Arterial and portal venous ammonia concentrations were
627 significantly increased in BDL-LPS rats, which was significantly reduced in Yaq-001-
628 treated animals [$(p = 0.007)$ and $(p = 0.017)$ respectively] (**Fig.1B**).

629

630 *Kidneys:* Creatinine concentrations were significantly higher in BDL-LPS animals
631 ($p = 0.004$), which was significantly reduced by Yaq-001 ($p = 0.03$) (**Fig.1B**).

632

633 *Cytokines:* BDL-LPS group had a significantly higher plasma IL-1 β , which was
634 significantly reduced with Yaq-001 ($p = 0.003$). Plasma IL-10 was higher in BDL-LPS and
635 was significantly reduced with Yaq-001 ($p = 0.028$) (**Fig.1B**). No significant differences
636 were observed in IL-6 or TNF- α concentrations between any of the groups (**Table S1**).

637

638 ***Effect of Yaq-001 on peripheral blood cells and Kupffer cells***

639 Significant increase in total leucocyte, neutrophil and monocyte counts in the artery and
640 portal vein were observed with BDL rats (**Fig.S8A, B**) ($p = 0.008$ and $p = 0.016$

641 respectively), which was significantly reduced with Yaq-001 in the arterial blood and
642 insignificantly reduced in the portal vein (**Fig. S8B**). To determine whether Yaq-001
643 impacts on the response of peripheral inflammatory cells and Kupffer cells to generate
644 reactive oxygen species (ROS) to LPS *ex vivo*, studies using isolated cells incubated with
645 LPS, were performed. Yaq-001 was associated with significantly lower LPS-induced ROS
646 production in CD163⁺ Kupffer cells in BDL rats ($p=0.036$) and portal venous CD43^{hi}
647 monocyte populations of BDL rats ($p=0.029$) (**Fig.S8C**).

648

649 ***Transcriptomic analysis of gene expression profiles from the Liver, Colon, Brain*** 650 ***and Kidneys***

651 Multiorgan transcriptomic analysis was performed to determine the possible molecular
652 mechanisms underlying the clinical effects of Yaq-001. The four groups studied were as
653 follows: Sham (n=3), Sham-Yaq-001 (n=3), BDL (n=3) and BDL-Yaq-001 (n=4) (**Fig.2A**).
654 All differentially expressed genes (DEGs) and related pathways in the liver, colon, kidney
655 and brain are listed in **Table S2**. The top 20 and significant DEGs are listed in **Table S3**.

656

657 Effect of Yaq-001 on gene expression profiles in the liver and gut in BDL rats

658 *Liver*: Analysis of liver tissue showed 82 DEGs at the threshold of 1.2-fold change and
659 $p=0.1$ in the four groups (**Fig.2B**). Compared with the Sham group, expression of 62-
660 genes was upregulated, and 15-genes were downregulated in BDL. These significantly
661 changed genes were associated with inflammation, cell death and senescence.
662 Compared to the untreated BDL group, the expression of 7-genes was upregulated and
663 12-genes were downregulated in the Yaq-001-treated BDL group, indicating the potential
664 role of Yaq-001 in reducing inflammation, cell death and cell senescence. Furthermore,
665 2-genes were upregulated, and 4-genes downregulated in Sham-Yaq-001 group in
666 comparison to Sham group (**Fig.2C**). Functional analysis demonstrated that BDL rats had
667 enriched pathways related to inflammation, cell senescence, cell death, TLR signaling
668 and other related signaling pathways in comparison with Sham (**Fig.S9A**). Yaq-001
669 treatment targeted the altered pathways compared with untreated BDL group. Additionally,
670 Yaq-001 treatment also changed the pathways in the liver when compared to Sham group,
671 demonstrating its effect in rats even without cirrhosis (**Fig.S9A**).

672
673 *Colon:* 43 DEGs were identified from the colonic tissue (**Fig.2D**). 5-genes that correlated
674 with inflammation and cell death were upregulated and 15-genes were downregulated in
675 BDL compared with the Sham group. Moreover, the expression of 10-genes was
676 upregulated, and 13-genes were downregulated with Yaq-001 treatment. Only 1-gene
677 was upregulated in the Sham-Yaq-001 group, and 16-genes were downregulated with
678 Yaq-001 compared with the untreated Sham group (**Fig.2E**). Functional analysis
679 indicated that inflammation, cell senescence, cell death, TLR signaling and intracellular
680 signaling were associated with BDL in comparison with the Sham group (**Fig.S9B**). Yaq-
681 001 targeted the altered pathways, indicating the potential mechanisms in the prevention
682 of gut dysfunction and permeability (**Fig.S9B**).

683

684 Effect of Yaq-001 on gene expression profiles in the brain and kidney in BDL rats

685 *Brain:* 17 DEGs were identified from the brain tissue (**Fig.2F**). Compared with Sham
686 group, expression of 2-genes was upregulated and 13-genes were downregulated in BDL
687 animals. These significantly changed genes were associated with inflammation, cell death,
688 and cell senescence. Compared to the untreated-BDL group, the expression of 5-genes
689 was upregulated and 2-genes were downregulated in the Yaq-001-treated BDL group
690 (**Fig.2G**). Functional analysis demonstrated that BDL rats had enriched pathways related
691 to inflammation, cell senescence, cell death, TLR signaling and intracellular signaling
692 (**Fig.S9C**). Yaq-001 targeted cytokine-cytokine receptor interaction, cytosolic DNA-
693 sensing pathway, TLR signaling pathway, NOD-like receptor signaling pathway,
694 neutrophil extracellular trap formation, TGF-beta signaling pathway and cytokine-cytokine
695 receptor interaction pathways compared to untreated-BDL group (**Fig.S9C**).

696

697 *Kidneys:* 30 DEGs were identified from kidney tissue (**Fig.2H**). 9-genes that correlated
698 with inflammation were downregulated in BDL. The expression of 5-genes was
699 upregulated and 4-genes were downregulated with Yaq-001 treatment compared to
700 untreated-BDL group. 5-genes were upregulated in Sham-Yaq-001 group, and 3-genes
701 were downregulated with Yaq-001 compared with untreated-Sham group (**Fig.2I**).
702 Functional analysis indicated that inflammation and TLR signaling were associated with

703 BDL in comparison with Sham (**Fig.S9D**). Compared with the untreated-BDL group, Yaq-
704 001 targeted the altered pathways, indicating the potential mechanisms in the prevention
705 of renal dysfunction (**Fig.S9D**).

706

707 ***Effect of Yaq-001 on the gut microbiome profile***

708 The effects of Yaq-001 on the microbiome bacterial composition was assessed by
709 metataxonomics. At the family level, an abundance of six bacteria were significantly
710 changed at the threshold of 2-fold change and *Porphyromonadaceae* was significantly
711 changed ($p < 0.05$) comparing BDL with Sham (**Fig.3A**). At genus level, 19 bacteria
712 including were significantly changed in abundance. *Barnesiella* was significantly changed
713 ($p < 0.05$) comparing BDL with Sham group (**Fig.3B**). These changes were reversed with
714 Yaq-001 treatment compared to untreated-BDL rats (**Fig.S10A, B, Table S4 and**
715 **Fig.S10C, D**). For between groups sample diversity, PERMANOVA analysis revealed a
716 significant difference in beta diversity between groups ($R^2 = 0.32$, $p = 0.001$). Yaq-001
717 appeared to moderately restore the beta diversity in the BDL group especially in PCoA2
718 axis (**Fig.S10E, F**).

719

720 To further investigate the potential importance of the changes in the microbiome induced
721 by Yaq-001, we correlated these with all significantly changed DEGs and the top 20 DEGs
722 in the four organs. Circos plots indicated a significant correlation between them (**Fig.3C,**
723 **D and Fig.S11A, B, C**). *Porphyromonadaceae*, was observed to positively correlate with
724 three DEGs - TGFB2 and CASP1 in liver tissue, and FOS in colonic tissue. Also, it
725 correlated negatively with five DEGs- TGFB2, IL-18 and CCR5 in brain tissue, CXCL10
726 in colon tissue and CCL24 in kidney tissue.

727

728 ***Effect of Yaq-001 on metabolomic profile***

729 Significant difference of acetate/creatinine, glycine/creatinine, lactate/creatinine,
730 betaine/creatinine, trimethylamine oxide/creatinine and bile acid/creatinine ratio were
731 observed in BDL compared to Sham. Treatment of BDL rats with Yaq-001 resulted in
732 significant resolution of acetate/creatinine, glycine/creatinine and lactate/creatinine
733 compared to the untreated BDL animals (**Fig.S12**).

734

735 **Studies in CCl₄ mice**

736 Effect of Yaq-001 on liver injury and fibrosis

737 6-week and 12-week CCl₄ mice models were used to further confirm the effect of Yaq-
738 001 in cirrhosis (**Fig.4A, B**). Yaq-001 was associated with a significantly lower plasma
739 ALT ($p < 0.0001$, $p < 0.0001$) in both 6-week and 12-week CCl₄ models. ALP and TBIL were
740 reduced by Yaq-001 in 6-week CCl₄ mice ($p = 0.040$, $p = 0.001$) (**Fig.4A, B**). CPA was
741 significantly higher in both CCl₄ mice compared with control animals ($p = 0.0001$,
742 $p = 0.0001$), which was significantly reduced with Yaq-001 ($p = 0.024$, $p = 0.012$) (**Fig.4A, B**).
743 TUNEL assay showed significantly more intense staining in the liver tissue of CCl₄
744 compared to Control mice ($p < 0.001$, $p < 0.001$), which was significantly reduced in Yaq-
745 001-treated CCl₄ mice compared to untreated-CCl₄ mice ($p = 0.021$, $p = 0.017$) (**Fig.4A, B**).
746

747 Effect of Yaq-001 on ammonia, organ dysfunction and endotoxemia

748 *Ammonia*: Ammonia concentrations were significantly increased in the 6-week and 12-
749 week CCl₄ mice compared with controls ($p = 0.002$, $p = 0.001$), which was significantly
750 reduced by Yaq-001 ($p = 0.025$, $p = 0.035$) (**Fig.4A, B**). None of the animals showed signs
751 of hepatic encephalopathy.

752

753 *Kidneys*: Higher plasma creatinine was significantly reduced by Yaq-001 treatment
754 ($p = 0.005$, $p = 0.003$) in 6-week and 12-week CCl₄ animals (**Fig.4A, B**).

755

756 *Endotoxemia*: Both 6-week and 12-week CCl₄ mice exhibited marked endotoxemia
757 compared with controls ($p = 0.007$, $p = 0.007$), which was significantly reduced with Yaq-
758 001 ($p = 0.007$, $p = 0.043$) (**Fig.4A, B**).

759

760 ***In vitro* studies in intestinal organoids to assess gut permeability.**

761 Intestinal organoids were successfully derived and cultured from small intestine of
762 C57BL/6 mice. Intestinal organoids underwent eversion into apical-out polarity in the first
763 12h of suspension culture and collected for identification and subsequent experiments
764 (**Fig.5A**). Immunostaining of the microvilli (mv; F-actin) demonstrated that intestinal

765 organoids in suspension had reversed polarity such that the apical surface faces outward
766 (**Fig.5A**). Apical-out intestinal organoids possessed goblet cells, which were identified
767 with MUC2 staining (**Fig.5B**). Gut permeability of apical-out intestinal organoids was
768 significantly increased by coculturing with fecal water from CCl₄ group compared with the
769 control group (p=0.003) (**Fig.5C,D**). The gut permeability was significantly decreased with
770 fecal water from Yaq-001 treated CCl₄ animals compared with the CCl₄ group (p=0.001)
771 (**Fig.5C,D**).

772

773 **CARBALIVE-SAFETY Clinical Trial**

774 The data regarding safety and tolerability are reported here. Other secondary and
775 exploratory end points will be described elsewhere.

776

777 ***Patient Characteristics***

778 Thirty-four-patients were screened for this study at 8-European centers. 28-patients met
779 the study entry criteria and were randomized to either active or placebo groups. Six-
780 patients screened did not meet the study entry criteria. Dosing was not initiated in 2-
781 patients randomized to placebo due to withdrawal of consent (**Fig.S3, CONSORT**). 3-
782 patients were included for the second dosing cohort of 8 g. This part of the study was
783 terminated prematurely due to the coronavirus pandemic with none of the patients
784 completing the study duration (data not included).

785

786 In accordance with study entry criteria, all patients had cirrhosis with diuretic-responsive
787 ascites and Child-Pugh score of 7-8. The baseline demographics were similar across
788 treatment groups. The ratio of male patients to female patients was reflective of the
789 disease state. Compliance in the active and placebo groups was 92.9% and 66.7%
790 respectively (**Table 1**).

791

792 ***Safety and Tolerability***

793 Of the 14-patients enrolled in the Yaq-001 treatment group, 13 (93%) completed 12-
794 weeks of therapy. The median duration of exposure was 83 (6-94) days. Ten of the 12
795 (83%) patients who received placebo completed the treatment. The median duration of

796 exposure was 83 (14-86) days. No deaths or serious adverse events were reported in the
797 study. The difference in treatment-emergent adverse events (TEAEs) in patients treated
798 with Yaq-001 and those treated with placebo is presented in **Table 2**. The most frequently
799 reported TEAEs were gastrointestinal in nature in both the active and placebo groups. Of
800 these, only constipation and diarrhea were evaluated by the clinical investigator as
801 possibly related to the investigational product. One placebo-treated patient withdrew from
802 the study due to diarrhea.

803

804 Across both treatment groups, 40/51(78%) of the reported TEAEs were evaluated by the
805 clinical investigator as not related or unlikely related (32/38; 84% for the active treatment
806 group; 8/13; 62% for the placebo group). The incidence of adverse events reported is
807 reflective of the targeted subject population for this clinical investigation. The majority of
808 the TEAEs reported were not considered by the clinical investigator to be related to
809 treatment and were mild in intensity. Systemic antibiotics were administered for the
810 following TEAEs in the active arm: Amoxicillin – acute bronchitis; Clarithromycin – acute
811 bronchitis; Phosphomycin – urinary tract infection. None of these infections were related
812 to the administration of the investigational product. Drugs received by the patients at the
813 time of randomization and during follow up are listed in Supplementary Tables.
814 Treatment-emergent, clinically significant laboratory abnormalities are listed in **Table 3**.
815 None were deemed treatment-related by the investigator.

816

817 ***Clinical, hematological and biochemical variables***

818 The data are summarized in **Table 3**. No significant changes in any of the clinical
819 parameters were observed in any of the groups. Although there was a trend towards a
820 reduction in the white cell count and C-reactive protein in the Yaq-001 group, the
821 differences were not significant.

822

823 ***Nutritional status***

824 No significant differences were observed in either treatment group with regards to global
825 nutritional status, vitamin B12 and folate, Vitamin A or E, or copper zinc, and selenium.
826 Median vitamin A, zinc and baseline vitamin D concentrations were below the limit of

827 normal range but no differences between treatment groups were observed. No changes
828 were observed in any of the micronutrient parameters with treatment and these were
829 evenly matched between groups. Any baseline abnormalities were attributable to the
830 underlying natural history of cirrhosis.

831

832

833

834 **Discussion**

835 The results of the study showed that Yaq-001 prevented progression of liver injury and
836 fibrosis in animal models of cirrhosis and significantly reduced the mortality of ACLF
837 animals. This was associated with positive impact on markers of gut permeability, liver
838 injury, portal pressure, brain and kidneys. These pleiotropic effects of Yaq-001 were
839 associated with partial restoration of the composition of the microbiome bacterial
840 community, reduction in the severity of endotoxemia, ammonia, inflammation, cell death,
841 signaling pathways and LPS sensitivity. A Phase 2 equivalent, double-blind, multicenter,
842 placebo-controlled, randomized clinical trial in patients with cirrhosis confirmed regulatory
843 compliance and, safety and tolerability of Yaq-001, thereby, providing evidence of clinical
844 translatability. The data provide the rationale to proceed to further clinical trials.

845
846 Translocation of bacteria, its products and metabolites are critically important in the
847 progression of hepatic fibrosis and pathogenesis of complications of cirrhosis^{1,17-20}.
848 Indeed, selective gut decontamination using norfloxacin or rifaximin are the current
849 standard of care for secondary prophylaxis of patients with spontaneous bacterial
850 peritonitis and hepatic encephalopathy respectively²¹⁻²². However, the use of these
851 antibiotic strategies is limited to patients with advanced cirrhosis and induces the risk of
852 antibiotic resistance.²³ The data presented here provide a safe, gut-restricted, non-
853 antibiotic strategy, Yaq-001, which has the potential to diminish translocation and prevent
854 the progression of hepatic injury, fibrosis and, prevent extrahepatic organ injury in models
855 of cirrhosis. The *in vitro* studies demonstrate that Yaq-001 has the optimal pore size
856 distribution to bind intraluminal factors such as free endotoxin. We also tested *in vitro*
857 bacterial growth kinetics of two species, which were not affected by Yaq-001, an
858 observation that was subsequently confirmed with studies in the BDL animal model where
859 no change diversity were observed.

860
861 Endotoxemia has also been implicated in immune dysfunction resulting in a dysregulated
862 systemic inflammatory response, which is strongly associated with the progression of
863 fibrosis, cirrhosis and occurrence of ACLF^{24,25}. Yaq-001 reduced the severity of
864 endotoxemia and bacterial DNA positivity, which was associated with attenuated systemic

865 inflammation. Significant improvements in LPS-induced ROS production were observed
866 in trafficking portal venous monocytes suggesting that Yaq-001 had attenuated the
867 primed state of monocyte/macrophage populations within the gut-liver axis. This observed
868 reduction in LPS-induced ROS production may be important in explaining the reduction
869 in plasma IL-1 β in LPS-treated BDL rats.

870

871 Plasma D-lactate, a marker of increased gut permeability was reduced in the Yaq-001
872 treated BDL rats²⁶. Elevated plasma D-lactate levels in cirrhosis is associated with
873 decompensation²². Transcriptomic analysis of colonic tissue demonstrated upregulation
874 of genes associated with necroptosis, apoptosis and inflammation in BDL animals.
875 Functional analyses pointed to modulation of colonic inflammation by Yaq-001, IL-17
876 signaling, which is known to have diverse biological functions, promoting protective
877 immunity against many pathogens, neutrophil recruitment, antimicrobial peptide
878 production and enhanced barrier function^{27, 28}. To further validate the potential effect of
879 Yaq-001 in modulating gut permeability, we performed experiments in intestinal organoids
880 that were incubated with fecal water²⁹. The data confirmed that even in *in vitro* settings,
881 fecal water obtained from the feces of CCL4-induced cirrhosis animals enhanced
882 permeability of the organoids, which was prevented in the fecal water obtained from
883 animals that were treated with Yaq-001. The data support the hypothesis that Yaq-001
884 impacts on the factors in the gut responsible for increasing gut permeability in cirrhosis.

885

886 Yaq-001 significantly reduced the severity of liver injury and portal hypertension in both
887 the BDL and BDL-LPS models of cirrhosis and ACLF. The lack of significant differences
888 in CPA between untreated and Yaq-001-treated BDL groups suggests that the reduction
889 in portal pressure is possibly due to modulation of the dynamic component of portal
890 hypertension, in which inflammation is known to play a role^{30,31} and proposes Yaq-001 as
891 a potential treatment for portal hypertension. Reduction in ALT levels and TUNEL staining
892 confirmed a reduction in liver injury in the Yaq-001 treated animals. The reduction in liver
893 injury in the LPS treated BDL animals suggests that Yaq-001 has a particular effect on
894 endotoxin sensitivity *in vivo*. This hypothesis was tested in isolated Kupffer cells, which

895 confirmed that LPS-induced ROS production was significantly impacted by Yaq-001
896 treatment.

897

898 Transcriptomic analysis of liver tissue demonstrated that the upregulated genes,
899 CXCL16, CASP1 and TGFB2 in BDL rats was prevented by Yaq-001 administration.
900 Silencing of CXCL16 alleviates hepatic ischemia reperfusion injury and CXCL16 variant
901 is also associated with Hepatitis B virus related acute liver failure³². CASP1 mediates pro-
902 inflammatory cytokine release and pyroptotic cell death in cirrhosis and its inhibition has
903 been shown to prevent ACLF³³. TGFB2 is an important mediator of cellular senescence³⁴,
904 ³⁵. Of note, Yaq-001 also modified necroptosis and cytosolic DNA-sensing pathways
905 representing cell death, which are known to be activated by LPS that can trigger systemic
906 inflammation³⁶. These effects of Yaq-001 potentially explains the effect of Yaq-001 in
907 reducing liver injury^{37, 38}.

908

909 Yaq-001 administration had a significant impact on time to coma of ACLF rats, which is
910 considered as a surrogate for mortality compared to untreated controls. Yaq-001 also
911 significantly lowered portal venous and arterial ammonia levels, which was associated
912 with reduced brain water. Transcriptomic analysis of brain tissue showed that IL-18,
913 TGFB2, CCR5 and IL-23A were dysregulated in BDL rats and these were corrected by
914 Yaq-001. IL-18 is released during pyroptosis by activation of the inflammasome complex
915 in neuroinflammatory and neurodegenerative diseases³⁹. The effect of Yaq-001 on
916 TGFB2 may mean that it has an impact on senescence, which is known to be associated
917 with hepatic encephalopathy. CCR5 has been implicated in neuroprotection and is novel
918 therapeutic target in stroke⁴⁰. The impact of Yaq-001 on IL-23A indicates possible
919 reduction in neuroinflammation.

920

921 In both cirrhosis and ACLF models, Yaq-001 reduced renal dysfunction. Transcriptomic
922 analysis of kidney tissue showed that CCL24 was downregulated in BDL rats, which was
923 prevented in the Yaq-001-treated animals. CCL24 protects renal function in the
924 development of early diabetic nephropathy by exerting an anti-inflammatory effect⁴¹. Yaq-

925 001 impacted, on the cytokine-cytokine receptor interactions and chemokine and toll-like
926 signaling pathways, which were abnormal in the BDL rats.

927

928 BDL animals become sarcopenic and lose weight⁴², which was significantly abrogated by
929 Yaq-001. The possible mechanisms underlying this effect are likely multifactorial⁴³. Yaq-
930 001 reduced ammonia significantly, which has been shown to induce sarcopenia⁴⁴.
931 Weight loss in cirrhosis is also attributed to an increased catabolic state in the context of
932 systemic inflammatory response and thus the observed improvement in body weight may
933 reflect the diminished catabolic state with reduced inflammation⁴³.

934

935 The clinical effects of Yaq-001 observed in the BDL models was validated in the CCL4
936 induced liver injury animal models. Two models were studied. In the first (6-week model),
937 Yaq-001 was administered in a preventative mode starting its administration with the
938 onset of liver injury during parenteral administration of CCL4. The results showed
939 significant reduction in the severity of liver injury, fibrosis and progression to cirrhosis,
940 endotoxemia, creatinine and ammonia levels. In the second (12-week model), Yaq-001
941 was administered starting at 6-weeks when the animal already had advanced fibrosis/
942 cirrhosis. Again, significant reduction in markers of liver injury, fibrosis, endotoxemia,
943 creatinine and ammonia were observed. Extrapolating these observations to the humans,
944 the results from the 6-week model suggests that Yaq-001 may well be useful to prevent
945 the progression of fibrosis in patients without cirrhosis and, from the 12-week model, the
946 possibility of prevention of progression of liver disease in those with well compensated
947 cirrhosis.

948

949 Gut microbiota are important in modulating intestinal health, permeability, bacterial
950 translocation, systemic inflammation and complications of cirrhosis⁴⁵⁻⁴⁷. BDL was
951 associated with marked changes in the abundance of microbiota, which were reversed
952 by Yaq-001. In particular, the abundance of *Porphyromonadaceae* and *Barnesiella* were
953 significantly elevated in BDL rats and significantly decreased with Yaq-001. This change
954 is potentially important as *Porphyromonadaceae* is a pro-inflammatory bacterium that has
955 been positively correlated with hepatic encephalopathy⁴⁸ and, *Barnesiella* and

956 *Porphyromonadaceae* have been associated with liver cancer^{49,50}. Urinary NMR analysis
957 reflects the combined metabolic status of both host and microbiota. Yaq-001 was
958 associated with a distinct shift of acetate, glycine and lactate in metabolomic profile in
959 BDL rats. These metabolites are generated by mixed acid fermentation (MAF), typically
960 by bacteria such as *Enterobacter*. MAF is not the preferred metabolic pathway for
961 facultative anaerobes and may be indicative that *Enterobacter* populations are under
962 conditions of metabolic stress in Yaq-001 treated BDL animals. As these species are
963 often pathogenic in cirrhosis, this may represent a beneficial change. However, the exact
964 mechanisms by which the change in the microbiome results in improvement in distant
965 organ function and gene expression cannot be directly inferred from the data derived from
966 this study. One possibility is that alongside LPS adsorption and modulation of other
967 unmeasured toxins, the *milieu* of the gut is changed allowing proliferation of more
968 autochthonous bacteria⁴⁷, which impacts on gut inflammation that reduces gut
969 permeability. This hypothesis is supported by the organoid experiments. Reduction in
970 permeability would result in a reduction in endotoxemia, systemic inflammation,
971 improvement of organ function and LPS-sensitivity. In this study, most of these changes
972 have been described individually but whether this is happening in sequence has not been
973 investigated.

974

975 As Yaq-001 is completely excreted unchanged in the stool, it is regulated in Europe as a
976 device but as a drug in the US, the clinical trial was performed both according to ISO
977 standards and ICH-GCP guidance. The results of this first-in-man randomized, placebo-
978 controlled trial suggested that oral Yaq-001 at a dose of 4g nocte was well tolerated with
979 a favorable safety profile. Despite the rapid adsorption kinetics for bacterial toxins and
980 metabolites, Yaq-001 treatment had no negative impact on micronutrient levels or impact
981 on nutritional profile as assessed by the gold standard Royal Free Global Assessment
982 tool. This data must be interpreted keeping in mind that Yaq-001 was administered post
983 prandially, separated from meals by 4-hours and from drugs by about 6-hours as
984 necessitated by the protocol. It is important to note that the studies were performed in
985 stable cirrhosis patients, many of whom had minimal evidence of systemic inflammation
986 and therefore, any clinical effect of this intervention was difficult to gauge. However, future

987 analysis of the available samples from the blood and stool will provide answers as to
988 whether Yaq-001 modulates the gut microbiome, inflammation and endotoxemia.

989
990 These results must be considered in view of some limitations. First, the rodent
991 microbiome is not directly analogous to the human and further clinical studies will be
992 required to verify the effects on the gut microbiome's bacterial composition. Second,
993 although Yaq-001 was effective in adsorbing a variety of bile acids *in vitro* and reduced
994 bile acids significantly in Sham animals, no impact on bile acids was seen in BDL animals.
995 This possibly reflects the effect of the BDL model, where no increase in bile acids was
996 observed. Also, no changes in bile acids were observed in CCL4 animals but these
997 animals did not have elevated bile acid either. Third, although, Yaq-001 was observed to
998 impact positively on the gene expression profiles of multiple pathways, their exact
999 relevance at the protein or cellular level has not been explored. Fourth, as only one dose
1000 of Yaq-001 was tested in the clinical trial, further dose-ranging studies will be needed to
1001 define optimal dosing for safety and efficacy. However, the animal toxicity studies that
1002 were performed by an independent laboratory for regulatory purposes, showed evidence
1003 of safety in much larger doses than that administered in the present studies (summary in
1004 Supplementary).

1005
1006 In conclusion, the data provides compelling evidence for the potential of Yaq-001 as a
1007 novel therapy targeting the gut microbiome, bacterial translocation and gut permeability
1008 that impacts on systemic inflammation, liver injury and fibrosis and, organ function in
1009 models of cirrhosis and improves survival in ACLF. The placebo-controlled clinical trial of
1010 Yaq-001 in cirrhosis patients provides evidence of safety and tolerability allowing
1011 translation to next phase of clinical studies to define its potential as a novel therapeutic
1012 for patients with cirrhosis.

1013

1014 **Abbreviations**

1015 ACLF, acute-on-chronic liver failure; LPS, lipopolysaccharides; BDL, bile duct ligation;
1016 ALT, alanine aminotransferase; ALP, alkaline phosphatase; TBIL, total bilirubin; MAP,
1017 mean arterial pressure; CPA, collagen proportionate area; PSR, picosirus red ; PP, portal
1018 pressure; ROS, reactive oxidant species; DEGs, differential expressed genes; KEGG,
1019 Kyoto Encyclopedia of Genes and Genomes; TLR, toll-like receptor; TNF-a, tumor
1020 necrosis factor-a.

1021

1022 **Acknowledgements**

1023 We would like to thank Fraser Simpson (Department of Genetics, Evolution &
1024 Environment, University College London) for his support to perform the Nanostring. The
1025 urinary NMR studies were facilitated by a Medical Research Council research grant under
1026 the High-throughput "omic" Science and Imaging funding scheme [MC_PC_13045].

1027

1028

1029 **References**

- 1030 [1] Engelmann C, Adebayo D, Oria M, De Chiara F, Novelli S, Habtesion A et al.
1031 Recombinant alkaline phosphatase prevents acute on chronic liver failure. *Sci Rep*
1032 2020; 10: 389.
- 1033 [2] Michelena J, Alonso C, Martinez-Arranz I, Altamirano J, Mayo R, Sancho-Bru P et al.
1034 Metabolomics discloses a new non-invasive method for the diagnosis and prognosis
1035 of patients with alcoholic hepatitis. *Ann Hepatol* 2019; 18: 144-154.
- 1036 [3] Engelmann C, Sheikh M, Sharma S, Kondo T, Loeffler-Wirth H, Zheng YB et al. Toll-
1037 like receptor 4 is a therapeutic target for prevention and treatment of liver failure. *J*
1038 *Hepatol* 2020; 73: 102-112.
- 1039 [4] Moreau R, Claria J, Aguilar F, Fenaille F, Lozano JJ, Junot C et al. Blood metabolomics
1040 uncovers inflammation-associated mitochondrial dysfunction as a potential
1041 mechanism underlying ACLF. *J Hepatol* 2020; 72: 688-701.
- 1042 [5] Albillos A, Martin-Mateos R, Van der Merwe S, Wiest R, Jalan R, Alvarez-Mon M.
1043 Cirrhosis-associated immune dysfunction. *Nat Rev Gastroenterol Hepatol* 2022; 19:
1044 112-134.
- 1045 [6] Takaya H, Namisaki T, Sato S, Kaji K, Tsuji Y, Kaya D et al. Increased endotoxin
1046 activity is associated with the risk of developing acute-on-chronic liver failure. *J Clin*
1047 *Med* 2020; 9.
- 1048 [7] Bajaj JS, Thacker LR, Fagan A, White MB, Gavis EA, Hylemon PB et al. Gut microbial
1049 RNA and DNA analysis predicts hospitalizations in cirrhosis. *JCI Insight* 2018; 3.
- 1050 [8] Fernandez J, Prado V, Trebicka J, Amoros A, Gustot T, Wiest R et al. Multidrug-
1051 resistant bacterial infections in patients with decompensated cirrhosis and with acute-
1052 on-chronic liver failure in Europe. *J Hepatol* 2019; 70: 398-411.
- 1053 [9] Piano S, Singh V, Caraceni P, Maiwall R, Alessandria C, Fernandez J et al.
1054 Epidemiology and effects of bacterial infections in patients with cirrhosis worldwide.
1055 *Gastroenterology* 2019; 156: 1368-1380 e1310.
- 1056 [10] Shah N, Mohamed FE, Jover-Cobos M, Macnaughtan J, Davies N, Moreau R et al.
1057 Increased renal expression and urinary excretion of TLR4 in acute kidney injury
1058 associated with cirrhosis. *Liver Int* 2013; 33: 398-409.

- 1059 [11] Macnaughtan J, Ranchal I , Soeda J, Sawhney R, Oben J, Davies N et al. O091:
1060 Oral therapy with non-absorbable carbons of controlled porosity (YAQ-001)
1061 selectively modulates stool microbiome and its function and this is associated with
1062 restoration of immune function and infammasome activation. *J Hepatol*
1063 2015;62:S240.
- 1064 [12] Macnaughtan J, Ranchal I, Soeda J, Sawhney R, Oben J, Davies N et al. PTH-095
1065 Oral carbon therapy is associated with a selective modulation of the microbiome in
1066 cirrhotic rats which is associated with a significant reduction in inflammatory
1067 activation. *Gut* 2015;64:A449-A450.
- 1068 [13] Macnaughtan J, Albillos A, Kerbert A, Vargas V, Durand F, Gine P et al. O09 A double
1069 blind, randomised, placebo-controlled study to assess safety and tolerability of oral
1070 enterosorbent Carbalive (Yaq-001) in cirrhotic patients. *Gut* 2021;70:A5-A6.
- 1071 [14] Bajaj JS, Sheikh MY, Chojkier M, Balart L, Sherker AH, Vemuru R et al. AST-120
1072 (spherical carbon adsorbent) in covert hepatic encephalopathy: results of the
1073 ASTUTE trial. *J Hepatol* 2013;58:S84.
- 1074 [15] den Daas SA, Soffientini U, Chokshi S, Mehta G. A permeability assay for mouse
1075 intestinal organoids. *STAR Protoc* 2022;3:101365[16] Riva A, Gray EH, Azarian S,
1076 Zamalloa A, McPhail MJW, Vincent RP *et al.* Faecal cytokine profiling as a marker
1077 of intestinal inflammation in acutely decompensated cirrhosis. *JHEP Rep* 2020; 2:
1078 100151.
- 1079 [17] Jalan R, Fernandez J, Wiest R, Schnabl B, Moreau R, Angeli P et al. Bacterial
1080 infections in cirrhosis: a position statement based on the EASL Special Conference
1081 2013. *J Hepatol* 2014; 60: 1310-1324.
- 1082 [18] Borzio M, Salerno F, Piantoni L, Cazzaniga M, Angeli P, Bissoli F et al. Bacterial
1083 infection in patients with advanced cirrhosis: a multicentre prospective study. *Dig*
1084 *Liver Dis* 2001; 33: 41-48.
- 1085 [19] Wong F, Piano S, Singh V, Bartoletti M, Maiwall R, Alessandria C et al. Clinical
1086 features and evolution of bacterial infection-related acute-on-chronic liver failure. *J*
1087 *Hepatol* 2021; 74: 330-339.

- 1088 [20] Engemann C, Habtesion A, Hassan M, Kerbert AJ, Hammerich L, Novelli S et al.
1089 Combination of G-CSF and a TLR4 inhibitor reduce inflammation and promote
1090 regeneration in a mouse model of ACLF. *J Hepatol* 2022; 77: 1325-1338.
- 1091 [21] Praharaj DL, Premkumar M, Roy A, Verma N, Taneja S, Duseja A et al. Rifaximin vs.
1092 norfloxacin for spontaneous bacterial peritonitis prophylaxis: a randomized controlled
1093 trial. *J Clin Exp Hepatol* 2022; 12: 336-342.
- 1094 [22] Patel VC, Lee S, McPhail MJW, Da Silva K, Guilly S, Zamalloa A et al. Rifaximin-
1095 alpha reduces gut-derived inflammation and mucin degradation in cirrhosis and
1096 encephalopathy: RIFSYS randomised controlled trial. *J Hepatol* 2022; 76: 332-342.
- 1097 [23] Shenep JL, Barton RP, Mogan KA. Role of antibiotic class in the rate of liberation
1098 of endotoxin during therapy for experimental gram-negative bacterial sepsis. *J Infect*
1099 *Dis* 1985; 151: 1012-1018.
- 1100 [24] Wasmuth HE, Kunz D, Yagmur E, Timmer-Stranghoner A, Vidacek D, Siewert E et
1101 al. Patients with acute on chronic liver failure display "sepsis-like" immune paralysis.
1102 *J Hepatol* 2005; 42: 195-201.
- 1103 [25] Scarpellini E, Abenavoli L, Cassano V, Rinninella E, Sorge M, Capretti F, Rasetti C,
1104 et al. The Apparent Asymmetrical Relationship Between Small Bowel Bacterial
1105 Overgrowth, Endotoxemia, and Liver Steatosis and Fibrosis in Cirrhotic and Non-
1106 Cirrhotic Patients: A Single-Center Pilot Study. *Front Med (Lausanne)*
1107 2022;9:872428.
- 1108 [26] Grootjans J, Thuijls G, Verdam F, Derikx JP, Lenaerts K, Buurman WA. Non-invasive
1109 assessment of barrier integrity and uncton of the human gut. *World J Gastrointest*
1110 *Surg* 2010; 2: 61-69.
- 1111 [27] Mills KHG. IL-17 and IL-17-producing cells in protection versus pathology. *Nat Rev*
1112 *Immunol* 2023; 23: 38-54.
- 1113 [28] He S, Cui S, Song W, Jiang Y, Chen H, Liao D et al. Interleukin-17 weakens the
1114 NAFLD/NASH process by facilitating intestinal barrier restoration depending on the
1115 gut microbiota. *mBio* 2022; 13: e0368821.
- 1116 [29] Puschhof J, Pleguezuelos-Manzano C, Martinez-Silgado A, Akkerman N, Saftien A,
1117 Boot C, de Waal A, et al. Intestinal organoid cocultures with microbes. *Nat Protoc*
1118 2021;16:4633-4649.

- 1119 [30] Mookerjee RP, Sen S, Davies NA, Hodges SJ, Williams R, Jalan R. Tumour
1120 necrosis factor alpha is an important mediator of portal and systemic haemodynamic
1121 derangements in alcoholic hepatitis. *Gut* 2003; 52: 1182-1187.
- 1122 [31] Mehta G, Gustot T, Mookerjee RP, Garcia-Pagan JC, Fallon MB, Shah VH *et al.*
1123 Inflammation and portal hypertension - the undiscovered country. *J Hepatol* 2014;
1124 61: 155-163.
- 1125 [32] Ajmera V, Huang H, Dao D, Feld JJ, Lau DT, Patel K *et al.* Host genetic variant in
1126 CXCL16 may be associated with hepatitis B virus-related acute liver failure. *Cell Mol*
1127 *Gastroenterol Hepatol* 2019; 7: 477-479 e474.
- 1128 [33] Kondo T, Macdonald S, Engelmann C, Habtesion A, Macnaughtan J, Mehta G *et al.*
1129 The role of RIPK1 mediated cell death in acute on chronic liver failure. *Cell Death*
1130 *Dis* 2021; 13: 5.
- 1131 [34] Tominaga K, Suzuki HI. TGF-beta signaling in cellular senescence and aging-related
1132 pathology. *Int J Mol Sci* 2019; 20.
- 1133 [35] Dewidar B, Meyer C, Dooley S, Meindl-Beinker AN. TGF-beta in hepatic stellate cell
1134 activation and liver fibrogenesis-updated 2019. *Cells* 2019; 8.
- 1135 [36] Bertheloot D, Latz E, Franklin BS. Necroptosis, pyroptosis and apoptosis: an intricate
1136 game of cell death. *Cell Mol Immunol* 2021; 18: 1106-1121.
- 1137 [37] Kondo T, Macdonald S, Engelmann C, Habtesion A, Macnaughtan J, Mehta G *et al.*
1138 The role of RIPK1 mediated cell death in acute on chronic liver failure. *Cell Death*
1139 *Dis* 2021; 13: 5.
- 1140 [38] Soffientini U, Beaton N, Baweja S, Weiss E, Bihari C, Habtesion A *et al.* The
1141 lipopolysaccharide-sensing caspase(s)-4/11 are activated in cirrhosis and are
1142 causally associated with progression to multi-organ injury. *Front Cell Dev Biol* 2021;
1143 9: 668459.
- 1144 [39] Voet S, Srinivasan S, Lamkanfi M, van Loo G. Inflammasomes in neuroinflammatory
1145 and neurodegenerative diseases. *EMBO Mol Med* 2019; 11.
- 1146 [40] Joy MT, Ben Assayag E, Shabashov-Stone D, Liraz-Zaltsman S, Mazzitelli J, Arenas
1147 M *et al.* CCR5 is a therapeutic target for recovery after stroke and traumatic brain
1148 injury. *Cell* 2019; 176: 1143-1157 e1113.

- 1149 [41] Wang Y, Wu X, Geng M, Ding J, Lv K, Du H et al. CCL24 protects renal function by
1150 controlling inflammation in podocytes. *Dis Markers* 2021; 2021: 8837825.
- 1151 [42] Rosa CGS, Colares JR, da Fonseca SRB, Martins GDS, Miguel FM, Dias AS et al.
1152 Sarcopenia, oxidative stress and inflammatory process in muscle of cirrhotic rats -
1153 action of melatonin and physical exercise. *Exp Mol Pathol* 2021; 121: 104662.
- 1154 [43] Ebadi M, Burra P, Zanetto A, Montano-Loza AJ. Current treatment strategies and
1155 future possibilities for sarcopenia in cirrhosis. *J Hepatol* 2023; 78: 889-892.
- 1156 [44] Lee PC, Lee KC, Yang TC, Lu HS, Cheng TY, Chen YJ, Chiou JJ, et al. Sarcopenia-
1157 related gut microbial changes are associated with the risk of complications in people
1158 with cirrhosis. *JHEP Rep* 2023;5:100619.
- 1159 [45] Trebicka J, Bork P, Krag A, Arumugam M. Utilizing the gut microbiome in
1160 decompensated cirrhosis and acute-on-chronic liver failure. *Nat Rev Gastroenterol*
1161 *Hepatol* 2021; 18: 167-180.
- 1162 [46] Tilg H, Cani PD, Mayer EA. Gut microbiome and liver diseases. *Gut* 2016; 65: 2035-
1163 2044.
- 1164 [47] Albillos A, de Gottardi A, Rescigno M. The gut-liver axis in liver disease:
1165 Pathophysiological basis for therapy. *J Hepatol* 2020; 72: 558-577.
- 1166 [48] Ahluwalia V, Betrapally NS, Hylemon PB, White MB, Gillevet PM, Unser AB et al.
1167 Impaired gut-liver-brain axis in patients with cirrhosis. *Sci Rep* 2016; 6: 26800.
- 1168 [49] Jiang N, Song X, Peng YM, Wang WN, Song Z. Association of disease condition with
1169 changes in intestinal flora, and plasma endotoxin and vascular endothelial growth
1170 factor levels in patients with liver cancer. *Eur Rev Med Pharmacol Sci* 2020; 24:
1171 3605-3613.
- 1172 [50] Ma J, Li J, Jin C, Yang J, Zheng C, Chen K et al. Association of gut microbiome and
1173 primary liver cancer: a two-sample Mendelian randomization and case-control study.
1174 *Liver Int* 2023; 43: 221-233.
- 1175

1176 **Figure legends**

1177 **Fig.1. Effect of Yaq-001 on organ dysfunction, endotoxemia and bacterial**
1178 **translocation in BDL and ACLF rats.**

1179 (A) Rats underwent bile duct ligation for 4-weeks as a model of cirrhosis (n=23-37/group).
1180 Treatment groups received Yaq-001 for 2 weeks before sacrifice.

1181 *4-week body weight in four groups:* Sham (n=31), Sham-Yaq-001 (n=24), BDL (n=31)
1182 and BDL-Yaq-001 (n=38). Significantly lower final body weights were observed in BDL
1183 compared to Sham controls (p<0.001). Yaq-001-treated BDL rats had a significantly
1184 higher body weights compared to untreated-BDL rats (p<0.05).

1185 *Plasma alanine transaminase (ALT) concentrations* in Sham (n=17), Sham-Yaq-001
1186 (n=14), BDL (n=17) and BDL-Yaq-001 (n=26) groups and *Portal pressure (PP)*
1187 *measurements in Sham (n=17), Sham-Yaq-001 (n=19), BDL (n=14) and BDL-Yaq-001*
1188 *(n=26) groups.* Significantly higher ALT and PP were observed in BDL compared to Sham
1189 controls (p<0.0001). Yaq-001-treated BDL rats had a significantly lower ALT and PP
1190 compared to untreated-BDL rats (p<0.01, p<0.05).

1191 *TUNEL assay of liver tissue with quantification of staining by digital image analysis.*
1192 Significantly higher TUNEL staining was observed in BDL compared to Sham controls
1193 (p<0.0001). Yaq-001-treated BDL rats had a significantly lower TUNEL staining
1194 compared to untreated-BDL rats (p<0.05) indicative of a reduction in liver cell death with
1195 Yaq-001 treatment.

1196 *Arterial ammonia concentrations* in Sham (n=7), Sham-Yaq-001 (n=5), BDL (n=19), BDL-
1197 Yaq-001 (n=21) groups and *Portal venous ammonia concentrations* in Sham (n=6),
1198 Sham-Yaq-001 (n=5), BDL (n=13), BDL-Yaq-001 (n=18) groups. Significantly increased
1199 arterial ammonia concentrations and portal venous ammonia concentrations were
1200 observed in BDL compared to Sham controls (p<0.0001, p=0.0001). Yaq-001 significantly
1201 decreased arterial and portal venous ammonia concentrations in BDL rats (p<0.01 for
1202 both).

1203 *Serum creatinine* in Sham (n=19), Sham-Yaq-001 (n=17), BDL (n=20), BDL-Yaq-001
1204 (n=17) and *serum urea* in Sham (n=28), Sham-Yaq-001 (n=23), BDL (n=30), BDL-Yaq-
1205 001 (n=34) groups. Yaq-001 markedly decreased serum creatinine levels in BDL rats
1206 (p<0.05).

1207 *Plasma D-lactate* in Sham (n=7), Sham-Yaq-001 (n=8), BDL (n=6), BDL-Yaq-001 (n=7).
1208 Plasma D-lactate was significantly increased in the BDL group compared with Sham
1209 animals ($p<0.05$). Yaq-001 resulted in a significant reduction in plasma D-lactate in BDL
1210 rats ($p<0.05$).

1211 *Portal venous endotoxin* [Sham (n=6), Sham-Yaq-001 (n=5), BDL (n=12) and BDL-Yaq-
1212 001 (n=7)] and *arterial endotoxin* [Sham (n=6), Sham-Yaq-001 (n=5), BDL (n=12) and
1213 BDL-Yaq-001 (n=7)].

1214 *Portal venous bacterial DNA positivity* [Sham (n=6), Sham-Yaq-001 (n=5), BDL (n=12)
1215 and BDL-Yaq-001 (n=13)] and *arterial plasma bacterial DNA positivity* [Sham (n=6),
1216 Sham-Yaq-001 (n=6), BDL (n=12) and BDL-Yaq-001 (n=7)]. Significantly higher portal
1217 venous endotoxin and arterial endotoxin were observed in BDL rats compared to Sham
1218 rats ($p<0.0001$). Significantly higher portal venous plasma bacterial DNA positivity was
1219 observed in BDL rats compared to Sham rats ($p<0.05$). Yaq-001 administration was
1220 associated with a significant reduction of portal venous and arterial endotoxin compared
1221 to untreated-BDL rats ($p<0.0001$, $p<0.01$). Yaq-001 administration reduced bacterial DNA
1222 positivity, which was not statistically different ($p>0.05$).

1223

1224 (B) Rats underwent sham biliary surgery or BDL for 4-weeks. The treated group received
1225 Yaq-001 for 2 weeks prior to LPS injection. Animals were sacrificed either at coma stages
1226 or 6 hours after LPS injection (n=9-16/group).

1227 *Kaplan-Meier analysis* of BDL-LPS rats with (n=16) or without (n=12) Yaq-001 treatment.
1228 Yaq-001 treatment significantly improved the survival of BDL-LPS rats compared to
1229 untreated-BDL-LPS rats (log rank test, $p=0.003$).

1230 *Plasma ALT concentrations* in Sham-LPS (n=7), Sham-LPS-Yaq-001 (n=5), BDL-LPS
1231 (n=10) and BDL-LPS-Yaq-001 (n=9) groups and *PP measurements* in Sham-LPS (n=8),
1232 Sham-LPS-Yaq-001 (n=10), BDL-LPS (n=9) and BDL-LPS-Yaq-001 (n=9) groups. Yaq-
1233 001-treated BDL-LPS rats had a significantly lower ALT and PP compared to untreated-
1234 BDL-LPS rats ($p<0.005$).

1235 *Brain water percentage* in Sham-LPS (n=4), Sham-LPS-Yaq-001 (n=4), BDL-LPS (n=7),
1236 BDL-LPS-Yaq-001 (n=13) groups. *Arterial ammonia concentrations* in Sham-LPS (n=5),
1237 Sham-LPS-Yaq-001 (n=5), BDL-LPS (n=7), BDL-LPS-Yaq-001 (n=7) groups. *Portal*

1238 venous ammonia concentrations in Sham-LPS (n=5), Sham-LPS-Yaq-001 (n=5), BDL-
1239 LPS (n=6), BDL-LPS-Yaq-001 (n=5) groups. Yaq-001 decreased brain water percentage
1240 and arterial/portal venous ammonia concentrations in BDL-LPS rats compared to
1241 untreated rats ($p<0.05$, $p<0.01$, $p<0.05$).

1242 Serum creatinine in Sham-LPS (n=4), Sham-LPS-Yaq-001 (n=3), BDL-LPS (n=12) and
1243 BDL-LPS-Yaq-001 (n=6) groups. Serum urea in Sham-LPS (n=8), Sham-LPS-Yaq-001
1244 (n=4), BDL-LPS (n=12) and BDL-LPS-Yaq-001 (n=8) groups. Yaq-001 significantly
1245 decreased creatinine levels in BDL-LPS rats ($p<0.05$).

1246 Plasma cytokines in Sham-LPS (n=6), Sham-LPS-Yaq-001 (n=9), BDL-LPS (n=8) and
1247 BDL-LPS-Yaq-001 (n=8) groups. Yaq-001 significantly decreased plasma IL-1 β and IL-
1248 10 concentrations in BDL-LPS groups ($p<0.01$, $p<0.05$).

1249

1250 **Fig.2.Effect of Yaq-001 on gene expression profiles in the multiorgans in BDL rats.**

1251 (A) Rats underwent BDL for 4-weeks as a model of cirrhosis (n=3-4/group) and the
1252 treatment groups received Yaq-001 for 2-weeks before sacrifice. Liver, colon, brain and
1253 kidney were collected for transcriptomic analysis. (B, D, F, H) Heatmap of differentially
1254 expressed genes (DEGs) in different organs between Sham (n=3), Sham-Yaq-001 (n=3),
1255 BDL (n=3) and BDL-Yaq-001 (n=4) groups. DEGs were identified at 1.2-fold change and
1256 $p=0.1$ threshold in three pairwise groups (BDL versus Sham, BDL-Yaq-001 versus BDL,
1257 Sham-Yaq-001 versus Sham). (C, E, G, I) Volcano plot of pairwise DEGs in four organs
1258 among Sham (n=3), Sham-Yaq-001 (n=3), BDL (n=3) and BDL-Yaq-001 (n=4) groups.
1259 The vertical dashed lines indicated the threshold for 1.2-fold change. The horizontal
1260 dashed line indicated the adjusted $p=0.05$ and $p=0.1$ threshold. The right part indicates
1261 up-regulation of gene expression, and the left part indicates down-regulation of gene
1262 expression. The top 20 genes are indicated by gene names.

1263

1264 **Fig.3. Effect of Yaq-001 treatment on the microbiome composition.** (A) Heatmap of

1265 gut microbiome associated with the effect of Yaq-001 as determined by 16S PCR at the
1266 family level. The Family *Porphyromonadaceae* with asterisk was statistically differently
1267 abundant between BDL (n=7) vs Sham (n=6), and between BDL-Yaq-001 (n=7) vs BDL
1268 groups (n=7) (Wilcoxon rank sum test, $p<0.05$). The abundance of this family was

1269 statistically higher in BDL group than in Sham group, and its abundance statistically
1270 decreased in the BDL-Yaq-001 group than in the BDL group. The other six families in the
1271 heatmap were with marked fold changes between BDL vs Sham, and between BDL-Yaq-
1272 001 vs BDL groups ($|\log_2FC|>2$). Of these, five were more abundant in the BDL group
1273 than in the Sham group. The abundance largely decreased in the Yaq-001-treated group.
1274 In addition, of these, one family was less abundant in the BDL group than in the Sham
1275 group. The abundance increased in the Yaq-001-treated group. (B) Heatmap of gut
1276 microbiome at the Genus level. The Genus *Barnesiella* with asterisk was statistically
1277 differently abundant between BDL vs Sham, and between BDL-Yaq-001 vs BDL groups
1278 (Wilcoxon rank sum test, $p<0.05$). The abundance of this genus was statistically higher
1279 in BDL group than in the Sham group, and its abundance statistically decreased in the
1280 BDL-Yaq-001 group. The other 19 genera in the heatmap represent those with significant
1281 fold change values between BDL vs Sham, and between BDL-Yaq-001 vs BDL groups
1282 ($|\log_2FC|>2$). Of these, 14 were more abundant in the BDL group compared with the
1283 Sham group. The abundance decreased in the Yaq-001-treated BDL group. In addition, 5
1284 genera were less abundant in the BDL group than in the Sham group. Their abundance
1285 increased in the Yaq-001-treated BDL animals. (C, D) Correlation plots between markedly
1286 changed genes and gut microbiome at family/genus. The genes were from amongst the
1287 top 20 changed genes in BDL animals with Yaq-001 treatment. Nodes represent either
1288 genes (lower semi-circular part) or bacteria (upper semi-circular part) at the family and
1289 genus level. The nodes are colored based on the log-fold change for the differential gene
1290 expression and differences in the bacterial abundance. The red nodes indicate an
1291 increase and blue nodes indicate a decrease. Edges represent the correlation coefficient
1292 calculated between genes and microbial genus or family with red indicating a positive
1293 correlation and blue a negative correlation. Correlation coefficients greater or equal to 0.4
1294 were plotted in plot C (Spearman's coefficient ≥ 0.4), and D shows all correlations.

1295

1296 **Fig.4. Effect of Yaq-001 on organ dysfunction, ammonia and endotoxemia in CCl₄**
1297 **mice.**

1298 (A) Mice underwent CCl₄ injection for 6-weeks as a model of cirrhosis (n=6-12/group) and
1299 the treatment groups received Yaq-001 for 6 weeks before sacrifice.

1300 *Plasma ALT, ALP and TBIL concentrations* in Control (n=6), Control-Yaq-001 (n=6), 6-
1301 week CCl₄ (n=12) and 6-week CCl₄-Yaq-001 (n=12) groups. Significantly higher ALT, ALP
1302 and TBIL were observed in CCl₄ compared to controls (p=0.0001, p=0.0007, p=0.012).
1303 Yaq-001-treated CCl₄ mice had a significantly lower ALT, ALP and TBIL compared to
1304 untreated-CCl₄ mice (p<0.0001, p=0.040, p=0.001).

1305 *H&E and PSR staining of liver tissue.* CCl₄ mice were associated with a significant
1306 increase in CPA compared to Controls (p=0.0001). Yaq-001 had significant effect on CPA
1307 in CCl₄-Yaq-001 compared to CCl₄ mice(p=0.024).

1308 *TUNEL staining liver tissues.* Significantly greater staining was observed in CCl₄
1309 compared to Controls (p=0.0001). Yaq-001-treated CCl₄ mice had a significantly lower
1310 TUNEL staining compared to untreated-CCl₄ (p=0.021) with Yaq-001 treatment.

1311 *Venous ammonia concentrations and serum creatinine levels* in Control (n=6), Control-
1312 Yaq-001 (n=6), 6-week CCl₄ (n=12) and 6-week CCl₄-Yaq-001 groups(n=12).
1313 Significantly increased ammonia concentrations were observed in CCl₄ compared to

1314 Controls (p=0.0020). Yaq-001 significantly decreased venous ammonia concentrations
1315 and serum creatinine levels in CCl₄ mice (p=0.025, p=0.005).

1316 *Venous endotoxin concentrations* in Control (n=3), Control-Yaq-001 (n=3), 6-week CCl₄
1317 (n=10) and 6-week CCl₄-Yaq-001 groups(n=10). Significantly higher venous endotoxin
1318 was observed in CCl₄ mice compared to Control mice (p=0.007). Yaq-001 administration
1319 was associated with a significant reduction of venous endotoxin compared to untreated-
1320 CCl₄ mice (p=0.007).

1321

1322 (B) Mice underwent CCl₄ injection for 12-weeks as a model of cirrhosis (n=6-12/group)
1323 and the treatment groups received Yaq-001 for 6-weeks before sacrifice.

1324 *Plasma ALT, ALP and TBIL concentrations* in Control (n=6), Control-Yaq-001 (n=6), 12-
1325 week CCl₄ (n=12) and 12-week CCl₄-Yaq-001 (n=12) groups. Significantly higher ALT,
1326 ALP and TBIL were observed in CCl₄ compared to controls (p=0.0001, p=0.0008,
1327 p=0.007). Yaq-001-treated CCl₄ mice had a significantly lower ALT compared to
1328 untreated-CCl₄ mice (p<0.0001).

1329 *H&E and PSR staining of liver tissue* in CCl₄ mice. CCl₄ mice were associated with a
1330 significant increase in CPA compared to Controls (p=0.0001). Yaq-001 had significant
1331 effect on CPA in CCl₄-Yaq-001 compared to CCl₄ mice(p=0.012).

1332 *TUNEL staining of liver tissues.* Significantly higher TUNEL staining was observed in CCl₄
1333 compared to Controls (p=0.0001). Yaq-001-treated CCl₄ mice had a significantly lower
1334 TUNEL staining compared to untreated-CCl₄ (p=0.017) indicative of a reduction in liver
1335 cell death with Yaq-001 treatment.

1336 *Venous ammonia.* Significantly increased ammonia concentrations were observed in
1337 CCl₄ compared to Controls (p=0.001). Yaq-001 significantly decreased venous ammonia
1338 concentrations in CCl₄ mice (p=0.035).

1339 *Serum creatinine:* Yaq-001 significantly decreased serum creatinine levels in CCl₄ mice
1340 (p=0.003).

1341 *Venous endotoxin concentrations* in Control (n=3), Control-Yaq-001 (n=3), 12-week CCl₄
1342 (n=10) and 12-week CCl₄-Yaq-001 groups(n=10). Significantly higher venous endotoxin
1343 was observed in CCl₄ mice compared to Control mice (p=0.007). Yaq-001 administration
1344 was associated with a significant reduction of venous endotoxin compared to untreated-
1345 CCl₄ mice (p=0.043).

1346

1347 **Fig.5. Effect of Yaq-001 on gut permeability in intestinal organoids.**

1348 (A) Intestinal organoids derived and cultured from small intestine of C57BL/6 mice
1349 underwent eversion into apical-out polarity in the first 12 h of suspension culture.
1350 Immunostaining of the microvilli (mv; F-actin) demonstrated that intestinal organoids in
1351 suspension have reversed polarity from basolateral-out to apical-out.

1352 (B) Apical-out intestinal organoids in suspension culture generate goblet cells (MUC2).

1353 (C)Gut permeability of apical-out intestinal organoids was significantly increased by
1354 coculturing with fecal water from CCl₄ group than control group (p=0.003). Gut
1355 permeability was notably decreased in fecal water from CCl₄-Yaq-001 group compared
1356 to CCl₄ group (p=0.001).

1357 (D)Quantification of the integrated density/area of each group.

1358

Figure 1

A

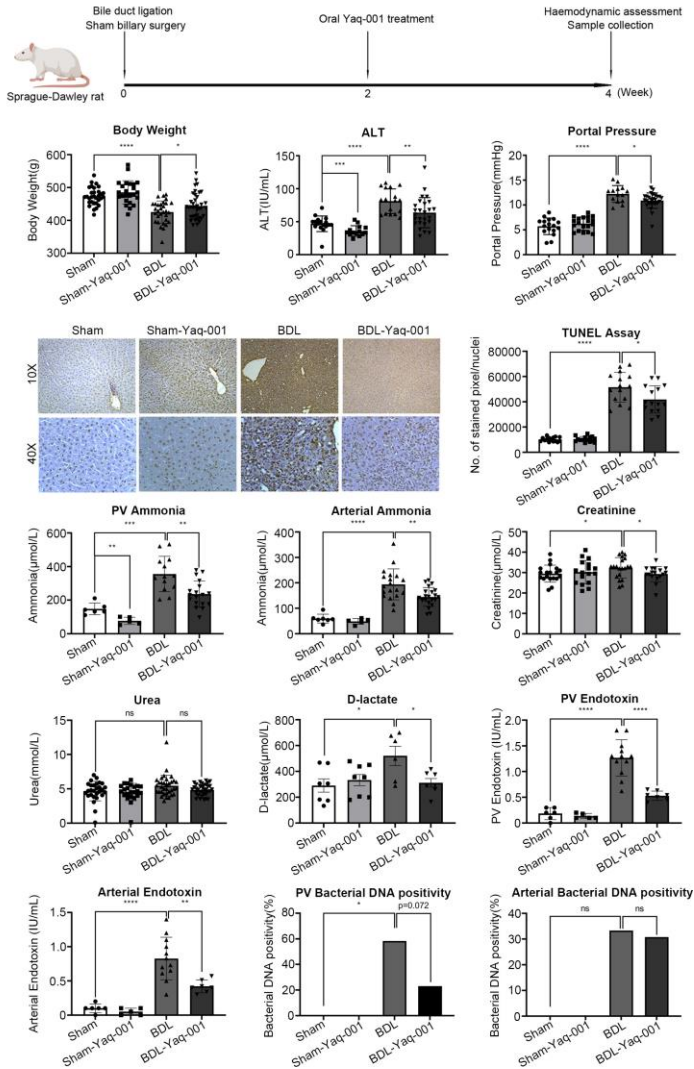


Figure 1

B

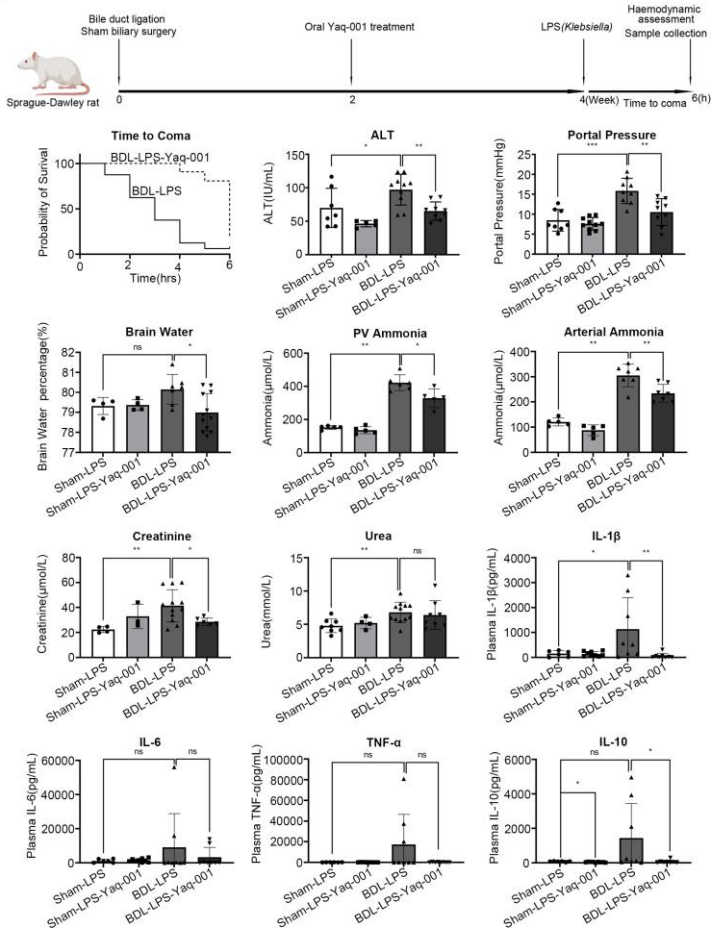


Figure 2

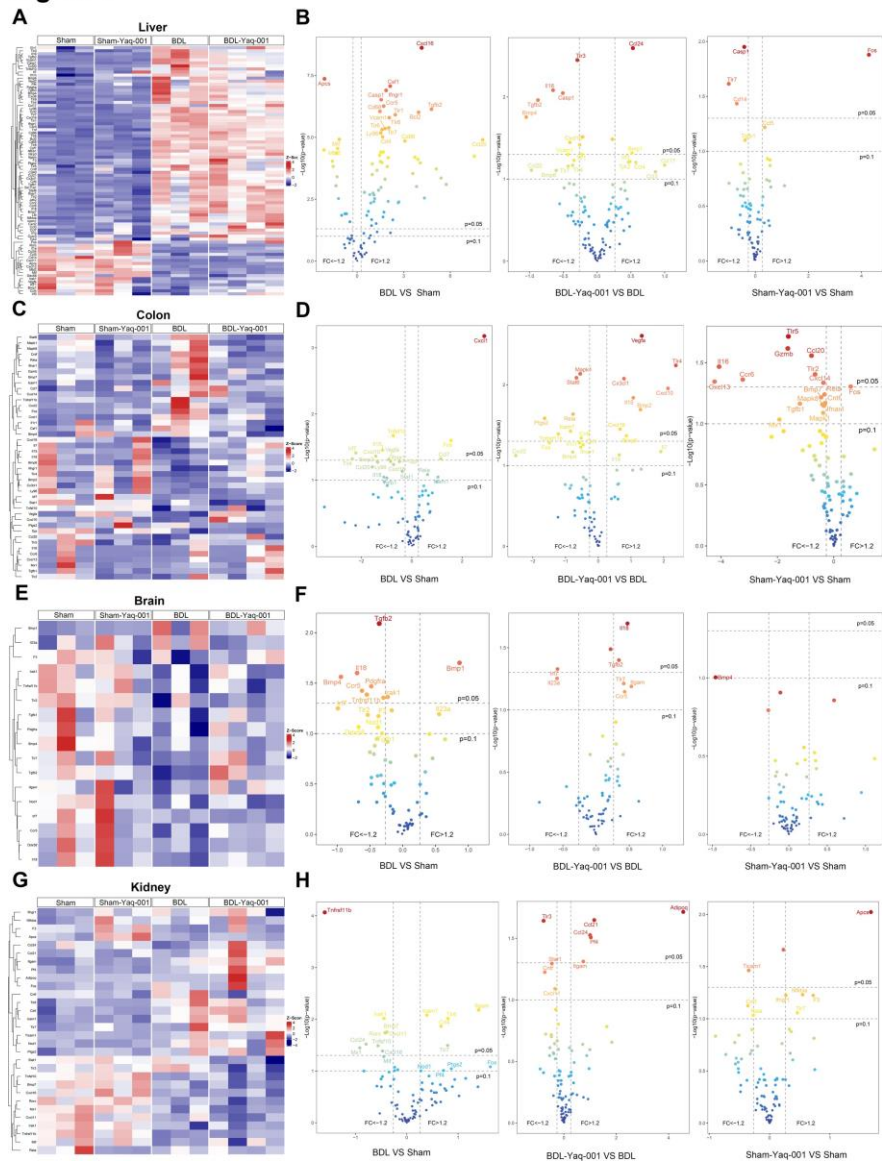


Figure 3

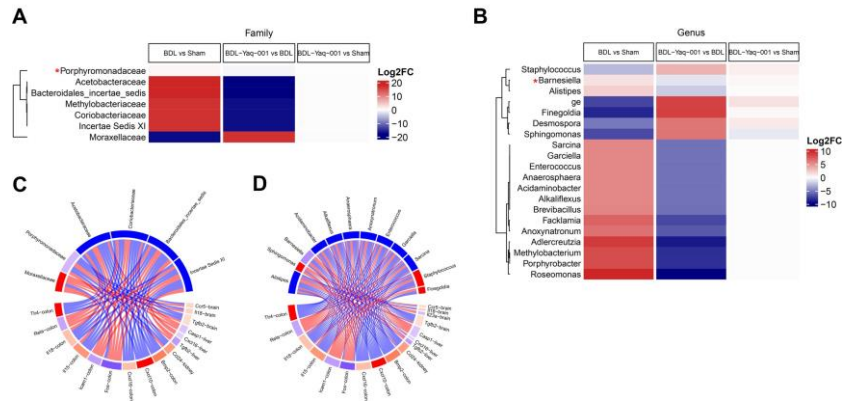


Figure 4

A

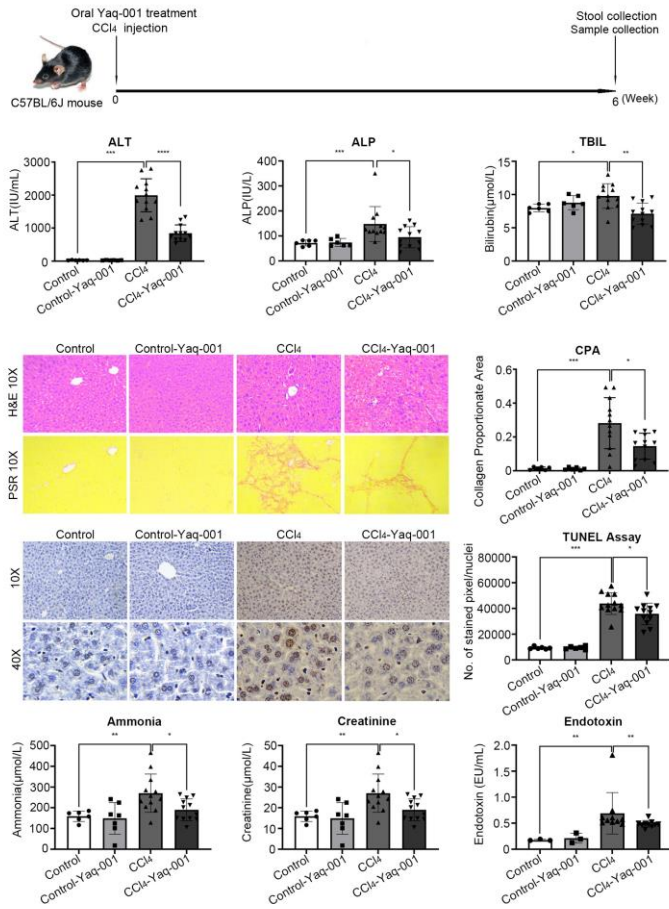


Figure 4

B

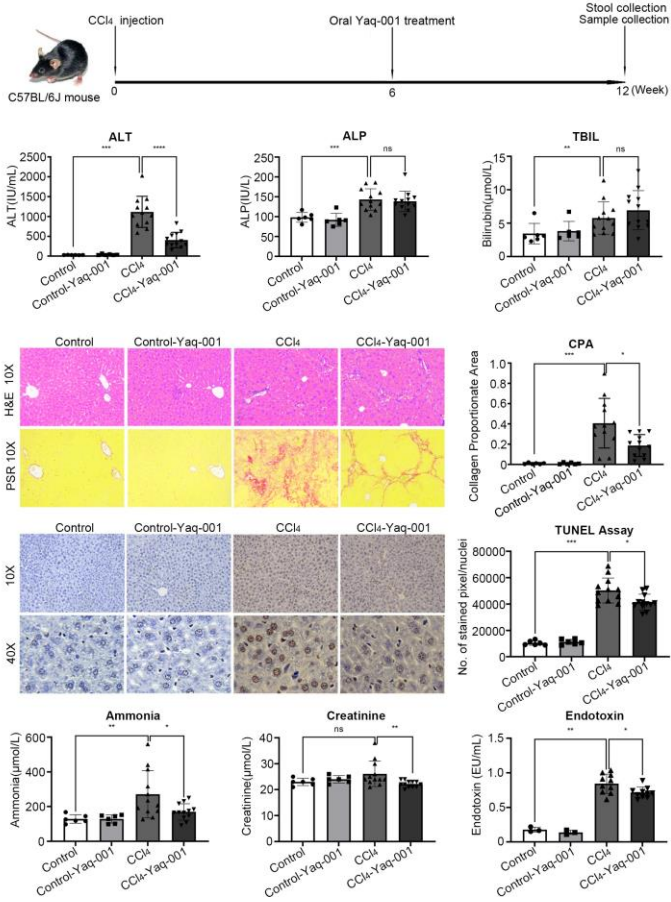


Figure 5

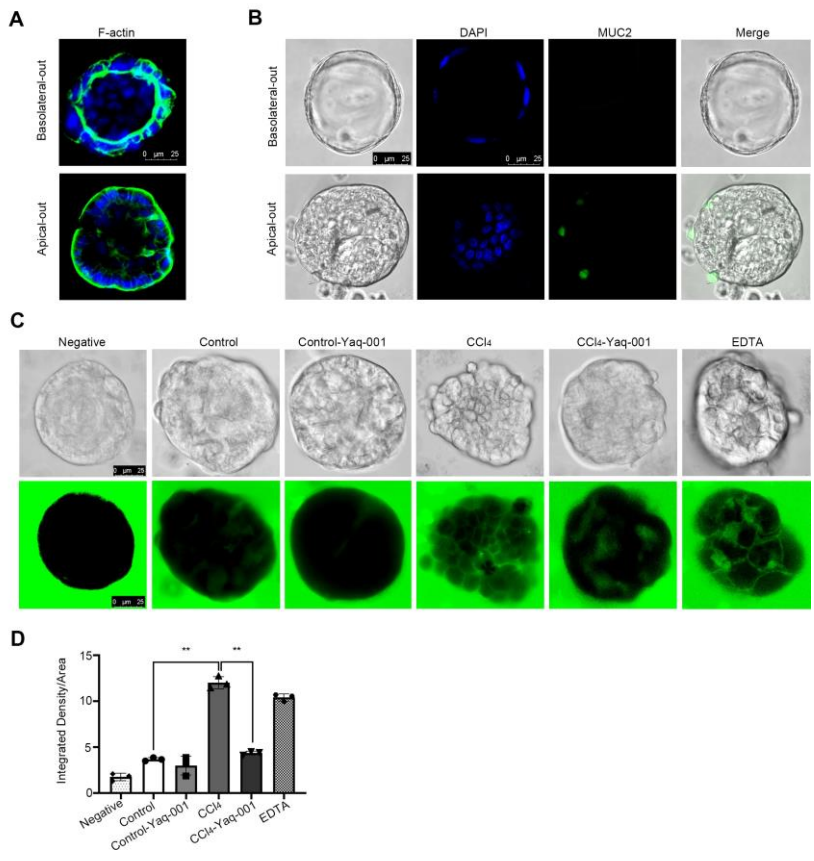


Table 1. Patient Characteristics

	Placebo	Active
Number	12	14
Age(years)	58.5(35-75)	58.5(47-68)
Male sex	9(75%)	10(71.4%)
Race/ethnicity		
Black	0(0%)	0(0%)
Other	12(100%)	14(100%)
BMI (kg/m ²) (SD)	27.8(22.9-32.8)	26.3(20.4-32.8)
Child Pugh Score	7(7-8)	7(7-8)
MELD score	13.2(10.2-16.1)	12.6(9.7-13.5)
Decompensation history		
Alcoholic hepatitis	2(16.7%)	0(0.0%)
Ascites	8(66.7%)	11(78.6%)
Hepatic encephalopathy	5(41.6%)	2(14.3%)
Varices	2(16.7%)	6(42.9%)
Laboratory values		
Bilirubin(μ mol/L)	31(19-40)	41(17-68)
Albumin(g/L)	34(30-38)	34(30-39)
Creatinine (μ mol/L)	65(53-83)	70(60-81)
Sodium(mmol/L)	136.5(134-137)	137.2(134-140)

1366
1367
1368
1369
1370
1371
1372
1373
1374
1375

Table 2. Adverse and Serious Events

Adverse Event	Placebo n (%)	Active n (%)
Constipation	3(25%)	2(14%)
Epigastric pain	0(0%)	1(7%)
Nausea	0(0%)	1(7%)
Meteorism	0(0%)	1(7%)
Osophageal reflux	0(0%)	1(7%)
Diarrhoea	2(17%)	0(0%)
Diuresis	0(0%)	1(7%)
Serious Event		
Death	0(0%)	0(0%)
50% increase in MELD	0(0%)	0(0%)
100% increase in creatinine	0(0%)	0(0%)
50% reduction in BMI	0(0%)	0(0%)
Acute decompensation	0(0%)	0(0%)
Episode of ACLF	0(0%)	0(0%)

1376

1377

1378

1379

1380

1381

1382

1383

1384

Table 3. Safety Parameters: Clinical laboratory assessments, Royal Free Global Assessment and Micronutrient concentrations

Parameters	Placebo			Active		
	Baseline	4 Weeks	12 Weeks	Baseline	4 Weeks	12 Weeks
Laboratory Parameters	Median(range)			Median(range)		
Haemoglobin(g/dL)	122(108-143)	119(107-145)	119(105-139)	122(113-134)	123(113-133)	121(110-137)
Leucocyte count($10^9/L$)	4.22(3.47-4.97)	4.05(3.43-4.84)	3.94(3.20-4.90)	5.22(4.20-6.99)	5.12(4.21-6.09)	4.70(4.26-5.52)
Platelets ($10^9/L$)	81(75-113)	93(83-104)	93(85-100)	95(81-127)	84(75-107)	93(75-105)
Bilirubin($\mu\text{mol/L}$)	41(17-68)	31(16-79)	47(20-71)	31(19-40)	42(21-52)	31(17-47)
ALT(IU/L)	25.9(19.8-8-8.55)	22.89(18.0-7-31.33)	24.1(18.0-7-31.33)	25.9(24.1-34.94)	27.11(24.1-33.13)	28.31(19.8-8-34.94)
ALP(IU/L)	49.2(37.8-52.8)	37.2(24-54)	40.8(27-56.4)	46.8(31.2-51)	46.8(30-69)	45(30-61.2)
Albumin(g/L)	34(30-39)	35(31-40)	32(31-38)	34(30-38)	35(30-38)	32(30-38)
Sodium(mmol/L)	137(135-140)	138(132-140)	138(135-140)	137(134-137)	136(132-137)	137(136-138)
Creatinine ($\mu\text{mol/L}$)	70(60-81)	77(63-83)	65(63-90)	65(53-83)	64(53-75)	70(57-72)
INR	1.4(1.3-1.6)	1.3(1.2-1.5)	1.4(1.3-1.5)	1.3(1.2-1.4)	1.4(1.2-1.4)	1.3(1.2-1.4)
Child Pugh Score	7(7-8)	7(6-8)	8(6-8)	7(7-8)	8(6.7-9)	8(7-9)

MELD score	12(10-17)	12(10-17)	13(12-16)	13(11-14)	14(11-14)	13(9-15)
MELD Na score	15(10-21)	13(9-19)	16(12-21)	15(13-17)	16(15-17)	14(13-19)
Nutritional Status		n (%)			n (%)	
Adequate	10(83%)	8(73%)	10(91%)	13(93%)	13(100%)	10(77%)
Moderate malnourishment	2(17%)	3(27%)	1(9%)	1(7%)	0(0%)	3(23%)
Severe malnourishment	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)
Micronutrient Concentrations		Median(range)		Median(range)		
Vitamin B12(ng/L)	533(290-765)	530(384-970)	524(275-815)	503(337-790)	483(365-694)	462(326-749)
Folate(µg/L)	16(9-22)	17(13-19)	23(21-26)	22(20-30)	20(18-24)	21(17-27)
Vitamin D(nmol/L)	28(20-51)	45(25-57)	43(26-61)	32(25-72)	57(22-72)	70(37-72)
Vitamin A(µmol/L)	0.48(0.13-2.18)	0.47(0.12-1.90)	0.39(0.09-1.80)	0.46(0.1-5-1.84)	0.39(0.16-1.86)	0.62(0.17-1.82)
Vitamin E(µmol/L)	28.4(17.7-42.0)	32.5(17.6-36.5)	29.9(20.8-36.1)	30.0(19.5-45.6)	30.3(18.3-46.2)	29.4(17.7-37.9)
Vitamin K(µg/L)	0.58(0.16-3.28)	0.33(0.12-2.5)	0.35(0.17-4.2)	0.58(0.1-4-4.3)	0.41(0.12-5.7)	0.60(0.14-2.8)
Copper(µmol/L)	13.6(8.30-22.6)	13.6(7.10-23.7)	13.5(11.2-24.1)	13.7(10.0-30.1)	15.5(9.10-26.7)	15.1(7.80-37.6)

Zinc($\mu\text{mol/L}$)	9.10(5.60-15.3)	8.85(5.00-13.7)	8.40(5.70-14.3)	8.20(5.60-12.9)	8.60(4.70-15.0)	9.00(5.00-15.8)
Selenium($\mu\text{mol/L}$)	0.78(0.56-1.10)	0.86(0.47-1.12)	0.90(0.65-1.04)	0.77(0.55-1.14)	0.93(0.48-1.23)	0.91(0.49-1.46)

