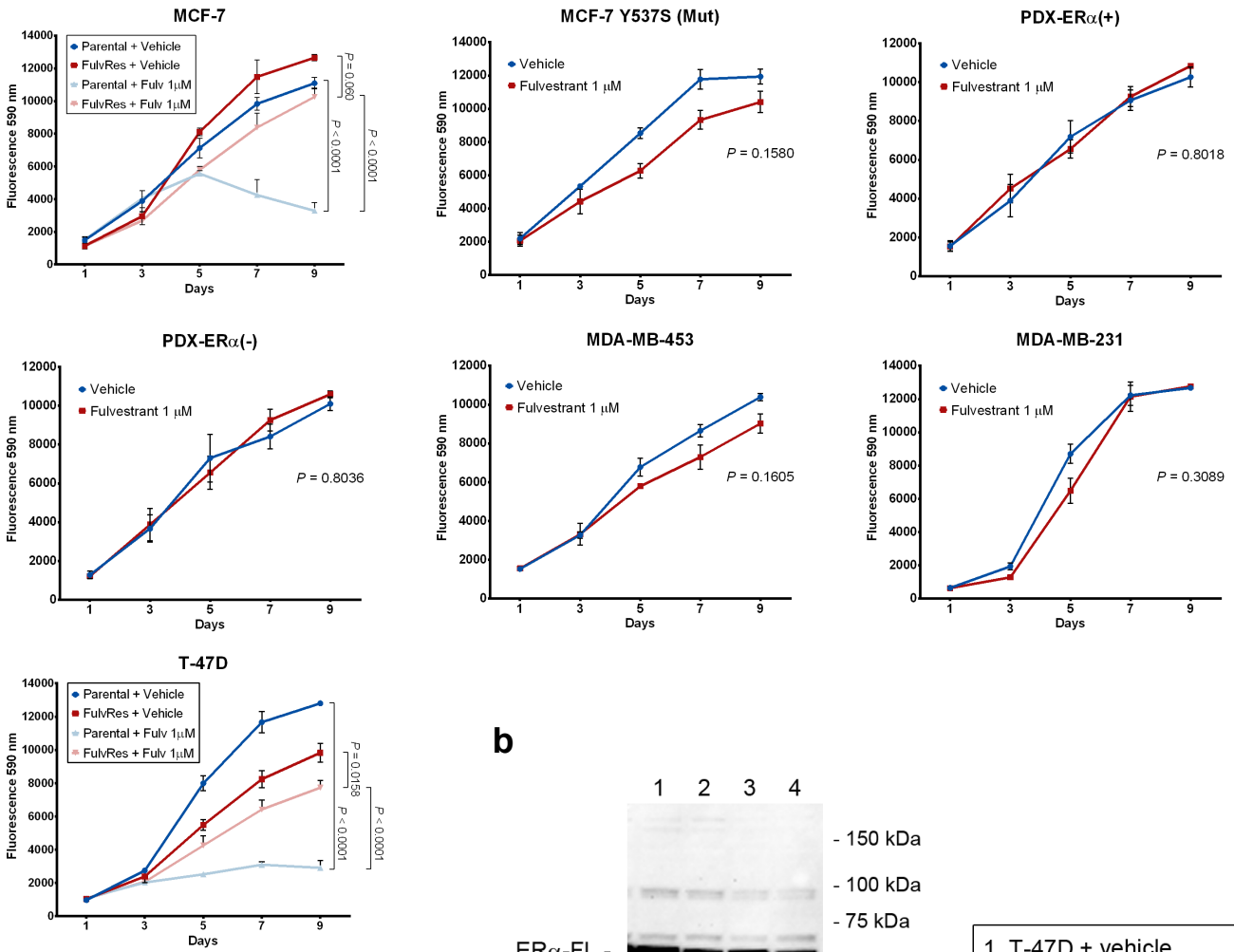
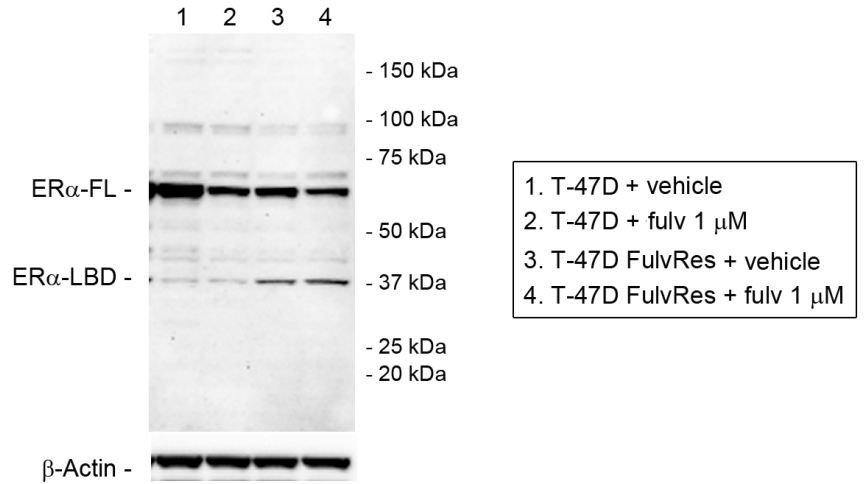


Supplementary Figure 1

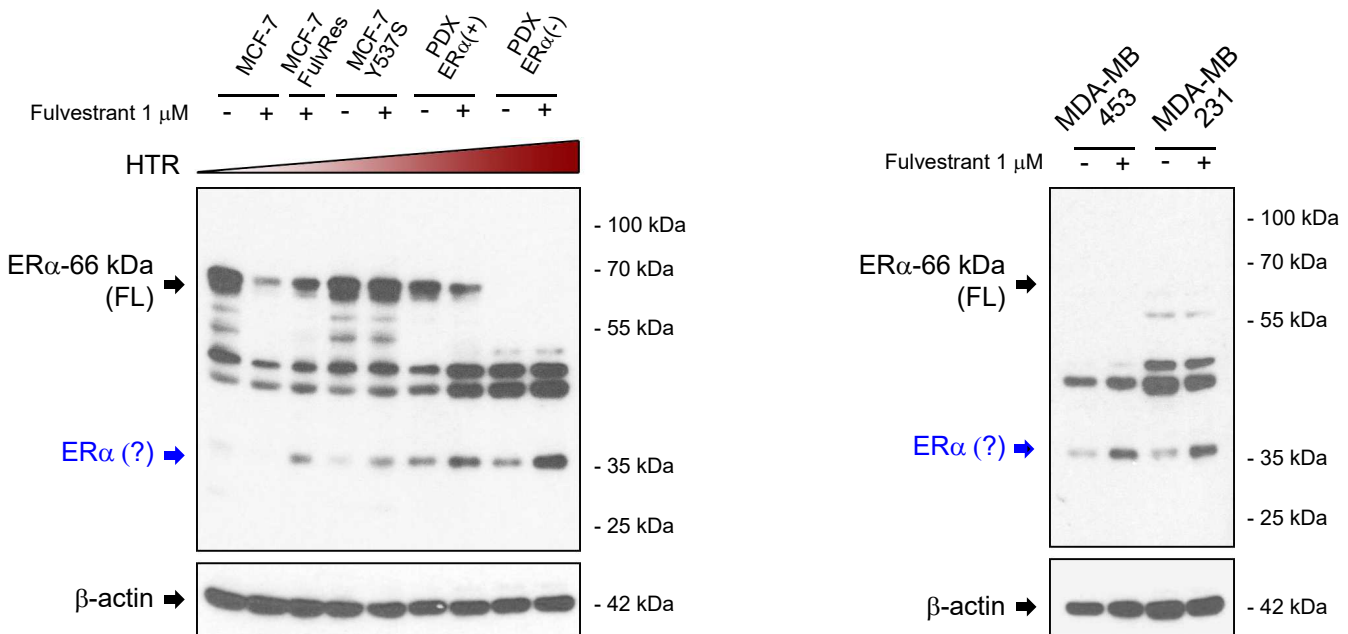
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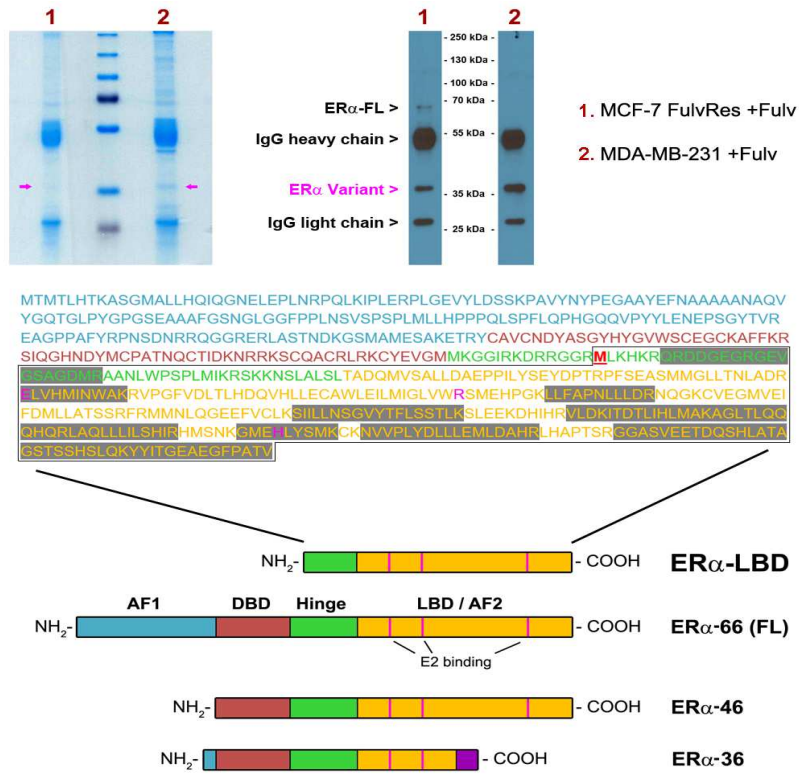
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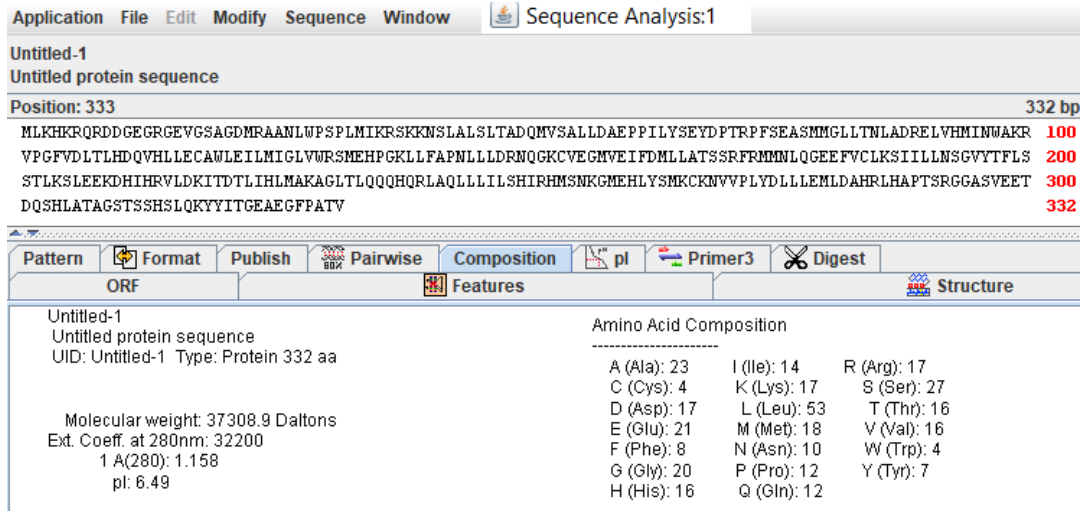
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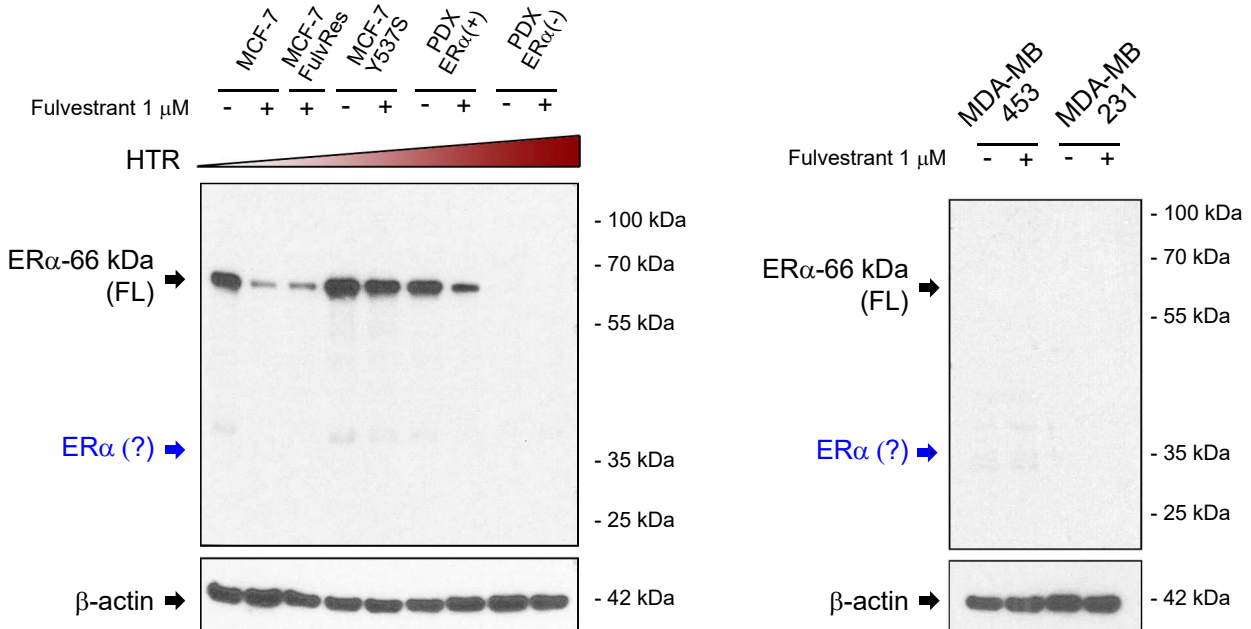
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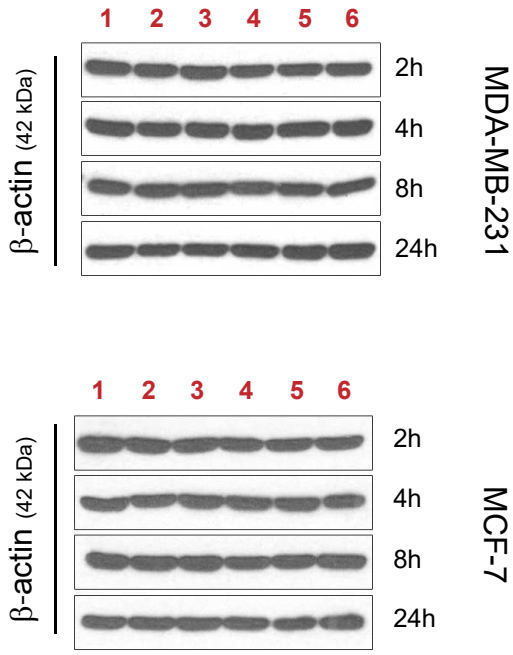
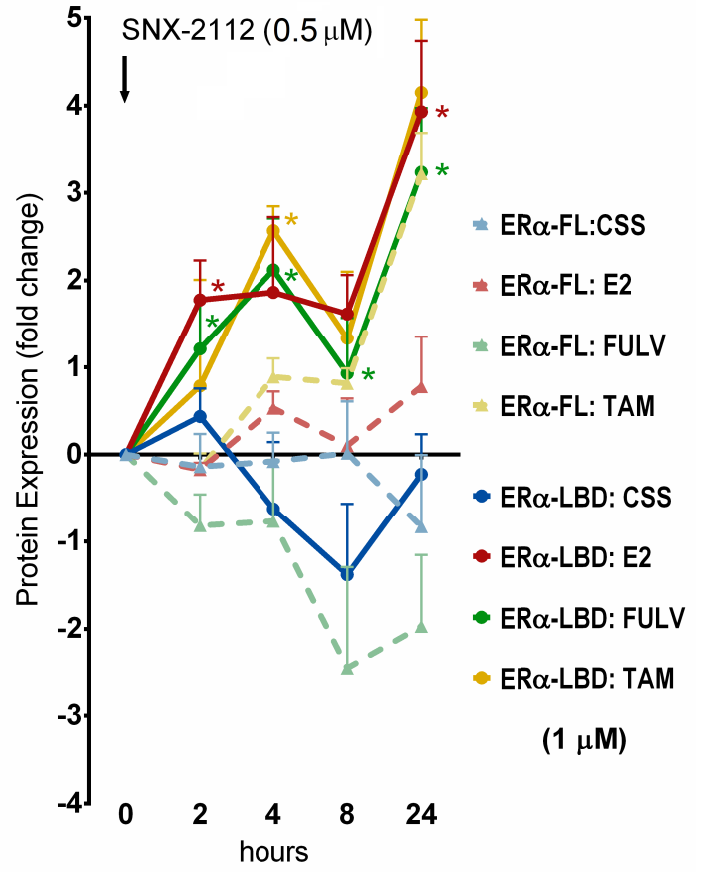


e



f



g**h**

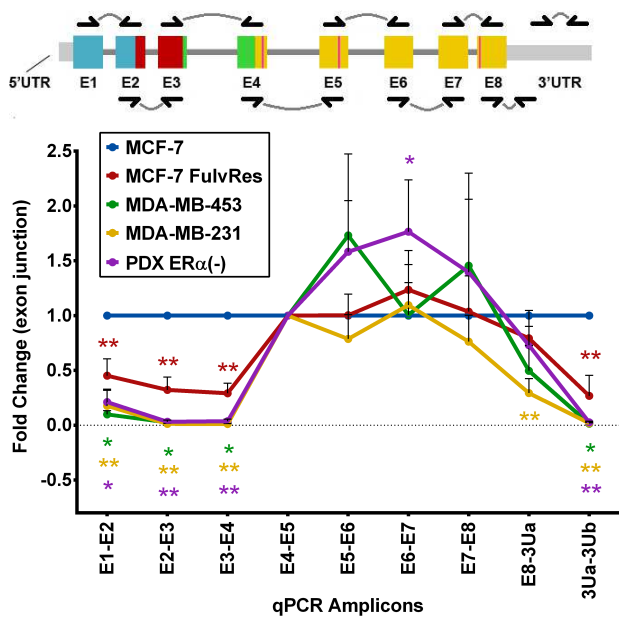
Supplementary Figure 1. a Proliferation of different breast cancer (BC) cell lines, in the presence or absence of fulvestrant 1 μ M treatment, assayed using resazurin reagent and expressed as fluorescence intensity (absorbance at 590 nm), taken as index of cell growth. Data are shown as mean \pm s.e.m. ($n = 3$ independent experiments). P values are shown (two-way ANOVA, Sidak's test for multiple comparisons). FulvRes = fulvestrant resistant; vehicle = DMSO. **b** Western blot analysis of ER α protein expression in T-47D cells (Parental vs. FulvRes). Cells were cultured for 24 h in the presence of vehicle (DMSO) or fulvestrant 1 μ M before lysis. ER α protein expression was normalized against β -actin. **c** Western blot analysis of ER α protein expression in breast BC cell lines. Cells were cultured for 24 h in the presence of vehicle (-) or fulvestrant 1 μ M (+) before lysis. Vehicle = DMSO. ER α protein was detected by using an antibody raised against the C-terminal of ER α protein and expression was normalized against β -actin. Increasing levels of hormonal therapy resistance (HTR) are indicated. **d** Protein lysates from MCF-7 FulvRes and MDA-MB-231 (both treated with fulvestrant 1 μ M for 24h) were immunoprecipitated using anti-ER α antibody. IP samples were run on two electrophoresis gels, one used for staining and band extraction, the other for WB check. Extracted bands were analyzed by capLC-MS/MS and results are shown. Sequenced/identified peptides from ER α protein are highlighted in dark grey and compared to the full-length ER α AA sequence. Different ER α domains with specific color code are also shown. AF1: transcription Activation Function-1 (cyan); DBD: DNA Binding Domain (red); Hinge (green); LBD/AF2: Ligand Binding Domain and transcription Activation Function-2 (yellow); amino acids involved in E2 binding (magenta); ER α -36 unique C-terminal (purple). **e** Analysis on ER α -LBD sequence. **f** As in (c). ER α protein was detected by using an antibody raised against the N-terminus of ER α protein. **g, h** ER α protein stability assay. ER α -FL and ER α -LBD protein levels were analyzed by western blot in MCF-7 and MDA-MB-231 cells, respectively. Protein samples were collected at different time points (2-24 h), after treatment. ER α protein levels were quantified and normalized against β -actin expression (**g**), and plotted as fold-change (\log_2), relative to SNX-2112 treatment alone (lane 2, medium containing physiological levels of E2 in pM range) (**h**). CSS = charcoal stripped serum. Data are presented as mean \pm s.e.m. ($n = 3$ independent experiments); * $P < 0.01$, two-way ANOVA (Tukey's correction), ER α -LBD vs. ER α -FL.

Supplementary Figure 2

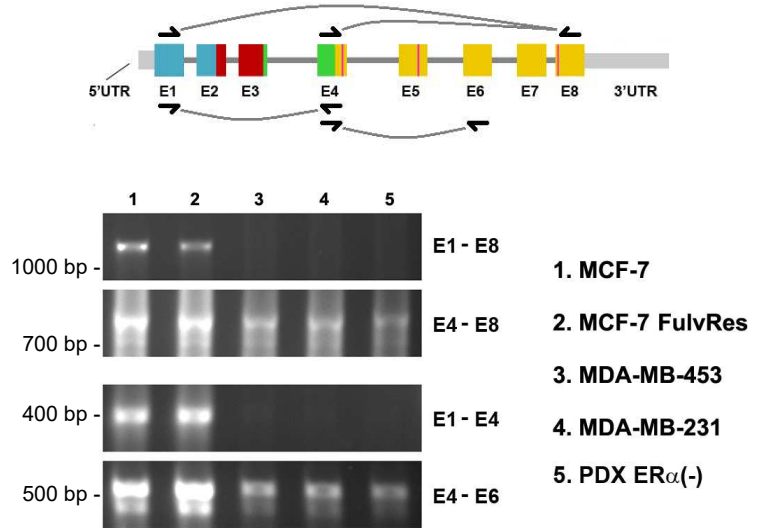
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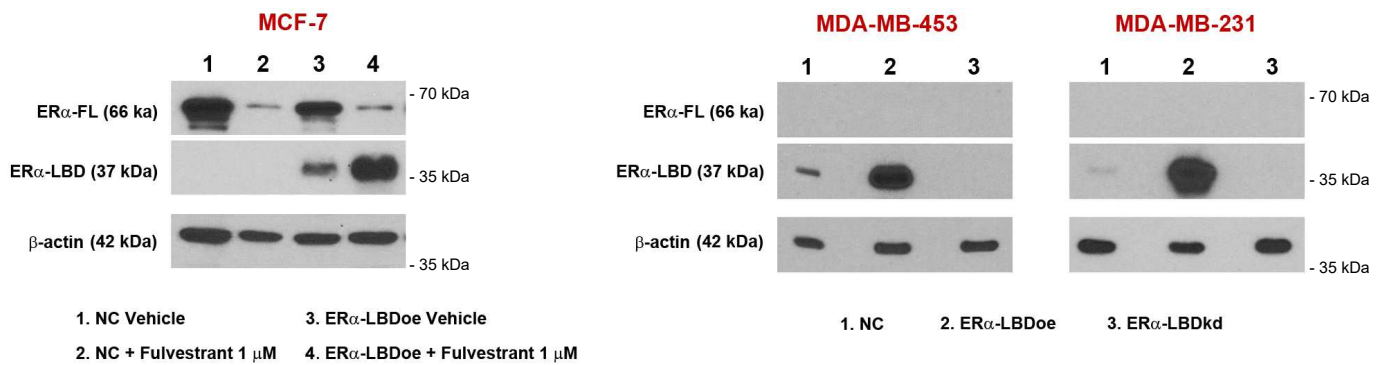
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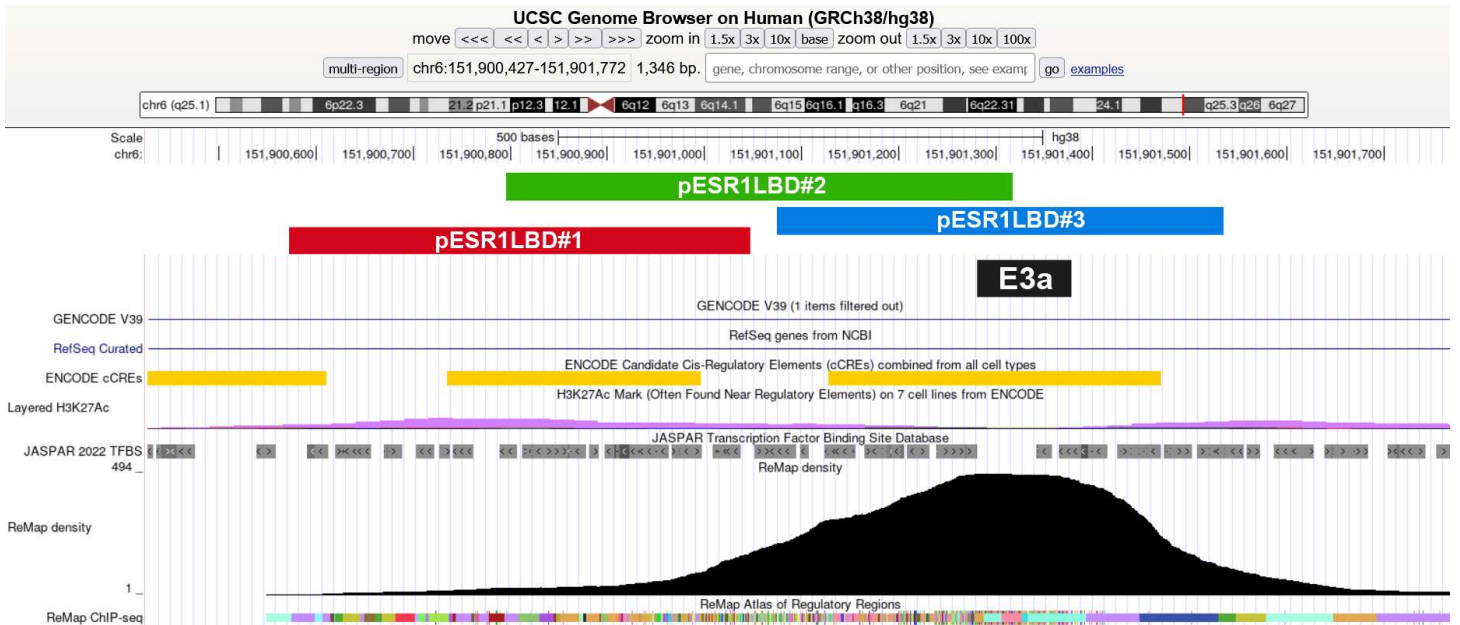
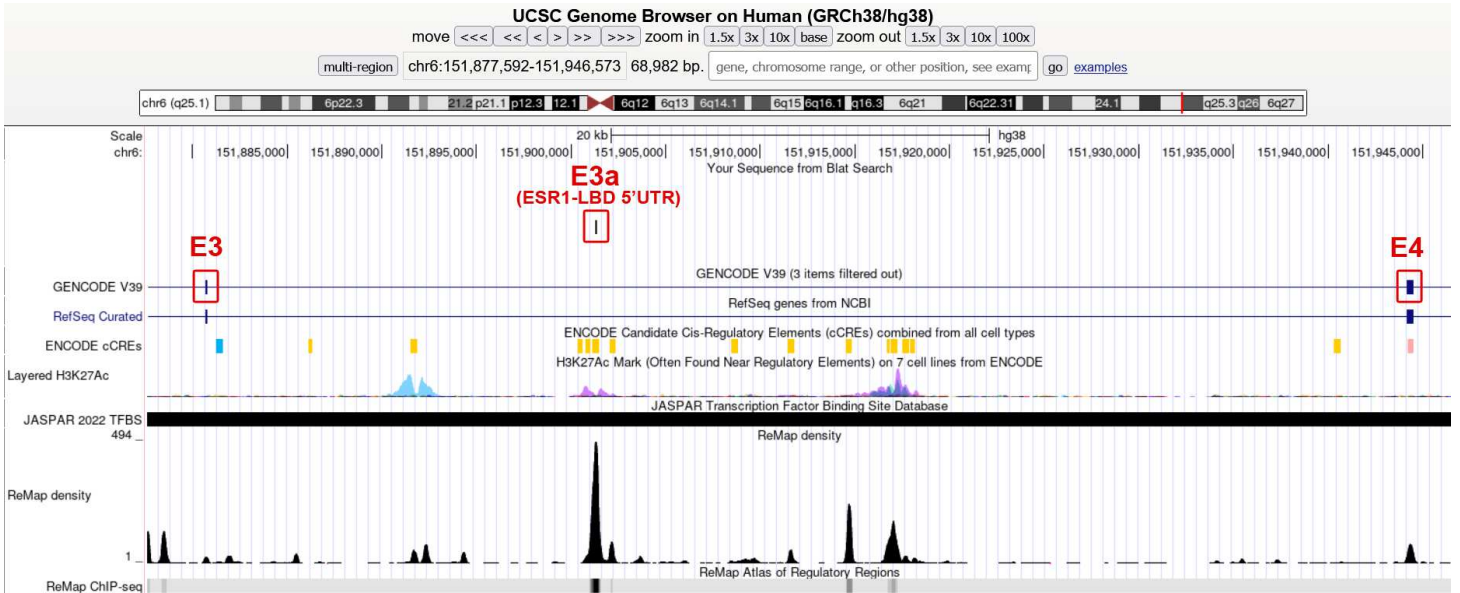
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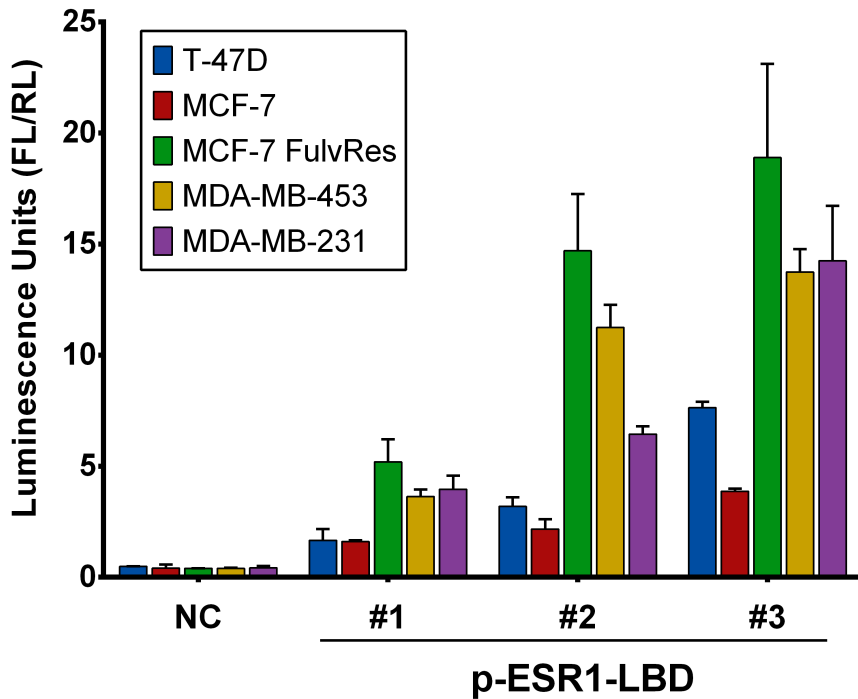
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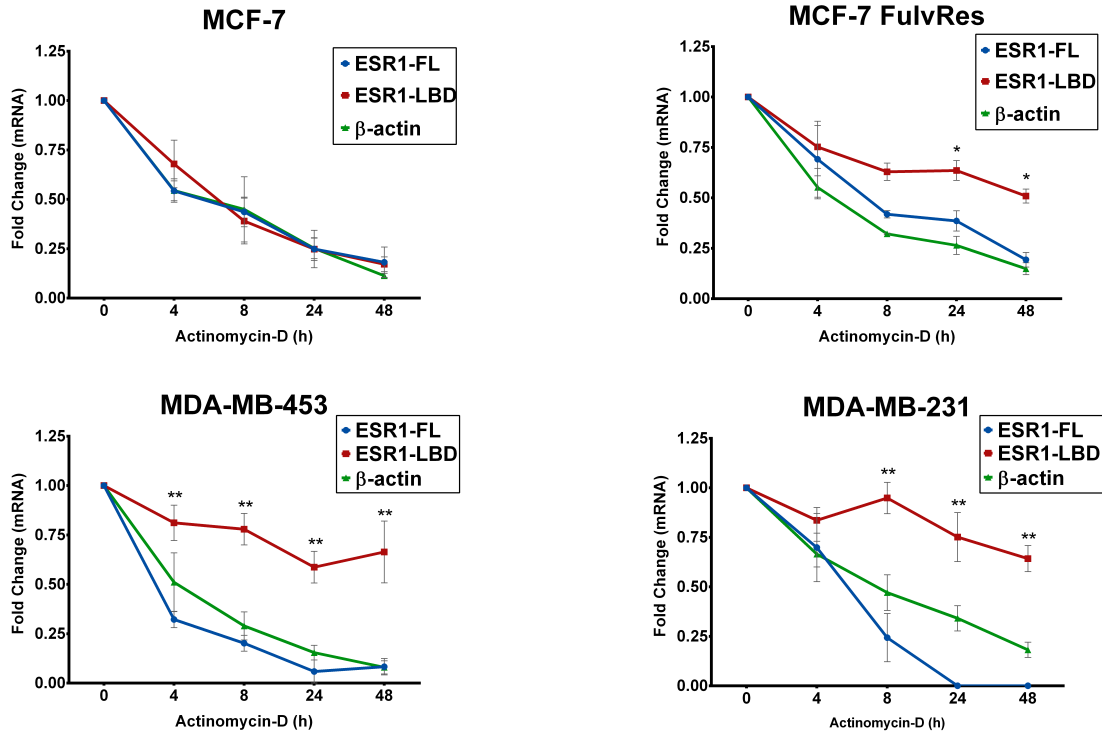
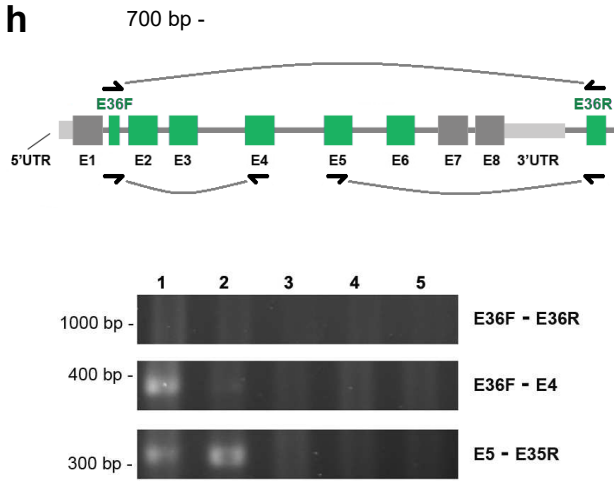


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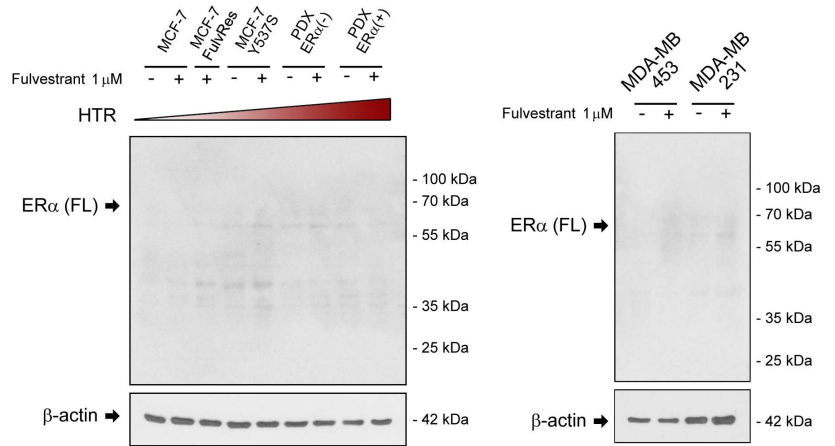


f



g**h**

1. MCF-7
2. MCF-7 FulvRes
3. MDA-MB-453
4. MDA-MB-231
5. PDX ER α (-)

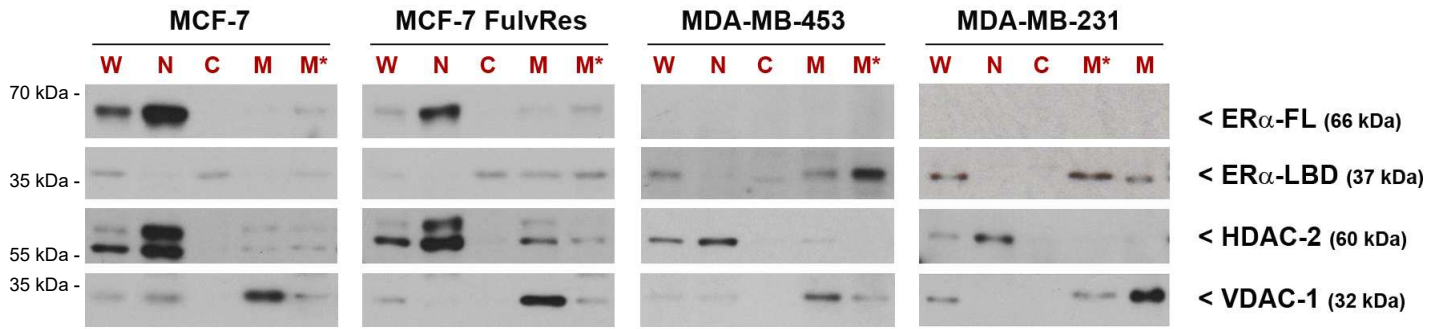
i

Supplementary Figure 2. a Screenshot of ZEMBU Genome Browser showing data from FANTOM5/FANTOM CAT analyses collected on ESR1 gene. Exon 3 (E3) and exon 4 (E4) of ESR1 gene are highlighted by blue arrows, together with a new putative transcription starting site (TSS) mapped in the intronic region between the two exons (E3a, red arrow). **b** Expression of ESR1 mRNA exon junctions in BC cell lines, analyzed by qPCR. Plot shows expression fold change, normalized on RPLP0 and relative to MCF-7 sample. Color-code of exons is based on different ER α protein domain. Data are presented as mean \pm s.e.m. ($n = 3$ independent experiments). * $P < 0.05$, ** $P < 0.01$, two-way ANOVA (Fisher's LSD test). **c** Expression of different ESR1 mRNA regions in BC cell lines, analyzed by RT-PCR. The scheme above depicts PCR amplicons and their position on ESR1 transcript. Color-code of exon as in (b). **d** Western blot analysis of ER α protein variants expression (ER α -FL vs. ER α -LBD) in different BC cell lines (stable clones). Vehicle (DMSO 0.01%) or fulvestrant treatment (1 μ M, 24 h) was added to MCF-7 cells (on the left). NC: cells transduced with control vector. ER α -LBDoe: cells transduced with ESR1-LBD CDS sequence (exon 4 to exon 8), inducing ER α -LBD protein overexpression. ER α -LBDkd: cells transduced with CRISPR/CAS9 vector targeting ESR1 exon 4 and promoting ER α -LBD protein knockdown. **e** Screenshots of UCSC Genome Browser showing regulatory elements and transcription binding site mapping on ESR1 exon E3a genomic region (putative ESR1-LBD 5'UTR). The zoomed area below shows three different genomic DNA regions representing putative ESR1-LBD promoter (pESR1-LBD) and screened for transcriptional activity. **f** pESR1-LBD promoter-driven luciferase reporter assay in transfected T-47D, MCF-7, MCF-7 FulvRes, MDA-MB-453 and MDA-MB-231 cells, respectively. Firefly luciferase (FL) levels were normalized to Renilla luciferase (RL). Controls (NC) = cells transfected with empty pGL3.basic plasmid. Data are presented as mean \pm s.e.m. ($n = 2$ independent experiments). **g** BC cells lines were treated with actinomycin-D and tested by qPCR at different time points for ESR1 and β -actin mRNA expression. ESR1-FL: PCR amplicon ranging from ESR1 exon E1 to E3. ESR1-LBD: PCR amplicon ranging from ESR1 exon E4 to E5. β -actin transcript was taken as positive control for mRNA decay. Plots shows mRNA fold change, normalized on RPLP0 and relative to time 0. Data are presented as mean \pm s.e.m. ($n = 3$ independent experiments). * $P < 0.05$, ** $P < 0.01$, two-way ANOVA (Fisher's LSD test). **h** Expression of different ESR1 mRNA regions in BC cell lines, analyzed by RT-PCR. The scheme above depicts PCR amplicons and their position on ESR1 transcript. Exons of ER α -36 transcript are in green. **i** Western blot analysis of ER α -36 protein expression in BC cell lines. Cells were cultured for 24 h in the presence of vehicle (-) or fulvestrant 1 μ M (+) before lysis.

Vehicle = DMSO. ER α protein expression was normalized against β -actin. Increasing levels of hormonal therapy resistance (HTR) are indicated.

Supplementary Figure 3

a



W = whole lysate
N = Nuclear fraction
C = Cytosolic fraction
M = Mitochondrial fraction
M* = Mitochondrial fraction + proteinase K

b

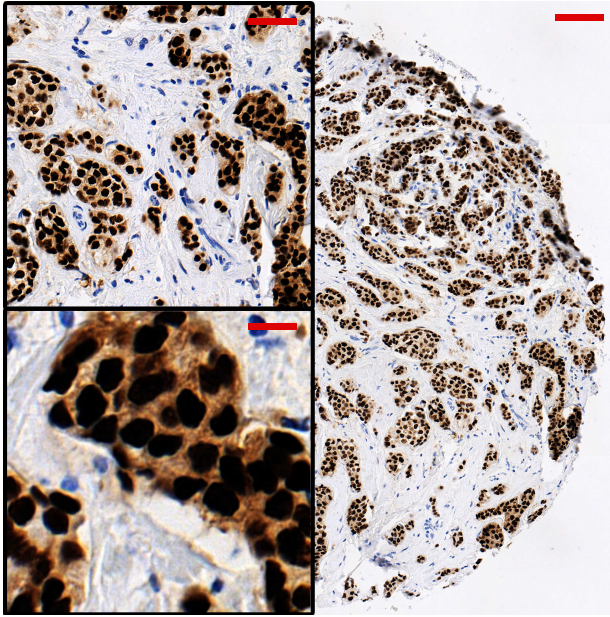
<i>Anti-ERα (Cell Signaling)</i>	MCF-7		MCF-7 +Fulv		MCF-7 FulvRes +Fulv	
	Nucl	Mito	Nucl	Mito	Nucl	Mito
threshold A	50	50	50	50	50	50
threshold B	50	50	50	50	50	50
number of colocalized voxels	535701	28128	75341	39987	49694	96212
% of dataset colocalized	1.7	0.09	0.27	0.14	0.23	0.44
% of ROI colocalized	1.7	0.09	0.27	0.14	0.23	0.44
% of volume A above threshold colocalized (DAPI / OXPHOS)	34.71	4.4	29.1	10.63	30.14	46.75
% of volume B above threshold colocalized (ERα)	53.89	2.83	31.28	16.6	13.55	26.24
% of material A above threshold colocalized (DAPI / OXPHOS)	35.48	4.91	29.52	11.89	29.24	50.07
% of material B above threshold colocalized (ERα)	56.09	2.06	34.49	14.63	14.94	25.42
% of ROI material A colocalized (DAPI / OXPHOS)	10.77	2.85	7.46	8.42	7.66	37.34
% of ROI material B colocalized (ERα)	31.29	1.15	15.58	6.61	9.89	16.82
Pearson's coefficient in dataset volume	0.5687	0.0975	0.5852	0.3261	0.452	0.4067
Pearson's coefficient in ROI volume	0.5687	0.0975	0.5852	0.3261	0.452	0.4067
Pearson's coefficient in colocalized volume	-0.0016	-0.1769	0.0433	-0.0152	-0.0603	0.0343
original Mander's coefficient A	0.7252	0.8992	0.7513	0.9647	0.6997	0.9805
original Mander's coefficient B	0.996	0.4609	0.9714	0.6228	0.9063	0.5377
thresholded Mander's coefficient A	0.1586	0.0551	0.145	0.1061	0.2041	0.4425
thresholded Mander's coefficient B	0.4157	0.0532	0.2268	0.1976	0.132	0.2136

<i>Anti-ERα (Cell Signaling)</i>	MDA-MB-453		MDA-MB-231		PDX ERα(-)	
	Nucl	Mito	Nucl	Mito	Nucl	Mito
threshold A	50	50	50	50	50	50
threshold B	50	50	50	50	50	50
number of colocalized voxels	2634	90200	23806	81216	4412	78345
% of dataset colocalized	0.01	0.3	0.11	0.37	0.02	0.31
% of ROI colocalized	0.01	0.3	0.11	0.37	0.02	0.31
% of volume A above threshold colocalized (DAPI / OXPHOS)	3.77	33.87	6.65	45.95	2.79	31.79
% of volume B above threshold colocalized (ERα)	0.67	22.8	10.9	37.19	2.16	38.28
% of material A above threshold colocalized (DAPI / OXPHOS)	3.77	39.11	6.43	53.84	2.73	35.49
% of material B above threshold colocalized (ERα)	0.53	22.47	9.27	41.03	1.8	38.27
% of ROI material A colocalized (DAPI / OXPHOS)	0.31	23.7	4.55	40.54	0.61	23.12
% of ROI material B colocalized (ERα)	0.26	11.13	3.66	16.19	0.79	16.89
Pearson's coefficient in dataset volume	0.2851	0.4097	0.3069	0.5341	0.3525	0.5341
Pearson's coefficient in ROI volume	0.2851	0.4097	0.3069	0.5341	0.3525	0.5341
Pearson's coefficient in colocalized volume	-0.0453	0.0104	-0.0515	0.1891	-0.0215	-0.0183
original Mander's coefficient A	0.7753	0.9737	0.9518	0.9927	0.7565	0.9925
original Mander's coefficient B	0.9236	0.6657	0.7661	0.5258	0.9522	0.8155
thresholded Mander's coefficient A	0.0735	0.3069	0.0774	0.4472	0.0621	0.2852
thresholded Mander's coefficient B	0.0127	0.1644	0.1685	0.2169	0.0576	0.2711

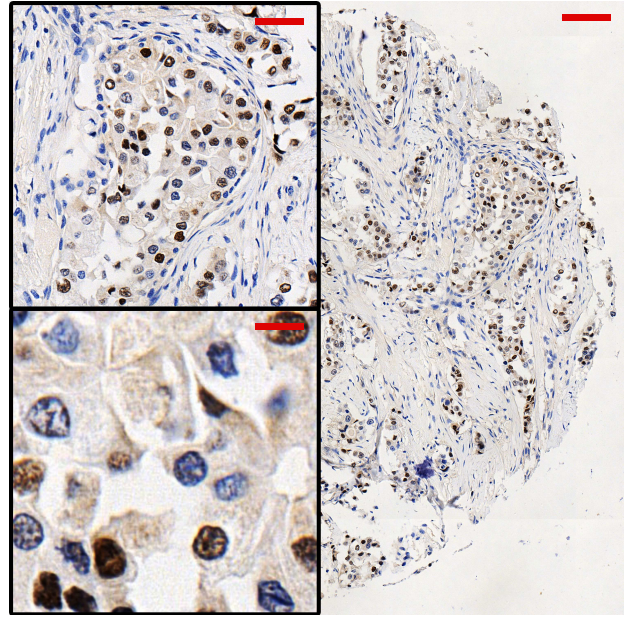
C

ER α staining with C-terminal antibody (D8H8)

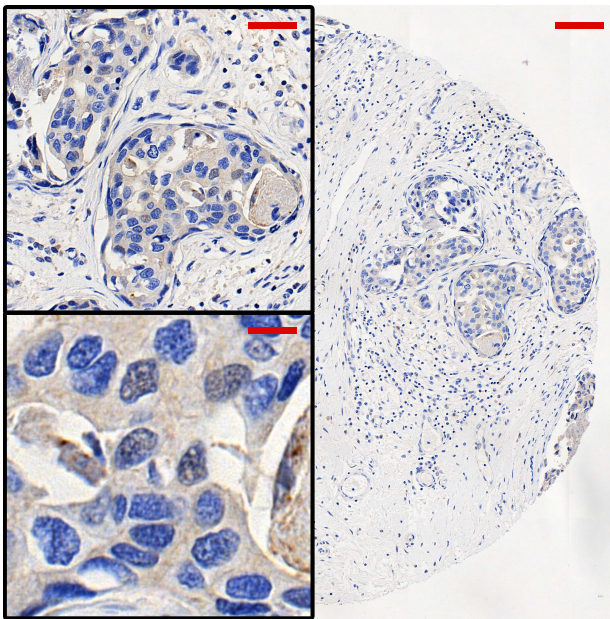
Nuclear (+++) / Cytosolic (+++)



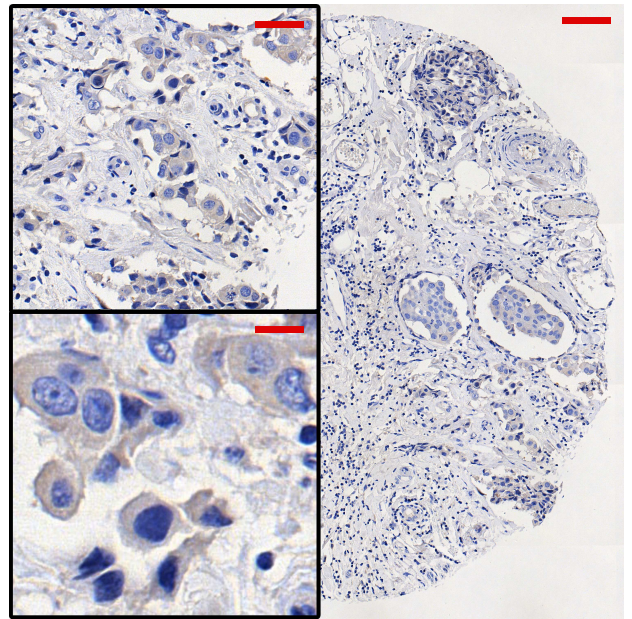
Nuclear (++) / Cytosolic (++)



Nuclear (+) / Cytosolic (++)



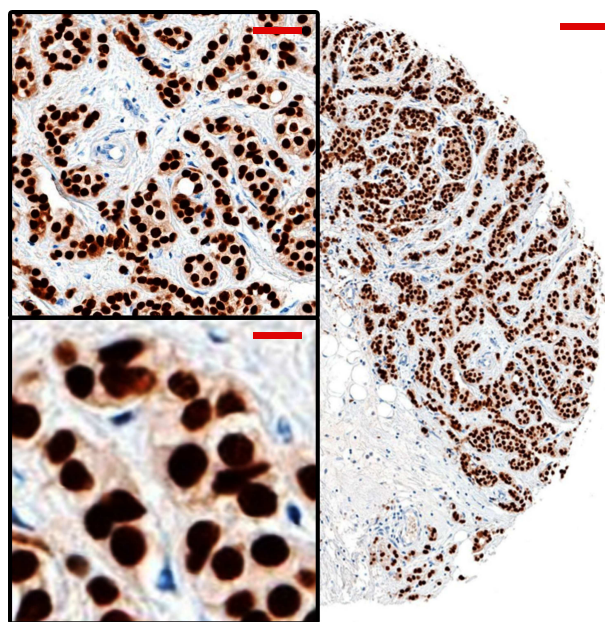
Nuclear (-) / Cytosolic (++)



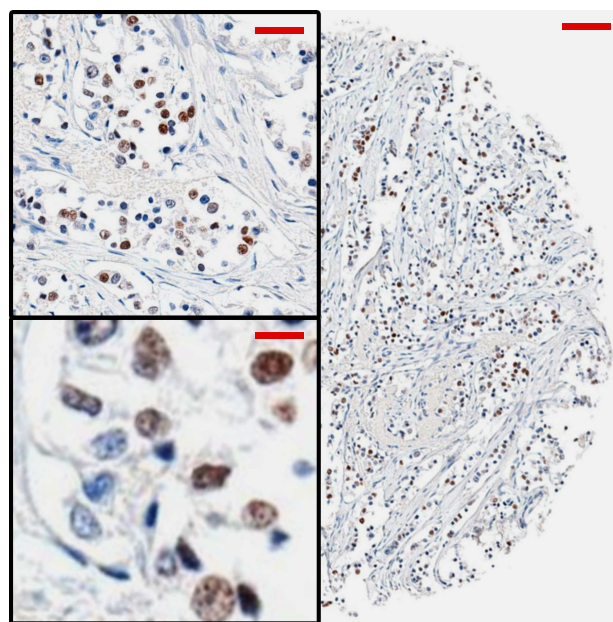
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ER α staining with N-terminal antibody (6F11)

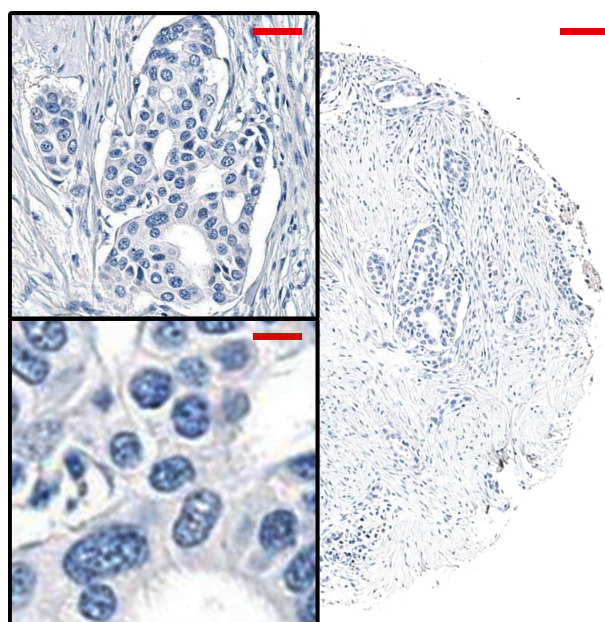
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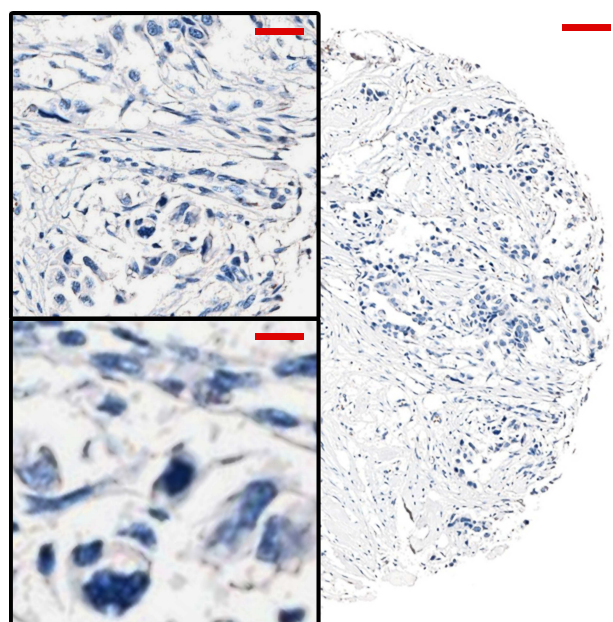
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Nuclear (+) / Cytosolic (-)

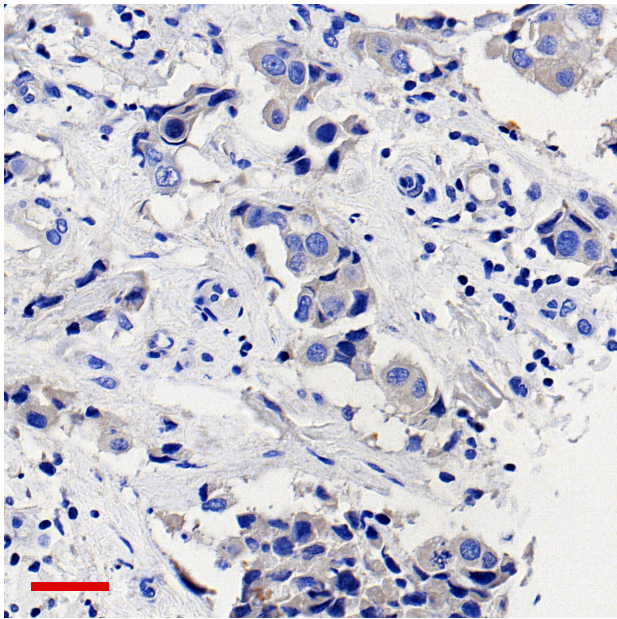


Nuclear (-) / Cytosolic (-)

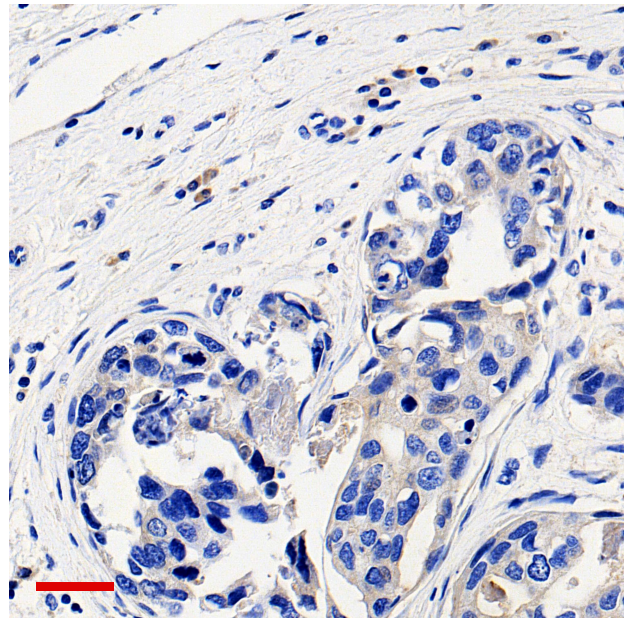


e ER α staining with C-terminal antibody (D8H8) - Cytosolic Grading

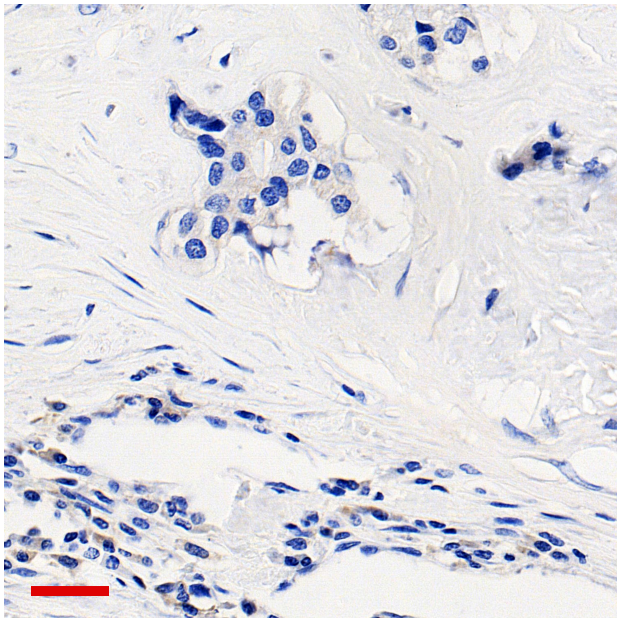
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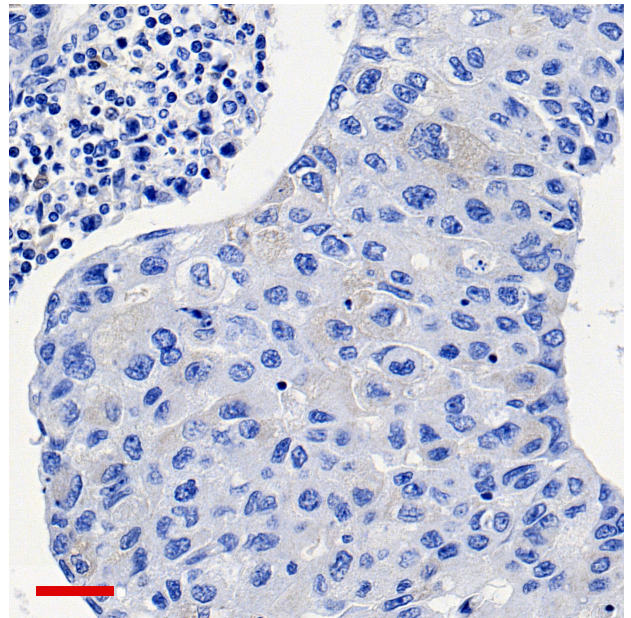
Cytosolic (++)



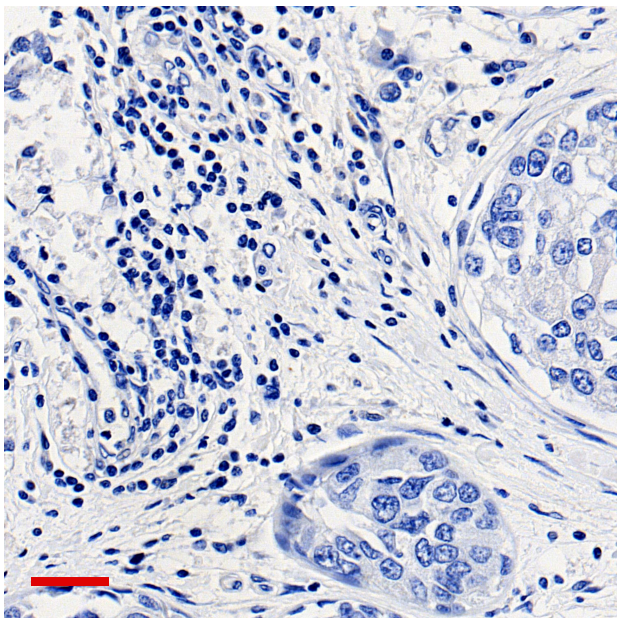
Cytosolic (+)



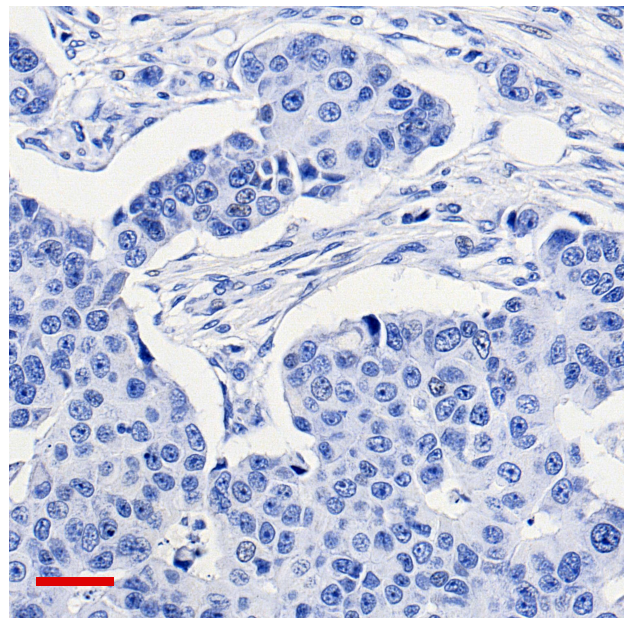
Cytosolic (+)

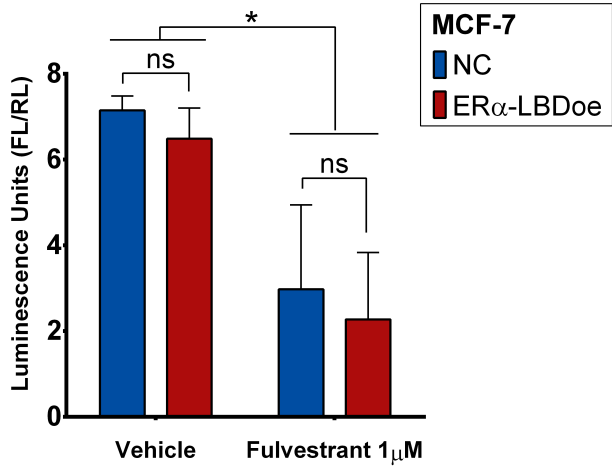
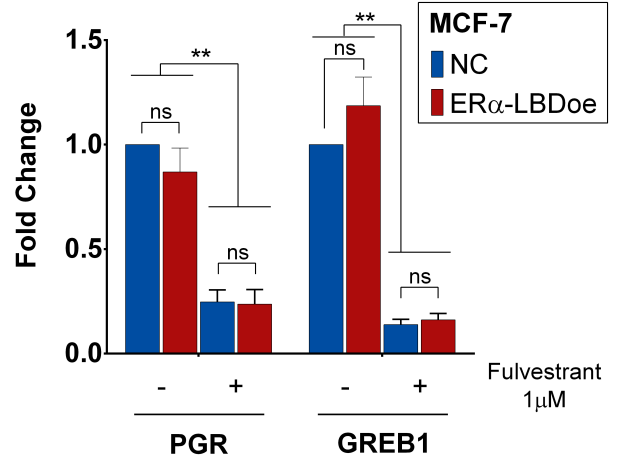
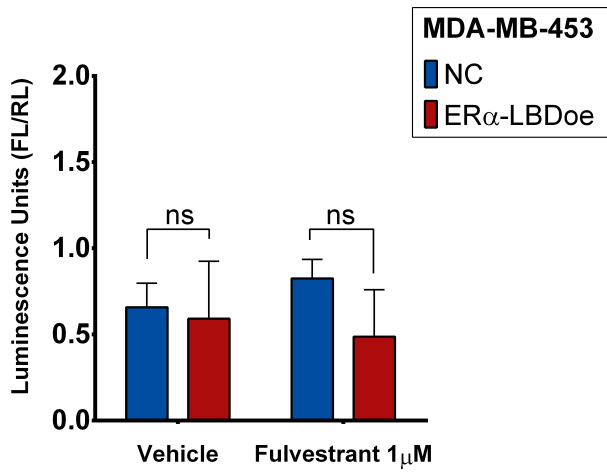
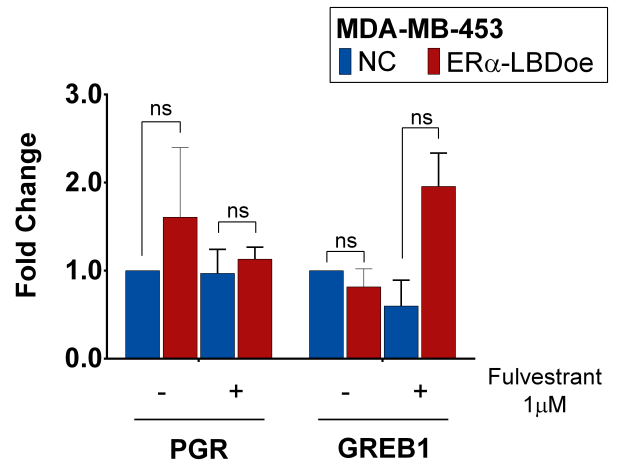
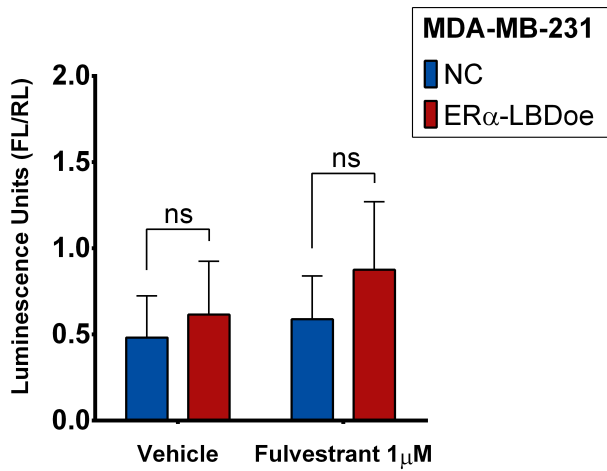
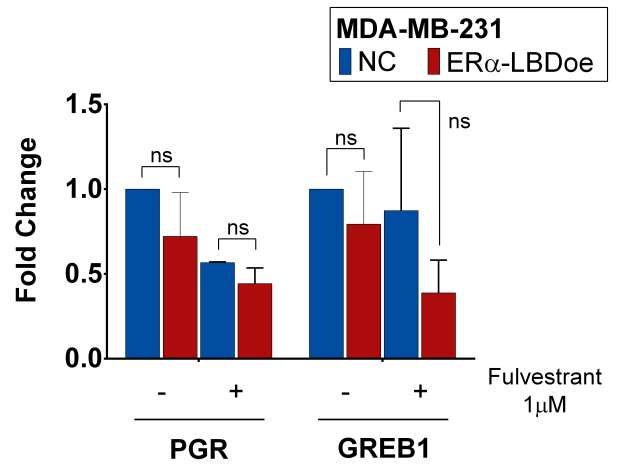


Cytosolic (-)



Cytosolic (-)

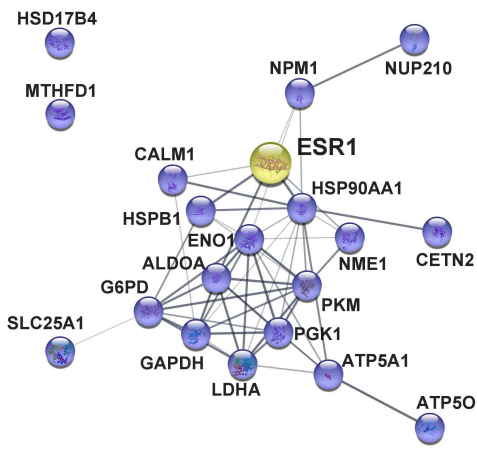


f**g****h****i****j****k**

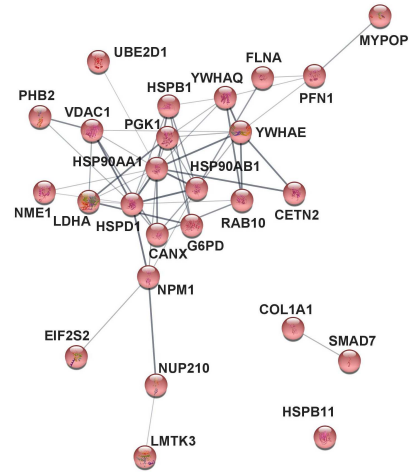
Supplementary Figure 3. a Western Blot analysis of ER α protein variants and their expression in BC cell fractions. HDAC2 was taken as nuclear protein marker whereas VDAC1 was taken as mitochondrial protein marker. Samples from mitochondrial fractions were also treated with proteinase K (50 μ g/ml). **b** Table summarizing statistics from confocal analysis on different BC cell lines. In particular, colocalization data are shown. Blue and red numbers indicate colocalization level of ER α protein into the nucleus or mitochondria of cells, respectively. **c, d** Representative images of ER α staining by immunohistochemistry (IHC) in breast cancer tissue microarray (TMA). Two different ER α antibodies were used: D8H8 (against the C-terminal of the protein) (**c**) and 6F11 (against the N-terminal of the protein) (**d**). All histological sections were counterstained with hematoxylin. Scale bars and magnification: 160 μ m (15X); 40 μ m (63X); 10 μ m (250X). **e** Representative images of ER α cytosolic staining by IHC in BC-TMA by using ER α D8H8 antibody. Different grading is described (++ vs. + vs. -). All histological sections were counterstained with hematoxylin. Scale bar and magnification: 40 μ m (63X). **f, h, j** ERE (Estrogen Responsive Element) promoter-driven luciferase reporter assay in MCF-7, MDA-MB-453 and MDA-MB-231 cells, respectively. Firefly luciferase levels were normalized to Renilla luciferase. Controls (NC) were compared to cells overexpressing ER α -LBD (oe), either in the presence or absence of fulvestrant 1 μ M treatment (24 h). Vehicle = DMSO 0.01%. Data are presented as mean \pm s.e.m. ($n = 3$ independent experiments). * $P < 0.05$, ns = not significant; two-way ANOVA (Sidak's correction). **g, i, k** Expression of PGR and GREB1 mRNA in MCF-7, MDA-MB-453 and MDA-MB-231 cells, respectively. Analysis was carried out by qPCR. Samples and treatments: same as above. Plot shows fold change expression, relative to NC without treatment. In all panels, blue indicates NC samples and red indicates ER α -LBDoe/kd samples. Data are presented as mean \pm s.e.m. ($n = 6$ or $n = 4$ independent experiments). ** $P < 0.01$, ns = not significant; two-way ANOVA (Sidak's correction).

Supplementary Figure 4

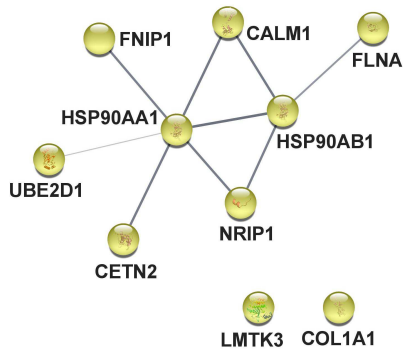
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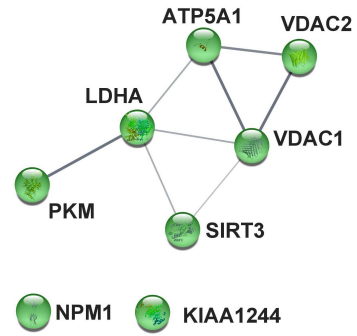
Glycolysis and Gluconeogenesis



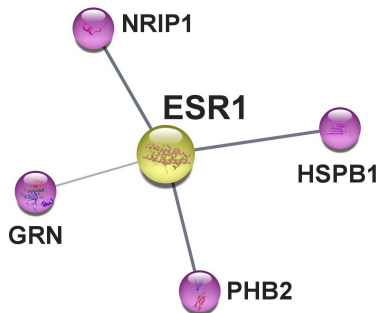
Signaling



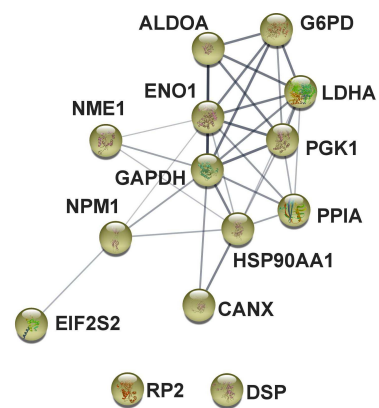
Hypoxia and Angiogenesis



Mitochondrial Metabolism and Respiration

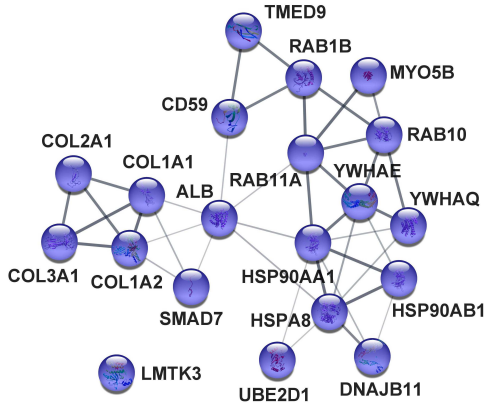


ER α Signaling

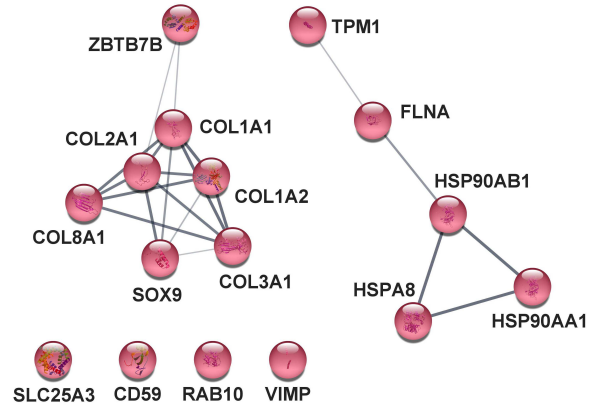


MYC Signaling

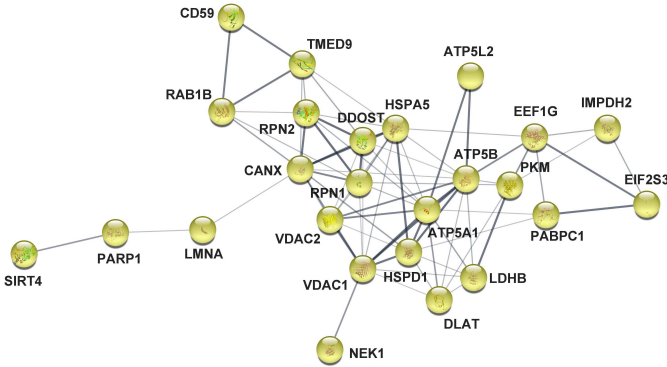
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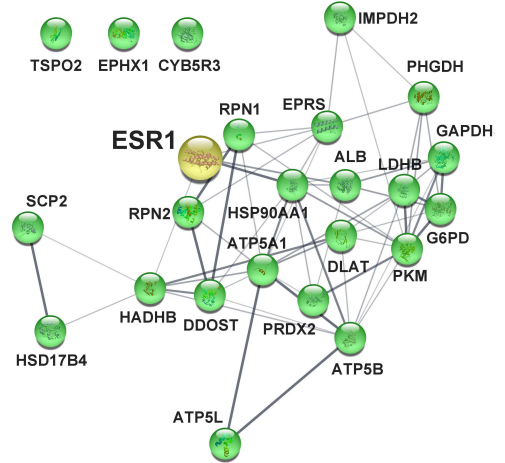
Signaling



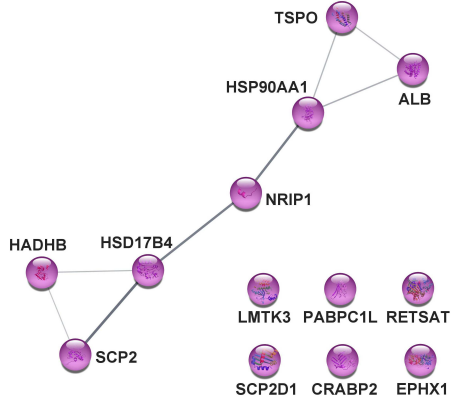
Integrins, Angiogenesis and EMT



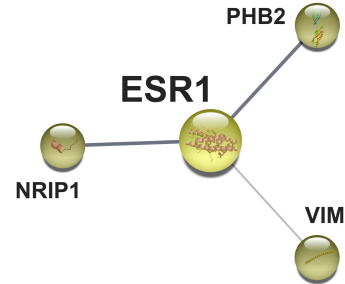
Mitochondrial Metabolism and Respiration



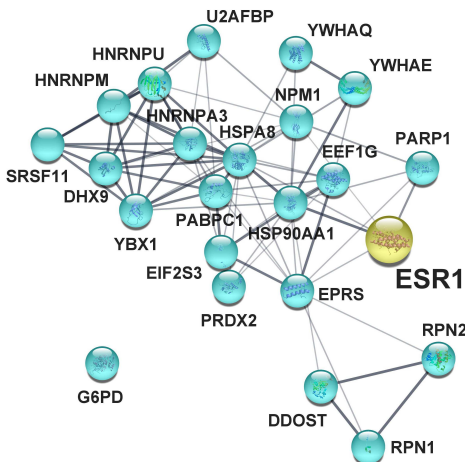
Glycolysis and Gluconeogenesis



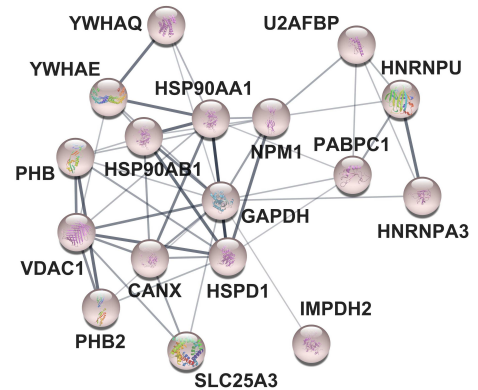
Fatty Acid Metabolism



ER α Signaling



mRNA Splicing

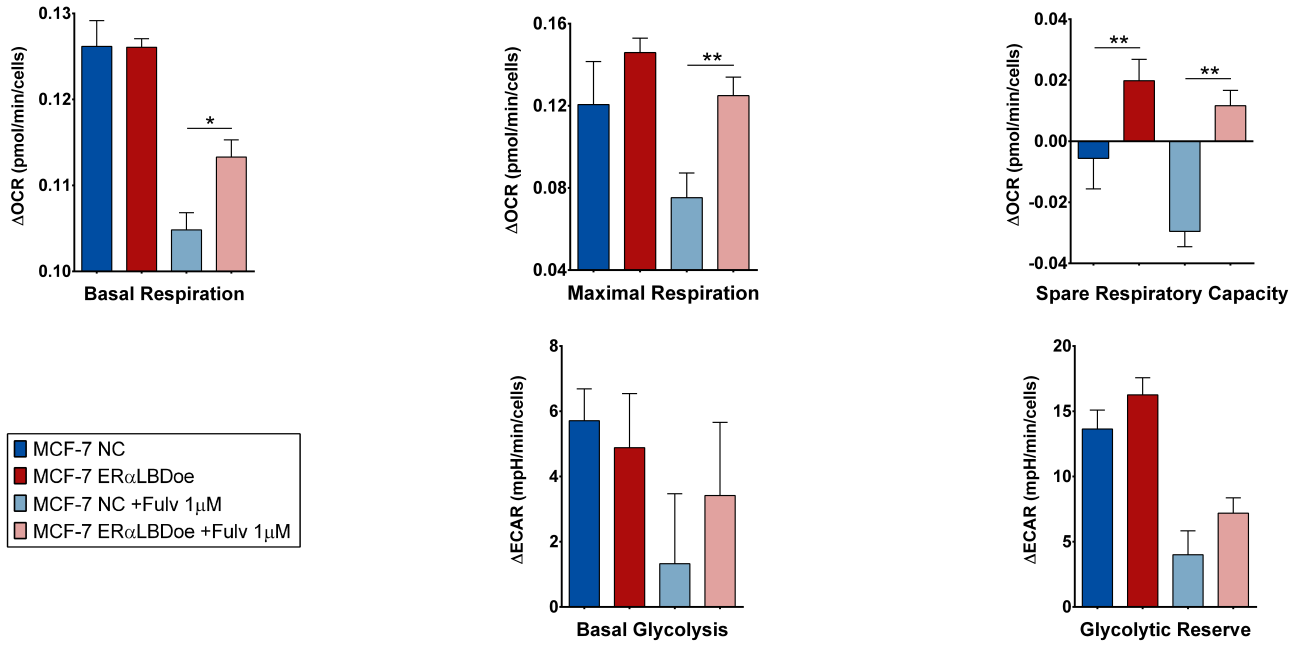


MYC Signaling

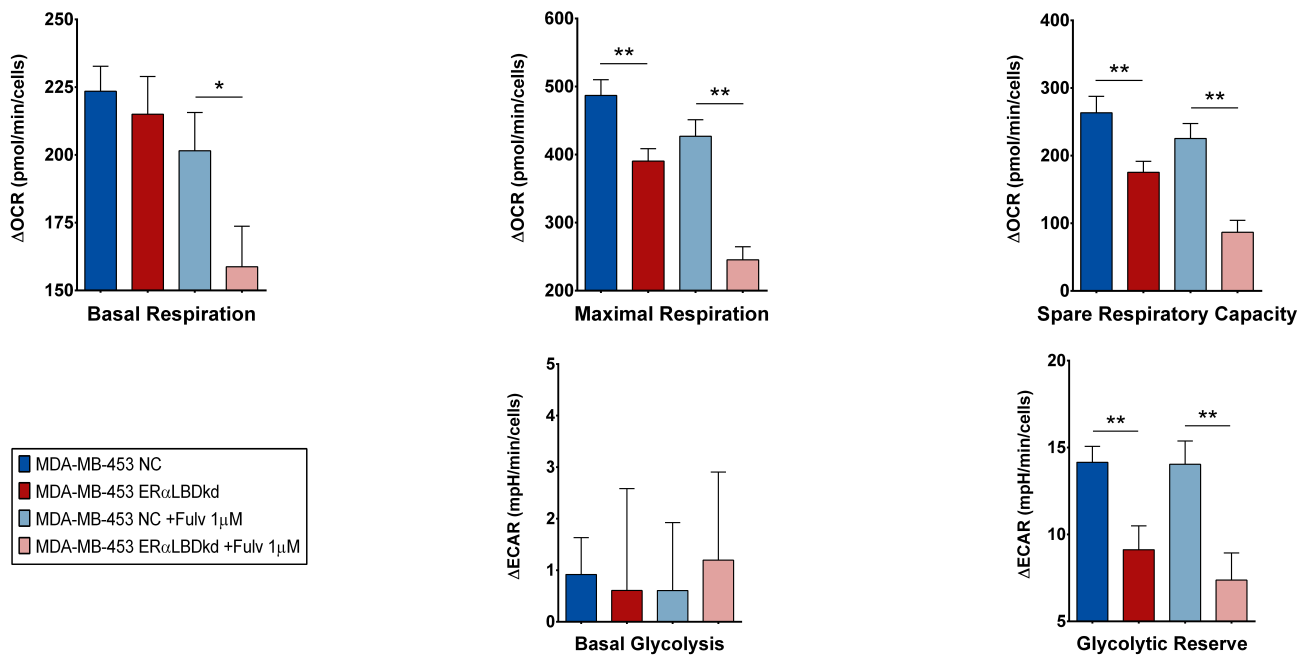
Supplementary Figure 4. a, b Analysis of ER α -LBD protein-protein interaction (PPI) network in MCF-7 and TNBC models respectively, using Cytoscape and STRING software. Proteins are presented as nodes. PPIs networks associated to specific biological functions are presented by using different colors.

Supplementary Figure 5

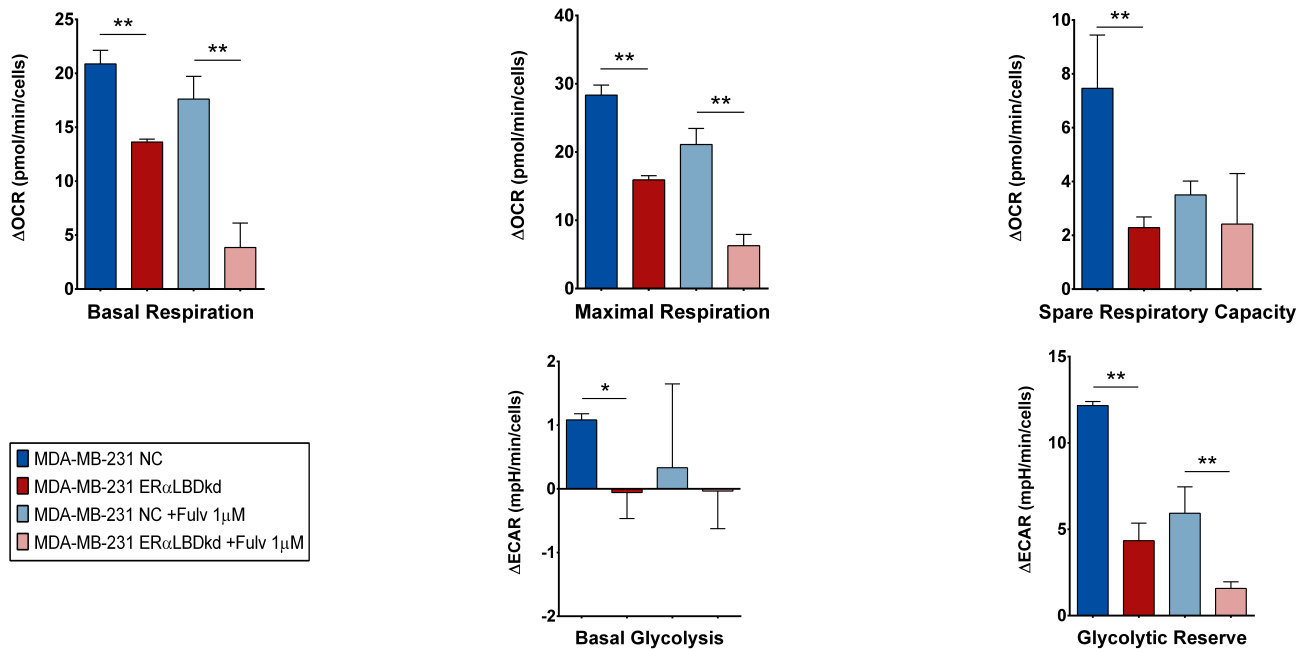
a



b



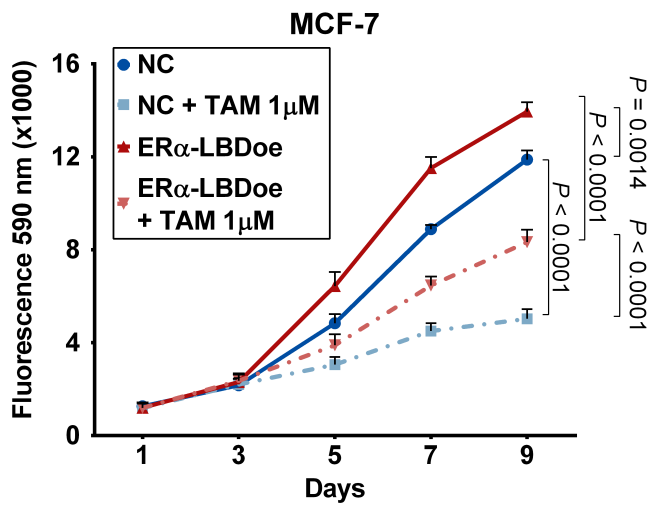
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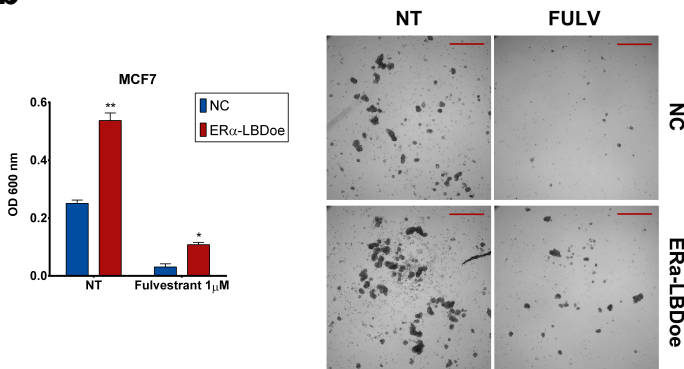
Supplementary Figure 5. a Evaluation of respiratory and glycolytic parameters in MCF-7 cell clones. Fulvestrant 1 μM (24 h pre-treatment) or vehicle (DMSO) was added to cells. ER α -LBD overexpressing (oe) cells were compared to controls (NC). Analyses were carried out by using XF Cell Mito Stress Kit (Agilent). Following manufacturer's guidelines, the calculation of all parameters was based on ΔOCR and ΔECAR values collected during the assay. **b, c** Evaluation of respiratory and glycolytic parameters in TNBC cells (MDA-MB-453/-231). Fulvestrant 1 μM (24 h pre-treatment) or vehicle (DMSO) was added to cells. Cells with ER α -LBD knockdown (kd) were compared to controls (NC). Analysis was carried out as described above. Data in the figures are presented as mean \pm s.e.m. ($n = 2$ independent experiments). * $P < 0.05$, ** $P < 0.01$; unpaired t test, one-sided.

Supplementary Figure 6

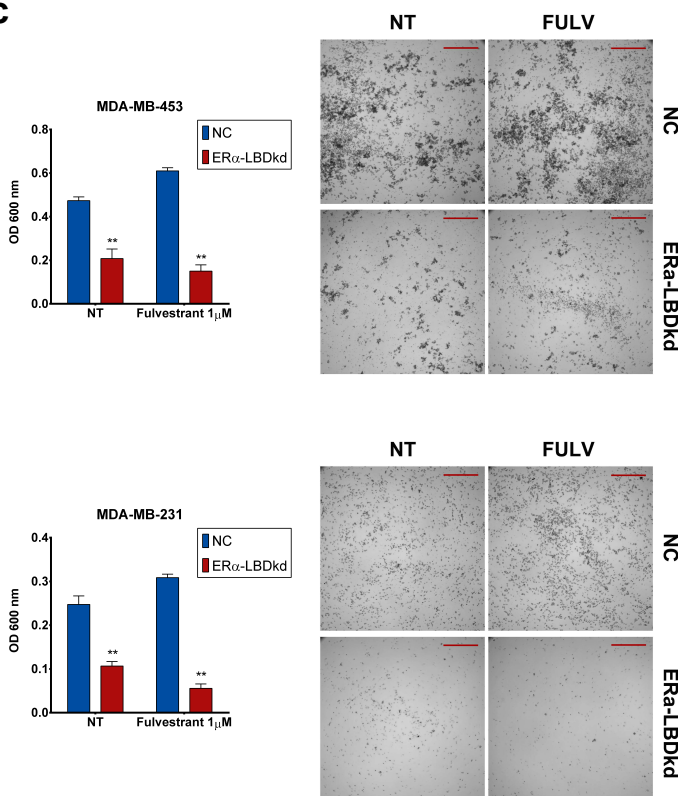
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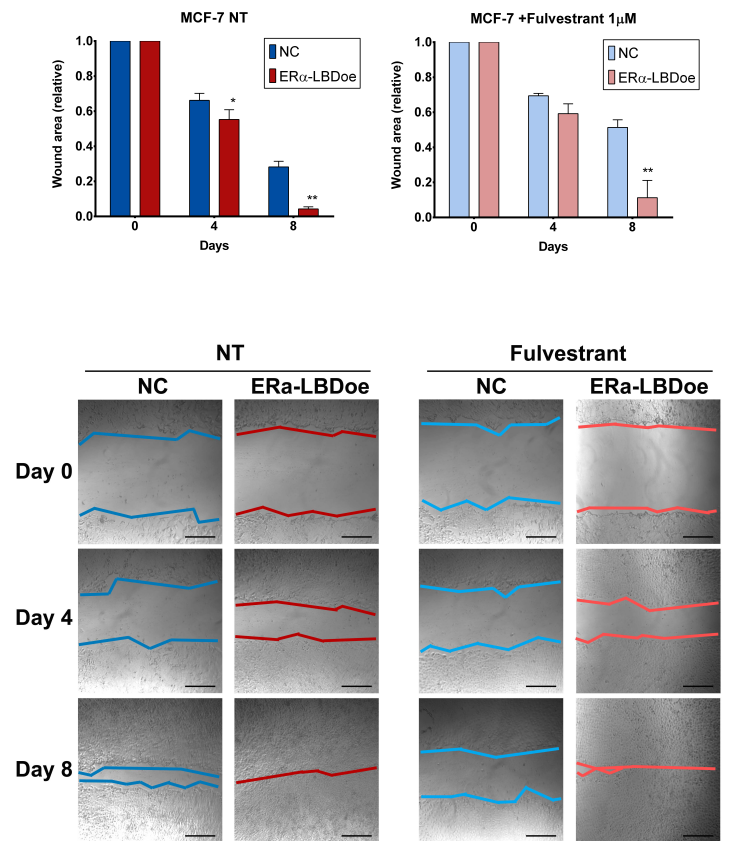
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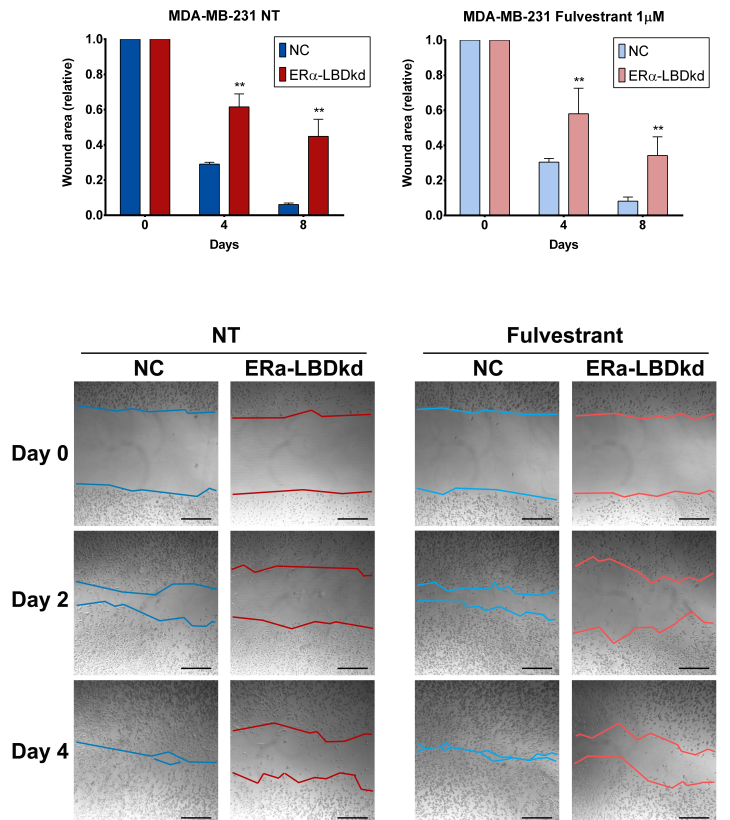
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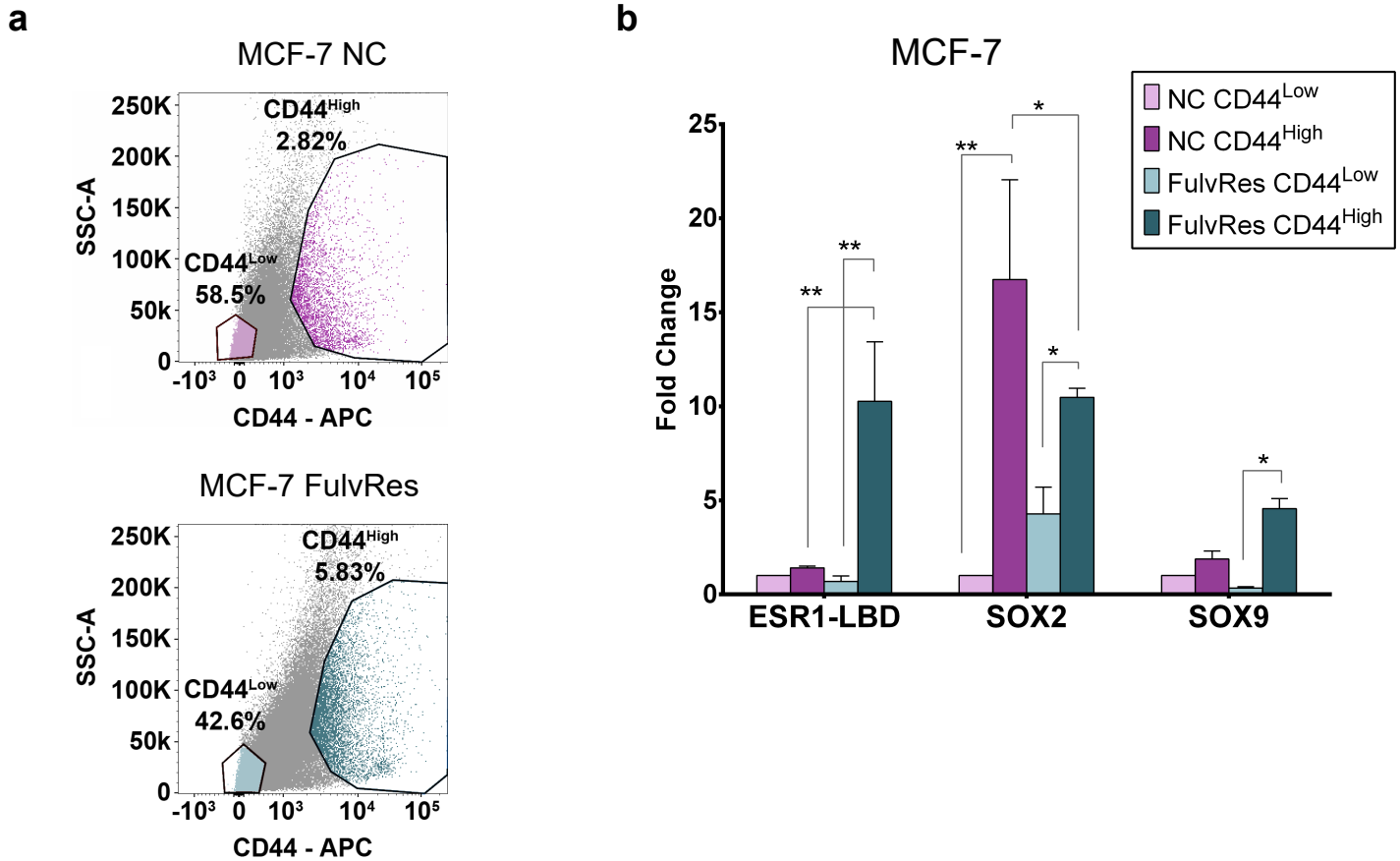


e



Supplementary Figure 6. a *In vitro* proliferation of MCF-7 cell clones, using resazurin reagent and expressed as fluorescence intensity (absorbance at 590 nm). ER α -LBD overexpression was compared to controls (NC) and cell proliferation was tested in the presence or absence of tamoxifen 1 μ M treatment ($n = 3$ independent experiments). All P values describing statistical differences between samples are shown. **b, c** 3-D growth of BC stable clones, measured as optical density (OD) at 600 nm of cell suspensions. ER α -LBD overexpression (MCF-) or knockdown (MDA-MB-453 & 231) was compared to controls (NC), either in the absence (NT) or presence of fulvestrant 1 μ M treatment. Representative images of cell 3-D growth are also shown. Fulv = fulvestrant 1 μ M. Scale bar: 200 μ m. Magnification: 10X. **d, e** Cell migration of BC stable clones was tested by wound healing assay, at different time points (days). Samples and treatments: same as in (**b**). In all panels, blue indicates NC samples and red indicates ER α LBD_{oe/kd} samples. Representative images of scratches are also shown, with wound edges highlighted by different colored lines. Fulv = fulvestrant 1 μ M. Scale bar: 200 μ m. Magnification: 10X. All data in the figure are presented as mean \pm s.e.m. ($n = 3$ independent experiments), * $P < 0.05$, ** $P < 0.01$, two-way ANOVA (Sidak's correction).

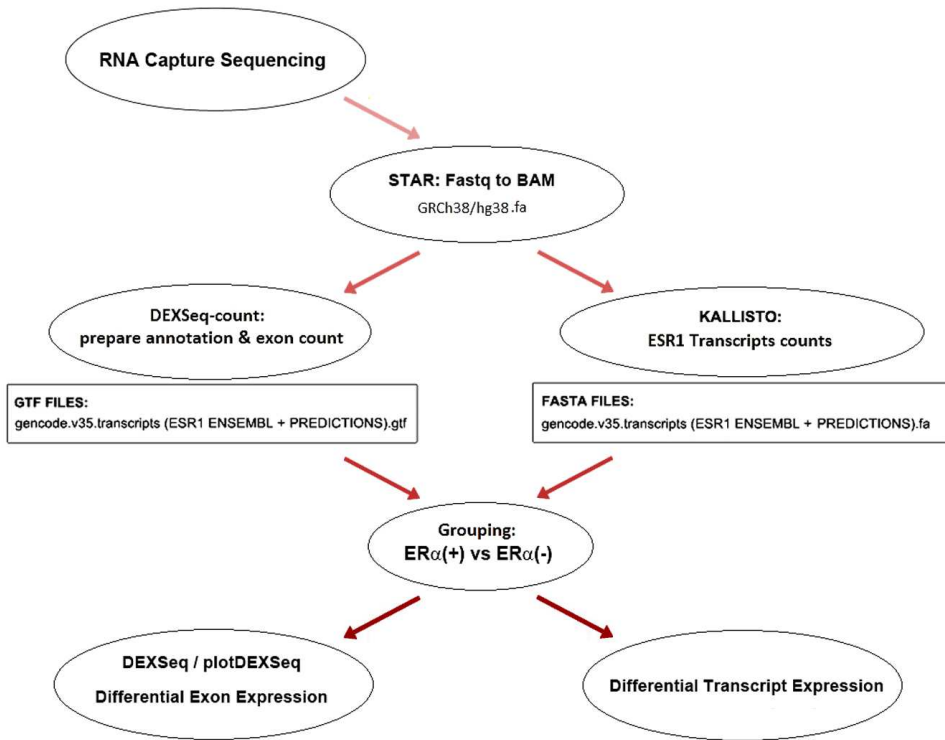
Supplementary Figure 7



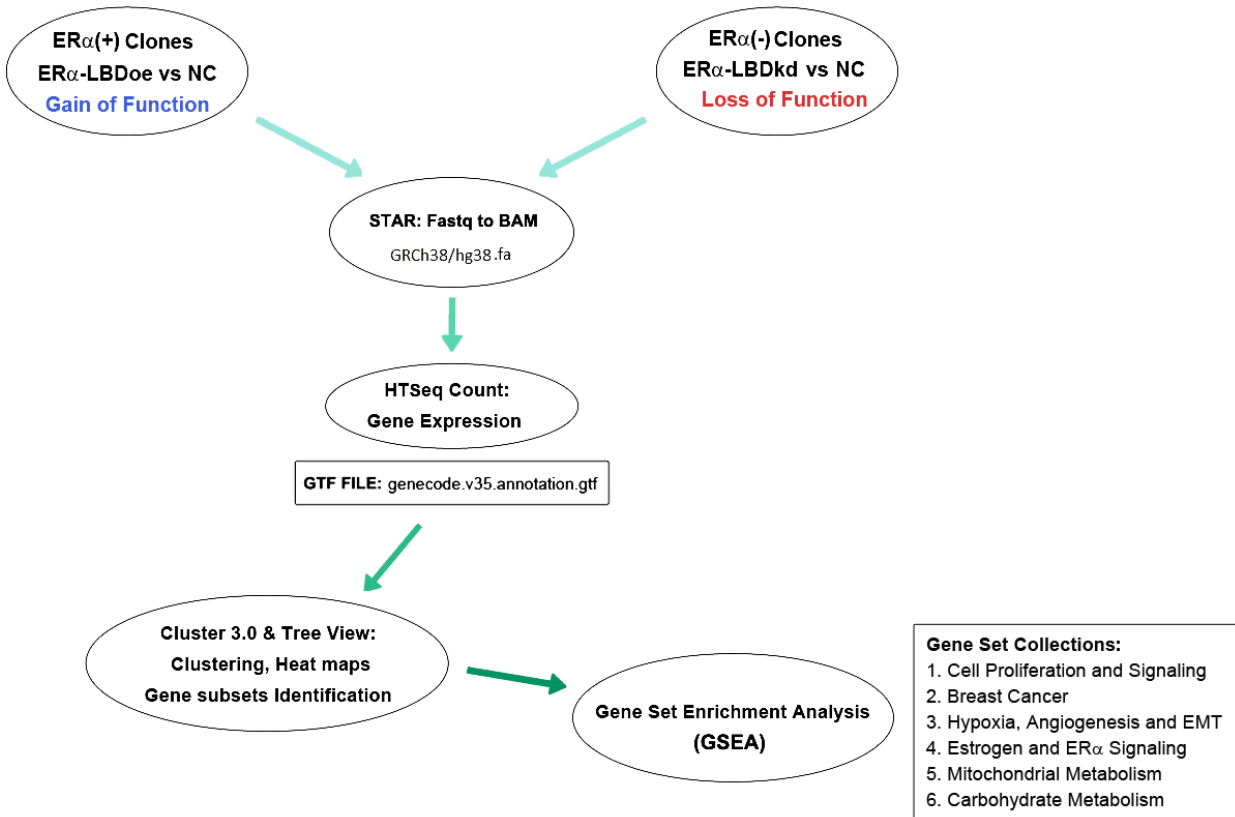
Supplementary Figure 7. **a** Representative images of flow cytometry analysis of CD44^{Low} and CD44^{High} cells, isolated from BC murine xenografts models (MCF-7 and MCF-7 FulvRes). Color code: purple, MCF-7 NC; teal, MCF-7 FulvRes; light, CD44^{Low}; dark, CD44^{High}. **b** Total RNA was extracted from CD44^{Low/High} sorted BC cells and analyzed by qPCR. Plot shows levels of ESR1-LBD (normalized first on RPLP0, then on ESR1-FL) and stem-cell markers Sox2 and Sox9 (normalized on RPLP0), expressed as fold change (CD44^{High} vs. CD44^{Low}). Color code as in (a). Data are presented as mean \pm s.e.m. ($n = 2$ independent experiments; $n = 3$ replicates, each experiment). * $P < 0.05$, ** $P < 0.01$, two-way ANOVA (Fisher's LSD test) and unpaired t test, two-tailed.

Supplementary Figure 8

a



b



Supplementary Figure 8. a Schematic summary of the pipeline used to analyze raw data obtained from RNA capture-sequencing experiment. **b** Schematic summary of the pipeline used to analyze raw data from RNA sequencing experiment on BC cell clones.