



ALMA MATER STUDIORUM
UNIVERSITÀ DI BOLOGNA

ARCHIVIO ISTITUZIONALE
DELLA RICERCA

Alma Mater Studiorum Università di Bologna Archivio istituzionale della ricerca

White grape pomace extracts, obtained by a sequential enzymatic plus ethanol-based extraction, exert antioxidant, anti-tyrosinase and anti-inflammatory activities

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

White grape pomace extracts, obtained by a sequential enzymatic plus ethanol-based extraction, exert antioxidant, anti-tyrosinase and anti-inflammatory activities / Ferri, Maura; Rondini, Greta; Calabretta, Maria Maddalena; Michelini, Elisa; Vallini, Veronica; Fava, Fabio; Roda, Aldo; Minnucci, Giordano; Tassoni, Annalisa. - In: NEW BIOTECHNOLOGY. - ISSN 1871-6784. - STAMPA. - 39:A(2017), pp. S1871678416323901.51-S1871678416323901.58. [10.1016/j.nbt.2017.07.002]

Availability:

This version is available at: <https://hdl.handle.net/11585/615828> since: 2022-03-08

Published:

DOI: <http://doi.org/10.1016/j.nbt.2017.07.002>

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>).
When citing, please refer to the published version.

(Article begins on next page)

This is the final peer-reviewed accepted manuscript of:

*Maura Ferri, Greta Rondini, Maria Maddalena Calabretta, Elisa Michelini, Veronica Vallini, Fabio Fava, Aldo Roda, Giordano Minnucci, Annalisa Tassoni, **White grape pomace extracts, obtained by a sequential enzymatic plus ethanol-based extraction, exert antioxidant, anti-tyrosinase and anti-inflammatory activities**, New Biotechnology, Volume 39, Part A, 2017, Pages 51-58, ISSN 1871-6784*

The final published version is available online at:

<https://doi.org/10.1016/j.nbt.2017.07.002>

Rights / License:

The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>)

When citing, please refer to the published version.

1 **Title**

2 White grape pomace extracts, obtained by a sequential enzymatic plus ethanol-based extraction, exert
3 antioxidant, anti-tyrosinase and anti-inflammatory activities

4

5 **Authors**

6 Maura Ferri^{a,b}, Greta Rondini^a, Maria Maddalena Calabretta^c, Elisa Michelini^c, Veronica Vallini^d,
7 Fabio Fava^b, Aldo Roda^c, Giordano Minnucci^d, Annalisa Tassoni^{a*}

8

9 ^aDepartment of Biological, Geological and Environmental Sciences, University of Bologna, via Irnerio
10 42, 40126 Bologna, Italy;

11 ^bDepartment of Civil, Chemical, Environmental and Materials Engineering, University of Bologna, via
12 Terracini 28, 40131 Bologna, Italy;

13 ^cDepartment of Chemistry G. Ciamician, University of Bologna, via Selmi 2, 40126, Bologna;

14 ^dSadam Engineering Srl, via Agresti 6, 40123 Bologna, Italy;

15

16

17 ***Corresponding author:** Dr. Annalisa Tassoni, Department of Biological, Geological and
18 Environmental Sciences, University of Bologna, via Irnerio 42, 40126 Bologna, Italy.

19 E-mail: annalisa.tassoni2@unibo.it

20

21

22

23

24

25

26

1 **Abstract**

2 The present work aimed at optimizing a two-step enzymatic plus solvent-based process for the
3 recovery of bioactive compounds from white grape (*Vitis vinifera* L., mix of Trebbiano and Verdicchio
4 cultivars) pomace, the winemaking primary by-product. Phenolic compounds solubilised by water
5 enzyme-assisted and ethanol-based extractions of wet (WP) and dried (DP) pomace were characterized
6 for composition and tested for antioxidant, anti-tyrosinase and anti-inflammatory bioactivities. Ethanol
7 treatment led to higher phenol yields than water extraction, while DP samples showed the highest
8 capacity of releasing polyphenols, most probably as a positive consequence of the pomace drying
9 process. Different compositions and bioactivities were observed between water and ethanol extracts
10 and among different treatments and for the first time the anti-tyrosinase activity of *V. vinifera* pomace
11 extracts, was here reported. Enzymatic treatments did not significantly improve the total amount of
12 solubilised compounds; Celluclast in DP led to the recovery of extracts enriched in specific
13 compounds, when compared to control. The best extracts (enzymatic plus ethanol treatment total
14 levels) were obtained from DP showing significantly higher amounts of polyphenols, flavonoids,
15 flavanols and tannins and exerted higher antioxidant and anti-tyrosinase activities than WP total
16 extracts. Conversely, anti-inflammatory capacity was only detected in water (with and without
17 enzyme) extracts, with WP samples showing on average a higher activity than DP. The present
18 findings demonstrate that white grape pomace constitute a sustainable source for the extraction of
19 phytochemicals that might be exploited as functional ingredients in the food, nutraceutical,
20 pharmaceutical or cosmetic industries.

21

22 **Keywords:** Antioxidant activity; anti-tyrosinase activity; by-product; enzymatic digestion; white grape
23 pomace; phenols.

24 **Abbreviations:** AA: ascorbic acid; Abs: absorbance; ABTS: 2'-azino-bis-3-ethylbenzothiazoline-6-
25 sulfonic acid; CAT: catechin; DP: dried pomace; DW: dry weight; GA: gallic acid; KA: kojic acid;
26 WP: wet pomace.

1 **1. Introduction**

2 The large volumes of both liquid and solid by-products generated by the food processing industry
3 poses major environmental and economic challenges. The waste load at a food processing plant can be
4 significantly reduced through the application of new or modified industrial processes and reuse
5 methods especially developed to convert by-products into added-value biomolecules and biofuels [1].
6 In fact, food by-products contain numerous high-value compounds, which could be reused in the food,
7 feed, cosmetic and pharmaceutical industries. Grape (*Vitis* sp.) pomace constitutes the major by-
8 product of the wine making process, representing generally 20-30% of the processed grapes weight
9 [1]. It is mainly made up of pressed skins, seeds and stems of the fruit with a composition that varies
10 considerably depending on grape variety and winemaking technology. Winemakers sometimes
11 produce spirit from grape by-products, but pomace is also traditionally used as animal feed or fertilizer
12 [1, 2]. It has been estimated that more than 9 million tons of grape pomace are generated annually [3],
13 which causes a serious environmental and disposal problem for wineries. However, solid by-products
14 from wine industry are rich in phenolic compounds, in particular flavonoids (such as anthocyanins and
15 flavanols), phenolic acids and stilbenes [4-6], many of which have been shown to be beneficial to
16 human health due to their antioxidant capacity which is believed to contribute to the prevention of
17 cancer and cardiovascular diseases [7]. Anti-inflammatory, anti-microbial and anti-cholesterol
18 properties have also been attributed to grape polyphenols [4, 5, 8]. Due to these properties, there is a
19 vast array of potential applications for grape pomace extractable components: as ingredients of
20 functional foods and feeds, cosmetics and nutraceuticals; and as natural colorants and preservatives of
21 foods [4, 6]. Moreover, nowadays there is a growing interest in finding phytochemicals than can be
22 used as an alternative to synthetic substances, which are sometimes perceived by consumers as
23 harmful to human health [1, 2].

24 Solvent (mainly water, ethanol and methanol) and supercritical fluid extractions are the most
25 efficient in recovering phenolic compounds from grape pomace [2, 9, 10]. Available studies regarding
26 phenolic compositions, extractions and applications are mainly focused on pomace from red grape

1 varieties, whereas little attention has been devoted to white grape pomace, which also contains a wide
2 spectrum of potentially bioactive phenols. Moreover, to the authors' knowledge, phenolic extraction
3 by means of cell wall polysaccharide degrading enzyme mixtures has only been reported for red
4 pomace [3, 5, 11]. In contrast to red wines, white wines are usually made by immediately removing,
5 after grape pressing, the pomace from the must. As a consequence, a much lower phenolic content is
6 present in white wines with respect to red ones. The differences between red and white wine
7 production processes may also affect the amount and availability of polyphenols in the resulting
8 pomace. In fact, as white grape must is not usually fermented in contact with the solid parts of the
9 grape, higher contents of phenolic compounds are present in white compared to red pomace [12, 13].

10 In the present work, enzyme-assisted and ethanol-based extractions were combined in a two-step
11 process aimed at the recovery of phenolic compounds from wet pomace (WP) and dried pomace (DP)
12 of white grapes (*Vitis vinifera* L., mixture of Trebbiano and Verdicchio cultivars). The extracts with
13 the highest amount of compounds were characterised for phenolic composition and tested for
14 antioxidant, anti-tyrosinase and anti-inflammatory bioactivities.

15

16 **2. Materials and Methods**

17

18 *2.1. Materials*

19 White pomace, derived from a mix (60:40) of *V. vinifera* cv. Trebbiano and Verdicchio, was
20 supplied by the Cantine Moncaro wineries (Jesi, Ancona, Italy) immediately after wine production.
21 Pomace was either frozen (wet pomace, WP) or dried (dried pomace, DP) in an industrial vented oven
22 (60°C for 24h) and stored at -20°C until used for analyses. Different WP and DP lots were pulled
23 together before grinding in order to minimise the biological differences.

24

25 *2.2. Enzyme-assisted and ethanol-based pomace extractions*

1 White pomace was treated as previously described by Ferri et al. [5]. WP was ground in a kitchen
2 blender with the addition of distilled water (1:5 g/mL), while DP was ground directly and rehydrated
3 with distilled water (1:5 g/mL for 1h) just before enzymatic digestion. Enzyme-assisted extractions of
4 WP and DP pomace suspensions (20 mL aliquots) were carried out by adding different concentrations
5 (0.5, 1 or 2% enzyme volume/pomace DW) of Pectinex 3XL, Pectinex Ultra SPL, Termamyl,
6 Fungamyl, Pentopan 500BG or Celluclast (Sigma-Aldrich, Milan, Italy). The enzymatic treatments
7 were incubated on an orbital shaker (150 rpm) at different incubation times (2, 6, 24h) and at each
8 enzyme's optimal working temperature (24, 30 or 37°C) [5]. Controls without the addition of enzymes,
9 were also prepared. After incubation, the supernatant (water extract) and pellet fractions were collected
10 [5]. The pellet was extracted overnight with 30 mL of 95% v/v ethanol at 24°C, the supernatant was
11 separated from the pellet and stored at -20°C until further analyses.

12

13 2.3. Extract characterisation

14 Water and ethanol extracts were characterised spectrophotometrically for phenolic [14, 15],
15 flavonoid [15, 16] and flavanol [17] contents. The results were expressed, respectively, as gallic acid
16 (GA) and catechin (CAT) equivalents by means of calibration curves. The amount of tannins was
17 analysed according to Porter et al. [18]; briefly, two aliquots of 0.3 mL of each sample were mixed
18 with 0.9 mL of reagent (50% *n*-butanol, 50% 12N HCl, 0.015% FeCl₃) and incubated for 30 min, one
19 at room temperature and the second at 100°C. Absorbance (Abs) was measured at 550 nm and tannin
20 content (g/L) was calculated as $(Abs_{100^{\circ}C} - Abs_{\text{Room temperature}}) \times 0.1736$.

21 Both water and ethanol supernatants were analysed for phenolic compounds by HPLC-DAD as
22 previously described [5, 19]. The adopted HPLC-DAD separation procedure allowed for the
23 simultaneous analysis of 28 different compounds among stilbenes, phenolic acids and flavonoids [19]:
24 *trans*- and *cis*-resveratrol, *trans*- and *cis*-piceid, *trans*- and *cis*-resveratrolside, piceatannol; gallic,
25 protocatechuic, syringic and vanillic acids; caffeic, chlorogenic, *p*-coumaric, ferulic, sinapic and *trans*-

1 cinnamic acids; catechin, epicatechin, epigallocatechin gallate (EGCG), epicatechin gallate,
2 epigallocatechin; vanillin, naringenin, quercetin, rutin, myricetin and kaempferol.

3

4 *2.4. Determination of biological activities*

5 *In vitro* antioxidant activities were measured using the 2'-azino-bis-3-ethylbenzothiazoline-6-
6 sulfonic acid (ABTS) method with minor modifications [15]. The results were expressed as ascorbic
7 acid (AA) equivalents by means of a dose-response calibration curve.

8 Anti-tyrosinase activity was assessed by an optimised tyrosinase enzyme inhibition assay [20]. The
9 kinetics of brown colour formation was evaluated by absorbance measurement (490 nm) in reactions
10 containing 10 U of tyrosinase and 2 mM L-DOPA (the substrate) in the presence of the sample. The
11 results were expressed as kojic acid (KA, a well-known tyrosinase inhibitor) equivalents by means of a
12 calibration curve (between 1 and 10 µg of KA).

13 A bioluminescent cell-based assay for anti-inflammatory activity was performed using human
14 embryonic kidney HEK293 cells ATCC (American Type Culture Collection, Manassas, VA, USA)
15 routinely grown in Dulbecco modified essential medium (DMEM high glucose 4.5 g/L, GE
16 Healthcare) supplemented with 10% fetal bovine serum, 2 mM L-glutamine, 50 U/µL penicillin, and
17 50 µg/mL streptomycin. HEK293 cells were plated in 96-well plates the day before transfection at a
18 density of 2×10^4 cells/100 µL of growth medium per well. Co-transfection was performed with
19 plasmid pGL4.32[luc2P/NF-κB-RE/Hygro] containing five copies of the NF-κB response element
20 (NF-κB-RE) driving transcription of the luc2P reporter protein (Promega, Madison, WI) and with
21 plasmid pcDNA.3.1-mcherryPRET9 expressing a thermostable mutant *P. pyralis* luciferase [21], and
22 by using FuGENE®HD (Promega) according to the manufacturer's instructions. During co-
23 transfection, cells were incubated for 24h at 37°C with 5% CO₂. After transfection cells were co-
24 incubated for 5h with 100 µL of fresh medium containing sample (1:5 and 1:20 dilutions) and 1 ng/mL
25 TNFα (Sigma Aldrich, Saint Louis, Missouri, USA). Bioluminescent (BL) measurements were
26 performed with a Varioskan™ Flash Multimode Reader acquiring BL signals with band pass green

1 and red filters after injection of 100 μ L substrate BrightGlo (Promega). Corrected values were
2 obtained as previously described [5].

3

4 *2.5. Statistical analyses*

5 All the treatments and assay procedures were performed twice with two biological replicates each.
6 Each repetition was analysed in two technical replicates. The results are expressed as the mean (n=2) \pm
7 SD. Cell-based assays were performed in triplicate, with at least three technical replicates ((n=3) \pm
8 SD). Statistically significant differences between data sets were analysed by using [one-way ANOVA](#)
9 [test followed by post-hoc corrected two tail t-student test assuming equal variance.](#)

10

11 **3. Results**

12

13 *3.1. Enzyme-assisted and ethanol white pomace extractions*

14 Different concentrations (0.5, 1 or 2% v/w) of six commercial enzymes (Pectinex 3XL, Pectinex
15 Ultra SPL, Termamyl, Fungamyl, Pentopan, Celluclast) were tested in enzyme-assisted extractions of
16 both wet pomace (WP) and dried white pomace (DP) in 2h incubations, followed by ethanol
17 extraction. Subsequently, the Folin-Ciocalteu assay was used to quantify phenolic compound content
18 of both aqueous and ethanol extracts (Suppl. Fig. 1). As expected, 95% (v/v) ethanol was able to
19 improve the extraction efficiency of phenolic compounds from the residual pellet with respect to pure
20 water-based (with or without enzyme) treatments (Suppl. Fig. 1). In WP samples, the phenolic content
21 was on average 2.2-fold higher in ethanol compared to water extracts (Suppl. Fig. 1A), while only a
22 1.7-fold average increase was obtained in ethanol extracts of DP samples (Suppl. Fig. 1B). In general,
23 DP contained 2-fold more extractable phenols than WP. Larger variations were observed among WP
24 samples, with total amounts (water plus ethanol extracts) ranging between 940 and 1494 mg GA eq/L
25 (Suppl. Fig. 1A), while DP total phenols were between 1995 and 2437 mg GA eq/L (Suppl. Fig. 1B).
26 Overall, the best treatments (enzymatic plus ethanol extracts total levels) for phenolic compounds were

1 2% Pectinex 3XL 24°C, 2% Pentopan 30°C and 2% Celluclast 37°C (Suppl. Fig. 1). To increase
2 extraction yields, the best enzymatic treatments and related controls for WP and DP were performed at
3 2, 6 and 24h incubation times, and the total level of phenolic compounds in water supernatant and
4 ethanol extracts, were measured (Suppl. Fig. 2). For WP the highest total phenolic contents were
5 detected in samples incubated at 2h or 6h (on average 1316 and 1272 mg GA eq/L, respectively),
6 while a 25% decrease was found in 24h treated samples (Suppl. Fig. 2A). On the other hand, in DP a
7 no statistically significant difference was detected among samples at incubated 2, 6 and 24h (on
8 average 2635, 2659 and 2412 mg GA eq/L, respectively) (Suppl. Fig. 2B).

9

10 3.2. Total phenolic contents

11 On the basis of phenol yield, the following best conditions (two-step enzymatic plus ethanol
12 treatments) for both WP and DP, were selected (Suppl. Fig. 2): for WP 2h 2% Pectinex 3XL 24°C, 2h
13 control 30°C, 6h control 30°C, 6h 2% Pentopan 30°C and 2h control 37°C for WP; for DP 2h control
14 37°C, 2h 2% Celluclast 37°C and 6h 2% Celluclast 37°C.

15 Both water and ethanol extracts were characterised by spectrophotometric assays (Fig. 1). On
16 average, the phenolic content was 1.5-fold higher in ethanol with respect to water extracts and 2.1-fold
17 higher in DP than in WP samples (Fig. 1A). Similar trends were obtained for flavonoids (Fig. 1B),
18 flavanols (Fig. 1C) and tannins (Fig. 1D) contents. Flavanols (such as flavan-3-ols or catechins), a
19 subclass of flavonoids, constituted the majority of flavonoids extracted from Trebbiano and Verdicchio
20 pomace, especially in DP (Fig. 1B-C), confirming that they represent the most abundant phenolic
21 group present in white grape pomace [22]. Moreover, the level of flavanols seemed to decrease with
22 increasing incubation time and/or temperature and following enzyme digestion (Fig. 1C). This can
23 probably be explained considering the fact that the structure of phytochemicals, such as phenolic acid
24 and flavonoids present in fruits, vegetables and grains both in the free and bound forms, is degraded or
25 modified during thermal and non-thermal processing [23]. The maximum level of solubilised total
26 (water plus ethanol extracts) tannins was detected for DP in 2h control 37°C (2120 mg/L) and for WP

1 in 2h control 30°C (1477 mg/L) (Fig. 1D), with a general pattern similar to that observed for flavanols
2 (tannin monomers) (Fig. 1C-D).

3

4 3.3. Characterization of individual phenolic compounds by HPLC-DAD

5 Water and ethanol extracts of the eight best treatments (Fig. 1) were also analysed by HPLC-DAD
6 to quantify specific phenolic compounds (Table 1). In WP samples, the detected phenols were more
7 abundant in ethanol than in water extracts (up to a maximum 36.2-fold difference for *cis*-
8 resveratrolside), with the only exception of gallic acid. In WP, the incubation time increase
9 negatively affected the metabolite extraction yield, while no effect was detected in relation to
10 temperature. In fact, WP 2h control 30°C and WP 2h control 37°C results were not statistically
11 significantly different, while most of the detected compounds in WP decreased after 6h at 30°C when
12 compared to 2h control 30°C. In DP, some phenolic compounds (in particular those belonging to the
13 catechin family) were extracted at higher concentrations in water samples as a consequence of the
14 enzymatic digestions (2h or 6h 2% Celluclast 37°C) when compared to 2h Control 37°C (Table 1).
15 Both water and ethanol solvents were generally able to extract phenolics more efficiently from DP
16 than from WP, such as the up to 110-fold higher amount of catechin in DP water samples compared to
17 the corresponding WP samples. Moreover, a wider spectrum of compounds was detected in DP
18 extracts compared to WP (Table 1). In all the analysed samples, a large amount of the unidentified
19 chromatographic peaks showed an absorbance spectrum characteristic of flavanols [24], thus
20 confirming that these compounds are the main phenolic group present in white grape pomace, as also
21 observed by spectrophotometric analyses (Fig. 1).

22 While most of the detected phenols (Table 1) have been previously found in white pomace extracts
23 from other grape cultivars [11-13, 22, 25], the presence of mono-glycoside stilbenes (*cis*-piceid and
24 *cis*-resveratrolside) is here reported for the first time.

25

26 3.3. Biological activities of the extracts

1 In agreement with spectrophotometrical analyses (Fig. 1), WP samples showed on average a 1.7-
2 fold lower total (water plus ethanol extracts) antioxidant activity than DP samples, while ethanol
3 extracts of both WP and DP showed a higher capacity than water extracts (Fig. 2A). The highest
4 antioxidant activity was detected in the DP 6h 2% Celluclast 37°C sample (water plus ethanol extracts
5 total activity of 7.82 g AA eq/L) (Fig. 2A).

6 The anti-tyrosinase activity of *V. vinifera* pomace extracts was also determined (Fig. 2B). In
7 agreement with biochemical (Fig. 1) and antioxidant capacity results (Fig. 2A), ethanol extracts were
8 more active than water ones (up to 5.3-times in WP 2h 2% Pectinex 24°C), and DP samples were more
9 active than WP ones (on average 2.8-times; 686.3 and 243.3 mg KA eq/L, respectively) (Fig. 2B). The
10 percentage of tyrosinase enzyme inhibition ranged from 43 to 71% in water and from 63 to 79% in
11 ethanol samples.

12 Anti-inflammatory activity of both water and ethanol extracts (1:20 and 1:5 dilutions) was
13 investigated using a cell-based assay. Significant bioactivity was identified for most of the water
14 extracts at the 1:5 dilution (Fig. 3), where a decrease in reporting gene transcription was observed in
15 treated cells in comparison with control. Water WP samples showed on average a higher anti-
16 inflammatory activity than DP. In particular, WP 2h control 37°C was able to decrease TNF α -induced
17 inflammation by 62% (Fig. 3). No activity was reported for ethanol extracts at both dilutions and for
18 water extracts at the 1:20 dilution that showed no variation in transcription levels respect to controls.

19

20 **4. Discussion**

21 In the present paper, the recovery of bioactive phenolic compounds from wet (WP) and dried (DP)
22 white grape (*Vitis vinifera* L.) pomace was investigated by means of different optimised processes
23 including a first extraction step in water (with or without enzymes) followed by a second one in
24 ethanol.

25 DP samples showed the highest capacity of releasing plant compounds, which included high
26 amounts of phenols, flavonoids, flavanols and tannins (Fig. 1). The biochemical compositions of

1 extracts depended on the kind of solvent (water and ethanol) and treatment (with or without enzymes)
2 used (Fig.1 and Table 1).

3 Previous published data showed that the feedstock preserving method had a significant impact on
4 the recovery of phenolic compounds from berry pomace. In particular, drying of red grape pomace
5 caused a significant reduction in the amount of extractable molecules, indicating that moisture content
6 is important to facilitate solvent diffusion and reaching a kinetic equilibrium [5, 10]. In contrast to
7 what has been reported for red pomace [5, 10], the present data (Fig. 1) indicate that, at least for
8 Trebbiano and Verdicchio pomace, drying could be a useful technique to preserve white grape pomace
9 until further industrial application, with the positive consequence of increasing phenol yields after
10 extraction. However, the reduction of moisture content after dehydration, the severity of heat
11 application and the rate of evaporation may influence the stability of water-soluble phenolic
12 compounds, which can result in opposite effects (increase or decrease of recovery) in different samples
13 [23]. These considerations, together with the different metabolite composition of red and white grape
14 winery residues, may explain the different results obtained, after similar extraction treatments, between
15 white (Fig. 1) and red pomace [5].

16 To the authors' knowledge, the present work represents the first study on enzyme-assisted
17 extraction of phenols from white grape pomace. Results indicated that the enzyme treatments did not
18 significantly improve the total amount of solubilised compounds, in agreement to previous results on
19 red grape pomace [5]. Nevertheless, some specific enzyme-treatments produced extracts enriched in
20 specific compounds, such as 2h and 6h 2% Celluclast to DP, and 6h 2% Pentopan to WP (Table 1). In
21 particular, Celluclast is a commercial mixture containing several enzymes (such as ferulic acid esterase
22 and glycosidase) that may selectively release or modify the structure of certain polyphenols thereby
23 also promoting their antioxidant activity [3]. Previous research using cell wall degrading enzymes to
24 solubilise phytochemicals from red grape pomace reported different results depending on plant sample
25 (e.g. cultivar, agricultural and oenological practices) and on adopted extraction process [2, 3, 5, 11].
26 For Sangiovese and Montepulciano [5] and for Merlot red WP [3], increasing incubation times

1 negatively affected metabolite extraction probably due to metabolite degradation during extended
2 incubation. In particular, increasing incubation times (8 to 48h) led to a loss of up to 27% of phenols
3 extracted from Merlot pomace after treatment with or without Celluclast [3]. Present data report a
4 decrease in flavonoids, flavanols and tannins recovery (Fig. 1B-D) during extended incubations,
5 therefore, confirming previous findings.

6 Overall, a comparison between the present results (Fig. 1) and data on red grape pomace obtained
7 by similar methodologies [5], indicate that white grape pomace extracts possess higher levels of
8 phenols (1.4 and 7.7-fold, respectively, for WP and DP; red WP total phenols were on average 990 mg
9 GA eq/L and red DP ones were 393 mg GA eq/L), flavonoids (1.4 and 5.1-fold, respectively, for WP
10 and DP; red WP total flavonoids were on average 882 mg CAT eq/L and red DP ones were 420 mg
11 CAT eq/L) and tannins (2.1 and 5.4-times, respectively, for WP and DP; red WP total tannins were on
12 average 577 mg/L and red DP ones were 350 mg/L) with respect to red grape pomace samples. The
13 present findings are agreement with previous studies performed on four different Chilean varieties and
14 on Vidal Blanc and Viognier cultivars, in which higher polyphenols [12, 13], flavonoids [13] and total
15 proanthocyanidins [12] concentrations, were also detected in white with respect to red grape pomace.
16 Besides the obvious differences related to the used grape cultivars, these results may also be ascribed
17 to a more efficient phenol extraction during the red wine making process in which the fermentation is
18 performed in the presence of the pomace. In contrast, white wine fermentation is generally performed
19 in the absence of berry skins and seeds, thus resulting in a higher content of phenols still present in the
20 pomace.

21 All the extracts exerted antioxidant and anti-tyrosinase activities (Fig. 2). On average, WP samples
22 showed a 1.7-fold lower total antioxidant activity (water plus ethanol extracts) than DP samples. This
23 result could be explained by the fact that high temperature drying treatments have significant effects
24 on the stability and composition of naturally occurring antioxidants [23]. The antioxidant capacity of
25 different white grape pomace extracts was previously measured by using several *in vitro* methods [2].
26 In particular, using the ABTS method, antioxidant activities were reported for aqueous/ethanol pomace

1 extracts of four different white grape varieties from the Balearic Islands (72-134 mg Trolox/gDW
2 equivalent to 0.03-0.05 g Trolox/L) [25], methanol extracts of Palomino Fino and Moscatel marcs (20
3 mM Trolox equivalent corresponding to 5 g Trolox/L) [26], and 80% v/v acetone samples from Vidal
4 Blanc and Viognier cultivars (334-951 μmol Trolox/gDW equivalent to 21-60 g Trolox/L) [13]. In all
5 these studies ABTS scavenging capacity was positively correlated with phenol, flavonoid and
6 condensed tannin contents. A comparison between the antioxidant activity of white (Fig. 2A) and red
7 pomace [5] extracted by the same process, demonstrated on average a 1.6 and 4.9-times higher level
8 respectively, for WP and DP white samples (antioxidant capacities were on average 2.7 and 1.5 g AA
9 eq/L in red WP and DP extracts respectively). This finding is also in agreement with recent
10 investigations of de la Cerda-Carrasco [12], in which the strong oxygen radical scavenging capacity of
11 a white pomace extract (Chardonnay, 80% v/v ethanol extract) and its flavonoid and phenolic contents
12 was positively correlated to the selective *in vivo* inhibition of intestinal α -glucosidases and to the
13 suppression of the postprandial hyperglycemia in diabetic mice, thus also suggesting the potential
14 medical applications of pomace extracts [27].

15 For the first time, the anti-tyrosinase activity of *V. vinifera* pomace extracts is here reported.
16 Tyrosinase is a multifunctional oxidase which catalyses the first two steps of mammalian
17 melanogenesis and is responsible for the enzymatic browning reactions in damaged fruits during post-
18 harvest handling and processing [28]. Both phenomena (hyperpigmentation in human skin and
19 browning in fruits) are considered not desirable and, therefore, new potent tyrosinase inhibitors are
20 actually being sought. One important endpoint for bioactive phytochemical ingredients is in fact their
21 ability to decrease skin pigmentation and/or to inhibit fruit browning during post-harvest processing.
22 This feature has been related to many different phenolic compounds. Most of the identified compounds
23 in the present extracts (Table 1) are known tyrosinase inhibitors [25-27], such as quercetin and gallic
24 acid, having the highest inhibitory activity, followed by resveratrol and catechin derivatives and
25 protocatechuic acid with the lowest inhibitory effect. However, depending on their structure,
26 polyphenols may act both as inhibitors or substrates of tyrosinase [28]. Regarding grape extracts, anti-

1 tyrosinase capacity was previously measured in aqueous/ethanol extracts from pomace of two cultivars
2 of *Vitis rotundifolia* [29] and in *V. vinifera* seed and peel extracts [30]. In particular, the latter showed
3 an inhibition of tyrosinase ranging from 15 to 35%, about half of that measured in Verdicchio and
4 Trebbiano pomace extracts (between 43 and 71% inhibition in water extracts and from 63 to 79% in
5 ethanol samples).

6 Anti-inflammatory capacity was only detected in water extracts (Fig. 3), probably as a consequence
7 of a different phenolic composition of the samples. These data are consistent with previous studies on
8 animal models demonstrating that white and red grape pomace extracts were able to suppress, in a
9 dose-dependent manner, chronic inflammation induced by lipopolysaccharide (LPS) and
10 galactosamine (GalN) in N13 microglia cells [8] and in Sprague-Dawley rats [31]. Although more
11 investigations are required to evaluate if the reported anti-inflammatory activity is concentration-
12 dependent, the present results seem promising and lead us to hypothesise a possible use of pomace
13 extracts as ingredients or supplements for functional foods. Future sample characterization will enable
14 to investigate if the single components, such as quercetin, catechin, epicatechin and gallic acid, are
15 responsible for such activity.

16 In conclusion, the present results strongly support a possible application of white grape pomace
17 extracts, obtained by a two-step process, as an inexpensive, easily available and alternative source of
18 bioactive phytochemicals, which could be advantageously used as ingredients in the food,
19 nutraceutical, pharmacological or cosmetic fields.

20

21 **Acknowledgment**

22 This work was supported by Sadam Engineering (Sadam Eridania Spa group, Bologna, Italy). Prof.
23 Aldo Roda was financed by the Emilia Romagna Region (Italy) project “Dai Distretti Produttivi ai
24 Distretti Tecnologici”, DGR 1631/2009. We are very thankful to Prof. Bruce R. Branchini
25 (Connecticut College, New London, CT) for providing the pcDNA.3.1-mcherryPRET9 plasmid and to
26 Dr. Marianne Louise van Buuren for the editing of the English language.

1 **References**

- 2
- 3 [1] Djilas S, Canadanovic-Brunet J, Cetkovic G. By-products of fruits processing as a source of
4 phytochemicals. *Chem Ind & Chem Eng Q* 2009; 15:191-202.
- 5 [2] Fontana AR, Antonioli A, Bottini R. Grape pomace as a sustainable source of bioactive compounds:
6 extraction, characterization and biotechnological applications of phenolics. *J Agric Food Chem* 2013;
7 61:8987-9003.
- 8 [3] Meyer AS, Jepsen SM, Sørensen NS. Enzymatic release of antioxidants for human low-density
9 lipoprotein from grape pomace. *J Agric Food Chem* 1998; 46:2439-2446.
- 10 [4] Yu J, Ahmedna M. Functional components of grape pomace: their composition, biological properties
11 and potential applications. *Int J Food Sci Technol* 2013; 48:221-237.
- 12 [5] Ferri M, Bin S, Vallini V, Fava F, Michelini E, Roda A, Minnucci G, Bucchi G, Tassoni A. Recovery
13 of polyphenols from red grape pomace and assessment of their antioxidant and anti-cholesterol activities.
14 *New Biotech* 2016; 33:338-344.
- 15 [6] Ignat I, Volf I, Popa VI. A critical review of methods for characterisation of polyphenolic compounds
16 in fruit and vegetables. *Food Chem* 2011; 126:1821-1835.
- 17 [7] Del Rio D, Rodriguez-Mateos A, Spencer JP, Tognolini M, Borges G, Crozier A. Dietary
18 (poly)phenolics in human health: structures, bioavailability, and evidence of protective effects against
19 chronic diseases. *Antiox Redox Signal* 2013; 18:1818-1892.
- 20 [8] Rodriguez-Morgado B, Candiracci M, Santa-Maria C, Revilla E, Gordillo B, Parrado J, Castaldo A.
21 Obtaining from grape pomace and enzymatic extract with anti-inflammatory properties. *Plant Foods*
22 *Hum Nutr* 2015; 70:42-49.
- 23 [9] Brazinha C, Cadima M, Crespo JG. Optimization of extraction of bioactive compounds from
24 different types of grape pomace produced at wineries and distilleries. *J Food Sci* 2014; 79:E1142-E1149.
- 25 [10] Monrad JK, Srinivas K, Howard LR, King JW. Design and optimization of a semicontinuous hot-
26 cold extraction of polyphenols from grape pomace. *J Agric Food Chem* 2012; 60:5571-5582.

- 1 [11] Kammerer D, Claus A, Schieber A, Carle R. A novel process for the recovery of polyphenols from
2 grape (*Vitis vinifera* L.) pomace. J Food Sci 2005; 70:C157-C163.
- 3 [12] de la Cerda-Carrasco A, Lopez-Solis R, Nunez-Kalasic H, Pena-Neira A, Obreque-Slier E. Phenolic
4 composition and antioxidant capacity of pomaces from four grape varieties (*Vitis vinifera* L.). J Sci Food
5 Agric 2015; 95:1521-1527.
- 6 [13] Xu Y, Burton S, Kim C, Sismour E. Phenolic compounds, antioxidant, and antibacterial properties
7 of pomace extracts from four Virginia-grown grape varieties. Food Sci Nutr 2016; 4:125-133.
- 8 [14] Singleton VL, Orthofer R, Lamuela-Raventos RM. Analysis of total phenols and other oxidation
9 substrates and antioxidants by means of Folin-Ciocalteu reagent. Methods Enzymol 1999; 299:152-178.
- 10 [15] Ferri M, Gianotti A, Tassoni A. Optimisation of assay conditions for the determination of
11 antioxidant capacity and polyphenols in cereal food components. J Food Comp Anal 2013; 30:94-101.
- 12 [16] Zhishen J, Mengcheng T, Jianming W. The determination of flavonoid in mulberry and their
13 scavenging effects on superoxide radicals. Food Chem 1999; 64:555-559.
- 14 [17] McMurrough I, McDowell J. Chromatographic separation and automated analysis of flavanols.
15 Anal Biochem 1978; 91:92-100.
- 16 [18] Porter LJ, Hrstich LN, Chan BG. The conversion of procyanidins and prodelphinidins to cyanidin
17 and delphinidin. Phytochemistry 1986; 25:223-230.
- 18 [19] Ferri M, Tassoni A, Franceschetti M, Righetti L, Naldrett MJ, Bagni N. Chitosan treatment induces
19 changes of protein expression profile and stilbene distribution in *Vitis vinifera* cell suspensions.
20 Proteomics 2009; 9:610-624.
- 21 [20] Masmoto Y, Iida S, Kubo M. Inhibitory effect of Chinese crude drugs on tyrosinase. Planta Med
22 1980; 40:361-365.
- 23 [21] Branchini BR, Ablamsky DM, Davis AL, Southworth TL, Butler B, Fan F, Jathoul AP, Pule MA.
24 Red-emitting luciferases for bioluminescence reporter and imaging applications. Anal Biochem 2010;
25 396:290-297.

- 1 [22] Jara-Palacios MJ, Hernanz D, Escudero-Gilete ML, Heredia FJ. Antioxidant potential of white
2 grape pomaces: phenolic composition and antioxidant capacity measured by spectrophotometric and
3 cyclic voltammetry methods. *Food Res Int* 2014; 66:150-157.
- 4 [23] Nayak B, Liu RH, Tang J. Effect of processing on phenolic antioxidants of fruits, vegetables, and
5 grains--a review. *Crit Rev Food Sci Nutr* 2015; 55:887-919.
- 6 [24] Atomssa T, Gholap AV. Characterization and determination of catechins in green tea leaves using
7 UV-visible spectrometer. *J Eng Technol Res* 2015; 7:22-31.
- 8 [25] Gonzalez-Centeno MR, Jourdes M, Femenia A, Simal S, Rossello C, Teissedre PL. Characterization
9 of polyphenols and antioxidant potential of white grape pomace by-products (*Vitis vinifera* L.). *J Agric*
10 *Food Chem* 2013; 61:11579-11587.
- 11 [26] Alonso AM, Guillen DA, Barroso CG, Puertas B, Garcia A. Determination of antioxidant activity
12 of wine by-products and its correlation with polyphenolic content. *J Agric Food Chem* 2002; 50:5832-
13 5836.
- 14 [27] Hogan S, Zhang L, Li JR, Sun S, Canning C, Zhou KQ. Antioxidant rich grape pomace extract
15 suppresses postprandial hyperglycemia in diabetic mice by specifically inhibiting alpha-glucosidase.
16 *Nutr Metab* 2010; 7:71-80.
- 17 [28] Chang TS. An updated review of tyrosinase inhibitors. *Int J Mol Sci* 2009; 10:2440-2475.
- 18 [29] Plundrich N, Grace MH, Raskin I, Lila MA. Bioactive polyphenols from muscadine grape and
19 blackcurrant stably concentrated onto protein-rich matrices for topical applications. *Int J Cosmetic Sci*
20 2013; 35:394-401.
- 21 [30] Hsu CK, Chou ST, Huang PJ, Mong MC, Wang CK, Hsueh YP, Jhan JK. Crude ethanol extracts
22 from grape seeds and peels exhibit anti-tyrosinase activity. *J Cosmet Sci* 2012; 63:225-232.
- 23 [31] Nishiumi S, Mukai R, Ichianagi T, Ashida H. Suppression of lipopolysaccharide and
24 galactosamine-induced hepatic inflammation by red grape pomace. *J Agric Food Chem* 2012; 60:9315-
25 9320.

26

1 **Supplementary material captions**

2

3 **Supplementary Figure 1.** Levels of phenolic compounds in water supernatants and ethanol extracts of
4 (A) wet pomace (WP) and (B) dried pomace (DP) samples treated for 2h with different concentrations
5 (0.5, 1 or 2% enzyme volume/pomace DW) of six commercial enzymes and controls. The data are
6 expressed as mg of gallic acid (GA) equivalent per litre of extract (mg GA eq/L). The star symbol
7 indicates a statistically significant difference (one way ANOVA followed by post hoc two tail
8 Student's *t*-test, $p < 0.05$) between the total level (water plus ethanol extracts) of each enzymatic treated
9 sample with respect to the control incubated at the same temperature (24, 30 or 37°C). Data are the
10 mean \pm SD (n=2).

11

12 **Supplementary Figure 2.** Levels of phenolic compounds in water supernatants and ethanol extracts of
13 the three best treatments and the three controls of (A) wet pomace (WP) and (B) dried pomace (DP)
14 samples incubated for increasing times (2, 6 and 24h). The data are expressed as mg of gallic acid
15 (GA) equivalent per litre of extract (mg GA eq/L). The star symbol indicates a statistically significant
16 difference (one way ANOVA followed by post hoc two tail Student's *t*-test, $p < 0.05$) between the total
17 level (water plus ethanol extracts) of the treatments incubated for 6h and 24h in comparison to the
18 same treatment at 2h. Data are the mean \pm SD (n=2).

19

1 **Figure Captions**

2

3 **Figure 1.** Levels of total (water plus ethanol extracts) (A) phenols, (B) flavonoids, (C) flavanols and
4 (D) tannins in water and ethanol extracts of the five best wet pomace (WP) and the three best dried
5 pomace (DP) samples. Phenol data are expressed as mg of gallic acid (GA) equivalent per L of extract
6 (mg GA eq/L); flavonoid and flavanol data are expressed as mg of catechin (CAT) equivalent per L of
7 extract (mg CAT eq/L); tannins are expressed as mg of tannin per L. Different letters indicate
8 statistically significant difference (one way ANOVA followed by post hoc two tail Student's *t*-test,
9 $p < 0.05$) between data total levels (water plus ethanol extracts). Data are the mean \pm SD (n=2).

10

11 **Figure 2.** (A) Antioxidant and (B) anti-tyrosinase activities in selected wet pomace (WP) and dried
12 pomace (DP) extracts. Antioxidant data were obtained by means of the ABTS method and are
13 expressed as g of ascorbic acid (AA) equivalent per L of extract (g AA eq/L). Anti-tyrosinase data are
14 expressed as mg of kojic acid (KA) equivalent per L of extract (mg KA eq/L). Different letters indicate
15 statistically significant difference (one way ANOVA followed by post hoc two tail Student's *t*-test,
16 $p < 0.05$) between data total levels (water plus ethanol extracts). Data are the mean \pm SD (n=2).

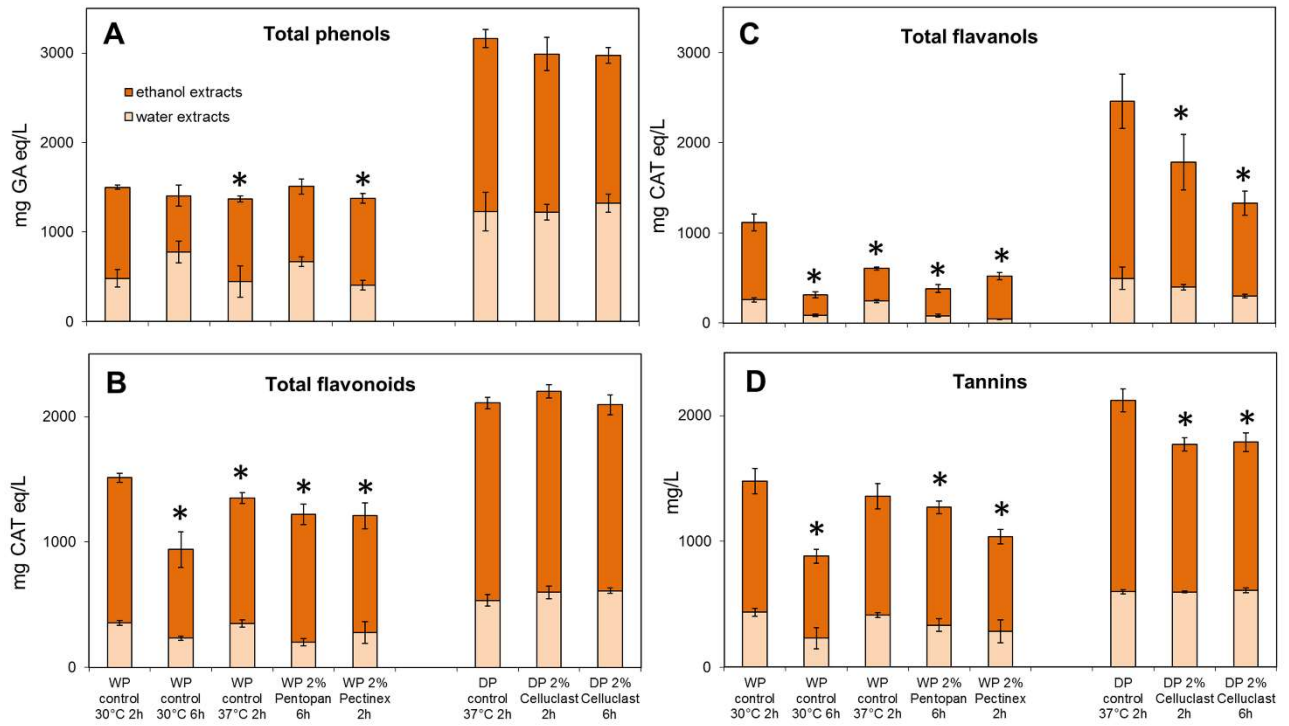
17

18 **Figure 3.** Anti-inflammatory activity obtained with HEK293 cells co-transfected with
19 pGL4.32[luc2P/NF- κ B-RE/Hygro] and pcDNA.3.1-mcherryPRET9 and co-incubated for 5h at 37°C
20 with 1:5 WP and DP water sample dilutions in the presence of 1 ng/mL TNF α . Control sample (H₂O
21 Control) for inflammatory activity was obtained by incubating cells with 1 ng/mL TNF α and the BL
22 signal of the control well was set at 1 to normalize results and enable both evaluation of anti- and pro-
23 inflammatory activities. Different letters indicate statistically significant difference (one way ANOVA
24 followed by post hoc two tail Student's *t*-test, $p < 0.05$) between data total levels (water plus ethanol
25 extracts) and the H₂O Control. Data are the mean \pm SD (n=3).

1 **Table 1.** Polyphenol levels (mg/L) of water and ethanol extracts of wet (WP) and dried (DP) white grape pomace determined by HPLC-DAD analysis.
 2 Different letters indicate statistically significant difference (one way ANOVA followed by post hoc two tail Student's *t*-test, $p < 0.05$) between data.
 3 Data are the mean \pm SD (n=2).

Compound	Type of extract	WP 2h 2% Pectinex 3XL 24°C	WP 2h Control 30°C	WP 6h Control 30°C	WP 6h 2% Pentopan 30°C	WP 2h Control 37°C	DP 2h Control 37°C	DP 2h 2% Celluclast 37°C	DP 6h 2% Celluclast 37°C
Catechin	Water	0.26 \pm 0.02 ^a	0.65 \pm 0.02 ^b	0.34 \pm 0.03 ^a	0.48 \pm 0.06 ^c	0.67 \pm 0.01 ^b	45.66 \pm 2.01 ^d	55.43 \pm 5.42 ^{d,e}	57.61 \pm 1.29 ^e
	Ethanol	3.39 \pm 0.22 ^a	5.76 \pm 1.79 ^b	3.38 \pm 0.14 ^a	3.20 \pm 0.24 ^a	5.18 \pm 0.93 ^b	31.62 \pm 0.94 ^c	32.29 \pm 3.50 ^c	31.64 \pm 2.25 ^c
Epicatechin	Water	0.62 \pm 0.04 ^a	0.86 \pm 0.04 ^b	0.70 \pm 0.06 ^a	0.86 \pm 0.05 ^b	0.83 \pm 0.15 ^b	31.86 \pm 0.17 ^c	41.39 \pm 3.92 ^d	42.90 \pm 1.30 ^d
	Ethanol	2.97 \pm 0.22 ^a	4.64 \pm 0.70 ^b	2.69 \pm 0.20 ^a	3.04 \pm 0.01 ^a	4.28 \pm 0.75 ^b	26.96 \pm 0.03 ^c	27.47 \pm 1.39 ^c	27.26 \pm 0.34 ^c
Epigallocatechin gallate	Water	1.32 \pm 0.51 ^a	2.99 \pm 0.08 ^b	1.47 \pm 0.12 ^a	2.03 \pm 0.08 ^a	2.28 \pm 1.32 ^{a,b}	19.27 \pm 1.27 ^c	45.06 \pm 18.60 ^d	48.69 \pm 5.58 ^d
	Ethanol	8.31 \pm 0.72 ^a	13.58 \pm 2.86 ^b	8.15 \pm 2.21 ^{a,b}	9.62 \pm 0.67 ^a	10.62 \pm 1.44 ^{a,b}	35.42 \pm 0.47 ^c	36.23 \pm 3.16 ^c	37.39 \pm 1.80 ^c
Epicatechin gallate	Water	0.45 \pm 0.03 ^a	0.65 \pm 0.02 ^b	0.49 \pm 0.04 ^a	0.57 \pm 0.05 ^b	0.68 \pm 0.11 ^b	2.12 \pm 0.76 ^c	2.81 \pm 0.22 ^c	3.03 \pm 0.14 ^c
	Ethanol	7.15 \pm 0.12 ^a	10.00 \pm 1.17 ^{a,b}	8.05 \pm 1.64 ^a	10.45 \pm 0.11 ^b	10.50 \pm 0.20 ^b	13.97 \pm 0.13 ^c	13.33 \pm 0.75 ^c	12.90 \pm 0.36 ^c
Epigallocatechin	Water	0.22 \pm 0.01 ^a	0.31 \pm 0.01 ^b	0.26 \pm 0.08 ^{a,b}	0.30 \pm 0.02 ^b	0.32 \pm 0.01 ^b	4.82 \pm 0.16 ^c	5.44 \pm 3.36 ^{c,d}	7.82 \pm 0.56 ^d
	Ethanol	7.94 \pm 0.05 ^a	9.66 \pm 3.75 ^a	8.22 \pm 1.70 ^a	10.64 \pm 0.81 ^{a,b}	10.74 \pm 0.09 ^b	35.74 \pm 0.18 ^c	39.33 \pm 2.53 ^c	36.37 \pm 0.05 ^c
Gallic acid	Water	0.20 \pm 0.01 ^a	0.45 \pm 0.05 ^{b,c}	0.25 \pm 0.01 ^a	0.37 \pm 0.03 ^b	0.56 \pm 0.06 ^c	22.73 \pm 0.66 ^d	23.70 \pm 0.63 ^d	26.62 \pm 0.76 ^d
	Ethanol	-	-	-	-	-	-	-	-
Protocatechuic acid	Water	-	-	-	-	-	1.89 \pm 0.09 ^a	2.74 \pm 0.54 ^b	2.41 \pm 0.02 ^b
	Ethanol	-	-	-	-	-	1.01 \pm 0.13 ^a	1.48 \pm 0.03 ^b	1.50 \pm 0.18 ^b
Chlogenic acid	Water	-	-	-	-	-	9.21 \pm 0.61 ^a	10.85 \pm 1.38 ^{a,b}	12.57 \pm 1.08 ^b
	Ethanol	-	-	-	-	-	-	-	-
Cis-Piceid	Water	0.33 \pm 0.01 ^a	0.48 \pm 0.03 ^b	0.35 \pm 0.01 ^a	0.42 \pm 0.01 ^b	0.48 \pm 0.07 ^b	1.81 \pm 0.65 ^c	2.30 \pm 0.16 ^c	2.42 \pm 0.13 ^c
	Ethanol	10.56 \pm 0.82 ^a	13.61 \pm 0.18 ^b	9.20 \pm 1.21 ^a	13.98 \pm 0.22 ^b	12.54 \pm 0.05 ^b	20.32 \pm 1.16 ^c	23.69 \pm 0.30 ^c	24.22 \pm 0.19 ^c
Cis- Resveratrolside	Water	0.15 \pm 0.01 ^a	0.18 \pm 0.01 ^a	0.16 \pm 0.01 ^a	0.19 \pm 0.01 ^a	0.18 \pm 0.01 ^a	1.45 \pm 0.01 ^b	0.74 \pm 0.09 ^c	0.70 \pm 0.21 ^c
	Ethanol	5.44 \pm 0.30 ^a	6.89 \pm 0.69 ^b	4.79 \pm 0.61 ^a	7.38 \pm 0.09 ^b	6.83 \pm 0.51 ^b	11.68 \pm 0.24 ^c	13.31 \pm 0.89 ^c	12.21 \pm 0.23 ^c
Quercetin	Water	-	-	-	-	-	0.46 \pm 0.07 ^a	0.55 \pm 0.05 ^{a,b}	0.67 \pm 0.05 ^b
	Ethanol	-	-	-	-	-	-	-	-

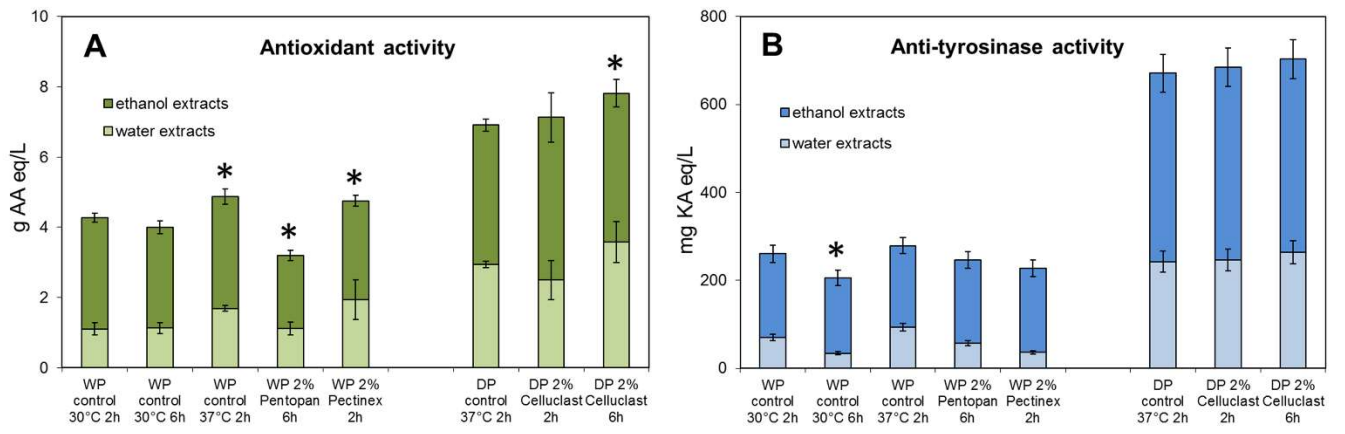
1 **Figure 1.**



2

3

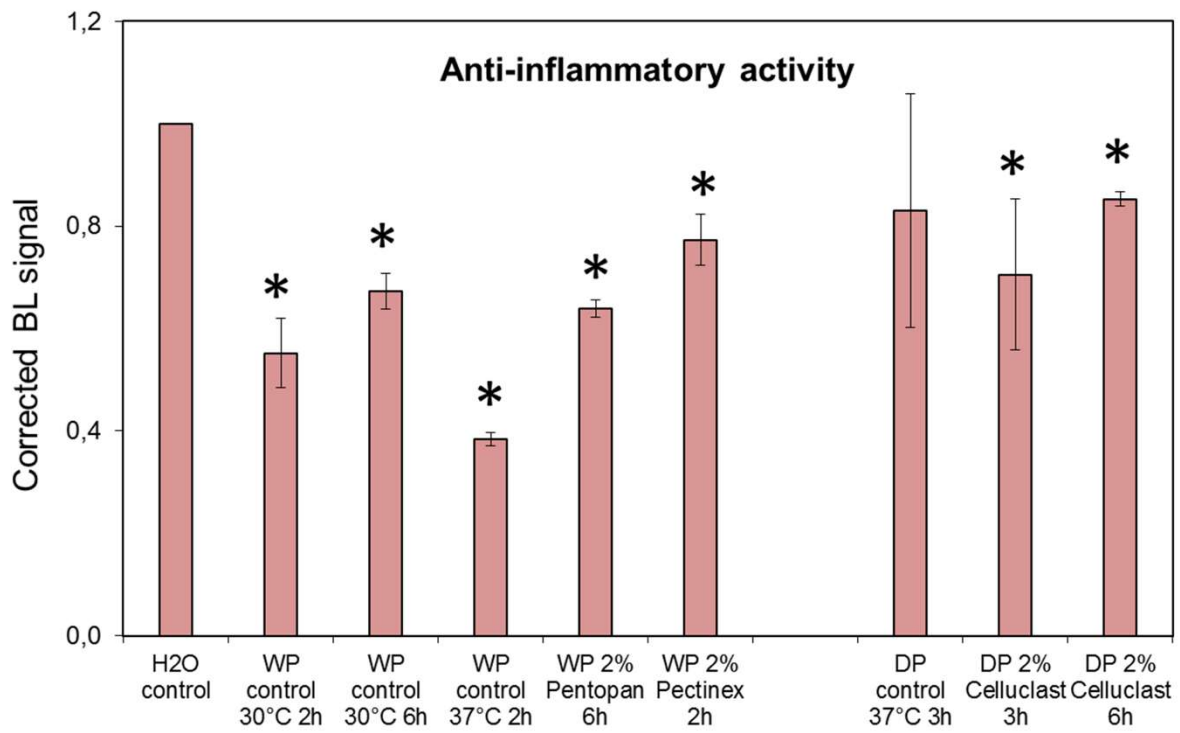
1 **Figure 2.**



2

3

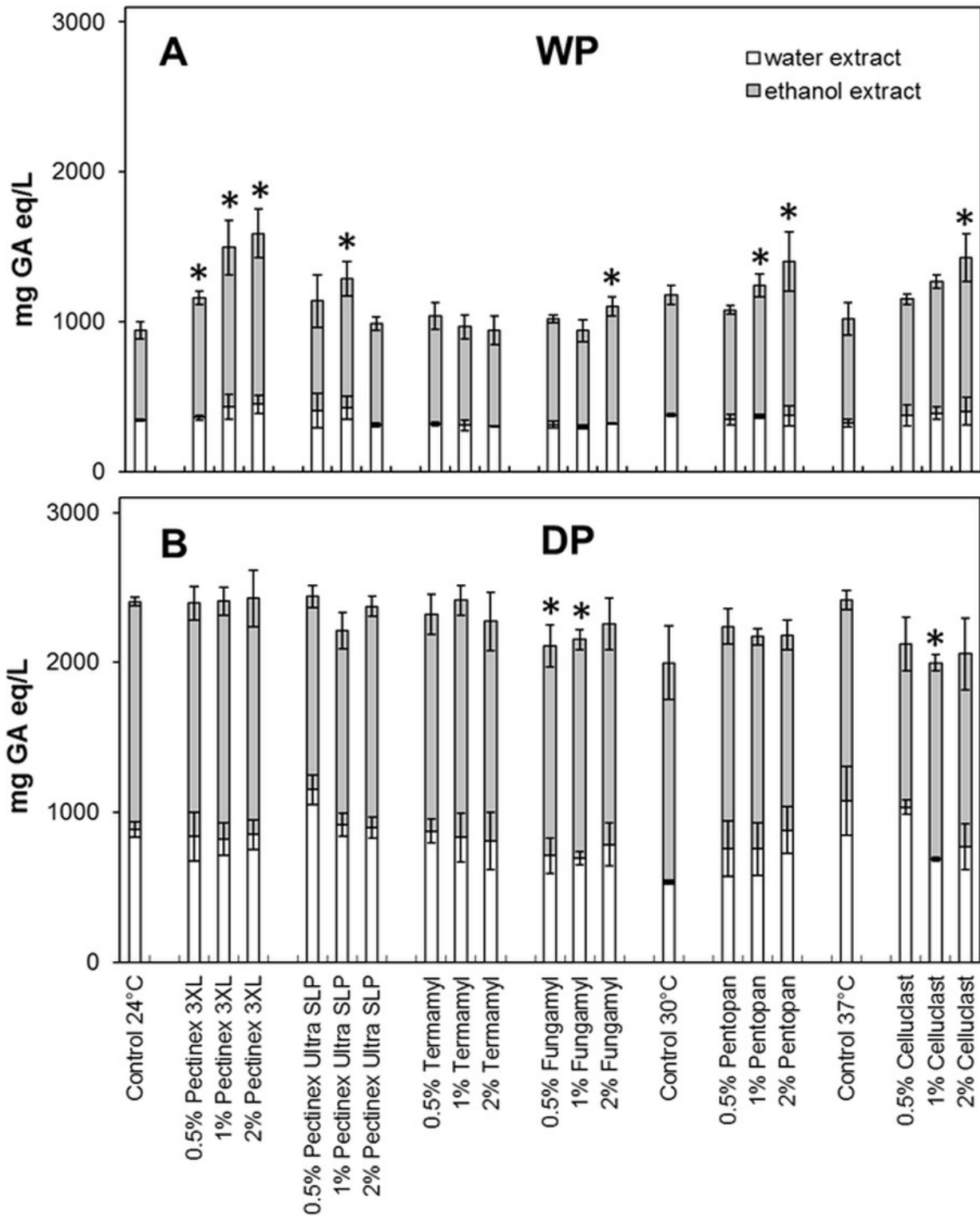
1 **Figure 3.**



2

3

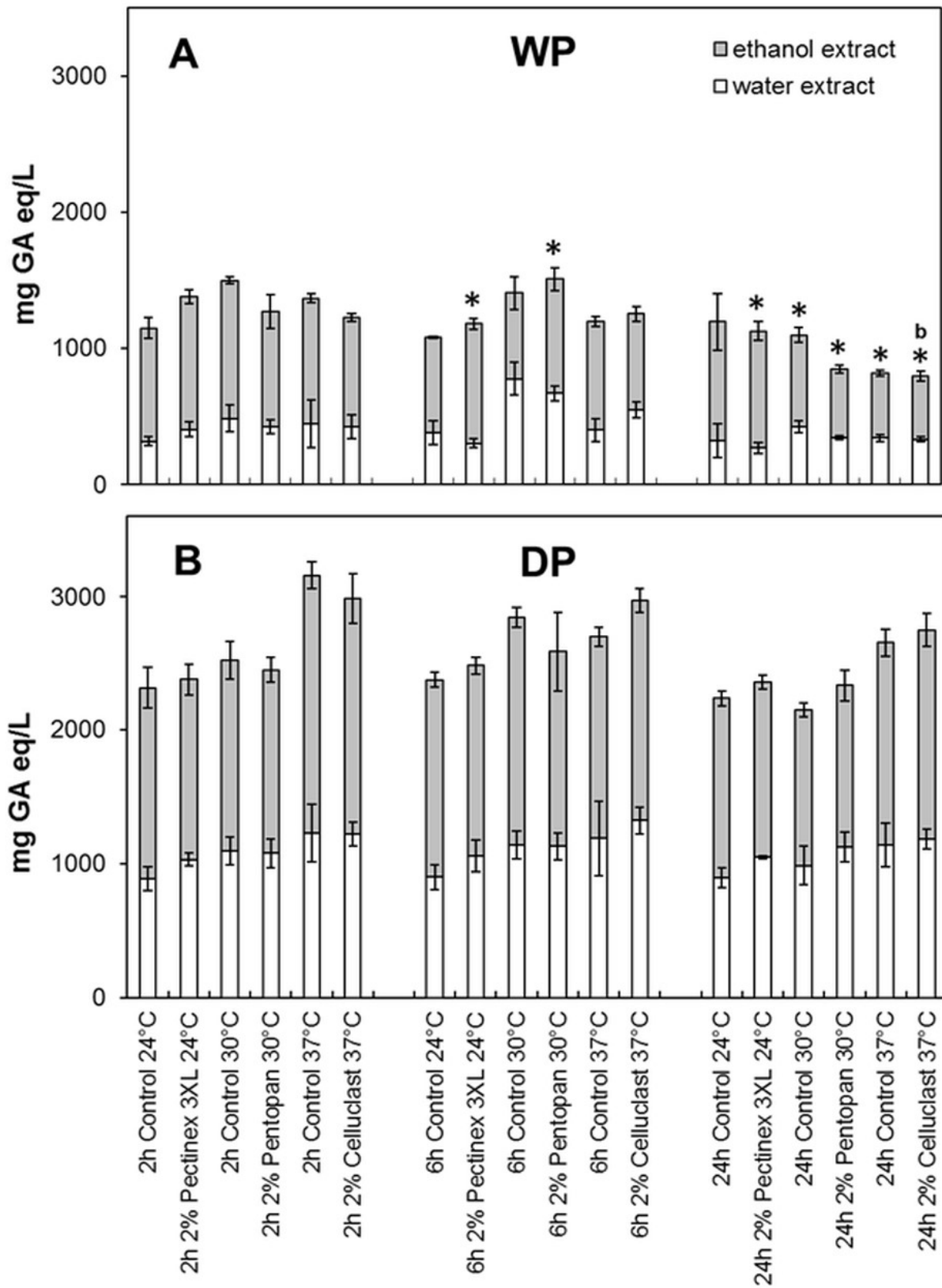
1 **Supplementary Figure S1.**



2

3

1 Supplementary Figure S2.



2

3