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Mercury and protein thiols: Stimulation of mitochondrial F1FO-ATPase and inhibition of respiration

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Mercury and protein thiols: stimulation of mitochondrial F_1F_0 -ATPase and inhibition of respiration

*Salvatore Nesci, Fabiana Trombetti, Maurizio Pirini, Vittoria Ventrella, Alessandra Pagliarani**

Department of Veterinary Medical Sciences, University of Bologna, Italy

*Corresponding author: Alessandra Pagliarani alessandra.pagliarani@unibo.it Tel +390512097017

Fax +390512097037, Department of Veterinary Medical Sciences (DIMEVET), University
of Bologna, via Tolara di Sopra, 50, 40064, Ozzano Emilia, Bologna, Italy

Abstract

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3 In spite of the known widespread toxicity of mercury, its impact on mitochondrial bioenergetics is a
4
5 still poorly explored topic. Even if many studies have dealt with mercury poisoning of
6
7 mitochondrial respiration, as far as we are aware Hg^{2+} effects on individual complexes are not so
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9 clear. In the present study changes in swine heart mitochondrial respiration and F_1F_0 -ATPase (F-
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11 ATPase) activity promoted by micromolar Hg^{2+} concentrations were investigated. Hg^{2+} was found
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13 to inhibit the respiration of NADH-energized mitochondria, whereas it was ineffective when the
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15 substrate was succinate. Interestingly, the same micromolar Hg^{2+} doses which inhibited the NADH-
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17 O_2 activity stimulated the F-ATPase, most likely by interacting with adjacent thiol residues.
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19 Accordingly, Hg^{2+} dose-dependently decreased protein thiols and all the elicited effects on
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21 mitochondrial complexes were reversed by the thiol reducing agent DTE. These findings clearly
22
23 indicate that Hg^{2+} interacts with Cys residues of these complexes and differently modulate their
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25 functionality by modifying the redox state of thiol groups. The results, which cast light on some
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27 implications of metal-thiol interactions up to now not fully explored, may contribute to clarify the
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29 molecular mechanisms of mercury toxicity to mitochondria.
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Keywords

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42 F_1F_0 -ATPase; Mitochondria; Oxidative phosphorylation; Mercury; Thiol groups.
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Abbreviations

$\Delta\mu_{H^+}$, Transmembrane electrochemical gradient of protons; OXPHOS, oxidative phosphorylation; DTE, 1,4-Dithioerythritol; DNP, 2,4-Dinitrophenol; PAO, Phenylarsine oxide; NEM, *N*-Ethylmaleimide; MBBr, Monobromobimane; DTNB, 5,5'-Dithiobis(2-nitrobenzoic acid); MTRs, monothiol reagents

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1. Introduction

Mercury is a toxic heavy metal and mercury contamination causes pathological symptoms primarily in brain but also in lungs, kidney and heart [1-4]. The different chemical forms of mercury have been related to their different toxicity. Inorganic mercury comprises the elemental forms metallic mercury and mercury vapor (Hg^0), and the ionic forms mercurous (Hg^+) and mercuric (Hg^{2+}) ions. Organic mercury, in which the metal is covalently bound to an organic moiety, mainly stems from biological sources. The organic forms have long been considered highly dangerous, being more easily absorbed and life-long accumulated especially in animals at the top of the food chain. Organic mercury can be also converted into inorganic salts, and especially into mercuric chloride (HgCl_2) [4]. This inorganic salt has raised concern as a significant source of acute and chronic poisoning by mercury [5]. From a molecular insight, the chemical interaction between Hg^{2+} and biomolecules, only partially known, may substantially contribute to the highly varied mercury toxicity [3].

Due to their biological features, mitochondria are extremely vulnerable to pollutants [6] including highly toxic heavy metals [7]. Exposure to mercury, mainly through contaminated seafood and dental amalgam [8], has been associated with mitochondrial dysfunctions in aging [6], cardiovascular [2,4,9] and neurodegenerative diseases [7,10] and even with the development of autoimmune diseases [11] and autism [12].

All mercury compounds have a great affinity for sulphhydrylic groups of proteins and other biomolecules [3,5]. The inorganic Hg^{2+} [8] enters mitochondria by exploiting the negative charge in the mitochondrial matrix built by the respiratory chain [6] and uncouples oxidative phosphorylation (OXPHOS) [7,13,14]. Mercury mainly localizes in the protein fraction of mitochondria [15]. Hg^{2+} covalently binds to thiol ($-\text{SH}$) groups of Cys residues of proteins and forms adducts which change the protein conformation [3,16,17]. Moreover, mercury may bind to intracellular reduced glutathione (GSH) and deplete GSH stores, thus contributing to an increase in reactive oxygen species (ROS) [18]. Furthermore Hg^{2+} formation from organomercury compounds in the

1 mitochondrial matrix is stimulated by superoxide production. Additionally, oxidative stress greatly
2 amplifies mercury capability to bind Cys thiols [19]. As a response to cellular stress, mitochondria
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4 can dictate the cell fate by increasing the mitochondrial membrane permeability, an event that
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6 triggers apoptosis [20], another process in which mercury has been involved [4,7,11]. So, a tight
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8 link exists between mercury, oxidative stress and cell death. Being among mercury targets and ROS
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10 producers, the mitochondrion plays a key role in this interaction network.
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14 The primary role of mitochondria is ATP production through OXPHOS. Basically OXPHOS
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16 involves four respiratory complexes in the inner mitochondrial membrane (IMM) which transfer the
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18 electrons withdrawn from the reduced substrates $\text{NADH} + \text{H}^+$ and FADH_2 , in turn generated by
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20 nutrient oxidation, to molecular oxygen [21]. The energy released by this electron cascade through
21
22 the so-called electron transport chain is converted into an electrochemical proton gradient ($\Delta\mu_{\text{H}^+}$)
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24 across the IMM by three of these complexes (complex I, III and IV, respectively), which pump
25
26 protons. The ATP synthase or F_1F_0 -ATPase is able to dissipate $\Delta\mu_{\text{H}^+}$ to synthesize ATP; under
27
28 pathological conditions when IMM is depolarized, the same enzyme complex can work in reverse,
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30 namely it hydrolyzes ATP. In this case the enzyme complex exploits ATP hydrolysis to pump
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32 protons and re-energize the membrane [22]. The two interlocked domains of the F_1F_0 -ATPase, F_1
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34 and F_0 , work together matched by a torque generation mechanism [23]: the hydrophilic F_1 builds
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36 ATP, while the membrane-embedded domain F_0 rotates as a H^+ turbine. The rotation direction of
37
38 these engines and their catalytic task (synthesis/hydrolysis) depend on the thermodynamic balance
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40 between $\Delta\mu_{\text{H}^+}$ and ΔG_p (the Gibbs free energy for ATP synthesis). $\Delta\mu_{\text{H}^+}$ drives ATP synthesis
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42 (counterclockwise rotation, viewed from the matrix side), and *vice versa* an overwhelming ΔG_p
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44 leads to ATP hydrolysis (clockwise rotation) [24].
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53 Post-translational modifications altering the redox state of thiols concur to physiologically rule the
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55 mitochondrial functions associated with signal transduction pathways triggered by ROS production
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57 [25]. Since antioxidant defenses, mainly GSH pool, the related regenerating systems and ROS
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59 sources, are differently distributed, mitochondria contain different redox compartments.
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1 Interestingly, OXPHOS complexes lie on the boundary line of two redox environments: the
2 reducing matrix and the intermembrane space with cristae lumen which constitutes a relatively
3 oxidizing environment [26]. Within this boundary line, protein Cys residues may act as a “sulfur
4 switch” [25], thus behaving as chemical interface between the mitochondrial environment and the
5 enzyme function. The protein thiol redox state, already involved in the conformational and/or
6 activity changes of Complex I [27] and F₁F₀-ATPase [28] under pathological conditions, may also
7 play a role in the mercury-driven mitochondrial dysfunction. The latter leads to a decrease in ATP
8 production and to an increase in oxidative stress [4,7]. In spite of the wealth of studies on mercury
9 effects on the respiratory chain, the modulation of the enzyme complexes, with the exclusion of any
10 interference of membrane potential driven by mercury cations, has not been investigated yet. The
11 molecular mechanisms of metal interaction with protein thiols, here investigated in uncoupled
12 mitochondria, reveal that the same basic chemical mechanism differently modulates the F₁F₀-
13 ATPase (henceforth defined as F-ATPase) and other OXPHOS complexes. The findings may
14 contribute to understand the complex and varied pattern of mercury toxicity by casting light on
15 some implications of metal-thiol interactions which up to now have not been fully explored.
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39 2. Materials and methods

40 2.1. Chemicals

41 Na₂ATP, oligomycin mixture (A:B:C 64:15:17%), 1,4-Dithioerythritol (DTE), 2,4-Dinitrophenol
42 (DNP), Phenylarsine oxide (PAO), N-Ethylmaleimide (NEM), NADH, sodium succinate, rotenone,
43 mercury(II) chloride and 5,5'-Dithiobis(2-nitrobenzoic acid) (DTNB) were obtained from Sigma-
44 Aldrich (Milan, Italy). Monobromobimane (MBr) was purchased by Vinci-Biochem (Vinci, Italy).
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46 All other chemicals were reagent grade. Quartz double distilled water was used for all reagent
47 solutions except when differently stated.
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61 2.2. Preparation of the mitochondrial fraction

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1 Swine hearts (*Sus scrofa domesticus*) were collected at a local abattoir and transported to the lab
2 within 2 h in ice buckets at 0-4°C. After removal of fat and blood clots as much as possible,
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4 approximately 30-40 g of heart tissue from the left ventricle were rinsed in ice-cold washing Tris-
5
6 HCl buffer consisting of 0.25 M sucrose, 10 mM Tris(hydroxymethyl)-aminomethane (Tris), pH
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8 7.4 and finely chopped into fine pieces with scissors. Each preparation was made from one heart.
9
10 Mitochondria were isolated by stepwise centrifugation as described previously [29]. The protein
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12 concentration was determined according to the spectrophotometric method of Bradford [30] by Bio-
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14 Rad Protein Assay kit II. The mitochondrial preparations were then stored in liquid nitrogen until
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16 analyses.
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24 2.3. Assay of the mitochondrial F₁F₀-ATPase (F-ATPase) activity

25 Immediately after thawing, the mitochondrial fractions were used for the F-ATPase activity assays.
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27 The capability of ATP hydrolysis was assayed in a reaction medium (1 mL) containing 75 mM
28
29 ethanolamine-HCl buffer pH 9.3, 0.15 mg mitochondrial protein and 2.0 mM MgCl₂ for F₁F₀-
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31 ATPase assay. After 5 min preincubation at 37 °C, the reaction, carried out at the same temperature,
32
33 was started by the addition of 6.0 mM Na₂ATP as substrate and stopped after 5 min by the addition
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35 of 1 mL of ice-cold 15% (w/w) aqueous solution trichloroacetic acid. Once the reaction was
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37 stopped, vials were centrifuged for 15 min at 5000 rpm (ALC 4225 Centrifuge). In the supernatant,
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39 the concentration of inorganic phosphate (P_i) hydrolyzed by known amounts of mitochondrial
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41 protein, which is an indirect measure of ATPase activity, was colorimetrically evaluated [31].
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48 The ATPase activity was routinely measured by subtracting, from the P_i hydrolyzed by known
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50 amounts of mitochondrial protein (which indirectly indicates the total ATPase activity); the P_i
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52 hydrolyzed in the presence of 3 µg/mL oligomycin. To this aim, in vials run in parallel, 1 µL from a
53
54 mother solution of 3 mg/mL oligomycin in dimethylsulfoxide (DMSO) was directly added to the
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56 reaction mixture before starting the reaction. The employed dose of oligomycin, a specific inhibitor
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58 of F-ATPases which selectively blocks the F₀ subunit [32-34], ensured maximal enzyme activity
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1 inhibition and was currently used in ATPase assays [29,32-34]. The total ATPase activity was
2 calculated by detecting the P_i in control tubes run in parallel and containing 1 μ L DMSO per mL
3 reaction system. In each experimental set, control tubes were alternated to the condition to be
4 tested.. Small volumes of the effectors in aqueous solutions were directly added to the reaction
5 system so as to obtain the required concentrations. In all experiments the enzyme activity was
6 calculated as μ mol P_i ·mg protein⁻¹·min⁻¹.
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17 2.4. Preincubation and treatment of mitochondria

18 When combined effects of different effectors, *i.e.* HgCl₂ and mono- or di-thiol reagents were tested,
19 mitochondrial suspensions were preincubated for 30 min with appropriate concentrations of these
20 effectors in Eppendorf vials on ice. This procedure was designed to ensure the interaction of the
21 effectors in Eppendorf vials on ice. This procedure was designed to ensure the interaction of the
22 effectors to be tested with the mitochondrial proteins [34]. After this preincubation time, the
23 ATPase reaction was carried out as described in Section 2.3.
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34 2.5. Assay of the mitochondrial respiration

35 Immediately after thawing, the mitochondrial fractions were used to evaluate the mitochondrial
36 respiration. The oxygen consumption rates were polarographically evaluated by Clark-type
37 electrode using a thermostated Oxytherm System (Hansatech Instruments) equipped with a 1 mL
38 polarographic chamber. The reaction mixture (120 mM KCl, 10 mM Tris-HCl buffer pH 7.2),
39 maintained under Peltier thermostataion at 37 °C and continuous stirring, contained 0.25 mg
40 mitochondrial protein.
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51 To evaluate the NADH-O₂ activity, the mitochondrial oxidation was run under saturating substrate
52 conditions (75 μ M NADH) after 2 min of stabilization of oxygen signal. Preliminary tests assessed
53 that under these conditions the O₂ consumption was suppressed by the presence of 2.5 μ M rotenone,
54 known inhibitor of Complex I [21]. The succinate-O₂ activity by Complex II was determined by the
55 oxidation of succinate in the presence of 2.5 μ M rotenone. The reaction was started by the addition
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1 of 10 mM succinate after 2 min of stabilization of oxygen signal. Also in this case preliminary tests
2 assessed that, under the conditions applied, succinate oxidation was suppressed by 1µg/mL of
3 antimycin A, a selective inhibitor of complex III [21].
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5
6 To evaluate Hg²⁺-effects, HgCl₂ solutions of adequate concentrations, prepared immediately before
7 the experiments by dissolving the contaminant as chloride salt in water, were added 5 min before to
8 start the reaction at 37 °C at the same time of mitochondria to the polarographic chamber. The rate
9 of mitochondrial respiratory activities were evaluated automatically by O₂view software as nmoles
10 O₂·mg protein⁻¹·min⁻¹. In routine experiments, reagents were added by a syringe to the
11 polarographic cell containing the reaction mixture and the mitochondrial protein suspension in the
12 following order: HgCl₂ when required, rotenone for succinate-O₂ activity, substrate (NADH or
13 succinate) and DTE. Polarographic assays were run at least in triplicate on mitochondrial fractions
14 from distinct animal pools.
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31 2.6. Quantitative evaluation of free thiols

32 Free thiols in mitochondrial suspensions in the absence and in the presence of 1.0, 2.0, 3.0, 5.0, 10.0
33 µM HgCl₂ were colorimetrically quantified by Ellman's reagent [35]. The selected HgCl₂
34 concentrations were added to the mitochondrial suspensions immediately before the colorimetric
35 analysis. The widely used Ellman's method is based on the capability of 5,5'-Dithiobis(2-
36 nitrobenzoic acid) (DTNB) to react with free thiol groups by disulphide bonds with thionitrobenzoic
37 acid (TNB). As the ratio of protein thiols to TNB formed is 1:1, TNB formation is currently taken
38 as a measure of the number of free thiol groups in mitochondrial suspensions. After the addition of
39 15% w/v trichloroacetic acid solution (250 µL/0.15 mg protein) to precipitate proteins, the
40 mitochondrial suspensions were centrifuged at 12,000 g for 5 min at 4 °C. After removal of the
41 supernatant, the mitochondrial pellet was carefully resuspended with potter Eppendorf pestle. Then,
42 400 µL of reagent solution containing 0.5 M phosphate buffer (KH₂PO₄/K₂HPO₄, pH 7.4), 0.2 mM
43 DTNB, were added to the suspensions and incubated for 20 min at 4 °C. The absorbance at 412 nm
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1 (maximum TNB absorption wavelength) of the supernatant from a second centrifugation at 12,000
2 g for 5 min at 4 °C was read on a Perkin–Elmer lambda 45 spectrophotometer. Mitochondrial thiol
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4 groups were quantitatively evaluated by interpolating the absorbance values in a calibration curve
5
6 built by employing known cysteine concentrations as -SH standard. In each experiment set, data
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8 were expressed as % free thiol/mg protein in mitochondria detected in HgCl₂-free medium taken as
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10 control.
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13 14 15 16 17 2.7. Kinetic analyses

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19 The mechanism of the enzyme activation by HgCl₂ on the F-ATPase was explored by the aid of the
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21 graphical method of Lineweaver-Burk (double reciprocal plots). In all kinetic analyses the enzyme
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23 specific activity was taken as the expression of the reaction rate (v). Briefly, to build double
24
25 reciprocal plots the reciprocal of the reaction rate $1/v$ (y axis) was plotted as a function of the
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27 reciprocal of concentration of ATP substrate or Mg²⁺ cofactor (x axis) [36]. By plotting the enzyme
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29 activity data, distinct straight lines, each of which corresponded to a fixed concentration of HgCl₂
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31 were obtained by linear regression. The R² value was never lower than 0.98, thus confirming the
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33 linearity of these plots.
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39 At least three independent experiments were carried out to build each plot.
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43 44 2.8. Statistical Analysis

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46 The data represent the mean \pm SD (shown as vertical bars in the figures) of the number of
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48 experiments reported in the figure captions and table legends. In each experimental set, the analyses
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50 were carried out on different mitochondrial preparations. The differences between data were
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52 evaluated by one way ANOVA followed by Students–Newman–Keuls' test when F values indicated
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54 significance ($P \leq 0.05$).
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60 61 3. Results 62 63 64 65

3.1. Mercury effects on selected mitochondrial enzyme activities

The present work highlights a differential effect of HgCl₂ on some enzyme activities involved in OXPHOS. Accordingly, as shown in Figure 1, micromolar concentrations of HgCl₂ up to 10 μM, evaluated by detecting the oxygen consumption in the presence of two substrates which elicit the activity of Complex I (NADH) and Complex II (succinate), respectively, yielded substrate-dependent effects. The NADH-O₂ activity was progressively inhibited by increasing HgCl₂ concentrations up to attain 10% of the activity in a mercury-free medium at 10 μM HgCl₂. The calculated IC₅₀ value was 1.7±0.5 μM. Conversely, the succinate-O₂ activity was unaffected by the HgCl₂ concentrations tested up to 10 μM HgCl₂.

A biphasic effect was shown on the F-ATPase (Fig. 2). The enzyme activity was stimulated by low micromolar HgCl₂ concentrations: at 2 μM HgCl₂ the enzyme activity was 42% higher than the control (no HgCl₂). Only at HgCl₂ concentrations ≥ 10 μM was the F-ATPase activity decreased: 50% enzyme inhibition occurred at 22.6±4.0 μM HgCl and 100% inhibition was detected in the presence of 100 μM HgCl₂ (Fig. S1).

3.2. Hg²⁺ and thiols

On considering the ability of mercury to form complexes with Cys thiols, its putative binding to mitochondrial proteins was investigated by evaluating the thiol content in swine heart mitochondria incubated in the absence or in the presence of increasing concentrations of HgCl₂. The amount of free thiol groups in mitochondria significantly decreased at concentrations of HgCl₂ ≥ 3 μM up to plateau to 20% in the range 5-10 μM HgCl₂. Therefore, at least under the conditions applied, only 20% of the free mitochondrial thiols were targeted by mercury (Fig. 3).

In order to check whether HgCl₂ effects on mitochondrial enzyme activities could be reversed, the thiol reagent DTE, known to reduce thiols, was tested (Fig. 4). The NADH-O₂ activity *per se* was unaffected by 50 μM DTE. Conversely, when the enzyme activity was inhibited by 2 μM HgCl₂,

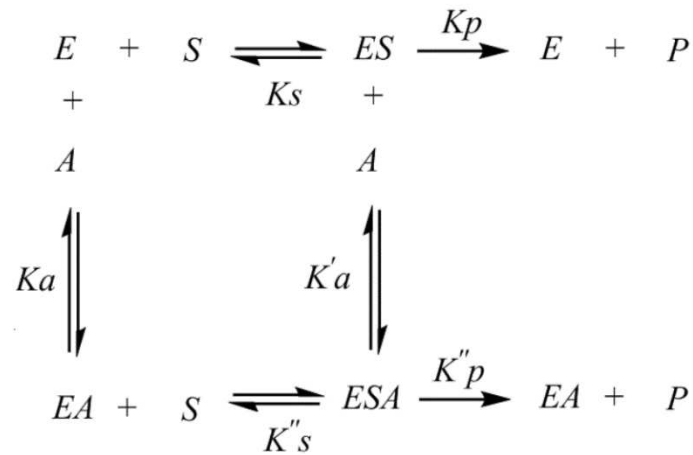
1 DTE removed the enzyme inhibition and restored the mitochondrial respiration to control values
2 (Fig. 4A). Likewise, the F-ATPase activity was *per se* unaffected by 50 μ M DTE, a dose which
3
4 suppressed the enzyme activation by 2 μ M HgCl₂ (Fig. 4B).
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6

7 Interestingly, the mitochondrial F-ATPase was also activated by the presence of micromolar
8
9 concentrations of phenylarsine oxide (PAO), a vicinal dithiol-binding agent. Conversely binary
10
11 mixtures of increasing micromolar concentrations of PAO plus 50 μ M DTE were ineffective (Fig.
12
13 S2). To check if the F₁F₀-ATPase activation was due to an uncoupler-like effect of mercury, the
14
15 known uncoupler DNP was tested. However, no significant change in the enzyme activity was
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17 detected (Fig. S3).
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21 The mechanisms involved in the F-ATPase activation by HgCl₂ were deepened by assaying the
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23 effects of PAO and of the thiol reagents (MTRs) *N*-Ethylmaleimide (NEM) and monobromobimane
24
25 (MBBr), already used in a previous work [34] (Table 1). In these sets of experiments two different
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27 protocols were applied, namely mitochondria were either preincubated with HgCl₂ or with
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29 individual MTRs in the absence of other effectors. When mitochondria were preincubated with
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31 HgCl₂, individual MTRs were added to the reaction system immediately after the addition of
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33 HgCl₂-preincubated mitochondrial suspensions. In the alternative mode, HgCl₂ was added to the
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35 reaction system immediately after the addition of MTR-preincubated mitochondrial suspensions.
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38 The two procedures resulted in the same F-ATPase activation by HgCl₂ which was suppressed by
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40 DTE addition. In the absence of mercury, all the MTRs, both in the presence and in the absence of
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42 DTE, did not affect the F-ATPase activity. Conversely, in HgCl₂-preincubated mitochondria the
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44 subsequent addition of PAO to the reaction system did not result in any enzyme activity
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46 stimulation. In PAO-preincubated mitochondria, HgCl₂ added to the reaction system did not
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48 stimulate the F-ATPase activity. In all cases, DTE restored the enzyme activity detected in the
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50 control.
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61 3.3. Kinetics 62 63 64 65

Finally, the F-ATPase activation kinetics by Hg^{2+} was studied by evaluating the F-ATPase activities raised by different concentrations of ATP substrate at fixed cofactor Mg^{2+} concentration, or by different concentrations of Mg^{2+} at fixed ATP concentration, in the presence and in the absence of 2 μM HgCl_2 . This approach, adequately adapted from kinetic analysis methods [36], aimed at evaluating the kinetic activation by HgCl_2 of the F-ATPase with respect to ATP substrate and Mg^{2+} cofactor, respectively. The unparallel lines of the obtained–double reciprocal plots (Figure 5), showed that in the presence of Hg^{2+} the line was below the control line in both cases. Therefore Hg^{2+} increases both the V_{max} and K_m values for the ATP substrate and the Mg^{2+} cofactor (Table 2). On these bases mixed-type or non-competitive activation are suggested. Mixed-type activation indicates that the activator (A) binds both to the free enzyme, yielding the binary complex EA and to ES , forming the ternary complex ESA . Non-competitive activation means that these two possibilities are equal. The whole pattern of mixed-type enzyme activation by the modulator A is illustrated below:



To explore this point, once obtained the modified K_m and V_{max} values from the double reciprocal plot in the presence of Hg^{2+} , the factor α , which represents the proportionality constant which modifies the slope in the presence of a modulator, was calculated from the relationship $\alpha = \text{slope} \cdot V_{\text{max}} / K_m$. Since in mixed-type activation the intercept on y axis is modified by the constant α' , namely y intercept = α' / V_{max} , α' was obtained by multiplying the y intercept per V_{max} . In other words, α refers to the formation of EA complex, while α' refers to the formation of ESA complex.

1
2 Therefore, the activation constants K_a and K'_a , which represent the equilibrium constants of the
3 dissociation of the *EA* and *ESA* complexes respectively, were obtained from the relationships:
4 $\alpha=1+[A]/K_a$ and $\alpha'=1+[A]/K'_a$. Since K_a is higher than K'_a (Table 3), *ESA* is more stable than *EA*.
5
6 This means that Hg^{2+} mainly forms the ternary enzyme-substrate-activator complex.
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10 11 4. Discussion

12
13 Mercury toxicity renewed concern, especially for the cardiovascular consequences of the chronic
14 human exposure [2-4]. The cationic form Hg^{2+} which alters the mitochondrial integrity and
15 metabolism [4,37,39] is long known to perturb OXPHOS in mammalian [4,7,15] and non
16 mammalian [39] species. Given the documented wide range of action of the toxicant [3,5] and the
17 interconnection between membrane and enzyme perturbations, it is quite difficult to elucidate the
18 deleterious mechanisms triggered by the toxicant. As far as we are aware, up to now no study has
19 considered mercury effects on the mitochondrial respiratory complexes and F_1F_0 -ATPase activity
20 independently of IMM potential and structural perturbations. So, the present work on freeze-thawed
21 mitochondria had the main goal of clarifying mercury effects on selected enzyme activities of the
22 IMM, under conditions which rule out any possible concomitant effect of $\Delta\mu_{H^+}$ changes. With the
23 aim of casting light on the molecular mechanisms involved, we primarily focused on the metal
24 interaction with protein thiols. Recent advances hint that this is probably the main molecular
25 mechanism of mercury toxicity [3,4,17], now recognized as connected with oxidative damage [7,
26 9]. Accordingly, thiol-based redox changes in OXPHOS complexes are accompanied by ROS
27 production [7] and/or come from a reduction potential of a critical redox couple [40]. Moreover,
28 independently of major redox changes within the membrane, under appropriate conditions, post-
29 translational changes lead to a variety of thiol modifications such as sulfenic acid formation,
30 glutathionylation, sulfenyl amides, *S*-nitrosothiols and to the onset of intra- and inter-protein
31 disulfide bridges [41]. Thiol modifications rule Complex I functions in heart failure: inactive
32 Complex I is likely to be *S*-nitrosylated, while the active Complex I is unaffected. This reversible
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1 manipulation would inhibit superoxide production by reverse electron transfer [42]. Redox changes
2 in some thiol groups in the F_O portion of the mitochondrial F-ATPase result in uncoupling [43], loss
3 of sensitivity to oligomycin and other macrolide antibiotics [31]. The reversible thiol modification
4 by mercurials prevents the F-ATPase from being irreversibly blocked by DCCD [44], thus hinting
5 that some susceptible thiols occur in critical proton binding points. Past *in vitro* research pointed out
6 a widespread HgCl₂ toxicity on OXPHOS. Various effects were recorded: Hg²⁺ mainly acted as
7 uncoupler by dissipating $\Delta\mu_{H^+}$, enhanced basal respiration [7,45], while state 3 respiration was
8 unaffected or depressed [7], inhibited the succinate dehydrogenase and ATP synthase activities
9 [39], suppressed the membrane cation selectivity [46] and prevented the membrane re-energization
10 [14]. Concomitantly, Hg²⁺ promoted oxidative stress [7], leading to lipid peroxidation [47], in turn
11 compromising the IMM structural integrity [48]. In heart mitochondria the ATP synthase reversible
12 inhibition by nanomolar Hg²⁺ concentrations was already reported, without elucidating the
13 mechanism(s) involved [49]. Conversely, methylmercury stimulated the F-ATPase activity in the
14 presence of K⁺, which by entering mitochondria would collapse the membrane potential [15].

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34 In the present study the NADH-O₂ inhibition by micromolar HgCl₂ concentrations, mirrors the
35 activities of three respiratory complexes (I, III and IV) which act as proton pumps [21]. Since the
36 mercury-driven inhibition is confined to the NADH-O₂ activity, being the succinate-O₂ activity
37 unaffected (Fig. 1), Complex I is confirmed as main target of mercury among OXPHOS complexes
38 and its inhibition may *per se* lead to mitochondrial dysfunction [4]. In contrast, the Complex II
39 features may explain the succinate oxidase activity refractoriness to the tested Hg²⁺ concentrations.
40 Accordingly, in spite of its content of thiols and dithiols [50], Complex II contains less reactive or
41 less accessible thiol groups than Complex I [51].

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1 presence of $\mu\text{molar HgCl}_2$, we can infer that only the succinate- O_2 activity contributes to $\Delta\mu_{\text{H}^+}$
2 generation since the electron transfer from Complex I to Complex 3 is prevented by mercury, which
3 blocks Complex I by thiol complexation. By this chemical mechanism, mercury would poison the
4 respiratory chain in a different way from other mechanisms already described in mitochondria such
5 as the displacement of Fe^{2+} and Cu^+ ions from redox centers and membrane lipid peroxidation [2, 4,
6 7]. To sum up, it seems likely that the direct binding of Hg^{2+} to critical thiols of Complex I may
7 lessen Complex I functionality and significantly concur to the mercury-driven mitochondrial
8 dysfunction.

9 An opposite effect, namely the enzyme activity stimulation, was shown on the mitochondrial F-
10 ATPase. As bifunctional enzyme [53,54], the F-ATPase not only synthesizes ATP but also can
11 exploit ATP hydrolysis to pump protons when the IMM is insufficiently polarized, to overcome the
12 reversal potential threshold [54] and prevent the depolarization of mitochondria. In this context
13 HgCl_2 , by stimulating the ATPase activity, may improve the coupling between glycolytic and $\Delta\mu_{\text{H}^+}$
14 oscillations in non-respiring mitochondria [55]. Interestingly, mercury complexation to thiols can
15 produce either inhibition (NADH-O_2) or activation (F-ATPase), depending on the localization and
16 the role of the susceptible thiols in the mitochondrial complexes. Consistently, even if mercury
17 binding to enzyme thiols is believed a common motif for enzyme inactivation [3], it activates some
18 phospholipases [9,56].

19 Finally, the failed effect of the protonophore DNP on the F-ATPase activation rules out any
20 uncoupling effect of Hg^{2+} under our experimental conditions, even if Hg^{2+} is a recognized ionophore
21 [46].

22 The reversibility of these modifications of thiols shoulders the thiol role as sulfur switch to
23 modulate enzyme function [3]. Since the F-ATPase activation by mercury is not sensitive to MTRs
24 but is prevented by PAO, which specifically binds to dithiols, two adjacent thiol groups, known to
25 play a significant role in mitochondrial proteins [57] are required for mercury binding to the F-
26 ATPase. Mercury complexation to the F-ATPase thiols is facilitated when ATP and Mg^{2+} are bound
27

1 to the enzyme, since mercury preferentially binds to the enzyme-substrate complex, namely it
2 shows mixed-type activation kinetics (Figure 5). It seems likely that the MgATP-complex
3 formation may induce conformational changes which somehow make dithiols more accessible to
4 mercury. Consistently, with respect to controls, the reaction rate increases in spite of a decreased
5 affinity for ATP and Mg²⁺ (Table 2).
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10 This work shoulders the concept that enzyme alterations play a great role in mercury toxicity [3].
11 From a literature overview, this is probably the first report which shows that Hg²⁺, by targeting
12 crucial thiols, acts differently on the mitochondrial complexes, producing either inhibition or
13 stimulation of IMM enzyme activities or even no effect, as on Complex II. Even if we must be
14 cautious to transpose *in vitro* to *in vivo* responses, it is intriguing to speculate that the shown duality
15 of mercury yielding Complex I inhibition and F-ATPase stimulation could make mitochondria
16 resilient to mercury contamination. Accordingly, the stimulation of the reverse functioning mode of
17 the ATP synthase (F-ATPase) would preserve, at least to a certain extent, the IMM polarization and
18 ionic homeostasis under conditions of respiratory chain inhibition. If this mechanism also works *in*
19 *vivo*, mercury-contaminated mitochondria would maintain at least partially their functionality.
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35 Even if vulnerable to pollutants, mitochondria are at the same time robust [6], being endowed with
36 multiple mechanisms to ensure their function. Consistently, the cysteine proportion in proteins
37 increases with organism complexity [58] and this thiol abundance on the protein surface may be
38 there for a reason [59]. So, among the multiple targets of mercury in mitochondria, the most
39 reactive thiols of Complex I and F-ATPase could work together to minimize the mitochondrial
40 damage by mercury.
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6. References

- 1
2
3 [1]World Health Organisation Geneva, Inorganic mercury. Sources of human and
4 environmental exposure, Environmental Health Criteria 118 (1991) 29-33.
5
6
7 [2]M.C. Houston , Role of mercury toxicity in hypertension, cardiovascular disease and stroke.
8
9 J. Clin. Hyper. 8 (2011) 621-627.
10
11
12 [3]R. Ynalvez, J. Gutierrez, H. Gonzalez-Cantu, Minireview: toxicity of mercury as a
13 consequence of enzyme alteration, Biometals (2016), in press doi; 10.1007/s10534-015-
14 9967-8.
15
16
17
18
19 [4] M.C. Houston, The role of mercury in cardiovascular disease, J. Cardiovasc. Dis. Diagn. 2
20 (2014) 170.
21
22
23
24 [5]R.A. Bernhoft, Mercury toxicity and treatment: a review of the literature. J. Environ. Public
25 Health (2012) 2012, ID 460508 <http://dx.doi.org/10.1155/2012/460508>.
26
27
28
29 [6]J.N. Meyer, M.C.K. Leung, J.P. Rooney, A. Sandoel, M.O. Hengartner, G.E. Kisby, A.S.
30 Bess, Mitochondria as a target of environmental toxicants, Toxicol. Sci. 134 (2013) 1-17.
31
32
33 [7]E.A. Belyaeva, T.V. Sokolova, L.V. Emelyanova. I.O. Zakharova, Mitochondrial electron
34 transport chain in Heavy metal-induced neurotoxicity: effects of cadmium, mercury and
35 copper, The Scientific World J. 2012 (2012) Article ID 136063 doi: 10.1100/2012/136063.
36
37
38
39 [8]T. Syversen, P. Kaur, The toxicology of mercury and its compounds, J. Trace El. Med. Biol.
40 26 (2012) 215-26.
41
42
43
44
45 [9]T.J. Hagele, J.N. Mazerik, A. Gregory, B. Kaufman, U. Magalang, M.L. Kuppusamy, C.B.
46 Marsch, P. Kuppusamy, N.L. Parinandi, Mercury activates vascular endothelial cell
47 phospholipase D through thiols and oxidative stress, Int. J. Toxicol. 26 (2007) 57-69.
48
49
50
51 [10]A. Carocci, N., Rovito, M.S. Sinicropi, G., Genchi, Mercury toxicity and neurodegenerative
52 effects, Rev. Environ.Contam. Toxicol. 229 (2014) 1-18.
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- [11]M.P. Waalkes, D.A. Fox, J.C. States, S.R. Patierno, M.J. McCabe Jr., Metals and disorders of cell accumulation: modulation of apoptosis and cell proliferation, *Toxicol. Sci.* 56 (2000) 255-61.
- [12]S. Rose, R. Wynne, R.E. Frye, S. Melnyk, S.J. James, Increased susceptibility to ethylmercury-induced mitochondrial dysfunction in a subset of autism lymphoblastoid cell lines, *J. Toxicol.* 2015 (2015) 573701
- [13]A.C. Santos, S.A. Uyemura, N.A. Santos, F.E. Mingatto, C. Curti, Hg(II)-induced renal cytotoxicity: *in vitro* and *in vivo* implications for the bioenergetic and oxidative status of mitochondria, *Mol. Cell. Biochem.* 177 (1997) 53-9.
- [14]H. Reyes-Vivas, M. Lopez-Moreno, E. Chavez, Protective effect of dithiocarbamate on mercury-induced toxicity in kidney mitochondria, *Comp. Biochem. Physiol.* 113C (1996) 349-352.
- [15]N. Sone, M.K. Larstuvold, Y. Kagawa, Effect of methyl mercury on phosphorylation, transport and oxidation in mammalian mitochondria, *J. Biochem.* 82 (1977) 859-68.
- [16]P.R. van Iwaarden, A.J. Driessen, W.N. Konings, What we can learn from the effects of thiol reagents on transport proteins, *Biochim. Biophys. Acta* 1113 (1992) 161-70.
- [17]M. Farina, D.S. Avila, J.B. Teixeira da Rocha, M. Aschner, Metals, oxidative stress and neurodegeneration: a focus on iron, manganese and mercury. *Neurochem. Int.* 62 (2013) 575-591.
- [18]J.M. Hansen, H. Zhang, D.P. Jones, Differential oxidation of thioredoxin-1, thioredoxin-2, and glutathione by metal ions, *Free Radic. Biol. Med.* 40 (2006) 138-45.
- [19]R.J. Mailloux, E. Yumvihoze, H.M. Chan, Superoxide produced in the matrix of mitochondria enhances methylmercury toxicity in human neuroblastoma cells, *Toxicol. Appl. Pharmacol.* 289 (2015) 371-80.
- [20]D.R. Green, L. Galluzzi, G. Kroemer, Cell biology. Metabolic control of cell death, *Science* 345 (2014) 1250256.

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- [21] D.G. Nicholls, S.J. Ferguson, *Bioenergetics* 4, fourth ed., Academic Press, Amsterdam, 2013.
- [22] P. Dimroth, C. von Ballmoos, T. Meier, Catalytic and mechanical cycles in F-ATP synthases. Fourth in the Cycles Review Series, *EMBO Rep.* 7 (2006) 276-82.
- [23] W. Junge, H. Sielaff, S. Engelbrecht, Torque generation and elastic power transmission in the rotary F(O)F(1)-ATPase, *Nature* 459 (2009) 364-70.
- [24] S. Nesci, F. Trombetti, V. Ventrella, A. Pagliarani, Opposite rotation directions in the synthesis and hydrolysis of ATP by the ATP synthase: hints from a subunit asymmetry, *J. Membr. Biol.* 248 (2015) 163-9.
- [25] Y.M. Go, D.P. Jones, The redox proteome, *J. Biol. Chem.* 288 (2013) 26512-20.
- [26] S. Dröse, U. Brandt, I. Wittig, Mitochondrial respiratory chain complexes as sources and targets of thiol-based redox-regulation, *Biochim. Biophys. Acta.* 1844 (2014) 1344-54.
- [27] M. Babot, A. Birch, P. Labarbuta, A. Galkin, Characterisation of the active/de-active transition of mitochondrial complex I, *Biochim. Biophys. Acta.* 1837 (2014) 1083-92.
- [28] A. Pagliarani, S. Nesci, F. Trombetti, V. Ventrella, Thiol-related regulation of the mitochondrial F₁F₀-ATPase activity, in: S. Chakraborti, N.S. Dhalla, (Eds.), *Regulation of Ca²⁺-ATPases, V-ATPases and F-ATPases Series: Advances in Biochemistry in Health and Disease Vol. 14*, Springer International Publishing, Switzerland, (2016), pp 441-458.
- [29] S. Nesci, V. Ventrella, F. Trombetti, M. Pirini, A. Pagliarani, Preferential nitrite inhibition of the mitochondrial F₁F₀-ATPase activities when activated by Ca²⁺ in replacement of the natural cofactor Mg²⁺, *Biochim. Biophys. Acta* 1860 (2016) 345-53.
- [30] M.M. Bradford, A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding, *Anal. Biochem.* 72 (1976) 248-54.
- [31] C.G. Fiske, Y. Subbarow, The colorimetric determination of phosphorus, *J. Biol. Chem.* 66 (1925) 375-400.

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- [32] S. Nesci, V. Ventrella, F. Trombetti, M. Pirini, A. Pagliarani, Thiol oxidation is crucial in the desensitization of the mitochondrial F_1F_0 -ATPase to oligomycin and other macrolide antibiotics, *Biochim. Biophys. Acta* 1840 (2014) 1882-91.
- [33] S. Nesci, V. Ventrella, F. Trombetti, M. Pirini, A. Pagliarani, The mitochondrial F_1F_0 -ATPase desensitization to oligomycin by tributyltin is due to thiol oxidation, *Biochimie* 97 (2014) 128-37.
- [34] S. Nesci, V. Ventrella, F. Trombetti, M. Pirini, A. Pagliarani, Tri-*n*-butyltin binding to a low-affinity site decreases the F_1F_0 -ATPase sensitivity to oligomycin in mussel mitochondria, *Appl. Organomet. Chem.* 26 (2012) 593–599.
- [35] G.L. Ellman, Tissue sulfhydryl groups, *Arch. Biochem. Biophys.* 82 (1959) 70-7.
- [36] I.H. Segel, *Enzyme kinetics*, John Wiley & Sons Inc, New York, 1975.
- [37] T. Toimela, H. Tähti, Mitochondrial viability and apoptosis induced by aluminum, mercuric mercury and methylmercury in cell lines of neural origin, *Arch. Toxicol.* 78 (2004) 565-74.
- [38] P.B. Pal, S. Pal, J. Das, P.C. Sil, Modulation of mercury-induced mitochondria-dependent apoptosis by glycine in hepatocytes, *Amino Acids* 42 (2012) 1669-83.
- [39] C.L. Mieiro, M. Pardal, A. Duarte, E. Pereira, C.M. Palmeira, Impairment of mitochondrial Energy metabolism of two marine fish by in vitro mercuric chloride exposure, *Mar. Poll. Bull.* 97 (2015) 488-93.
- [40] Y.M. Janssen-Heininger, B.T. Mossman, N.H. Heintz, H.J. Forman, B. Kalyanaraman, T. Finkel, J.S. Stamler, S.G. Rhee, A. van der Vliet, Redox-based regulation of signal transduction: principles, pitfalls, and promises, *Free Radic. Biol. Med.* 45 (2008) 1-17.
- [41] M.P. Murphy, Mitochondrial thiols in antioxidant protection and redox signaling: distinct roles for glutathionylation and other thiol modifications, *Antioxid. Redox. Signal.* 16 (2012) 476-95.
- [42] E.T. Chouchani, C. Methner, S.M. Nadtochiy, A. Logan, V.R. Pell, S. Ding, A.M. James, H.M. Cochemé, J. Reinhold, K.S. Lilley, L. Partridge, I.M. Fearnley, A.J. Robinson, R.C.

- 1 Hartley, R.A. Smith, T. Krieg, P.S. Brookes, M.P. Murphy, Cardioprotection by S-
2 nitrosation of a cysteine switch on mitochondrial complex I, *Nat. Med.* 19 (2013) 753-9.
3
4
5 [43] T. Yagi, Y. Hatefi, Thiols in oxidative phosphorylation: inhibition and energy-potentiated
6 uncoupling by monothiol and dithiol modifiers, *Biochemistry* 23 (1984) 2449-55.
7
8
9 [44] T. Yagi, Y. Hatefi, Thiols in oxidative phosphorylation: thiols in the F_O of ATP synthase
10 essential for ATPase activity, *Arch. Biochem. Biophys.* 254 (1987) 102-9.
11
12
13 [45] C.M. Palmeira, V.M.C. Madeira, Mercuric chloride toxicity in rat liver mitochondria and
14 isolated hepatocytes, *Toxicol. Appl. Pharmacol.* 143 (1997) 338-47.
15
16
17 [46] L.P. Karniski, Hg²⁺ and Cu⁺ are ionophores, mediating Cl⁻/OH⁻ exchange in liposomes and
18 rabbit renal brush border membranes, *J. Biol. Chem.* 267 (1992) 19218-25.
19
20
21
22 [47] B.O. Lund, D.M. Miller, J.S. Woods, Studies on Hg(II)-induced H₂O₂ formation and
23 oxidative stress *in vivo* and *in vitro* in rat kidney mitochondria, *Biochem. Pharmacol.* 45
24 (1993) 2017-24.
25
26
27 [48] J.H. Southard, J.T. Penniston, D.E. Green, Induction of transmembrane proton transfer by
28 mercurials in mitochondria. I. Ion movements accompanying transmembrane proton
29 transfer, *J. Biol. Chem.* 248 (1973) 3546-50.
30
31
32 [49] H. Shara, Z. Drahotka, Inhibition of mitochondrial ATPase by Hg⁺⁺ ions, *Physiol.*
33 *Bohemoslov.* 27 (1978) 193-8.
34
35
36 [50] S. Pagani, F. Bonomo, P. Cerletti, Sulfhydryl and disulfide content of succinate
37 dehydrogenase, *FEBS Lett.* 39 (1974) 139-43.
38
39
40 [51] T.K. Lin, G. Hughes, A. Muratovska, F.H. Blaikie, P.S. Brookes, D. Darley-Usmar, R.A.J.
41 Smith, M.P. Murphy, Specific modification of mitochondrial protein thiols in response to
42 oxidative stress—a proteomics approach, *J. Biol. Chem.* 277 (2002) 17048-56.
43
44
45 [52] P.R. Rich, A. Maréchal, Electron transfer chains: Structures, mechanisms and energy
46 coupling, *Comp. Biophys.* 8 (2012) 73-93.
47
48
49
50
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60
61
62
63
64
65
- [53] S. Nesci, F. Trombetti, V. Ventrella, A. Pagliarani, The *c*-Ring of the F₁F₀-ATP Synthase: Facts and Perspectives, *J. Membr. Biol.* 249 (2016) 11-21.
- [54] J.E. Walker, The ATP synthase: the understood, the uncertain and the unknown, *Biochem. Soc. Trans.* 41 (2013) 1-16.
- [55] L.F. Olsen, A.Z. Andersen, A. Lunding, J.C. Brasen, A.K. Poulsen, Regulation of glycolytic oscillations by mitochondrial and plasma membrane H⁺-ATPases, *Biophys. J.* 96 (2009) 3850-61.
- [56] J.N. Mazerik, H. Mikkilineni, V.A. Kuppusamy, E. Steinhour, A. Peltz, C.B. Marsch, P. Kuppusamy, N.L. Parinandi, Mercury activates phospholipase A₂ and induces formation of arachidonic acid metabolites in vascular endothelial cells, *Toxicol. Mechan. Methods* 17 (2007) 1-17.
- [57] R. Requejo, E.T. Chouchani, A.M. James, T.A. Prime, K.S. Lilley, J.M. Fearnley, M.P. Murphy, Quantification and identification of mitochondrial proteins containing vicinal dithiols, *Arch. Biochem. Biophys.* 504 (2010) 228-235.
- [58] A. Miseta, P. Csutora, Relationship between the occurrence of cysteine in proteins and the complexity of organisms, *Mol. Biol. Evol.* 17 (2000) 1232-9.
- [59] S.M. Marino, V.N. Gladyshev, Cysteine function governs its conservation and degeneration and restricts its utilization on protein surfaces, *J. Mol. Biol.* 404 (2010) 902-16.

Table 1. Effects of 2.0 μM HgCl_2 , thiol reagents (1.5 μM NEM, 1.0 μM MBBr and 50 μM PAO) and of the reducing agent DTE on the mitochondrial F-ATPase activity.

CONDITION	no DTE	DTE
Control	2.67 \pm 0.17	2.69 \pm 0.09
HgCl_2	3.34 \pm 0.23*	2.73 \pm 0.14
NEM	2.52 \pm 0.03	2.67 \pm 0.04
MBBr	2.68 \pm 0.31	2.63 \pm 0.27
PAO	3.21 \pm 0.29*	2.55 \pm 0.18
HgCl_2 + (NEM)	3.22 \pm 0.16*	2.74 \pm 0.03
NEM + (HgCl_2)	3.31 \pm 0.04*	2.79 \pm 0.05
HgCl_2 + (MBBr)	3.30 \pm 0.17*	2.58 \pm 0.19
MBBr + (HgCl_2)	3.22 \pm 0.34*	2.77 \pm 0.29
HgCl_2 + (PAO)	3.45 \pm 0.29*	2.80 \pm 0.46
PAO + (HgCl_2)	2.62 \pm 0.18	2.37 \pm 0.13

Thiol reagents were either incubated with mitochondria as detailed in Section 2.4 or, if in brackets, directly added to the reaction system; 50 μM DTE was added to the reaction system as reported in Section 2.5. ATPase activity values ($\mu\text{mol Pi} \cdot \text{mg protein}^{-1} \cdot \text{hr}^{-1}$) are the mean \pm SD of three different determinations on distinct mitochondrial preparations. The asterisk (*) indicates significantly different values from the control ($P \leq 0.05$).

Table 2. Effect of HgCl₂ on kinetic parameters of the mitochondrial F-ATPase activity.

HgCl ₂ (μM)	0.0	2.0	0.0	2.0
	V_{\max}		K_m	
	(μmol P _i ·mg protein ⁻¹ ·min ⁻¹)		(μM)	
Mg ²⁺	3.4±0.2a	5.0±0.1b	0.3±0.0a	0.4±0.1a
ATP	5.9±1.5a	8.2±1.1b	4.9±0.8a	6.0±0.6a

Data are the mean ± SD of n=3 (for Mg²⁺ parameters) and n=4 (for ATP parameters) sets of experiments carried out on different mitochondrial preparations. In each row different letters indicate significantly different values ($P \leq 0.05$).

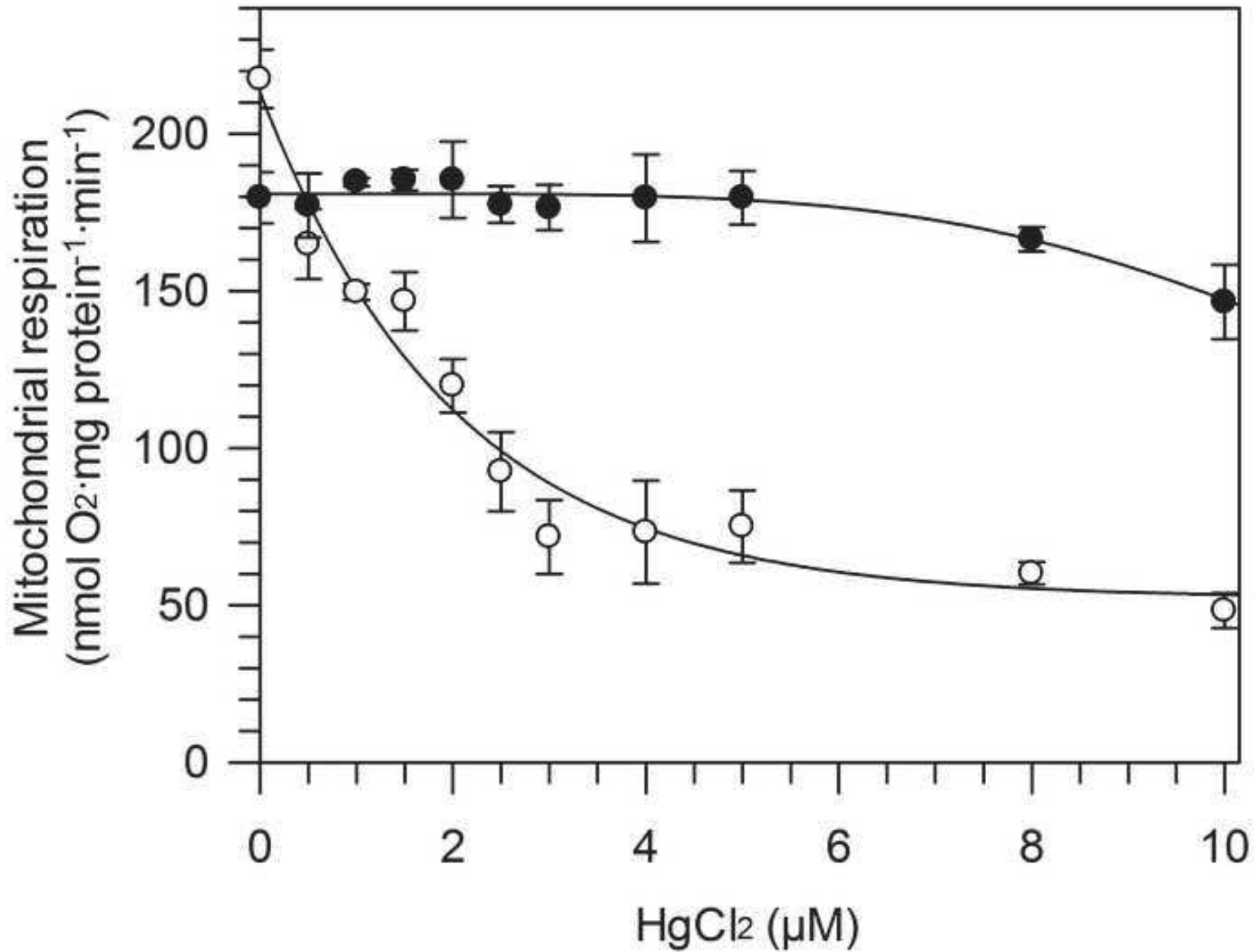
Table 3. Kinetic parameters of the F-ATPase activation by Mg^{2+} and ATP

	α	α'	K_a (μM)	K'_a (μM)
Mg^{2+}	0.9	0.7	17.6	6.3
ATP	0.9	0.7	17.3	6.6

These parameters were calculated as detailed in the text.

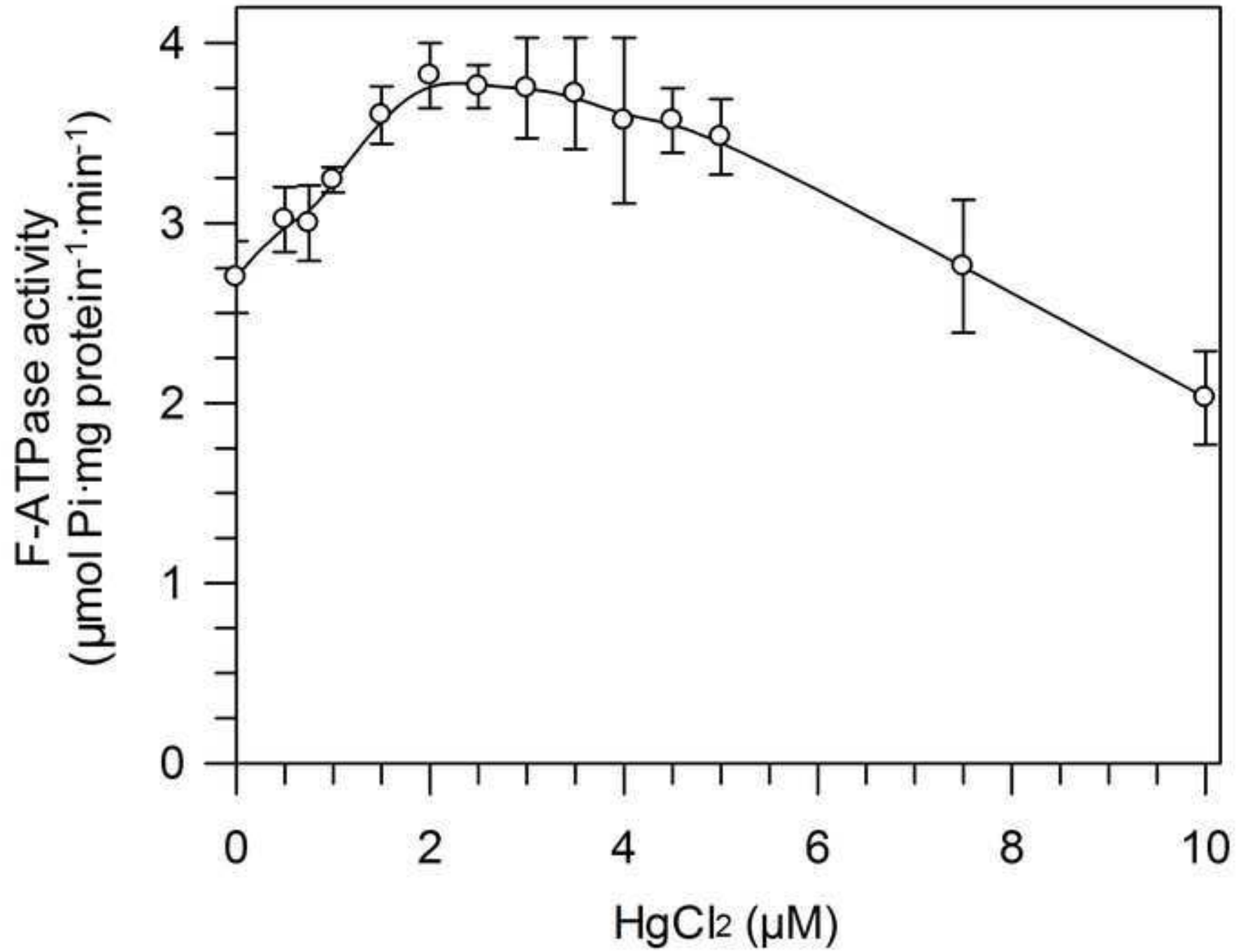
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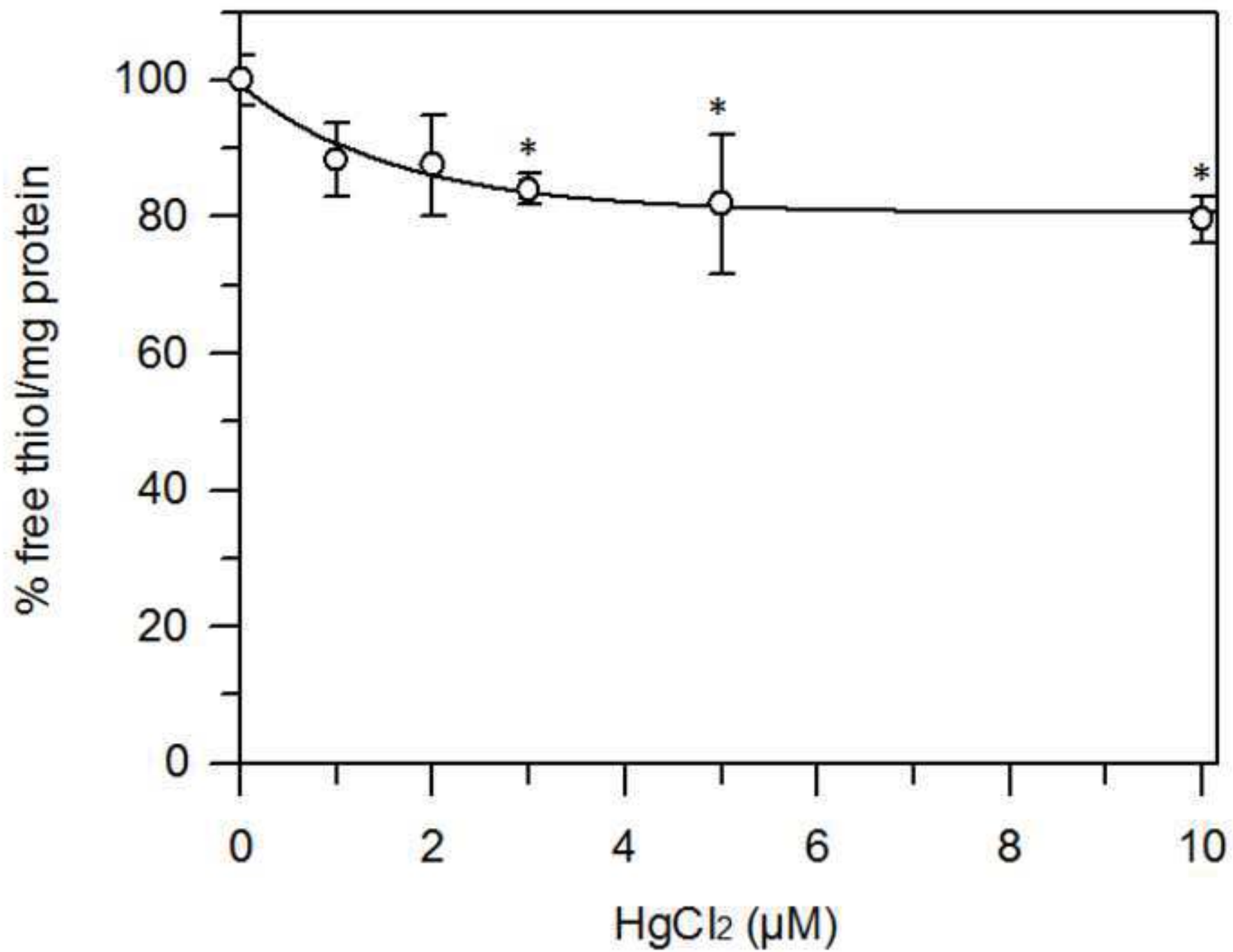
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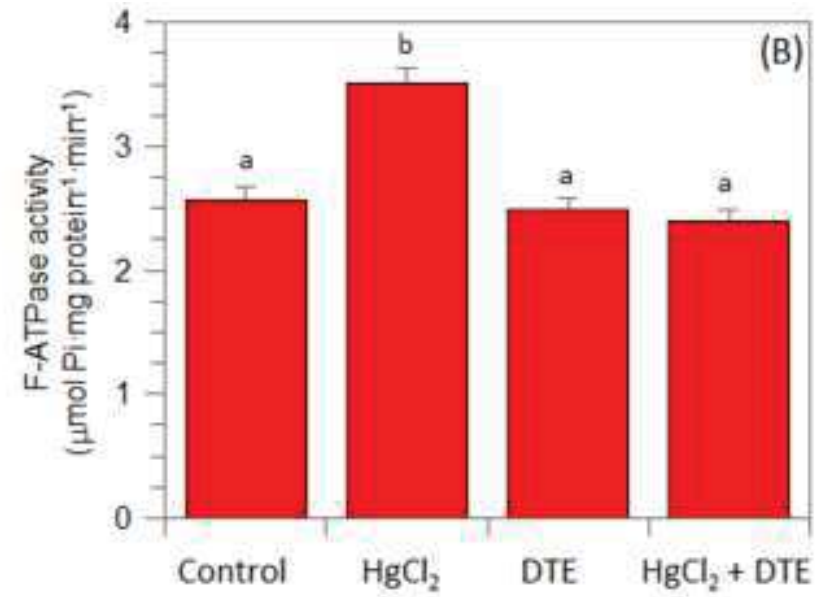
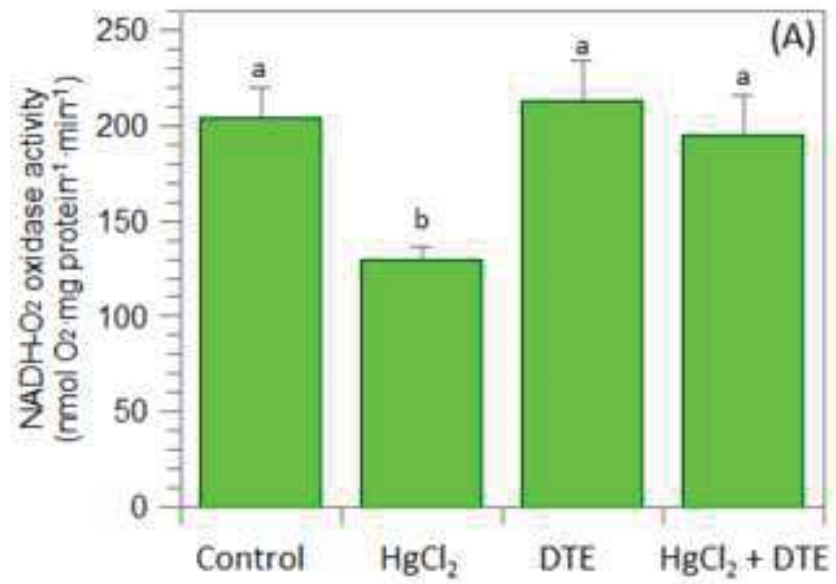
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