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An image-based kinematic model of the tibiotalar and subtalar joints and its application to gait analysis in children with Juvenile Idiopathic Arthritis

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

An image-based kinematic model of the tibiotalar and subtalar joints and its application to gait analysis in children with Juvenile Idiopathic Arthritis / Montefiori, Erica; Modenese, Luca; Di Marco, Roberto; Magni-Manzoni, Silvia; Malattia, Clara; Petrarca, Maurizio; Ronchetti, Anna; de Horatio, Laura Tanturri; van Dijkhuizen, Pieter; Wang, Anqi; Wesarg, Stefan; Viceconti, Marco; Mazzà, Claudia; MD-PAEDIGREE Consortium. - In: JOURNAL OF BIOMECHANICS. - ISSN 0021-9290. - STAMPA. - 85:(2019), pp. 27-36.

This version is available at: https://hdl.handle.net/11585/726505 since: 2021-08-18

Published:

DOI: http://doi.org/10.1016/j.jbiomech.2018.12.041

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Montefiori, E. et al., 2019. An image-based kinematic model of the tibiotalar and subtalar joints and its application to gait analysis in children with Juvenile Idiopathic Arthritis. *Journal of biomechanics*, 85, pp.27–36.

The final published version is available online at:

https://doi.org/10.1016/j.jbiomech.2018.12.041

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1	An image-based kinematic model of the tibiotalar and subtalar joints and
2	its application to gait analysis in children with Juvenile Idiopathic Arthritis
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19	SUBMITTED TO JOURNAL OF BIOMECHANICS
20	ON 13 th February 2018
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30 Abstract

31 In vivo estimates of tibiotalar and the subtalar joint kinematics can unveil unique information about gait 32 biomechanics, especially in the presence of musculoskeletal disorders affecting the foot and ankle complex. 33 Previous literature investigated the ankle kinematics on ex vivo data sets, but little has been reported for 34 natural walking, and even less for pathological and juvenile populations. This paper proposes an MRI-based 35 morphological fitting methodology for the personalised definition of the tibiotalar and the subtalar joint axes during gait, and investigated its application to characterise the ankle kinematics in twenty patients 36 37 affected by Juvenile Idiopathic Arthritis (JIA). The estimated joint axes were in line with in vivo and ex 38 vivo literature data and joint kinematics variation subsequent to inter-operator variability was in the order 39 of 1°. The model allowed to investigate, for the first time in patients with JIA, the functional response to 40 joint impairment. The joint kinematics highlighted changes over time that were consistent with changes in 41 the patient's clinical pattern and notably varied from patient to patient. The heterogeneous and patient-42 specific nature of the effects of JIA was confirmed by the absence of a correlation between a semiquantitative MRI-based impairment score and a variety of investigated joint kinematics indexes. In 43 44 conclusion, this study showed the feasibility of using MRI and morphological fitting to identify the 45 tibiotalar and subtalar joint axes in a non-invasive patient-specific manner. The proposed methodology 46 represents an innovative and reliable approach to the analysis of the ankle joint kinematics in pathological juvenile populations. 47

48

Key words: Biomechanics, Ankle joint axis, Musculoskeletal modelling, Gait analysis, Patient-specific
 modelling

52 Introduction

53 Functional anatomy literature describes the ankle joint as a very complex structure allowing for multiple 54 movements due to the combination of various mechanically coupled joints, including the tibiotalar (i.e. 55 between tibia and talus) and subtalar (i.e. between talus and calcaneus) joints (Hicks et al., 1953; Siegler et 56 al., 1988; Dettwyler et al., 2004). The biomechanical behaviour of the ankle during locomotion and its 57 relationship with the anatomy have been investigated since the beginning of the last century (Fick, 1911; 58 Manter, 1941; Barnett and Napier, 1952; Isman and Inman, 1969; Inman, 1976) and many authors have 59 also estimated the kinematics of the tibiotalar and subtalar joints ex vivo (Hicks et al., 1953; Rasmussen and 60 Tovborg-Jensen, 1982; van Langelaan, 1983; Siegler et al., 1988). The possibility of estimating the kinematics of the ankle's intrinsic joints from *in vivo* data is of interest when investigating musculoskeletal 61 62 diseases. Nonetheless, a comprehensive understanding of the joint's intrinsic movement during walking is 63 still lacking. This is because measuring the motion associated to foot inversion/eversion is not trivial and 64 most literature has focused on the quantification of articular range of motion (ROM) for the various joint's 65 degrees of freedom (DOFs) under controlled conditions (Lundberg et al., 1989; Mattingly et al., 2006; 66 Lewis et al., 2009).

67 In vivo tracking of the relative movement of the talus relative to the calcaneus using skin markers and a 68 standard gait analysis technique is complicated by the small size of these bones and the absence of visible 69 superficial landmarks (Scott et al., 1991; Di Marco et al., 2016). Few studies have investigated the 70 kinematics of the intrinsic joints of the ankle during walking and running (Arndt et al., 2004 and 2006) 71 using intracortical bone pins, and compared the results to those from using superficial markers (Westblad 72 et al., 2002). These studies clearly showed a description of plantar/dorsiflexion is possible with traditional 73 gait analysis methods, however, estimates of inversion/eversion movement are still far from being accurate. Intracortical pin-based studies partially overcome this lack of accuracy but, due to the invasiveness of the 74 75 technique, the number of participants is usually limited to few healthy volunteers, whose natural gait pattern 76 can be altered by the possible pain and discomfort related to the implant. Both in vivo and ex vivo studies 77 reported high intra-subject and inter-subject variability in the subtalar joint kinematics with ROM up to 60° 78 (Roaas and Anderson, 1982; Sepic et al., 1986; Lundberg, 1989).

79 The functional complexity of the subtalar joint led to a number of different modelling approaches, from the 80 attempt to capture its mobility through multi-segmental foot models where the subtalar articulation was 81 interpreted as a motion between hind-foot and fore-foot (Prinold et al., 2016; Saraswat et al., 2010), to a 82 more anatomical representation as a universal or hinge joint (Delp et al., 1990; Malaquias et al., 2017). The 83 hinge-like schematisation also applies to the tibiotalar joint and this approach is currently used within 84 widely adopted musculoskeletal models (Delp et al., 1990). When simultaneously modelling both joints as 85 hinges (Dul and Johnson, 1985), a reasonable simplification is made with respect to their real functional role (Siegler et al., 1988), according to which the tibiotalar and subtalar joints describe 86 87 the plantar/dorsiflexion and inversion/eversion motions, respectively. This latter motion, despite its 88 simplified appearance, is justified because the predominant motion occurs about a single axis of rotation 89 (Scott and Winter, 1991). However, this DOF has been reported to be less accurately described with current 90 musculoskeletal modelling approaches, mainly due to the difficulties in identifying the joint functional axis 91 in vivo (Van den Bogert et al., 1994; Dettwyler et al., 2004; Parr et al., 2012). A high variability within-92 and between-subjects has been observed in the modelled joint axes, which is also related to the specific 93 locomotion task (Leitch et al., 2010). In the presence of musculoskeletal disorders, the adoption of image-94 based patient-specific modelling approaches has been previously proposed (Prinold et al., 2016; Hannah et 95 al., 2017) and proved to increase anatomical modelling accuracy (Correa and Pandy 2011; Durkin et al., 96 2006; Schevs et al., 2009). The use of this technique accounts for patients' anatomical features and 97 peculiarities, crucial when impairments and gait limitations affect the subjects. In this study, we propose an 98 image-based modelling procedure to define the tibiotalar and subtalar joints axes, avoiding operator-99 dependent steps and related variability issues (Prinold et al., 2016; Hannah et al., 2017). Once compared 100 against literature, the procedure will be used as part of a patient-specific musculoskeletal modelling 101 approach to investigate the gait ankle kinematics in children with Juvenile Idiopathic Arthritis (JIA), a

paediatric group of diseases of unknown aetiology characterised by joint inflammation potentially leading to cartilage damage. Altered gait patterns and physical disabilities (Ravelli and Martini, 2007) are possible outcomes in JIA. This longitudinal study will prove whether our modelling approach is capable of detecting clinical changes observed in the tibiotalar and the subtalar joint functions and quantify for the first time the relationship between these changes and the underlying joint impairments.

107 Methods

108 **2.1 Subjects and data acquisition**

Twenty participants (5 males, 15 females, age: 11.6±3.1 years, mass: 47.6±18.2 kg, height: 148±17 cm, 11 new onsets) affected by Juvenile Idiopathic Arthritis (JIA) of various sub-types (oligoarticular onset JIA, polyarticular JIA, psoriatic arthritis, and undifferentiated arthritis) (Ravelli and Martini, 2007) were recruited among those referred to two different children's hospitals (Istituto Giannina Gaslini, Genoa (Lab 1), and "Bambino Gesù" Children's Hospital, Rome (Lab 2)). The study was conducted following Helsinki's declaration on human rights and was approved by the ethical committee of both hospitals. Written informed consent was obtained by patients' parents.

Medical resonance images (MRI) and gait analysis data were collected at three time-points (6 months apart) 116 117 to follow the disease progression. The imaging performed at month 0 (M0) and month 12 (M12) included 118 a foot and ankle regional MRI (multi-slice multi-echo 3D Gradient Echo (mFFE) with water-only selection 119 (WATS) with 0.5 mm in-plane resolution and 1 mm slice thickness). The month 6 (M6) imaging included 120 a full lower limb MRI (3D T1-weighted fat-suppression sequence (e-THRIVE) with 1mm in-plane 121 resolution and 1mm slice thickness). The core set of basic sequences and definitions suggested by the 122 Outcome Measure in Rheumatology (OMERACT) MRI Working Group (Ostergaard et al., 2003; Nusman 123 et al., 2016) was used to provide an MRI-based evaluation of the joints (Table I). A weighted, average index (I_{MRI}) was used to quantify the overall level of impairment of the foot and ankle region. 124

Table I - MRI scoring.

Index	MRI sequence	Scale	Sites
Bone erosion	T1-weighted fat-saturated	Range 0-10 % of eroded articular surface <i>(Ostergaard et al., 2003)</i> 0 = no erosion; 1 = 1–10%; 2 = 11-20%; 3 = 21–30%; 4 = 31–40%; 5 = 41–50%; 6 = 51–60%; 7 = 61–70%; 8 = 61–80%; 9 = 81–90%; 10 = 91–100%	Distal tibial epiphysis Distal fibula epiphysis Tarsal bones Metatarsal bases
Cartilage damage	WATS	Range 0-3 % of damaged cartilage surface 0 = no damage; 1 = 1-33%; 2 = 34-66%; 3 = 67-100%; 4 = extensive damage causing ankyloses	Tibiotalar Between distal talus and calcaneus, Talonavicular Calcaneocuboid Cuneonavicular Between cuneiforms and I, II and III metatarsal bones Between cuboid and IV and V metatarsal bones
Synovitis	T1-weighted fat-saturated	Range 0-3 Degree of synovial enhancement and synovial thickness (Ostergaard et al., 2003; Malattia et al., 2011) 0 = normal; 1 = mild; 2 = moderate; 3 = severe	Tibio-peroneo-talar Subtalar Talonavicular Calcaneocuboid I-V tarsometatarsal Cuneonavicular
Tenosynovitis	T1-weighted fat-saturated with enhancement	Range 0-3 Degree of peritendinous effusion or synovial proliferation 0 = normal; 1 = mild (< 2 mm); 2 = moderate (2 -5 mm); 3 = severe (> 5 mm)	Anterior tibial Extensor digitorum longus Extensor hallucis longus Posterior tibial Flexor digitorum longus Flexor hallucis longus Peroneal tendons

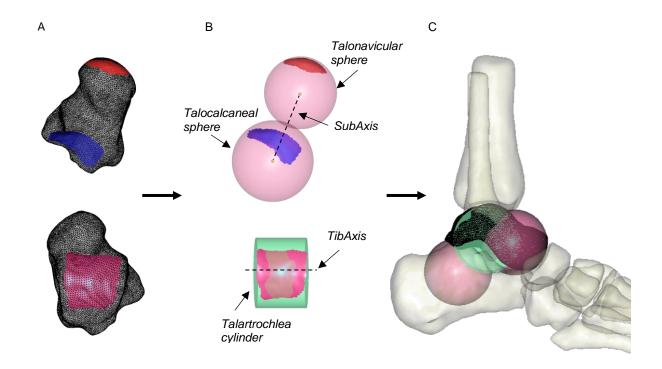
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Gait analysis was based on stereophotogrammetry and data were collected using a 6-camera system (BTS, Smart DX, 100Hz) with two force plates (Kistler, 1kHz) in Lab 1, and an 8-camera system (Vicon, MX, 200Hz) and two force plates (AMTI, OR6, 1kHz) in Lab 2. Five walking trials at self-selected speed were performed and a minimum of three trials were used for the analysis. The marker set included forty-four markers from the Vicon Plug in gait protocol (Vicon Motion System) and the modified Oxford Foot Model (mOFM) protocol (Stebbins et al., 2006). A subset of MRI-visible markers (twenty-eight in the lower limb MRI and six in the regional MRI scans) was retained during the imaging acquisition for data registration.

Despite being collected in different centres and with different equipment, the raw-data underwent the same pre-processing in terms of labelling, gap-filling (spline algorithm built in Vicon Nexus 1.8.5 (Woltring et al., 1986)), and smoothing (4th-order Butterworth filter, 6Hz cut-off (Barlett et al., 2007)).

137 **2.2 Anatomical model**

138 A statistical shape modelling approach (Steger et al., 2012) was used to segment the lower limb bones from 139 the MRI and subject-specific anatomical models were produced using specialised software (NMSBuilder, 140 Valente et al., 2017). For each patient, two bilateral three-segment anatomical models were built using the 141 M0 and M12 datasets, resulting in 80 foot models. Twelve of these were excluded due to incompleteness 142 of the experimental dataset, resulting in a final dataset of 68 feet. The joints' reference frames, namely 143 tibiotalar joint (between tibia and talus) and subtalar (between talus and foot) were defined according to the 144 ISB conventions (Baker et al., 2003) and the joint axes were identified through morphological fitting of 145 articular surfaces (Figure 1A-C). The subtalar joint axis (SubAxis) was defined as the axis connecting the 146 centres of the spheres fitted to the anterior (Talonavicular sphere) and to the posterior-inferior 147 (*Talocalcaneal sphere*) facets of the talus respectively (Figure 1B). This was similar to that proposed by 148 Parr et al., 2012, who, however, used the anterior-inferior portion of the talus surface to define the 149 Talonavicular sphere. To define the tibiotalar joint axis (*TibAxis*), a cylinder was fitted to the entire trochlea 150 (*Talartrochlea cylinder*) as a simplification of the approach proposed by Siegler et al., 2014 (Modenese et 151 al., 2018). The fitting was implemented in Meshlab (Cignoni et al., 2008) by identifying the articular 152 surfaces from the segmented geometries and minimising the least squares distance between the identified 153 surface and the corresponding best fitting analytical shape (Least Squares Geometric Elements library, 154 Matlab). The distal tibia (segmented from the M0/12 MRI) was afterwards registered to the entire tibia (M6 155 dataset) using the Iterative Closest Point algorithm in Meshlab to obtain a full lower limb model. A 156 comprehensive description of the modelling procedure is available as supplementary material in Modenese 157 el al. (2018). The subset of instructions used for producing the proposed ankle kinematic model is also 158 available on Figshare together with the data and models (doi:).



160Figure 1 - (A) Plantar (top) and dorsal (bottom) views of the right talus (black wireframe) with161highlighted articular regions: anterior facet (red), posterior-inferior facet (blue), trochlea (fuchsia). (B)162Fitting of analytical shapes to the selected articular regions: two spheres (light pink) identify the axis of163the subtalar joint (SubAxis) as the axis connecting the centres of the spheres and a cylinder (light green)164identifies the axis of the tibiotalar joint (TibAxis) as the cylinder axis. (C) Example of the fitted165geometries integrated within the ankle anatomical model.

166

167 **2.3 Joint kinematics**

The OpenSim's (Delp et al., 2007) Inverse Kinematics (IK) tool was run to estimate the tibiotalar and subtalar joint angles starting from a set of sixteen skin markers (five on the tibia, eleven on the foot, Figure 2), eight were also virtually palpated on the medical images. The difference between the virtual and experimental markers estimated by the IK tool was less than 1cm on average over all the time-steps, as suggested in the OpenSim best practice recommendations (Hicks et al., 2015).

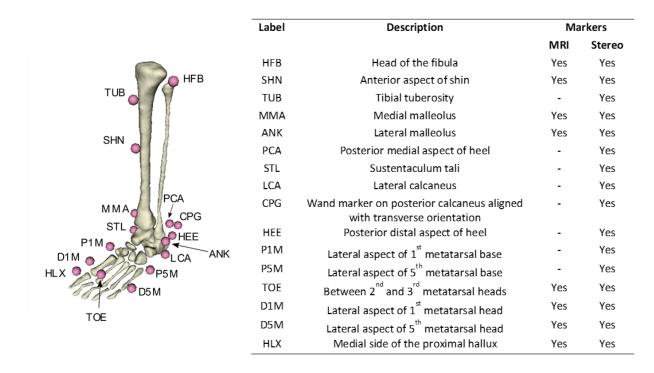


Figure 2 - Experimental markers used in the imaging (MRI) and stereo-photogrammetric (Stereo)
 measurements.

176 **2.4 Model evaluation**

177 Sensitivity to operator-dependent input

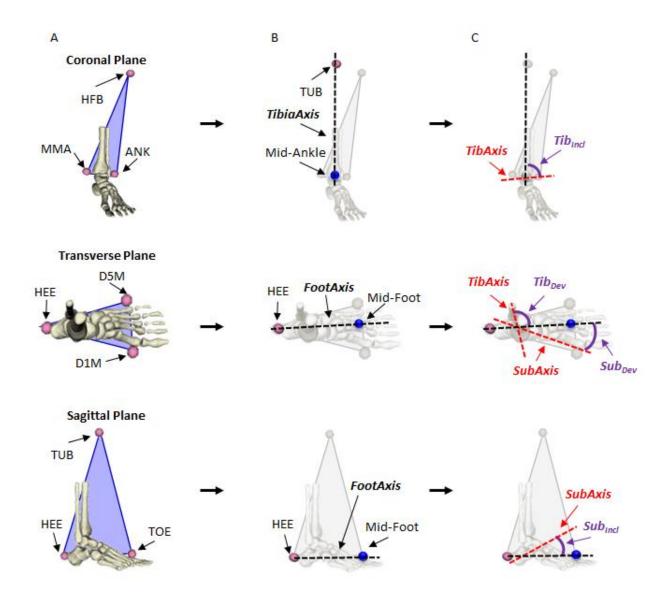
The bone segmentations from three randomly chosen patients were used to investigate the effect of operator-dependent variability in the definition of *TibAxis* and *SubAxis*. Three operators repeated the morphological fitting three times and the coordinates of the *Talartrochlea cylinder*, *Talocalcaneal sphere* and *Talonavicular sphere* centres were used for the comparison. A 3D quantification of their variability (SD_{3d}) was calculated from the standard deviation of the point coordinates (sd_x,sd_y,sd_z) as:

183
$$SD_{3d} = \sqrt{sd_x^2 + sd_y^2 + sd_z^2}$$

For the foot that led to the worst-case scenario (higher inter-operator SD_{3d}), a second level of analysis was conducted to quantify the propagation of this error on the joint kinematics. The nine models built by the three operators were then used to estimate the tibiotalar and subtalar joint kinematics using data from one randomly selected gait trial from the same patient. The maximum value of the mean and standard deviation
calculated over the nine repetitions for each point of the gait cycle was then used to quantify the maximum
expected error.

190 **Consistency with literature data**

191 Among the 68 available models, 38 were selected (19 per side, preferentially from M12) to conduct the 192 following analysis. A standing trial collected during the gait analysis session was used to identify the pose 193 of each subject and the resulting neutral position of the foot. The transverse, sagittal, and coronal anatomical 194 planes, the midline of the foot (FootAxis) and the long axis of tibia (TibiaAxis) were identified using the 195 standing trial markers (Figure 3A-B). These allowed quantifying the tibiotalar inclination (*Tib_{Incl}*) and 196 deviation (Tib_{Dev}), and the subtalar inclination (Sub_{Incl}) and deviation (Sub_{Dev}) as shown by the angles in 197 Figure 3C. Tib_{Incl}, Tib_{Dev}, Sub_{Incl} and Sub_{Dev} were compared to literature data from ex vivo cadaveric 198 specimens (Isman and Inman., 1969; Inman, 1976) and from healthy adults (Van den Bogert et al., 1994). 199 The estimations of *TibAxis* and *SubAxis* at M0 and M12 were also compared. All 26 models for which the 200 3D anatomy was available at both time-points (52 models) were used for a between-session comparison. 201 For this analysis, the angle between the two joint axes (*InterAxis*) was preferred over the measures of *Tiblaci*, 202 Tib_{Dev} , $Sub_{Incl.}$ and Sub_{Dev} to avoid the effect of experimental markers repositioning (between the two sessions) on these angles. Mean and maximum between-session variations were quantified, and a paired-203 204 two-sided Wilcoxon signed-rank test ($\alpha = 0.05$) was performed under the null hypothesis showed that no 205 statistical difference existed between the two repeated measures. This was intended as a repeatability 206 assessment of the proposed method, assuming in the investigated age range, and within 12 months, neither 207 disease progression (Ravelli and Martini, 2007) nor growth (Evans, 2010) would cause changes in the joint 208 morphology.



²⁰⁹ 210

211 Figure 3 - (A) Identification of anatomical planes (blue triangles) as defined using the virtual markers 212 (pink) corresponding to the experimental markers listed in Figure 2. (B) Definition of the anatomical 213 axes (midline of the foot = FootAxis, long axis of the tibia = TibiaAxis, black dashed lines) by calculating 214 average points (blue markers) between virtual marker pairs (Mid-Foot = midpoint between D1M and 215 D5M; Mid-Ankle = midpoint between ANK and MMA). (C) Quantification of the inclination (TibIncl) 216 and deviation (TibDev) of tibiotalar joint and inclination (SubIncl) and deviation (SubDev) of subtalar 217 joint as the angles (purple arches) between the anatomical axes and the joint axes (red dashed lines) as 218 defined through morphological fitting (Figure 1).

219 Effect of clinical impairment on joint kinematics

220 The models from 13 subjects (3 males, 10 females, age: 11.0 ± 3.1 years, mass: 44.5 ± 16.9 kg, height: 143

 \pm 13 cm, 8 new onsets), for whom both clinical and biomechanical information was available, were used to

222 test the link between changes in the kinematics and impairment of the ankle as measured from the MRI. 223 The I_{MRI} scores were used to classify the disability level of each ankle and identify better and worse time-224 points. They were then placed into "low-involvement" and "high-involvement" groups accordingly. The 225 joint kinematics of the two groups were then compared using a non-parametric 1D two-tailed paired *t*-test 226 $(\alpha=0.05)$ (Nichols and Holmes, 2002) based on Statistical Parametric Mapping (SPM) in MATLAB (v9.1, 227 R2016b, Mathworks, USA), using the SPM1D package (Pataky et al., 2012). This was chosen since the 228 data were not normally distributed. The following kinematic parameters were also calculated to investigate 229 the correlation with the I_{MRI} : area under the curves of the tibiotalar and subtalar joint angles, maximum 230 plantarflexion (PF) and dorsiflexion (DF) angles, maximum inversion (Inv) and eversion (Ev) angles, and 231 joint ROM. Furthermore, the asymmetry between the left and right foot kinematics was quantified using 232 the Root Mean Square Deviation (RMSD) and Mean Absolute Variability (MAV) (Di Marco et al., 2018), 233 as well as the between-side difference of ROM and standard deviations (SD). RMSD, MAV, ROM and SD 234 were measured at the two time-points and compared using a two-sided Wilcoxon signed-rank test (α =0.05). 235 The absolute difference (ΔI_{MRI}) between left and right I_{MRI} was also calculated and a correlation analysis 236 was used to assess whether an asymmetry in the clinical score, namely higher ΔI_{MRI} , corresponded to higher values of the kinematic parameters. 237

238 **Results**

239 Sensitivity to operator-dependent input

 SD_{3d} of *Talonavicular sphere* and *Talocalcaneal sphere's* centres are reported in Table II, as well as the resulting maximum angular variability of the *TibAxis* and *SubAxis*, whose maximum value (9.6°) was found for the inclination of *SubAxis* in patient P3. For this patient, the propagation of inter-operator variability on the articular kinematics introduced a maximum standard deviation of 0.6° and 1.3° for the tibiotalar and subtalar joints respectively, both occurring at 63% of the gait cycle.

	Talartrochlea center	Talonavicular center	Talocalcaneal center	TibAxis	SubAxis
Patients	<i>SD</i> _{3<i>d</i>} [mm]	<i>SD</i> _{3<i>d</i>} [mm]	<i>SD</i> _{3<i>d</i>} [mm]	SD [°]	SD [°]
P1	0.4	0.4	1.4	0.6	1.7
P2	0.5	0.8	1.5	0.8	1.3
P3	0.8	2.1	5.1	2.0	5.6

247 Consistency with literature data

The residual error of the fitting algorithm (average (\pm SD) across the 52 models) was equal to 0.16 (\pm 0.05) mm, 0.48 (\pm 0.21) mm, and 0.28 (\pm 0.11) mm for the *Talonavicular*, *Talocalcaneal*, and *Talartrochlea* surfaces, respectively. The average (\pm SD) values of the measured foot angles (*Tib_{Incl}*, *Tib_{Dev}*, *Sub_{Incl}*, and *Sub_{Dev}*) (Table III) were found to be in line with the corresponding *ex vivo* (Isman and Inman., 1969; Inman, 1976) and *in vivo* (Van den Bogert et al., 1994) measurements available in the literature. The average absolute difference between the M0 and M12 measures of *InterAxes* was 2.2° \pm 2.1°, which was not statistically significant (Wilcoxon test p=0.648).

255 256

Table III - Inclination and deviation of tibiotalar and subtalar joint axes and comparison with published literature datasets (n = numebr of subjects).

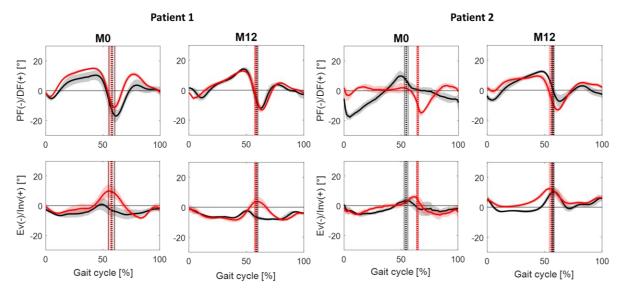
Angle	lsman and Inman, 1969 (n=46) mean (±SD) [°]	Inman, 1976 (n=104) mean (±SD) [°]	Van den Bogert, 1994 (n=14) mean (±SD) [°]	This study (n=38) mean (±SD) [°]
Gender	NA	NA	males	30 females/8 males
Age	Adults (age not specified)	Adults (age not specified)	Adults (age not specified)	11.2±3.1 years
TibIncl	80(±4)	82.7(±3.7) <i>(n=107)</i>	85.4(±7.4)	90.7(±4.1)
TibDev	84(±7)	-	89.0(±15.1)	82.7(±7.4)
SubIncl	41(±9)	42(±9)	35.3(±4.8)	41.1(±14.1)
SubDev	23(±11)	23(±11)	18.0(±16.2)	27.0(±9.0)

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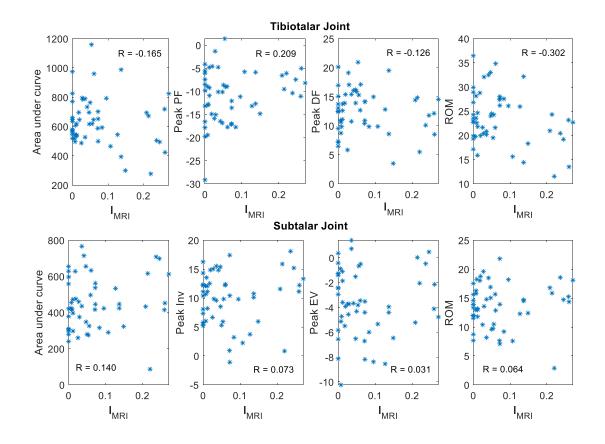
258 Effect of clinical impairment on joint kinematics

259 Figure 4 shows the estimated kinematics of two subjects with different clinical scoring: patient 1 was 260 similarly affected by the pathology at the two observations, whereas at M12 patient 2 was in total remission, as defined by Ravelli and Martini (2007). This example highlights how the models clearly capture different 261 262 kinematic patterns associated with different paths of disease progression. The observation of the joint angles 263 also clearly indicates the ability of the model to describe changes in the gait patterns happening between 264 the two time-points, which were also confirmed by consistent changes in the walking speed $(1.51\pm0.05 \text{ m/s})$ 265 at M0 and 1.22±0.05m/s at M12 for subject 1; 0.83±0.03m/s at M0 and 1.20±0.04m/s at M12 for subject 266 2). For the whole cohort, walking speed varied from 1.01 ± 0.24 m/s at M0 to 1.12 ± 0.13 m/s at M12, and was 1.14±0.17m/s and 0.93±0.33m/s at the "low-involvement" and "high-involvement" time-points 267 respectively, with no significant difference. Walking speed values did not correlate with the joint 268 269 impairment level, as measured with the I_{MRI} (R=-0.21 and R=0.16 at M0 and M12, respectively). Similarly, 270 no correlation was observed between I_{MRI} and the kinematic parameters (Figure 5). This was confirmed by 271 the absence of a group-wise statistically significant difference between the joint kinematics of the ankles at 272 the "low-involvement" and "high-involvement" time-points throughout the gait cycle (Figure 6). Figure 7 273 clearly shows the absence of a significant correspondence between the asymmetry of impairment (ΔI_{MRI}) and the RMSD, MAV, Δ ROM and Δ SD observed at M0 and M12. However, a smaller ΔI_{MRI} at M12 was 274

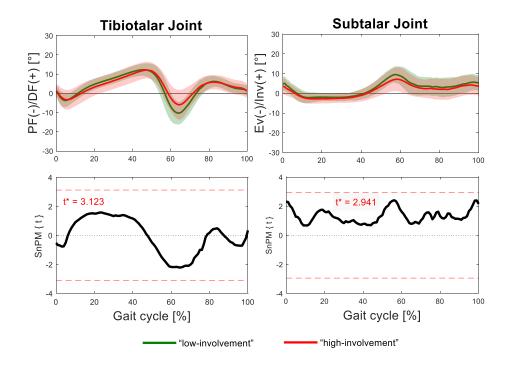
275 generally associated to a smaller value of the kinematics indices at that time-point, except for the Δ SD of 276 the tibiotalar joint and the Δ ROM of the subtalar joint.



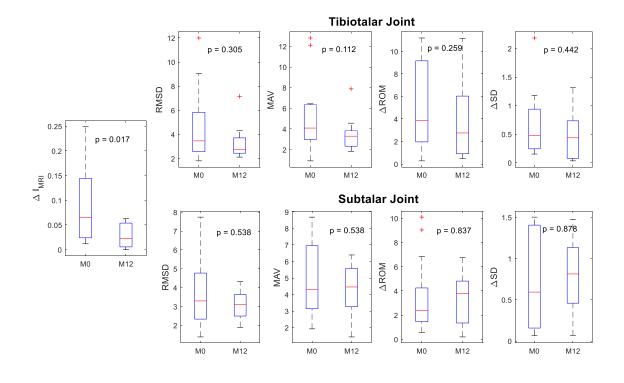
277Figure 4 - Tibiotalar (PF/DF) and subtalar (Ev/Inv) joints kinematics for two JIA patients at M0 and278M12. Average right (left) kinematics is shown with black (red) solid line with shadow representing ± 1 279standard deviation. Toe off is shown with dotted vertical lines ± 1 standard deviation (solid vertical lines).280Walking speed changed from 1.51 ± 0.05 m/s at M0 to 1.22 ± 0.05 m/s at M12 for patient 1 and from 0.83281 ± 0.03 m/s at M0 and 1.20 ± 0.04 m/s at M12 for patient 2.



283Figure 5 - Correlation between joint impairment level (I_{MRI}) and joint kinematics parameters (area284under the curve, peak of plantarflexion (Peak PF) and dorsiflexion (Peak DF), peak of Inversion (Peak285Inv) and eversion (Peak Ev), ROM) for all feet and observations.



291Figure 6 - Tibiotalar (PF/DF) and subtalar (Ev/Inv) joint kinematics of the 13 subjects as calculated at292the "low-involvement" (green) and "high-involvement" (red) time-point. Solid lines in the left graphs293represent mean values and bands represent ± 1 standard deviation. The right figures show the294corresponding distribution of *t*-values (SnPM{t}) throughout the gait cycle as obtained from the non-295parametric 1D paired *t*-test (Nichols and Holmes, 2002), calculated using the SPM1D package (Pataky296et al., 2012). Each group includes 24 mono-lateral models (2 models were excluded from the analysis).



300

Figure 7 – Boxplot distribution of ΔI_{MRI} and kinematics indices (RMSD, MAV, ΔROM and ΔSD) for 301 both tibiotalar and subtalar joints (n=13) at M0 and M12. p-values from two-sided Wilcoxon signed-302 rank test are reported in each plot. Data outliers are marked with a +.

Discussion 303

304 The aim of the study was to propose a kinematic model of the tibiotalar and subtalar joints, and to use this 305 model to investigate the ankle joint kinematics in a group of children with JIA. The anatomical model was 306 based on a morphological fitting approach and underwent repeatability analysis.

307 The procedure proved to be robust to the operator-dependent input. Even in the worst-case scenario, where 308 the definition of the subtalar axis was associated with high inter-operator error (9.6°) , the joint kinematics 309 varied less than 1.3°. The inter-operator variability was mainly associated with the quality of the segmented 310 images, i.e. low resolution, bias field or noise in the MRI, and to the complexity of segmenting bone tissue 311 in young subjects, where cortical bone is not completely ossified (Evans, 2010). Nonetheless, this error was 312 still acceptable when compared to other possible sources of variability coming from the experimental errors, 313 such as instrumental error and marker placement error (up to $6^{\circ}\pm 2^{\circ}$ at the toe off (Di Marco et al., 2016)), or soft tissue artefact (up to 20% of variability in the ankle kinematics (Lamberto et al., 2016)), confirming
the chosen morphological fitting approach is suitable in the presence of low quality images and/or poor
bone reconstructions.

317 An *in vivo* validation of the proposed technique was not possible within the framework of this project due 318 to ethics constraint in the use of approaches like dual-fluoroscopy in a paediatric population. However, the 319 comparison with ex vivo (Isman and Inman, 1969; Inman, 1976), and in vivo (Van den Bogert et al., 1994) 320 data certainly support the validity of the technique. Previous studies (Leitch et al., 2010; Van den Bogert et 321 al., 1994) reported the highest between-subject variabilities in the deviation angle (up to 15°); conversely, 322 we found the biggest differences in the inclination of the subtalar axis (14°). This could be ascribed to the 323 subtalar axis' definition relying on the identification of the anterior facet of the talus. In the youngest 324 children, in fact, this surface can present a layer of unossified cartilage (Evans et al., 2010), which can 325 complicate the identification of the bone contour in the MRI, consequently affecting the results of 326 segmentation and morphological fitting.

The second goal of the study involved the application of the modelling approach as part of the clinical gait assessment of patients with JIA. The between-session repeatability showed no statistically significant difference between the measures of *InterAxis* at M0 and M12, confirming our hypothesis.

330 The observed joint kinematics reflected the heterogeneous and patient-specific nature of the pathology, 331 which presents several sub-types, each with a specific progression (Ravelli and Martini, 2007). In fact, the 332 individual differences (Figure 4) were not representative of a group behaviour (Figure 6) as a consequence 333 of different possible evolutions of the disease. The absence of a recognisable group pattern was 334 demonstrated by the lack of a direct relationship between a joint's clinical impairment and its kinematics. 335 The inter-subject variability was probably exacerbated by the heterogeneity of the cohort in terms of age, 336 anthropometry, disease subtype and activity level. This explains the lack of correlation between joint kinematics (and their changes between time points) and the patient's I_{MRI} scores. This also held true for 337 338 the walking speed, which was not correlated with the MRI scores, but was found in line with the

1.17±0.02m/s reported by Esbjörnsson et al., 2015 for a group of JIA children with similar ankle
involvement. If group stratification needs to be pursued, then further investigation should aim at involving
larger subgroups for every sub-type of JIA and matching them by age and size.

342 The analysis of the between-limb asymmetry at the two time-points showed similar trends in the distribution 343 of ΔI_{MRI} and in the observed kinematics indices, despite none of the latter was significantly different 344 between the two time-points. In the tibiotalar articulation, lower ΔI_{MRI} at M12 corresponded to smaller 345 RMSD and MAV, confirming the asymmetry in the clinical involvement of the ankles is reflected by an 346 asymmetry in the biomechanics of gait. The subtalar kinematics was in general less informative and this is 347 probably associated to a smaller ROM of this joint when compared to the tibiotalar joint, potentially 348 resulting in smaller sensitivity to kinematics changes. Furthermore, disease-related alterations in the 349 movement are likely to be compensated by the tibiotalar joint being dominant in the ankle kinematics 350 (Lundberg et al., 1989) and therefore limiting the role of the subtalar joint. The lack of an independent 351 clinical assessment of the two joints must be considered as a limitation in the study. In fact, the present 352 work is based on the assumption that the I_{MRI} score, evaluating the overall condition of the ankle joint, is 353 representative of both tibiotalar and subtalar impairment level. Nonetheless, a different level of involvement 354 of the two joints could justify their different biomechanical response. Lastly, the assumption made in 355 schematising the joints as hinge-like mechanisms represents a substantial simplification of the true 356 articulating surfaces, potentially limiting the representation of their true 3D motion. However, the tibiotalar 357 kinematics was only marginally affected by this modelling choice, as this movement mainly occurs in the 358 sagittal plane (Roach et al., 2016). On the contrary, the subtalar joint might benefit from a more detailed 359 representation and further studies are needed to investigate this aspect.

In conclusion, this study showed the feasibility of using morphological fitting of MRI-based bone segmentation to identify the tibiotalar and subtalar joint axes in a non-invasive patient-specific manner. Including these joints in a musculoskeletal model of the lower limb, coupled with an appropriate marker set, can give a better understanding of their individual contribution to the ankle biomechanics. This supports the adoption of the proposed modelling procedure into the practice of lower limb musculoskeletal modelling for the quantification of ankle biomechanics. The application to a pathological population, children with JIA, unveiled for the first time the absence of correlation between ankle impairment and biomechanical function, confirming the heterogeneous and systemic nature of this disease.

368 **Conflict of interest**

369 The authors declare they do not have any financial or personal relationships with other people or 370 organizations that could have inappropriately influenced this study.

371 Acknowledgments

372 The authors want to acknowledge Dr Norman Powell for the writing assistance. This research was

- 373 supported by the European Commission (MD-PAEDIGREE project, FP7-ICT Programme, Project ID:
- 374 600932) and by the UK EPSRC (Multisim project, Grant number: EP/K03877X/1).

375 **References**

Arndt, A., Westblad, P., Winson, I., Hashimoto, T., Lundberg, A., 2004. Ankle and Subtalar Kinematics
Measured with Intracortical Pins During the Stance Phase of Walking. Foot Ankle Int. 25, 357-364.

Arndt, A., Wolf, P., Nester, C., Liu, A., Jones, R., Howard, D., Stacoff, A., Lundgren, P., Lundberg, A.,
2006. Intrinsic foot motion measured in vivo during barefoot running. Journal of Biomechanics 39, S182S182.

Baker, R., 2003. Letter to the editor: ISB recommendation on definition of joint coordinate systems for the
 reporting of human joint motion—part I: ankle, hip and spine. Journal of Biomechanics 36(2), 300-302

Bartlett, R., 2007. Introduction to sports biomechanics: analysing human movement patterns (2nd Ed.).
Routledge, Abingdon, England

Barnett, G.H. and Napier, J.R., 1952. The Axis of Rotation at the Ankle Joint in Man. Its Influence upon
the Form of the Talus and the Mobility of the Fibula. J. Anat. Lond., 86: 1-9.

Cignoni, P., Callieri, M., Corsini, M., Dellepiane, M., Ganovelli, F. and Ranzuglia, G., 2008. Meshlab: an
 open-source mesh processing tool. Eurographics Italian Chapter Conference.

Correa, T.A., Pandy, M.G., 2011. A mass-length scaling law for modelling muscle strength in the lower
 limb. Journal of Biomechanics 44 (16), 2782-2789

- 391 Delp, S.L., Loan, J.P., Hoy, M.G., Zajac, F.E., Topp, E.L. and Rosen, J.M., 1990. An interactive graphics-
- based model of the lower extremity to study orthopaedic surgical procedures. IEEE Transactions on
 Biomedical Engineering 37, 757-767.
- Delp, S. L., Anderson, F. C., Arnold, A. S., Loan, P., Habib, A., John, C. T., Guendelman, E. and Thelen,
 D. G., 2007. OpenSim: open-source software to create and analyze dynamic simulations of movement.
- 396 IEEE Transactions on Biomedical Engineering 54, 1940-1950.
- Dettwylera, M., Stacoffa, A., Kramers-de Quervaina, I.A., Stüssia, E., 2004. Modelling of the ankle joint
 complex. Reflections with regards to ankle prostheses. Foot Ankle Surg. 10, 109–119.
- 399 Di Marco, R., Rossi, S., Racic V., Cappa, P., Mazzà, C., 2016. Concurrent repeatability and reproducibility
- 400 analyses of four marker placement protocols for the foot-ankle complex. Journal of Biomechanics 49,401 3168–3176.
- 402 Di Marco, R., Scalona, E., Pacilli, A., Cappa, P., Mazzà, C., Rossi, S., 2018. How to choose and interpret 403 similarity indices to quantify the variability in gait joint kinematics. International Biomechanics 5(1), 1–8
- 404 Dul, J., Johnson, G.E., 1985. A kinematic model of the ankle joint. J Biomed Eng. 7, 137-143.
- Durkin, J.L., Dowling, J.J., 2006. Body Segment Parameter Estimation of the Human Lower Leg Using an
 Elliptical Model with Validation from DEXA. Annals of Biomedical Engineering 34, 1483–1493.
- 407 Evans, A., 2010. Paediatrics. The Pocket Podiatry Guide. Churchill Livingstone Elsevier.
- 408 Esbjörnsson, A. C., Iversen, M. D., André, M., Hagelberg, S., Schwartz, M. H., & Broström, E. W. (2015).
- 409 Effect of intraarticular corticosteroid foot injections on walking function in children with juvenile idiopathic
- 410 arthritis. Arthritis care & research, 67(12), 1693-1701.
- 411 Fick, R., 1911. Handbuch der Anatomie und Mechanik der Gelenke: III, Spezielle Gelenk-und
 412 Muskelmechanik, Gustav Fischer Verlag, Jena
- 413 Hannah, I., Montefiori, E., Modenese, L., Prinold, J., Viceconti, M. and Mazzà, C., 2017. Sensitivity of a
- 414 juvenile subject-specific musculoskeletal model of the ankle joint to the variability of operator-dependent
- 415 input. Proceedings of the Institution of Mechanical Engineers, Part H: Journal of Engineering in Medicine
- 416 231, 415-422.
- 417 Hicks, J.H., 1953. The mechanics of the foot: the joints. Journal of Anatomy 87, 345-357
- 418 Hicks, J., Uchida, T., Seth, A., Rajagopal, A. and Delp, S. L., 2015. Is my model good enough? Best
- 419 practices for verification and validation of musculoskeletal models and simulations of human movement.
- 420 Journal of Biomechanical Engineering 137(2):020905.
- 421 Inman, V.T., 1976. The Joints of the Ankle. Williams and Wilkins, Baltimore.
- Isman, R. E. and Inman, V. T., 1968. Anthropometric studies of the human foot and ankle. Bulletin of
 prosthetics research 10/11, 97-129.
- Lamberto, G., Martelli, S., Cappozzo, A., Mazzà, C., 2017. To what extent is joint and muscle mechanics predicted by musculoskeletal models sensitive to soft tissue artefacts? Journal of Biomechanics 62, 68-

- 426 76.Leitch, J., Stebbins, J., Zavatsky, A.B., 2010. Subject-specific axes of the ankle joint complex. Journal
 427 of Biomechanics 43, 2923–2928
- Leitch, J., Stebbins, J., Zavatsky, A.B., 2010. Subject-specific axes of the ankle joint complex. Journal of
 Biomechanics 43, 2923–2928
- Lewis, G.S., Cohen, T.L., Seisler, A.R., Kirby, K.A., Sheehan, F.T., Piazza, S.J., 2009. In vivo tests of an improved method for functional location of the subtalar joint axis. Journal of Biomechanics 42, 146–151.
- 432 Lundberg, A., 1989. Kinematics of the ankle and foot: in vivo roentgen stereophotogrammetry. Ph.D.
- 433 Thesis, Deptartment of Orthopaedics, Karolinska Hospital, Stolkholm, Sweden. Acta Orthopaedica
- 434 Scandinavia 60 (Suppl.), 233.
- 435 Manter, J.T., 1941. Movements of the subtalar and transverse tarsal joints. The Anatomical Record 80, 397.
- 436 Malaquias, T.M., Silveira, C., Aerts, W., De Groote, F., Dereymaeker, G., Vander Sloten, J., Jonkers, I.,
- 437 2017. Extended foot-ankle musculoskeletal models for application in movement analysis. Computer
- 438 Methods in Biomechanics and Biomedical Engineering 20:2, 153-159
- 439 Malattia, C., Damasio, M.B., Pistorio, A., Ioseliani, M., Vilca, I., Valle, M., Ruperto, N., Viola, S.,
- 440 Buoncompagni, A., Magnano, G.M., Ravelli, A., Tomà, P., Martini, A., 2011. Development and
- 441 preliminary validation of a paediatric-targeted MRI scoring system for the assessment of disease activity
- 442 and damage in juvenile idiopathic arthritis. Annals of the Rheumatic Diseases 70, 440-6.
- 443 Mattingly, B., Talwalkar, V., Tylkowski, C., Stevens, D.B., Hardy, P.A., Pienkowski, D., 2006. Three-444 dimensional in vivo motion of adult hind foot bones. Journal of Biomechanics 39 (4), 726-733.
- Modenese, L., Montefiori, E., Wang, A., Wesarg, S., Viceconti, M., Mazzà, C., 2018. Investigation of the
 dependence of joint contact forces on musculotendon parameters using a codified workflow for imagebased modelling. Journal of Biomechanics 73, 108-118.
- Nichols, T.E., Holmes, A.P., 2002. Nonparametric permutation tests for functional neuroimaging: a primer
 with examples. Human Brain Mapping 15(1), 1–25.
- 450 Nusman, C.M., Ording Muller, L.S., Hemke, R., Doria, A.S., Avenarius, D., Tzaribachev, N., et al., 2016.
- 451 Current Status of Efforts on Standardizing Magnetic Resonance Imaging of Juvenile Idiopathic Arthritis:
- 452 Report from the OMERACT MRI in JIA Working Group and Health-e-Child. The Journal of
- 453 Rheumatology 43:239-244.
- 454 Ostergaard, M., Peterfy, C., Conaghan, P., McQueen, F., Bird, P., Ejbjerg, B., et al., 2003. OMERACT
- 455 Rheumatoid Arthritis Magnetic Resonance Imaging Studies. Core set of MRI acquisitions, joint pathology
- definitions, and the OMERACT RA-MRI scoring system. The Journal of Rheumatology 30:1385-6.Parr,
- 457 W. C. H., Chatterjee, H. J. and Soligo, C., 2012. Calculating the axes of rotation for the subtalar and
- talocrural joint using 3D bone reconstructions. Journal of Biomechanics 45, 1103-1107.
- Parr, W. C. H., Chatterjee, H. J. and Soligo, C., 2012. Calculating the axes of rotation for the subtalar and
 talocrural joint using 3D bone reconstructions. Journal of Biomechanics 45, 1103-1107.
- 461 Pataky, T.C., 2012. One-dimensional statistical parametric mapping in Python. Comput. Methods Biomech.
- 462 Biomed. Eng. 15, 295–301

- Prinold, J. I., Mazzà, C., Di Marco, R., Hannah, I., Malattia, C., Magni-Manzoni, S., Petrarca, M.,
 Ronchetti, A., Tanturri de Horatio, L., van Dijkhuizen, E. H. P., Wesarg, S. and Viceconti, M., 2016. A
- 465 Patient-Specific Foot Model for the Estimate of Ankle Joint Forces in Patients with Juvenile Idiopathic
- 466 Arthritis. Annals of biomedical engineering 44, 247-257.
- 467 Rasmussen, O., Tovborg-Jensen, I., 1982. Mobility of the ankle joint: recording of rotator movements in
- the talocrural joint in vitro with and without the lateral collateral ligaments of the ankle. Acta Orthop Scand53, 155–60.
- 470 Ravelli A. and Martini A., 2007. Juvenile idiopathic arthritis. Lancet 369(9563), 767-778.
- 471 Roaas, A., Anderson, G.B., 1982. Normal range of motion of the hip, knee and ankle joints in male subjects,
- 472 30-40 years of age. Acta Orthop Scand 53(2), 205-8.
- 473 Roach, K.E., Wang, B., Kapron, A.L., Fiorentino, N.M., Saltzman, C.L., Foreman, K.B., Anderson, A.E.
- 474 In Vivo Kinematics of the Tibiotalar and Subtalar Joints in Asymptomatic Subjects: A High-Speed Dual
- 475 Fluoroscopy Study. ASME. J Biomech Eng. 2016;138(9):091006-091006-9.
- 476 Saraswat, P., Andersen, M.S., MacWilliams, B.A., 2010. A musculoskeletal foot model for clinical gait
 477 analysis. Journal of Biomechanics 43 (9), 1645-1652.
- 478 Scheys, L., Loeckx, D., Spaepen, A., Suetens, P. and Jonkers, I., 2009. Atlas-based non-rigid image
- registration to automatically define line-of-action muscle models: A validation study. Journal ofBiomechanics 42, 565-572.
- Scott, S.H., Winter, D.A., 1991. Talocrural and talocalcaneal joint kinematics and kinetics during the stance
 phase of walking. J. Biomech. 24, 743–752.
- Sepic, S. B., Murray, M. P., Mollinger, L. A., Spurr, G. B., & Gardner, G. M. 1986. Strength and range of
 motion in the ankle in two age groups of men and women. American Journal of Physical Medicine 65, 7584.
- 486 Siegler, S., Chen, J., Schneck, C.D., 1988. The three-dimensional kinematics and flexibility characteristics
 487 of the human ankle and subtalar joints. Part 1: kinematics. Journal of Biomechanical Engineering 110, 364–
 488 373.
- 489 Siegler, S., Toy, J., Seale, D., & Pedowitz, D. (2014). The clinical biomechanics award 2013 presented
- 490 by the international society of biomechanics: New observations on the morphology of the talar dome and 491 its relationship to ankle kinematics. Clinical Biomechanics, 29(1), 1-6.
- 492 Stebbins, J., Harrington, M., Thompson, N., Zavatsky, A. and Theologis, T., 2006. Repeatability of a model
 493 for measuring multi-segment foot kinematics in children. Gait & Posture 23, 401-410.
- 494 Steger, S., Kirschner, M. and Wesarg, S., 2012. Articulated atlas for segmentation of the skeleton from
 495 head & neck CT datasets. 9th IEEE International Symposium on Biomedical Imaging (ISBI). Barcelona,
 496 Spain.
- 497 Valente, G., Crimi, G., Vanella, N., Schileo, E. and Taddei, F., 2017. nmsBuilder: Freeware to create
- 498 subject-specific musculoskeletal models for OpenSim. Computer Methods and Programs in Biomedicine499 152, 85-92.

- van den Bogert, A.J., Smith, G.D., Nigg, B.M., 1994. In vivo determination of the anatomical axes of the
 ankle joint complex: an optimization approach. Journal of Biomechanics 27, 1477–1488
- van Langelaan, E.J., 1983. A kinematical analysis of the tarsal joints. An X-ray photogram-metric study.
 Ph.D. Thesis, Acta Orthopaedica Scandinavia 54 (Suppl.), 204.

504ViconMotionSystems,L.BiomechanicalResearch,2012.http://www.irc-505web.co.jp/vicon_web/news_bn/PIGManualver1.pdf

- 506
- 507 Westblad, P., Hashimoto, T., Winson, I., Lundberg, A., Arndt, A., 2002. Differences in ankle-joint complex
- 508 motion during the stance phase of walking as measured by superficial and bone-anchored markers. Foot 509 Ankle Int. 23 (9), 856–863.
- 510 Woltring, H.J., 1986. A FORTAN package for generalized cross-validatory spline smoothing and
- 511 differentiation. Advances in Engineering Software 8(2), 104-113.