

### VIRTUAL ISSUE EDITORIAL

## Improving crop performance under drought – cross-fertilization of disciplines

Better crop performance in dry environments is imperative for food security in the face of climate change. This has never been as true as in 2017, but the concern has existed for decades. The four InterDrought conferences held since 1995 have addressed issues associated with crop performance under drought with a clear multi-disciplinary approach. During this time Journal of Experimental Botany has been at the forefront in publishing the underlying experimental science encompassing the disciplines and scales of organization required in drought research. We hope that the papers highlighted here will be useful to, and instrumental for, broadening interdisciplinary understanding of drought tolerance.

One of the most productive ways of tackling the agricultural challenge of drought is through cross-fertilization between areas of research, in particular crop physiology, agronomy, genetics, breeding, and environmental characterization/modelling. Scientists need to become familiar with multi-scale approaches from cells to crops subjected to water deficit, and this has been a major achievement of the InterDrought network (Box 1). Indeed, drought tolerance involves cellular aspects such as detoxification (Missihoun et al., 2011) and osmotic adjustment (Blum, 2016), but also whole-plant signalling involved in the control of growth and transpiration under water deficit (Tardieu, 2016), the whole-plant control of shoot and root system architectures, and feedbacks between water capture, growth and transpiration at canopy level (Messina et al., 2015). The exploitation of native genetic variability provides invaluable opportunities for improving plant performance based on mechanisms at any of these scales, namely cell, organ, whole-plant and canopy, in particular through progress in phenotyping (Fiorani and Schurr, 2013; see also the special issue 'Phenotyping in plants', introduced by Pieruschka and Lawson, 2015) and genomicsassisted breeding (Reynolds and Langridge, 2016).

Improved knowledge of the physiological mechanisms involved in the control of transpiration and growth, and of their genetic make-up, paves the way for manipulating and eventually fine-tuning these controls in order to enhance their efficiency via genetic approaches (Habben *et al.*, 2014), possibly complemented by application of compounds that affect them

(Park et al., 2015). While agronomy is currently undergoing a major change in focus based on the widespread use of sensors, robots and imaging techniques resulting in precision agriculture, breeders are taking full advantage of our increasing ability to identify and tailor beneficial alleles able to enhance crop productivity and eventually mitigate the negative effects of drought.

## Rapid progress in molecular physiology and genomics applied to drought

Identification of the sources of tolerance and then cross-hybridization to recombine genomic segments form the basis of classical breeding for the development of drought-tolerant cultivars (Rauf *et al.*, 2016). However, the pace of development of improved drought-tolerant cultivars is slow, severely hampering the timely replacement of improved varieties for agriculture.

The controlling mechanisms for plant performance under drought are complex due to the multifaceted interplay between genetic components including genes, transcription factors, microRNAs (miRNAs), hormones, proteins, co-factors, ions and metabolites (reviewed by Janiak et al., 2016; Tardieu, 2016). Advances in cost-effective sequencing and high-throughput genotyping technologies now mean that sequencing/genotyping large amounts of genetic material can be achieved within a limited period of time. These sequencing and genotyping data together with information from multi-environment phenotyping have enabled high-resolution genetic mapping leading to genetic dissection of improved yield under drought. Quantitative trait loci (QTLs) and candidate genes have been identified in various crops including rice (Zhou et al., 2010), wheat (Maccaferri et al., 2016), maize (Millet et al., 2016) and chickpea (Jaganathan et al., 2015; Kale et al., 2015). Although gene editing has shown great potential as a powerful tool for improving any trait for which sequence variation is available (Bortesi et al., 2015), this technology has yet to contribute to an appreciable improvement in drought tolerance in crop plants.

In addition to genetic studies conducted at the DNA level, efforts have also been made in candidate gene discovery through RNA deep sequencing (Chang et al., 2016; Garg et al., 2016; Shankar et al., 2016) and microarray analysis

### Box 1. InterDrought: 25 years of progress

The link between crop performance and drought is now deeply embedded (Lobell et al., 2008; Tester and Langridge, 2010) and highlighted in the last IPCC report (Field et al., 2014). However, this wider appreciation by policymakers was being highlighted by scientists much earlier. InterDrought was created in 1992 with EEC funding, and at that time included European teams of researchers covering molecular biology, physiology, genetics and breeding. This quickly led on to the first international meeting for the network in Montpellier, in 1995 (Belhassen, 1997). Although the second InterDrought congress in 2005 was also in Europe (Rome) as the scope expanded so the meetings extended in 2009 to Shanghai, in 2013 to Perth and, in 2017, to Hyderabad.

Writing in the first contribution to a publication arising from InterDrought, Passioura (1997) noted that 'We can all tell that a cactus is more drought tolerant than a carnation. But when we look at crop plants, the features that confer drought tolerance are far from clear ... the traits we associate with xerophytes typically concern survival during drought, whereas with crops we are concerned with production – and insofaras the term "drought tolerance" has any useful meaning in an agricultural context, it must be defined in terms of yield in relation to a limiting water supply.' These words have been a clear 'marker' of InterDrought across its five congresses. We believe that the success of these meetings has enabled scientists from quite different disciplines to interact and acquire a broader view of the adaptive response of crops to such water scarcity. Image: Chickpea field trial in Patancheru, India, courtesy of Rajeev K. Varshney (Credit: L.Vidyasagar, ICRISAT).



(Mishra et al., 2016). For example, whole-genome transcriptome-profiling studies have identified a large number of transcripts encoding members of various gene families reported to play an important role in abiotic stress tolerance, including AP2/EREBP, bLHL, MYB, and auxin-related families (Chang et al., 2016; Garg et al., 2016). Similarly, transcriptome analysis has identified the molecular mechanism underlying the high degree of plasticity of the water-deficit response in maize (Opitz et al., 2016). Furthermore, induced expression of various transcription factors (e.g. MYB) have favourable effects under drought in maize (Casaretto et al., 2016) and Arabidopsis (Scarpeci et al., 2016).

Various studies have reported a role of miRNAs in abiotic stress tolerance (Shriram et al., 2016), as in the case of drought-induced expression of Hv-miR827 in barley (Ferdous et al.,

2016). Likewise, up- or down-regulation of different miRNAs was found to be associated with improved performance in rice (Chung et al., 2016) and barley (Hackenberg et al., 2015) subjected to water deficit. Hormones also play an important role in the regulation of drought acclimation/adaptation (Li et al., 2016 Tardieu, 2016; Visentin et al., 2016). For instance, recent studies suggest a role for cytokinins in barley (Pospíšilová et al., 2016), Arabidopsis (Nguyen et al., 2016), tomato (Farber et al., 2016) and rice (Talla et al., 2016), and for strigolactones in tomato (Visentin et al., 2016). These mechanisms have a crucial role for phenotypic plasticity. Indeed, the latter is a key trait for dealing with complex G×E interactions, as shown by Sadras et al. (2016), who explored the genetic control of phenotypic plasticity in chickpea. A systems-based approach which allows the integration of 'omic' technologies

using computer-assisted theoretical and molecular biology would therefore help capture a global view of the complex mechanisms involved in the phenotypic plasticity associated with drought responses (Hussain *et al.*, 2016).

# Exploitation of genetic resources based on association mapping and genomic selection

Root features play a pivotal role in crop performance under water deficit, as well as in optimizing use of the available water resources (Messina *et al.*, 2015). In the past, roots have received limited experimental attention due to difficulties in phenotyping, particularly under field conditions. However, recent technical advances in root phenotyping and the utilization of high-throughput platforms have led to the publication of an impressive number of papers.

Particular attention has been devoted to the characterization of root mutants and the dissection of the genetic makeup governing root system architecture (RSA) and its effects on crop performance under different water regimes. The work of Jiang et al. (2016) highlights the role of strigolactones in the hormonal landscape that shapes RSA through the modulation of lateral root development via a tight interplay with auxins and cytokinins. Additional physiological work in Arabidopsis by Kircher and Schopfe (2016) supports the existence of periodic priming signals influencing lateral root formation along the growing root (see also the Insight article by Scheres and Laskowski, 2016). A valuable example of how to leverage molecular knowledge on lateral root growth to enhance the field performance of a drought-stressed crop is presented by Li et al. (2016). In rice, overexpression of transcription factor gene MORE ROOT (TaMOR) from wheat results in more roots and higher grain yield. TaMOR, a plant-specific transcription factor belonging to the ASYMMETRIC LEAVES2/ LATERAL ORGAN BOUNDARIES (AS2/LOB) protein family, is highly conserved in wheat and its wild relatives. Notably, ITaMOR-D-overexpressing lines had larger root systems in Arabidopsis and rice, and produced a higher grain yield per plant. Therefore, TaMOR offers an opportunity to improve root architecture and increase yield in crops.

Among crops, cereals have a particularly complex and plastic root system whose components play different adaptive roles according to the growth stage and prevailing soil conditions (Hochholdinger, 2016). In maize, the QTL study of Gao and Lynch (2016) shows that a reduced crown root number is associated with greater root depth and improved water acquisition from drying soil. Previous studies have shown that major QTLs for RSA influence yield in maize grown under different water regimes (Landi *et al.*, 2007, 2010). Similar results have also been reported in rice (Price *et al.*, 2002; Uga *et al.*, 2011) and chickpea (Varshney *et al.*, 2013).

From an agronomic standpoint, the work of Lilley and Kirkegaard (2016) shows the importance of the time–space interplay between root depth and water capture as related to soil depth and annual resetting of soil water. This modelling study shows that capturing more water from deeper soil layers

is not always the best option. Additionally, the simulation shows a greater impact of earlier sowing than modified root systems on water uptake, indicating that crop sequence must be managed tactically to optimize overall system benefits.

### 'Whole-plant mechanisms' which affect yield may differ between environments

Causal relationships between potential mechanisms and plant behaviour under drought are often far from straightforward or unidirectional due to multiple feedbacks at different timescales (Tardieu and Parent, 2017). For instance, it is well-known that early-flowering genotypes tend to escape drought compared with later genotypes, because flowering time and physiological maturity occur earlier in the season when the soil water reserve is not depleted. This is at the expense of potential biomass accumulation because of a shorter period in which photosynthesis can occur over the plant's life cyle. Kazan and Lyons (2016) show that evolution may well have resulted in elaborate mechanisms in Arabidopsis plants subjected to water deficit which finetune the escape strategy and avoid its negative trade-offs. In particular earliness is controlled differentially in the case of drought in late or early-maturing accessions via differential expression of genes involved in the floral transition. This results in interesting feedback loops between floral transition, water uptake and growth. Another interesting example arises from the study of Christopher et al. (2016) (see also the Insight article by Rebetzke et al., 2016b). Stay-green is often considered as a trait per se that confers drought tolerance. Christopher et al. considered the relationship between yield and different stay-green traits in eight contrasting environments. They found marked differences in relationships in well-watered conditions, and in water deficit occurring during either flowering or grain filling. Hence, indicators of staygreen have an effect on yield that is dependent on context. This may well be a general case for most traits involved in drought tolerance (Tardieu, 2012).

Similarly, there has been a long-standing debate about the effects of awns on yield, particularly under water deficit. Rebetzke *et al.* (2016a) propose a Solomon-like judgment: the presence or absence of awns has opposite effects on grain number and grain size, resulting in no major effect on yield (see also the Insight article by Guo and Schnurbusch, 2016). Indeed, the allocation of assimilates to awns decreases floret fertility, but favours grain filling particularly in dry environments. However, the respective effects on grain number and grain yield may have amplitudes that are context-dependent.

Hence, Blum (2016) states that drought tolerance needs to be re-defined in a better way by distinguishing the environment as sensed by plants (stress), the early plant reactions (strain) that trigger signalling processes and, eventually, acclimation processes. It is noteworthy that evolutionary geneticists usually reserve the term 'adaptation' for the selection of these acclimation processes over generations. As stated by Maron *et al.* (2016), fine-tuning concepts has large consequences for annotating so-called stress-resistance genes with

#### Box 2. Journal of Experimental Botany and research on environmental change

Journal of Experimental Botany (JXB) has had a long association with research on global environmental change, including drought and food security, not least through the influence of Bill Davies, Editor in Chief from 1995 to 2007 (see Dodd *et al.*, 2015). The JXB publications which followed on from the InterDrought conferences over the past decade chart our developing understanding of crop responses: InterDrought II (Araus *et al.*, 2007); InterDrought III (Boyer, 2010); and InterDrought IV (Tuberosa *et al.*, 2014).

A number of related special issues of *JXB* have also recently been produced: 'Roots to global food security' (Dodd *et al.*, 2015); 'Plant roots: new challenges in a changing world' (Price, 2016); 'From inspiration to impact: delivering value from global root research' (Rebetzke, 2016), with links made between research, breeding and environmental challenges (e.g. see Reyes *et al.*, 2015); and 'Breeding plants to cope with future climate change' (Halford and Foyer, 2015), which includes consideration of food security planning (Halford and Foyer, 2015; McKersie, 2015). Image: Pixabay, CC0 Public Domain.



markedly different effects in time and scales of organization (Tardieu and Parent, 2017). The latter considerations may considerably complicate early screening for drought tolerance based on root and shoot (Avramova *et al.*, 2016).

### **Acknowledgements**

We sincerely thank *Journal of Experimental Botany* (Box 2) for continued support of drought studies over the long period of time covered by the InterDrought conferences.

Key words: Climate change, crop productivity, drought tolerance, food security, water deficit.

### Francois Tardieu<sup>1</sup>, Rajeev K. Varshney<sup>2</sup> and Roberto Tuberosa<sup>3</sup>

<sup>1</sup> INRA LEPSE, 2 Place Viala 34060, Montpellier, France
<sup>2</sup> Research Programme – Genetic Gains, International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), Patancheru-502 324, India

<sup>3</sup> Department of Agricultural Sciences, Viale Fanin 44, 40127 Bologna, Italy

Correspondence: francois.tardieu@inra.fr; r.k.varshney@cgiar.org; roberto.tuberosa@unibo.it

### References

**Araus JL, Ferrio JP, Buxó R, Voltas J.** 2007. Preface (Integrated approaches to sustain and improve plant production under drought stress). Journal of Experimental Botany **58,** 131–145.

**Avramova V, Nagel KA, AbdElgawad H, Bustos D, DuPlessis M, Fiorani F, Beemster GT.** 2016. Screening for drought tolerance of maize hybrids by multi-scale analysis of root and shoot traits at the seedling stage. Journal of Experimental Botany **67,** 2453–2466.

**Belhassen E.** 1997. Drought tolerance in higher plants: genetic, physiological and molecular biological analysis. Springer.

**Blum A.** 2016. Stress, strain, signaling, and adaptation–not just a matter of definition. Journal of Experimental Botany **67,** 562–565.

**Bortesi L, Fischer R.** 2015. The CRISPR/Cas9 system for plant genome editing and beyond. Biotechnology Advances **33**, 41–52.

**Boyer JS.** 2010. Drought decision-making. Journal of Experimental Botany **61**, 3493–3497.

Casaretto JA, El-Kereamy A, Zeng B, Stiegelmeyer SM, Chen X, Bi YM, Rothstein SJ. 2016. Expression of OsMYB55 in maize activates stress-responsive genes and enhances heat and drought tolerance. BMC Genomics 17, 312.

Chang YL, Li WY, Miao H, Yang SQ, Li R, Wang X, Li WQ, Chen KM. 2016. Comprehensive genomic analysis and expression profiling of the NOX gene families under abiotic stresses and hormones in plants. Genome Biology and Evolution 8, 791–810.

**Christopher JT, Christopher MJ, Borrell AK, Fletcher S, Chenu K.** 2016. Stay-green traits to improve wheat adaptation in well-watered and water-limited environments. Journal of Experimental Botany **67,** 5159–5172.

**Chung PJ, Jung H, Jeong DH, Ha SH, Choi YD, Kim JK.** 2016. Transcriptome profiling of drought responsive noncoding RNAs and their target genes in rice. BMC Genomics **17,** 563.

- **Dodd I, Griffiths H, Sharp R, Traynor M, Zhang J.** 2015. Preface (Roots to global food security). Journal of Experimental Botany **66**, 6351–6362.
- **Farber M, Attia Z, Weiss D.** 2016. Cytokinin activity increases stomatal density and transpiration rate in tomato. Journal of Experimental Botany **67,** 6351–6362.
- **Ferdous J, Whitford R, Nguyen M, Brien C, Langridge P, Tricker PJ.** 2016. Drought-inducible expression of *Hv*-miR827 enhances drought tolerance in transgenic barley. Functional & Integrative Genomics. doi:10.1007/s10142-016-0526-8.
- Field CB, Barros VR, Mastrandrea MD, et al. 2014. Climate change 2014. Cambridge University Press.
- **Fiorani F, Schurr U.** 2013. Future scenarios for plant phenotyping. Annual Review of Plant Biology **64,** 267–291.
- **Gao Y, Lynch JP.** 2016. Reduced crown root number improves water acquisition under water deficit stress in maize (*Zea mays* L.). Journal of Experimental Botany **67**, 4545–4557.
- **Garg R, Shankar R, Thakkar B, Kudapa H, Krishnamurthy L, Mantri N, Varshney RK, Bhatia S, Jain M.** 2016. Transcriptome analyses reveal genotype-and developmental stage-specific molecular responses to drought and salinity stresses in chickpea. Scientific Reports **6,** 19228.
- **Guo Z, Schnurbusch T.** 2016. Costs and benefits of awns. Journal of Experimental Botany **67,** 2533–2535.
- **Habben JE, Bao X, Bate NJ, et al.** 2014. Transgenic alteration of ethylene biosynthesis increases grain yield in maize under field drought-stress conditions. Plant Biotechnology Journal **12,** 685–693.
- **Hackenberg M, Gustafson P, Langridge P, Shi BJ.** 2015. Differential expression of microRNAs and other small RNAs in barley between water and drought conditions. Plant Biotechnology Journal **13**, 2–13.
- **Halford NG, Foyer C.** 2015. Preface (Breeding plants to cope with future climate change): Producing a road map that enables plants to cope with future climate change. Journal of Experimental Botany **66**, 2433–3434.
- **Hochholdinger F.** 2016. Untapping root system architecture for crop improvement. Journal of Experimental Botany **67**, 4431–4433.
- **Hussain SS, Ahsan MA, Sornaraj P, Ali M, Shi BJ.** 2016. Toward integration of a systems-based approach for understanding drought stress in plants. In: Parvaiz A, ed. Water stress and crop plants: a sustainable approach. John Wiley & Sons, 227–247.
- **Jaganathan D, Thudi M, Kale S, et al.** 2015. Genotyping-by-sequencing based intra-specific genetic map refines a "*QTL-hotspot*" region for drought tolerance in chickpea. Molecular Genetics and Genomics: MGG **290**, 559–571.
- **Janiak A, Kwaśniewski M**, Szarejko I. 2016. Gene expression regulation in roots under drought. Journal of Experimental Botany **67**, 1003–1014.
- **Jiang L, Matthys C, Marquez-Garcia B, et al.** 2016. Strigolactones spatially influence lateral root development through the cytokinin signaling network. Journal of Experimental Botany **67,** 379–389.
- **Kale SM, Jaganathan D, Ruperao P, et al.** 2015. Prioritization of candidate genes in "QTL-hotspot" region for drought tolerance in chickpea (*Cicer arietinum* L.). Scientific Reports **5,** 15296.
- **Kazan K, Lyons R.** 2016. The link between flowering time and stress tolerance. Journal of Experimental Botany **67,** 47–60.
- **Kircher S, Schopfer P.** 2016. Priming and positioning of lateral roots in Arabidopsis. An approach for an integrating concept. Journal of Experimental Botany **67,** 1411–1420.
- **Landi P, Giuliani S, Salvi S, Ferri M, Tuberosa R, Sanguineti MC.** 2010. Characterization of *root-yield-*1.06, a major constitutive QTL for root and agronomic traits in maize across water regimes. Journal of Experimental Botany **61,** 3553–3562.
- Landi P, Sanguineti MC, Liu C, Li Y, Wang TY, Giuliani S, Bellotti M, Salvi S, Tuberosa R. 2007. *Root-ABA1* QTL affects root lodging, grain yield, and other agronomic traits in maize grown under well-watered and water-stressed conditions. Journal of Experimental Botany **58**, 319–326.
- **Li W, Herrera-Estrella L, Tran LS.** 2016. The Yin-Yang of cytokinin homeostasis and drought acclimation/adaptation. Trends in Plant Science **21,** 548–550.
- **Lilley JM, Kirkegaard JA.** 2016. Farming system context drives the value of deep wheat roots in semi-arid environments. Journal of Experimental Botany **67,** 3665–3681.

- **Lobell DB, Burke MB, Tebaldi C, Mastrandrea MD, Falcon WP, Naylor RL.** 2008. Prioritizing climate change adaptation needs for food security in 2030. Science **319**, 607–610.
- Maccaferri M, El-Feki W, Nazemi G, Salvi S, Canè MA, Colalongo MC, Stefanelli S, Tuberosa R. 2016. Prioritizing quantitative trait loci for root system architecture in tetraploid wheat. Journal of Experimental Botany 67, 1161–1178.
- Maron LG, Piñeros MA, Kochian LV, McCouch SR. 2016. Redefining 'stress resistance genes', and why it matters. Journal of Experimental Botany **67**, 5588–5591.
- **McKersie B.** 2015. Planning for food security in a changing climate. Journal of Experimental Botany **66**, 3435–3450.
- Messina CD, Sinclair TR, Hammer GL, Curan D, Thompson J, Oler Z, Gho C, Cooper M. 2015. Limited-transpiration trait may increase maize drought tolerance in the US corn belt. Agronomy Journal 107.
- **Millet E, Welcker C, Kruijer W, et al.** 2016. Genome-wide analysis of yield in Europe: allelic effects as functions of drought and heat scenarios. Plant Physiology **172**, 749–764.
- **Mishra U, Rai A, Kumar R, Singh M, Pandey HP.** 2016. Gene expression analysis of *Solanum lycopersicum* and *Solanum habrochaites* under drought conditions. Genomics Data **9,** 40–41.
- **Missihoun TD, Schmitz J, Klug R, Kirch HH, Bartels D.** 2011. Betaine aldehyde dehydrogenase genes from Arabidopsis with different subcellular localization affect stress responses. Planta **233,** 369–382.
- **Nguyen KH, Van Ha C, Nishiyama R, et al.** 2016. Arabidopsis type B cytokinin response regulators ARR1, ARR10, and ARR12 negatively regulate plant responses to drought. Proceedings of the National Academy of Sciences, United States of America **13,** 3090–3095.
- **Opitz N, Marcon C, Paschold A, Malik WA, Lithio A, Brandt R, Piepho HP, Nettleton D, Hochholdinger F.** 2016. Extensive tissue-specific transcriptomic plasticity in maize primary roots upon water deficit. Journal of Experimental Botany **67,** 1095–1107.
- Park SY, Peterson FC, Mosquna A, Yao J, Volkman BF, Cutler SR. 2015. Agrochemical control of plant water use using engineered abscisic acid receptors. Nature **520**, 545–548.
- **Passioura JB.** 1997. Drought and drought tolerance. In: Belhassen E, ed. Drought tolerance in higher plants: genetic, physiological and molecular biological analysis. Springer, 1–5.
- **Pieruschka R, Lawson T.** 2015. Phenotyping in Plants. Preface. Journal of Experimental Botany **66**, 5385–5387.
- **Pospíšilová H, Jiskrová E, Vojta P, et al.** 2016. Transgenic barley overexpressing a cytokinin dehydrogenase gene shows greater tolerance to drought stress. New Biotechnology **33**, 692–705.
- **Price AH.** 2016. Preface (Plant roots: new challenges in a changing world). Journal of Experimental Botany **67,** 991–993.
- **Price AH, Cairns JE, Horton P, Jones HG, Griffiths H.** 2002. Linking drought-resistance mechanisms to drought avoidance in upland rice using a QTL approach: progress and new opportunities to integrate stomatal and mesophyll responses. Journal of Experimental Botany **53**, 989–1004.
- Rauf S, Al-Khayri JM, Zaharieva M, Monneveux P, Khalil F. 2016. Breeding strategies to enhance drought tolerance in crops. In: Al-Khayri JM, Jain SM, Johnson DV, eds. Advances in plant breeding strategies: agronomic, abiotic and biotic stress traits. Springer International Publishing. 397–445.
- **Rebetzke G.** 2016. Preface (From inspiration to impact: delivering value from global root research). Journal of Experimental Botany **67**, 3601–3603.
- **Rebetzke GJ, Bonnett DG, Reynolds MP.** 2016a. Awns reduce grain number to increase grain size and harvestable yield in irrigated and rainfed spring wheat. Journal of Experimental Botany **67,** 2573–2586.
- **Rebetzke GJ, Jimenez-Berni JA, Bovill WD, Deery DM, James RA.** 2016b. High-throughput phenotyping technologies allow accurate selection of stay-green. Journal of Experimental Botany **67,** 4919–4924.
- Reyes A, Messina CD, Hammer GL, Liu L, van Oosterom E, Lafitte R, Cooper M. 2015. Soil water capture trends over 50 years of single-cross maize (*Zea mays* L.) breeding in the US corn-belt. Journal of Experimental Botany **66**, 7339–7346.
- **Reynolds M, Langridge P.** 2016. Physiological breeding. Current Opinion in Plant Biology **31,** 162–171.

Sadras VO, Lake L, Li Y, Farquharson EA, Sutton T. 2016. Phenotypic plasticity and its genetic regulation for yield, nitrogen fixation and  $\delta 13C$  in chickpea crops under varying water regimes. Journal of Experimental Botany **67**, 4339–4351.

Scarpeci TE, Frea VS, Zanor MI, Valle EM. 2016. Overexpression of AtERF019 delays plant growth and senescence and improves drought tolerance in Arabidopsis. Journal of Experimental Botany doi: https://doi.org/10.1093/jxb/erw429

**Scheres B, Laskowski M.** 2016. Root patterning: it takes two to tangle. Journal of Experimental Botany **67**, 1201–1203.

**Shankar R, Bhattacharjee A, Jain M.** 2016. Transcriptome analysis in different rice cultivars provides novel insights into desiccation and salinity stress responses. Scientific Reports **6,** 23719.

**Shriram V, Kumar V, Devarumath RM, Khare TS, Wani SH.** 2016. MicroRNAs as potential targets for abiotic stress tolerance in plants. Frontiers in Plant Science **7,** 817.

**Talla SK, Panigrahy M, Kappara S, Nirosha P, Neelamraju S, Ramanan R.** 2016. Cytokinin delays dark-induced senescence in rice by maintaining the chlorophyll cycle and photosynthetic complexes. Journal of Experimental Botany **67,** 1839–1851.

**Tardieu F.** 2012. Any trait or trait-related allele can confer drought tolerance: just design the right drought scenario. Journal of Experimental Botany **63**, 25–31.

**Tardieu F.** 2016. Too many partners in root-shoot signals. Does hydraulics qualify as the only signal that feeds back over time for reliable stomatal control? The New Phytologist **212**, 802–804.

**Tardieu F, Parent B.** 2017. Predictable 'meta-mechanisms' emerge from feedbacks between transpiration and plant growth and cannot be simply deduced from short-term mechanisms. Plant, Cell & Environment doi: 10.1111/pce.12822

**Tester M, Langridge P.** 2010. Breeding technologies to increase crop production in a changing world. Science **327**, 818–822.

**Tuberosa R, Turner NC, Cakir M.** 2014. Preface (Two decades of InterDrought conferences: are we bridging the genotype-to-phenotype gap?) Journal of Experimental Botany **65**, 6137–6139.

**Uga Y, Okuno K, Yano M.** 2011. *Dro1*, a major QTL involved in deep rooting of rice under upland field conditions. Journal of Experimental Botany **62**, 2485–2494.

**Varshney RK, Gaur PM, Chamarthi SK, et al.** 2013. Fast-track introgression of "QTL-hotspot" for root traits and other drought tolerance traits in JG 11, an elite and leading variety of chickpea. The Plant Genome **6,** 3.

**Visentin I, Vitali M, Ferrero M, et al.** 2016. Low levels of strigolactones in roots as a component of the systemic signal of drought stress in tomato. The New Phytologist **212,** 954–963.

**Zhou L, Liu Y, Liu Z, Kong D, Duan M, Luo L.** 2010. Genome-wide identification and analysis of drought-responsive microRNAs in *Oryza sativa*. Journal of Experimental Botany **61,** 4157–4168.