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Superovulation protocols for dairy cows bred with SexedULTRA™ sex-sorted semen

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13	Superovulation protocols for dairy cows bred with SexedULTRA TM sex-sorted semen
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Summary

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The objective was to compare embryo yield and quality in lactating dairy cows superovulated (SO) 27 28 with varying amounts of gonadotropins and FSH:LH ratios and inseminated with SexedULTRATM 29 sex-sorted semen. The SO treatments (n=77) involved 3 protocols: groups F700 and F1000 were 30 given total doses of 700 and 1000 IU of Folltropin (FSH:LH ratio 49:1), respectively, whereas 31 group F700P300 was given 700 IU of Folltropin + 300 IU of Pluset (FSH:LH ratio 1:1). Cows were 32 artificially inseminated 3 times over a 10-h interval with frozen-thawed SexedULTRATM sex-sorted semen (total of 10 x 10 sex-sorted sperm), starting 18 h after onset of estrus, with embryos/ova 33 34 recovered 7 d after estrus. Total number of recovered structures and transferable embryos were 35 lower (P<0.05) in F700 (4.7 \pm 3.0 and 1.9 \pm 1.7, respectively; mean \pm SD) compared to F1000 (8.1 36 \pm 3.8 and 4.4 \pm 2.6) and F700P300 (8.5 \pm 6.4 and 4.5 \pm 3.3). Percentage of cows ovulating >50% of 37 follicles \geq 0.8 cm in diameter was lower (P<0.05) in F700 (35.5%) than in F1000 (82.4%) and 38 F700P300 (73.1%). Percentage of unfertilized oocytes was higher (P<0.05) in F700 (45.0 vs 27.7%) 39 for F1000 and 29.0% for F700P300) whereas percentage of morulae was higher (P<0.05) in F1000 40 (19.3 vs 8.7% for F700 and 12.2% for F700P300). Embryo quality was similar among groups 41 (P>0.05). In conclusion, embryo production in lactating dairy cows was improved by increasing 42 total dose of gonadotropins from 700 to 1000 IU, with SexedULTRATM sex-sorted semen yielding 43 satisfactory fertilization rates and embryo quality. 44

Keywords: superovulation, gonadotropins, sexedUltra, sex-sorted semen, dairy cow

1. Introduction

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Numerous superovulation (SO) protocols have been intensively studied to improve embryo 47 48 production and embryo quality in cattle. As many donors produce no, few or poor-quality embryos, 49 various FSH treatments have been attempted, including varying total doses of gonadotropins and 50 the FSH:LH ratio. Although both FSH and LH are required in physiologic reproductive processes, 51 limiting exogenous LH in SO regimens in cattle has been advocated (Kanitz et al. 2002, Mapletoft 52 et al. 2002), with benefits including reduced variability in the SO response (Moor et al 1984), 53 enhanced embryo production due to higher ovulation rates and improvements in fertilization rate 54 and embryo quality (Donaldson et al. 1987, Yamamoto et al. 1993, Quaresma et al. 2003). Excess 55 LH seemed to have deleterious effects due to premature oocyte activation (Hyttel et al. 1991). 56 premature ovulation (Callesen et al. 1987) and luteinization of FSH-stimulated follicles (Boland et 57 al. 1991). In many studies, increasing LH content in the SO regimen decreased proportion of 58 transferable embryos; despite a higher ovulation rate, a large proportion of ova/embryos were 59 unfertilized or degenerate (Donaldson and Ward 1986, Donaldson et al. 1986, Kelly et al. 1997). 60 Treatment with a purified FSH preparation resulted in greater embryo production than treatment 61 with equine chorionic gonadotropin (eCG), which has high LH activity (Goulding et al. 1991). 62 Notwithstanding the deleterious effects of excessive exogenous LH, inadequate exogenous LH 63 reduced embryo yield (Chupin et al. 1984), as highly purified FSH preparations significantly 64 reduced ovulation rates compared to FSH supplemented with LH (Chupin et al. 1987, Ereno et al. 1988, Schmidt et al. 1988, Herrler 1991). Impaired follicular maturation in heifers superovulated 65 66 with a recombinant human FSH preparation (Takagi et al. 2001) was attributed to a lack of 67 exogenous LH activity and severe suppression of LH pulsatility. 68 The role of LH in SO protocols is controversial; outcomes could depend on many factors, including 69 subspecies, breed or genetic differences, general health condition and energy balance, acute or 70 chronic stress and various management or environmental conditions. It is well established that SO regimens alter LH secretion, including reductions in pulse amplitude and frequency, a reduced basal 71

12	secretion of Lift and an affered (absent, ininioned, premature of face) preovulatory Lift surge which
73	reduces ovulation rate, fertilization rates, egg/embryo quality and embryo production (Greve et al.
74	1984, Roberge et al. 1995, Price et al. 1999, Gosselin et al. 2000). In 1 study (Ben Jabara et al.
75	1994), lesser amounts of exogenous LH administered throughout the entire SO protocol resulted in
76	greatest suppression of endogenous LH. However, providing more exogenous LH, only near the end
77	of the SO regimen, increased transferable embryos (Barcelos et al. 2006, Cifuentes et al. 2009).
78	Fertilization rates with sexed semen have always been severely reduced, particularly in
79	superovulated cows versus single-ovulating females or superovulated heifers. Reduced fertilization
80	with sexed semen may be due to low doses of sperm, an abnormal uterine environment or atypical
81	sperm transport in superovulated cows, damage to sperm during sex sorting, or some combination.
82	Superovulated cows inseminated with sex-sorted semen produced a significantly smaller proportion
83	of transferable embryos and significantly larger proportions of unfertilized oocytes and/or
84	degenerate embryos than heifers or cows inseminated with unsorted semen (Peippo et al 2009,
85	Monteiro et al 2016, Mikkola et al 2017). With 4M SexedULTRA™ sex-sorted semen, fertility rates
86	between conventional semen and sex-sorted semen in single-ovulating heifers approached
87	equivalence (Vishwanath and Moreno 2018); however, no data have been published regarding
88	superovulated cattle. Although the economic impact of using sexed semen in an embryo transfer
89	program was deemed profitable only in heifers (Mikkola et al 2017, Hayakawa et al 2009), 4M
90	SexedULTRA™ sex-sorted semen may be appropriate in cows.
91	The objective was to determine whether embryo yield in stressed lactating dairy cows would be
92	altered by changing the FSH:LH ratio and inseminating with SexedULTRATM sex-sorted semen.
93	Three SO protocols were tested using the 2 products commercially available in Italy: Folltropin,
94	with a high FSH:LH ratio (49:1) (Henderson et al. 1990) and Pluset, with a low FSH:LH ratio (1:1)
95	(Kelly et al. 1995).

2. Materials and methods

- 98 2.1 Donors
- 99 The study was conducted on a dairy farm in Emilia Romagna, Italy, with the consent of the owner.
- No ethical approval was needed for the routine veterinary procedures and drugs used. This herd had
- 101 300 lactating Holstein cows, housed in a free-stall barn with cooling systems (ventilation and
- shower system), fed *ad libitum* and with an average daily milk yield of 40 L. The SO protocols were
- performed during spring or late autumn/winter to avoid the hottest part of the year.
- Donor cattle (36 primiparous and 41 multiparous cows) had a body condition score between 2.75
- and 3.5 (scale, 1-5) and had at least 2 physiologic estrous cycles after calving (the SO treatment was
- initiated 78 ± 15 d after calving). Cows with clinical illness, e.g. mastitis, lameness or
- gastrointestinal disorders with a considerable reduction in milk production and impaired general
- health after calving, were not used.
- 109 2.2 Superovulation protocols
- Potential donor cows were observed at least twice daily for behavioral signs of estrus and SO was
- induced by 9 im injections of decreasing dosages of gonadotropins at 12-h intervals over 4.5 d,
- beginning 9 to 11 d after the onset of standing estrus (Day 0). Concurrent with the seventh and
- eighth injections of gonadotropins, 150 µg d-cloprostenol (Dalmazin, Fatro, Ozzano dell'Emilia,
- 114 Italy), a PGF2a analog, was given im.
- Donors were randomly allocated to receive one of 3 SO protocols: 1) F700 (n=17), a total dose of
- 116 700 IU of Folltropin (Vetoquinol, Bertinoro, Italy) administered morning and evening as follows:
- st 117 1 day 150 IU and 127 IU, 2 day 108 IU and 87 IU, 3 day 74 IU and 63 IU, 4 day 45 IU and 29
- 118 IU, 5 day 17 IU; 2) F1000 (n=34), a total dose of 1000 IU of Folltropin administered morning and
- evening as follows: 1 day 170 IU and 155 IU, 2 day 135 IU and 125 IU, 3 day 115 IU and 100 IU,
- 120 4 day 88 IU and 77 IU, 5 day 35 IU; and 3) F700P300 (n=26) cows received 700 IU of Folltropin
- for the first 5 injections administered as follows: 1 day 170 IU + 155 IU, 2 day 135 IU + 125 IU,
- 122 3 day morning 115 IU + 300 IU of Pluset (Calier Italia, Milan Italy), for the last 4 injections

administered as follows: 3 day evening 100 IU, 4 day 88 IU + 77 IU, 5 day 35 IU.

2.3 Artificial insemination and semen

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At the time of the first insemination, a transrectal ultrasonographic examination was done to determine number and size of ovarian follicles present. Only cows with at least 3 ovarian follicles ≥ 0.8 cm in diameter were included in the study. Estrus was detected both with pedometers and visually by the herdsman who recorded behavioral estrous signs, including cow standing and being mounted or mounting other cows and vulvar discharge. On the basis of these observations, cows with signs of estrus within 36 h after the last PGF 2α analog injection were artificially inseminated. Inseminations were initiated 18 h after the onset of standing estrus and consisted of 3 inseminations, 5 h apart. For the first 2 inseminations, 2 straws of SexedULTRATM sex-sorted semen, containing 2 x 10° sperm each, were used, whereas on the third and final insemination, only 1 straw was used (total of 5 straws and 10 x 10 sex-sorted sperm). SexedULTRATM sex-sorted semen from 4 bulls were randomly used, so that each cow received semen from at least 3 bulls. At 5 h after the last insemination, ultrasound examinations were conducted to determine number of follicles ≥0.8 cm in diameter persistent on the ovaries and, based on this data, it was calculated if the cow was ovulating at least 50% of the follicles \ge 0.8 cm in diameter present on the ovaries at the time of the first insemination.

140 2.4 Embryo collection

Transcervical uterine flushing was done 7 d after onset of estrus. Embryos were evaluated under a stereomicroscope and classified according to the IETS classification guidelines (Robertson et al. 2010) for quality (Grade 1: excellent or good; Grade 2: fair; Grade 3: poor; Grade 4: dead or degenerating) and developmental stage (1: 1-cell; 2: 2 to 16-cell, 3: early morula; 4: morula; 5: early blastocyst; 6: blastocyst; 7: expanded blastocyst; 8: hatched blastocyst; and 9: expanded

147 2.5 Statistical analyses

hatched blastocyst).

Data were analyzed for normality using a Shapiro-Wilk test. Homogeneity of groups for parity, days in milk (DIM), milk production and BCS was evaluated using a one-way ANOVA or a Kruskall-Wallis ANOVA. Statistical differences in total recovery, transferable and non-transferable embryos and mean embryo grade were assessed by one way-ANOVA or Kruskal-Wallis ANOVA, using a Tukey HSD test for *post hoc* comparison or a Wilcoxon-Mann-Whitney test. A Chi-square test was used for analysis of embryo grade, embryo stage, embryo collection yield and ovulation rate. All statistical analyses were performed using IBM SPSS Statistics 23 (IBM Corporation, Milan, Italy). For all analyses, P<0.05 was considered significant.

3. Results

Groups were homogeneus for parity, DIM, milk production and BCS (P>0.05). Data regarding embryo collection are summarized in Table 1. Total number of recovered structures and transferable embryos were lower (P<0.05) in F700 versus F1000 and F700P300; however, there was no difference (P>0.05) for mean number of non-transferable structures. Although proportion of collections yielding no transferable embryos was not different (P>0.05) among groups (3/17 - 17.6% F700; 1/34 - 2.9% F1000; 2/26 - 7.7% F700P300), low embryo collections yielding <3 transferable embryos per flushing were higher (P<0.05) in F700 group (11/17 - 64.7%) than in F1000 (9/34 - 26.5%) or F700P300 (7/26 - 26.9%) groups. Percentage of cows ovulating \geq 50 % of follicles \geq 0.8 cm in diameter present at the first insemination was lower (P<0.05) in F700 (35.5%) than in F1000 (82.4%) and F700P300 (73.1%) groups. Quality grades were similar among groups, with no difference (P<0.05) in mean grade of transferable embryos (Table 2). Unfertilized oocytes were higher (P<0.05) in F700 than in F1000 and F700P300 (Table 3). Regarding transferable embryos stages, the only significant difference was percentage of morulae, which was highest in the F1000 group (P<0.05).

4. Discussion

174 In the present study, embryo yield was better in cows given 1000 versus 700 IU of gonadotropins. Group F700 was given 700 IU, the dose recommended by the manufacturer of Folltropin, a drug 175 with a high FSH:LH ratio (49:1) (Henderson et al. 1990), purified to avoid detrimental effects of 176 177 excessive LH. The other 2 groups received 1000 IU of gonadotropins, using the same drug (1000 IU of Folltropin), without modifying FSH:LH ratio throughout the entire SO treatment, or reducing the 178 179 FSH:LH ratio in the final part of the SO treatment, switching from Folltropin (700 IU) to Pluset 180 (last 300 IU), a drug with a 1:1 FSH:LH ratio. The rationale for keeping LH low from the beginning 181 to the end of the protocol was based on the premise that exogenous LH is not needed in the SO 182 process, as endogenous LH would support growth of ovarian follicles (Kanitz et al. 2002, Mapletoft 183 et al. 2002). Adding an extra dose of gonadotropin (300 IU) was prompted by reports that high 184 hepatic blood flow and metabolism in dairy cows promoted clearance of steroid hormones 185 (Sangsritavong et al. 2002, Wiltbank et al. 2006), thereby diminishing gonadotropin effect and the 186 SO response. The rationale for an extra dose of gonadotropin with a higher LH content was not only 187 the LH suppression linked to SO treatment, but that LH seems to be required, especially if there is 188 stress and/or negative energy balance, which attenuate or suppress the LH surge (Butler et al. 2003, 189 Matteri et al. 1982). The farm where the study was conducted had greater than average 190 overcrowding and competition, which along with high production and consequent diseases, are 191 important sources of stress for cows, with negative impacts on feed intake, BCS, and reproductive 192 performance. 193 In the F700 group, the number of recovered structures and transferable embryos were the lowest 194 among all 3 protocols. Cows in the F700 group had a good superovulatory response with several 195 follicles reaching pre-ovulatory size by first insemination, although many failed to ovulate, despite displaying estrous behaviors. In F700 group there were very few ovulations and many anovulatory 196 197 follicles persisting on the ovary 5 h after the last insemination. It is likely that, for the few follicles 198 which reached ovulation, that was a very asynchronous and prolonged process that yielded poor 199 quality, aged oocytes, resulting in the higher percentage of unfertilized eggs and degenerate

embryos in the F700 group.

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The success rate of SO did not differ between 1000 IU groups; the only significant difference was 201 202 that the F1000 group had more embryos at the morula stage, attributed to a more synchronous and 203 slightly delayed ovulation compared to the F700P300 group. These findings supported the notion 204 that less exogenous LH reduces premature ovulations. 205 We inferred that the better outcome of F700P300 and F1000 groups compared to F700 was due to 206 an increased total amount of gonadotropins administered, irrespective of the FSH:LH ratio in the 207 last part of the SO treatment. Further investigations are needed to clarify why a relatively high dose 208 of gonadotropins was needed to produce a satisfactory SO response. In F700 group, multiple 209 ovarian follicles developed until ovulatory size, so the first part of the SO process was considered 210 efficacious; however, ovulation was either delayed or failed to occur. Based on ovarian follicular 211 dynamics in this group, an altered LH release from the hypothalamus-pituitary axis most likely 212 caused development of these anovulatory structures (VanHolder et al. 2006). We inferred that 213 disturbed LH secretion in stressed cows was a critical point; this must be considered, along with 214 choosing an appropriate FSH dosage. It is noteworthy that a low dose of exogenous LH has been believed responsible for a lack of ovulation rate improvement in previous studies (Martinez et al. 215 1999, Ree et al 2009, Rosa et al. 2010). A notable lack of synchrony in ovulation was noted in dairy 216 217 heifers given 5.0 mg pLH (Ambrose et al. 2005); therefore, as 5.0 mg of pLH was considered inadequate to consistently synchronize ovulation, in subsequent experiments using higher doses 218 (12.5 and 25.0 mg) of pLH, it was determined that synchronization of ovulation was satisfactory 219 220 with at least 12.5 mg of pLH. A tendency toward a decreased ovulation rate in the group where LH 221 dose administered was 2.0 mg compared to 4.0 mg and eCG groups was reported (Oliveira et al. 222 2014), despite no significant difference among the protocols in average number of viable embryos 223 recovered. The authors had 2 potential explanations: the LH dose was too low to enhance ovulation 224 rate or a small quantity of pFSH on the last day of superstimulatory treatment may be necessary (in that study, on the last day of the SO, only pLH and no FSH were given; Oliveira et al. 2014). In the 225

226 present study, we inferred that: 1) superior results in both 1000 IU protocols were due to more FSH; 2) the amount of LH administered in the F1000 protocol (least amount) was adequate to increase 227 228 ovulation rate, thus additional LH was not warranted; 3) additional LH in the F700P300 protocol 229 (highest amount) induced more precocious ovulations as compared to F1000 group, as confirmed by 230 a higher percentage of morula-stage embryos in this group. In superovulated cows, SexedULTRATM sex-sorted semen yielded acceptable results (average of 4.5 231 232 embryos per flush), which seemed better than most studies involving traditional XY sex-sorted 233 semen in superovulated lactating dairy cows (3.1 embryos Schenk et al 2006; 2.4 embryos 234 Hayawaka et al 2009; 2.1 embryos Peippo et al 2009; and 2.4 embryos Monteiro et al 2016). 235 To our knowledge, only 2 studies with XY sorted semen had better results, with 6.4 (Soares 2011) 236 and 5.4 (Mikkola 2017) embryos per flush. It is noteworthy that cows used in both studies 237 responded successfully to the SO treatment with a standard dose of Folltropin. In Soares' study SO 238 protocol included also P4 and pLH, and the best results were achieved inseminating 18 and 30 h 239 versus 12 and 24 h after pLH (6.4 vs 4.6 embryos; Soares 2011). Comparisons of these studies are 240 challenging and must be done with caution, as there were many differences and many critical points 241 difficult to analyze and consider. It is noteworthy that in the present study, although environmental 242 and management conditions were somewhat stressful, the mean number of DIM was quite low and semen quality was not assessed before insemination, we considered that the SexedULTRATM sex-243 244 sorted semen yielded satisfactory outcomes.

5. Conclusions

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In conclusion, 700 IU of highly purified gonadotropins provided inadequate stimulation for SO of lactating dairy cows in this study. However, a better SO response was achieved by increasing total gonadotropin dose from 700 to 1000 IU, irrespective of increasing only FSH or both FSH and LH.

The use of SexedULTRATM sex-sorted semen for insemination of superovulated lactating dairy cows was considered satisfactory in terms of proportion of transferable embryos produced.

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256	7. Conflict of Interest Statement
257	There was no conflict of interest that could be perceived as prejudicing impartiality of the research
258	reported.

259 260	8. References
261	Ambrose JD, Kastelic JP, Rajamahendran R, Aali M, Dinn N. Progesterone (CIDR)-based timed AI
262	protocols using GnRH, porcine LH or estradiol cypionate for dairy heifers: ovarian and endocrine
263	responses and pregnancy rates. Theriogenology 2005;64:1457-1474.
264	
265	Barcelos ACZ, Satrapa RA, Nogueira MFG, Barros CM. Superstimulatory protocol P-36 in
266	Bonsmara breed: use of eCG and delay on induction of ovulation with LH. Acta Scientiae
267	Veterinariae 2006;34:s513.
268	
269	Ben Jabara MK, Carriere PD, Price CA. Decreased pulsatile LH secretion in heifers superovulated
270	with eCG or FSH. Theriogenology 1994;42:685-694.
271	
272	Boland MP, Goulding D, Roche JF. Alternative gonadotropins for superovulation in cattle.
273	Theriogenology 1991;35:5-17.
274	
275	Butler WR. Energy balance relationship with follicular development, ovulation and fertility in
276	postpartum dairy cows. Livestock Production Science 2003;83:211-218.
277	
278	Callesen H, Greve T, Hyttel P. Premature ovulations in superovulated cattle. Theriogenology
279	1987;28:155-166.
280	
281	Chupin D, Combarnous Y, Procureur R. Antagonistic effect of LH on FSH-induced superovulation
282	in cattle. Theriogenology 1984;21:229.
283	
284	Chupin D, Cognie Y, Combarnous Y, Procureur R, Saumande J. Effect of purified LH and FSH on
285	ovulation in the cow and ewe. In: Roche JF, O'Callaghan D, editors. Follicular Growth and

286 Ovulation Rate in Farm Animals, The Hague: Martinus Nijhoff; 1987, p. 66-72. 287 288 Cifuentes E, Quevedo L, Hoyos A, Carballo D, Piccardi M, Bó GA. Efecto de la aplicacion de eCG 289 en vacas donates de embriones superovuladas con Folltropin-V. Proceedings of VIII Simpósio 290 Internacional de Reproduccion Animal, Córdoba, Argentina 2009. 291 292 Donaldson LE, Ward DN. Effects of luteinising hormone on embryo production in superovulated 293 cows. Vet Rec 1986;119:625-626. 294 295 Donaldson LE, Ward DN, Glenn SG. Use of porcine follicle stimulating hormone after 296 chromatographic purification in superovulation of cattle. Theriogenology 1986;25:747-757. 297 298 Donaldson LE, Ward DN. LH effects on superovulation and fertilisation rates. Theriogenology 299 1987;27:225. 300 301 Ereno RL, Rosa FS, Oliveira ACS, Nogueira MFG and Barros CM. Use of LH or eCG in the last 302 day of superstimulatory treatment. Acta Scientiae Veterinariae 2010; 38 (Supl 2): s277-s315. 303 304 Gosselin N, Price CA, Roy R, Carriere PD. Decreased LH pulsatility during initiation of 305 gonadotropin superovulation treatment in the cow: evidence for negative feedback other than 306 estradiol and progesterone. Theriogenology 2000;54:507-521. 307 308 Goulding D, Williams DH, Roche JF and Boland MP. Superovulation in heifers using pregnant

mare's serum gonadotropin or follicle stimulating hormone during the mid luteal stage of the estrus

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cycle. Theriogenology 1991;36:949-958.

312 Greve T, Callesen H, Hyttel P. Characterization of plasma LH profiles in superovulated 313 dairy cows. Theriogenology 1984;21:237. 314 315 Hayakawa H, Hirai T, Takimoto A, Ideta A, Aoyagi Y. Superovulation and embryo transfer in 316 Holstein cattle using sexed sperm. Theriogenology 2009;71:68-73. 317 318 Henderson K, Weaver A, Wards ARL, Ball K, Lun S, Mullin C, and McNatty KP. Oocyte 319 production and ovarian steroid concentrations of immature rats in response to some commercial 320 gonadotrophin preparations. Reprod Fertil Dev 1990;2:671-682. 321 322 Herrler A, Elsaesser F, Parvizi N, Niemann H. Superovulation of dairy cows with purified FSH 323 supplemented with defined amounts of LH. Theriogenology 1991;35:633-643. 324 Hyttel P, Callesen H, Greve T, and Schmidt M. Oocyte maturation and sperm transport in 325 326 superovulated cattle. Theriogenology 1991;35:91-108. 327 Kanitz W, Becker F, Schneider F, Kanitz E, Leiding C, Nohner HP, Pohland R. Superovulation in 328 329 cattle: practical aspects of gonadotropin treatment and insemination. Reprod Nutr Dev 2002;42:587-330 599. 331 332 Kelly P, Duffy P, Baguisi A, Dobrinsky JR, Overstrom EW, Duby RT, Roche JF, and Boland MP. 333 Effect of FSH type and number of injections on peripheral FSH concentrations, follicle numbers 334 and embryo yield in heifers. Theriogenology 1995;43:245. 335 336 Kelly P, Duffy P, Roche JF, Boland MP. Superovulation in cattle: effect of FSH type and method of 337 administration on follicular growth, ovulatory response and endocrine patterns. Anim. Reprod Sci.

338 1997;46:1-14. 339 340 Mapletoft RJ, Bennett Steward K, Adams GP. Recent advances in the superovulation in cattle. 341 Reprod Nutr Dev 2002;42:601-611. 342 343 Martinez MF, Adams GP, Bergfelt DR, Kastelic JP, Mapletoft RJ. Effect of LH or GnRH on the 344 dominant follicle of the first follicular wave in beef heifers. Anim Reprod Sci 1999;57:23-33. 345 346 Matteri RL and Moberg GPJ. Effect of Cortisol or adrenocorticotrophin on release of luteinizing 347 hormone induced by luteinizing hormone releasing hormone in the dairy heifer. Endocrinology 348 1982;92:141-146. 349 Mikkola M, Taponen J. Quality and developmental rate of embryos produced with sex-sorted and 350 351 conventional semen from superovulated dairy cattle. Theriogenology 2017;87:135-140. 352 Monteiro PLJ, Batista AM, Almeida FC, Figueirêdo AES, Soares PC, Carneiro GF, Guerra MMP. 353 354 Fertilization rate and embryo production of superovulated dairy cows after insemination with non-355 sorted and sex-sorted semen. Animal Reproduction 2016;13:112-116. 356 357 Moor RM, Kruip ThAM, Green D. Intraovarian control of folliculogenesis: Limits to 358 superovulation? Theriogenology 1984;21:103-116. 359 360 Oliveira ACS, Mattos MCC, Bastos MR, Trinca LA, Razza EM, Satrapa RA, Sartori R, Barros CM. 361 Efficiency of superstimulatory protocol P-36 associated with the administration of eCG and LH in

Nelore cows. Theriogenology 2014;82:715-719.

362

364 Peippo J, Vartia K, Kananen-Anttila K, Räty M, Korhonen K, Hurme T, Myllymäki H, Sairanen A, 365 Mäki-Tanila A. Embryo production from superovulated Holstein-Friesian dairy heifers and cows 366 after insemination with frozen-thawed sex-sorted X spermatozoa or unsorted semen. Anim Reprod 367 Sci 2009;111:80-92. 368 369 Price CA, Carriere PD, Gosselin N, Kohram H and Guilbault LA. Effects of superovulation on 370 endogenous LH secretion in cattle, and consequences for embryo production. Theriogenology 371 1999;51:37-46. 372 373 Ouaresma MA, Lopes da Costa L, Robalo Silva J. Superovulation of Mertolenga cows with two 374 FSH preparations (FSH-P and FOLLTROPIN). Revista Portuguesa de Ciencias Veterinàrias 375 2003;98:81-84. 376 Ree TO, Colazo MG, Lamont AGA, Kastelic JP, Dyck MK, Mapletoft RJ, Ametaj BN, Ambrose DJ. 377 378 The effect of porcine luteinizing hormone in the synchronization of ovulation and corpus luteum 379 development in nonlactating cows. Theriogenology 2009;72:120-128. 380 381 Roberge S, Rieger D. Rawlings NC. Periovulatory LH, FSH and steroid hormone profiles in 382 superovulated and unstimulated Holstein heifers. Theriogenology 1995; 44:59-70. 383 384 Robertson I, Nelson RE. Certification and identification of embryos. In: Stringfellow D, Givens 385 MD, editors. Manual of the International Embryo Transfer Society. Fourth edition. Savoy, IL: IETS; 386 2010. p.86-105. 387 388 Rosa FS, Bonotto ALM, Trinca LA, Nogueira MFG, Barros CM. Eficiência do protocolo

supervovulatório P-36, associado à administração eCG ou LH em doadoras da raça Angus. Acta Sci

390 Vet 2010;37:711. 391 392 Sangsritavong S, Combs DK, Sartori R, Armentano LE, Wiltbank MC. High feed intake increases liver blood flow and metabolism of progesterone and estradiol 17b in dairy cattle. J Dairy Sci 393 394 2002;85:2831-2842. 395 396 Schenk JL, Suh TK, Seidel GE Jr. Embryo production from superovulated cattle following 397 insemination of sexed sperm. Theriogenology 2006;65:299-307. 398 399 Schmidt M, Greve T, Callesen H. Superovulation of cattle with FSH containing standardized LH 400 amount. Proceedings of 1st International Congress on Animal Reproduction and A.I., Dublin, 1988; 401 Vol. 2, p. 191. 402 403 Soares JG, Martins CM, Carvalho NA, Nicacio AC, Abreu-Silva AL, Campos Filho EP, Torres 404 Júnior JR, Sá Filho MF, Baruselli PS. Timing of insemination using sex-sorted sperm in embryo 405 production with Bos indicus and Bos taurus superovulated donors. Anim Reprod Sci 2011;127:148-406 153. 407 Takagi M, Kim IH, Izadyar F, Hyttel P, Bevers MM, Dieleman SJ, Hendriksen PJM, Vos PLAM. 408 409 Impaired final follicular maturation in heifers after superovulation with recombinant human FSH. 410 Reproduction 2001;121:941-951. 411 412 VanHolder T, Opsomer G, De Kruif A. Aetiology and pathogenesis of cystic ovarian follicles in 413 dairy cattle: a review. Reprod Nutr Dev 2006;46:105-119. 414 415 Vishwanath R, Moreno JF. Review: Semen sexing - current state of the art with emphasis on bovine species. Animal 2018;12:s85-s96.
Wiltbank M, Lopez H, Sartori R, Sangsritavong S, Gümen A. Changes in reproductive physiology
of lactating dairy cows due to elevated steroid metabolism. Theriogenology 2006;65:17-29.
Yamamoto M, Ooe M, Fujii C, Suzuki T. Superovulation of Japanese black heifers treated with
FSH-P and FSH-R. J Vet Med Sci 1993;55:133-134.