

*The following supplement accompanies the article*

**Chasing genetic structure in coralligenous reef invertebrates: patterns, criticalities and conservation issues**

**RUNNING HEAD: Connectivity and conservation of coralligenous reefs**

**Federica Costantini<sup>1, 2, 3\*</sup>, Filippo Ferrario<sup>4</sup>, Marco Abbiati<sup>1, 2, 3, 5</sup>**

**1 Dipartimento di Scienze Biologiche, Geologiche e Ambientali, Università di Bologna, UOS  
Ravenna**

**2 Centro Interdipartimentale di Ricerca per le Scienze Ambientali, Università di Bologna, Via  
S. Alberto 163, I – 48123 Ravenna, Italy**

**3 CoNISMa, Piazzale Flaminio 9, 00197 Roma, Italy**

**4 Québec-Océan, Université Laval, Québec, QC, Canada**

**5 Consiglio Nazionale delle Ricerche, Istituto di Scienze Marine, ISMAR, Via P. Gobetti 101,  
40129 Bologna, Italy**

## Supplementary methods

Combinations of keywords in the “Topic” field used to systematically search the literature from 2000 to 2016: “mediterranean” AND [“population genetic\*” OR “population genetic structur\*” OR “genetic structur\*” OR “phylogeography” OR “dispersal”] AND [“molecular marker\*” OR “microsatellite\*” OR “mitochondrial” OR “allozyme\*”] AND “marine invertebrate\*”.

Additionally, to ensure the inclusion of relevant articles on typical species of coralligenous habitat, the focus of our research, the main query was refined by replacing “marine invertebrate\*” in turn with 1) [“gorgonian\*” OR “coral” OR “cnidaria\*” OR “anthozoa\*”], 2) [“echinoderm\*” OR “sea star\*”], 3) [“sponge\*” OR “porifer\*”], and 4) [“tunicata\*” OR “ascidia\*”]. Six additional records identified through other sources were added (Figure 6).

### Data extraction:

Each entry in the full dataset included the following variables (Supplementary Table S1): (i) taxa analysed, (ii) dispersal potential (low vs. high), (iii) mating system (brooders vs. broadcaster), (iv) nutritional mode of the larvae (lecitotrophic, planktotrophic, direct development), (v) maximum pelagic larval duration (when data were not available in the analysed paper we obtained this information from other publications on the same species; see Supplementary data Table S2), (vi) molecular markers used, (vii) sampling area considered (ranked in four groups: Atlantic-Mediterranean transition, Mediterranean scale (thousand of kilometres), regional scale (hundreds of kilometres), local (tens of kilometres) and micro scale (meters)), (viii) total number of the populations, (ix) maximum geographic distance at Mediterranean scale, (x) global  $F_{ST}$  as carried out from each study, (xi) slope and standard errors of the regression line between pairwise  $F_{ST}$  and geographical distance. Pairwise  $F_{ST}$  were transformed using the formula by Baco et al.<sup>1</sup> and the slope of the regression were calculated using transformed  $F_{ST}$ , (xii) pattern of spatial genetic structure, and (xiii) basin from where the pairwise  $F_{ST}$  values were extrapolated.

## **Quantitative assessment of the genetic structure within the Mediterranean Sea:**

### **Meta-analyses**

The observation from Ledoux et al.<sup>2</sup> appeared to be an outlier within META-set because its ES standard error was at least an order of magnitude smaller than the others, conferring it with an excessive weight, and it was therefore excluded from the analysis (IBD slope  $\pm$  standard error =  $0.0079 \pm 0.0008$ ).

We first we analysed the effect of the single factors “Marker” and “Maximum PLD” and their additive effect. We did not include the interaction term because PLD values were unevenly distributed between levels of the factor “Marker”. We then tested the effect of “Phylum”, excluding the only observation belonging to tunicates. Because of a potential confounding effect in the analysis due to the overlap between groups “Marker” and “Phylum”, the last analysis was performed only after verifying the absence of a marker effect.

Since the Cochran's Q-test for heterogeneity for the null model was significant, we always used random effects models (where residual heterogeneity is calculated and tested against a  $\chi^2$ -distribution with n-1 degrees of freedom<sup>3</sup>;  $Q_{df30} = 147.69$ ,  $P < 0.0001$  for META-set;  $Q_{df20} = 127.17$ ,  $P < 0.0001$  for META-set for long PLD only). All models were fitted using the most popular DerSimonian-Laird estimator<sup>3</sup> and AIC was used to choose among competing models.

### **ANCOVA analysis**

We did not include the interaction “Marker  $\times$  Maximum Distance” because geographical distances within mtDNA groups were almost all clustered between 300 and 600 km (distance range 0 – 1200 km).

Contrary to meta-analyses, we did not test the factor “Phylum” in the ANCOVA, as the potential confounding effect of “Marker” could not be excluded because of its significance in terms that included it (either alone or in interaction, see Results).

Models were first fitted with maximum likelihood methods to allow model selection, based on AICc and significance assessment, via LRT. The best model was then refitted using the restricted maximum likelihood.

### **R Packages used**

The package *metafor*<sup>4</sup> was used for meta-analyses while *nlme*<sup>5</sup> for ANCOVA. Model selection was performed using packages *AICcmodavg*<sup>6</sup> and *glmulti*<sup>7</sup>.

### References

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**Supplementary Table S2.** Dataset of marine invertebrate pelagic larval duration and their references.

<i>Species</i>	Pelagic larval duration (PLD)	Reference
<i>Corallium rubrum</i>	4-12 d	Vighi (1972)
<i>Eunicella singularis</i>	122 d	Theodor (1967); Weinberg and Weinberg 1979
<i>Eunicella cavolinii</i>	4-12 d	Costantini F, Personal communication
<i>Paramuricea clavata</i>	6-23 d	Linares et al. (2008)
<i>Astroides calycularis</i>	2-3 d	Goffredo et al. (2010)
<i>Cladocora caespitosa</i>	2-3 d	Goffredo et al. (2010)
<i>Leptopsamnia pruvoti</i>	7-42 d	Goffredo et al. (2005)
<i>Balanophyllia europaea</i>	7-42 d	Goffredo and Zaccanti (2004)
<i>Echinaster sepositus</i>	3 d	Bailly and Flammang (2008)
<i>Marthasterias glacialis</i>	127 d	Barker and Nichols (1983)
<i>Amphipholis squamata</i>	absent	Boissin et al. (2008)
<i>Ophioderma longicauda</i>	weeks	Boissin et al. (2011)
<i>Ophiothrix fragilis</i>	21–26 d	Perez-Portela et al. (2013)
<i>Chondrosia reniformis</i>	2-3 d	Uriz et al. (1998)
<i>Crambe crambe</i>	2-3 d	Maldonado and Bergquist (2002)
<i>Paraleucilla magna</i>	2-3 d	Uriz et al. (1998)
<i>Phorbast fictitius</i>	2-3 d	Uriz et al. (1998)
<i>Scopalina lophyropoda</i>	2-3 d	Uriz et al. (1998)
<i>Ircinia fasciculata</i>	2-3 d	Uriz et al. (1998)
<i>Spongia lamella</i>	2-3 d	Maldonado and Bergquist (2002)
<i>Spongia officinalis</i>	2-3 d	Maldonado and Bergquist (2002)
<i>Botryllus schlosseri</i>	hours	Svane and Young (1989)
<i>Pseudodistoma crucigaster</i>	10 h	Tarjuelo et al. (2004)
<i>Clavelina lepadiformis</i>	2-3 d	Svane and Young (1989)
<i>Cystodytes dellechiaiei</i>	10 h	Tarjuelo and Turon (2004)
<i>Halocynthia papillosa</i>	2 weeks	Kim et al. 2012
<i>Pycnoclavella communis</i>	hours	Svane and Young (1989)

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**Supplementary Table S3.** Estimate dispersal distance for each species that demonstrated significant IBD using the method of Kinlan and Gaines (2003). For *Corallium rubrum* and *Crambe crambe* we used the already published values of mean dispersal distance calculated by Calderon et al. (2007)\* and Ledoux et al. (2010)\*\* using an “individual based” sampling. To calculate the estimated dispersal scale we used the formula used by Chust et al. (2016): dispersal distance =  $0.0016 (\text{IBD slope})^{-1.0001}$ .

Species	IBD	Slope IBD	Estimated dispersal distance (m)	References
<i>Crambe crambe</i>	yes	-	0.2	from Calderon et al. (2007)*
<i>Corallium rubrum</i>	yes	-	0.3	from Ledoux et al. (2010)**
<i>Astroides calycularis</i>	yes	0.0208	76.95	Casado-Amezua et al. (2012)
<i>Ophioderma longicauda</i> C5	yes	0.0275	58.21	Weber et al. 2015
<i>Paramuricea clavata</i>	yes	0.0382	41.9	Pilczynska et al. (2016)
<i>Eunicella cavolinii</i>	yes	0.0184	86.99	Masmoudi et al. (2016)
<i>Ophiothrix fragilis</i>	yes	0.0059	271.32	Taboada & Perez-Portela (2016)

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**Supplementary Figure S1.** Boxplots of maximum geographical distance per Phylum. Each boxplot describes the variability of the recorded maximum distance for each study included in the ANCOVA and/or Meta analysis. The thick line in the box represents the median, the box includes the 1st and 3rd quantile and the dotted lines extend to the minimum and maximum values. For each Phylum it is reported the sample size (N) and the distance range in km. Tunicates are presented for the sake of completeness, as they were not included in the meta-analysis testing the phylum effect.

