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1 **Survey on the presence of *Leishmania* sp. in peridomestic rodents from the**  
2 **Emilia-Romagna Region (North-Eastern Italy)**  
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11

12 **Abstract**

13 Leishmaniasis is a neglected vector-borne parasitic disease caused in Italy only by the  
14 species *Leishmania infantum* of the *Leishmania donovani* complex, which is the causative agent  
15 of the zoonotic visceral leishmaniasis (VL), and the sporadic cutaneous leishmaniasis (CL)  
16 in humans, and of the canine leishmaniasis (CanL). The disease is considered endemic in  
17 southern, central, and insular Italian regions and recognizes phlebotomine sand flies as  
18 vector and dogs as main reservoir. However, a specific north-eastern region, namely Emilia-  
19 Romagna, always showed a peculiar epidemiological situation when compared to the other  
20 northern Italian regions and recent studies are indeed questioning the role of dog as main  
21 reservoir of *L. infantum*. Due to their synanthropic relationship with humans, rodents have  
22 been tested for *Leishmania* spp. in several European countries. The aim of this study was to  
23 assess the presence of *Leishmania* spp. in peridomestic rodents in the Emilia-Romagna

24 Region. The study was carried out on 136 peridomestic rodents collected by professional  
25 rodent control services: 47 brown rats (*Rattus norvegicus*), 39 black rats (*Rattus rattus*) and 50  
26 mice (*Mus musculus*). Specimens of earlobe skin, spleen, liver and prescapular lymph nodes  
27 were tested with a real-time PCR. Fifteen (11 %) rodents, tested positive for *L. infantum*.  
28 Positivity was obtained from different target organs; notably 33% of the rodents tested  
29 positive in earlobe skin samples. These findings revealed the presence of *Leishmania* spp. in  
30 peridomestic rodents of the Emilia-Romagna Region, also in two species never tested before  
31 in Italy, namely *R. norvegicus* and *M. musculus*.

32 **Keywords:** Leishmaniasis, Italy, *Mus musculus*, *Rattus norvegicus*, *Rattus rattus*

33

## 34    **Background**

35    Leishmaniasis is a neglected vector-borne parasitic disease endemic in southwestern  
36    Europe. With reference to Italy, *Leishmania infantum* of the *Leishmania donovani* complex is  
37    the only species responsible for visceral leishmaniasis (VL), for sporadic cutaneous  
38    leishmaniasis (CL) in humans and for canine leishmaniasis (CanL) (Gramiccia and Gradoni  
39    2005; Rugna et al. 2018). The parasite is transmitted by phlebotomine sand flies, and in Italy  
40    dogs have long been recognized as the major reservoir in southern, central and insular  
41    regions, where the disease is considered endemic. Among the northern Italian regions,  
42    Emilia-Romagna has always had a different epidemiological scenario: CL has been widely  
43    reported since 1934, and between 1950-1958 up to 2,670 cases were diagnosed in the  
44    province of Forlì (Pampiglione 1975). In contrast, until the early 1970's, in this region VL  
45    appeared sporadically, with only 4 autochthonous cases observed, one in the province of  
46    Bologna and 3 in the province of Forlì. In the same period and within the same area, no  
47    ascertained autochthonous cases of CanL were reported (Pampiglione 1975). In 1971-1972,  
48    in two municipalities of Bologna province located in a foothill area a dramatic outbreak of  
49    VL was reported, involving 60 patients with a lethality of 21.7% (Pampiglione 1975). Since  
50    then, the geographic distribution of human and canine leishmaniasis has notably increased  
51    and the disease spread even in other regions of northern Italy, where many autochthonous  
52    cases of VL, CL and CanL have been reported (Gaspari et al. 2017; Mendoza-Roldan et al.  
53    2020). This epidemiological change may be due to environmental issues, occurrence of  
54    competent insect vectors and movement of infected dogs from endemic areas (Santi et al.  
55    2014). However, such changes might not be sufficient to explain the recurrent VL and CL

56 foci recorded in Emilia-Romagna Region (Gaspari et al. 2017), especially considering that  
57 molecular studies carried out on strains isolated from autochthonous cases of VL are  
58 questioning the role of dogs as reservoirs of *L. infantum* in this region, as earlier suggested  
59 (Pampiglione 1975; Rugna et al. 2018).

60 The role of wildlife has long been recognized as crucial in the transmission and maintenance  
61 of zoonotic agents and several sylvatic species are known to be susceptible to leishmaniasis.  
62 Considering their synanthropic relationship with humans and their abundance the role of  
63 rodents as possible leishmaniasis reservoirs has been questioned in different European  
64 countries (Alcover et al. 2021). Several studies established the presence of *L. infantum* in  
65 these hosts in Greece (Papadogiannakis et al. 2009; Tsakmakidis et al. 2017), Portugal  
66 (Helhazar et al. 2013) and Spain (Navea-Pérez et al. 2015; Galán-Puchades et al. 2019;  
67 Martín-Sánchez et al. 2020).

68 In Italy, a study performed in Sicily detected *L. infantum* by PCR in 45% of black rats, even  
69 if in this region the role of the dog as reservoir has been well established (Di Bella et al.  
70 2003). However, a study performed in Montecristo Island (Tuscany), revealed the presence  
71 of *L. infantum* in the 15.5% of rodents examined, even in the absence of domestic carnivores  
72 (Zanet et al. 2014).

73 The aim of this survey was to assess the presence of *Leishmania* spp. in peridomestic rodents  
74 collected in the Emilia-Romagna Region, Italy.

75

76

77

78 **Materials and Methods**

79 From June 2019 to June 2021, 136 peridomestic rodent carcasses were sampled during pest  
80 control programs from the provinces of Ferrara, Forlì-Cesena and Ravenna (Emilia  
81 Romagna) (figure 1): 47 brown rats, *Rattus norvegicus* (20 females and 27 males), 39 black  
82 rats, *R. rattus* (21 females and 18 males), 50 mice, *Mus musculus* (22 females and 28 males)  
83 were collected from the territory by professional rodent control services and stored at -20  
84 °C before processing.

85 The entire carcass was examined; species, sex and age classes were identified by  
86 morphological and metrical evaluation (CDC). Necropsies and samples collection were  
87 performed with sterile surgical instruments and when possible, according to the state of the  
88 carcasses, 25 mg of tissue were collected from earlobe skin, prescapular lymph node and  
89 liver, and 10 mg from the spleen (Helhazar et al. 2013). Due to the corruption of the remains,  
90 lymph nodes were not collected from 16 subjects. Samples were placed in sterile 1.5 ml tubes  
91 and stored at -20 °C.

92 DNA extraction was performed with PureLink® Genomic DNA Mini Kit (Invitrogen,  
93 ThermoFisher Scientific) following the manufacturer's instructions. For DNA amplification  
94 a real-time PCR protocol was performed targeting a 71-bp region of the minicircle  
95 kinetoplast DNA using primer pair Leish71Up (5'-  
96 CCAAACTTTTCTGGTCCTYCGGGTAG-3') and Leish71Do (5'-  
97 TGAACGGGATTCTGCACCCATTTTTC -3') (Tsakmakidis et al. 2017). Reactions were  
98 carried out in a total volume of 20 µL with 10 µL of PowerUP™ SYBR™ Green master mix  
99 (2X), 0.3 µM of each primer and 2 µL of DNA. The amplification was performed in

100 StepOnePlus Real-Time PCR System (Applied Biosystems) and the thermal cycling profile  
101 was as follows: 95 °C for 5 min, followed by 40 cycles at 95 °C for 5 sec., 60 °C for 30 sec. At  
102 the end of the amplification, a melting curve analysis was performed from 60 °C to 95 °C,  
103 with a slope of 0.3 °C to monitor primer dimers of non-specific product formation. Each  
104 sample was amplified in triplicate, the average temperature of melting (T<sub>m</sub>) observed was  
105 79.39 ± 0.15 °C and the average standard deviation observed in cts was 0.65. The standard  
106 curve was created with serial dilution of *L. infantum* DNA ranging from 10,000 to 0.1  
107 parasites per reaction. Each reaction was carried out by three replicates per dilution, in three  
108 independent experiments. The ct value cut-off was settled at mean ct value of 39.3 which  
109 corresponds to 1 parasite per mL of the original parasite suspension.

110 As a positive control the reference strain *L. infantum* MHOM/TN/80/IPT1, kindly provided  
111 by the Unit of Clinical Microbiology, Regional Reference Centre for Microbiological  
112 Emergencies (CRREM), St. Orsola-Malpighi University Hospital, Bologna, Italy, was used.  
113 Confidence intervals were calculated by R Studio (RStudio Team 2020).

114 **Results and Discussion**

115 Out of 136 subjects examined, 15 (11 %; 95% CI=5.7-16.3) were positive for *Leishmania* spp.  
116 In particular, 10.6% (95% CI=1.8-19.4) of brown rats, 12.8% (95% CI=2.5-23.7) of black rats  
117 and 10% (95% CI=1.7-18.3) of mice (Table 1). Of the five positive mice, three tested positive  
118 in two target organs - spleen and earlobe skin or spleen and liver or spleen and lymph nodes  
119 - the remaining two subjects tested positive only in lymph nodes or liver, respectively. The  
120 geographical distribution of the positive subjects appears homogeneous between the  
121 sampled sites (figure 1).



122 The present survey assessed the presence of *Leishmania* spp. in synanthropic rodents of the  
123 Emilia-Romagna Region. The conditions settled by the WHO (2010) for a species to be  
124 recognized as reservoir is the prevalence of infection > 20% and the availability of the  
125 parasite in blood and skin in sufficient amount to be ingested by a sand fly. In the  
126 Mediterranean area such conditions were globally assessed only for *M. musculus*, while *R.*  
127 *norvegicus* and *R. rattus* showed lower prevalence of infections (16.4% and 9.9%,  
128 respectively) (Alcover et al. 2021).

129 The prevalence values observed in the current study are below the average found in  
130 Portugal or Spain (Barcelona) where the 33.3% of examined rodents (*M. musculus* and *R.*  
131 *norvegicus*, Helhazar et al. 2013; *R. norvegicus*, Galán-Puchades et al. 2019) resulted positive,  
132 or the one reported in Granada (Spain) in mice (88.9%) (Martín-Sánchez et al. 2020) or in  
133 different rodent species (*R. rattus*, *M. musculus* and *Apodemus sylvaticus*) (27%) by Navea-  
134 Pérez et al. (2015), whilst it is higher than the prevalence observed in brown rats (5.9%) in  
135 Greece (Papadogiannakis et al. 2009). Further studies performed in Greece by Tsakmakidis  
136 et al. (2017) on spleen of *R. norvegicus*, *R. rattus* and *M. musculus* revealed a prevalence of  
137 19.58% comparable to the one herein reported. The majority of the studies evaluated the  
138 presence of the parasite in more than one target organ including skin, liver, spleen and blood  
139 (Helhazar et al. 2013; Martín-Sánchez et al. 2020; Navea-Pérez et al. 2015; Tsakmakidis et al.  
140 2017) while few studies examined only the spleen as target organ (Galán-Puchades et al.  
141 2019; Papadogiannakis et al. 2009). Testing more than one target tissue allow to increase the  
142 possibility to detect *Leishmania* spp. as observed also in our study. Three *M. musculus* here  
143 examined showed the presence of the parasite DNA in two different target organs (spleen

144 + lymph nodes and spleen + liver). Although the spleen is traditionally recognized as  
 145 *Leishmania* spp. target organ for PCR in different animal species (Papadogiannakis et al.  
 146 2009), our results showed the presence of *Leishmania* spp. in the earlobe skin samples from  
 147 33 % of the positive rodents pointing out that this tissue should be also considered. In fact,  
 148 wild animals are frequently collected in decomposition state and the putrefaction of the  
 149 target tissues, like visceral organs, may affect the integrity of the kinetoplast DNA (Múnoz-  
 150 Madrid et al. 2013).

151 In Italy, the role of black rats in the transmission of *L. infantum* has long been investigated,  
 152 starting from surveys performed in Tuscany in the 1980's (Pozio et al. 1985). Further studies  
 153 showed that *Phlebotomus perniciosus* and *P. perfiliewi* are attracted to *R. rattus* and that these  
 154 sand fly species become infected when they feed on black rats experimentally infected with  
 155 *L. infantum* (Pozio et al. 1985). More recent study carried out in Calabria (Italy) by Di Bella  
 156 et al. (2003) showed 45% positivity in the spleen of 22 *R. rattus* although in this region the  
 157 role of dogs as reservoirs has long been established. Zanet et al. (2014), reported 15.5%  
 158 prevalence in black rats examined in the Montecristo Island (Tuscany, Italy) where *L.*  
 159 *infantum* was recorded even in absence of domestic carnivore hosts. This value is similar to  
 160 the one (12.8%) obtained in the same host in our study, that moreover provided also data  
 161 on *R. norvegicus* and *M. musculus* (10.6% and 10% respectively) species not previously tested  
 162 for *L. infantum* in Italy.

163 Leishmaniasis in Emilia-Romagna has a peculiar epidemiological scenario compared to the  
 164 other Northern Italian regions. Recently Rugna et al. (2018), by Multilocus Microsatellite  
 165 Typing (MLMT) detected differences between *Leishmania* strains from men and sand flies to

166 the ones from dogs. The MLMT profiles showed all canine samples belonged to one group  
167 genetically related to Mediterranean MON-1 strain and similar to the VL samples from other  
168 Italian regions, while all but one VL Emilia-Romagna case, and the isolates from sand fly  
169 fell into a different group. Therefore, in this region the co-circulation of two distinct groups  
170 of *L. infantum* seems to occur, and the VL in humans could have different cycles involving  
171 *P. perfliewi* as a vector (Rugna et al. 2018; Calzolari et al. 2019) and might include other  
172 vertebrates, besides dogs, as reservoirs.

173 In two of the three provinces studied, Ravenna and Forlì-Cesena, foci of VL, usually located  
174 in hilly areas, were historically described. The rodent samples analyzed were collected in  
175 an area not higher than 50 m above sea level, where the density of phlebotomines is scant  
176 and, according to leishmaniasis regional control plan, in 2020 only CL cases have been  
177 reported (Santi et al. 2021). Further research should focus on studying which strains  
178 circulate in this area.

179 Also notable is the presence of a positive brown rat in the province of Ferrara, where  
180 autochthonous cases of leishmaniasis in both dogs and humans have never been recorded:  
181 the specimen was collected in a locality on the border between the provinces of Ferrara and  
182 Ravenna where the phlebotomine population is recorded as being moderate (Santi et al.  
183 2021). This finding, considering the consistent increase in geographical distribution of the  
184 disease and its vector, will require further investigation.

185 *L. infantum* is a vector-borne parasite and in its epidemiology many mammal species are  
186 involved, hence identifying which one may act as a reservoir in the Emilia-Romagna Region  
187 is an ambitious task due to the presence of different environments i.e. hilly or flatlands and

188 different distribution of sylvatic and peridomestic animals, which may possibly be involved  
189 in the parasite cycle. Even if the presence of the parasite in mammalian hosts is crucial, in  
190 order to fully understand his meaning as main reservoir or epiphenomena it should be  
191 associated with studies on the blood preferences of the phlebotomine vector.

192 The total prevalence observed in the present study (11%), despite being lower to the one  
193 required from WHO (2010) to establish a role of reservoir is comparable to the  
194 Mediterranean's one. As reported in previous studies, this value is far from being trivial:  
195 considering their close relationship with humans, their ability to colonize new environments  
196 and their impact on human health, rodents should not be neglected for their potential role  
197 in the transmission of Leishmaniasis, especially in urban areas (Alcover et al. 2021).

198 Although these preliminary findings are not sufficient to prove the role of peridomestic  
199 rodents as reservoirs of *L. infantum*, they nevertheless indicate the opportunity to further  
200 investigate their possible role in the epidemiology of different strains of *L. infantum*  
201 circulating in the Emilia-Romagna Region.

202

## 203 **List of abbreviations**

204 VL = Visceral Leishmaniasis

205 CanL = Canine Leishmaniasis

206 CL = Cutaneous Leishmaniasis

207

## 208 **Declarations:**

209 *-Ethics approval and consent to participate*

210 No ethical approval is officially required since the rodents examined had been subjected to  
211 pest control are considered pest species.

212 - *Consent for publication*

213 Not applicable

214 - *Availability of data and materials*

215 The datasets generated during and/or analyzed during the current study are available from  
216 the corresponding author on reasonable request

217 - *Competing interests*

218 The authors declare that they have no competing interests

219 - *Funding*

220 This study received no external funding

221 - *Authors' contributions*

222 MF and RG conceived the study. AM performed field work. AM and MC performed  
223 laboratory work and analyzed data. AM and MC wrote the first draft of the manuscript. MF  
224 and RG reviewed the manuscript. All authors read and approved the final manuscript

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Table 1: Real time PCR positive samples

ID	Specimen	Locality	Real-Time PCR			
			Earlobe Skin	Spleen	Liver	Lymph Node
57	<i>Mus musculus</i>	Bizzuno (RA)	ct= 32.7 (87)	ct= 29.68 (676)	Negative	NA
59	<i>Mus musculus</i>	Bizzuno (RA)	Negative	ct= 30.77 (316)	ct= 31.97 (143)	NA
67	<i>Mus musculus</i>	Bizzuno (RA)	Negative	Negative	Negative	ct= 33.61 (47)
98	<i>Mus musculus</i>	S. Alberto (RA)	Negative	ct= 36.71 (5.8)	Negative	ct= 37.07 (4.5)
111	<i>Mus musculus</i>	Bizzuno (RA))	Negative	Negative	ct= 35.9 (10)	Negative
4	<i>Rattus norvegicus</i>	Ravenna (RA)	ct= 34.25 (30.9)	Negative	Negative	Negative
86	<i>Rattus norvegicus</i>	Godo (RA)	Negative	Negative	ct= 36.47 (6.8)	Negative
141	<i>Rattus norvegicus</i>	Ravenna (RA)	ct= 37.75 (2.9)	Negative	Negative	Negative
175	<i>Rattus norvegicus</i>	Forlì (FC)	Negative	Negative	ct= 36.27 (7.8)	Negative
178	<i>Rattus norvegicus</i>	Argenta (FE)	Negative	Negative	ct= 36.67 (5.8)	Negative
37	<i>Rattus rattus</i>	Forlì (FC)	Negative	ct= 36.47 (6.8)	Negative	Negative
60	<i>Rattus rattus</i>	San Pietro in Campiano (RA)	ct= 36.86 (6.2)	Negative	Negative	Negative
95	<i>Rattus rattus</i>	Montaletto di Cervia (RA)	Negative	ct= 37.44 (6.2)	Negative	Negative
179	<i>Rattus rattus</i>	Montaletto di Cervia (RA)	Negative	Negative	Negative	ct= 36.63
206	<i>Rattus rattus</i>	Longastrino (RA)	ct= 37.89 (2.6)	Negative	Negative	Negative

Legend: Ct values are reported as mean ct of observed in different target organs with the estimated quantity of parasites/ml (mean standard deviation observed  $\pm 0.65$ ). Localities are as well reported with reference to the province: Ferrara (FE), Forlì-Cesena (FC) and Ravenna (RA).

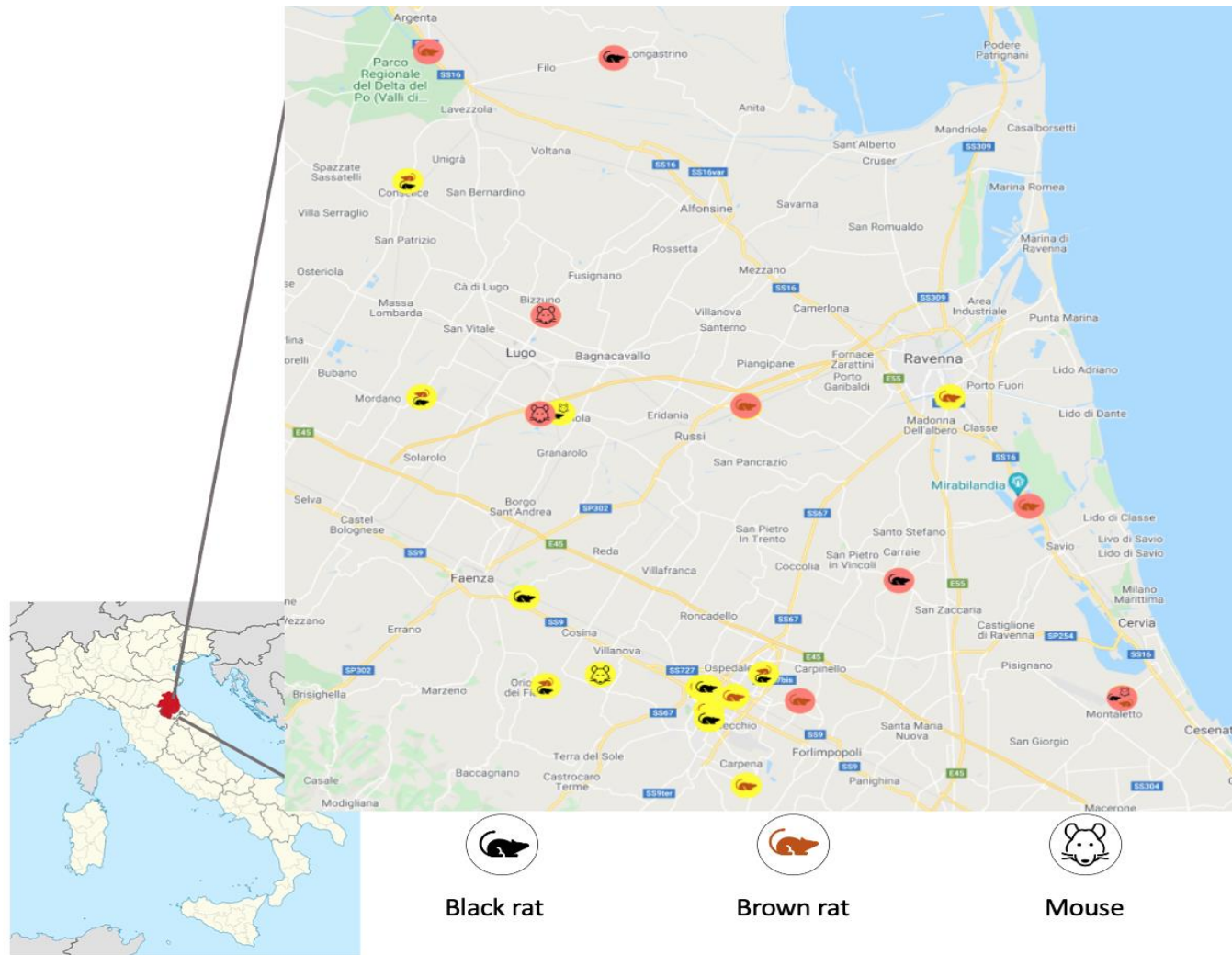


Figure 1. Map of the sampling area in the Emilia-Romagna Region. Dots are representative for sampling sites; red dots: at least one specimen positive, yellow dots: all the specimen negative.