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Antimicrobial resistance patterns in *Salmonella enterica* subsp. *enterica* and *Escherichia coli* isolated from bivalve molluscs and marine environment

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ABSTRACT

The current study presents data on the antimicrobial resistance (AMR) patterns of 102 *S. enterica* subsp. *enterica* (72 *Salmonella* ser. Typhimurium and 30 monophasic *S.* Typhimurium serovar) and 79 *Escherichia coli* (and their phylogenetic group determination) isolates from different species of bivalve molluscs and from the water samples collected from the sub-areas of a mollusc production area near Ferrara (Italy). These areas were classified as Long-line, Lupini, B-Out, B-in, and Sacca. A retrospective evaluation was performed to assess the spatial trends of the resistance patterns of *Salmonella* and *E. coli* and the temporal trend for *Salmonella*; the role of molluscs as AMR indicators and the potential use of *E. coli* as a microorganism indicator of AMR occurrence in a seawater environment were also investigated. Overall, 81% of *Salmonella* spp. and 75% of *E. coli* isolates were resistant to, at least, one antimicrobial agent (AA) and 44% and 38% of the isolates were multidrug resistant (MDR), respectively. The resistance levels of *Salmonella* were influenced by the investigated serovars. Monophasic *S.* Typhimurium serovar showed the highest resistance value with 70% of MDR isolates, in contrast with only 33% in *S.* Typhimurium. In monophasic *S.* Typhimurium versus *S.* Typhimurium, twofold resistance levels were observed to streptomycin (97 versus 43%), ampicillin (80 versus 40%) and tetracyclines (67 versus 36%). Regarding the temporal trend for *Salmonella*, strains were resistant to, at least, one AA, but this resistance was significantly lower during the first years of this 17-year sampling; however, in parallel MDR isolates, the resistance increased from 23% to a maximum level of 57% during the 2008-2012 period. On assessing the spatial trends, the Sacca area was found to show the lowest number of *Salmonella* spp. and *E. coli* strains resistant to, at least, one AA and MDR. For *E. coli*, the most commonly observed resistance was towards ampicillin (56%), streptomycin (52%), sulphonamides (30%) and ceftiofur (24%). The great majority (65%) of *E. coli* isolates belonged to the commensal phylogroups A and B1, with B1 as the dominant one, whereas most MDR isolates belonged to phylogroup C. Molluscs may be an efficient tool for antimicrobial resistance monitoring, and *E. coli* could be used as a microorganism indicator of the occurrence of antimicrobial resistance in seawater environment.

1. Introduction

Bivalve molluscs represent an important tool for monitoring antibacterial-resistant *Escherichia coli* and other members of the Enterobacteriaceae family in a coastal environment (Bighiu et al., 2019; Grevskott et al., 2017). Antimicrobial resistance (AMR) is a natural phenomenon, but the misuse and overuse of antibacterial agents in human and veterinary medicine, as well as in agriculture, have increased the release of these substances to the environment, which threatens global public health (Davies & Davies, 2010). Marine environments play an important role in accentuating antimicrobial resistance, as an unknown amount of these drugs ends up either indirectly in the receiving waters, or directly, as a result of intensive fish farming. As a consequence, living organisms could be exposed to a variety of these compounds present in the environment at low concentrations (Chiesaet al., 2018). Besides, coastal areas are subjected to faecal contamination of human and animal waste coming from a variety of sources, including rivers, runoff from agricultural and industrial activities and urban wastewater, resulting in the pollution of marine habitats (Vignaroli et al., 2016; Grevskott et al., 2017). In this context, marine ecosystems are not only an important reservoir for AMR, but also drive its emergence (Al-Sarawi et al., 2018; Taylor, Verner-Jeffreys, & Baker-Austin, 2011; Williams et al., 2016).

In the European Union (EU), the microbiological quality of coastal waters (Directive 2006/7/EC), as well as the sanitary control of shellfish produced and sold for human consumption, are consistently monitored by measuring the abundance of faecal indicator bacteria, *E. coli* and enterococci, in waters and the content of *E. coli* in the soft parts, flesh and intravalvular liquid of harvested bivalves for the classification of the mollusc production areas as A, B or C (Regulation EC 853/2004; Regulation EC 2017/265; Regulation EC 2019/265; Regulation EC 2019/267). In addition, the food safety criteria concerning bivalves entering the market states that the absence of *Salmonella* and the enumeration of *E. coli* are requested (Regulation EC 2073/2005; Regulation EC 2285/2015; Regulation 229/2019). In the European Union, *Salmonella* is the second most common

78 cause of human gastroenteritis (EFSA & ECDC, 2018). A considerable amount of epidemiological
79 data regarding the presence of *Salmonella* in seafood and its related illnesses is available, and the
80 risks of foodborne diseases associated with *Salmonella* in molluscs are classified as low (Davies &
81 Davies, 2010; NACMCF, 1992). Even if the microorganisms of the genus *Salmonella* are not
82 natural inhabitants of aquatic environments, several *Salmonella* serovars are widely distributed in
83 water (sea, estuarine, river) and in a variety of seafood, with the highest prevalence in molluscs,
84 shrimps, clams, and various fish species (Novoslavskij et al., 2016). *E. coli* is a common inhabitant
85 of the human and animal intestinal tract. *E. coli* may inhabit a host as a harmless symbiont or,
86 depending on the spectrum of encoded virulence factors, it can cause either intestinal or
87 extraintestinal infections (Logue et al., 2017). *E. coli* is a common indicator organism of faecal
88 contamination in aquatic systems, and it is recognized as an important player in the spread of
89 antibiotic resistance (Henriques et al., 2006; Szmolka & Nagy, 2013) due to its plasticity and high
90 aptitude to acquire genetic information through horizontal gene transfer. These characteristics
91 enable *E. coli* to exchange genetic material with other bacterial species (Araújo et al., 2017).
92 In Italy, a continuous microbiological monitoring of compliance with both the shellfish harvesting
93 areas and food safety criteria has been established by the official Veterinary Authorities, as required
94 by the EU regulations. Therefore, a continuous baseline data on the presence of *Salmonella* and *E.*
95 *coli* enumeration in water, shellfish flesh and intravalvular liquid is available. However, this
96 monitoring does not include the isolation and typing of *E. coli* isolates, as well as the AMR
97 evaluation of *Salmonella* and *E. coli.* The current study presents data on the AMR patterns of
98 *Salmonella* isolated with a continuous sampling history, from 2001 to 2017, and of *E. coli*, isolated
99 from 2016 to 2018, in the mollusc production area in the province of Ferrara, Emilia-Romagna
100 Region, Northern Italy. Here, even the distribution of *E. coli* in the phylogenetic groups was
101 investigated. A retrospective evaluation was performed with the following objectives: i) to
102 determine and compare the antimicrobial susceptibility among *Salmonella* serovars and the most
103 prevalent *E. coli* phylogenetic groups identified from bivalve molluscs and sea and brackish water,

as well as from different sampling areas; ii) to assess the spatial trends of resistance patterns of *Salmonella* and *E. coli* and the temporal trend for *Salmonella* in the mollusc production area of the province of Ferrara; iii) to investigate the role of molluscs as AMR indicators and to evaluate the potential use of *E. coli* as a microorganism indicator of AMR occurrence in a seawater environment.

2. Materials and methods

2.1 Salmonella spp. strain collection

In the present study, 102 *S. enterica* subsp. *Enterica* isolates from a strain collection of *Salmonella* isolates were selected: a total of 72 *Salmonella* ser. Typhimurium and 30 monophasic *S.* Typhimurium serovar (4,[5],12:i:-), isolated from 2001 to 2017, were considered. All these strains were isolated during the official monitoring performed by the regional Veterinary Authority and during a shellfish monitoring program using live bivalve molluscs and water samples from the seashore and inland channels of the province of Ferrara, Emilia-Romagna region, Italy. Most of these strains belong to a previous study (Rubini et al., 2018), which involved a total of 237 *Salmonella* isolates identified as *S. enterica* subsp. *enterica* (collected from a total of 10,757 seawater and molluscs samples from 1997 to 2015), in which 53 different serovars were observed. *S.* Typhimurium was the dominant serovar (26.9%), followed by its monophasic variant 4,[5],12:i:- (11.8%), Derby (6.3%), Newport (5.5%) and Thompson (4.6%). The *Salmonella* isolates considered in this study represent about 50% of isolates and almost all the serovars monophasic *S.* Typhimurium and *S.* Typhimurium collected in the investigated molluscs area. Each isolate refers to a specific area of classification (A or B), a different area of sampling (namely, specific sampling points within the considered area), a different source (Manila clams (*Ruditapes philippinarum*) (n = 30 isolates), striped clams (*Chamelea gallina*) (n = 1 isolate), mussels (*Mytilus galloprovincialis*) (n = 3 isolates) and water samples (n = 68 isolates), and a different time of sampling (see Table 1 of supplemental material). All the *Salmonella* strains were isolated in the North Western area of the

Adriatic Sea, facing the Southern area of the Po river delta, the major Italian river which, from spring to estuary, flows through the Po Valley (Pianura Padana) for a total of 652 km. The Po Valley is a densely populated area with a high number of large intensive animal farms. The Po river, near its end, in the Adriatic Sea, creates a wide delta with a surface area of 31 km^2 and an average depth of 1.5 m; its hydrographic network is mostly artificially regulated and, as a consequence, its freshwater flows are partially independent of rainfalls. More than one third of the lagoon surface is exploited for clam farming, with an annual production that has reached a maximum of 87,000 t/year in 2011 (Bison, 2012). This area has been divided into five sub-areas, following the mollusc production areas: I) long-line: the marine class A area used to breed mostly mussels and secondary oysters; ii) Lupini: the coastal marine area, including seawater between 1 and 2 nautical miles, that is classified as a class A area in which natural banks of striped clams are present and harvested; iii) B-Out: the narrow sea coastal area and inland waters, classified as a class B area; iv) B-In: class B area that includes the inner channels directly connected to the sea, together with the internal waters; v) Sacca: the class B area included between the Po river and the marine coastline (figure 1). These last three sub-areas are used to breed mussels, Manila clams and oysters.

2.2. E. coli isolation and phylogenetic group determination

The isolation of *E. coli* was performed starting from the official samplings of Manila clams (*R. philippinarum*) (n = 54), striped clams (*C. gallina*) (n = 3) and mussels (*M. galloprovincialis*) (n = 11), collected between 2016 and 2018 in the same aforesaid areas, and analysed for *E. coli* enumeration using the Most Probable Number (MPN) technique specified in ISO 16649-3, and from water samples (n = 11), using a membrane filtering method (APAT, 2003a. 2003b, 2003c). Each sample refers to a different area of classification, namely A and B, a different area of sampling, namely specific sampling points within the considered areas, a different source and a different time of sampling (see Table 2 of supplemental material). The MPN method utilizes Minerals Modified Glutamate Broth (MMGB) as the growth medium, and the material from

positive tubes, i.e., tubes whose colour has changed due to acid production, which was confirmed on Tryptone Bile with X-glucuronide (TBX) agar (Oxoid, Basington, UK); the suspected *E. coli* isolates were streaked onto MacConkey agar (Oxoid, Basington, UK) and incubated aerobically at 37 ± 1°C. After 24 hours of incubation, colonies of Gram-negative rods were streaked onto Tryptone Soya Agar (Oxoid, Basington, UK). All the isolates were subjected to a DNA extraction using the REDExtract-N-Amp tissue PCR kit (Sigma, St Louis, MO) and were identified using the PCR procedure described by Clermont and colleagues (Clermont et al., 2008). Overall, 79 *E. coli* strains were considered in this study and were subjected to phylogenetic group determination. *E. coli* isolates were assigned to one of the 7 phylogroups (A, B1, B2, C, D, E or F) based on the PCR amplification of *chuA*, *yjaA*, *arpA* and *trpA* genes and of the TspE4.C2 DNA fragment (Clermont et al., 2013). The reference *E. coli* strain ATCC 25922 and sterile water were used as the quality and negative control samples throughout the procedure, respectively.

2.3 Antibiotic susceptibility testing

All the isolates were tested for their antimicrobial susceptibility to twelve antibiotic agents, according to the agar disk diffusion method described by the Clinical and Laboratory Standard Institute (CLSI, 2016). The antimicrobial panel was chosen considering the importance of antimicrobial classes in the treatment of human and animal infections and the intrinsic resistance of *E. coli* reported by the European Committee on Antimicrobial Susceptibility Testing (EUCAST). The antimicrobials tested and the resistant breakpoints used in this study are reported in Table 1. For the breakpoint selection, when available, the epidemiological cut-off values proposed by EUCAST were used as the first choice. The EUCAST clinical breakpoints for Enterobacteriaceae were used as the second choice, and finally, for antimicrobials/bacterial species breakpoints not defined by EUCAST, the CLSI breakpoints for Enterobacteriaceae were chosen. Strains were considered multidrug resistant (MDR) when they showed resistance to three or more antimicrobial classes (Schwarz et al., 2010).

182 *2.4 Statistical analysis*

183 Descriptive statistics (absolute frequencies and percentages) have been provided regarding the
184 number of isolates: isolates resistant to, at least, one AA and MDR isolates. Findings were
185 presented by area of production, period of sampling, tested antimicrobial agent, *Salmonella*
186 serovars, and phylogenetic group of *E. coli*. The monitoring part was not planned and designed in
187 this study because it followed the real official activities performed by the Veterinary Authority.
188 Given the long period of official monitoring considered in this study, these 17 years of sampling
189 were arbitrarily divided into three different periods, namely, from 2001 to 2007 (17 isolates), from
190 2008 to 2012 (28 isolates) and from 2013 to 2017 (57 isolates), in order to have similar periods with
191 enough data to be compared. Furthermore, a limited number of samplings was performed for Long-
192 line, Lupini and B-out areas (11 for *Salmonella* and 19 for *E. coli* isolates), and therefore, the
193 overall isolates belonging to these areas were merged and, in the aggregate analysis, three definitive
194 areas were considered to assess the spatial resistance differences, namely in B-in, Sacca and the
195 area including Long-line, Lupini and B-out areas. Significant caution should be exerted for data
196 belonging to this merged area.

197 Pearson's chi-squared test and Chi-square test for trend were used to compare the temporal trends of
198 resistance patterns of *Salmonella*, whereas Chi-square test or Fisher's exact test were used to
199 compare the spatial trends for both *Salmonella* spp. and *E. coli* in the mollusc production areas of
200 the province of Ferrara. The significance limit was set at a $p < 0.05$.

201

202 **3. Results and Discussion**

203 In total, the antibiotic sensitivity tests showed that 19 (18.62%) *Salmonella* and 20 (25.31%) *E. coli*
204 isolates were susceptible to all the antimicrobials tested. Among the 102 *Salmonella* isolates,
205 encompassing the two different serovars, a total of 83 isolates (81.37%) showed resistance to, at
206 least, one antimicrobial agent and 45 isolates (44.12%) were MDR. More specifically, detailing the
207 patterns of resistance associated with the two considered serovars, monophasic *S.* Typhimurium

serovar showed the highest rates of resistance, with 96.67% and 70% of the isolates being resistant to, at least, one antimicrobial agent and having MDR, respectively, whereas 75% and 33.33% of *Salmonella* ser. Typhimurium isolates were resistant to, at least, one antimicrobial agent and possessed MDR, respectively. Tables 2 and 3 detail the antimicrobial susceptibility findings of *Salmonella* isolates of both serovars. The most common resistance in *Salmonella* isolates was towards streptomycin (58.8%), ampicillin (52%) and tetracyclines (45.1%), but different resistance levels were observed in the two considered serovars. The monophasic variant and *S.* Typhimurium showed resistance to streptomycin (97 and 43%, respectively), ampicillin (80 and 40%, respectively) and tetracyclines (67 and 36%, respectively). A very different behaviour was observed for other AAs: very high rates of resistance to sulphonamides (60%) were reported for the monophasic variant, whereas high resistance to carbapenems (23.6%) and chloramphenicol (21%) was observed in *S.* Typhimurium isolates. Over the years, the trend of resistance to, at least, one antimicrobial agent, significantly decreased ($p < 0.05$) from 94% to 72% and, in parallel, the proportions of MDR *Salmonella* isolates increased from 23.53% (4/17) between 2001 – 2007 to 57.14% (16/28) and 43.86% (25/57) in the 2008-2012 and 2013-2017 periods, respectively, even though no significant differences were observed. More details are presented in Table 4.

Among the 79 *E. coli* considered from 2016 to 2018, a total of 59 isolates (74.68%) showed resistance to, at least, one antimicrobial agent and 30 isolates (37.97%) were MDR. The most common types of resistance of *E. coli* isolates were to ampicillin (56%), streptomycin (52%), gentamycin (35,4%), sulphonamides (30%) and ceftiofur (23%). Regarding the phylogenetic group determination, most isolates belonged to phylogroup B1 (n = 40; 50.63%), followed by phylogroups A (n = 11; 13.92%) and C (n = 11; 13.92%), and even to other less frequent phylogroups, D (n = 7; 8.86%), B2 (n = 6; 7.59%) and E (n = 2; 2.53%). Two isolates were untypable and were classified as "unknown" (2.53%). The highest resistances were observed in phylogroup C, in which 90.91% and 72.73% of the *E. coli* isolates showed resistance to, at least, one antimicrobial agent and were MDR, respectively. Phylogroup C showed the highest resistances to almost all the considered

antimicrobial agents (data not shown). Tables 5 and 6 detail the antimicrobial susceptibility findings of *E. coli* isolates, also regarding the distribution of the frequent *E. coli* phylogroups.

No differences were observed regarding the resistance proportions of both *Salmonella* and *E. coli* isolates collected from molluscs and water samples, and from isolates belonging to classes A and B areas (data not shown). However, considering the spatial trends, the *E. coli* and *Salmonella* isolates in the area of Sacca had the lowest resistance for both serovars. Significant differences ($p < 0.05$) among the three different areas were observed, probably mostly regarding the lower percentage in Sacca, for MDR *Salmonella* spp. (20% in Sacca versus 51% in B-in and 73% in Long-line, Lupini B-out) and *S.* Typhimurium strains (9% in Sacca versus 67% in B-in and 39% in Long-line, Lupini B-out), but not for its monophasic variant. No significant differences were found regarding *Salmonella* spp. and *E. coli* strains resistant to, at least, one antimicrobial agent and even MDR among the three different areas. More details are reported in Table 7.

Extremely high resistance levels to, at least, one antimicrobial agent and high multi-resistant levels were observed in *Salmonella* spp. and *E. coli* isolates from molluscs, sea and brackish water in the investigated production area of Ferrara. The *Salmonella* data presented in this study comprise almost half of the overall *Salmonella* isolates of the last 20 years, from molluscs and water collected from the area of Ferrara, that is an important mollusc production area in Italy (Rubini et al., 2018), and therefore, they represent relevant data regarding the occurrence of antimicrobial resistance in *Salmonella* spp. isolates in molluscs. By plotting the amount of resistance of *Salmonella* for each period of the considered 17-year sampling, an evident rise in resistance from the first years of sampling with a doubling of the multi-resistance level in the following years must be mentioned, confirming the global antimicrobial resistance public health concern (ECDC, 2018; EFSA &ECDC, 2019). This increasing resistance is worrisome, particularly since several antimicrobial agents that were considered in this study are empiric or mere antibiotics which are commonly used in cases of serious human and animal infections.

Serovars monophasic *S.* Typhimurium and *S.* Typhimurium significantly contribute to the overall numbers of *Salmonella* isolates: in the European framework, they are among the most commonly reported serovars in human cases, as well as in food and animals, with the difference that *S.* Typhimurium has been associated with many food and animal sources, whereas monophasic *S.* Typhimurium has been mainly associated with pig and broiler sources (EFSA &ECDC, 2018). In molluscs, few data are available in the literature and the positive findings are usually reported as *Salmonella* spp.; however, the complexity of the global epidemiology of *Salmonella* requires improved monitoring data of those serovars which are of the highest epidemiologic importance (Rene et al., 2011).

Based on our findings, and in line with literature (EFSA &ECDC, 2019), the resistance levels for *Salmonella* spp. are greatly influenced by the serovars investigated, and therefore, the characterization at the serovar level is indispensable to address the temporal, geographic and source trends. The MDR level was high, overall, but monophasic *S.* Typhimurium had the highest MDR levels (70%) and the resistance level is twice as high for streptomycin, ampicillin, tetracyclines, and sulphonamides (see Table 3); in contrast, the highest proportion of resistance for meropenem and chloramphenicol were reported in *S.* Typhimurium isolates (see Table 3). *S.* Typhimurium and monophasic *S.* Typhimurium were the second and third most common *Salmonella* serovars identified in 2017 from human cases in Europe, respectively, in which the highest proportion of resistance was observed for ampicillin, sulphonamides and tetracyclines, with occurrences of 53, 48 and 44% in *S.* Typhimurium and of 87, 87 and 88% in monophasic *S.* Typhimurium (EFSA & ECDC, 2019). These data, even if with clear differences, is in line with our resistance findings, whereas for meropenem, a non-negligible level of resistance was observed in our study, compared to a full susceptibility found in human isolates. From a clinical point of view, fluoroquinolones and third-generation cephalosporins are classified as critically important antimicrobials (CIA) of the highest priority and represent the most important antimicrobial classes for treatment of

284 salmonellosis. Our findings showed moderate resistances to third-generation cephalosporins, but
285 which were still higher than data reported in humans, pigs, calves or cattle (EFSA & ECDC, 2019).
286 *E. coli* is an important foodborne pathogen. In addition, it is used worldwide as an indicator of
287 faecal contamination and as a hygiene indicator regarding sanitary quality or unsanitary conditions
288 (Metz, Sheehan, & Feng, 2020). In antimicrobial resistance, *E. coli* is also considered as a sensor of
289 the situation at each moment, and it has emerged as a major player in resistance: *E. coli* is typically
290 chosen as the representative indicator of antimicrobial resistance in Gram-negative bacteria, and its
291 monitoring in a specific population provides valuable data on the resistance occurring in that
292 population (EFSA & ECDC, 2019). Regarding the data on antimicrobial resistance in *E. coli*
293 isolated from molluscs and seafood or marine environments, available in the literature, comparisons
294 are arduous due to the differences in antimicrobial agents and breakpoints used; in most studies, the
295 most common resistances were observed for tetracycline, trimethoprim-sulfamethoxazole,
296 ampicillin and streptomycin, with lower prevalence values than those found here (Changkaew et al.,
297 2014; Grevskott et al., 2017; Ryu et al., 2012; Van et al., 2008; Vignaroli et al., 2015; Vignaroli et
298 al., 2016; Wang et al., 2011) or with values in line with ours (Al-Sarawi et al., 2018; Bighiu et al.,
299 2019). The prevalence of MDR strains in our study is higher than in most of the aforementioned
300 studies, but is still in line with those of other studies (Bighiu et al., 2019; Boss, Overesch, &
301 Baumgartner, 2016; Changkaew et al., 2014; Kumaran et al., 2010; Van et al., 2008). In addition, in
302 the European Union, the monitoring of antimicrobial resistance using the indicator *E. coli* in food-
303 producing animals (pigs and calves) and in their food products, has been mandatory since 2014: in
304 both animal species, tetracycline resistance was the most common trait, followed by resistance to
305 sulfamethoxazole and ampicillin (EFSA & ECDC, 2019).
306 *E. coli* is a common commensal inhabitant of the gastrointestinal tract, but it is also a common
307 cause of severe infections in humans, being the most frequent cause of bloodstream infections and
308 urinary tract infections among the Gram-negative bacteria. *E. coli* is involved in infections of both
309 community and healthcare origin, as well as being associated with intra-abdominal infections and

causing neonatal infections, such as early and late neonatal sepsis (ECDC, 2018; Wang et al., 2011). *E. coli* is among the infectious agents whose antimicrobial resistance has reached an extremely worrisome situation (WHO, 2014). According to the last report of the European Center for Disease Control (ECDC) (ECDC, 2018) on antimicrobial resistance obtained through the European Antimicrobial Resistance Surveillance Network database, that included only data from invasive isolates (blood and cerebrospinal fluid), in Italy, in 2017, the prevalence of *E. coli* resistant to aminopenicillins (ampicillin or amoxicillin), fluoroquinolones and to third-generation cephalosporins was 67% (EU mean value, 58%), 45% (EU mean value, 26%) and 29.5% (EU mean value, 15%), respectively. These data roughly reflect our findings regarding the resistance to ampicillin (56%) and to third-generation cephalosporins (24%), except for enrofloxacin (5%) (see Table 5). By contrast, while carbapenem resistance remains rare in Europe, and in Italy (0.3%) (ECDC, 2018), in our study, worrisome percentages (above 10%) of this type of resistance were observed. Carbapenems are among the most frequently prescribed antibiotics for the treatment of bacterial infections in humans, even if their utility is being threatened by the worldwide rise of carbapenem-resistant Enterobacteriaceae. Although the use of carbapenems is prohibited in food-producing animals and restricted to pets in most European countries, these findings illustrate the continuous spread of these highly resistant bacteria, accompanied by emerging public health problems (Roschanski et al., 2017). In our study, resistance to ceftiofur was the fifth most common resistance type. Ceftiofur is used in human and veterinary medicine and it has an activity against both Gram positive and negative bacteria; however, third-generation cephalosporins have been categorized as CIA and substances of the highest priority, while 3rd and 4th-generation cephalosporins, as well as fluoroquinolones and polymyxins, are included in category 2, according to the European Medicines Agency. Thus, those veterinary antimicrobials represent a higher estimated risk for public health than other classes of antimicrobials. Our findings confirm that bivalve molluscs are an efficient tool for identify antimicrobial resistance, being able to detect antimicrobials used in the past and still used in food-producing animals and human beings, as well

as a useful opportunity to drive emergent threats to humans in terms of both antimicrobial resistance and clinical aspects.

Given the worldwide phylogenetic analyses, *E. coli* showed an apparent clonal population structure and a clear phylogenetic signal that appropriately reflect the relationship between strains (Tenaillon et al., 2010). Different phylogenetic groups play distinct ecological roles and, therefore, the classification of *E. coli* in the phylogenetic groups is important to understand its pathogenesis and interaction with hosts. Hitherto, a robust phylogeny, was built and 8 phylogenetic groups were identified (A, B1, B2, C, D, E, F, and the *Escherichia* cryptic clade I). The *E. coli* strains responsible for extra-intestinal infections were far more likely to be members of phylogroups B2 or D, rather than of A or B1, that usually lack a distinct virulence profile and are classified as commensal or diarrheagenic strains (Clermont et al., 2013). In general, strains belonging to different phylogenetic groups show different phenotypic and genotypic traits (Gordon & Cowling, 2003); differences in the phylogroup distribution are mainly ascribable to geographical (location and climate) and host (diet, gut morphology, body mass) factors, explaining variable animals and human phylogroup identification in several studies in the literature. The phylogroups that are considered to contain highly virulent extra-intestinal *E. coli* strains are the B2 phylogroup, whose microorganisms are a major cause of bacteraemia and neonatal sepsis (Cole et al., 2019; Vila et al., 2016) and, to a lower extent, the D phylogroup, albeit it was detected as the most common in urinary tract infections in humans (Farajzadeh Sheikh, 2019). Particular *E. coli* clones, more frequently belonging to the phylogroup B2, are more prone than others to cause bacteraemia, but they cannot be considered as having a deterministic causation relationship with extra-intestinal diseases (Vila et al., 2016); the B2 group exhibits the highest diversity at both the nucleotide and the gene content level, supporting its early emergence in the species lineage and suggesting that it has a subspecies status. Considering the findings obtained in our study using Clermont typing, the great majority (65%) of *E. coli* isolates belong to the commensal phylogroups A and B1 (these groups appear as sister groups), but phylogroup B1 strains dominated with a prevalence of 51%,

both in general and in each of the three considered sampling areas. These data are in agreement with several studies reporting a higher prevalence of A and B1 phylogroups in molluscs, with A being the dominant one (Luna et al., 2010; Vignaroli et al., 2016; Vignaroli et al., 2012) in surface water, in several animals (Johnson et al., 2017; Tomazi et al., 2018) and in water and vegetables (Araújo et al., 2017). Unlike the studies performed by Vignaroli in *Chamelea gallina* clam in Marche Region, Italy, and in marine sediments in the Adriatic Sea, in which the prevalence of MDR strains was higher in phylogroup A, in our study, most MDR isolates belonged to phylogroup C, followed by B2. Phylogroup C was the only one resistant to chloramphenicol and trimethoprim-sulfamethoxazole, and for which the highest resistance values were reported for ampicillin, streptomycin, sulphonamides, and tetracycline (data not shown). In addition, it should be noted that, whereas isolates belonging to A and B1 phylogroups were observed to be MDR, depending on the considered area of sampling, isolates belonging to phylogroup C were MDR, in spite of the area considered. It has been observed that strains of these phylogroups vary in their phenotypic and genotypic characteristics, ecological niche, lifestyle, and propensity to cause disease (Tenaillon et al., 2010); therefore, these findings confirm that waters receive contamination from a variety of sources. In this context, there is a growing body of evidence suggesting that *E. coli* can not only survive for extended periods of time, but also proliferate in several different environments, such as water, soil, algae, plants, and manure, also in the absence of faecal inputs, thus supporting its use as a water quality indicator (Nanayakkara, O'Brien, & Gordon, 2019) and as an indicator organism of faecal contamination (Vignaroli et al., 2015). In addition, *E. coli* isolates that are responsible for elevated counts (blooms) in freshwater reservoirs carry a capsule originating from *Klebsiella* spp.; overall, about 7% of *E. coli* strains have acquired these capsules, which were observed to be non-random distributed and restricted to A, B1 and C phylogroups (Nanayakkara, O'Brien, & Gordon, 2019). Given the limited number of isolates used in this study, and also the absence of any isolate characterization, authors could not resolve these doubts. However, the diversity in both the

distribution of phylogroups among the different areas and the antimicrobial resistance patterns remains.

Regarding the spatial trends of resistance, the area of Sacca was the one with the lowest number of *Salmonella* spp. and *E. coli* isolates resistant to, at least, one antimicrobial agent and MDR; conversely, the B-in area was the area with the highest number of *Salmonella* spp. and *E. coli* strains resistant to, at least, one antimicrobial agent and MDR. However, whereas for *Salmonella* spp., isolates collected from the Sacca area were susceptible to 6 out of the 12 antimicrobials tested and showed the lowest levels of resistance, for *E. coli*, resistance was observed for all the antimicrobials, even if with the lowest levels of resistance observed. In contrast, the B-in area was the area with the highest number of *Salmonella* spp. And *E. coli* strains resistant to, at least, one antimicrobial agent and MDR, as well as having the highest levels of resistance to the antimicrobial agents. See Tables 2 and 5 for more details. Even if neither the area nor the environment could be considered truly pristine, owing to the transfer of antibiotics and resistance genes via the wind, tides, bird migration and other environmental elements (Barkovskii et al., 2012), the B-in area is directly subjected to anthropogenic activities and it is exposed to industrial, municipal, agricultural and zootechnical impacts, whereas the Sacca area is certainly less anthropogenically impacted than the others. The aforesaid occurrence frequencies of antimicrobial resistance, as well as the distributions of resistant *E. coli* isolates among the phylogroups, identify the B-in area as the main source of antimicrobial resistance. *E. coli* strains that can presumably cause human extraintestinal infections are not prominent within the *E. coli* population of the aquatic environment, but, at the same time, our findings allow us to speculate on the fact that molluscs, and therefore, water environment sites, cannot be considered risk-free. Obviously, further studies should be performed to understand and verify the ecology of allochthonous and indigenous bacteria, as well as the pathogens in aquatic environments.

4. Conclusion

In conclusion, bivalve molluscs are confirmed to be an efficient tool to detect antimicrobial resistance. This study presents data on the antimicrobial resistance in *Salmonella* and *E. coli* strains isolated from molluscs and water and showed high multi-resistant levels, as well as spatial and temporal trends of resistance; however, for trend analysis of resistance, further studies are needed. The overlap of the antimicrobial resistance data collected in this study with the previously reported data of isolates from food animals and human beings suggests that testing the isolates from water and molluscs could be a useful tool to monitor the evolution of AMR of some bacterial species. In this context, routine antimicrobial susceptibility testing could be included as a parameter to be investigated by official laboratories using isolates collected from different species of bivalve molluscs, sea and brackish water by the official Veterinary Authorities during the official monitoring of molluscs. Starting from the strength of the European microbiological monitoring of molluscs and their production areas, the implementation of this step could further allow the development of an official network at the national and/or European level, which could be able to optimize the data referring to activities already planned and which could be performed by official veterinary authorities. This monitoring could be useful to raise a constant and ongoing awareness of antimicrobial resistance in the environment.

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References

853/2004/EC, R. (2004). Commission Regulation (EC) No. 853/2004 of the European Parliament and of the Council laying down specific hygiene rules for food of animal origin. In E. Union

(Ed.), *853/2004*. Brussels, Belgium: The European parliament and the council of the European Union.

2073/2005/EC, R. (2005). Commission Regulation (EC) No. 2073/2005 on microbiological criteria for foodstuffs. In E. Union (Ed.), *2073/2005*. Brussels, Belgium: The European parliament and the council of the European Union.

2285/2015/EC, R. (2015). Commission Regulation No. 2285/2015. (2015). Amending Annex II to Regulation (EC) No 854/2004 of the European Parliament and of the Council laying down specific rules for the organisation of official controls on products of animal origin intended for human consumption as regards certain re- quirements for live bivalve molluscs, echinoderms, tunicates and marine gas- tropods and Annex I to Regulation (EC) No 2073/2005 on microbiological criteria for foodstuffs. In E. Union (Ed.), *2285/2015*. Brussels, Belgium: The European parliament and the council of the European Union.

625/2017/EC, R. (2017). Commission Regulation (EU) No. 2017/625. Regulation of the European Parliament and of the Council of 15 March 2017 on official controls and other official activities performed to ensure the application of food and feed law, rules on animal health and welfare, plant health and plant protection products, amending Regulations (EC) No 999/2001, (EC) No 396/2005, (EC) No 1069/2009, (EC) No 1107/2009, (EU) No 1151/2012, (EU) No 652/2014, (EU) 2016/429 and (EU) 2016/2031 of the European Parliament and of the Council, Council Regulations (EC) No 1/2005 and (EC) No 1099/2009 and Council Directives 98/58/EC, 1999/74/EC, 2007/43/EC, 2008/119/EC and 2008/120/EC, and repealing Regulations (EC) No 854/2004 and (EC) No 882/2004 of the European Parliament and of the Council, Council Directives 89/608/EEC, 89/662/EEC, 90/425/EEC, 91/496/EEC, 96/23/EC, 96/93/EC and 97/78/EC and Council Decision 92/438/EEC (Official Controls Regulation). In E. Union (Ed.), *265/2017. Official Journal of the European Union, L 95*.

625/2019/EC, R. (2019). Commission Delegated Regulation (EU) No. 2019/625. Commission Delegated Regulation of 4 March 2019 supplementing Regulation (EU) 2017/625 of the

European Parliament and of the Council with regard to requirements for the entry into the Union of consignments of certain animals and goods intended for human consumption. In E. Union (Ed.), *265/2019*. *Official Journal of the European Union, L 131*.

627/2019/EC, R. (2019). Commission Implementing Regulation (EU) No. 2019/627. Commission Implementing Regulation of 15 March 2019 laying down uniform practical arrangements for the performance of official controls on products of animal origin intended for human consumption in accordance with Regulation (EU) 2017/625 of the European Parliament and of the Council and amending Commission Regulation (EC) No 2074/2005 as regards official controls. *Official Journal of the European Union, L 131*.

229/2019/EC, R. (2019). Commission Regulation (EU) N0. 2019/229. Commission Regulation of 7 February 2019 amending Regulation (EC) No 2073/2005 on microbiological criteria for foodstuffs as regards certain methods, the food safety criterion for Listeria monocytogenes in sprouted seeds, and the process hygiene criterion and food safety criterion for unpasteurised fruit and vegetable juices (ready-to-eat). *Official Journal of the European Union, 37/106.*

7/2006/EC, D. (2006). Directive 2006/7/EC of the European Parliament and of the Council of 15 February 2006 concerning the management of bathing water quality and repealing Directive 76/160/EEC. *Official Journal of the European Union,* 37 r1.

Al-Sarawi, H. A., Jha, A. N., Baker-Austin, C., Al-Sarawi, M. A., & Lyons, B. P.. (2018). Baseline screening for the presence of antimicrobial resistance in *E. coli* isolated from Kuwait's marine environment. *Marine Pollution Bulletin*, 129, 893–898.

APAT CNR IRSA. (2003a). APAT *Manuali e Linee Guida 29/2003* (p. 883). Metodi analitici per le acque- Volume 3, 7000-Metodi per la determinazione di microorganismi indicatori di inquinamento e di patogeni - APAT CNR IRSA 7030 *Escherichia coli* http://www.irsa.cnr.it/Docs/Capitoli/1000.pdf.

APAT CNR IRSA. (2003b). APAT *Manuali e Linee Guida 29/2003* (p. 875). Metodi analitici per le acque- Volume 3, 7000-Metodi per la determinazione di microorganismi indicatori di

490 inquinamento e di patogeni - APAT CNR IRSA 7020 Coliformi fecali
491 http://www.irsa.cnr.it/Docs/Capitoli/1000.pdf.
492 APAT CNR IRSA. (2003c). APAT *Manuali e Linee Guida 29/2003* (p. 927). Metodi analitici per le
493 acque- Volume 3, 7000-Metodi per la determinazione di microorganismi indicatori di
494 inquinamento e di patogeni - APAT CNR IRSA 7080 *Salmonella* spp.
495 http://www.irsa.cnr.it/Docs/Capitoli/1000.pdf.
496 Araújo, S., Silva, I. A.T., Tacão, M., Patinha, C., Alves, A., & Henriques, I. (2017).Characterization
497 of antibiotic resistant and pathogenic *Escherichia coli* in irrigation water and vegetables in
498 household farms. *International Journal of Food Microbiology*, 257, 192–200.
499 Barkovskii, A. L., Thomas, M., Hurley, D., & Teems, C. (2012). Environmental factors responsible
500 for the incidence of antibiotic resistance genes in pristine Crassostrea virginica reefs. *Marine*
501 *Pollution Bulletin*, 64, 2692–2698.
502 Bighiu, M. A., Norman Haldén, A., Goedkoop, W., & Ottoson, J. (2019). Assessing microbial
503 contamination and antibiotic resistant bacteria using zebra mussels (Dreissena polymorpha).
504 *Science Total Environment*, 650,2141–2149.
505 Bison, G. O. (2012). Gli obiettivi per il futuro: Fidelizzare il consumatore al brand "Vongola di
506 Goro" e commercializzare confezioni sottovuoto con sugo pronto a fianco. *Il Pesce*, 2, 52.
507 http://www.pubblicitaitalia.com/ilpesce/2012/2/ 11664.html.
508 Boss, R., Overesch, G., & Baumgartner, A. (2016). Antimicrobial resistance of *Escherichia coli*,
509 enterococci, *Pseudomonas aeruginosa*, and *Staphylococcus aureus* from raw fish and
510 seafood imported into Switzerland. *Journal of Food Protection*, 79, 1240–1246.
511 Changkaew, K., Utrarachkij, F., Siripanichgon, K., Nakajima, C., Suthienkul, O., & Suzuki Y.
512 (2014). Characterization of antibiotic resistance in *Escherichia coli* isolated from shrimps
513 and their environment. *Journal of Food Protection*, 77, 1394–1401.
514 Chiesa, L. M., Nobile, M., Malandra, R., Panseri, S., & Arioli F. (2018). Occurrence of antibiotics
515 in mussels and clams from various FAO areas. *Food Chemistry*, 240, 16–23.

516 Clermont, O., Christenson, J. K., Denamur, E., & Gordon, D. M. (2013). The Clermont *Escherichia*
517 *coli* phylo-typing method revisited: Improvement of specificity and detection of new phylo-
518 groups. *Environmental Microbiology Reports,* 5, 58–65.
519 Clermont, O., Lescat, M., O'Brien, C. L., Gordon, D. M., Tenaillon, O., & Denamur, E. (2008).
520 Evidence for a human-specific *Escherichia coli* clone. *Environmental Microbiology,* 10,
521 1000–1006.
522 CLSI. (2016). Clinical and Laboratory Standards Institute. (2016). Performance standards for
523 antimicrobial susceptibility testing. *CLSI Supplement.*
524 Cole, B. K., Ilikj, M., McCloskey, C. B., & Chavez-Bueno, S. (2019). Antibiotic resistance and
525 molecular characterization of bacteremia *Escherichia coli* isolates from newborns in the
526 United States. *PLoS One,* 14:e0219352.
527 Davies, J., & Davies, D. (2010). Origins and Evolution of Antibiotic Resistance. *Microbiology*
528 *Molecular Biology Reviews,* 74(3, 417-33.
529 ECDC. (2018). Surveillance of antimicrobial resistance in Europe Annual report of the European
530 Antimicrobial Resistance Surveillance Network (EARS-Net) 2017. *ECDC: Surveillance*
531 *Report*. European Centre for Disease Prevention and Control (ECDC).
532 EFSA & ECDC. (2019). The European union summary report on antimicrobial resistance in
533 zoonotic and indicator bacteria from humans, animals and food in 2017. *EFSA Journal,*
534 *17(2), 5598.* European Food Safety Authority and European Centre for Disease Prevention
535 and Control (EFSA and ECDC).
536 EFSA & ECDC. (2018). The European Union summary report on trends and sources of zoonoses,
537 zoonotic agents and food-borne outbreaks in 2017. *EFSA Journal,* 16(12), 5500. European
538 Food Safety Authority and European Centre for Disease Prevention and Control (EFSA and
539 ECDC).
540 Farajzadeh Sheikh, A., Goodarzi, H., Yadyad, M. J., Aslani, S., Amin, M., Jomehzadeh, N.,
541 Ranjbar, R., Moradzadeh, M., Azarpira, S., Akhoond, M. R., & Hashemzadeh, M. (2019).

542 Virulence-associated genes and drug susceptibility patterns of uropathogenic *Escherichia*
543 *coli* isolated from patients with urinary tract infection. *Infection Drug Resistance,* 12, 2039-
544 2047.
545 Gordon, D.M., & Cowling, A. (2003). The distribution and genetic structure of *Escherichia coli* in
546 Australian vertebrates: host and geographic effects. *Microbiology*, 149, 3575-3586.
547 Grevskott, D. H., C. S. Svanevik, M. Sunde, A. L. Wester, and B. T. Lunestad. (2017). Marine
548 bivalve mollusks as possible indicators of multidrug-resistant *Escherichia coli* and other
549 species of the Enterobacteriaceae family. *Frontiers Microbiology,* 18(8), 8-24.
550 Henriques, I. S., Fonseca, F., Alves, A., Saavedra, M. J., & Correia, A. (2006). Occurrence and
551 diversity of integrons and β-lactamase genes among ampicillin-resistant isolates from
552 estuarine waters. *Research Microbiology,* 157, 938–947.
553 ISO. (2005). *ISO/TS 16649-3:2005. Microbiology of food and animal feeding stuffs - Horizontal*
554 *method for the enumeration of beta-glucuronidase-positive Escherichia coli - Part 3: Most*
555 *probable number technique using 5-bromo-4-chloro-3-indolyl- beta-D-glucuronide*. Geneva,
556 Switzerland: International Organization for Standardization (ISO).
557 Iwamoto, M., Ayers, T., Mahon, B. E, & Swerdlow, D. L. (2010). Epidemiology of seafood-
558 associated infections in the United States. *Clinical Microbiolical Review,* 23, 339-410.
559 Johnson, J. R., Johnston, B. D., Delavari, P., Thuras, P., Clabots, C., & Sadowsky, M. J. (2017).
560 Phylogenetic backgrounds and virulenceassociated traits of *Escherichia coli* isolates from
561 surface waters and diverse animals in Minnesota and Wisconsin. *Applied Environmental*
562 *Microbiology,* 1, 83(24).
563 Kumaran, S., Deivasigamani, B., Alagappan, K., Sakthivel, M., & Karthikeyan, R. (2010).
564 Antibiotic resistant *Esherichia coli* strains from seafood and its susceptibility to seaweed
565 extracts. *Asian Pacific Journal of Tropical Medicine,* 3, 977–981.
566 Logue, C. M., Wannemuehler, Y., Nicholson, B. A., Doetkott, C., Barbieri, N. L., & Nolan, L. K.
567 (2017). Comparative analysis of phylogenetic assignment of human and avian ExPEC and

568 fecal commensal *Escherichia coli* using the (previous and revised) clermont phylogenetic
569 typing methods and its impact on avian pathogenic *Escherichia coli* (APEC) classification.
570 *Frontiers Microbiology*, 8, 283.
571 Luna, G. M., Vignaroli, C., Rinaldi, C., Pusceddu, A., Nicoletti, L., Gabellini, M., Danovaro, R., &
572 Biavasco, F. (2010). Extraintestinal *Escherichia coli* carrying virulence genes in coastal
573 marine sediments. *Applied Environmental Microbiology*, 76, 5659–5668.
574 Metz, M., Sheehan, J., & Feng, P. C. H. (2020). Use of indicator bacteria for monitoring sanitary
575 quality of raw milk cheeses – A literature review. *Food Microbiology*, 85, 103283.
576 Nanayakkara, B. S., O'Brien, C. L., & Gordon, D. M. (2019). Diversity and distribution of
577 *Klebsiella* capsules in *Escherichia coli*. *Environmental Microbiology Reports*, 11, 107–117.
578 NACMCF. (1992). Microbiological criteria for raw molluscan shellfish. *Journal of Food*
579 *Protection*, 55, 463-480. (National Advisory Committee on Microbiological Criteria for
580 Foods).
581 Novoslavskij, A., Terentjeva, M., Eizenberga, I., Valcin̦a, O., Bartkevics, V., & Be"rzin̦, s, A.
582 (2016). Major foodborne pathogens in fish and fish products: a review. *Annals of*
583 *Microbiol*ogy, 66(1), 1-15.
584 Rene, S. H., Vieira, A. R., Karlsmose, S., Lo Fo Wong, D. M.A., Jensen, A. B., Wegener, H. C., &
585 Aarestrup, F. M. (2011). Global monitoring of *Salmonella* serovar distribution from the
586 world health organization global foodborne infections network country data bank: results of
587 quality assured laboratories from 2001 to 2007. *Foodborne Pathogones and Disease*, 8(8),
588 887-900.
589 Roschanski, N., Guenther, S., Vu, T. T. T., Fischer, J., Semmler, T., Huehn, S., Alter, T., &
590 Roesler, U. (2017). VIM-1 carbapenemase-producing *Escherichia coli* isolated from retail
591 seafood, Germany 2016. *Eurosurveillance*, 22, 43.
592 Rubini, S., Galletti, G., D'Incau, M., Govoni, G., Boschetti, L., Berardelli, C., Barbieri, S.,
593 Merialdi, G., Formaglio, A., Guidi, E., Bergamini, M., Piva, S., Serraino, A., & Giacometti,

F. (2018). Occurrence of *Salmonella enterica* subsp. *enterica* in bivalve molluscs and associations with *Escherichia coli* in molluscs and faecal coliforms in seawater. *Food Control*, 84, 429–435.

Ryu, S. H., Park, S. G., Choi, S. M., Hwang, Y. O., Ham, H. J., Kim, S. U., Lee, Y. K., Kim, M. S., Park, G. Y., Kim, K. S., & Chae, Y. Z. (2012). Antimicrobial resistance and resistance genes in *Escherichia coli* strains isolated from commercial fish and seafood. *International Journal of Food Microbiology*, 152, 14–18.

Schwarz, S., Silley, P., Simjee, S., Woodford, N., van Duijkeren, E., Johnson, A. P., & Gaastra, W. (2010). Editorial: Assessing the antimicrobial susceptibility of bacteria obtained from animals. *Journal of Antimicrobial Chemotherapy*, 65, 601-604.

Szmolka, A., & Nagy, B. (2013). Multidrug resistant commensal *Escherichia coli* in animals and its impact for public health. *Frontiers Microbiology*, 4, 258.

Taylor, N. G. H., Verner-Jeffreys, D. W., & Baker-Austin, C.. (2011). Aquatic systems: maintaining, mixing and mobilising antimicrobial resistance? *Trends in Ecology and Evolution*, 26, 278–284.

Tenaillon, O., Skurnik, D., Picard, B., & Denamur, E. (2010). The population genetics of commensal *Escherichia coli. Nature Reviews Microbiology*, 8(3), 207-217.

Tomazi, T., Coura, F. M., Gonçalves, J. L., Heinemann, M. B., & Santos, M. V. (2018). Antimicrobial susceptibility patterns of *Escherichia coli p*hylogenetic groups isolated from bovine clinical mastitis. *Journal of Dairy Science*, 101, 9406–9418.

Van, T. T. H., Chin, J., Chapman, T., Tran, L. T., & Coloe, P. J. (2008). Safety of raw meat and shellfish in Vietnam: An analysis of *Escherichia coli* isolations for antibiotic resistance and virulence genes. *International Journal of Food Microbiology*, 124, 217–223.

Vignaroli, C., Di Sante, L., Leoni, F., Chierichetti, S., Ottaviani, D., Citterio, B., & Biavasco, F. (2016). Multidrug-resistant and epidemic clones of *Escherichia coli* from natural beds of Venus clam. *Food Microbiology*, 59, 1–6.

620 Vignaroli, C., Luna, G. M., Rinaldi, C., Di Cesare, A., Danovaro, R., & Biavasco, F. (2012). New
621 sequence types and multidrug resistance among pathogenic *Escherichia coli* isolates from
622 coastal marine sediments. *Applied Environmental Microbioogy*, 78, 3916–3922.
623 Vignaroli, C., Di Sante, L., Magi, G., Luna, G. M., Di Cesare, A., Pasquaroli, S., Facinelli, B., &
624 Biavasco, F. (2015). Adhesion of marine cryptic *Escherichia* isolates to human intestinal
625 epithelial cells. *The ISME Journal*, 9, 508–515.
626 Vila, J., Sáez-López, E., Johnson, J. R., Römling, U., Dobrindt, U., Cantón, R., Giske, C. G., Naas,
627 T., Carattoli, A., Martínez-Medina, M., Bosch, J., Retamar, P., Rodríguez-Banõ, J., Baquero,
628 F., & Soto, S. M. (2016). *Escherichia coli*: An old friend with new tidings. *FEMS*
629 *Microbiology Reviews*, 40(4), 437-463.
630 Wang, F., Jiang, L., Yang, Q., Han, F., Chen, S., Pu, S., Vance, A., & Ge, B. (2011). Prevalence
631 and antimicrobial susceptibility of major foodborne pathogens in imported seafood. *Journal*
632 *of Food Protection*, 74, 1451–1461.
633 Williams, M. R., Stedtfeld, R. D., Guo, X., & Hashsham, S. A. (2016). Antimicrobial resistance in
634 the environment. *Water Environment Research*, 88, 1951–1967.
635 WHO. (2014). Antimicrobial resistance: global report on surveillance. World Health Organization
636 ISBN 978 92 4 156474 8. https://www.who.int/antimicrobial-
637 resistance/publications/surveillancereport/en/.
638

Highlights

- Bivalve molluscs are confirmed as efficient tool for antimicrobial resistance monitoring
- High multi-resistant levels were observed in *Salmonella* spp. and *E. coli* from molluscs and water
- Resistance levels for *Salmonella* are influenced by the investigated serovars
- Spatial trends of MDR *Salmonella* strains are observed

Figure 1

Figure 1. Scheme and classification of the five sub-areas in which bivalve molluscs and water samples were collected in the Ferrara area, Italy, from 2001 to 2018.

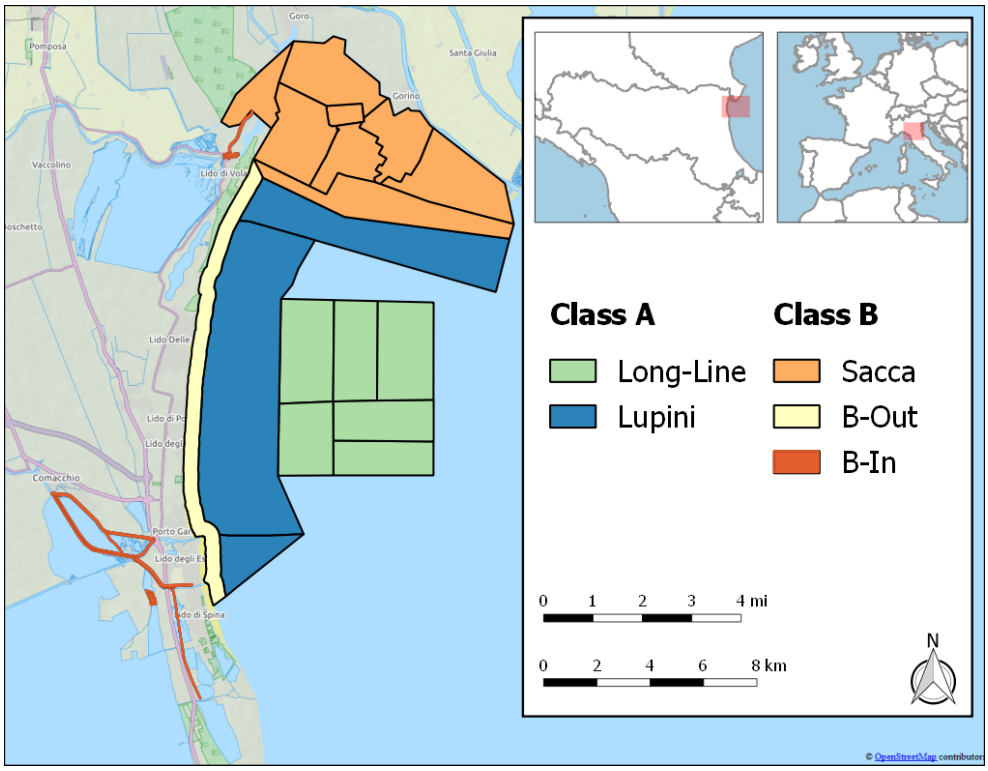

Table 1. List of tested antimicrobials and resistant breakpoints used.

Antimicrobials	Abbreviations	Disk content (µg)	Resistance breakpoint (zone diameter mm)	
			Salmonella enterica subsp. *enterica*	*Escherichia coli*
Ampicillin	AMP	10	≤17	≤13
Ceftazidime	CEFTZ	10	≤19	≤21
Ceftiofur	CEFT	30	≤17	≤17
Chloramphenicol	CHL	30	≤18	≤16
Enrofloxacin	ENR	5	≤16	≤16
Gentamicin	GENT	10	≤15	≤15
Meropenem	MER	10	≤26	≤24
Nalidixic Acid	NA	30	≤13	≤13
Streptomycin	ST	10	≤11	≤11
Sulfisoxazole	SULFAX	250	n.p.	≤12
Sulphonamides	SULFA	250	≤12	n.p.
Tetracycline	TETRA	30	≤16	≤11
Trimethoprim-sulfamethoxazole	T-SULFA	25	≤10	≤15

n.p. not performed; EUCAST breakpoints were chosen for all antimicrobials agents except for ceftiofur, enrofloxacin, nalidixc acid, streptpmycin for both *Salmonella enterica* subsp. *enterica* and *Escherichia coli,* as well as for sulfonamides for *Salmonella enterica* subsp. *enterica* and sulfisoxazole and tetracycline for *Escherichia coli,* for which CLSI breakpoints were used.

Table 2. Results of susceptibility testing to antimicrobial agents of *Salmonella* spp. isolates collected in different areas of mollusk production in the province of Ferrara.

Area	No. of isolates	No. of resistant isolates (%)													
		PEN	CEP III		CARB	QUIN		AMIN		PHEN	TETRA	SULFA	POT SULFA	R at least one AA	MDR
		AMP	CEFTZ	CEFT	MER	NA	ENR	GENT	ST	CHL	TETRA	SULFA	T-SULFA		
B-in	61	37(60.7)	-	7(11.5)	13(21.3)	2(3.8)	1(1.6)	2(1.9)	41(67.2)	11(18)	28(45.9)	20(32.8)	2(3.3)	53(86.9)	31(50.8)
Long-line, Lupini, B-Out	11	8(72.7)	-	-	1(9)	-	-	-	9(81.8)	5(45.4)	8(72.7)	4(36.4)	1(9)	9(81.8)	8(72.7)
Sacca	30	8(26.7)	-	2(6.7)	6(20)	-	-	-	10(33.3)	-	10(33.3)	4(13.3)	-	21(70)	6(20)
Total	102	53(52)	-	9(8.8)	20(19.6)	2(1.9)	1(0.9)	2(1.9)	60(58.8)	16(15.7)	46(45.1)	28(27.4)	3(2.9)	83(81.4)	45(44.1)

PEN: Penicillins; CEP III; cephalosporins III; CARB: Carbapenems; QUIN: Quinolones; AMIN: Aminoglycosides; PHEN: Phenicols; TETRA: Tetracyclines; SULFA: Sulphonamides; POT SULFA: Potentiated sulphonamides; R: resistant; AA: antimicrobial agent; MDR: multidrug resistant strains; - : not detected. The results are expressed as number of resistant isolates and their percentage value (%).

Table 3. Results of susceptibility testing to antimicrobial agents of *Salmonella* ser. Typhimurium and monophasic *S.* Typhimurium isolates

Serovar	No. of isolates	No. of resistant isolates (%)													
		PEN	CEP III		CARB	QUIN		AMIN		PHEN	TETRA	SULFA	POT SULFA	R at least one AA	MDR
		AMP	CEFTZ	CEFT	MER	NA	ENR	GENT	ST	CHL	TETRA	SULFA	T-SULFA		
*S.*Typhimurium	72	29(40.3)	-	7(9.7)	17(23.6)	1(1.4)	1(1.4)	2(2.8)	31(43.1)	15(20.8)	26(36.1)	10(13.9)	3(4.2)	54(75)	24(33.3)
monophasic *S.* Typhimurium	30	24(80)	-	2(6.7)	3(10)	1(3.33)	-	-	29(96.7)	1(3.3)	20(66.7)	18(60)	-	29(96.7)	21(70)

PEN: Penicillins; CEP III; cephalosporins III; CARB: Carbapenems; QUIN: Quinolones; AMIN: Aminoglycosides; PHEN: Phenicols; TETRA: Tetracyclines; SULFA: Sulphonamides; POT SULFA: Potentiated sulphonamides; R: resistant; AA: antimicrobial agent; MDR: multidrug resistant strains; - : not detected. The results are expressed as number of resistant isolates and their percentage value (%).

Table 4. Temporal trends of resistance to antimicrobial agents of *Salmonella* spp. isolates collected in different years of mollusk production in the province of Ferrara.

Period	No. of isolates	No. of resistant isolates (%)												
		PEN	CEP III		CARB	QUIN		AMIN		PHEN	TETRA	SULFA	POT SULFA	R at least one AA	MDR
		AMP	CEFTZ	CEFT	MER	NA	ENR	GENT	ST	CHL	TETRA	SULFA	T-SULFA		
2001-2007	17	4(23.5)	-	-	10(58.8)	-	-	-	91(52.9)	4(23.5)	4(23.5)	4(23.5)	2(3.3)	16(94.1)	4(23.5)
2008-2012	28	18(64.2)	-	2(7.1)	7(25)	-	1(1.6)	-	22(78.6)	6(28.6)	13(46.4)	8(28.6)	-	26(92.9)	16(57.1)
2013-2017	57	31(54.4)	-	7(12.8)	3(5.3)	2(3.8)	-	2(1.9)	29(50.9)	6(28.1)	29(50.9)	16(28.1)	1(9)	41(71.9)	25(43.9)
Total	102	53(52)	-	9(8.8)	20(19.6)	2(1.9)	1(0.9)	2(1.9)	60(58.8)	16(15.7)	46(45.1)	28(27.4)	3(2.9)	83(81.4)	45(44.1)

PEN: Penicillins; CEP III; cephalosporins III; CARB: Carbapenems; QUIN: Quinolones; AMIN: Aminoglycosides; PHEN: Phenicols; TETRA: Tetracyclines; SULFA: Sulphonamides; POT SULFA: Potentiated sulphonamides; R: resistant; AA: antimicrobial agent; MDR: multidrug resistant strains; - : not detected. The results are expressed as number of resistant isolates and their percentage value (%).

Table 5. Results of susceptibility testing to antimicrobial agents of *E. coli* isolates collected in different areas of mollusk production in the province of Ferrara.

Area	No. of isolates	No. of resistant isolates (%)												
		PEN	CEP III		CARB	QUIN		AMIN		PHEN	TETRA	SULFA	POT SULFA	R at least one AA	MDR
		AMP	CEFTZ	CEFT	MER	NA	ENR	GENT	ST	CHL	TETRA	SULFAX	T-SULFA		
B-in	27	17(63)	-	9(33.3)	4(14.8)	2(14.3)	2(7.4)	12(44.4)	18(66.7)	-	2(7.4)	13(48.1)	-	23(85.1%)	15(55.6%)
Long-line, Lupini, B-Out	19	8(42.1)	-	2(10.5)	1(5.2)	3(15.8)	-	7(36.8)	12(63.2)	-	3(15.8)	5(26.3)	-	14(73.7%)	6 (31.6%)
Sacca	33	19(57.6)	1(3)	7(21.2)	4(12.1)	3(9.1)	2(6.1)	9(27.3)	11(33.3)	1(3)	2(6.1)	6(18.2)	2(6.1)	22(66.7%)	9(27.3%)
Total	79	44(55.7)	1(1.3)	18(22.8)	9(11.4)	8(10.1)	4(5.06)	28(35.4)	41(51.9)	1(1.3)	7(8.9)	24(30.4)	2(2.5)	59(74.7)	30(38)

PEN: Penicillins; CEP III; cephalosporins III; CARB: Carbapenems; QUIN: Quinolones; AMIN: Aminoglycosides; PHEN: Phenicols; TETRA: Tetracyclines; SULFA: Sulphonamides; POT SULFA: Potentiated sulphonamides; R: resistant; AA: antimicrobial agent; MDR: multidrug resistant strains; - : not detected. The results are expressed as number of resistant isolates and their percentage value (%).

Table 6. Results of susceptibility testing to antimicrobial agents in relation to the inclusion of *E. coli* isolates in more and less frequent phylogenetic groups and the sampling area in the mollusk production area in the province of Ferrara; findings were expressed as number of resistant isolates and their percentage value (%).

	Phylogenetic group							
	A		**B1**		**C**		**B2, D, E and unknown**	
Area	R at least one AA	MDR	R at least one AA	MDR	R at least one AA	MDR	R at least one AA	MDR
B-in	7(100)	5(71.4)	9(81.8)	7(63.6)	4(100)	2(50)	3(60)	1(20)
Long-line, Lupini, B-Out	1(100)	-	6(75)	2(25)	2(66.7)	2(66.7)	5(71.4)	2(28.6)
Sacca	3(33)	-	13(61.9)	3(14.3)	4(100)	4(100)	4(80)	2(40)
Total	11(81.8)	5(45.4)	28(70)	12(30)	10(90.91)	8(72.7)	12(70.6)	5(29.4)

R: resistant; AA: antimicrobial agent; MDR: multidrug resistant strains

Table 7. Spatial trends of resistance to at least one antimicrobial agent and multi-resistant strains expressed as number of resistant isolates and their percentage value (%).

	***Salmonella* ser. Typhimurium**		**monophasic *S.* Typhimurium serovar**		***Salmonella* spp.**		***Escherichia coli***	
Area	R to at least one AA	MDR	R to at least one AA	MDR	R to at least one AA	MDR	R to at least one AA	MDR
B-In	33(80.5%)	16(39%)*	20(100%)	15(75%)	53(86.9%)	31(50.8%)*	23(85.1%)	15(55.6%)
Long-line, Lupini, B-Out	7(77.8%)	6(66.7%)*	2(100%)	2(100%)	9(81.8%)	8(72.7%)*	14 (73.7%)	6 (31.6%)
Sacca	14(63.6%)	2(9.1%)*	7(87.5%)	4(50%)	21(70%)	6(20%)*	22(66.7%)	9(27.3%)

R: resistant; AA: antimicrobial agent; MDR: multidrug resistant strains. Number of isolates in columns bearing * are significantly different ($p < 0.05$).