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Compost improves plant and soil macronutrient content in a 14-years orchard

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Baldi, E., Mazzon, M., Cavani, L., Quartieri, M., Toselli, M., Marzadori, C. (2023). Compost improves plant and soil macronutrient content in a 14-years orchard. NUTRIENT CYCLING IN AGROECOSYSTEMS, 125, 425-435 [10.1007/s10705-023-10258-0].

Availability: This version is available at: https://hdl.handle.net/11585/923184 since: 2023-04-17

Published:

DOI: http://doi.org/10.1007/s10705-023-10258-0

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This is the peer-reviewed accepted manuscript of: Baldi, E., Mazzon, M., Cavani, L. et al. *Compost improves plant and soil macronutrient content in a 14-years orchard*. *Nutr Cycl Agroecosyst* 125, 425–435 (2023). The final published version is available online at: <u>https://doi.org/10.1007/s10705-023-10258-0</u>

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1	The fate of generally unconsidered macronutrients in plant and soil: effect of different
2	fertilization strategy on K, Ca, Mg and S
3	Compost improves plant and soil macronutrient content in a 14-years orchard
4	
5	
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8	
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11	
12	Abstract
13	Adequate plant nutritional status and soil fertility preservation should can be achieved through the
14	employment of sustainable agricultural management techniques. The challenge of intensive orchard
15	cultivation, besides the risk of nutrient decline, is the to prevention of the loss of soil fertility that
16	could lead to soil degradation with a consequent negative impact on yield and fruit quality. The use
17	of organic amendments could be a sustainable strategy to combine elevated high plant performance
18	with soil fertility improvement.
19	This work aims at shedding light on the effects of compost addition with respect to an unfertilized
20	control and a mineral fertilization treatment on macronutrient (K, Ca, Mg, and S) dynamics in plants
21	and soil of a commercial nectarine orchard planted in 2001. Therefore, the manuscript aims at
22	shading light on macronutrient (K, Ca, Mg and S) dynamics in plants and soil of a commercial
23	nectarine orchard planted in 2001 and in which three fertilization treatments were compared:
24	unfertilized control, mineral fertilization, and compost.

25	In the first 0.15 m of soil, compost addition resulted in higher values (26 - 42%) of all the parameters.
26	Both fertilization treatments induced a 28% increase of in roots' S content compared to the control
27	but did not induce macronutrients content variation in plant skeleton, pruned wood, and thinned
28	fruits. In autumn leaves, all the macronutrients resulted <u>in</u> higher <u>values</u> (24 - 45%) with both mineral
29	and compost fertilization, and the same was observed in fruit at harvest (increases of 15 - 31%).
30	Finally, in the first 0.15 m of soil, compost addition resulted with the higher values (+ 26 42%) of all
31	the parameters.
32	In our study, the treatment with compost satisfied plants' nutrient demands as much as the mineral
33	fertilizer. In addition, compost treatment also improved soil nutrient content while preserving yield.
34	Our results show that it is possible to reconcile plant nutrient needs with the preservation of soil
35	fertility with the aim of reaching agriculture sustainability.
36	Thus, the long term use of mineral fertilizer, even if meeting plant's nutrient demand, could lead to
37	soil macronutrient depletion. Whereas, organic amendments addition could not only satisfy plant's
38	nutrient demand but also maintain soil quality and fertility through the higher macronutrients
39	storage in the soil; this would lead to both higher plant performances and to economic and
40	environmental benefits.
41	

42 Keywords

Prunus persica, soil nutrient availability, compost, mineral fertilization, nutrient removal, soil
macronutrient content.

45 1. Introduction

46 The rising increasing demand for high-quality nutritional food related to the constant steady 47 increase of in population (El-Jendoubi et al., 2013) is leading to the necessity to maximise maximize yield by while minimizing ecosystem impacts deriving from agriculture. According to the report of 48 49 the Food and Agriculture Organization of the United Nations (FAO, 2017), the requirements for adequate food supplies have to pass through sustainable agricultural management techniques. This 50 issue also deals with adequate plant nutritional status and preservation/increase of soil fertility 51 52 (Toselli et al., 2019a; Zhang et al., 2020). Plant nutrition depends on is the result of the nutrient cycle 53 in and out of the orchard ecosystem. The inputs to the soil, for example, include mineral and/or organic fertilizers, atmospheric deposition, and biological nitrogen (N) fixation, while the outputs 54 55 are represented by harvested fruits, nutrients lost by leaching, gaseous losses, and erosion (Toselli 56 et al., 2019b). In case of a negative balance between inputs and outputs, the soil would come across 57 a nutrient depletion that, in the long_-term, would lead to an unsustainable farming system (El-58 Jendoubi et al., 2013). The challenge of intensive orchard cultivation is, besides the risk of nutrient 59 decline, the loss of soil fertility (both chemical and biological) that could lead to soil degradation (i.e. 60 loss of soil organic matter, erosion, acidification, and pollution) with a consequent negative impact on plant performances (Zhang et al., 2020). Thus, the great challenge for modern farmers is to 61 maintain and/or increase soil fertility in a sustainable way. This could be reached through the 62 63 application of organic amendments (i.e. composts, biochar, and manures) that are widely recognised 64 recognized to be cheapinexpensive, to release slowly release the nutrients through mineralization 65 slowly, and to enhance soil carbon (C) and organic matter increase (Mazzon et al., 2018; Sciubba et 66 al., 2015) thus stimulating carbon dioxide sequestration. In addition, the application of organic 67 matter is able tocan improve soil physical properties (Chatzistathis et al., 2020) as, for example, bulk 68 density reduction and aggregate stability and water holding capacity increase (Adugna, 2016),

69 positively affecting soil microbial community (Fawzi et al., 2010; Safaei Khorram et al., 2019) and 70 increasing macro- and micro-nutrient availability (Baldi et al., 2021a; Fawzi et al., 2010). Organic amendments thus represent not only a source of N, phosphorus (P), and potassium (K) but also of 71 72 calcium (Ca), magnesium (Mg), and sulphur (S) that which are equally considered as essential plant 73 macronutrients (Barreto et al., 2021; Shiwakoti et al., 2020). In the study of Shiwakoti et al. (2020), 74 the farmyard manure long-term (approximatively 64 years) addition of farmyard manure (at the rate of 11.2 Mg ha⁻¹ year⁻¹) evidenced higher macronutrient (K, S, and Mg) content in soil than the 75 76 other treatments (pea vine and wheat residues, with and without N addition) mainly due to the high 77 amounts of these nutrients that directly contribute to soil chemical fertility. Shiwakoti et al. (2020) also highlighted that manure could have activated soil cation exchange sites releasing organic 78 79 colloids and consequently adsorbing K to the exchangeable sites and increasing its availability.

Macronutrient soil availability throughout the growing season is fundamental for fruit trees. In a 80 81 study on pomegranate nutrient dynamics, Maity et al. (2019) demonstrated that most of plant needs 82 were <u>mostly</u> satisfied by uptake from <u>the</u> soil more than <u>from the</u> mobilization from plant reserves; as a consequence, if soil nutrients are not replenished through appropriate fertilization management, 83 84 fruit yield and quality could be severely impaired. In a different study on various fruit trees (i.e. apple, peach, and mandarin), Cruz et al. (2019) showed that adequate K supply at fruit set is of 85 fundamental importance for the final fruit quality. Maity et al. (2019) evidenced that a great amount 86 87 of Mg was remobilized from leaves to fruit at the maturity stage, while S was mainly concentrated 88 in shoots and Ca in the woody organs of pomegranate. El-Jendoubi et al. (2013) found that K, P, and N mainly accumulate in fruits, while Mg and Ca in abscised leaves. Moreover, it is estimated that 89 90 peach trees in commercial orchards have a macronutrient requirement accounting for 74 - 425 g K 91 tree⁻¹, 25 - 518 g Ca tree⁻¹, and 9 - 74 g Mg tree⁻¹ (Baldi et al., 2021b) every year.

92 The key issue for fruit tree nutrition is the availability of nutrients in the soil during the entire 93 vegetative season, consequently, the use of organic amendments, that gradually release nutrients 94 through mineralization, could be a sustainable strategy able to combine elevated plant performance 95 with the improvement of soil fertility (Baldi et al., 2021b).

This manuscript follows 3-<u>three</u> previous publications on C (Baldi et al., 2018), N (Toselli et al., 2019b), and micronutrients <u>on-in</u> the same experiment (Baldi et al., 2021a), and aims at <u>shading</u> shedding light on <u>other nutrients such a the macronutrients</u> K, Ca, Mg, and S. Indeed, the goal of the present study was to determine the effects of <u>the long term</u> mineral fertilization and compost addition in <u>the long terma nectarine orchard</u> (14 years) on: i) soil macronutrient content at the end of the 14-years life-time of the <u>nectarine</u> orchard, ii) macronutrient content in different plant organs, and iii) the relation between plant and soil macronutrient content.

103

104 2. Materials and methods

105 *2.1 Orchard description and treatments*

106 The experiment was carried out on a commercial nectarine orchard [Prunus persica, Batsch var. 107 nucipersica (Bockh.) Schn.] planted in 2001 (Table 1). The orchard was located in the Po valley (Italy) near Ravenna (44°27' N;12°13' E), an area characterized by a temperate climate and a silt-loam soil 108 109 (Calcaric Cambisol) with a total carbonate content (% CaCO₃) of 31 ± 1 and an active carbonate 110 content (% CaCO₃) of 13 ± 1 (Baldi et al., 2021a, 2018). The pPlanting layout and main orchard management strategies are reported in Table 1. Since orchard plantation, three fertilization 111 112 treatments were compared with four replicates (4 trees each) according to a complete randomized block design: unfertilized control (CK); mineral fertilization (MIN); and compost (COM). Specific 113 114 information regarding the treatments are is reported in Table 1. Fertilizers were applied on to the tree row and tilled into the soil to a depth of 0.25 m, while pruned wood was left into the ground

116 and chopped (Baldi et al., 2021*a*, 2018; Toselli et al., 2019b).

- 117
- **Table 1** Nectarine orchard main characteristics and management strategies (Baldi et al., 2021a, 2018; Toselli
 et al., 2019b).

Management	Description
Variety and rootstock	Stark RedGold, grafted on hybrid GF677 (<i>P. persica</i> × <i>P. dulcis</i>)
Training system	Delayed vasette, distance of 5 m between the rows and 3.8 m between trees along
	the row
Irrigation	Drip irrigation system from June to September (vegetation season).
Phytosanitary treatments	Done aAccording to Crop Management Guideline of the region Emilia-Roma-
	gna
	(www.regione.emilia-romagna.it).
Soil management	Tree row was tilled superficially for weed control.
	Alleys were covered with spontaneous grass managed by mowing it 3 times a
	year.
Fertilization treatments	Mineral fertilization (MIN)
	Nitrogen (N) applied every year at the rate of 70 kg ha ⁻¹ yr ⁻¹ (60% in May and
	40% in September); in 2004, N rate was increased to 120 kg ha^{-1} yr^{-1} and from
	2006 to 130 kg ha ⁻¹ yr ⁻¹ . <u>Nitrogen was applied as ammonium nitrate (N = 35 %)</u> ,
	while P and K were applied as a binary fertilizer (P = 10 %; K = 20 %); no micro-
	nutrients was provided in mineral fertilization.
	Phosphorus (P) at 100 and potassium (K) at 200 kg ha ⁻¹ (applied only at plant-
	ing). Rate established according to Integrated Crop Management Guideline of
	the region Emilia-Romagna (<u>www.regione.emilia-romagna.it</u>).
	Nitrogen was applied as ammonium nitrate (N = 35%), while P and K were ap-
	plied as a binary fertilizer (P = 10%; K = 20%); no micronutrients was provided
	with mineral fertilization.

Compost (COM)

Applied at a rate of 10 t DW ha⁻¹ yr⁻¹, equal to 240 kg N ha⁻¹ yr⁻¹.

Compost was obtained from domestic organic wastes (50%) mixed with pruning material (50%) after a 3-month stabilization. Average characteristics: DW of 73%, pH 9, EC 2.96 mS cm⁻¹, C/N ratio 10.2, and (in g kg_{DW}⁻¹) organic C 234, N 21.1, P 4.8, K 15.2.

120

121 2.2 Plant sampling and analysis

122 In 2014, after 14 years of life, 4 trees per treatment were harvested and divided into organs -as 123 described by Baldi et al. (2021a, 2018) and Toselli et al. (2019b). Briefly, thinned fruits were collected 124 in spring and weighted, and a representative sample was-then oven-dried and milled (2 mm). In July, a sample of 40 young fully expanded leaves was collected from the apical part of shoots, and 125 126 the leaf area was measured by a portable area meter (Li-3000, LiCor inc., Lincoln, Nebraska). Leaves 127 were then washed, oven-dried, and milled at 2 mm. At harvest, in August, plant yield was recorded; 128 afterwards, on a representative sample of fruits, fresh weight (FW) and dry weight (DW) of flesh 129 and kernel were measured on a representative sample of fruits; dried flesh and kernel were weighted 130 and milled. In September, one tree per plot was enclosed into a plastic net to collect autumn leaves 131 that were weighted, leaf area measured, dried, and milled. In December 2014, at the end of the 132 commercial life of the orchard, the same trees were harvested, divided into roots, trunks, branches 133 $(age > 2 year_{\underline{s}})_{t}$ and current year shoots $(twigs)_{t}$ and weighted. A subsample of each organ was oven-134 dried, weighted, and milled. The sample of rRoots was were washed with deionized water to 135 remove soil residues.

A sample (0.3 g) of each plant organ was mineralized according to the US EPA Method 3052 (Kingston and Jassie, 1988) in an Ethos TC microwave (Milestone, Bergamo, Italy), filtered (Whatman 42®), and analyzsed for Ca, K, Mg, and S by Inductively inductively Coupled coupled

139 Plasma plasma Optical optical Emission emission Spectrometer spectrometer (ICP-OES; Ametek 140 Spectro, Arcos, Kleve, Germany). Blank and certified reference materials (NIST standard reference 141 material SRM_1573a tomato leaves and SRM_1570a spinach leaves) analyses were performed. Relative uncertainty, calculated as the relative deviation of the measured element concentration to 142 143 its certified value, was typically better than ± 5%. 144 Mineral The mineral content in different parts of the plant (leaves, fruits, branches, trunk, and roots) was calculated by multiplying each mineral concentration by the DW of the specific organ. 145 146 The biomass of the skeleton was calculated as the sum (without pruning wood) of trunk, branch,

and twigs > 2 years (identified according to their insertion into branches) measured at the end of the
experiment. Skeleton and root nutrient content were divided by the age of the orchard (13) assuming
a constant annual increase of weight and nutrient accumulation, and considering the increase of the
first year (2001) was negligible since orchard plantation was done planted at the end of the year.

151

152 2.3 Soil sampling and analysis

In December 2014, an 80 cm deep soil core (70 mm diameter) was collected in the row of each plot 153 154 with a soil column cylinder auger that was inserted into the soil using a tractor. The core was carefully removed from the auger and divided into four parts according to depth: 0 - 0.15 m, 0.16 -155 156 0.25 m, 0.26 - 0.45 m and 0.46 - 0.65 m. Soil from each depth was separately weighted and oven dried 157 at 105°C for 24 hours to evaluate soil bulk density (BD) that which was calculated as the ratio between DW and the volume of each core. In addition, soil samples were collected, always in the 158 159 row, with an auger at 0-0.15 m, 0.16-0.25 m, 0.26-0.45 m, and 0.46-0.65 m of depth. Each sample (made of 4 sub-samples) was sieved (2 mm), cleaned from roots and visible plant residues, and air-160 161 dried or stored at 4°C. A sub-sub-sample was then used to evaluate soil total concentration of Ca, K, 162 Mg_{μ} and S. Briefly, samples were subjected to wet mineralization by treating 0.5 g of dry sample 163 with 6 mL of hydrochloric acid (37%), 2 mL of nitric acid (65%) and 2 mL of hydrogen peroxide 164 (30%) in an Ethos TC microwave lab station (Milestone, Bergamo, Italy) according to the methods ISO 12914:2012 and 22036:2008. Solutions were filtered (Whatman 42®) and the element 165 concentration was determined by plasma spectrometer (ICP-OES; Ametek Spectro, Arcos, Kleve, 166 167 Germany). Blank and certified reference material (BCR reference material No 141R calcareous loam soil) analyses were performed; relative uncertainty, calculated as the relative deviation of the 168 169 measured element concentration to its certified value, was typically better than ± 5%. Soil pseudo-170 total mineral element content at the end of the orchard life-time was calculated by multiplying the 171 nutrient concentration by the soil bulk density at the respective depth intervals. Electrical conductivity (EC) was measured on a suspension of 10 g of fresh sample and 20 mL of deionised 172 water that was stirred for 120 min at 25°C and filtered before measurement. Soil potential cation 173 174 exchange capacity (CEC) was estimated using the ammonium acetate method (Sumner and Miller, 175 1996).

- 176
- 177 2.4 Statistical analysis

178 After assumption verification (Shapiro-Wilk for normality and Bartlett for homogeneity of variance), 179 <u>p</u>Plant organ data were <u>analysed analyzed</u> with a one-way ANOVA with treatment as <u>a</u> factor (three 180 levels: unfertilized control, mineral fertilization, and compost) after assumption verification 181 (Shapiro Wilk for normality and Bartlett for homogeneity of variance). Whether necessary, data were transformed using the Box-Cox procedure to fit the ANOVA assumption. Similarly, data on 182 183 macronutrient content forof autumn and summer leaves macronutrient content were analysed with 184 a twoone-way ANOVA with treatment and season as <u>a</u> factors. When significant differences occur 185 $(P \leq 0.05)$, an HSD post-hoc test (Tukey's test) was applied to separate the means. A Principal 186 Component Analysis (PCA) was carried out using plant organ data that showeding significant differences <u>for-in</u> the treatment<u>ss</u>. Soil data were <u>analysed-analyzed</u> using a split-plot design, with
treatments (three levels: unfertilized control, mineral fertilization, and compost) as the main factor
and sampling depths (four levels: 0 - 0.15, 0.16 - 0.25, 0.26 - 0.45, 0.46 - 0.65 m) as the sub-factor.
ANOVA assumption verification and means separation were <u>done asperformed as described</u> -for
plant organs (R Core Team, 2021).

192

193 3. Results

194 Fertilization treatments, no matter which one, induced on average a 28% increase of in roots' S 195 content compared to the control (Table 2). The fertilization treatments did not induce <u>a</u> variation of 196 Ca, K, Mg, and S content in <u>the</u> plant skeleton, pruned wood, and thinned fruits (Table 2). On the 197 contrary, all the considered macronutrients considered (Ca, K, Mg, and S) were increased in autumn 198 leaves (in a range from 24 to 45%) by both mineral and compost fertilizations. Similar results in 199 correspondence of the two fertilization treatments (with increases between 15 and 31%) was were 200 observed in fruit at harvest with the exception of Ca content that did not show any differences 201 among treatments (Table 2).

202

203Table 2 – Macronutrient content (g pt⁻¹) in plant organs ± standard error (based on field replicates). Different204lowercase letters indicate significant (P < 0.05) differences between treatments (CK = unfertilized control, MIN205= mineral fertilization, COM = compost).

Plant organs	Treatments	Ca	К	Mg	S
	CK	26 ± 1	9.9 ± 0.7	2.2 ± 0.2	1.4 ± 0.1
Duran o di succio di	MIN	43 ± 1	17 ± 1	3.5 ± 0.3	2.5 ± 0.1
Pruned wood	COM	30 ± 7	13 ± 3	2.6 ± 0.6	1.9 ± 0.4
	p.value	0.150	0.098	0.178	0.097
	СК	0.62 ± 0.14	2.6 ± 0.6	0.29 ± 0.07	0.27 ± 0.06
Thinned fruits	MIN	0.44 ± 0.12	2.0 ± 0.6	0.22 ± 0.06	0.21 ± 0.06
	COM	0.87 ± 0.17	3.5 ± 0.7	0.38 ± 0.07	0.36 ± 0.07

	p.value	0.168	0.257	0.259	0.251
	CK	2.3 ± 0.2	43 ± 1 ^b	3.6 ± 0.1 b	2.0 ± 0.1 b
Fruit at harvest	MIN	2.4 ± 0.1	51 ± 2 ^{ab}	4.8 ± 0.3 a	2.8 ± 0.1 a
Truit at haivest	COM	2.4 ± 0.2	57 ± 1 a	4.8 ± 0.2 a	2.8 ± 0.1 a
	p.value	0.867	0.010	0.012	0.006
	CK	118 ± 3 b	$43 \pm 2^{\text{b}}$	14 ± 0.3 ^b	2.6 ± 0.2 b
Autumn leaves	MIN	181 ± 23 a	70 ± 7 a	21 ± 2^{a}	4.7 ± 0.5 a
Autumn leaves	COM	156 ± 9 ª	66 ± 4 ab	19 ± 2^{a}	4.5 ± 0.4 a
	p.value	0.010	0.026	0.012	0.013
	CK	21 ± 3	5.6 ± 0.2	3.0 ± 0.5	0.32 ± 0.02 b
Roots	MIN	21 ± 4	5.2 ± 0.5	3.0 ± 0.5	0.43 ± 0.03 a
Roots	COM	24 ± 0.3	6.1 ± 0.7	3.6 ± 0.2	0.45 ± 0.03 a
	p.value	0.771	0.313	0.659	0.001
		(= 0	10 1	• • • • •	
	СК	45 ± 2	12 ± 1	3.8 ± 0.3	1.8 ± 0.1
Skeleton	MIN	60 ± 7	16 ± 2	5.2 ± 0.7	2.4 ± 0.2
	COM	56 ± 7	17 ± 1	5.0 ± 0.6	2.5 ± 0.2
	p.value	0.328	0.220	0.329	0.126

No significant differences between fertilization treatments were observed in the concentration of nutrients in summer leaves (Table 3); the only exception was Mg which resulted in a 13% higher concentration in the control compared to the two fertilization strategies (Table 3).

Table 3 – Macronutrient concentration (g 100 g⁻¹ DW) ± standard error (based on field replicates) in leaves sample in summer. Different lowercase letters indicate significant (P < 0.05) differences between treatments

213	(CK = unfertilized control, MIN = mineral fertilization, COM = compost).
	· · · · · · · · · · · · · · · · · · ·

Treatments	nents Ca K Mg		S	
СК	2.2 ± 0.1	1.2 ± 0.04	0.44 ± 0.01 a	0.16 ± 0.03
MIN	1.9 ± 0.1	1.1 ± 0.01	0.39 ± 0.01 ^b	0.15 ± 0.01
COM	1.9 ± 0.1	1.2 ± 0.04	0.39 ± 0.01 ^b	0.15 ± 0.01
p.value	0.088	0.088	0.013	0.947

The comparison of macronutrients content in autumn and summer leaves highlighted the significant impact of <u>the season-phenological state</u> for all <u>the</u>-four macronutrients considered in this study (Figure 1). Specifically, higher concentrations of Ca and K were observed in autumn than in summer leaves; the opposite was observed for Mg and S.

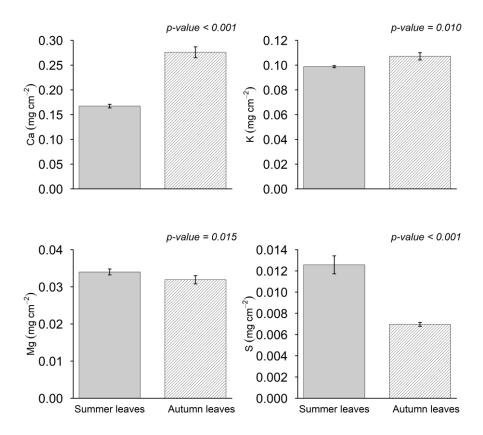


Figure 1– Means of summer and autumn leaves content of calcium (Ca), potassium (K), magnesium (Mg) and sulphur (S). Error bar represent the standard error <u>(based on four leaf samples collected in field replicates)</u> and the significant differences ($P \leq 0.05$) are reported.

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A PCA (Figure 2) was <u>done-performed using with macronutrients content indata from</u> those organs whose macronutrient content was <u>that were</u> significantly affected by the fertilization treatments (Table 2). The treatment<u>s clusterized were split</u> resulted into two main <u>cluster groups</u> (according to PC1 ANOVA results): the unfertilized control <u>in-on</u> the left side of the plot, and the two fertilization strategies (mineral and compost) <u>in-on</u> the right side of the plot. These two groups showed to be clearly defined with-by the Mg content in summer leaves that characterized the unfertilized control
group (Figure 2). A clear separation in plot space was also evident between macronutrients content
in the autumn leaves (bottom-right side of the plot) and in the fruit at harvest and roots (upper-right
side of the plot).

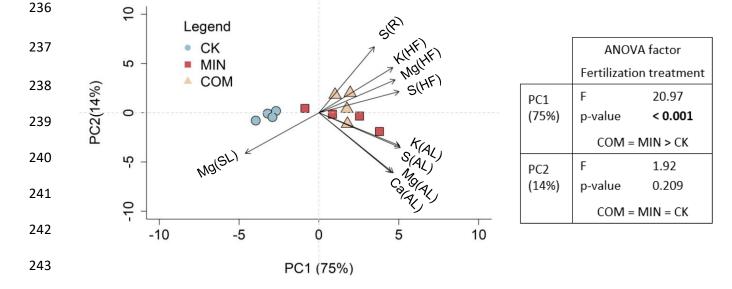


Figure 2– Principal component (PC) analysis with macronutrients content <u>(calcium (Ca), potassium (K),</u> magnesium (Mg) and sulfursulphur (S)) in the most relevant plant organs: harvested fruits (HF), roots (R), autumn leaves (AL), and summer leaves (SL). <u>TIn the table are reported reports</u> the statistical output of the ANOVA done on the PC <u>(Treatments: CK = unfertilized control, MIN = mineral fertilization, COM = compost)</u> with t-he significant differences (P < 0.05).

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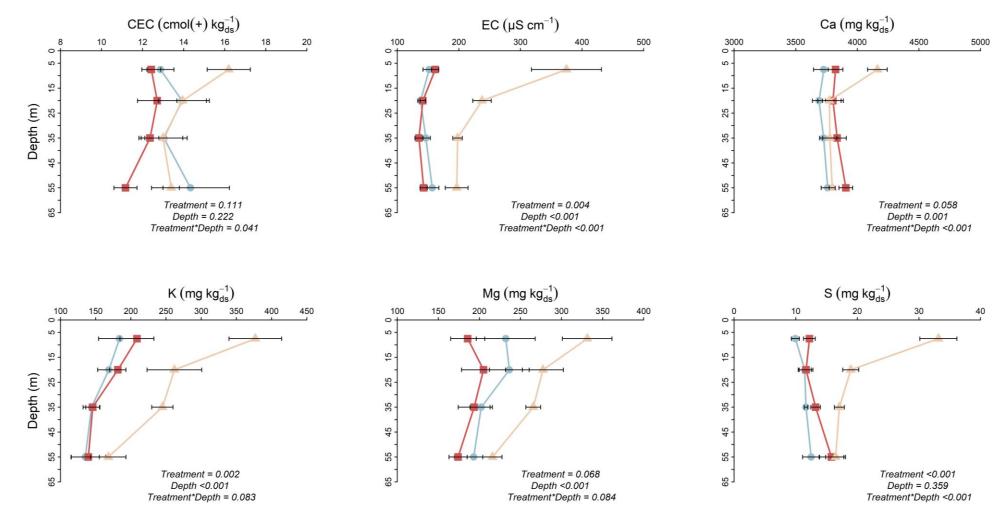
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In the first 0.15 m of soil, the addition of compost increased CEC, EC, Ca, K, Mg₂ and S compared with control and mineral fertilization (Fig. 3). With the exception of CEC and Ca, the positive effect of compost was observed also in other soil layers (Fig. 3) and in particular EC was higher in all soil

profiles, while the concentration of K, Mg and S were higher than in the control and mineral 255 256 fertilization between 0 and 45 cm of depth. The effect of compost decreased with depth for all the 257 investigated parameters investigated; however, while the concentration of K and Mg decreased 258 constantly with depths, the EC decreased until 25 cm and then was remained steady from 26 to 65 259 cm, and the concentration of Ca, S and CEC decreased until 15 cm and then was-remained steady 260 from 16 to 65 cm (Fig. 3). Unfertilized control and mineral fertilization did not show significant 261 differences in nutrient concentrations and in both treatments only a slightly decreasing trend with 262 depth was observed for K and Mg concentration. In control and mineral fertilization, S concentration 263 was higher in the deepest layer than in the superficial shallowest one (Fig. 3).



Unfertilized Control (CK)
 Mineral Fertilization (MIN)
 Compost (COM)

264

Figure 3 – Means of soil cation exchange capacity (CEC), electrical conductivity (EC), and soil calcium (Ca), potassium (K), magnesium (Mg) and sulphur (S) content at four sampling depth (0 - 0.15, 0.16 - 0.25, 0.26 - 0.45, 0.46 - 0.65 m) for the three fertilization treatments (CK = unfertilized control, MIN = mineral fertilization, COM = compost). Error bars represent the standard error (based on data on field replicates) and the significant differences (P ≤ 0.05) between "Treatment", "Depth", and "Treatment*Depth" interaction are reported.

269 4. Discussion

270 In this study and as already observed previously (El-Jendoubi et al., 2013) plants macronutrients 271 were mainly allocated in autumn leaves and in fruits at harvest. However, while in roots, skeleton, 272 pruned wood, autumn leaves, and summer leaves Ca contents were highest among all the other 273 macronutrients, K was the most important macronutrient in thinned fruit and fruit at harvest. 274 Similarly, El-Jendoubi et al. (2013) showed that each nutrient was characterized by a precise 275 allocation pattern: fruits were the largest sink for K, while Mg and Ca were mainly accumulated in 276 abscised leaves. 4.1 Calcium and potassium returned to soil with leaf abscission-abundance and allocation 277 In this study and as already observed previously (El-Jendoubi et al., 2013) plants macronutrients 278 279 were mainly allocated in autumn leaves and in fruits at harvest. However, while in roots, skeleton, 280 pruned wood, autumn leaves, and summer leaves Ca contents were highest among all the other 281 macronutrients, K was the most important macronutrient in thinned fruit and fruit at harvest. Similarly, El-Jendoubi et al. (2013) showed that each nutrient was characterized by a precise 282 283 allocation pattern: fruits were the largest sink for K, while Mg and Ca were mainly accumulated in

284 <u>abscised leaves.</u>

285 The difference between the amount of nutrients in summer leaves (sampled in summer July) and at natural abscission gives an estimation of the fraction of nutrients remobilized at the end of the 286 287 vegetative season and stored inside the woody part of the plant. In the present experiment, Ca and K showed higher concentrations in autumn leaves than in those sampled in summer indicating no 288 289 net remobilization through the season. SA similar behavior was reported in almond trees (Muhammad et al., 2015). As a consequence, Ca and K allocated in autumnto leaves returned into to 290 291 the soil after abscission and degradationdecomposition, thus becoming againreturning partially 292 available for root uptake (Baldi et al., 2021b) after mineralization-. On the other hand, little quantities

293 of Ca and K reservation are available for spring new growth. However, despite what was expected 294 and what was observed in other study studies (Dang et al., 2022; do Carmo et al., 2016), in this our 295 case, soil Ca and K concentration increased in the surface horizon and in the whole soil profile 296 respectively, only when compost was applied (CaO and K mean content of compost was were of 297 $\frac{8.25.8}{1.0} \pm 1.0$ % and $\frac{1.5 \pm 0.2}{0.2}$ %, respectively), and not in control and mineral fertilized plots, meaning that the source of additional Ca was the organic fertilizer rather than the litter made offormed by 298 abscised leaves. This The lack of effect fact that abscised leaves and/or mineral fertilizer have no such 299 300 effect on soil Ca content was probably in relation to the natural soil's high abundance in total and 301 active carbonate content (CaCO₃) (Baldi et al., 2018), indicating a large soil endowment in carbonates 302 thus making it impossible to observe variation in soil Ca content even in the long term.

303

304 *4.2 K and Mg in plant and soil<u>Compost contributed to soil and plants K and Mg content</u>*

305 Differently from Ca, the fertilization treatments (both mineral fertilizer and compost) induced an 306 increased of K and Mg content also in fruits at harvest (Table 2). This effect was also observed by 307 Increase of K content in fruits was observed also in another study on nectarine (Delian et al., (2012), 308 who reported a higher concentration of potassium than magnesium in nectarine fruits at harvestwhere at harvest macronutrients concentration in fruits followed the order K > Ca > Mg. 309 310 Potassium is not an essential element for organic molecules and it is not considered a structural 311 element (Delian et al., 2012); however, it is involved in many physiological and biochemical processes related to plant growth, crop quality, and plant response to stress factors (Delian et al., 312 313 2012; Wang et al., 2018). An excess in available Ppotassium is also known to induce a Mg-deficiency 314 in the plant due to the unidirectional competition for uptake between K and Mg for which an 315 increase in K concentration reduces Mg uptake (Xie et al., 2021). In this our study, independently 316 from the fertilization treatment (mineral or organic), the lower lowest K/Mg ratio (approx. 1.8) was

317 determined measured in plant roots with respect to the other plant organs, (approx. 1.8) thus 318 indicating that the increase in K concentration in plant organs did not inhibit plants' efficiency to 319 uptake Mg (Xie et al., 2021) was not limited by the high plant K concentrations. The apparently similar plant uptake of K and Mg is confirmed also by the content of these elements in soil samples 320 321 at the end of the commercial orchard life-time. Indeed, soil samples were not depleted in K nor Mg with values that ranged between 150 – 400 mg kg dw⁻¹, considering that optimum soil concentrations 322 lie between 240 and 300 mg kg dw⁻¹ for K (Xie et al., 2021) and between 25 and 180 mg kg dw⁻¹ for 323 Mg (Fox and Piekielek, 1984). Moreover, at the end of the experiment, significantly higher values of 324 325 both K and Mg content were measured with compost fertilization treatment (compost K2O and MgO mean concentration was of 1.4 ± 0.2 % and 0.72 43 ± 0.07 % respectively); this may indicating indicate 326 327 a its greater contribution of compost to soil nutrient content with respect to the other mineral 328 fertilization treatments. Similar results for these macronutrients were also observed in previous 329 studies (Acharya et al., 2019; do Carmo et al., 2016) and could be also related to compost chemical 330 characteristics.

331

4.3 <u>SulfurSulphur content had a different trend with compost and mineral fertilization destiny in plant roots</u>
and soil: fertilization treatment effects

Mineral and compost fertilization enhanced S content in fruit<u>s</u> at harvest and in summer leaves more than in autumn leaves. At the end of <u>the</u> nectarine orchard commercial life-time, S content in roots was still high with <u>the twoboth</u> treatments indicating a potential availability for bud <u>sprout break in</u> the next <u>vegetation vegetative</u> season. Indeed, S is a component of some secondary metabolites used for plant's physiological functions and development (Narayan et al., 2022). This <u>pP</u>lant activity production of secondary metabolites is <u>supposed to be</u> supported by soil S content which indeed resulted particularly high in correspondence of compost <u>additiontreatment</u>. Compost as an organic 341 matter supply increased increases soil organic matter, the largest reservoir of S (in organic form) into 342 soil-, and Thus compost (in our study characterized by a $0.44 \cdot 18 \pm 0.04$ % of SO₃ mean content) or soil 343 organic matter decomposition could result in organic sulfur mineralization into the SO_{4²⁻}, which is 344 available to plants (do Carmo et al., 2016; Narayan et al., 2022). However, while sSoil S-sulfur content 345 decreased with depth in correspondence of the plots treated with compost addition, while it slightly 346 increased or did not change with depth in the control and mineral fertilized plots. This different 347 trend could be ascribed to the different S forms present in the soil: the organic one related to 348 compost, and thus mainly present in the upper soil layers, and the inorganic one, which accumulate 349 moves deeper in the soil profile, and is probably less available to plants and more subjected to leaching and/or co-precipitation as calcium, magnesium or sodium sulphate sulfate (Scherer, 2001). 350

351

352 4.4 Compost effects on soil chemistryincreased soil CEC and EC

353 Compost contributes to the increase of exchangeable cations (i.e. Ca²⁺, K⁺, and Mg²⁺) creating 354 favorable conditions for cation exchange (Acharya et al., 2019; Dang et al., 2022). Fourteen years of compost addition significantly increased soil CEC and EC not only in the first (0 - 0.15 m) but also 355 356 in the deeper soil layers. Changes in the CEC of soils are directly linked to the negative charges in the SOM and in the humified compounds (do Carmo et al., 2016) and to the colloidal nature of 357 organic matter (Kumar Bhatt et al., 2019). Similarly, soil EC increase can be explained by the inputs 358 359 of nutrients and salts contained in the compost and by the soil organic matter mineralization rate. Notwithstanding, the increase of in soil EC values needs to be considered carefully, since -above a 360 361 critical range of 750 - 3490 µS cm⁻¹ plant growth could be damaged (do Carmo et al., 2016).

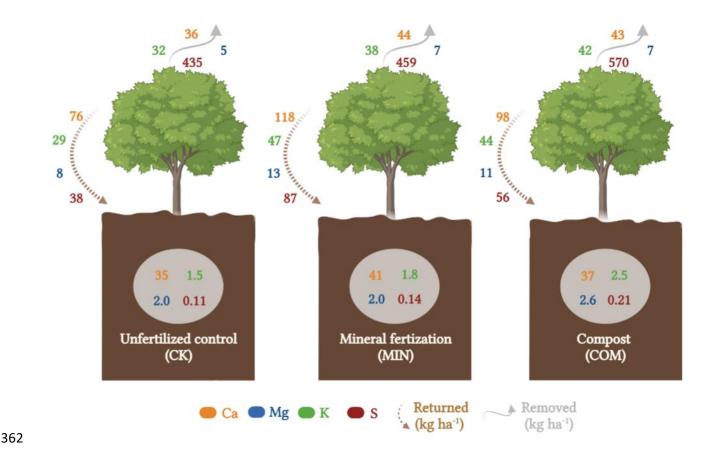


Figure 4 - Macronutrient dynamics in the soil-plant system. Ca, K, Mg and S content removed from
the plant (solid grey arrow) and recycled from plant to soil (dashed brown arrow) are expressed in
kg ha⁻¹, while Ca₂ content in soil is expressed in g over kg dry weight and K, Mg, and S soil-content
in soil are-is expressed in mg over kg dry weight Mg ha⁻¹.

367

368 5. Conclusion

369 Considering the goals of this study we have found that: (i) Longlong-term compost addition 370 facilitates macronutrients storage in the soil and thus indicating that it would favour soil macronutrients availability for the next crops (Figure 4). Moreover, organic amendment addition, 371 not only maintainsthis is functional both for the current crops and for future ones (Fig. 4); (ii) **T**the 372 supply of compost-addition, besides maintaining soil quality and fertility, but-also meets plant's 373 nutrient demand thus leading to higher plant performances and to economic and environmental 374 375 benefits .-; (iii) The synchronization between plant needs and nutrient soil availability is 376 fundamental for a correct fertilization management since it avoid plants' nutritional imbalance and,

377	at the same time, reduce the risk of nutrient leaching. The use of compost makes it difficult to
378	guarantee the mentioned synchronization, but significantly contributes to soil nutrient content and
379	soil quality increase. On the other hand, tThe exclusive use of mineral fertilizer, if not carefully
380	managed, even if meeting <u>the plant</u> 's nutrient demand, could lead <u>to macronutrient depletion due</u>
381	to scarce reserve creation in the soil; on the other hand, mineral fertilizer represents a source of
382	nutrients readily available to plant uptake. Therefore, the choice of the fertilizer to be used needs to
383	be calibrated on soil and plant requirements taking into account their potential effects (either
384	positive or negative, i.e. the increase of soil organic matter content or the increase of nutrient
385	leaching) on the environment
386	Future studies should take into consideration the effects of other organic amendments not only in
387	relation to soil and plant nutrient availability, but also on the effects that compost decomposition
388	could have on the amount of CO2 emitted or sequestered by the orchard, and the impact of
389	macronutrient dynamics on soil microbial communities structure and activity.
390	
391	
392	Acknowledgments
393	We would like to thank Dr. Andrea Simoni and Dr. Giampaolo Di Biase for performing the ICP-AES
394	measurements.
395	
396	Authors Contribution
397	Conceptualization: Elena Baldi, Moreno Toselli; Methodology: Elena Baldi; Formal analysis and
398	investigation: Elena Baldi, Martina Mazzon, Maurizio Quartieri, Luciano Cavani; Writing – original
399	draft preparation: Martina Mazzon; Writing – review and editing: Martina Mazzon, Elena Baldi,

400 Luciano Cavani; Resources: Moreno Toselli, Claudio Marzadori; Supervision: Moreno Toselli,
401 Claudio Marzadori.

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