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Isolation and Biological Characterization of Homoisoflavanoids and the Alkylamide *N-p*-Coumaroyltyramine from *Crinum biflorum* Rottb., an Amaryllidaceae Species Collected in Senegal

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Abstract: *Crinum biflorum* Rottb. (syn. *Crinum distichum*) is an Amaryllidaceae plant used in African traditional medicine but very few studies have been performed on this species from a chemical and applicative point of view. Bulbs of *C. biflorum*, collected in Senegal, were extracted with ethanol by Soxhlet and the corresponding organic extract was purified using chromatographic methods. The pure compounds were chemically characterized by spectroscopic techniques (1D and 2D ¹H and ¹³C NMR, HR MS and ECD) and X-ray analysis. Four homoisoflavanoids (**1–4**) and one alkylamide (**5**) were isolated and characterized as 5,6,7-trimethoxy-3-(4-hydroxybenzyl)chroman-4-one (**1**), as 3-hydroxy-5,6,7-trimethoxy-3-(4-hydroxybenzyl)chroman-4-one (**2**), as 3-hydroxy-5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one (**3**) and as 5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one (**4**), and the alkylamide as (*E*)-*N*-(4-hydroxyphenethyl)-3-(4-hydroxyphenyl)acrylamide (**5**), commonly named *N-p*-coumaroyltyramine. The relative configuration of compound **1** was verified thanks to the X-ray analysis which also allowed us to confirm its racemic nature. The absolute configurations of compounds **2** and **3** were assigned by comparing their ECD spectra with those previously reported for urginianins A and B. Flavanoids **1**, **3** and **4** showed promising anticancer properties being cytotoxic at low micromolar concentrations towards HeLa and A431 human cancer cell lines. The *N-p*-coumaroyltyramine (**5**) was selectively toxic to A431 and HeLa cancer cells while it protected immortalized HaCaT cells against oxidative stress induced by hydrogen peroxide. Compounds **1–4** also inhibited acetylcholinesterase activity with compound **3** being the most potent. The anti-amylase and the strong anti-glucosidase activity of compound **5** were confirmed. Our results show that *C. biflorum* produces compounds of therapeutic interest with anti-diabetic, anti-tumoral and anti-acetylcholinesterase properties.

Keywords: *Crinum biflorum*; homoisoflavanoids; alkylamides; cytotoxicity; antioxidant; antidiabetic and anti-acetylcholinesterase activities

1. Introduction

Plants and microorganisms are well-known sources of bioactive metabolites which have only been partly investigated [1]. Among the plants' kingdom, the Amaryllidaceae is a plant family extensively studied essentially for its alkaloids and related isocarbostryls content which show a broad spectrum of biological activities [2–5]. These plants are principally diffused in tropical and subtropical regions of the world, as Andean South America, the Mediterranean basin and Southern Africa [6], and include ca 1600 species classified into about 75 genera [7]. Hundreds of Amaryllidaceae alkaloids with different structures and biological activities were isolated and reported in several reviews. A Special Issue of the journal *Molecules* was edited by Bastida J. and Berkov S. in 2020 on different aspects of Amaryllidaceae alkaloids including biodiversity, chemocology, pharmacology, in vitro production, structural characterization, mass spectrometry and molecular modeling [8]. Advances on the chemical and biological characterization of the alkaloids and analogs isolated in the last decade from this plant family were reviewed by Masi et al. (2020) [9] and embedded in this Special Issue [8].

Crinum is a well-known subgroup of Amaryllidaceae studied for a long time and shown to be very rich in crinine-type alkaloids [10], one of the 12 ring-type in which are grouped the Amaryllidaceae alkaloids [11]. Three undescribed alkaloids, gigantelline, gigantellinine and gigancrinine with anti-acetylcholinesterase activity, were recently isolated from *Crinum jagus*, together with the already known lycorine-, cherylline-, galanthamine- and crinine-type alkaloids [12]. We also showed that *C. jagus* crude alkaloid extract inhibited Dengue virus (DENV) infection [13]. Among the alkaloids isolated from this plant, cherylline inhibited efficiently of both DENV ($EC_{50} = 8.8 \mu\text{M}$) and Zika virus (ZIKV) replication ($EC_{50} = 20.3 \mu\text{M}$), whereas it was ineffective on human immunodeficiency virus type 1 (HIV-1) infection. Thus, cherylline could be optimized for new therapeutic approaches against flaviviruses [13].

Amaryllidaceae plants, and in particular the subfamily Amaryllidoideae, also produce metabolites belonging to other classes of natural compounds such as flavonoids, lignans, chromones, terpenoids and ceramides. However, metabolites different from alkaloids are less studied. Still, up to now about 223 compounds, essentially flavan and phytosterols, were isolated from Amaryllidoideae, all with interesting biological activities and taxonomical importance [14]. In particular, flavonoids are a well-known class of natural compounds possessing protective action against oxidative stress, heart disease and some cancers, and are considered healthy for humans and livestock [15,16].

Examples of non-alkaloids bioactive metabolites used in folk medicine include 2(S)-4'-hydroxy-7-methoxyflavan with cytotoxic activity [17] and vanillin [18] from *Crinum bulbispermum*, while amabiloside and its 4-glucoside was isolated from *Crinum amabile* [19]. *Crinum latifolium*, a rare species growing in Vietnam, produced a coumarin derivative as 4-[(seneciolyloxy)methyl]-6,7-dimethoxycoumarin showing strong antiangiogenic activity [20]. *Crinum aurea* produced (7S)-7-(4-hydroxyphenyl)-7-hydroxypropyl)-2'-methylbenzene-3',6'-diol exhibiting neuroprotection against $\text{H}_2\text{O}_2/\text{CoCl}_2$ -induced neuronal cell death in dopaminergic neuroblastoma SH-SY5Y cells [21]. *Crinum asiaticum* L. var. *sinicum* produced several benzoic and cinnamic acid derivatives [22]. *Crinum purpurascens* produced β -sitosterol showing a weak antibiotic activity [23] while *Crinum ensifolium* produced parthenicin, a sesquiterpene exhibiting strong cytotoxic activity [24]. *Crinum angustum* produced aliphatic hydroxylketones [25] while *Crinum yemense* produced 6-hydroxy-2H-pyran-3-carbaldehyde showing strong inhibition of tyrosinase enzyme [26]. Acetovanillone and 4-hydroxyacetophenone, also known, respectively, as apocynin and piceol, were recently isolated from *Crinum buphonoides*, while only the former was isolated from *Crinum graminicola* [27].

Crinum biflorum Rottb. (syn. *Crinum distichum*) is found in seasonally flooded places in savannas from Senegal to Nigeria and extending to Sudan [28] but few studies have been carried out on its constituents and their biological properties [29].

Thus, the present study aimed to identify compounds with interesting biological activities from bulbs of *C. biflorum* collected in Senegal. Four homoisoflavonoids and one alkylamide were isolated. The chemical and biological properties were characterized to envisage their potential nutraceutical and pharmacological applications. This study expands the library of compounds isolated from *C. biflorum* and shows for the first time that this Amaryllidaceae species produces compounds of therapeutic interest against diabetes, cancer and Alzheimer's disease.

2. Materials and Methods

2.1. General Experimental Procedures

A JASCO P-1010 digital polarimeter (Jasco, Tokyo, Japan) was used to measure the optical rotations in CH₃OH. Electronic circular dichroism (ECD) spectra were recorded on a JASCO J-815 spectrometer (Jasco, Tokyo, Japan) in CH₃OH. 400 Anova Advance (Bruker, Karlsruhe, Germany) and Inova 500 MHz (Varian, Palo Alto, CA, USA) instruments were used to record ¹H and ¹³C NMR spectra in CDCl₃ or CD₃OD at 400/100 or 500/125 MHz, respectively. The same solvents were used as internal standards. Correlation spectroscopy with a 45° mixing pulse (COSY-45), Nuclear Overhauser effect spectroscopy (NOESY), heteronuclear single quantum correlation (HSQC) and heteronuclear multiple bond correlation (HMBC) experiments were performed using Bruker or Varian microprograms. Electrospray ionization (ESI) mass spectra and liquid chromatography/mass spectrometry (LC/MS) analyses were carried out using the LC/MS time-of-flight (TOF) system Agilent 6230B, high-performance liquid chromatography (HPLC) 1260 Infinity. A Phenomenex LUNA [C₁₈ (2) 5 μm 150 × 4.6 mm column] was utilized to perform the HPLC separations. Analytical and preparative thin-layer chromatography (TLC) were performed on silica gel (Kieselgel 60, F₂₅₄, 0.25 and 0.5 mm, respectively, Merck) plates. The spots were visualized by exposure to ultraviolet (UV) radiation (254) or iodine vapors. Column chromatography (CC) was performed using silica gel (Kieselgel 60, 0.063–0.200 mm, Merck). Sigma-Aldrich Co. (St. Louis, MO, USA) supplied all the reagents and the solvents.

2.2. Plant Material

Bulbs of *C. biflorum* were collected in Senegal, in Kaffrine department, in December 2018. A senior scientist from the Herbarium of IFAN of University Cheikh Anta Diop of Dakar taxonomically identified the plant materials.

2.3. Extraction and Purification of Compounds 1–5

Fresh bulbs of *C. biflorum* were dried at room temperature and then finely powdered. The resultant powder (545 g) was extracted with ethanol by Soxhlet obtaining a semisolid brown extract (450 mg). The latter was fractionated by CC eluted with CHCl₃/EtOAc/CH₃OH (3:1.5:0.5 *v/v/v*), affording eight groups of homogeneous fractions (F1–F8). The residue (102.8 mg) of fraction F1 was further purified by CC eluted with CHCl₃/*iso*PrOH (97:3 *v/v*) yielding ten groups of homogeneous fractions (F1.1–F1.10). The residue (15.8 mg) of F1.3 was purified by preparative TLC eluting with CH₂Cl₂/MeOH (97:3 *v/v*) yielding compound 4 (3.04 mg). The residue (14.3 mg) of F1.5 was purified by preparative TLC eluting with *n*-hexane/EtOAc (6:4 *v/v*) yielding compound 1 (1.42 mg) and compound 3 (3.52 mg). Compound 1 was crystallized from a CHCl₃-*iso*PrOH 9:1 *v/v* solution. The residue (6.7 mg) of F1.6 was purified by TLC eluted with CHCl₃/*iso*PrOH (9:1 *v/v*) yielding compound 2 (1.61 mg). The residue (15.2 mg) of F4 was further purified by preparative TLC eluting with CHCl₃/*iso*PrOH (95:5 *v/v*) yielding compound 5 (1.19 mg).

2.4. Spectroscopic Data of Compounds 1–5

5,6,7-trimethoxy-3-(4-hydroxybenzyl)chroman-4-one (1). [α]_D²⁵: 0 (c 0.3 CH₃OH); ¹H NMR data are very similar to those reported by Sylao et al. 1999 [30]; ESIMS (+) *m/z*: 727 [2M + K]⁺, 719 [2M + Na]⁺, 383 [M + K]⁺, 367 [M + Na]⁺, 345 [M + H]⁺.

3-hydroxy-5,6,7-trimethoxy-3-(4-hydroxybenzyl)chroman-4-one (2). ECD (*c* 0.010 mM, CH₃OH) $\Delta\epsilon$ 255 (-1.1), 280 (+2.5), 317 (-1.6); lit. [31]: ECD (*c* 0.031 mM, CH₃OH) $\Delta\epsilon$ 281 (+10.4), 316 (-7.3); ¹H NMR data are very similar to those reported for urginianin B [31]; ESIMS (+) *m/z*: 383 [M + Na]⁺, 361 [M + H]⁺.

3-hydroxy-5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one (3). ECD (*c* 0.010 mM, CH₃OH) $\Delta\epsilon$ 255 (-1.1), 281 (+2.5), 317 (-1.6); lit. [31]: ECD (*c* 0.031 mM, CH₃OH) $\Delta\epsilon$ 280 (+10.8), 315 (-7.2); ¹H NMR data are very similar to those reported for unginianin A [31]; ESIMS (+) *m/z*: 397 [M + Na]⁺, 375 [M + H]⁺.

5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one (4). [α]_D²⁵: 0 (*c* 0.2 CH₃OH); ¹H NMR data are very similar to those reported by Sidwell and Tamm (1970) [32]; ESIMS (+) *m/z*: 381 [M + Na]⁺, 359 [M + H]⁺.

(*E*)-*N*-(4-hydroxyphenethyl)-3-(4-hydroxyphenyl)acrylamide (*N-p*-coumaroyltyramine) (5). ¹H and ¹³C NMR data are very similar to those reported by Bhatti et al. (1992) [33]; ESIMS (+) *m/z*: 306 [M + Na]⁺; ESIMS (-) *m/z*: 282 [M - H].

2.5. Crystal Structure Determination of Compound 1

Single crystals of **1** suitable for X-ray structure analysis were obtained by slow evaporation of a CHCl₃-*iso*PrOH 9:1 *v/v* solution. One selected crystal was mounted at ambient temperature on a Bruker–Nonius KappaCCD diffractometer (Bruker-Nonius, Delft, The Netherlands) (graphite monochromated MoK α radiation, λ = 0.710 73 Å, CCD rotation images, thick slices and φ and ω scan to fill the asymmetric unit). A semi-empirical absorption correction (multiscan, SADABS) was applied. The structure was solved by direct methods using the SIR97 program [34] and anisotropically refined by the full-matrix least-squares method on F² against all independently measured reflections using the SHELXL-2018/3 program [35] with the aid of program WinGX [36]. Water solvent crystallization molecules are present in the structure. The hydroxy and water H atoms were located in different Fourier maps and freely refined with Uiso(H) equal to 1.2 Ueq of the carrier atom. All of the other hydrogen atoms were introduced in calculated positions and refined according to the riding model with C–H distances in the range of 0.93–0.96 Å and with Uiso(H) equal to 1.2 Ueq or 1.5 Ueq (Cmethyl) of the carrier atom. One stereogenic center is present in the compound that crystallizes in the centrosymmetric P-1 space group as a racemate. The E-statistics indicate that the structure is centrosymmetric. Two independent X-ray structure analyses performed on different crystals confirmed the result. Unitary cell parameters were checked on several crystals. Figures were generated using ORTEP-3 [36] and Mercury-CSD-3.9. [37]. Crystallographic data of **1**: empirical formula: C₁₉H₂₀O₆·H₂O; formula weight: 362.36 g mol⁻¹; triclinic, P-1; a: 8.368(2) Å; b: 10.183(2) Å; c: 12.0420(6) Å; α : 104.470(8)°; β : 108.252(13)°; γ : 100.010(19)°; V: 907.4(3) Å³; Z: 2, Dx: 1.326 Mg/m³. All homoisoflavanoid crystallographic data for (**1**) were deposited in the Cambridge Crystallographic Data Centre with deposition number CCDC 2092030. These data can be obtained free of charge from www.ccdc.cam.ac.uk/data_request/cif.

2.6. Cell Culture and Reagents

HaCaT, spontaneously immortalized keratinocytes from adult skin, were purchased from Service Cell Line (GmbH, Eppelheim, CLS, Germany) and cultured as described [38,39]. HeLa cervical cancer cells (CCL-2) and A431 (ATCC-CRL1555) human epidermoid carcinoma cells were from American Type Culture Collection (ATCC, Manassas, VA, USA). According to the p53 compendium database (<http://p53.fr/tp53-database/the-tp53-cell-line-compendium>, 05/05/2021), HaCaT cells contain mutant p53 (H179Y/R282W), HeLa have p53 impaired function by viral infection while A431 cells contain only one p53 mutated allele (R273H). All mentioned cell lines were cultured in Dulbecco's Modified Eagle's Medium (DMEM, Sigma Chemical Co, St. Louis, MO, USA) supplemented with 10% fetal bovine serum (FBS, Hyclone Laboratories, Inc. Logan, UT, USA) at 37 °C in a

humidified atmosphere of 5% CO₂. All cell lines were routinely tested for mycoplasma contamination and were not infected.

2.7. MTT Assay

Cell viability was assessed by 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay following the published procedure [40]. Briefly, cells were seeded at 10⁵/cm² density in 96-well plates. Twenty-four hours later, the medium was changed and supplemented with the specified concentrations of metabolite (from 0.5 to 10 mM in DMSO) for 24 and 48 h. MTT solution 1:10 (stock solution 5 mg/mL) was added to each well and the absorbance was measured in dual-wavelength mode (570 nm and 630 nm). The percentage of cell viability was calculated as follows: mean (A570-A630) and compared to cells supplemented with DMSO alone. Values shown in the plot are mean ± SD of sixfold determinations. Mean and the standard deviation was calculated on biological triplicates using GraphPad Prism8 software (GraphPad, San Diego, CA).

2.8. Detection of DNA Damage

Cells were seeded in 35 mm dishes on micro cover glasses (BDH) and treated with the metabolite at a concentration of 10 µM. At 48 h after treatment, cells were washed with cold phosphate-buffered saline (PBS) and fixed with 4% paraformaldehyde Sigma-Aldrich (Merck Life Science, Milan, Italy) for 15 min at RT. Cells were permeabilized with ice-cold 0.5% Triton X-100 for 5 min and then washed with PBS. Cells were then incubated with phospho-histone H2A.X (Ser139) antibody (from Cell Signaling Technologies 9542, Boston, MA, USA) for 1 h, followed by DAPI (Sigma-Aldrich) for 3 min and washed with PBS/0.05% Tween. Coverslip was mounted with Ibbidi mounting medium (Ibbidi GmbH, Martinsried, Germany). Images were taken with a Zeiss confocal laser-scanning microscope Axio Observer (Zeiss, Ostfilden, Germany) (scale bar, 20 µm). A 40× objective was used and image analysis was performed using Fiji ImageJ open source software project (<https://imagej.net/imagej/>). All the images were taken with the same setting [39].

2.9. Western Blot Analysis

Western blot was performed as previously reported [39,41]. Briefly, 30 µg of whole-cell extracts were separated by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE), subjected to Western blot and incubated overnight at 4 °C with antibodies. Antibodies against p21WAF, Poly [ADP-ribose] polymerase 1 (PARP1) and actin were from Cell Signaling Technologies 9542, Boston, MA, USA. Each experiment was run in triplicate. Signal intensities of Western blot bands were quantified by Quantity One analysis software (Version Number 2, Biorad Laboratories, London, UK) and analyzed by GraphPad Prism 8.0.2 software (GraphPad, San Diego, CA).

2.10. DCFDA Assay

N-p-coumaroyltyramine antioxidant activity was measured using 2'-7'-dichlorofluorescein diacetate (DCFDA), a non-fluorescent compound permeable to the cell membrane, which can be oxidized by reactive oxygen species (ROS) giving a fluorescent compound. Cells were seeded at 2.5 × 10⁴ in 96 well and pre-treated with *N-p*-coumaroyltyramine (10 and 100 mM). The medium was removed after 4 h and 1 mM (3%) H₂O₂ was added for 45 min, 1.5 and 2.0 h. Cells were washed with PBS and a fresh medium with DCFDA (30 mM) was added for 45 min, then DCFDA was removed by washing in PBS and the cells were harvested. The measurement of ROS was obtained using the Sinergy H4 microplate reader (Gen5 2.07, Thermofisher, Waltham MA, USA). The fluorescence emitted from the cells treated with DCFDA was compared to the untreated cells. Trolox was used as a positive control. Values shown in the plot are mean ± SD of sixfold determinations. The mean and the standard deviation were calculated on biological triplicates using GraphPad Prism 8.0.2 software (GraphPad, San Diego, CA).

2.11. 2Pseudotyped HIV-1_{GFP} Infectivity Assay

The anti-HIV-1 activity of compounds 1–5 was evaluated using VSV-G pseudotyped NL4_{3GFP} infection of human monocytic THP-1 cells. THP-1 and NL4–3_{GFP} were generously provided by Lionel Berthoux and Amita Singh and are described in Ka et al. (2021) [13]. THP-1 cells were seeded at 2.0×10^4 cells per well in 96 well-plates. The next day, cells were treated with 4 concentrations of each compound (12.5, 25, 50 and 100 mM) and then infected with HIV_{GFP} at a MOI of 1. After 72 h, cells were stained with propidium iodide (PI, 0.5 mg/mL) and both PI⁺ and HIV-1_{GFP}⁺ infected cells frequencies were assessed on a FC500 MPL cytometer (Beckman Coulter, Inc., Mississauga, ON, Canada) and analyzed using FlowJo software (FlowJo LLC, BD Biosciences, Ashland, OR, USA). Matched concentrations of dimethyl sulfoxide (DMSO) were used as negative controls. All infection assays were performed in triplicate.

2.12. α -Glucosidase and α -Amylase Inhibitor Assay

α -glucosidase and α -amylase inhibitor screening kits (colorimetric) were purchased from Biovision (Milpitas, CA, USA). In total, 10 mM of stock solution of all the tested compounds were dissolved in DMSO and serially diluted in the assay buffer of each kit. Experiments were performed according to the manufacturer's protocol. Briefly, for the α -glucosidase assay, 10 μ L of serially diluted compounds at the corresponding concentration (10 nm–1 mM) were added into designated wells of clear 96 well-plates. Subsequently, 10 μ L of the α -glycosidase enzyme was added to each well and volume was adjusted to 80 μ L and plates were incubated for 15-min at room temperature in dark condition. Then, 20 μ L of α -glycosidase substrate mixture was added in all wells and kinetic of reaction was measured at OD: 410 nm for 60 min at 2 min intervals by using a multiplate reader, Biotek instrument, Inc., Canada. Enzyme control (no inhibitor), background control (no enzyme), solvent control (DMSO) and inhibitor control (acarbose) were included in the plates. For the α -amylase assay, 50 μ L of serially diluted compounds (3.25 μ M to 500 μ M) were added into a clear 96-well plate with 50 μ L of assay buffer and 50 μ L of α -amylase enzymes. The plate was incubated at room temperature in the dark for 10 min. Then 50 μ L of the α -amylase substrate was added in all wells. The kinetic of reaction was measured at OD:410 nm for 26 min at intervals of 2 min by using a multiplate reader. Control α -amylase inhibitor was provided by the manufacturer, enzyme control, background control and solvent control were all included. Enzyme inhibition was calculated according to Zhang et al. (2014) [42]. In summary, ODs were plotted according to the time for each sample. Areas under the curve (AUC) were calculated, and enzyme inhibition was measured as $100 - (AUC_{\text{compound}}/AUC_{\text{enzyme}}) \times 100$ for each dilution of each compound.

2.13. Anti-Acetylcholinesterase Assays

In vitro acetylcholinesterase (AChE) activity was assessed exactly as in Ka et al. (2020) [12] following Ellman's colorimetric protocol [43] with the Acetylcholinesterase Assay Kit (Abcam Inc., Boston, MA, USA). Briefly, 50 μ L serial dilutions (3.9–500 μ M) of compounds 1–5 were prepared in Tris-HCl pH = 7.9 buffer into designated wells of a clear 96 well-plate. A total of 5 μ L of DTNB was added in each well, then 50 μ L of diluted acetylcholinesterase was added. The plate was incubated for 10 min in the dark. Matched concentrations of DMSO were used as a negative control. Kinetic of reaction was measured in a multiple plate reader at 410 nm in kinetic mode for 40 min at room temperature. The percentage of anti-AChE inhibition was calculated according to the following formula: $100 - [(E - S)/E] \times 100$, where E is the activity of the enzyme with matched concentrations of DMSO and S is the activity of the enzyme with the test sample.

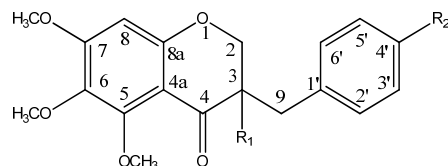
2.14. Statistical Analysis

Statistical analyses were carried out using the GraphPad Prism version 8.1.2 (<https://www.graphpad.com/scientific-software/prism/>). Data were represented as the mean \pm standard deviation and analyzed for statistical significance using ordinary one-way analysis of variance (ANOVA) and multiple comparisons. For all tests, $p < 0.05$ was considered to indicate a statistically significant difference.

3. Results

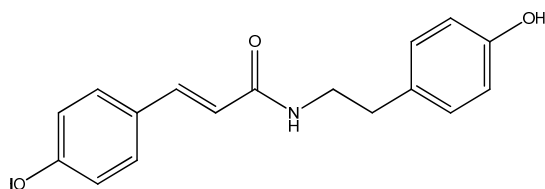
3.1. Identification of Metabolites Isolated from *C. biflorum*

The purification of the crude organic extract from bulbs of *C. biflorum* allowed us to isolate four homoisoflavanoids ((1)–(4), Figure 1) identified using spectroscopic (essentially 1D and 2D ^1H and ^{13}C NMR and HR MS) methods as 5,6,7-trimethoxy-3-(4-hydroxybenzyl)chroman-4-one (1), as 3-hydroxy-5,6,7-trimethoxy-3-(4-hydroxybenzyl)chroman-4-one (2), as 3-hydroxy-5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one (3) and as 5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one (4).



- 1, $R_1 = \beta\text{H}$, $R_2 = \text{OH}$
 2, $R_1 = \beta\text{OH}$, $R_2 = \text{OH}$
 3, $R_1 = \beta\text{OH}$, $R_2 = \text{OCH}_3$
 4, $R_1 = \alpha\text{H}$, $R_2 = \text{OCH}_3$

- 1, 5,6,7-trimethoxy-3-(4-hydroxybenzyl)chroman-4-one
 2, 3-hydroxy-5,6,7-trimethoxy-3-(4-hydroxybenzyl)chroman-4-one = urgieanin B
 3, 3-hydroxy-5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one = urgieanin A
 4, 5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one



5, (*E*)-*N*-(4-hydroxyphenethyl)-3-(4-hydroxyphenyl)acrylamide

Figure 1. Structure of homoisoflavanoids (1)–(4) and alkylamide (5) isolated from *C. biflorum* bulbs.

The identification of 1 was performed comparing its ^1H NMR spectrum with that reported in the literature when it was isolated for the first time from *Scilla nervosa*, subsp. *rigidifolia* collected in Botswana where it is used in Zulu folk medicine to treat rheumatic fever and as a purge for children [30]. ESI MS spectrum showed the potassium $[2\text{M} + \text{K}]^+$ and sodium $[2\text{M} + \text{Na}]^+$ dimeric, the potassium $[\text{M} + \text{K}]^+$ and sodium $[\text{M} + \text{Na}]^+$ and the protonated $[\text{M} + \text{H}]^+$ adduct ions at m/z : 727, 711, 383, 367 and 345.

The structure of the homoisoflavanoid (1) was confirmed by X-ray analysis of a single crystal obtained by slow evaporation of a CHCl_3 -isoPrOH 9:1 *v/v* solution. Compound 1 crystallizes in the P-1 space group with one molecule of 1 and one H_2O solvent molecule contained in the independent unit. All bond lengths and angles are in a normal range. The molecule consists of a substituted chromanone system. The six-membered chroman-4-one ring assumes an envelope conformation with C2 atom at the flap. The 4-hydroxybenzyl substituent at C3 is in the equatorial position and places nearly parallel to the benzene

ring of the chromanone system (Figure 2A,B). The crystal packing is stabilized by strong OH...O hydrogen bonds involving **1** and solvent water molecules (Figure 3A,B).

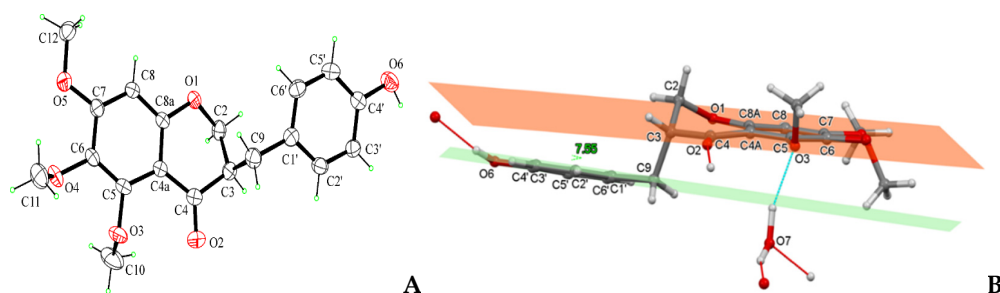


Figure 2. (A) ORTEP view of the homoisoflavanoid (**1**) molecular structure, with ellipsoids drawn at the 30% probability level. (B) Independent unit of compound **1** in a perspective view showing a quite parallelism between hydroxybenzene plane (green) and benzene ring of chromanone system (orange) (angle of 7.55° between the two planes). Ball-and-stick style. Hydrogen bonds drawn as cyan and red dashed lines.

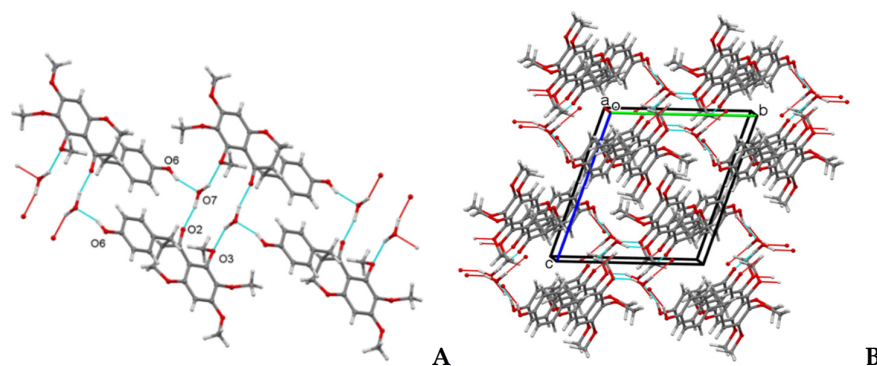


Figure 3. (A) Details of hydrogen bonding pattern in compound **1**. (B) Perspective view of the crystal packing of compound **1** with hydrogen bond pattern drawn as cyan and red lines. Ball-and-stick style.

The homoisoflavanoid (**1**) molecule contains one stereogenic center at C3 atom and crystallizes in the centrosymmetric P-1 space group as a racemate. The racemic nature of crystals was confirmed by performing two independent X-ray structure analyses on different crystals (Figure 2B). This result can be ascribed to easy inversion at the chirality center, because of the presence at C-3 of a proton which could exchange by keto-enol tautomerism. The ECD spectrum of **1** (Figure 4, black line) cannot be interpreted due to the presence of a racemic mixture. These results were confirmed by the optical inactivity found by measuring the specific optical rotation of compound **1**.

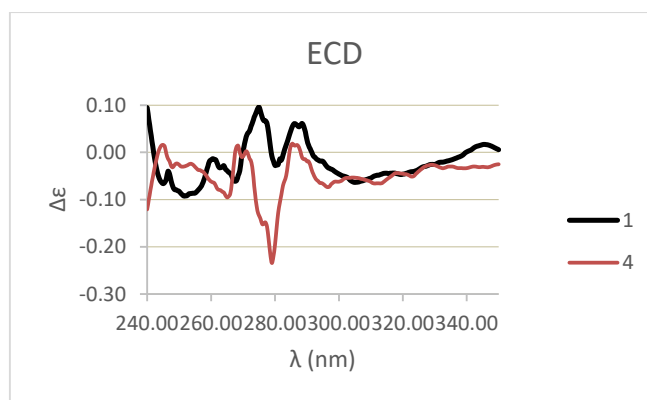


Figure 4. Experimental ECD spectra of homoisoflavanoids (1) (black solid line), and 4 (red line).

Homoisoflavanoids **2** and **3** were identified by comparing their physic (ECD) and spectroscopic data (^1H NMR) with those reported in the literature for urgineanins B and A, isolated from *Urginea depressa*. This is an Asparagaceae collected in South Africa and used for its antiproliferative activity against the A2780 ovarian cancer cell line [31]. The ESI MS spectra of both **2** and **3** confirmed their identification showing the sodiated and protonated adduct ions $[\text{M} + \text{Na}]^+$ and $[\text{M} + \text{H}]^+$, respectively, at m/z 383 and 361 and 397 and 375. The absolute configuration of homoisoflavanoids **2** and **3** was the same of urgineanins B and A as appeared by comparison of their ECD spectra (Figure 5, black line for **2** and red line for **3**) with those already reported in the literature by Dai et al. (2013) [31] which assigned a *R* configuration at C-3 of the two homoisoflavanoids by comparison of their experimental ECD spectra with those reported in literature for caesalpiniafenol A [31].

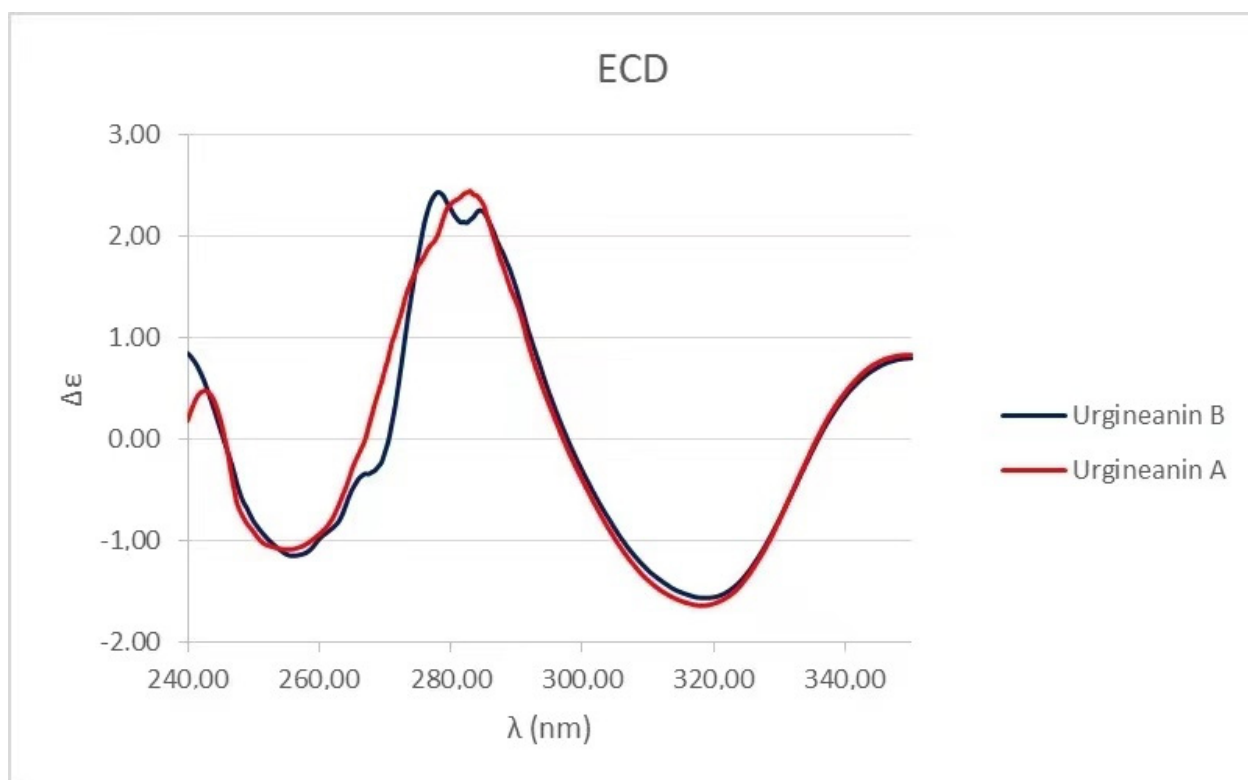


Figure 5. ECD spectra of homoisoflavanoids (2) (black solid line), and 3 (red line).

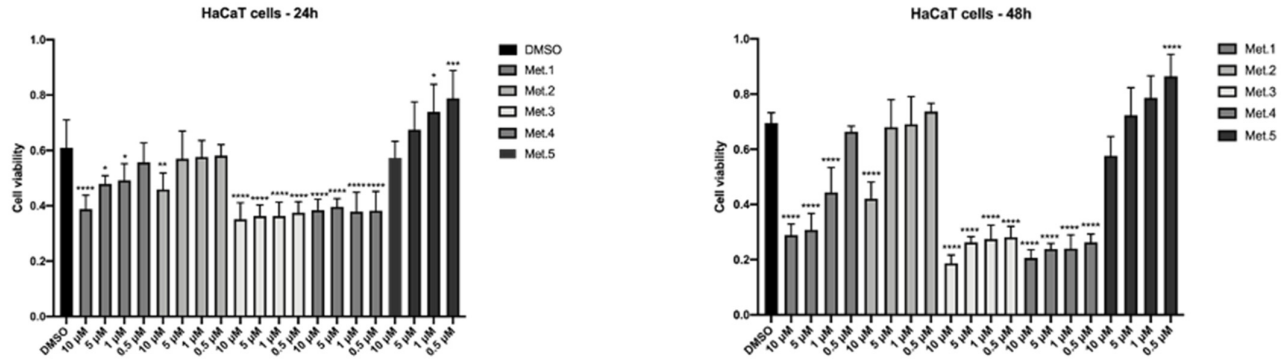
The fourth homoisoflavanoid (**4**) was identified as the 5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one by comparing its physic and spectroscopic data with those reported for the trimethyl derivative of 3,9-dihydroautumnalin obtained together to other homoisoflavanoids from *Eucomis autumnalis* (Liliaceae) [32]. The same derivative was also obtained from 3,9-dihydroeucomsalin, which resulted to be identical to 3,9-dihydroautumnalin, which was isolated from the bulbs of *Muscari comsum* (commonly named lampascioni) collected in Basilicata region, Italy, where they are used in traditional cuisine as bitter plants [44]. The identification of compound **4** was also supported by ESI MS data which showed the sodiated $[M + Na]^+$ and protonated $[M + H]^+$ adduct ions at m/z 381 and 359. The ECD spectrum of **4** (Figure 4, red line) cannot be interpreted, as in the case of compound **1**, due to the presence of a racemic mixture, as confirmed by the optical inactivity found by measuring the specific optical rotation of **4**.

From the ethanolic extract of the *C. biflorum* bulbs and alkylamide was also isolated and identified compound **5**, or (*E*)-*N*-(4-hydroxyphenethyl)-3-(4-hydroxyphenyl)acrylamide (**5**, Figure 1). Compound **5** was identified by comparison of its spectroscopic data with those reported in the literature when isolated from the first time from crude Chinese drug “Xiebai”, the tuber of *Allium bakeri* Reg. (Liliaceae) and used for inhibition on human platelet aggregation [45]. A similar comparison was done with the data reported when **5** was isolated together with some alkaloids from *Fumaria indica*, collected in Multan City (Punjab, Pakistan) and inappropriately indicated as an alkaloid instead of amide [33]. The identification of compound **5** was also supported by ESI MS data which showed the sodiated $[M + Na]^+$ adduct ion at m/z 306. When the same spectrum was recorded in negative modality, the pseudomolecular ion $[M - H]^-$ at m/z 282 was observed.

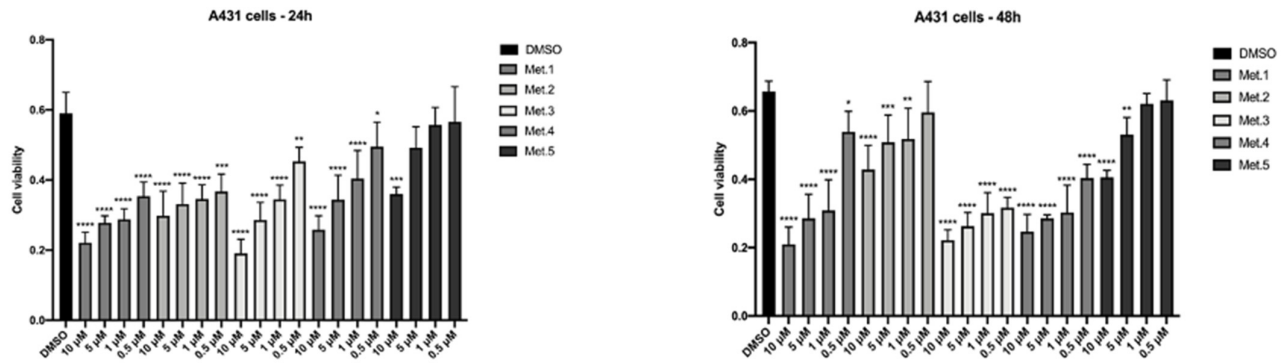
3.2. In Vitro Cytotoxicity

Considering that the anti-tumor activities of homoisoflavanoids (**1–4**) from *C. biflorum* have not yet been addressed, we evaluated the in vitro cytotoxicity of isolated homoisoflavanoids towards HaCaT, A431 and HeLa human cell lines using the MTT viability assay. The *N-p*-coumaroyltyramine (**5**) alkylamide was also included in these experiments. Cells were plated and supplemented with each metabolite, at low micromolar concentrations ranging from 0.5 to 10 μ M in DMSO, and incubated at 37 °C for 24 and 48 h. As shown in Figure 6, all tested metabolites were toxic for cancer cell lines, in a dose and time-dependent way, with HeLa cells being more sensitive than A431. Remarkably, 48 h treatment with 5 μ M metabolite **1** reduced HeLa cells viability to less than 20% of the untreated control. All metabolites were less active on HaCaT immortalized keratinocytes at 24 h of incubation over the range of concentration tested although a significant reduction of HaCaT cell viability was caused by metabolites **3** and **4** when the incubation was extended for 48 h. Table 1 lists the IC_{50} values obtained with the test compounds are means of triplicates at 24 h. The *N-p*-coumaroyltyramine (**5**) was also found to be toxic on A431 and HeLa cancer cells while at the lower concentration tested, 0.5 and 1 μ M, it was shown to significantly increase HaCaT cell viability (Figure 6). Detection of nuclear γ -H2A.X foci provides indirect evidence of the occurrence of DNA double-strand breaks (DSB) and/or DNA replication stress [40]. Upon induction of a DNA double-strand break, the H2A.X histone becomes rapidly phosphorylated at serine 139 to form γ H2AX [46]. This phosphorylation event is dynamic, complex and depends on interactions between MDC1, H2AX and ATM and other kinases to persist [47]. This amplified response is easily detected using specific antibodies against γ -H2AX, manifesting discrete nuclear foci.

• HaCaT cells: 96-well, sixfold, 1×10^4



• A431 cells: 96-well, sixfold, 8×10^3



• Hela cells: 96-well, sixfold, 8×10^3

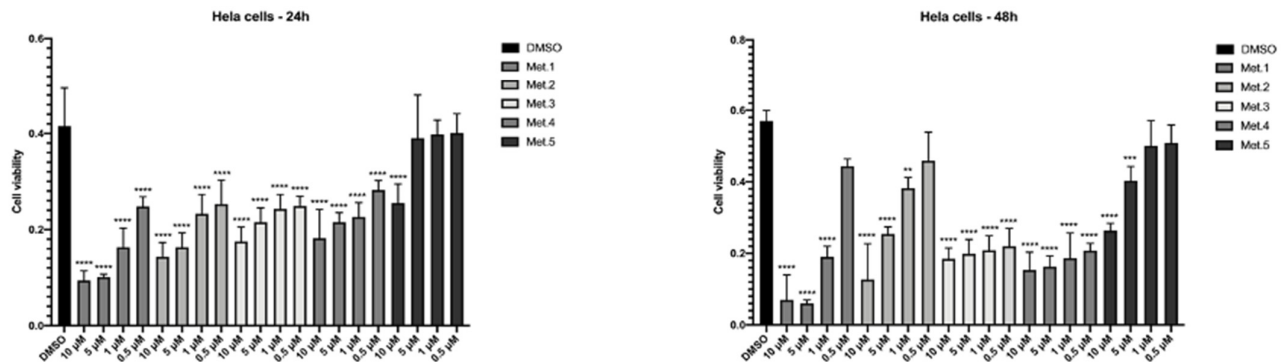


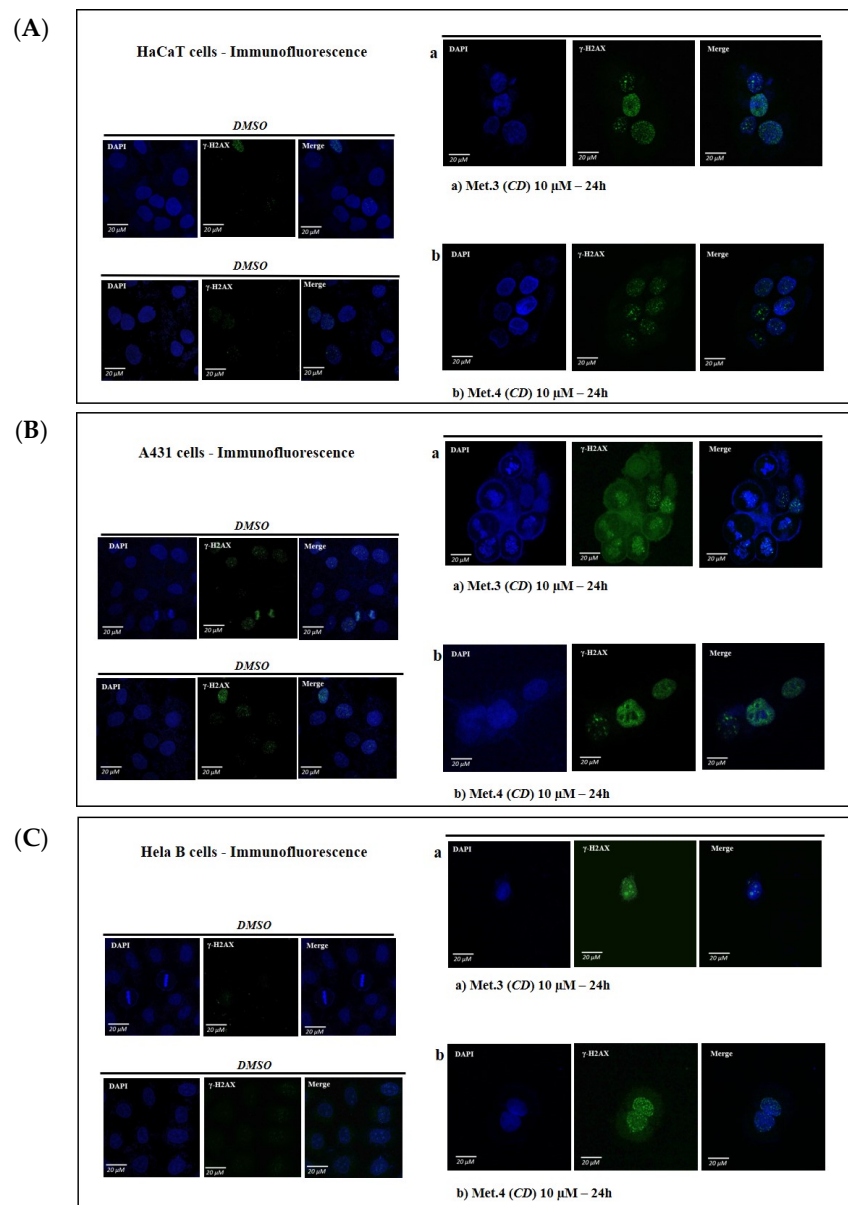
Figure 6. MTT viability assays with homoisoflavanoids 1–4 and *N-p*-coumaroyltyramine (5) from *C. biflorum* on HaCaT, A431 and Hela human cell lines. Statistical analyses were performed using one-way ANOVA and Dunnett’s multiple comparisons test. Levels of significance between points of expression are indicated (**** $p < 0.001$, *** $p < 0.01$, ** $p < 0.02$, * $p < 0.05$).

Table 1. Cytotoxic activities of metabolites 1–5. Compounds IC₅₀ (μM) ¹.

Compound	HaCaT	A431	HeLa
1	>10	6.0	1.0
2	>10	>10	4.0
3	>10	4.0	4.5
4	>10	6.5	5.5
5	Not detected	>10	7.5

¹ IC₅₀ was calculated after 24 h of incubation.

The formation of γ-H2AX foci by immunofluorescence using the antibody against the histone H2A.X phosphorylated in Serine 139 was monitored considering that the homoisoflavonoids isolated from *C. biflorum* reduced cell viability. HaCaT, HeLa and A431 cells were treated with 10 μM metabolites 3 and 4 for 24 h to detect DNA damage foci. As shown in Figure 7 a remarkable increase of nuclear γH2AX foci was observed in all tested cell lines.



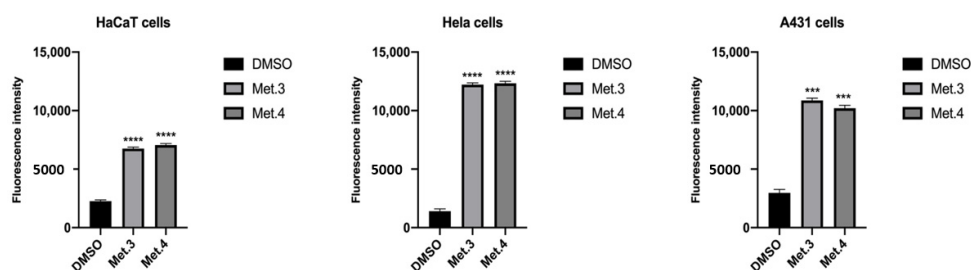
(D). Quantification of γ -H2AX mean fluorescence

Figure 7. Immunofluorescence microscopy showing γ -H2AX foci formation (green) in nuclei of HaCaT (panel (A)), A431 cells (panel (B)) or HeLa cells (panel (C)) treated with DMSO alone or 10 μ M **3** or **4** for 24 h. Nuclei were stained with DAPI (blue). Note the abnormal morphology of nuclei induced by **3** and **4** in A431 cells compared to HaCaT cells. Images from 3 fields per each experimental point were collected to obtain data for up to 150 cells. Quantitation of γ -H2AX foci fluorescence was performed by Image J software shown as mean \pm SD in graph bars of panels (D). Statistical analyses were performed using one-way ANOVA and Dunnett's multiple comparisons test. Levels of significance between points of expression are indicated (**** $p < 0.0001$, *** $p < 0.001$).

Determination of reactive oxygen species (ROS) induced by H_2O_2 in HaCaT cells treated with 10 and 100 μ M *N-p*-coumaroyltyramine alkylamide revealed that **5** has antioxidant activity comparable to Trolox, the water-soluble derivative of vitamin E used as a positive control (Figure 8). Remarkably, treatment of HaCaT cells with 10 μ M metabolite **3** or **4** did not increase the production of reactive oxygen species (ROS) implying that ROS generation was not responsible for the observed DNA damage (Figure 9A,B).

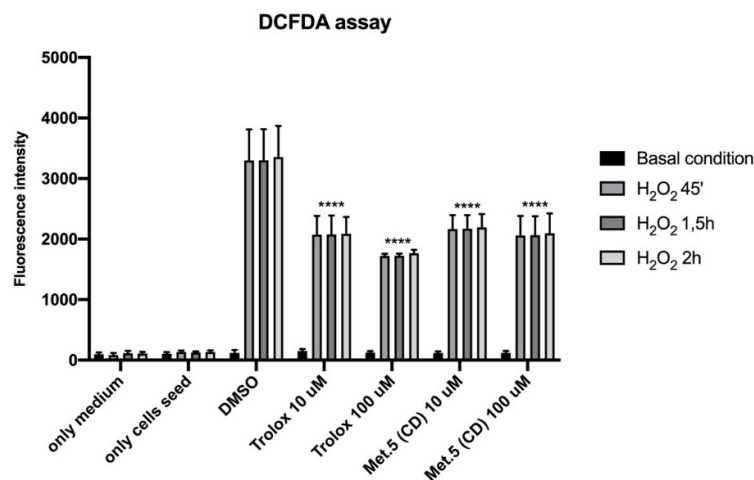


Figure 8. DCFDA assay. HaCaT cells were seeded and pre-treated for 4 h with 10 and 100 μ M *N-p*-coumaroyltyramine (**5**) from *C. biflorum*. H_2O_2 (1 mM; 3%) was added to the medium for 45', 1.5 and 2 h. The fluorescence intensity of DCFDA was read after 45' of incubation. Trolox was used as a positive control. Levels of significance between points of expression are indicated (**** $p < 0.0001$).

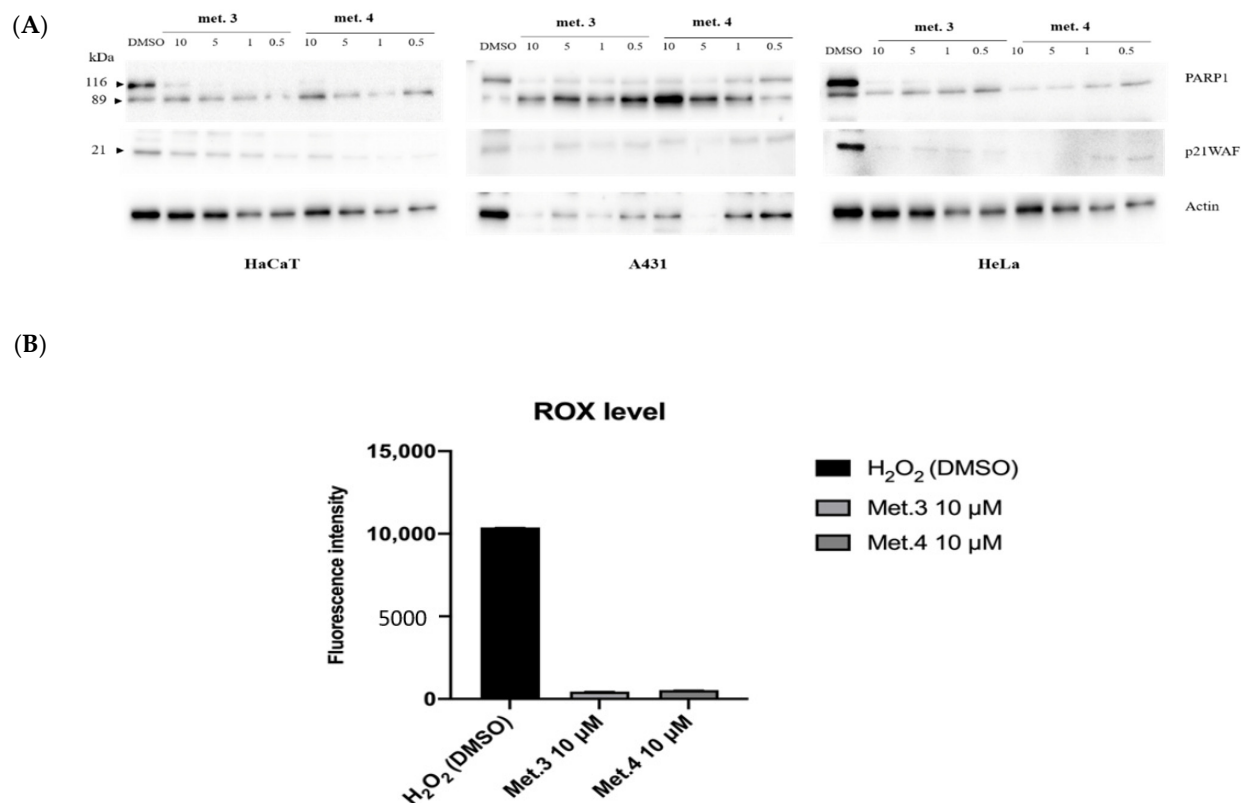


Figure 9. (A) Representative immunoblot analysis showing the effect of **3** and **4** on PARP1 activation and p21WAF protein levels. Full length and cleaved PARP1 bands were revealed by immunoblot with specific antibodies. The blots were then re-probed with actin antibody to check for protein loading. (B) DCFDA on HaCaT cells pre-treated for 4 h with 10 μ M **3** and **4** from *C. biflorum*. H₂O₂ (1 mM; 3%) was added to the medium for 45' and used as a positive control. Fluorescence intensity of DCFDA was read after 45' of incubation.

3.3. Antidiabetic and Anti-Acetylcholinesterase Properties

Next, we assessed the anti-acetylcholinesterase and antidiabetic (Figure 10A,B) properties of *C. biflorum*-isolated compounds. We confirmed that compound **5** was a strong inhibitor of α -amylase and α -glucosidase (Figure 10) [48]. Compound **1**, **2**, **3** also inhibited 50% of α -glucosidase at high concentration (1 mM) (Figure 10B). Altogether, the results support the antidiabetic properties of *C. biflorum*. Compounds **1–4** also showed anti-acetylcholinesterase activity. Interestingly, compound **5** weakly inhibited AChE catalyzed reaction at high concentrations, while **1**, **2** and **4** displayed increasing inhibition from 15.6 to 250 μ M up to 20% (Figure 11). Compound **3** was the most potent inhibitor, blocking from 15 to 30% of enzyme activity as concentrations increased from 3.9 to 50 μ M (Figure 11).

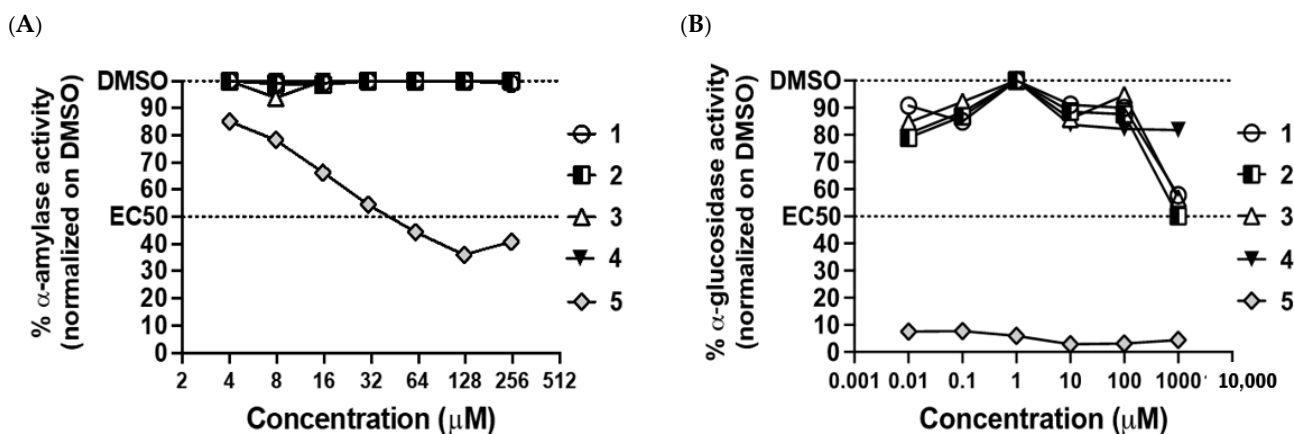


Figure 10. (A) Anti- α -amylase activity. Kinetics of α -amylase reaction was measured in presence of each compound at concentrations ranging from 3.9–250 μ M, and matched concentrations of DMSO solvent over a 26 min period. Areas under the curves (AUC) of OD (410 nm) vs. time were calculated according to Zhang et al. (2014) [42] and used to determine enzyme activity defined as the ratio of the AUC for each compound at each concentration normalized on the AUC in presence of DMSO matched concentration. Compound 5 strongly inhibited α -amylase catalyzed reaction with an IC_{50} of 40 μ M. (B) Anti- α -glucosidase activity. Kinetics of α -glucosidase reaction were measured in presence of each compound at concentrations ranging from 0.01–1000 μ M, and matched concentrations of DMSO solvent over a 40 min period. Areas under the curves (AUC) of OD (410 nm) vs. time were calculated and used to determine enzyme activity defined as the ratio of the AUC for each compound at each concentration normalized on the AUC in presence of DMSO matched concentration. Compound 5 nearly completely inhibited α -glucosidase catalyzed reaction with at all concentration tested. Compounds 1, 2 and 3 inhibited 50% of the reaction at 1 mM. For all assays, enzyme, no compound and inhibitor controls were included.

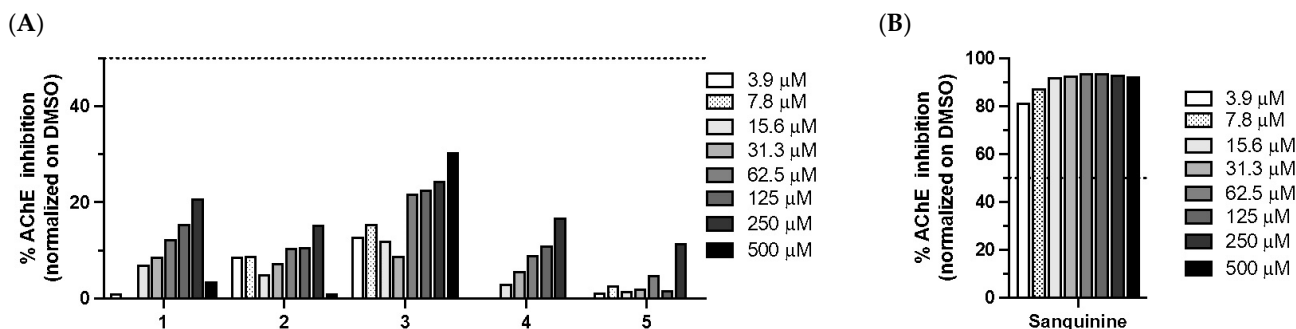


Figure 11. (A) Anti-AChE activity. AChE-catalyzed reaction was assessed in presence of each compound at concentrations ranging from 3.9–500 μ M and matched concentrations of DMSO after a 10 min incubation. The percentage of anti-AChE inhibition was calculated according to the following formula: $100 - [(E - S)/E \times 100]$, where E is the activity of the enzyme with matched concentrations of DMSO and S is the activity of the enzyme with the test sample. Compound 5 weakly inhibited AChE catalyzed reaction at high concentrations. Compounds 1, 2 and 4 displayed increasing inhibition from 15.6 μ M to 250 μ M up to 20%. Compound 3 was the most potent inhibitor of AChE, blocking 15–30% of enzyme activity as concentrations increased from 3.9 to 500 μ M. (B) Sanguinine (isolated from *C. jagus* [12]) was used as a positive control. For all assays, enzyme, no compounds and inhibition controls were included.

3.4. Antiviral Activity

Flavonoids and alkylamides have also been shown to exhibit antiviral activity [49]. Thus, we tested the anti-retroviral effect of compounds 1–5 using VSV-G pseudotyped HIV-1 particles. Interestingly, all compounds displayed anti-HIV-1 potential at 100 μ M (Figure 12A). Infection dropped from a mean of 8.16%, when cells were treated with DMSO, to means of 3.78, 4.03, 2.51, 2.76 and 2.12 %, when compounds 1 to 5 were added, respectively. These differences were statistically significant for compounds 3, 4 and 5. At 50 μ M, there was a 2.0, 2.4 and 1.6-fold decrease in infection when 3, 4 and 5 were added to the cell media, respectively. At lower concentrations only 3 and 4 decreased infection levels. Cytotoxicity

was verified following propidium iodide staining of THP-1 cells after an incubation of 72 h with compounds (Figure 12B). Compounds 1, 2 and 5 were weakly cytotoxic to THP-1 at 100 μ M with 8.74, 4.15 and 5.97% of PI⁺ cells, respectively. In total, 30–40% of cells treated with 3 and 4 were dead at all concentrations tested. Thus, although cytotoxicity could contribute to the observed antiviral activity, this experiment was performed using a single-cycle infection system, lessening the effect of cell loss on further cycles of viral replication. In conclusion, *C. biflorum* contains compounds with interesting anti-retroviral activity.

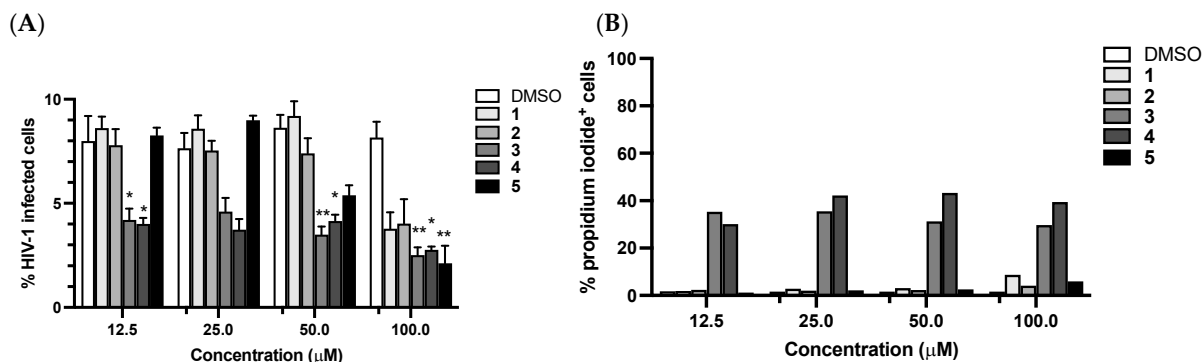


Figure 12. Antiviral activity of compounds 1–5. **(A)** THP-1 cells were treated with four concentrations (12.5, 25, 50 and 100 μ M) of each compound in triplicates and infected with VSV-G pseudotyped HIV-1_{GFP} at MOI = 1. Infection levels were assessed by flow cytometry 72 h post-infection. Bars show means with standard deviation. Kruskal–Wallis and uncorrected Dunn’s tests were performed to assess statistically significant differences between groups. * $p < 0.05$; ** $p < 0.01$. **(B)** Cell death (% propidium iodide⁺ cells) was measured by flow cytometry following PI staining 72 h post-treatment of THP-1 with 1, 2, 3, 4 and 5. Matched concentrations of DMSO were used as a negative control.

4. Discussion

Very few studies have been performed on *C. biflorum* (syn. *C. distichum*) from a chemical and applicative point of view. A phytochemical investigation on this Amaryllidaceae species was carried out on the ethanol extract of the whole plant using HPLC analysis. A new natural flavan, (2*S*)-4',7-dimethoxyflavan, was isolated together with the already known (2*S*)-4'-hydroxy-7-methoxyflavan, (2*R*)-4'-hydroxy-5,7-dimethoxyflavan, hippadine and hippacine. All the compounds, except the new flavan, showed moderate antibacterial activity [50]. Another study was carried out on the ethyl acetate extract from the whole plant of *C. biflorum* collected in Cameroon. Its phytochemical investigation allowed to isolate a new flavan-3-ol derivative, namely (2*R*,3*R*)-3-hydroxy-7-methoxy-3',4'-methylenedioxyflavan, together with (2*S*)-7-hydroxy-3',4'-methylenedioxyflavan, (2*R*,3*R*)-7-methoxy-flavan-3-ol, (2*S*)-7-hydroxy-3',4'-dimethoxyflavan, 3',7-dihydroxy-4'-methoxyflavan, 4',7-dimethoxy-3'-hydroxyflavan, farrerol, β -sitosterol, β -sitosterol-3-*O*- β -D-glucopyranoside, oleanolic acid, kaempferol, pancratistatin, lupeol, aurantiamide acetate, narciprimine and 2,3-dihydroxypropyl palmitate [29]. Some of them were also isolated from Muscari species among a series of homoisoflavanones containing the 3-benzylchroman-4-one skeleton [51].

Compounds 2 and 3 were characterized as 3-hydroxy-5,6,7-trimethoxy-3-(4-hydroxybenzyl)chroman-4-one (2), as 3-hydroxy-5,6,7-trimethoxy-3-(4-methoxybenzyl)chroman-4-one (3) and resulted to be the previously isolated homoisoflavanoids named urgineanins B (2) and A (3) [31].

No alkaloids were detected in the acid organic extract from bulbs of *C. biflorum*, object of the present study, using either the optimized extraction method [52] or the traditional extraction with ethanol by Soxhlet. However, the organic extract obtained with the latter method showed the presence of four homoisoflavanoids and one alkylamide. The racemate nature of compound 1 was never reported before; only the absolute configuration of its *p*-bromobezoyl derivative was previously determined by X-ray [53]

when configurations were also confirmed by ECD comparing their ECD spectra with those previously reported. Urgineanin A was previously reported to have antiproliferative activity at submicromolar concentration against ovarian carcinoma, melanoma and non-small lung cancer cells [31].

Compound **4** is also a racemic mixture and therefore its ECD spectrum could not be interpreted (Figure 4). This result differed from that previously reported for **4** when synthesized from the (3*R*)-3,9-dihydroeucomnalin and wrongly reported as *R* enantiomer [44]. Compound **4**, as well as the starting homoisoflavanoid above described for compound **1**, are a racemic mixture for the presence at C-3 of a proton which could exchange by keto-enol tautomerism.

Racemic natural products are rare and could be obtained from nonenzymatic reactions [54]. However, a chiral tertiary asymmetric carbon in α position to a carbonyl group is easily subject to racemization, as in compounds **1** and **4** [55]. To the best of our knowledge no literatures are available in which compounds **1** and **4** are reported as racemates. However, Sylao et al. 1990 [30] isolated for the first time homoisoflavanoid **1** (named compound **12**) along with other compounds from *Scilla nervosa* subsp. *rigidifolia*. The previously undescribed homoisoflavanoids were also fully characterized by spectroscopic data (UV, IR, ^1H and ^{13}C NMR and EIMS) but any experiments to determine the absolute configuration of the chiral compounds were neither carried out nor discussed. Among the homoisoflavanoids isolated as new compounds, only for some of them, including **12** (=1), was reported the optical specific activity. Based on our experience, these results did not surprise us as the speed of inversion of the configuration of carbon 3, due to the keto-enol tautomerism, could depend on the properties of the solution in which the measurement of the optical rotational power is measured and not from the plant source. Thus, it is possible to have also a scalemic mixture that has optical activity according to the percentage of the predominant enantiomer, as reported for some optical active homoisoflavanoids by Sylao et al. 1999 [30]. Scalemic mixtures of two enantiomers are reported in the literature for some different secondary metabolites isolated from several sources as i.e.: phantasmidine, an alkaloid found to be a 4:1 scalemic mixture, enriched in the (2*aR*,4*aS*,9*aS*) enantiomer isolated from the poison frog *Epipedobates anthonyi* [56]; α -pinene, 1-octen-3-ol linalool found as a scalemic mixture of 34% (*R*)-(+ to 66% (*S*)-(-), 95% (*R*)-(-) to 5% (*S*)-(+), 96% (*R*)-(-) to 4% (*S*)-(+ when isolated from the edible wild mushroom *Tricholoma magnivelare* [57]; a furoic acid derivative, containing a chiral center in benzylic position, was found to be a scalemic mixture with an excess of the (*S*) enantiomer, when obtained from the endophytic fungus *Coniothyrium* sp. was isolated from leaves of *Quercus robur* [58]; six pairs of new 6-monosubstituted dihydrobenzophenanthridine alkaloids were separated as scalemic mixtures from the aerial part of *Chelidonium majus*, a plant belonging to the Papaveraceae family, which is widely used in Chinese folk medicine [59].

Compounds **5**, the alkylamide, was isolated for the first time, together with *N*-*trans*-feruloyl octopamine, *N*-*trans*-*p*-coumaroyl octopamine, vanillin, isoscapoletin, ethyl caffeate, ferulic acid and *p*-aminonenzaldehyde, from the eggplant roots [60]. Successively, **5** was also isolated together with close alkylamides from stem parts of *Annona montana* (Annonaceae) and showed significant inhibition of rabbit platelets aggregation induced by thrombin, arachidonic acid, collagen and PAF (platelet-activating factor) and selective cytotoxicity against the P-388 cell line [61,62]. Compound **5** was also found together with other close alkylamides and several phenolic compounds in the methanol extract of basil, lemon thyme, mint, oregano, rosemary, sage and thyme showing antioxidant and anti-inflammatory activities [63].

Alkylamides are a group of bioactive natural compounds widely distributed in plant families and characterized by broad structural variability and a plethora of important biological activities, such as immunomodulatory, antimicrobial, antiviral, larvicidal, insecticidal, diuretic, pungent, analgesic, cannabimimetic and antioxidant activities. Furthermore, they have reinforced the efficacy of antibiotics and inhibited prostaglandin

biosynthesis, RNA synthesis and arachidonic acid metabolism [64]. In addition, alkylamides accumulate in rice plants as a defense against the harmful *Cochliobolus miyabeanus* and *Xanthomonas oryzae* pathogens [65].

Flavonoids were also reported to have cytotoxic and anticancer activity, although this aspect was not deeply addressed [66,67]. We found that all tested metabolites were toxic for cancer cells, in a dose- and time-dependent way, although the degree of inhibition of cell viability was cell-type specific. Remarkably, homoisoflavanoids **1**, **3** and **4** were more effective on A431 and HeLa cells compared to immortalized but not transformed HaCaT, thus suggesting that cancer cells were more sensitive to homoisoflavanoid cytotoxicity. The increase of nuclear γ H2AX foci upon treatment with metabolites **3** and **4** strongly suggests the occurrence of DNA damage by double-strand breaks (DSBs). However, both **3** and **4** did not increase the production of ROS indicating that a molecular mechanism different from ROS generation is responsible for the observed DNA damage induced by **3** and **4**. Additional experiments are needed to precisely define the molecular mechanism. Moreover, we also detected the signal of cleaved PARP-1 (89 kDa) by immunoblot while the level of the cell cycle inhibitor p21WAF, which causes cell growth arrest preventing the induction of apoptosis, was reduced, thus confirming that the decrease of cell viability was due to cell death rather than cell cycle arrest.

Isolated compounds displayed additional interesting biological properties. As previously described but isolated for the first time in a *Crinum* species, **5** is a strong anti- α -glucosidase and anti- α -amylase inhibitor. Recently, the efficacy of flavonoids for type 2 diabetes mellitus (T2DM) was shown in clinical therapies. T2DM is a metabolic disorder associated with the overproduction of free radicals and oxidative stress. Diabetes is increasing exponentially, and the World Health Organization estimates that by the year 2030, it could be the seventh cause of death worldwide [68,69]. The flavonoids appear to play a role in multiple processes involved in T2DM [70,71] such as the regulation of glucose metabolism, hepatic enzymes activities and a lipid profile [72]; thus, studies on nutritional flavonoids to manage diabetes and its complications are currently in progress [73].

Interestingly, the *N-p*-coumaroyltyramine alkylamide (**5**) was selectively cytotoxic against A431 and HeLa cancer cells while it protected immortalized HaCaT cells against oxidative stress induced by hydrogen peroxide. This result is highly relevant for a potential application of **5** in anticancer therapy. Furthermore, we observed the antidiabetic properties of **5** and the anti-acetylcholinesterase activity in compounds **1–4**. Homoisoflavanoids are also known for their anti-acetylcholinesterase properties, a key enzyme to Alzheimer's disease development [74]. We detected anti-acetylcholinesterase activity in compound **3**, a property that had not yet been reported for urgeineanins A and B, to our knowledge. Finally, as we recently showed that *Crinum jagus* contained antiviral compounds, we measured the antiretroviral activity of isolated compounds. We report that all compounds possessed anti-retroviral potential, **3** and **4** being the most potent inhibitors.

5. Conclusions

Four homoisoflavanoids and one alkylamide were isolated and characterized from *C. biflorum*, an Amaryllidaceae plant used in African traditional medicine, collected in Senegal. Flavonoids **1**, **3** and **4** showed promising anticancer properties being cytotoxic at low micromolar concentrations towards HeLa and A431 human cancer cell lines. The *N-p*-coumaroyltyramine (**5**) was selectively toxic to A431 and HeLa cancer cells, while it protected immortalized HaCaT cells against oxidative stress induced by hydrogen peroxide. Compounds **1–4** also inhibited acetylcholinesterase activity, with compound **3** being the most potent. The anti-amylase and the strong anti-glucosidase activity of compound **5** were confirmed. This study extends the chemical library of compounds that can be a potential candidate for the treatment of cancer, viral infections, diabetes and Alzheimer's disease.

Author Contributions: Conceptualization, M.M., A.E., I.D.-P. and V.C.; M.M., M.K., A.D., R.D.L., N.M., S.K., A.T., M.S. formal analysis and investigation; writing—original draft preparation, M.M., A.E., I.D.-P. and V.C.; writing—review and editing, M.M., A.E., I.D.-P. and V.C. All authors have read and agreed to the published version of the manuscript.

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References

1. Newman, D.J.; Cragg, G.M. Natural products as sources of new drugs over the nearly four decades from 01/1981 to 09/2019. *J. Nat. Prod.* **2020**, *83*, 770–803.
2. Hoshino, O. The amaryllidaceae alkaloids. *Alkaloids* **1998**, *51*, 323–376.
3. Evidente, A.; Kornienko, A. Anticancer evaluation of structurally diverse Amaryllidaceae alkaloids and their synthetic derivatives. *Phytochem. Rev.* **2009**, *8*, 449–459.
4. Nair, J.J.; van Staden, J. Pharmacological and toxicological insight to the South Africa Amaryllidaceae. *Food Chem. Toxicol.* **2013**, *62*, 262–275.
5. Jin, Z.; Yao, G. Amaryllidaceae and Scelletium alkaloids. *Nat. Prod. Rep.* **2019**, *36*, 1462–1488.
6. Nair, J.J.; Bastida, J.; Codina, C.; Viladomat, F.; van Staden, J. Alkaloids of the South African Amaryllidaceae: A review. *Nat. Prod. Commun.* **2013**, *8*, 1335–1350.
7. Christenhusz, M.J.; Byng, J.W. The number of known plants species in the world and its annual increase. *Phytotaxa* **2016**, *26*, 201–217.
8. Bastida, J.; Berkov, S. Amaryllidaceae Alkaloids. *Molecules* **2020**.
9. Masi, M.; Di Lecce, R.; Cimmino, A.; Evidente, A. Advances in the chemical and biological characterization of Amaryllidaceae alkaloids and natural analogues isolated in the last decade. *Molecules* **2020**, *25*, 5621.
10. Ghosal, S.; Saini, K.S.; Razdan, S. *Crinum* alkaloids: Their chemistry and biology. *Phytochemistry* **1985**, *24*, 2141–2156.
11. Kornienko, A.; Evidente, A. Chemistry, biology, and medicinal potential of narciclasine and its congeners. *Chem. Rev.* **2008**, *108*, 1982–2014.
12. Ka, S.; Masi, M.; Merindol, N.; Di Lecce, R.; Plourde, M.B.; Seck, M.; Marcin, G.; Pescitelli, G.; Desgagne-Penix, I.; Evidente, A. Gigantelline, gigantellinine and gigancrinine, cherylline-and crinine-type alkaloids isolated from *Crinum jagus* with anti-acetylcholinesterase activity. *Phytochemistry* **2020**, *175*, 112390.
13. Ka, S.; Merindol, N.; Sow, A.A.; Singh, A.; Landelouci, K.; Plourde, M.B.; Pepin, G.; Masi, M.; Di Lecce, R.; Evidente, A.; et al. Amaryllidaceae alkaloid cherylline inhibits the replication of dengue and Zika viruses. *Antimicrob. Agents Chemother.* **2021**, *65*, 9, doi:10.1128/AAC.00398-21.
14. Ibrakav, A.S.; Akinfewa, A.O.; Hussein, A.A. A comprehensive review on non alkaloids chemical constituents from Amaryllidaceae. *South Afr. J. Bot.* **2020**, In press.
15. Dewick, P.M. *Medicinal Natural Products: A Biosynthetic Approach*; John Wiley & Sons: Chichester, UK, 2009.
16. Osbourn, A.E.; Lanzotti, V. *Plant-Derived Natural Products Dordrecht*; Springer: Berlin, Germany, 2009; pp. 361–384.
17. Abd El-Hafiz, M.A. Aliphatic hydroxyketones from *Crinum augustum*. *Phytochemistry* **1991**, *30*, 3127–3129.
18. Watt, J.M.; Breyer-Brandwijk, M.G. The medicinal and poisonous plants of Southern and Eastern Africa. *Kew Bull.* **1964**, *17*, 463.
19. Likhitwitayawuid, K.; Ruangrunsi, N.; Cordell, G.A. Amabiloside, a new glycoside from *Crinum amabile*. *Nat. Prod. Lett.* **1993**, *3*, 1–4.
20. Nam, N.H.; Jae, Y.Y. NF- κ B Inhibitory activities of the methanol extracts and some constituents therein of some Vietnamese medicinal plants. *Sci. Pharm.* **2009**, *77*, 389–400.
21. Jin, A.; Li, X.; Zhu, Y.Y.; Yu, H.Y.; Pi, H.F.; Zhang, P.; Ruan, H.L. Four new compounds from the bulbs of *Lycoris aurea* with neuroprotective effects against CoCl₂ and H₂O₂-induced SH-SY5Y cell injuries. *Arch. Pharm. Res.* **2014**, *37*, 315–323.
22. Sun, Q.; Zhang, W.D.; Shen, Y.H.; Zhang, C.; Li, H.L. A new phenolic compound from *Crinum asiaticum* L. *Chin. Chem. Lett.* **2008**, *19*, 447–449.
23. Nkanwen, E.R.S.; Gatsing, D.; Ngamga, D.; Fodouop, S.P.C.; Tane, P. Antibacterial agents from the leaves of *Crinum purpurascens* herb (Amaryllidaceae). *Afric. Health Sci.* **2009**, *9*, 264–269.
24. Khoi, N.M.; Dat, N.T.; Na, M.K.; Thuong, P.T.; Min, B.S.; Bae, K.H. Cytotoxic activity of parthenin, a sesquiterpene isolated from a *Crinum ensifolium*. *Nat. Prod. Sci.* **2011**, *17*, 100–103.

25. Abd El Hafiz, M.A.; Ramadan, M.A.; Jung, M.L.; Beck, J.P.; Anton, R. Cytotoxic activity of Amaryllidaceae alkaloids from *Crinum augustum* and *Crinum bulbisperrum*. *Planta Med.* **1991**, *57*, 437–439.
26. Abdel-Halim, O.B.; Marzouk, A.M.; Mothana, R.; Awadh, N. A new tyrosinase inhibitor from *Crinum yemense* as potential treatment for hyperpigmentation. *Die Pharm. Int. J. Pharm. Sci.* **2008**, *63*, 405–407.
27. Masi, M.; Mubaiwa, B.; Cimmino, A.; Van Otterlo, W.A.L.; Green, I.R.; Evidente, A. First isolation of acetovanillone and piceol from *Crinum buphanoides* and *Crinum graminicola* (I. Verd.) Amaryllidaceae. *South Afric. J. Bot.* **2018**, *114*, 37–39.
28. Burkill, H.M. *The Useful Plants of West Tropical Africa*; Royal Botanic Gardens: London, UK, 1985; Volume 1, pp. 163.
29. Kianfé, B.Y.; Teponno, R.B.; Kühlborn, J.; Tchuenguem, R.T.; Ponou, B.K.; Helaly, S.E.; Dzoyemd, J.P.; Opatz, T.; Tapondjou, L.A. Flavans and other chemical constituents of *Crinum biflorum* (Amaryllidaceae). *Biochem. Syst. Ecol.* **2019**, *87*, 103953.
30. Silayo, A.; Ngadjui, B.T.; Abegaz, B.M. Homoisoflavonoids and stilbenes from the bulbs of *Scilla nervosa* subsp. *rigidifolia*. *Phytochemistry* **1999**, *52*, 947–955.
31. Dai, Y.; Harinantenaina, L.; Brodie, P.J.; Goetz, M.; Shen, Y.; TenDyke, K.; Kingston, D.G. Antiproliferative homoisoflavonoids and bufatrienolides from *Urginea depressa*. *J. Nat. Prod.* **2013**, *76*, 865–872.
32. Sidwell, W.T.L.; Tamm, C. The homo-isoflavones III). Isolation and structure of 4'-O-methyl-punctatin, autumnalin and 3,9-dihydro-autumnalin. *Tetrahedron Lett.* **1970**, *11*, 475–478.
33. Bhatti, M.K.; Akhtar, F.; Choudhary, M.I. Alkaloids of *Fumaria indica*. *Phytochemistry* **1992**, *31*, 2869–2872.
34. Altomare, A.; Burla, M.C.; Camalli, M.; Cacarano, G.L.; Giacobuzzo, C.; Guagliardi, A.; Moliterni, A.G.G.; Polidori, G.; Spagna, R. SIR97: A new tool for crystal structure determination and refinement. *J. Appl. Crystallogr.* **1999**, *32*, 115–119.
35. Sheldrick, G.M. Crystal structure refinement with SHELXL. *Acta Crystallogr. Sect. C Struct. Chem.* **2015**, *71*, 3–8.
36. Farrugia, L.J. WinGX and ORTEP for Windows: An update. *J. Appl. Crystallogr.* **2012**, *45*, 849–854.
37. Macrae, C.F.; Bruno, I.J.; Chisholm, J.A.; Edgington, P.R.; McCabe, P.; Pidcock, E.; Rodriguez-Monge, L.; Taylor, R.; van de Streek, J.; Wood, P.A. Mercury CSD 2.0-new features for the visualization and investigation of crystal structures. *J. Appl. Crystallogr.* **2008**, *41*, 466–470.
38. Amoresano, A.; Di Costanzo, A.; Leo, G.; Di Cunto, F.; La Mantia, G.; Guerrini, L.; Calabrò, V. Identification of Δ Np63 α protein interactions by mass spectrometry. *J. Proteome Res.* **2010**, *9*, 2042–2048.
39. Vivo, M.; Fontana, R.; Ranieri, M.; Capasso, G.; Angrisano, T.; Pollice, A.; Calabrò, V.; La Mantia, G. p14ARF interacts with the focal adhesion kinase and protects cells from anoikis. *Oncogene* **2017**, *36*, 4913–4928.
40. Sangermano, F.; Masi, M.; Vivo, M.; Ravindra, P.; Cimmino, A.; Pollice, A.; Evidente, A.; Calabrò, V. Higginsianins A and B, two fungal diterpenoid α -pyrones with cytotoxic activity against human cancer cells. *Toxicol. In Vitro* **2019**, *61*, 104614.
41. di Martino, O.; Troiano, A.; Guarino, A.M.; Pollice, A.; Vivo, M.; La Mantia, G.; Calabrò, V. DNp63a controls YB-1 protein stability: Evidence on YB-1 as a new player in keratinocyte differentiation. *Genes Cells* **2016**, *21*, 648–660.
42. Zhang, J.; Zhao, S.; Yin, P.; Yan, L.; Han, J.; Shi, L.; Zhou, X.; Liu, Y.; Ma, C. α -Glucosidase inhibitory activity of polyphenols from the burs of *Castanea mollissima* Blume. *Molecules* **2014**, *19*, 8373–8386.
43. Ellman, G.L.; Courtney, K.D.; Andres Featherstone, R.M. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol.* **1961**, *7*, 88–95.
44. Borgonovo, G.; Caimi, S.; Morini, G.; Scaglioni, L.; Bassoli, A. Taste-active compounds in a traditional Italian food: 'Lampascioni'. *Chem. Biodiver.* **2008**, *5*, 1184–1194.
45. Okuyama, T.; Shibata, S.; Hoson, M.; Kawada, T.; Osada, H.; Noguchi, T. Effect of oriental plant drugs on platelet aggregation; III. Effect of Chinese drug "Xiebai" on human platelet aggregation. *Planta Med.* **1986**, *52*, 171–175.
46. Rogakou, E.P.; Pilch, D.R.; Orr, A.H.; Ivanova, V.S.; Bonner, W.M. DNA doublestranded breaks induce histone H2AX phosphorylation on serine 139. *J. Biol. Chem.* **1998**, *273*, 5858–5868.
47. Dickey, J.S.; Redon, C.E.; Nakamura, A.J.; Baird, B.J.; Sedelnikova, O.A.; Bonner, W.M. H2AX: Functional roles and potential applications. *Chromosoma* **2009**, *118*, 683–692.
48. Nishioka, T.J.; Watanabe, J.; Kawabata, J.R.; Niki, I. Isolation and activity of *N-p*-coumaroyltyramine, an α -glucosidase inhibitor in welsh onion (*Allium fistulosum*). *Biosci. Biotech. Biochem.* **1997**, *61*, 1138–1141.
49. Pasetto, S.; Pardi, V.; Murata, R.M. Anti-HIV-1 activity of flavonoid myricetin on HIV-1 infection in a dual-chamber in vitro model. *PLoS ONE* **2014**, *9*, e115323.
50. Koagne, R.R.; Annang, F.; de la Cruz, M.; Bitchagno, G.T.M.; Perez-Victoria, I.; Konga, I.S.; Vincente, F.; Reyes, F.; Tane, P. Antibacterial activity of flavans from *Crinum distichum*. *Nat. Prod. Commun.* **2018**, *13*, 1637–1638.
51. Barone, G.; Corsaro, M.M.; Lanzetta, R.; Parrilli, M. Homoisoflavanones from *Muscari neglectum*. *Phytochemistry* **1988**, *27*, 921–923.
52. Evidente, A.; Iasiello, I.; Randazzo, G. An improved method for the large-scale preparation of lycorine. *Chem. Ind.* **1984**, *9*, 348–349.
53. Giordano, F.; Lanzetta, R. Structure and absolute configuration of (–)-(3R)-5, 7-dimethoxy-3,9-dihydro-4'-eucomminalinyl *p*-bromobenzoate: An uncommon case. *Acta Crystallogr. Sect. C Struct. Chem.* **1989**, *45*, 1603–1606.
54. Zask, A.; Ellestad, G. Biomimetic syntheses of race-mic natural products. *Chirality* **2018**, *30*, 157–164.
55. Di Lecce, R.; Masi, M.; Linaldeddu, B.T.; Pescitelli, G.; Maddau, L.; Evidente, A. Bioactive secondary metabolites produced by the emerging pathogen *Diplodia olivarum*. *Phytopathol. Mediterr.* **2021**, *60*, 129–138.
56. Fitch, R.W.; Snider, B.B.; Zhou, Q.; Foxman, B.M.; Pandya, A.A.; Yakel, J.L.; Olson, T.T.; Al-Muhtasib, N.; Xiao, Y.; Welch, K.D.; et al. Absolute configuration and pharmacology of the poison frog alkaloid phantasmidine. *J. Nat. Prod.* **2018**, *81*, 1029–1035.

57. Murray, A.F.; Moore, A.J.; Munafo, J.P., Jr. Key odorants from the american matsutake, *Tricholoma magnivelare*. *J. Agric. Food Chem.* **2020**, *68*, 9768–9775.
58. Tran-Cong, N.M.; Mándi, A.; Király, S.B.; Kurtán, T.; Lin, W.; Liu, Z.; Proksch, P. Furoic acid derivatives from the endophytic fungus *Coniothyrium* sp. *Chirality* **2020**, *32*, 605–610.
59. Deng, A.J.; Zhang, H.J.; Li, Q.; Li, Z.H.; Zhang, Z.H.; Wu, L.Q.; Li, L.; Qin, H.L. Six salemic mixtures of 6-monosubstituted dihydrobenzophenanthridine alkaloids from *Chelidonium majus* and optically active structures of enantiomers. *Phytochemistry* **2017**, *144*, 159–170.
60. Yoshihara, T.; Takamatsu, S.; Sakamura, S. Three new phenolic amides from the roots of eggplant (*Solanum melongena* L.). *Agric. Biol. Chem.* **1978**, *42*, 623–627.
61. Wu, Y.C.; Chang, G.Y.; Ko, F.N.; Teng, C.M. Bioactive constituents from the stems of *Annona montana*. *Planta Med.* **1995**, *61*, 146–149.
62. Barbalho, S.M.; de Alvares Goulart, R.; Vasques Farinazzi-Machado, F.M.; da Silva Soares de Souza, M.; Cincotto dos Santos Bueno, P.; Landgraf Guiguer, E.; Araújo, A.C.; Groppo, M. *Annona* sp: Plants with multiple applications as alternative medicine—A review. *Curr. Bioact. Compd.* **2012**, *8*, 277–286.
63. Park, J.B. Identification and quantification of a major anti-oxidant and anti-inflammatory phenolic compound found in basil, lemon thyme, mint, oregano, rosemary, sage, and thyme. *Int. J. Food Sci. Nutr.* **2011**, *62*, 577–584.
64. Rios, M.Y. Natural alkamides: Pharmacology, chemistry, and distribution. In *Drug Discovery: Research in Pharmacognosy*; Books on Demand: Norderstedt, Germany, 2012; pp. 107–144.
65. Morimoto, N.; Ueno, K.; Teraishi, M.; Okumoto, Y.; Mori, N.; Ishihara, A. Induced phenylamide accumulation in response to pathogen infection and hormone treatment in rice (*Oryza sativa*). *Biosci. Biotech. Bioch.* **2018**, *82*, 407–416.
66. Castelli, M.V.; López, S.N. Homoisoflavonoids: Occurrence, biosynthesis, and biological activity. In *Studies in Natural Products Chemistry*; Elsevier: Amsterdam, The Netherlands, 2017; pp. 315–354.
67. Abegaz, B.M.; Kinf, H.H. Naturally occurring homoisoflavonoids: Phytochemistry, biological activities, and synthesis (Part II). *Nat. Prod. Commun.* **2019**, *14*, doi:10.1177/1934578x19845813.
68. Danaei, G.; Finucane, M.M.; Lu, Y.; Singh, G.M.; Cowan, M.J.; Paciorek, C.J. Global burden of metabolic risk factors of chronic diseases collaborating, G. National, regional, and global trends in fasting plasma glucose and diabetes prevalence since 1980: Systematic analysis of health examination surveys and epidemiological studies with 370 country-years and 2.7 million participants. *Lancet* **2011**, *378*, 3140.
69. World Health Organization. *Definition and Diagnosis of Diabetes Mellitus and Intermediate Hyperglycemia: Report of a WHO/IDF Consultation*; World Health Organization: Geneva, Switzerland, 2006; pp. 1–50.
70. Mohan, S.; Nandhakumar, L. Role of various flavonoids: Hypotheses on novel approach to treat diabetes. *J. Med. Hypot. Ideas* **2014**, *8*, 1–6.
71. Testa, R.; Bonfigli, A.R.; Genovese, S.; De Nigris, V.; Ceriello, A. The possible role of flavonoids in the prevention of diabetic complications. *Nutrients* **2016**, *8*, 310.
72. AL-Ishaq, R.K.; Abotaleb, M.; Kubatka, P.; Kajo, K.; Büsselberg, D. Flavonoids and their anti-diabetic effects: Cellular mechanisms and effects to improve blood sugar levels. *Biomolecules* **2019**, *9*, 430.
73. Vinayagam, R.; Xu, B. Antidiabetic properties of dietary flavonoids: A cellular mechanism review. *Nutr. Metab.* **2015**, *12*, 60.
74. Li, Y.; Qiang, X.; Luo, L.; Yang, X.; Xiao, G.; Zheng, Y.; Cao, Z.; Sang, Z.; Su, F.; Deng, Y. Multitarget drug design strategy against Alzheimer's disease: Homoisoflavonoid Mannich base derivatives serve as acetylcholinesterase and monoamine oxidase B dual inhibitors with multifunctional properties. *Bioorg. Med. Chem.* **2017**, *25*, 714–726.