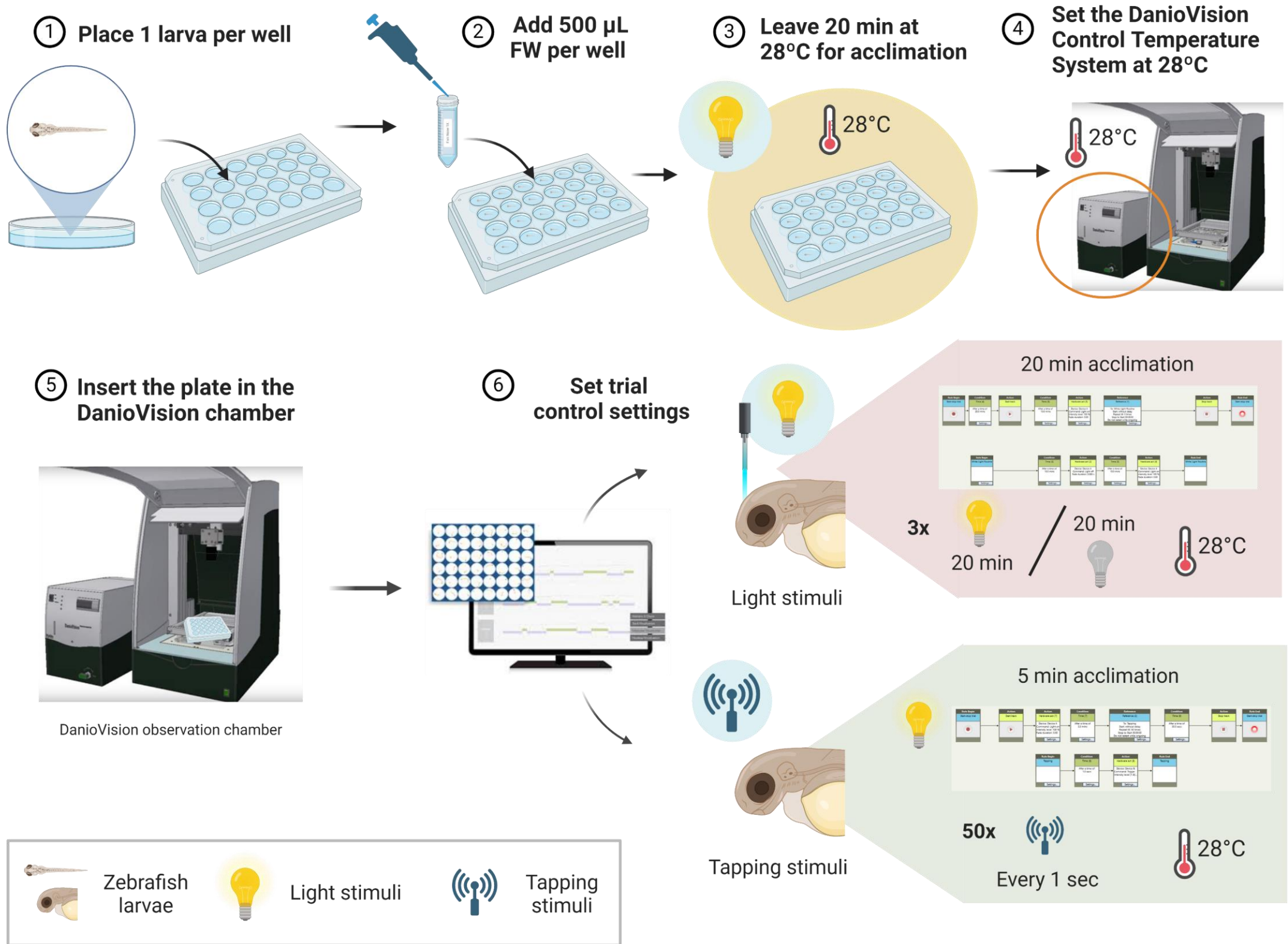
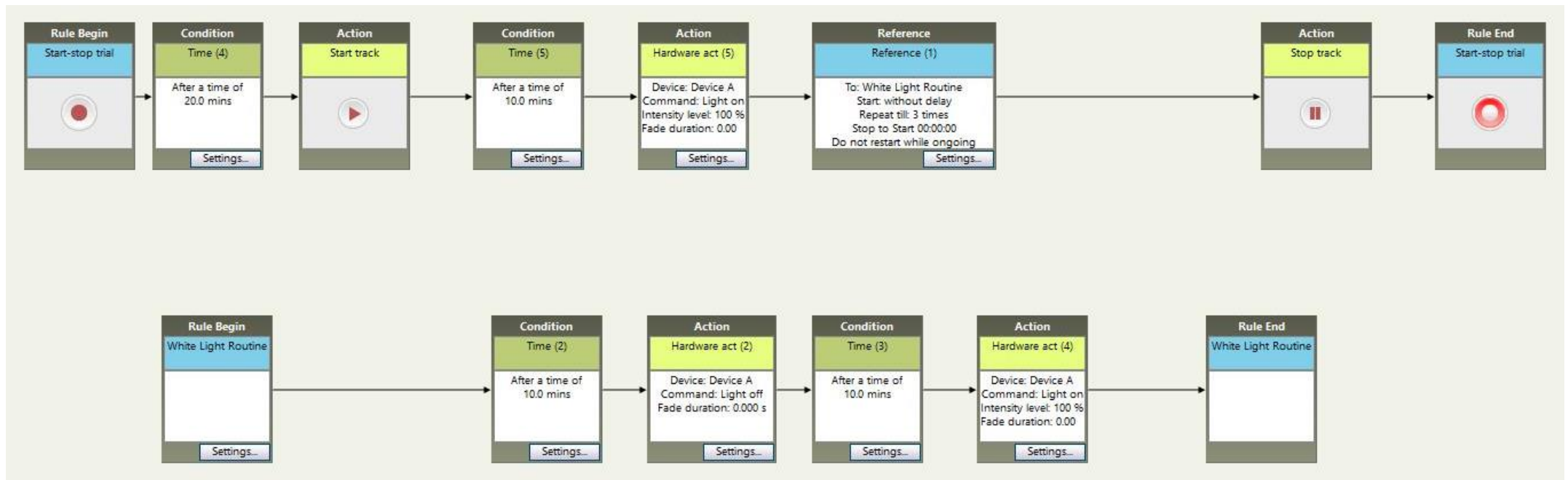


A



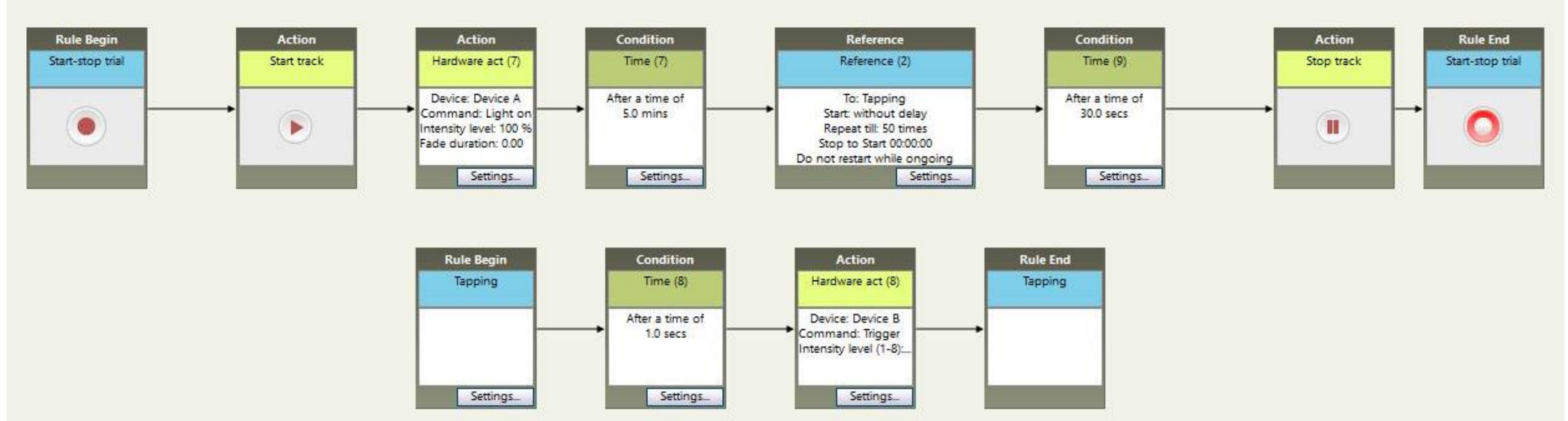
B

Light stimuli



C

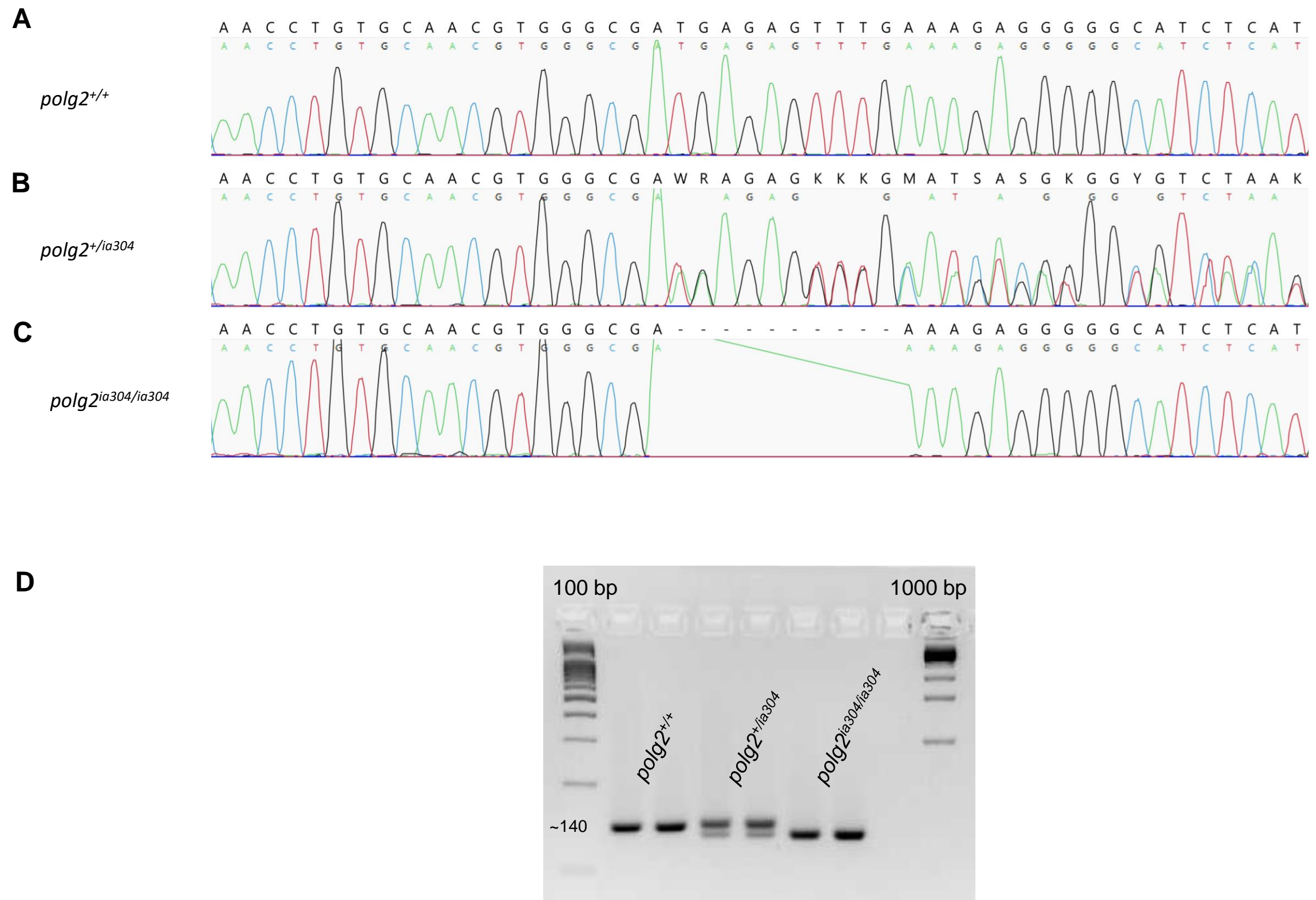
Tapping stimuli



Suppl. Figure 1: DanioVision protocol setting for behavioural analysis in zebrafish larvae

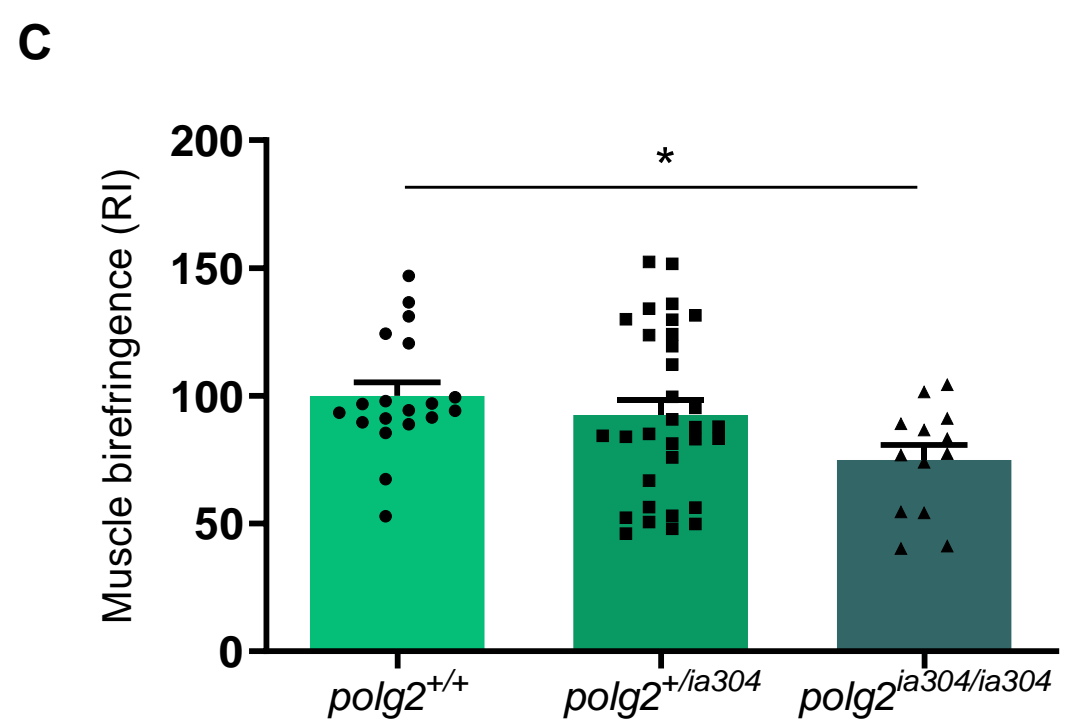
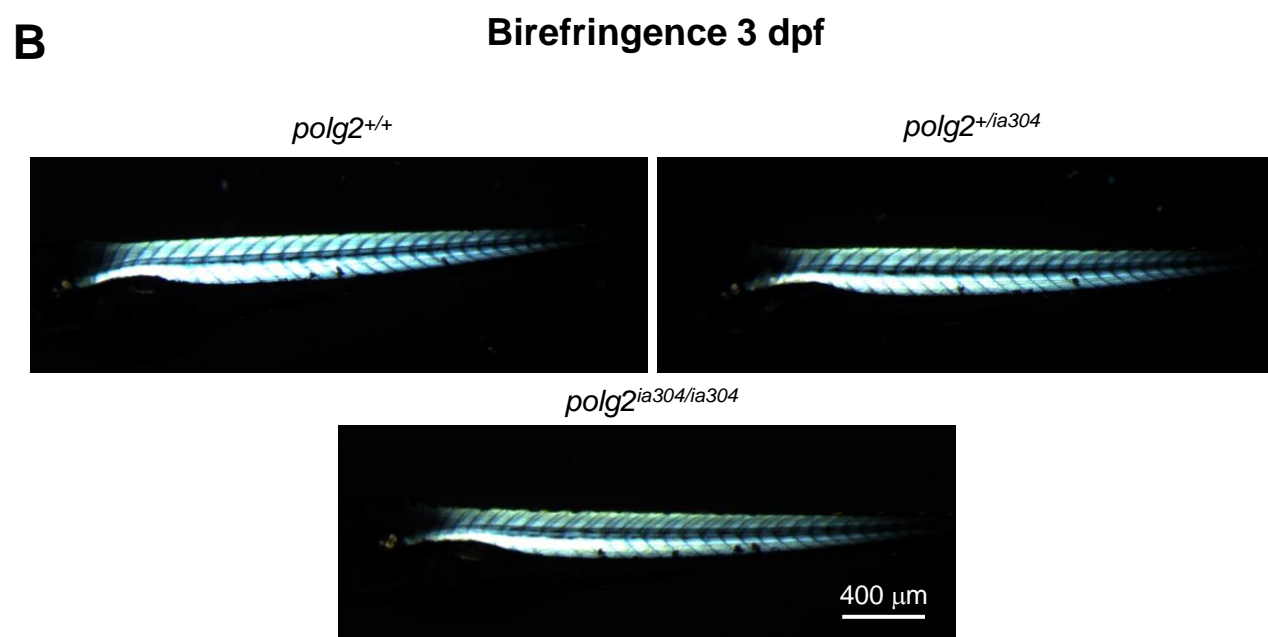
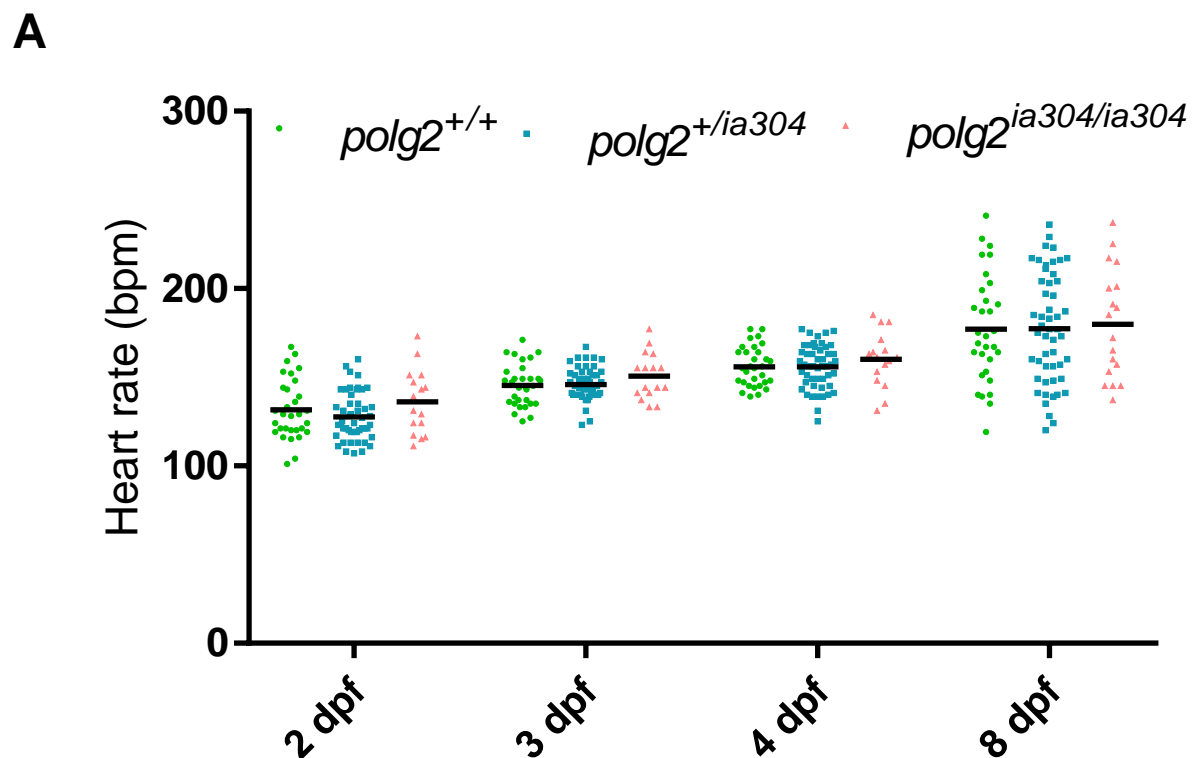
(A) Flowchart protocol for behavioural analysis in zebrafish larvae using a 24-well plate. Temperature was set to 28°C with the DanioVision Temperature Control Unit. FW: Fish Water. Created with BioRender.com.

(B) Trial Control Settings for the light stimuli protocol. (C) Trial Control Settings for the tapping stimuli protocol.



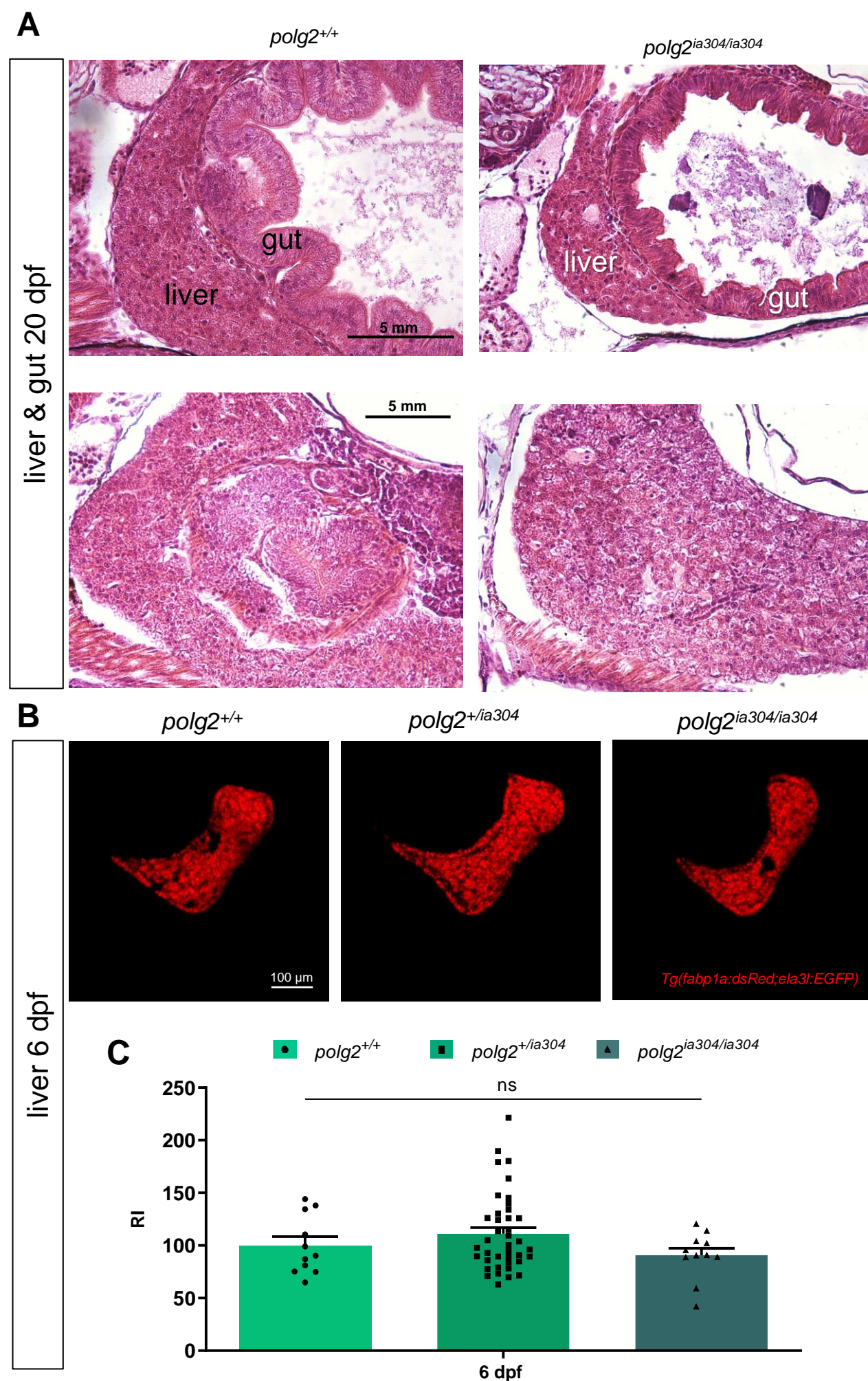
Suppl. Figure 2: Genotyping of the zebrafish *polg2*^{ia304} mutant line

(A-C) Chromatograms corresponding to *polg2*^{+/+}(A), *polg2*^{+/ia304} (B) and *polg2*^{ia304/ia304} (C) individuals, aligned with SeqMan Ultra, DNASTAR Lasergene. (D) Representative gel image of PCR genotyping using genomic DNA from tail fins of larvae from a cross between *polg2*^{+/ia304} heterozygotes.



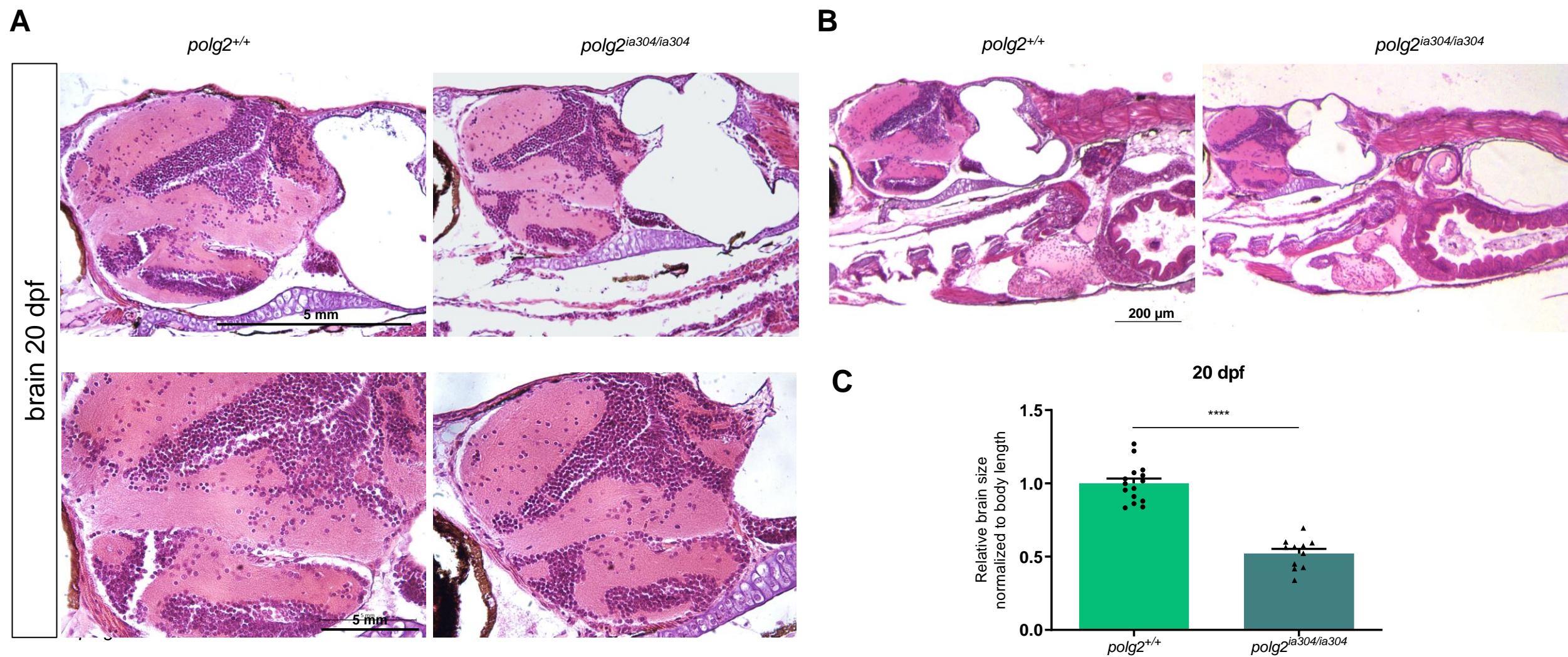
Suppl. Figure 3: Analysis of heart rate and skeletal muscle organization in *polg2* mutant larvae

(A) Heart rate measurements in *polg2*^{+/+}, *polg2*^{+/ia304} and *polg2*^{ia304/ia304} individuals at 2, 3, 4 and 8 dpf. Data are reported as heartbeats from 3 biological replicates and analysed by two-way ANOVA; *polg2*^{+/+} 2 dpf (n=30), *polg2*^{+/ia304} 2 dpf (n=50), *polg2*^{ia304/ia304} 2 dpf (n=17); *polg2*^{+/+} 3 dpf (n=30), *polg2*^{+/ia304} 3 dpf (n=50), *polg2*^{ia304/ia304} 3 dpf (n=16); *polg2*^{+/+} 4 dpf (n=30), *polg2*^{+/ia304} 4 dpf (n=50), *polg2*^{ia304/ia304} 4 dpf (n=16); *polg2*^{+/+} 8 dpf (n=31), *polg2*^{+/ia304} 8 dpf (n=51), *polg2*^{ia304/ia304} 8 dpf (n=20). (B) Light microscopy images of muscle birefringence in wt, heterozygous and homozygous *polg2*^{ia304} embryos at 3 dpf (scale bar: 400 μm). (C) Quantification of muscle birefringence in the three genotypes. Values from 3 independent biological replicates are shown as RI: Relative Intensity ± SEM and analysed by Kruskal-Wallis test followed by Dunn's multiple comparisons test; *polg2*^{+/+} (n=19), *polg2*^{+/ia304} (n=32), *polg2*^{ia304/ia304} (n=13); * p<0.05.



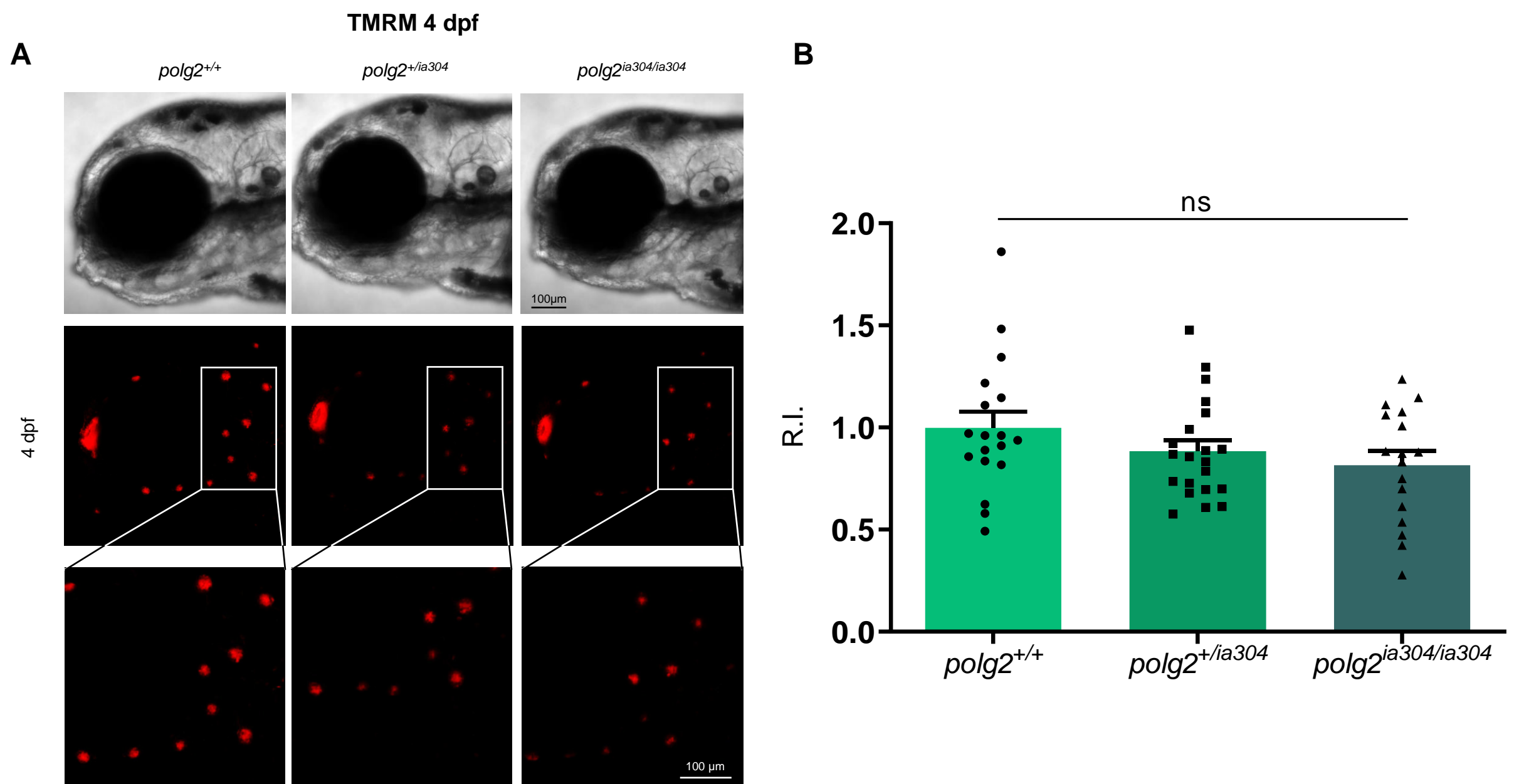
Suppl. Figure 4: Histological analysis of liver and gut in *polg2* mutants at 20 dpf.

(A) Histological analysis of gut and liver in 20 dpf zebrafish larvae. No significant alterations were found regarding their size (being isometric) and composition (scale bar: 5 mm). (B) Confocal images of the liver-expressed *Tg(fabp1a:dsRed;elaA:EGFP)^{gz15}* transgene at 6 dpf (scale bar 100 μ m). (C) Scatter-plot showing the relative quantification of the liver-expressed transgene in the three genotypes at 6 dpf. Data are expressed as the mean \pm SEM and analysed by Kruskal-Wallis test followed by Dunn's test for multiple comparisons; *polg2*^{+/+} (n=11), *polg2*^{+/ia304} (n=40), *polg2*^{ia304/ia304} (n=11).



Suppl. Figure 5: Histological analysis of brain in *polg2* mutants at 20 dpf

(A) Histological sections of *polg2*^{+/+} and *polg2*^{ia304/ia304} brain at 20 dpf, revealing allometric reduction of the brain size in mutants (scale bar: 5 mm). (B) Histological section at lower magnification of 20 dpf *polg2*^{+/+} and *polg2*^{ia304/ia304} larvae (scale bar: 200 μm). (C) Scatter-plot showing the quantification of brain size normalized to body length from different sections in *polg2*^{+/+} and *polg2*^{ia304/ia304} at 20 dpf larvae. Values are reported as the mean ± SEM and analysed by unpaired t-test; *polg2*^{+/+} (n=15), *polg2*^{ia304/ia304} (n=11); **** p<0.0001.



Suppl. Figure 6: Analysis of mitochondrial membrane potential in *polg2* mutant larvae.

(A) Fluorescence imaging of TetraMethylRhodamine Methyl ester (TMRM) accumulation in active mitochondria, monitored in the head of *polg2*^{+/+}, *polg2*^{+/ia304} and *polg2*^{ia304/ia304} individuals at 4 dpf (scale bar: 100 μm). (B) Quantification of mitochondrial TMRM accumulation in zebrafish embryos at 4 dpf. Data are expressed as mean ± SEM, analysed by ordinary One-way ANOVA and corrected by Tukey's multiple comparisons test; *polg2*^{+/+} (n=18), *polg2*^{+/ia304} (n=21), *polg2*^{ia304/ia304} (n=17); RI: Relative Intensity.

Oligomer name	Gene / Accession No.	Type	Sequence (5' - 3')	Effect; Product
<i>polg2</i> -specific oligo	<i>polg</i> / ZDB-GENE-060303-1	CRISPR/Cas9 DNA oligomer for gRNA (gRNA sequence)	ATTTAGGTGACACTATAGGGCGA TGAGAGTTTGAAAGGTTTTAGAG CTAGAAATAGCAAG	CRISPR/Cas9-induced <i>polg2</i> mutagenesis
<i>polg2</i> -F	<i>polg2</i> / ZDB-GENE-060810-116	DNA primer for <i>ia304</i> genotyping	TAGGCCCGTCTGACTTCAAC	Normal product: 137 bp Deleted product: 127 bp
<i>polg2</i> -R	<i>polg2</i> / ZDB-GENE-060810-116	DNA primer for <i>ia304</i> genotyping	TAGTTGTGTGTCTCCAGGG	
<i>polg2</i> -diagn-F	<i>polg2</i> / ZDB-GENE-060810-116	DNA primer for <i>ia304</i> sequencing	CTGACCACTGAAAGCCACTATG	Normal product: 252 bp Deleted product: 242 bp
<i>polg2</i> -diagn-R	<i>polg2</i> / ZDB-GENE-060810-116	DNA primer for <i>ia304</i> sequencing	ACGGATGTTTCTTGGTGAGTCT	
<i>nucl-polg</i> -F	<i>polg</i> / ZDB-GENE-060303-1	primer for mtDNA depletion analysis	GAGAGCGTCTATAAGGAGTAC	Reference nuclear gene Genomic DNA product: 81 bp
<i>nucl-polg</i> -R	<i>polg</i> / ZDB-GENE-060303-1	primer for mtDNA depletion analysis	GAGCTCATCAGAAACAGGACT	
<i>mt-nd1</i> -F	<i>mt-nd1</i> / ZDB-GENE-011205-7	primer for mtDNA depletion analysis	AGCCTACGCCGTACCAGTATT	Reference mitochondrial gene Mt DNA product: 143 bp
<i>mt-nd1</i> -R	<i>mt-nd1</i> / ZDB-GENE-011205-7	primer for mtDNA depletion analysis	GTTTCACGCCATCAGCTACTG	
<i>mt-nd2</i> -F	<i>mt-nd2</i> / ZDB-GENE-011205-8	primer for mtDNA depletion analysis	GCAGTAGAAGCCACCACAAA	Reference mitochondrial gene Mt DNA product: 173 bp
<i>mt-nd2</i> -R	<i>mt-nd2</i> / ZDB-GENE-011205-8	primer for mtDNA depletion analysis	GCTAGACCGATTTTGAGAGCC	
<i>zf-gapdh</i> -F	<i>gapdh</i> / ZDB-GENE-030115-1	DNA primer for Real Time RT-PCR	GTGGAGTCTACTGGTGTCTTC	Housekeeping gene cDNA control product: 161 bp
<i>zf-gapdh</i> -R	<i>gapdh</i> / ZDB-GENE-030115-1	DNA primer for Real Time RT-PCR	GTGCAGGAGGCATTGCTTACA	
<i>zf-eef1a1a</i> -F	<i>eef1a1a</i> / ZDB-GENE-030131-8278	DNA primer for Real Time RT-PCR	TGCAGAGATGGGAAAGGGT	Housekeeping gene cDNA control product: 161 bp
<i>zf-eef1a1a</i> -R	<i>eef1a1a</i> / ZDB-GENE-030131-8278	DNA primer for Real Time RT-PCR	GCTGGTCTCAAACCTCCACA	
<i>polg-ex3</i> -F	<i>polg</i> / ZDB-GENE-060303-1	DNA primer for Real Time RT-PCR	ATCTCATCCCGCTGGAAAC	Catalytic subunit gene cDNA target product: 320 bp
<i>polg-ex5</i> -R	<i>polg</i> / ZDB-GENE-060303-1	DNA primer for Real Time RT-PCR	GCTCATGGGAATGGGTTAAT	
<i>polg2-ex6</i> -F	<i>polg2</i> / ZDB-GENE-060810-116	DNA primer for Real Time RT-PCR	GCTCCATCCTGCTTTAACTCC	Accessory subunit gene cDNA target product: 141 bp
<i>polg2-ex7</i> -R	<i>polg2</i> / ZDB-GENE-060810-116	DNA primer for Real Time RT-PCR	GTGTCCAAGTATCCAGGCCA	

Suppl. Table 1: List of oligomers used in this study.