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Free radicals and neonatal encephalopathy: mechanisms of injury, biomarkers, and antioxidant treatment perspectives

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1	Free radicals and neonatal encephalopathy: mechanisms of injury, biomarkers and
2	antioxidant treatment perspectives.
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1 Abstract

2 Neonatal encephalopathy (NE), most commonly a result of the disruption of cerebral oxygen
3 delivery, is the leading cause of neurologic disability in term neonates.

Given the key role of free radicals in brain injury development following hypoxia-ischemiareperfusion, several oxidative biomarkers have been explored in pre-clinical and clinical models of NE. Among these, antioxidant enzyme activity, uric acid excretion, nitric oxide, malondialdehyde and non-protein-bound iron have shown promising results as possible predictors of NE severity and outcome. Due to high costs and technical complexity, however, their routine use in clinical practice is still limited.

10 Several strategies aimed at reducing free radical production or up-regulating physiological 11 scavengers have been proposed for NE. Room-air resuscitation has proved to reduce oxidative 12 stress following perinatal asphyxia and is now universally adopted. A number of medications 13 endowed with antioxidant properties, such as melatonin, erythropoietin, allopurinol or N-14 acetylcysteine, have also shown potential neuroprotective effects in perinatal asphyxia; 15 nevertheless, further evidence is needed before these antioxidant approaches could be implemented 16 as standard care.

1 Role of free radicals in the pathogenesis of hypoxic-ischemic encephalopathy

Neonatal encephalopathy (NE) most commonly results from an acute or subacute disruption of cerebral blood flow and oxygen delivery to the brain during the perinatal period. The incidence of NE ranges from 1 to 8 per 1000 live births in high-income countries to as high as 26 per 1000 live births in low-income countries (1). Despite the advent of therapeutic hypothermia, this condition is still a major cause of death and neurodevelopmental disability in term neonates worldwide (2).

7 Specific antepartum risk factors (e.g., maternal pyrexia, prolonged rupture of membranes, persistent 8 occipito-posterior position), or a well-recognized intrapartum event responsible for an acute 9 decrease of placental perfusion (e.g., placental abruption, prolapse of the umbilical cord, uterine 10 rupture, shoulder dystocia) can be often identified (3,4). The development of hypoxic-ischemic 11 brain damage is more likely in the presence of umbilical cord arterial pH <6.8, base excess <-20 12 mEq/l, Apgar score of ≤ 3 at 10 min and other negative prognostic factors, such as absent fetal heart 13 rate variability prior to birth, seizures in the first day of life, and multi-organ injury (5). The 14 characteristics of the asphyxiating insult (i.e., intermittent, persistent, chronic), together with the 15 infant's gestational age, prior metabolic and cardiovascular status and individual sensitivity to 16 oxidative stress further contribute to the severity of NE (5). The Sarnat staging system, based on the 17 combination of specific clinical signs (e.g., abnormalities of consciousness, tone, reflexes and/or 18 electrical brain activity), classifies NE into three stages of increasing severity: the higher the stage, 19 the lower the probability of survival without major neurological sequelae (6).

The main phases of NE, with the related mechanisms of injury and the therapeutic strategies currently proposed in research settings, are summarized in Figure 1. The event sequence leading to NE includes oxygen deprivation, energy depletion, and re-oxygenation. As illustrated in Figure 2, these events contribute to the generation of reactive oxygen (ROS) and nitrogen species (RNS), which harmfully interact with nearby proteins, nucleic acids or membrane lipids, altering their function and converting them into free radicals (7). ROS are finely regulated by specific antioxidant enzymes, such as superoxide dismutase (SOD), catalase (CAT), reduced glutathione (GSH) and glutathione peroxidase (GP); the imbalance between ROS production and clearance leads to
 oxidative stress (OS) in the newborn (8,9).

3 The metabolic consequence of acute cerebral hypoperfusion is the inhibition of oxidative 4 phosphorylation in the electron transport chain (ETC) of the mitochondria, resulting in anaerobic 5 metabolism. Glucose utilization in anaerobic glycolysis is highly inefficient and contributes to a 6 rapid depletion of cerebral glucose, the primary energy source for neural cells (10). The resulting 7 decrease in adenosine triphosphate (ATP) leads to the inactivation of ATP-dependent ion pumps, 8 with intracellular accumulation of sodium and water, progressive cell swelling and cellular necrosis. 9 This event cascade culminates into glutamate excitotoxicity, which not only activates apoptotic 10 pathways via N-methyl-D-aspartate (NMDA) and glutamate receptors, but also up-regulates nitric 11 oxide synthase (NOS) to induce a compensatory increase in cerebral blood flow (11). The ensuing 12 NO surge, however, triggers the production of potent RNS that actively contribute to brain damage 13 (12).

Following acute hypoxia-ischemia in term neonates, the deep grey nuclei appear most vulnerable. The enhanced susceptibility of this area is due to the presence of NOS–expressing (NOS+) striatal neurons that are paradoxically resistant to hypoxic-ischemic injury but, by producing high amounts of RNS, exert a harmful bystander effect on nearby neural and glial cells (13-16).

Following the acute insult, there is a restoration of cerebral perfusion, which, while essential for survival, paradoxically contributes to the so-called reperfusion injury, mediated by excessive free radical production (17). This results not only in an oxidative burst but also in a progressive disruption of the mitochondrial ETC, leading to secondary ATP depletion and subsequent apoptotic brain damage (18).

The progressive accumulation of hypoxanthine that follows ATP depletion, and the reoxygenationdriven conversion of xanthine dehydrogenase (XD) to xanthine oxidase (XO), a superoxideproducing enzyme which, using O₂ as a cofactor, produces •O2- and uric acid from xanthine or hypoxanthine, are primary sources of ROS after reperfusion (19). Non-protein-bound iron (NPBI), released from hemoglobin, enhances hydroxyl radical formation via the Fenton reaction, thus raising ROS levels exponentially (20). ROS and RNS further worsen the mitochondrial damage begun by primary ATP depletion and glutamate excitotoxicity, contributing to the progressive disruption of oxidative phosphorylation that ensues in secondary energy failure and neuronal apoptosis (19). These latter events, however, are preceded by a phase of latency, which may last up to 6 hours following reperfusion and represents a crucial therapeutic window (4).

7 Understanding the role of free radicals in NE has led to the discovery of several biomarkers of 8 oxidation or nitrosylation, with possible diagnostic and prognostic implications. Also, there is a 9 potential of developing novel neuroprotective approaches based on antioxidant therapies. This 10 review aims to provide an overview of the role of free radical biomarkers and antioxidant therapies 11 in NE. Literature search methods are provided in Supplemental Table S1 (online).

12

13 Free radical biomarkers in NE

14 The diagnosis of NE mainly relies on clinical, neurophysiological and neuroimaging abnormalities. 15 However, both the neurological status and cerebral electrical activity can be significantly altered by 16 sedatives, anticonvulsants, and therapeutic hypothermia (TH) (21). Moreover, diagnostic changes 17 on magnetic resonance (MR) imaging may take several days to become apparent. The validation of 18 molecular biomarkers of hypoxia-ischemia and ensuing free radical burden may help identifying 19 neonates at higher risk of moderate-to-severe NE, who could qualify for early neuroprotective 20 treatments. The following section focuses on current biomarkers for oxidative and nitrosative stress, 21 which are summarized in Table 1 (22).

Gas chromatography coupled to mass spectrometry has long been the gold-standard technique for a qualitative and quantitative estimation of oxidative compounds on different specimens (16). Recently, the development of high or ultra-performance liquid chromatography (LC-MS/MS), which allows a simultaneous measurement of different compounds on smaller sample volumes, has contributed to ease the assessment of oxidative status in the neonatal population (23). Nevertheless, high costs and need for trained personnel are major limitations to the routine clinical application of
these methods, which are therefore often confined to research facilities (24).

3

4 1. Antioxidant enzymes

5 SOD, GP and CAT are the first-line antioxidant defense against OS. Following a hypoxic-ischemic 6 hit, their activity acutely rises to counteract the ensuing ROS production and its harmful effects. 7 Significantly higher SOD, CAT and GP levels have been reported in cord blood samples of 8 asphyxiated newborns compared to controls (25); at 24 hours, however, only CAT and SOD, but 9 not GP, maintained higher plasma concentrations in infants with NE compared to controls (26–28). 10 A positive correlation between CAT and SOD cord levels and Sarnat stages was also observed (25), 11 suggesting that the up regulation of these enzymes may mirror NE severity. Given their acute 12 increase, these data suggest that the evaluation of antioxidant enzymes in cord blood may add 13 useful information on both timing and severity of perinatal asphyxia within the decision-making 14 window for TH. Nevertheless, it should be noted that they are based on small cohorts, thus needing 15 further confirmation in larger clinical trials.

Very little is known about antioxidant enzymatic activities in other biological fluids. One study has reported a significantly enhanced SOD activity in cerebrospinal fluid (CSF) between 0 and 72 hours in 30 term neonates with NE, whereas GP and CAT activity increased only in the severe NE subgroup (29). However, there was a wide time interval during which lumbar puncture was performed (anytime within 72 hours of life) making it difficult to interpret.

Most of these cited studies were performed before the routine introduction of TH, whose potential
modulatory effect on antioxidant enzymatic activities requires further evaluation.

23

24 2. Uric acid

25 The reoxygenation-driven conversion of XD to XO, which produces •O2- and uric acid, plays a key 26 role in oxidative brain damage. Urinary excretion of uric acid has thus been proposed as an 1 economical and non-invasive marker for XO-related ROS production in asphyxiated newborns.

2 Several studies have consistently reported an increased urinary uric-acid-to-urinary-creatinine ratio 3 within 48-72 hours of life in term and preterm asphyxiated newborns (30-35), and a positive 4 correlation between this ratio and Sarnat stage has also been reported (31). A cut-off of 2.3 mg/mg 5 over the first 72 hours has been proposed as diagnostic of perinatal asphyxia in a single study on 40 6 asphyxiated infants in a low/middle resource setting (35). In this study, however, the diagnosis of 7 perinatal asphyxia was mainly based on Apgar score and cord pH at birth; moreover, no post-natal 8 clinical data of the infants enrolled are available, other than none of them were undergoing TH at 9 the time of urine collection. Values $\geq 2.6 \text{ mg/mg}$ may predict impeding death in asphyxiated term 10 infants with good sensitivity and specificity (31), but again, this data was based on a small study of 11 only 20 infants and performed before the introduction of TH. Larger studies are therefore needed on 12 patients undergoing TH.

13

14 3. Nitric oxide (NO)

15 Ischemia-induced up-regulation of NOS enhances NO and RNS production. A raised concentration 16 of NO in blood and CSF NO, as well as a higher plasma nitrates/nitrites ratio (which serves as a 17 proxy for NO levels), have been detected in neonates with NE within the first 24 hours (27,36–38), 18 with higher NO concentration related to higher Sarnat stage (26,37,38). Furthermore, plasma NO 19 was increased in asphyxiated infants with early evidence of brain damage compared to those with 20 normal neuroimaging (37). However, these studies trials were carried out before the introduction of 21 TH. Future studies, though, could also help to evaluate the role of this biomarker in selecting those 22 infants who might benefit of specific antioxidant treatments such as xanthine oxidase inhibitors, 23 which hinder the formation of peroxynitrite from xanthine-oxidase-derived superoxide and NO and 24 the subsequent activation of downstream pathways that lead to cerebral endothelial and tissue injury 25 (38).

1 4. Non-protein-bound iron (NPBI)

2 Following hypoxia-reoxygenation, hemoglobin-released NPBI interacts with $\bullet O_2$ - and H_2O_2 and 3 forms highly reactive •OH. Studies conducted in the pre-cooling era reported increased 4 concentrations of plasma NPBI with increasing NE severity (39), although low or undetectable 5 NPBI levels were also seen in moderately and severely asphysiated infants. However, low or 6 undetectable concentrations values were significantly associated with normal neurological outcome 7 at 1 year, irrespective of NE stage; these latter data, however, were obtained from a small number 8 of infants and need to be confirmed on larger samples (39). Higher NPBI levels in plasma and CSF 9 have been described within the first 72 hours in neonates with moderate-to-severe NE who died or developed neurological sequelae (40,41). More recently, increased plasma NPBI levels at 4-6 hours, 10 11 but not at 24-72 hours and 5 days of life, were detected in 80 infants with severe NE who 12 underwent TH, compared to those with mild-moderate NE (42), suggesting not only a possible 13 prognostic value of NPBI in the earliest phases after the perinatal insult, but also a possible 14 influence of TH on this biomarker or, more generally, on OS levels. Nevertheless, diagnostic NPBI 15 levels for NE still have to be defined.

16

17 5. Lipid peroxidation markers

18 The extent of lipid peroxidation following hypoxia-ischemia can overwhelm the adaptive up-19 regulation of antioxidant systems, leading to harmful effects on membrane phospholipids, 20 functional loss and programmed cell death. Once this process is initiated, it generates a wide variety 21 of lipid peroxidation products that, given the rich lipid composition of the infant brain, might reflect 22 the extent of cerebral oxidative damage.

23

24 Malondialdehyde (MDA) and 4-hydroxynonenal (4-HNE)

MDA, a biomarker for n-3 and n-6 fatty acid peroxidation, has been widely investigated, as it can
be easily determined by thiobarbituric acid assay, a widely available spectrophotometric technique

(43). The current literature is consistent in reporting increased MDA levels in cord blood of
severely asphyxiated neonates (31,34,44) and in the serum of infants with NE (26-28,36,41,45)
compared to controls. As for prognostic implications, serum MDA at 24 hours positively correlates
with NE severity (27,28), with highest values in infants who died (41,45) or developed persistent
neurological abnormalities (40,41,45).

6 Being water-soluble, MDA is detected in urine, and its excretion rate has also been tested in the 7 context of NE. The ratio between urinary MDA and urinary creatinine (uMDA/uCr) was 8 significantly increased over the first 48 hours of life in asphyxiated neonates compared to controls, 9 and positively correlated with Sarnat stage (31,34). A higher uMDA/uCr ratio was also observed in 10 asphyxiated neonates who died compared to survivors, and values >3.49 µg/mg on day 1 have been 11 proposed to predict mortality following perinatal asphyxia in a small number of asphyxiated infants 12 (31). Since most of these studies did not include infants who underwent TH, it is unclear whether 13 this treatment can influence MDA levels.

14 A possible limitation of serum and urinary MDA is the lack of specificity for the cerebral tissue, as 15 it may also reflect the extent of lipid peroxidation following extensive multiorgan damage. MDA 16 concentration in CSF should provide a better estimate of oxidative brain damage in asphyxiated 17 infants. Significantly higher CSF levels of MDA have been reported at 24-48 hours in asphyxiated 18 term infants compared to controls, with a positive correlation with Sarnat stages (25), and also in 19 infants with NE who expired or developed neurological deficits compared to those with normal 20 neurological status at hospital discharge (41). However, further data are needed to validate CSF 21 MDA as a reliable marker for oxidative brain damage in NE.

As for 4-HNE, a metabolite of n-6 fatty acid peroxidation, Schmidt et al reported significantly risen cord blood levels following a perinatal hypoxic insult (46). To date, however, no additional data are available to add knowledge on this biomarker in the context of NE. This could be due the unstable nature of 4-HNE that contributes to the technical complexity of its determination, which benefits from the use of advanced liquid chromatography methods (47).

2 *Prostaglandin-like peroxidation products (PPPs)*

3 Isoprostanes, neuroprostanes and neurofurans are prostaglandin-like compounds derived from free
4 radical-catalyzed peroxidation of arachidonic (AA) and docosahexaenoic (DHA) acid.

5 F2-isoprostanes seem to best reflect OS extent after hypoxia-reperfusion (48) and, thanks to ultra-6 performance LC-MS/MS, can be determined even in the neonatal population. Higher cord blood 7 levels of 8-iso-15(R)-PGF2 α and total isoprostanes were detected in acidotic and depressed infants 8 compared to healthy neonates; 8-iso-15(R)-PGF2a positively correlated with the severity of 9 asphyxia (49). However, serial evaluation of serum F2-isoprostanes over the first 5 days showed no 10 difference between neonates who developed severe compared to mild-to-moderate NE nor any 11 correlation with brain damage at neuroimaging (42). Given this scarce and inconclusive evidence, 12 further studies are necessary to define whether PPPs may reflect PUFA oxidation after perinatal 13 asphyxia.

14

15 6. Protein oxidation markers

Protein carbonyls (PC) and advanced oxidation protein products (AOPP) result from protein carbonylation, nitration, cross-linking or loss of thiol groups by free radicals. Increased serum PC have been observed at birth and 48 hours in asphyxiated term neonates, with higher levels in those who developed seizures; however, no difference was observed in relation to Sarnat stage or developmental outcome at 9 months (45). Buonocore et al. previously reported increased cord levels of AOPP in hypoxic compared to normoxic preterm infants, and a positive correlation between this marker and plasma hydroperoxides (50).

Recently, higher plasma AOPP at 4-6 hours of life have been described in term infants with severe
NE, who required TH, compared to mild/moderate NE (42). Later measurements (i.e., 24-72 hours
and 5 days) showed no difference between cooled asphyxiated infants and controls. This is
consistent with previous findings by Mutlu et al, who compared AOPP levels between 30 cooled

NE infants and 30 healthy term controls at 6-24 hours and 5 days (28). Given this lack of difference in AOPP levels during and after TH, it suggests a possible role of cooling in modulating protein oxidation, which warrants further investigation. A significant independent association between MRI scores suggestive of brain injury and blood levels of AOPP over the first 5 days of life in neonates with NE has also been reported, with a stronger correlation in male infants, suggesting a possible role for AOPP as a biomarker of brain oxidative damage (42).

7

8 7. DNA peroxidation markers

9 Oxidative stress can lead to harmful peroxidative changes to nucleic acids. The oxidized DNA 10 nucleoside 8-hydroxydeoxyguanosine (8-OHdG), resulting from DNA peroxidation, has been 11 proposed as a biomarker for oxidative DNA damage in the neonatal population (51,52), and has 12 also been used to evaluate the efficacy of TH in reducing OS-related DNA damage (53). 13 Nevertheless, to the best of our knowledge, current clinical evidence on its diagnostic role in NE is 14 limited to a small, pilot study aimed at evaluating 8-OHdG concentration in urine and CSF samples 15 of children with brain damage, including a small subgroup with NE (54). Despite significantly 16 higher CSF and urinary 8-OHdG levels were observed in this population compared to control 17 subjects, the study sample is undersized to draw any conclusion, and data on the timing of specimen 18 collection are not available. Moreover, this biomarker is also increased in other causes of brain 19 injury (e.g., status epilepticus, central nervous system infections), thus suggesting a lack of 20 specificity for NE.

21

22 Antioxidant strategies for neuroprotection in NE

The role of free radicals in the development of brain injury following hypoxia-ischemia-reperfusion has provided the rationale to explore antioxidant therapeutic approaches for NE, aimed either at upregulating the physiological scavenging systems or hindering ROS and RNS production at different levels. While the use of room air for the resuscitation of depressed term infants has become part of 1 standard neonatal care, the use of other antioxidant molecules is mainly limited to research settings,

2 with only variable pre-clinical and/or clinical supportive evidence (see Table 2).

3 The main neuroprotective strategies currently adopted or proposed to dampen oxidative brain
4 damage after perinatal asphyxia will be are analyzed and discussed below.

5

6 a. Use of room air and novel inhaled antioxidant strategies for neonatal resuscitation

The beneficial effects of the use of 21% oxygen for the resuscitation of asphyxiated neonates on clinical outcomes and mortality was first reported two decades ago (55) and largely confirmed over the following years, thus becoming the standard of care for term infants' delivery room management since 2010 (56). In parallel, the relation between OS and the oxygen amount provided during resuscitation after perinatal asphyxia has been investigated in animal and human studies, producing a growing body of evidence towards a reduction of the oxidative burden following roomair resuscitation (57–60).

14 Based on this evidence, novel inhaled antioxidant strategies have been recently proposed for 15 perinatal resuscitation. The use of inhaled hydrogen with room air has shown an encouraging 16 attenuation of cerebral oxidative biomarkers in swine models of NE (61,62); clinical evidence, 17 however, is lacking. Due to its ability to cross the blood-brain barrier (BBB), inhaled xenon has 18 also attracted broad interest as a potential neuroprotective agent for NE, although its mechanism of 19 action is mainly ascribable to NMDA receptor antagonism rather than to antioxidant properties. 20 Current evidence from randomized-controlled trials, however, are limited to Azzopardi et al., who 21 did not demonstrate any added effect to TH in infants with NE (63,64), despite of the acknowledged 22 possible limitations (e.g., timing of the treatment start and duration of dose).

Recently, due to its cheaper costs compared to xenon, inhaled argon has also been proposed for neuroprotection following perinatal asphyxia. Encouraging preliminary evidence of reduced white matter lactate and N-acetyl aspartate at 24 and 48 hours, as well as of reduced apoptotic burden and faster recovery of amplitude-integrated electroencephalography has been observed in asphyxiated piglets undergone TH and 45-50% argon inhalation 2 hours after the insult, compared to a TH-only
group (65). Nevertheless, the safety of argon needs to be further assessed before translational
clinical trials can commence.

4

5 b. Therapeutic hypothermia

6 The efficacy of TH in decreasing brain injury (66) and reducing major neurocognitive sequelae (67) 7 in neonates with moderate-to-severe NE has been largely established, and is currently the standard 8 of care for term and late-preterm infants with NE (4). The beneficial effects of TH mainly result 9 from the down-regulation of cerebral energy metabolism, which dampens the apoptotic burden of 10 secondary cerebral energy failure (68). However, other suggested mechanisms through which TH 11 exerts neuroprotection include inhibiting neuronal cell death, limiting excitoxicity, modulating glial 12 cell activation (69) and activating cold-inducible RNAs (70). Moreover, a direct reduction of 13 oxygen-based free radicals following an ischemic insult and subsequent reperfusion has been 14 reported in both in vitro (71) and in vivo animal studies (72).

15 In animal models, TH has proved to effectively decrease striatal and cortical NO-mediated 16 production of cyclic GMP (73) and lipid peroxidation products in white matter (74), suggesting a 17 protective effect against not only oxidative but also nitrosative stress. In clinical studies, decreased 18 serum levels of MDA and PC and increased SOD, GP and glutathione S-transferase activities have 19 been documented in adults undergoing hypothermia after cardiac arrest (75), whereas no significant 20 difference in serum hydroperoxides was observed in hypothermic compared with normothermic 21 asphyxiated term infants over the first 3 days of life (76). The small amount of preliminary data 22 would suggest that TH has a beneficial effect on free radical production (28,42). Moreover, a 23 combined role of TH and other neuroprotective strategies with known antioxidant properties (e.g., 24 erythropoietin, melatonin, allopurinol, N-acetylcysteine, DHA) in reducing hypoxic-ischemic brain 25 damage has also been reported, suggesting a synergistic mechanism of action.

1 c. Erythropoietin (Epo)

By binding to its receptors (Epo-R), which are largely expressed in the central nervous system, Epo inhibits apoptotic pathways, reduces proinflammatory cytokines and dampens glutamate excitotoxicity (77), with promising therapeutic implications in NE (78). Under hypoxic conditions, Epo-R expression and Epo secretion are significantly up-regulated but, while the former increases promptly, the latter rises more slowly (79); hence, the potential therapeutic role of exogenous Epo is greatest during this period.

Pre-clinical research has provided robust evidence in support of the neuroprotective effects of Epo following perinatal hypoxia, reporting reduced neuronal apoptosis and inflammation, less damage to white and grey matter, lower mortality rates and long-term improvements in motor and cognitive functions (80-84). An *in vitro* suppression of ROS production in microglia and reduced lipid peroxidation products have also been reported in fetal murine brain following Epo administration (85,86), suggesting possible antioxidant properties.

14 Consistent evidence has been obtained from controlled clinical trials on asphyxiated infants treated 15 with Epo, either alongside TH (87) or alone (88-92). Epo administration has been associated with 16 decreased serum SOD and GP levels (91) fewer seizures (88,90), less abnormalities at 17 neuroimaging (87,90), better psychomotor outcomes up to 24 months of age (89–91), and, when compared to supportive care, reduced mortality (91,92). Current clinical evidence, however, is 18 19 derived from small studies and, as such, it is burdened by pronounced differences in terms of 20 settings (e.g., high or medium-low income), Epo dose ranges or timing of administration, outcome 21 evaluation etc. Hence, further data on a larger scale are needed to confirm these positive 22 preliminary results. To this regard, two multicenter randomized double-blind controlled trials are 23 currently recruiting (NCT02811263 and NCT03079167).

24

25 *d. Melatonin*

1 Melatonin has broad antioxidant, anti-inflammatory and anti-apoptotic properties. As an 2 antioxidant, it acts as a ROS scavenger and also enhances ETC efficiency and the enzymatic 3 activity of SOD, GP and glutathione reductase (93); moreover, by reducing NOS expression, it 4 contributes to decrease peroxynitrite formation (94). Due to its lipophilic nature, this molecule can 5 easily cross the placental interface and the BBB, with useful therapeutic implications.

6 Antioxidant benefits in NE are largely supported by pre-clinical evidence reporting lower brain 7 levels of ROS, lipid and protein peroxidation products, free iron and NO, increased GSH in 8 periventricular white matter, reduced neuronal apoptosis and improved long-term developmental 9 outcomes (95–99). Alongside TH, melatonin significantly enhanced hypothermic neuroprotection in asphyxiated newborn piglets by reducing the area under the p-MRS curve for lactate/N-acetyl 10 11 aspartate (NAA) and lactate/total creatine ratios in deep grey matter, improving cerebral energy 12 metabolism and decreasing the apoptotic burden in basal ganglia and internal capsule (100). In 13 order to enhance its solubility in aqueous vehicles, however, melatonin administered in the above 14 studies was diluted in ethanol, which, at low doses, has been associated with in vitro protective 15 effects against ischemia/reperfusion-mediated brain injury, and could have thus acted as a possible 16 confounder (101). This issue has been recently addressed by Robertson et al., who tested the 17 neuroprotective effects of an ethanol-free melatonin formulation, administered at 5 and 15 mg/kg, 18 compared to TH in a piglet model of NE (102). Reduced cell death in the sensorimotor cortex was 19 observed with 15 mg/kg melatonin, but not with lower concentrations; moreover, no between-group 20 difference in lactate/NAA was found at p-MRS evaluation.

In one small clinical study, lower serum levels of MDA, nitrosative biomarkers and reduced mortality rates have been reported in a small cohort of asphyxiated neonates given melatonin (103). In another trial, the combination of melatonin and TH resulted in decreased serum NO, fewer seizures, reduced white matter injury at neuroimaging and improved survival without neurodevelopmental abnormalities at 6 months (104); however, the different rates of severe NE in the two study groups (TH only vs. TH plus melatonin), together with the limited study sample, may

have biased these observed results. A targeted trial aimed at assessing the appropriate melatonin
dosage to achieve neuroprotective effects in infants with NE undergoing TH (NCT02621944) is
currently recruiting. Based on current literature, neonatal melatonin administration holds promising
potential for translation to standard practice in NE, although further studies are warranted for its
validation.

6 The ability of melatonin to cross the placenta has led to the evaluation of maternal intrapartum 7 administration to minimize the harm of intrauterine fetal asphyxia. A reduction of lipid peroxidation 8 products has been reported in the cerebral tissue of late-gestation lambs (105), but equivalent 9 clinical studies are not available yet. Perinatal asphyxia, however, often results from a sudden and 10 unpredictable event, thus possibly limiting the viability for research validation and subsequent 11 clinical implementation of this approach.

12

13 e. Allopurinol

The neuroprotective potential of allopurinol results from its antioxidant properties, such as XO inhibition and free radicals scavenging, combined with its ability to cross the BBB and the placenta. Significantly reduced acute cerebral edema and extent of brain injury have been reported in the treated arm of a murine model of NE (106,107), with possible gender-related effects (108).

18 When first translated into clinical research, allopurinol administration had been associated with 19 significantly lower serum levels of NPBI and uric acid, no rise in serum MDA, and more stable 20 cerebral blood flow and electrical activity in treated compared to untreated asphyxiated infants 21 (109). Nonetheless, no effect on mortality and morbidity in severe neonatal asphyxia was 22 documented (110). Decreased NO levels in serum, but not in CSF, were also reported at 72-96 23 hours of life following allopurinol treatment, commenced within 2 hours from the perinatal 24 asphyxiating insult (38). Importantly, no adverse effects on blood cell count, skin, and liver 25 enzymes have been reported (38,109,110).

Longer term effects of post-asphyxia allopurinol treatment on neurodevelopment have also been
 investigated. While Gunes et al. (38) observed improved outcomes at ≥12 months of age, the 5-year
 follow-up of Van Bel and Benders' trials demonstrated protective effects only for the moderate NE
 subgroup (111); these studies, however, were performed before the routine introduction of TH.

5 The efficacy of antepartum allopurinol administration in the presence of fetal hypoxia has been examined in animal (111,112) and human (113-115) studies. While reduced cord blood levels of 6 7 NPBI and neuronal damage biomarkers were preliminary reported in neonates born from treated 8 mothers (113), results from a multicenter randomized placebo-controlled trial (ALLO-trial) failed to 9 demonstrate any decrease in cord blood biomarkers of neuronal damage and lipid peroxidation 10 (114) or improved developmental and behavioral outcomes at 5 years in infants from the treated 11 arm (115). However, it should be noted that, as reported by the authors, none of the infants included 12 in the ALLO-trial had developed NE, and no significant differences in Apgar scores, cord pH and 13 base excess were noted between the treated and control arms (114). While these results would 14 suggest that further clinical studies are warranted, there remains an issue regarding the feasibility of 15 maternal intrapartum treatment in relation to the unpredictability of perinatal asphyxia, if the aim is 16 to prevent NE.

Available clinical evidence on the neuroprotective effects of antenatal or postnatal allopurinol in NE is still inconclusive (116). Data from an ongoing study (ALBINO, NCT03162653; http://www.albino-study.eu) evaluating the efficacy and safety of allopurinol administration immediately after birth to near-term infants with NE in addition to TH may help to clarify the validity of postnatal therapy in the upcoming years.

22

23 f. NOS inhibitors

NO production is mediated by different NOS isoforms: neuronal (nNOS), endothelial (eNOS) and inducible (iNOS). These isoforms, expressed in neurons, astrocytes and endothelial cells, are

significantly up-regulated under hypoxic conditions; as such, NOS inhibition has been proposed as
a possible neuroprotective strategy for NE (117).

In murine models of NE, pre-hypoxic administration of iNOS, nNOS and constitutive NOS inhibitors, such as aminoguanidine, 7-nitroindazole and N^G-nitro-L-arginine effectively reduced NO levels during hypoxia and re-oxygenation (118–126). Similar results, however, were not obtained after post-insult administration (120-122); given the unpredictability of perinatal asphyxia, this represents a significant limitation.

8 The selective nNOS and iNOS inhibitor 2-iminobiotin (2-IB), administered in a repeated dosing 9 regimen before and after the hypoxic insult, has shown neuroprotective effects in asphyxiated 10 female mice (14,123). Based on this pre-clinical evidence, 2-IB efficacy in human NE is currently 11 being tested in two clinical trials (EudraCT2015-003063-12, NTR5221), whereas another one 12 (NCT01626924) was prematurely terminated because TH, which was an exclusion criterion for the 13 trial enrolment, had soon become the standard of care. The mechanisms of action of 2-IB, 14 nevertheless, seem to ensue from the blockage of cytochrome C/caspase-3 apoptotic cascade rather 15 than from NOS inhibition (123–125).

Eventually, a novel class of computer-designed nNOS inhibitors has been tested in animal models of NE, proving better than 7-nitroindazole in nNOS down-regulation (126). These molecules have also shown protective effects against nitrosative stress on striatal neurons and improved motor and neurobehavioral outcomes (127–129), paving the way for its potential clinical translation.

20

21 g. N-Acetylcysteine (NAC)

NAC is a cysteine precursor endowed with antioxidant effects, such as ROS scavenging and GSH
replenishment in deficient cells (130). These properties, together with its lipophilic nature and lowtoxicity profile (131), have made NAC an interesting candidate for neuroprotection in NE.

Post-resuscitation NAC administration in asphyxiated piglets has been associated with decreased
 inflammatory markers in the prefrontal cortex and cerebellum (132,133), significantly attenuated

1 H₂O₂ surge in the cortex and decreased cortical levels of lipid hydroperoxide and oxidized 2 glutathione (134,135). Similar findings, together with reduced brain nitrotyrosine, were reported 3 also in a rodent model of lipopolysaccharide-sensitized hypoxia-ischemia (136). Moreover, when 4 combined with TH, NAC effectively reduced brain volume loss, increased myelin expression and 5 improved functional outcomes after hypoxic-ischemic brain injury in neonatal rats (137).

Preliminary clinical evidence from a small cohort of asphyxiated newborns who had previously
undergone TH has shown a significant GSH surge in basal ganglia on MR spectroscopy within 30
minutes from intravenous NAC administration (138). Nevertheless, further studies are needed to
evaluate NAC efficacy and safety in NE.

Encouraging neuroprotective effects after maternal intrapartum NAC administration have been
observed in newborns exposed to chorioamnionitis (139). Trials on asphyxiated neonates, however,
are not available yet.

13

14 h. Docosahexaenoic acid (DHA)

15 Based on its free radical scavenging ability, and following evidence of decreased glutamate 16 excitotoxicity, reduced NO and increased antioxidant enzymatic activities in DHA-enriched 17 neuronal cultures (140), a possible neuroprotective role for DHA has been hypothesized. In a swine 18 model of NE, post-insult DHA significantly reduced cortical and hippocampal lipid peroxidation 19 markers (141), increased hippocampal GSH levels and showed an added effect to TH in decreasing 20 urine F4-neuroprostanes (142) and cortical lactate/N-acetylaspartate ratio at MR spectroscopy (143). In rodent studies, pre-insult DHA reduced brain volume loss and improved functional 21 22 outcome after hypoxia-ischemia (144), whereas post-insult administration achieved similar results 23 only in combination with TH (145). Clinical trials on DHA administration in NE, however, are 24 lacking; hence, targeted studies are warranted to evaluate whether the above pre-clinical evidence 25 can be translated to clinical settings.

1 *i. Edaravone*

2 Edaravone (3-methyl-1-phenyl-pyrazolin-5-one) is a free radical scavenger whose multiple 3 antioxidant effects have been tested in different experimental settings, including animal models of 4 NE (146). In a murine study, pre- and post-insult edaravone administration effectively decreased the 5 burden of apoptosis and necrosis and dampened down mitochondrial injury (147), whereas its 6 administration before, during and after hypoxia led to a dose-dependent inhibition of lipid 7 peroxidation in the neonatal rat brain (148). Eventually, post-insult education both the 8 number of apoptotic neurons and the expression of 8-OHdG, a marker of DNA peroxidation, within 9 48 h after the hypoxic-ischemic insult (149). To date, however, clinical evidence on the 10 neuroprotective effects of edaravone is limited to a small cohort of pediatric patients with cerebral 11 infarction, who showed improved neurological outcome without significant adverse effects (150), 12 whilst data on its efficacy and safety in the neonatal population are still lacking.

13

14 Conclusion

15 Free radicals play a major role in the development of brain injury following hypoxia-ischemia-16 reperfusion in asphyxiated neonates. Over the past decades, a number of oxidative and nitrosative 17 biomarkers from different biological fluids have been proposed to assess the burden of free radical 18 damage following perinatal asphyxia. Among them, antioxidant enzymes, uric acid excretion rate, 19 NO, MDA and NPBI have been investigated more extensively in experimental and clinical 20 research, showing also a possible predictive value for NE severity and outcome, whereas other biomarkers (e.g., F2-isoprostanes, AOPP) have come into the spotlight more recently, and require 21 22 larger evaluation. Nevertheless, small and highly heterogeneous study samples together with the 23 lack of commercially available reference standards often hinder the clinical validation of these 24 biomarkers and call for larger, multicenter trials, aimed also at establishing normal values and 25 evaluating the influence of TH. Moreover, it is an open matter of debate whether plasma or urine 26 metabolites truly report brain oxidation extent or rather reflect the multiorgan oxidative damage that often accompanies severe NE. While CSF specimens are more specific, they are harder to collect, particularly in sick or unstable infants, and only few biomarkers have been currently assessed in this biological fluid. Eventually, by enhancing OS, several conditions different from perinatal asphyxia, either antenatal (e.g., maternal pre-eclampsia (151), exposure to maternal tobacco (152) etc.) or postnatal (e.g., sepsis (153), respiratory distress (154) etc.) may alter the oxidant status of newborn infants, and therefore should be taken into account when clinical trials evaluating oxidative or nitrosative biomarkers are designed.

8 Different antioxidant strategies have been explored as neuroprotective candidates for NE. While 9 room-air resuscitation of asphyxiated term neonates is universally recommended and TH has 10 become the standard of care for NE, a number of molecules endowed with antioxidant properties 11 are currently under investigation. Melatonin and Epo have shown mild beneficial effects in clinical 12 studies but require further large-scale validation before being introduced in routine neonatal care, 13 whereas evidence on allopurinol efficacy is still inconclusive. Encouraging neuroprotective effects 14 of NOS inhibitors, NAC and DHA have been shown in pre-clinical trials, thus calling for clinical 15 translation. In addition to the limitations already discussed for free radical biomarkers, such as 16 undersized and heterogeneous samples, often from the pre-cooling era, current literature evaluating 17 antioxidant treatments in clinical settings is further weakened by important differences in the 18 dosages adopted and in the outcomes examined. Large, prospective multicenter randomized 19 controlled clinical trials, assessing both the efficacy and safety of the above antioxidant treatments, 20 are required before they enter routine clinical use.

2	1.	Kurinczuk JJ, White-Koning M, Badawi N. Epidemiology of neonatal encephalopathy and
3		hypoxic-ischaemic encephalopathy. Early Hum Dev. 2010;86:329-338
4	2.	Natarajan G, Pappas A, Shankaran S. Outcomes in childhood following therapeutic
5		hypothermia for neonatal hypoxic-ischemic encephalopathy (HIE). Semin Perinatol
6		2016;40:549–55
7	3.	Nelson KB, Bingham P, Edwards EM, et al. Antecedents of neonatal encephalopathy in the
8		Vermont Oxford Network Encephalopathy Registry. Pediatrics 2012;130:878-86
9	4.	Douglas-Escobar M, Weiss MD. Hypoxic-ischemic encephalopathy: a review for the clinician.
10		JAMA Pediatr 2015;169:397–403
11	5.	Tonni G, Leoncini S, Signorini C, Ciccoli L, De Felice C. Pathology of perinatal brain damage:
12		background and oxidative stress markers. Arch Gynecol Obstet. 2014;290:13-20
13	6.	Sarnat HB, Sarnat MS. Neonatal encephalopathy following fetal distress. A clinical and
14		electroencephalographic study. Arch Neurol 1976;33:696–705
15	7.	Valko M, Leibfritz D, Moncol J, Cronin MTD, Mazur M, Telser J. Free radicals and
16		antioxidants in normal physiological functions and human disease. Int J Biochem Cell Biol
17		2007;39:44-84
18	8.	Lushchak VI. Free radicals, reactive oxygen species, oxidative stress and its classification.
19		Chem Biol Interact 2014;224:164–75
20	9.	Buonocore G, Perrone S, Tataranno ML. Oxidative Stress in the Newborn. Oxid Med Cell
21		Longev 2017;2017:1094247
22	10.	Brekke E, Berger HR, Widerøe M, Sonnewald U, Morken TS. Glucose and Intermediary
23		Metabolism and Astrocyte-Neuron Interactions Following Neonatal Hypoxia-Ischemia in Rat.
24		Neurochem Res 2017;42:115–32
25	11.	Iadecola C. Bright and dark sides of nitric oxide in ischemic brain injury. Trends Neurosci
26		1997;20:132–9

1	12.	Ferriero DM. Neonatal brain injury. N Engl J Med 2004;351:1985–95.
2	13.	McQuillen PS, Ferriero DM. Selective vulnerability in the developing central nervous system.
3		Pediatr Neurol 2004;30:227-35
4	14.	Peeters-Scholte C, Koster J, Veldhuis W, et al. Neuroprotection by selective nitric oxide
5		synthase inhibition at 24 hours after perinatal hypoxia-ischemia. Stroke 2002;33:2304–10
6	15.	Ferriero DM, Sheldon RA, Black SM, Chuai J. Selective destruction of nitric oxide synthase
7		neurons with quisqualate reduces damage after hypoxia-ischemia in the neonatal rat. Pediatr
8		Res 1995;38:912–8
9	16.	Ferriero DM, Holtzman DM, Black SM, Sheldon RA. Neonatal mice lacking neuronal nitric
10		oxide synthase are less vulnerable to hypoxic-ischemic injury. Neurobiol Dis 1996;3:64-71
11	17.	Hope PL, Cady EB, Chu A, Delpy DT, Gardiner RM, Reynolds EO. Brain metabolism and
12		intracellular pH during ischaemia and hypoxia: an in vivo 31P and 1H nuclear magnetic
13		resonance study in the lamb. J Neurochem 1987;49:75-82
14	18.	Wyatt JS, Edwards AD, Azzopardi D, Reynolds EO. Magnetic resonance and near infrared
15		spectroscopy for investigation of perinatal hypoxic-ischaemic brain injury. Arch. Dis. Child.
16		1989;64:953–63
17	19.	McCord JM, McCord JM. Oxygen-derived free radicals in postischemic tissue injury. N Engl J
18		Med 1985;312:159–63.
19	20.	Signorini C, Perrone S, Sgherri C, et al. Plasma Esterified F2-Isoprostanes and Oxidative Stress
20		in Newborns: Role of Nonprotein-Bound Iron. Pediatr Res 2008;63:287–91
21	21.	Thoresen M, Hellstrom-Westas L, Liu X, de Vries LS. Effect of Hypothermia on Amplitude-
22		Integrated Electroencephalogram in Infants With Asphyxia. Pediatrics 2010;126:e131-9
23	22.	Longini M, Belvisi E, Proietti F, Bazzini F, Buonocore G, Perrone S. Oxidative Stress
24		Biomarkers: Establishment of Reference Values for Isoprostanes, AOPP, and NPBI in Cord
25		Blood. Mediators Inflamm 2017;2017:1–6

1	23.	Torres-Cuevas I, Parra-Llorca A, Sánchez-Illana A, et al. Oxygen and oxidative stress in the
2		perinatal period. Redox Biol 2017;12:674–81

- 24. Kuligowski J, Escobar J, Quintás G, et al. Analysis of lipid peroxidation biomarkers in
 extremely low gestational age neonate urines by UPLC-MS/MS. Anal Bioanal Chem
 2014;406:4345–56
- 6 25. Kumar A, Ramakrishna SVK, Basu S, Rao GRK. Oxidative stress in perinatal asphyxia.
 7 Pediatr Neurol 2008;38:181–5
- 8 26. Singh SK, Dua T, Tandon A, Kumari S, Ray G, Batra S. Status of lipid peroxidation and
 9 antioxidant enzymes in hypoxic ischemic encephalopathy. Indian Pediatr 1999;36:561–6
- 10 27. Thorat VN, Suryakar AN, Sardeshmukh AS, Sarawade SS. Oxidants and antioxidants in
 11 hypoxic ischaemic encephalopathy. Indian J Clin Biochem 2004;19:32–5
- 12 28. Mutlu M, Sariaydin M, Aslan Y, et al. Status of vitamin D, antioxidant enzymes, and
 13 antioxidant substances in neonates with neonatal hypoxic-ischemic encephalopathy. J Matern
 14 Neonatal Med 2015;29:1–5
- Gulcan H, Ozturk IC, Arslan S. Alterations in Antioxidant Enzyme Activities in Cerebrospinal
 Fluid Related with Severity of Hypoxic Ischemic Encephalopathy in Newborns. Neonatology
 2005;88:87–91
- 30. Basu P, Som S, Choudhuri N, Das H. Correlation between Apgar score and urinary uric acid to
 creatinine ratio in perinatal asphyxia. Indian J Clin Biochem 2008;23:361–4
- 20 31. Banupriya C, Ratnakar, Doureradjou P, Mondal N, Vishnu B, Koner BC. Can urinary excretion
- 21 rate of malondialdehyde, uric acid and protein predict the severity and impending death in
- 22 perinatal asphyxia? Clin Biochem 2008;41:968–73
- 23 32. Bhongir AV, Yakama AVV, Saha S, Radia SB, Pabbati J. The Urinary Uric Acid/Creatinine
- Ratio is An Adjuvant Marker for Perinatal Asphyxia. Eur J Pharm Med Res 2015;2:520–8
- 25 33. Chen HJ, Yau KI, Tsai KS. Urinary uric acid/creatinine ratio as an additional marker of
- 26 perinatal asphyxia. J Formos Med Assoc 2000;99:771–4

1	34.	Mahmoud El Bana S, Esam Maher S, Fawzy Gaber A, Shaker Aly S. Serum and Urinary
2		Malondialdehyde (MDA), Uric acid, and Protein as markers of perinatal asphyxia. Electron
3		physician 2016;8:2614–9
4	35.	Patel KP, Makadia MG, Patel VI, Nilayangode HN, Nimbalkar SM. Urinary Uric
5		Acid/Creatinine Ratio - A Marker For Perinatal Asphyxia. J Clin Diagn Res 2017;11:SC08-
6		SC10
7	36.	Kumar A, Mittal R, Khanna HD, Basu S, Van de Bor M, Van Bel F. Free radical injury and
8		blood-brain barrier permeability in hypoxic-ischemic encephalopathy. Pediatrics
9		2008;122:e722-7
10	37.	Shi Y, Pan F, Li H, Pan J, Qin S, Shen C. Role of carbon monoxide and nitric oxide in newborn
11		infants with postasphyxial hypoxic-ischemic encephalopathy. Pediatrics 2000;106:1447-51
12	38.	Gunes T, Ozturk MA, Koklu E, Kose K, Gunes I. Effect of allopurinol supplementation on
13		nitric oxide levels in asphyxiated newborns. Pediatr Neurol 2007;36:17-24
14	39.	Dorrepaal CA, Berger HM, Benders MJ, van Zoeren-Grobben D, Van de Bor M, Van Bel F.
15		Nonprotein-bound iron in postasphyxial reperfusion injury of the newborn. Pediatrics
16		1996;98:883–9
17	40.	Yu T, Kui LQ, Ming QZ. Effect of asphyxia on non-protein-bound iron and lipid peroxidation
18		in newborn infants. Dev Med Child Neurol 2003;45:24-7
19	41.	Shouman BO, Mesbah A, Aly H. Iron metabolism and lipid peroxidation products in infants
20		with hypoxic ischemic encephalopathy. J Perinatol 2008;28:487-91
21	42.	Negro S, Benders MJNL, Tataranno ML, et al. Early Prediction of Hypoxic-Ischemic Brain
22		Injury by a New Panel of Biomarkers in a Population of Term Newborns. Oxid Med Cell
23		Longev 2018;2018:7608108
24	43.	Dasgupta A, Klein KL. Methods for Measuring Oxidative Stress in the Laboratory. In:
25		Antioxidants in Food, Vitamins and Supplements. 2014. p. 19-40

1	44.	Siciarz A, Weinberger B, Witz G, Hiatt M, Hegyi T. Urinary thiobarbituric acid-reacting
2		substances as potential biomarkers of intrauterine hypoxia. Arch Pediatr Adolesc Med
3		2001;155:718–22
4	45.	Mondal N, Bhat BV, Banupriya C, Koner BC. Oxidative stress in perinatal asphyxia in relation
5		to outcome. Indian J Pediatr 2010;77:515–7
6	46.	Schmidt H, Grune T, Müller R, Siems WG, Wauer RR. Increased Levels of Lipid Peroxidation
7		Products Malondialdehyde and 4-Hydroxynonenal after Perinatal Hypoxia. Pediatr Res
8		1996;40:15–20
9	47.	Zelzer S, Mangge H, Oberreither R, et al. Oxidative stress: Determination of 4-hydroxy-2-
10		nonenal by gas chromatography/mass spectrometry in human and rat plasma. Free Radic Res
11		2015;49:1233-8
12	48.	Sakamoto H, Corcoran TB, Laffey JG, Shorten GD. Isoprostanesmarkers of ischaemia
13		reperfusion injury. Eur J Anaesthesiol 2002;19:550–9
14	49.	Chafer-Pericas C, Cernada M, Rahkonen L, Stefanovic V, Andersson S, Vento M. Preliminary
15		case control study to establish the correlation between novel peroxidation biomarkers in cord
16		serum and the severity of hypoxic ischemic encephalopathy. Free Radic Biol Med
17		2016;97:244–9
18	50.	Buonocore G, Perrone S, Longini M, Terzuoli L, Bracci R. Total hydroperoxide and advanced
19		oxidation protein products in preterm hypoxic babies. Pediatr Res 2000;47:221-4
20	51.	Matsubasa T, Uchino T, Karashima S, et al. Oxidative stress in very low birth weight infants as
21		measured by urinary 8-OHdG. Free Radic Res. 2002;36:189-93
22	52.	Bandyopadhyay T, Bhatia BD, Khanna HD. A study of oxidative stress in neonates delivered
23		through meconium-stained amniotic fluid. Eur J Pediatr. 2017;176:317-325
24	53.	Gane BD, Bhat V, Rao R, Nandhakumar S, Harichandrakumar KT, Adhisivam B. Effect of
25		therapeutic hypothermia on DNA damage and neurodevelopmental outcome among term
26		neonates with perinatal asphyxia: a randomized controlled trial. J Trop Pediatr. 2014;60:134-40

1	54.	Fukuda M, Yamauchi H, Yamamoto H, et al. The evaluation of oxidative DNA damage in
2		children with brain damage using 8-hydroxydeoxyguanosine levels. Brain Dev. 2008;30:131-6
3	55.	Schwarcz R, Whetsell WO, Mangano RM. Quinolinic acid: an endogenous metabolite that
4		produces axon-sparing lesions in rat brain. Science 1983;219:316-8
5	56.	Santamaría A, Galván-Arzate S, Lisý V, et al. Quinolinic acid induces oxidative stress in rat
6		brain synaptosomes. Neuroreport 2001;12:871-4
7	57.	Solberg R, Andresen JH, Escrig R, Vento M, Saugstad OD. Resuscitation of Hypoxic Newborn
8		Piglets With Oxygen Induces a Dose-Dependent Increase in Markers of Oxidation. Pediatr Res
9		2007;62:559–63
10	58.	Solberg R, Longini M, Proietti F, Vezzosi P, Saugstad OD, Buonocore G. Resuscitation with
11		supplementary oxygen induces oxidative injury in the cerebral cortex. Free Radic Biol Med
12		2012;53:1061-7
13	59.	Vento M, Asensi M, Sastre J, García-Sala F, Pallardó F V, Viña J. Resuscitation with room air
14		instead of 100% oxygen prevents oxidative stress in moderately asphyxiated term neonates.
15		Pediatrics 2001;107:642-7
16	60.	Vento M, Asensi M, Sastre J, Lloret A, García-Sala F, Viña J. Oxidative stress in asphyxiated
17		term infants resuscitated with 100% oxygen. J Pediatr 2003;142:240-6
18	61.	Domoki F, Oláh O, Zimmermann A, et al. Hydrogen is neuroprotective and preserves
19		cerebrovascular reactivity in asphyxiated newborn pigs. Pediatr Res 2010;68:387-92
20	62.	Nemeth J, Toth-Szuki V, Varga V, Kovacs V, Remzso G, Domoki F. Molecular hydrogen
21		affords neuroprotection in a translational piglet model of hypoxic-ischemic encephalopathy. J
22		Physiol Pharmacol 2016;67:677–89
23	63.	Azzopardi D, Robertson NJ, Bainbridge A, et al. Moderate hypothermia within 6 h of birth plus
24		inhaled xenon versus moderate hypothermia alone after birth asphyxia (TOBY-Xe): a proof-of-
25		concept, open-label, randomised controlled trial. Lancet Neurol 2016;15:145-153.

1	64.	Rüegger CM, Davis PG, Cheong JL. Xenon as an adjuvant to therapeutic hypothermia in near-
2		term and term newborns with hypoxic-ischaemic encephalopathy. Cochrane database Syst Rev
3		2018;8:CD012753
4	65.	Broad KD, Fierens I, Fleiss B et al. Inhaled 45-50% argon augments hypothermic brain
5		protection in a piglet model of perinatal asphyxia. Neurobiol Dis. 2016;87:29-38
6	66.	Rutherford M, Ramenghi LA, Edwards AD, et al. Assessment of brain tissue injury after
7		moderate hypothermia in neonates with hypoxic-ischaemic encephalopathy: a nested substudy
8		of a randomised controlled trial. Lancet Neurol. 2010;9:39-45
9	67.	Azzopardi D, Strohm B, Marlow N, et al. Effects of hypothermia for perinatal asphyxia on
10		childhood outcomes. N Engl J Med. 2014;371:140-9
11	68.	Thoresen M, Penrice J, Lorek A, et al. Mild Hypothermia after Severe Transient Hypoxia-
12		Ischemia Ameliorates Delayed Cerebral Energy Failure in the Newborn Piglet. Pediatr Res
13		1995;37:667–70
14	69.	Drury PP, Gunn ER, Bennet L, Gunn AJ. Mechanisms of hypothermic neuroprotection. Clin
15		Perinatol 2014;41:161-75
16	70.	Ponnusamy V, Yip PK. The role of microRNAs in newborn brain development and hypoxic
17		ischaemic encephalopathy. Neuropharmacology 2019;149:55-65
18	71.	Tissier R, Chenoune M, Pons S, et al. Mild hypothermia reduces per-ischemic reactive oxygen
19		species production and preserves mitochondrial respiratory complexes. Resuscitation
20		2013;84:249-55.
21	72.	Zhao W, Richardson JS, Mombourquette MJ, Weil JA, Ijaz S, Shuaib A. Neuroprotective
22		effects of hypothermia and U-78517F in cerebral ischemia are due to reducing oxygen-based
23		free radicals: an electron paramagnetic resonance study with gerbils. J Neurosci Res
24		1996;45:282-8.

1	73.	Loidl CF, De Vente J, van Ittersum MM, et al. Hypothermia during or after severe perinatal
2		asphyxia prevents increase in cyclic GMP-related nitric oxide levels in the newborn rat
3		striatum. Brain Res 1998;791:303-7
4	74.	Huun MU, Garberg HT, Buonocore G, et al. Regional differences of hypothermia on oxidative
5		stress following hypoxia-ischemia: a study of DHA and hypothermia on brain lipid
6		peroxidation in newborn piglets. J Perinat Med 2018;47:82-9
7	75.	Hackenhaar FS, Medeiros TM, Heemann FM, et al. Therapeutic Hypothermia Reduces
8		Oxidative Damage and Alters Antioxidant Defenses after Cardiac Arrest. Oxid Med Cell
9		Longev 2017;2017:1–10
10	76.	Perrone S, Szabó M, Bellieni CV, et al. Whole Body Hypothermia and Oxidative Stress in
11		Babies With Hypoxic-Ischemic Brain Injury. Pediatr Neurol 2010;43:236–40
12	77.	Maiese K, Li F, Chong ZZ. New Avenues of Exploration for Erythropoietin. JAMA
13		2005;293:90-5
14	78.	Garg B, Sharma D, Bansal A. Systematic review seeking erythropoietin role for
15		neuroprotection in neonates with hypoxic ischemic encephalopathy: presently where do we
16		stand. J Matern Fetal Neonatal Med 2018;31:3214-24
17	79.	McAdams RM, Juul SE. Neonatal Encephalopathy: Update on Therapeutic Hypothermia and
18		Other Novel Therapeutics. Clin Perinatol 2016;43:485–500
19	80.	Wassink G, Davidson JO, Dhillon SK, et al. Partial white and grey matter protection with
20		prolonged infusion of recombinant human erythropoietin after asphyxia in preterm fetal sheep.
21		J Cereb Blood Flow Metab 2017;37:1080–94
22	81.	Kumral A, Genc S, Ozer E, et al. Erythropoietin Downregulates Bax and DP5 ProApoptotic
23		Gene Expression in Neonatal Hypoxic-Ischemic Brain Injury. Neonatology 2006;89:205–10
24	82.	Iwai M, Stetler RA, Xing J, et al. Enhanced oligodendrogenesis and recovery of neurological
25		function by erythropoietin after neonatal hypoxic/ischemic brain injury. Stroke 2010;41:1032-
26		7

1	83.	Traudt CM, McPherson RJ, Bauer LA, et al. Concurrent Erythropoietin and Hypothermia
2		Treatment Improve Outcomes in a Term Nonhuman Primate Model of Perinatal Asphyxia. Dev
3		Neurosci 2013;35:491–503
4	84.	McAdams RM, Fleiss B, Traudt C, et al. Long-Term Neuropathological Changes Associated
5		with Cerebral Palsy in a Nonhuman Primate Model of Hypoxic-Ischemic Encephalopathy. Dev
6		Neurosci 2017;39:124–40
7	85.	Pathipati P, Ferriero DM. The Differential Effects of Erythropoietin Exposure to Oxidative
8		Stress on Microglia and Astrocytes in vitro. Dev Neurosci 2017;39:310-22
9	86.	Solaroglu I, Solaroglu A, Kaptanoglu E, et al. Erythropoietin prevents ischemia-reperfusion
10		from inducing oxidative damage in fetal rat brain. Childs Nerv Syst 2003;19:19-22
11	87.	Wu YW, Mathur AM, Chang T, et al. High-Dose Erythropoietin and Hypothermia for
12		Hypoxic-Ischemic Encephalopathy: A Phase II Trial. Pediatrics 2016;137:e20160191
13	88.	Elmahdy H, El-Mashad A-R, El-Bahrawy H, El-Gohary T, El-Barbary A, Aly H. Human
14		Recombinant Erythropoietin in Asphyxia Neonatorum: Pilot Trial. Pediatrics 2010;125:e1135-
15		42
16	89.	Zhu C, Kang W, Xu F, et al. Erythropoietin Improved Neurologic Outcomes in Newborns With
17		Hypoxic-Ischemic Encephalopathy. Pediatrics 2009;124:e218–26
18	90.	Malla RR, Asimi R, Teli MA, Shaheen F, Bhat MA. Erythropoietin monotherapy in perinatal
19		asphyxia with moderate to severe encephalopathy: a randomized placebo-controlled trial. J
20		Perinatol 2017;37:596–601
21	91.	Avasiloaiei A, Dimitriu C, Moscalu M, Paduraru L, Stamatin M. High-dose phenobarbital or
22		erythropoietin for the treatment of perinatal asphyxia in term newborns. Pediatr Int
23		2013;55:589–93
24	92.	El Shimi MS, Awad HA, Hassanein SMA, et al. Single dose recombinant erythropoietin versus
25		moderate hypothermia for neonatal hypoxic ischemic encephalopathy in low resource settings.
26		J Matern Neonatal Med 2014;27:1295–300

- 93. Reiter RJ, Tan DX, Osuna C, Gitto E. Actions of melatonin in the reduction of oxidative stress.
 A review. J Biomed Sci 2000;7:444–58
- 94. Blanco S, Hernández R, Franchelli G, Ramos-Álvarez MM, Peinado MÁ. Melatonin influences
 NO/NOS pathway and reduces oxidative and nitrosative stress in a model of hypoxic-ischemic
 brain damage. Nitric Oxide 2017;62:32–43
- 6 95. Aridas JDS, Yawno T, Sutherland AE, et al. Systemic and transdermal melatonin
- administration prevents neuropathology in response to perinatal asphyxia in newborn lambs. J
 Pineal Res 2018;64:e12479
- 9 96. Kaur C, Sivakumar V, Ling EA. Melatonin protects periventricular white matter from damage
 10 due to hypoxia. J Pineal Res 2010;48:185–93
- 11 97. Signorini C, Ciccoli L, Leoncini S, et al. Free iron, total F 2 -isoprostanes and total F 4 -
- 12 neuroprostanes in a model of neonatal hypoxic-ischemic encephalopathy: neuroprotective

13 effect of melatonin. J Pineal Res 2009;46:148–54

- 14 98. Carloni S, Perrone S, Buonocore G, Longini M, Proietti F, Balduini W. Melatonin protects
- from the long-term consequences of a neonatal hypoxic-ischemic brain injury in rats. J Pineal
 Res 2008;44:157–64
- 17 99. Welin A-K, Svedin P, Lapatto R, et al. Melatonin reduces inflammation and cell death in white
- 18 matter in the mid-gestation fetal sheep following umbilical cord occlusion. Pediatr Res
- 19 2007;61:153–8
- 20 100.Robertson NJ, Faulkner S, Fleiss B, et al. Melatonin augments hypothermic neuroprotection in
 21 a perinatal asphyxia model. Brain 2013;136:90–105
- 22 101.Su F, Guo AC, Li WW, et al. Low-Dose Ethanol Preconditioning Protects Against Oxygen-
- 23 Glucose Deprivation/Reoxygenation-Induced Neuronal Injury By Activating Large
- 24 Conductance, Ca2+-Activated K+ Channels In Vitro. Neurosci Bull. 2017;33:28-40

- 1 102. Robertson NJ, Martinello K, Lingam I, et al. Melatonin as an adjunct to therapeutic
- 2 hypothermia in a piglet model of neonatal encephalopathy: A translational study. Neurobiol
 3 Dis. 2019;121:240-251
- 4 103.Fulia F, Gitto E, Cuzzocrea S, et al. Increased levels of malondialdehyde and nitrite/nitrate in
- 5 the blood of asphyxiated newborns: reduction by melatonin. J Pineal Res 2001;31:343–9
- 6 104.Aly H, Elmahdy H, El-Dib M, et al. Melatonin use for neuroprotection in perinatal asphyxia: a
 7 randomized controlled pilot study. J Perinatol 2015;35:186–91
- 8 105.Miller SL, Yan EB, Castillo-Meléndez M, Jenkin G, Walker DW. Melatonin Provides
- 9 Neuroprotection in the Late-Gestation Fetal Sheep Brain in Response to Umbilical Cord
- 10 Occlusion. Dev Neurosci 2005;27:200–10
- 106.Palmer C, Vannucci RC, Towfighi J. Reduction of Perinatal Hypoxic-Ischemic Brain Damage
 with Allopurinol. Pediatr Res 1990;27:332–6
- 13 107.Palmer C, Towfighi J, Roberts RL, Heitjan DF. Allopurinol administered after inducing
- 14 hypoxia-ischemia reduces brain injury in 7-day-old rats. Pediatr Res 1993;33:405–11
- 15 108.Rodríguez-Fanjul J, Durán Fernández-Feijóo C, Lopez-Abad M, et al. Neuroprotection with
- 16 hypothermia and allopurinol in an animal model of hypoxic-ischemic injury: Is it a gender
- 17 question? PLoS One. 2017;12:e0184643
- 18 109. Van Bel F, Shadid M, Moison RM, et al. Effect of allopurinol on postasphyxial free radical
- 19 formation, cerebral hemodynamics, and electrical brain activity. Pediatrics 1998;101:185–93
- 20 110.Benders MJ, Bos AF, Rademaker CMA, et al. Early postnatal allopurinol does not improve
- short term outcome after severe birth asphyxia. Arch. Dis. Child. Fetal Neonatal Ed.

22 2006;91:F163-5

- 23 111.Kaandorp JJ, van Bel F, Veen S, et al. Long-term neuroprotective effects of allopurinol after
- 24 moderate perinatal asphyxia: follow-up of two randomised controlled trials. Arch Dis Child -
- 25 Fetal Neonatal Ed 2012;97:F162–6

1	112.Kane AD, Hansell JA, Herrera EA, et al. Xanthine oxidase and the fetal cardiovascular defence
2	to hypoxia in late gestation ovine pregnancy. J Physiol. 2014;592:475-89
3	113.Torrance HL, Benders MJ, Derks JB, et al. Maternal allopurinol during fetal hypoxia lowers
4	cord blood levels of the brain injury marker S-100B. Pediatrics 2009;124:350-7
5	114.Kaandorp JJ, Benders MJNL, Schuit E, et al. Maternal allopurinol administration during
6	suspected fetal hypoxia: a novel neuroprotective intervention? A multicentre randomised
7	placebo controlled trial. Arch Dis Child - Fetal Neonatal Ed 2015;100:F216–23
8	115.Klumper J, Kaandorp JJ, Schuit E, et al. Behavioral and neurodevelopmental outcome of
9	children after maternal allopurinol administration during suspected fetal hypoxia: 5-year follow
10	up of the ALLO-trial. PLoS One 2018;13:e0201063
11	116.Chaudhari T, McGuire W. Allopurinol for preventing mortality and morbidity in newborn
12	infants with hypoxic-ischaemic encephalopathy. Cochrane Database Syst Rev.
13	2012;7:CD006817
14	117.Favié LMA, Cox AR, van den Hoogen A, et al. Nitric Oxide Synthase Inhibition as a
15	Neuroprotective Strategy Following Hypoxic-Ischemic Encephalopathy: Evidence From
16	Animal Studies. Front Neurol 2018;9:258
17	118. Trifiletti RR. Neuroprotective effects of NG-nitro-L-arginine in focal stroke in the 7-day old
18	rat. Eur J Pharmacol 1992;218:197–8
19	119. Tsuji M, Higuchi Y, Shiraishi K, Kume T, Akaike A, Hattori H. Protective effect of
20	aminoguanidine on hypoxic-ischemic brain damage and temporal profile of brain nitric oxide
21	in neonatal rat. Pediatr Res 2000;47:79-83
22	120.Hamada Y, Hayakawa T, Hattori H, Mikawa H. Inhibitor of Nitric Oxide Synthesis Reduces
23	Hypoxic-Ischemic Brain Damage in the Neonatal Rat. Pediatr Res 1994;35:10-4
24	121.Muramatsu K, Sheldon RA, Black SM, Täuber M, Ferriero DM. Nitric oxide synthase activity
25	and inhibition after neonatal hypoxia ischemia in the mouse brain. Brain Res Dev Brain Res
26	2000;123:119–27

1	122.Ishida A, Trescher WH, Lange MS, Johnston M V. Prolonged suppression of brain nitric oxide
2	synthase activity by 7-nitroindazole protects against cerebral hypoxic-ischemic injury in
3	neonatal rat. Brain Dev 2001;23:349–54
4	123.Nijboer CHA, Groenendaal F, Kavelaars A, Hagberg HH, van Bel F, Heijnen CJ. Gender-
5	specific neuroprotection by 2-iminobiotin after hypoxia-ischemia in the neonatal rat via a nitric
6	oxide independent pathway. J Cereb Blood Flow Metab 2007;27:282-92
7	124.van den Tweel ERW, van Bel F, Kavelaars A, et al. Long-term neuroprotection with 2-
8	iminobiotin, an inhibitor of neuronal and inducible nitric oxide synthase, after cerebral
9	hypoxia-ischemia in neonatal rats. J Cereb Blood Flow Metab 2005;25:67–74
10	125.Zitta K, Peeters-Scholte C, Sommer L, et al. 2-Iminobiotin Superimposed on Hypothermia
11	Protects Human Neuronal Cells from Hypoxia-Induced Cell Damage: An in Vitro Study. Front
12	Pharmacol 2017;8:971
13	126.Yu L, Derrick M, Ji H, et al. Neuronal nitric oxide synthase inhibition prevents cerebral palsy
14	following hypoxia-ischemia in fetal rabbits: comparison between JI-8 and 7-nitroindazole. Dev
15	Neurosci 2011;33:312–9
16	127.Rao S, Lin Z, Drobyshevsky A, et al. Involvement of Neuronal Nitric Oxide Synthase in
17	Ongoing Fetal Brain Injury following Near-Term Rabbit Hypoxia-Ischemia. Dev Neurosci
18	2011;33:288–98
19	128.Drury PP, Davidson JO, van den Heuij LG, et al. Partial neuroprotection by nNOS inhibition
20	during profound asphyxia in preterm fetal sheep. Exp Neurol 2013;250:282-92
21	129.Drury PP, Davidson JO, Mathai S, et al. nNOS inhibition during profound asphyxia reduces
22	seizure burden and improves survival of striatal phenotypic neurons in preterm fetal sheep.
23	Neuropharmacology 2014;83:62–70
24	130.Elbini Dhouib I, Jallouli M, Annabi A, Gharbi N, Elfazaa S, Lasram MM. A minireview on N-
25	acetylcysteine: An old drug with new approaches. Life Sci 2016;151:359-63

1	131.Jenkins DD, Wiest DB, Mulvihill DM, et al. Fetal and Neonatal Effects of N-Acetylcysteine
2	When Used for Neuroprotection in Maternal Chorioamnionitis. J Pediatr 2016;168:67-76
3	132.Benterud T, Ystgaard MB, Manueldas S, et al. N-Acetylcysteine Amide Exerts Possible
4	Neuroprotective Effects in Newborn Pigs after Perinatal Asphyxia. Neonatology 2017;111:12-
5	21
6	133.Benterud T, Manueldas S, Rivera S, et al. Cerebellum Susceptibility to Neonatal Asphyxia:
7	Possible Protective Effects of N-Acetylcysteine Amide. Dis Markers 2018;2018:5046372
8	134.Lee TF, Tymafichuk CN, Bigam DL, Cheung PY. Effects of postresuscitation N-acetylcysteine
9	on cerebral free radical production and perfusion during reoxygenation of hypoxic newborn
10	piglets. Pediatr Res 2008;64:256–61
11	135.Liu JQ, Lee TF, Chen C, Bagim DL, Cheung PY. N-Acetylcysteine Improves Hemodynamics
12	and Reduces Oxidative Stress in the Brains of Newborn Piglets with Hypoxia-Reoxygenation
13	Injury. J Neurotrauma 2010;27:1865–73
14	136.Wang X, Svedin P, Nie C, et al. N-acetylcysteine reduces lipopolysaccharide-sensitized
15	hypoxic-ischemic brain injury. Ann Neurol 2007;61:263–71
16	137.Jatana M, Singh I, Singh AK, Jenkins D. Combination of Systemic Hypothermia and N-
17	acetylcysteine Attenuates Hypoxic-Ischemic Brain Injury in Neonatal Rats. Pediatr Res
18	2006;59:684–9
19	138.Moss HG, Brown TR, Wiest DB, Jenkins DD. N-Acetylcysteine rapidly replenishes central
20	nervous system glutathione measured via magnetic resonance spectroscopy in human neonates
21	with hypoxic-ischemic encephalopathy. J Cereb Blood Flow Metab 2018;38:950-958.
22	139. Jenkins DD, Wiest DB, Mulvihill DM, et al. Fetal and Neonatal Effects of N-Acetylcysteine
23	When Used for Neuroprotection in Maternal Chorioamnionitis. J Pediatr. 2016;168:67-76.e6.
24	140.Wang X, Zhao X, Mao Z-Y, Wang X-M, Liu Z-L. Neuroprotective effect of docosahexaenoic
25	acid on glutamate-induced cytotoxicity in rat hippocampal cultures. Neuroreport
26	2003;14:2457–61

1	141.Solberg R, Longini M, Proietti F, et al. DHA Reduces Oxidative Stress after Perinatal
2	Asphyxia: A Study in Newborn Piglets. Neonatology 2017;112:1-8
3	142.Huun MU, Garberg HT, Escobar J, et al. DHA reduces oxidative stress following hypoxia-
4	ischemia in newborn piglets: a study of lipid peroxidation products in urine and plasma. J
5	Perinat Med 2018: 46:209-217
6	143.Huun MU, Garberg H, Løberg EM, et al. DHA and therapeutic hypothermia in a short-term
7	follow-up piglet model of hypoxia-ischemia: Effects on H+MRS biomarkers. PLoS One
8	2018;13:e0201895
9	144.Berman DR, Mozurkewich E, Liu Y, Barks J. Docosahexaenoic acid pretreatment confers
10	neuroprotection in a rat model of perinatal cerebral hypoxia-ischemia. Am J Obstet Gynecol
11	2009;200:305.e1-6.
12	145.Berman DR, Liu Y, Barks J, Mozurkewich E. Treatment with docosahexaenoic acid after
13	hypoxia-ischemia improves forepaw placing in a rat model of perinatal hypoxia-ischemia. Am
14	J Obstet Gynecol 2010;203:385.e1-385.e5
15	146.Yoshida H, Yanai H, Namiki Y, Fukatsu-Sasaki K, Furutani N, Tada N. Neuroprotective
16	effects of edaravone: a novel free radical scavenger in cerebrovascular injury. CNS Drug Rev
17	2006 Spring;12:9-20
18	147. Yasuoka N, Nakajima W, Ishida A, Takada G. Neuroprotection of edaravone on hypoxic-
19	ischemic brain injury in neonatal rats. Brain Res Dev Brain Res 2004;151:129-39
20	148. Noor JI, Ueda Y, Ikeda T, Ikenoue T. Edaravone inhibits lipid peroxidation in neonatal
21	hypoxic-ischemic rats: an in vivo microdialysis study. Neurosci Lett. 2007;414:5-9.
22	149. Takizawa Y, Miyazawa T, Nonoyama S, Goto Y, Itoh M. Edaravone inhibits DNA
23	peroxidation and neuronal cell death in neonatal hypoxic-ischemic encephalopathy model rat.
24	Pediatr Res 2009;65:636-41

1	150.Nakamoto H, Aihara Y, Yamaguchi K, Kawamata T, Okada Y. Efficacy, safety, and outcomes
2	in 17 pediatric cases treated with the free radical scavenger edaravone. Childs Nerv Syst
3	2015;31:1533-40
4	151.Bharadwaj S, Bhat VB, Vickneswaran V, Adhisivam B, Zachariah B, Habeebullah S.Oxidative
5	stress in preeclamptic mother - newborn dyads and its correlation with early neonatal outcome -
6	a case control study. J Matern Fetal Neonatal Med 2018;31:1548-1553
7	152.Chelchowska M, Ambroszkiewicz J, Gajewska J, Laskowska-Klita T, Leibschang J. The effect
8	of tobacco smoking during pregnancy on plasma oxidant and antioxidant status in mother and
9	newborn. Eur J Obstet Gynecol Reprod Biol 2011;155:132-6
10	153.Poggi C, Dani C. Sepsis and Oxidative Stress in the Newborn: From Pathogenesis to Novel
11	Therapeutic Targets. Oxid Med Cell Longev 2018; 2018: 9390140
12	154.Marseglia L, D'Angelo G, Granese R, et al. Role of oxidative stress in neonatal respiratory
13	distress syndrome. Free Radic Biol Med 2019 pii: S0891-5849(19)30070-X

1 Figure legends

3	Figure 1. Summary of the phases of neonatal encephalopathy with related mechanisms of injury and
4	proposed antioxidant strategies.
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6	Figure 2. Mechanisms of free radical production following hypoxia-ischemia and reperfusion.
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