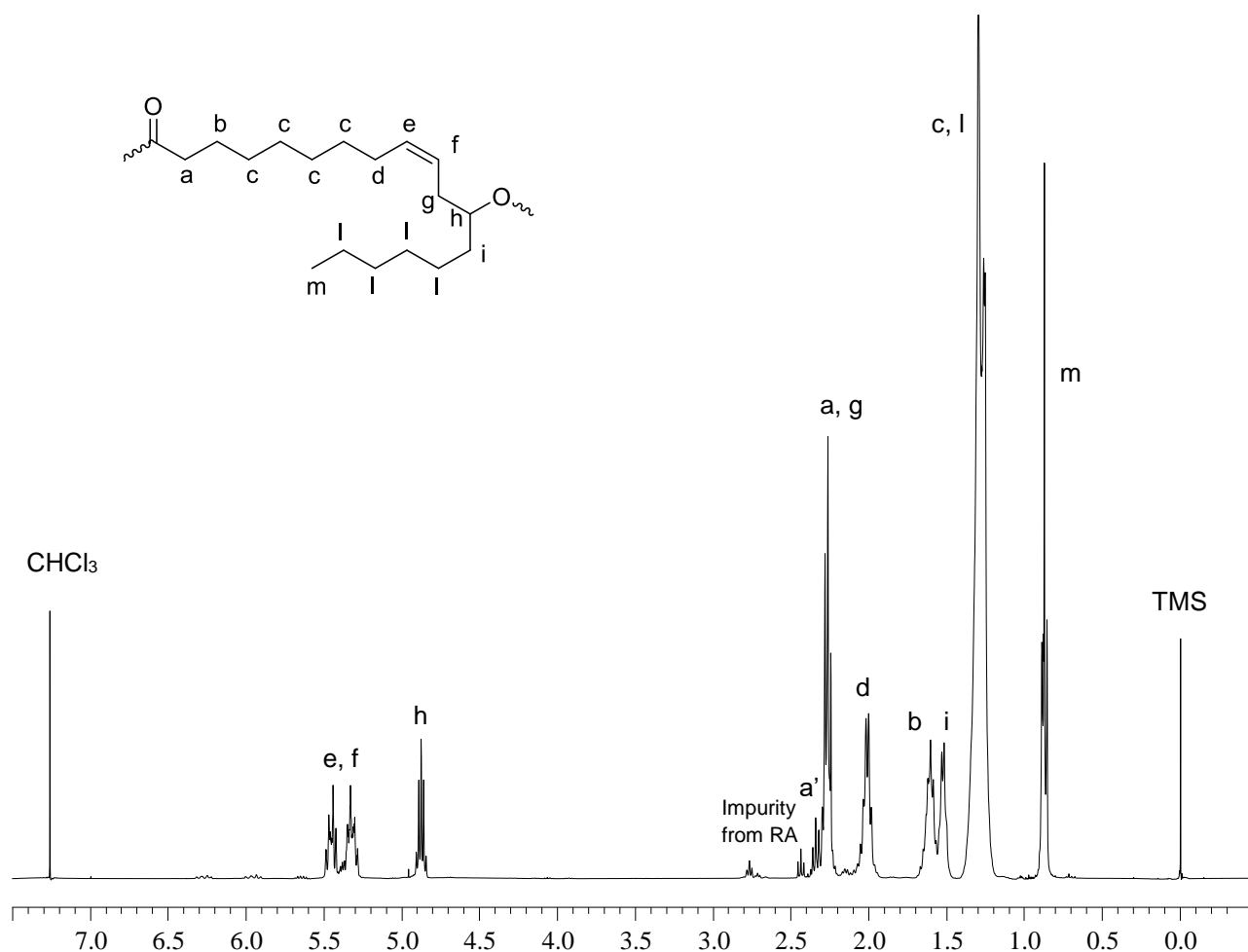


## Elastomeric/antibacterial properties in novel random *Ricinus communis* based-copolyesters

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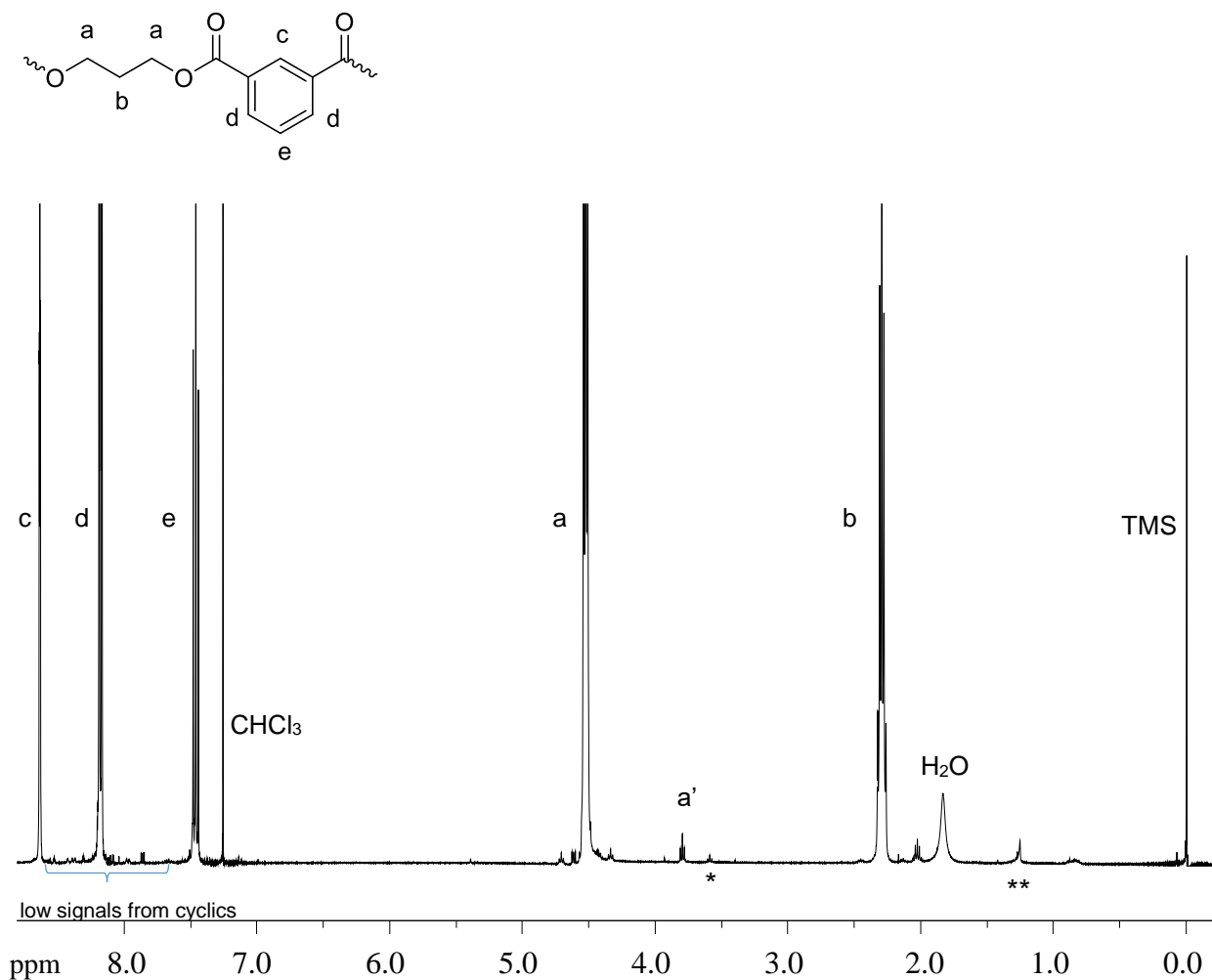
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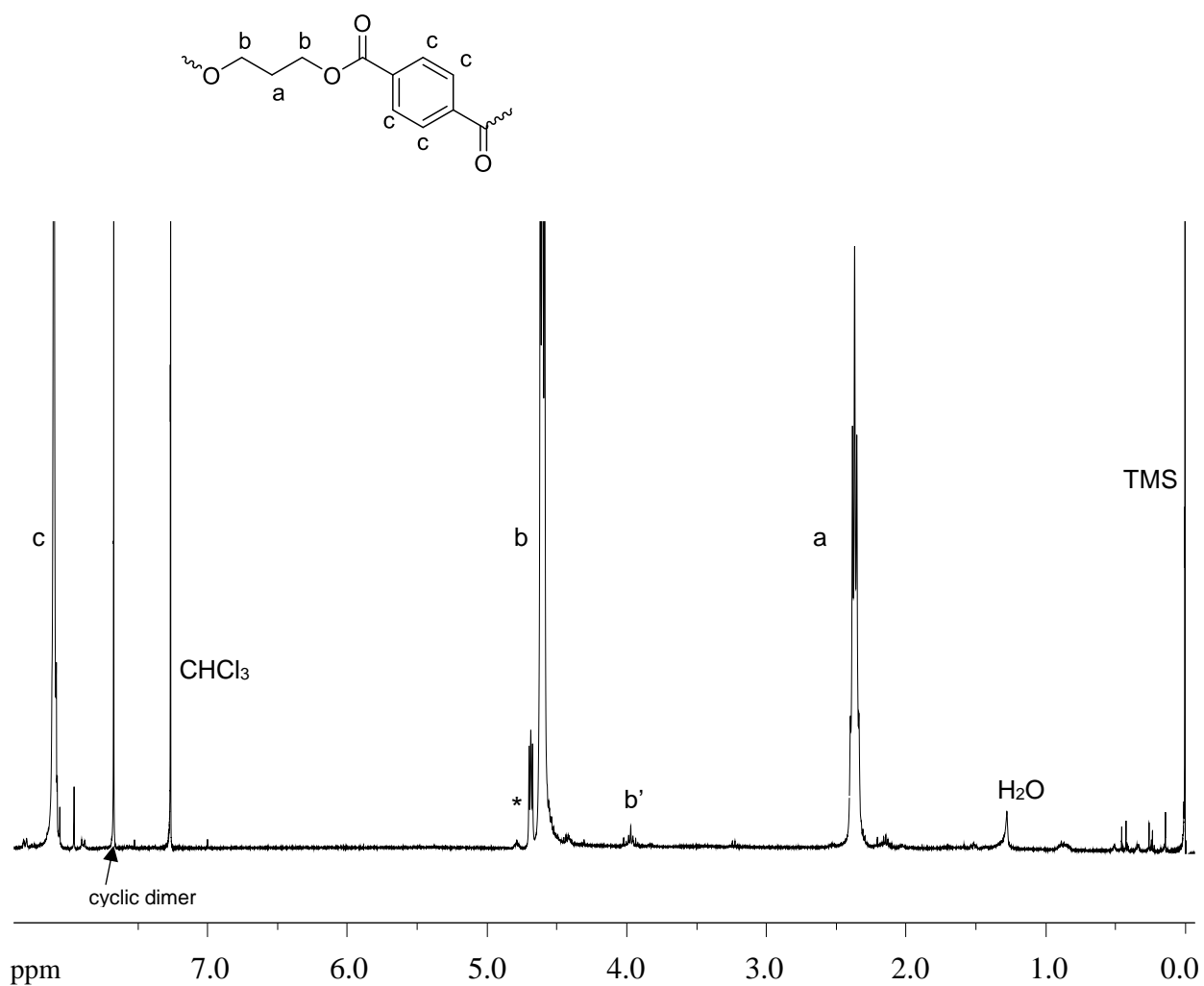
**Figure S1.** <sup>1</sup>H NMR (400 MHz) spectrum of poly(ricinoleic acid) in CDCl<sub>3</sub> (a' end group).

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>, δ): 0.80-0.88 (t, 3H; C<sup>m</sup>H<sub>3</sub>), 1.20-1.38 (m, 16H; C<sup>c</sup>H<sub>2</sub> and C<sup>l</sup>H<sub>2</sub>), 1.42-1.56 and 1.56-1.70 (m, 2H; C<sup>i</sup>H<sub>2</sub> and C<sup>b</sup>H<sub>2</sub>), 1.95-2.10 (m, 2H; C<sup>d</sup>H<sub>2</sub>), 2.23-2.40 (m, 4H; C<sup>a</sup>H<sub>2</sub> and C<sup>g</sup>H<sub>2</sub>), 4.82-4.92 (m, 1H; C<sup>h</sup>H), 5.28-5.36 and 5.40-5.50 (2m, 2H; C<sup>f</sup>H and C<sup>e</sup>H).



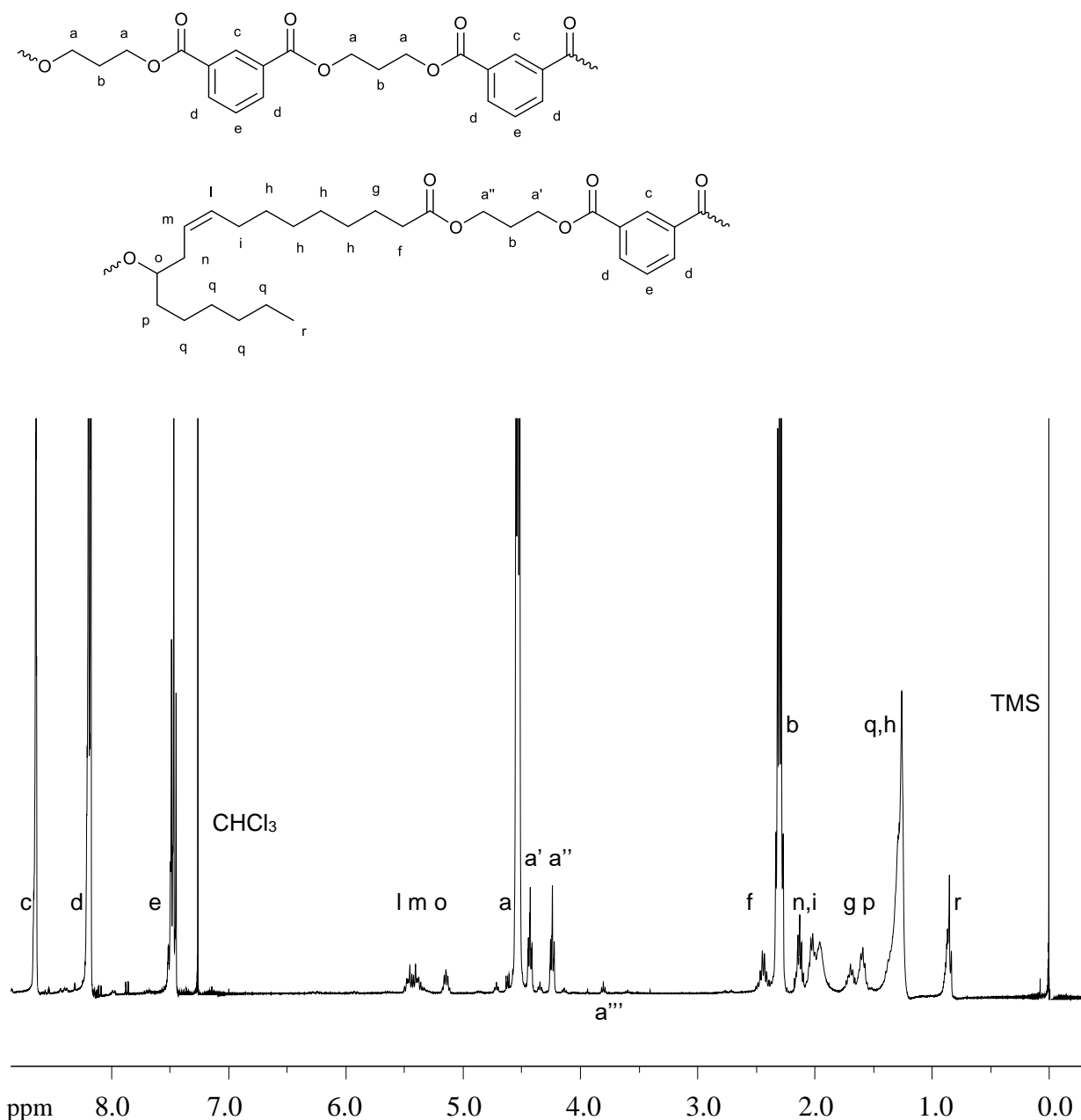
**Figure S2.**  $^1\text{H}$  NMR (400 MHz) spectrum of poly(propylene isophthalate) in  $\text{CDCl}_3$  (a' end group, \* ether linkages PD-PD, \*\* impurity).

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$   $\delta$ ): 2.22-2.34 (m, 2H;  $\text{C}^b\text{H}_2$ ), 4.47-4.56 (t, 4H;  $\text{C}^a\text{H}_2$ ), 7.43-7.49 (t, 1H;  $\text{C}^e\text{H}$ ), 8.15-8.20 (d, 2H;  $\text{C}^d\text{H}$ ), 8.62-8.66 (t, 1H;  $\text{C}^c\text{H}$ ).



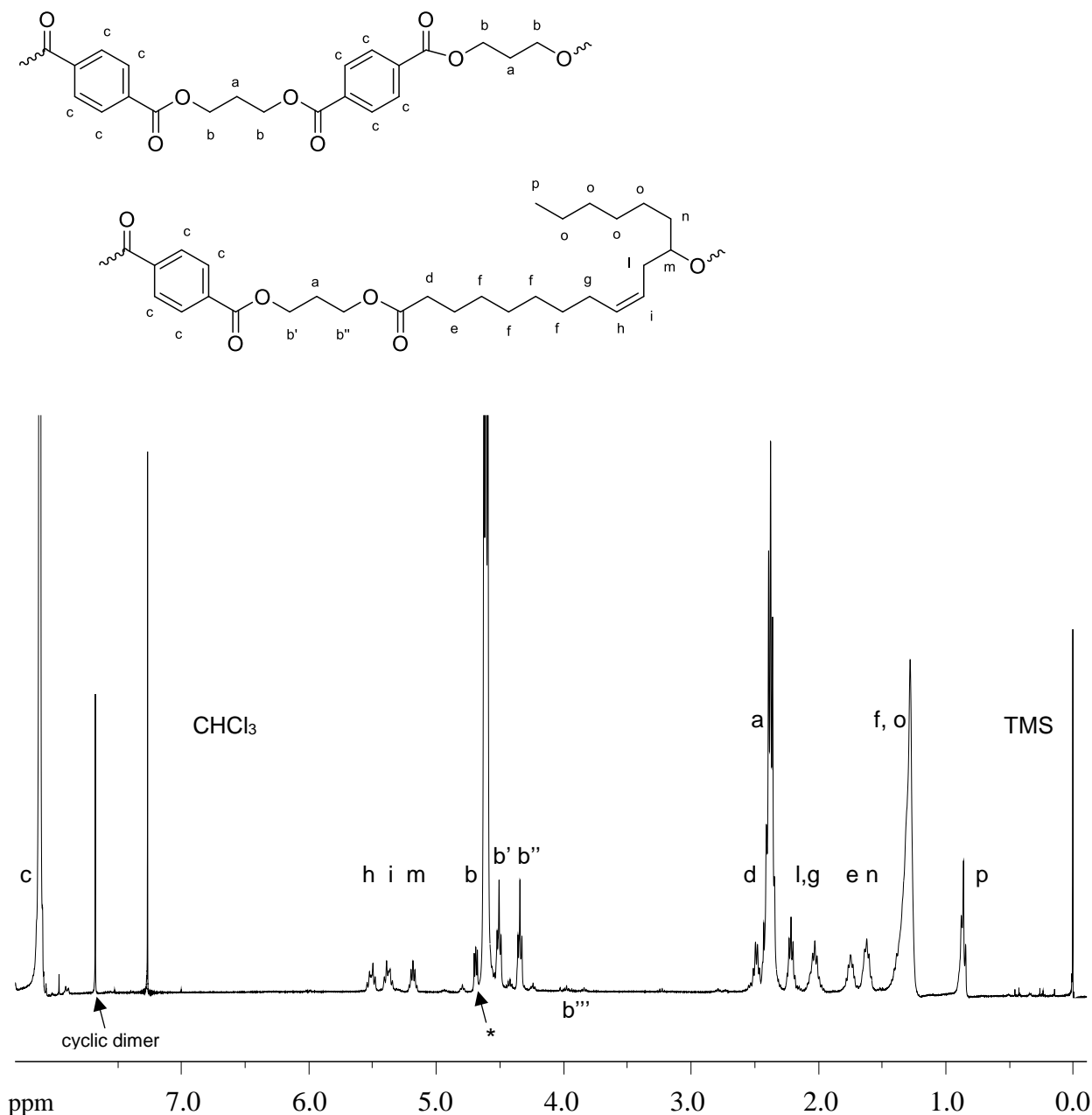
**Figure S3.** <sup>1</sup>H NMR (400 MHz) spectrum of poly(propylene terephthalate) in CDCl<sub>3</sub>/TFA (b' end group, \* cyclics) [1].

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>/TFA, δ): 2.24-2.52 (m, 2H; C<sup>a</sup>H<sub>2</sub>), 4.45-4.66 (m, 4H; C<sup>b</sup>H<sub>2</sub>), 7.98-8.26 (s, 4H; C<sup>c</sup>H<sub>2</sub>).



**Figure S4.**  $^1\text{H}$  NMR (400 MHz) spectrum of poly(propylene isophthalate-*co*-ricinoleic acid) (P(PI-*co*-RA) 90/10) in  $\text{CDCl}_3/\text{TFA}$  ( $\text{a}'''$  end group).

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3/\text{TFA}$ ,  $\delta$ ): 0.80-0.95 (t, 3H;  $\text{C}^{\text{r}}\text{H}_3$ ), 1.20-1.48 (m, 16H;  $\text{C}^{\text{q}}\text{H}_2$  and  $\text{C}^{\text{h}}\text{H}_2$ ), 1.55-1.65 and 1.65-1.75 (m, 2H;  $\text{C}^{\text{p}}\text{H}_2$  and  $\text{C}^{\text{g}}\text{H}_2$ ), 1.80-2.10 (m, 2H;  $\text{C}^{\text{l}}\text{H}_2$ ), 2.10-2.20 (m, 2H;  $\text{C}^{\text{n}}\text{H}_2$ ), 2.23-2.40 (m, 2H;  $\text{C}^{\text{b}}\text{H}_2$ ), 2.40-2.50 (m, 2H;  $\text{C}^{\text{f}}\text{H}_2$ ), 4.15-4.30 (m, 2H;  $\text{C}^{\text{a}''}\text{H}_2$ ), 4.35-4.45 (m, 2H;  $\text{C}^{\text{a}'}\text{H}_2$ ), 4.45-4.55 (m, 4H;  $\text{C}^{\text{a}}\text{H}_2$ ), 5.10-5.20 (m, 1H;  $\text{C}^{\text{o}}\text{H}$ ), 5.35-5.45 and 5.45-5.50 (2m, 2H;  $\text{C}^{\text{m}}\text{H}$  and  $\text{C}^{\text{l}}\text{H}$ ), 7.42-7.55 (m, 1H;  $\text{C}^{\text{e}}\text{H}$ ), 8.15-8.25 (d, 2H;  $\text{C}^{\text{d}}\text{H}$ ), 8.60-8.72 (s, 1H;  $\text{C}^{\text{c}}\text{H}$ ).



**Figure S5.**  $^1\text{H}$  NMR (400 MHz) spectrum of poly(propylene terephthalate-*co*-ricinoleic acid) (P(PT-*co*-RA) 90/10) in  $\text{CDCl}_3/\text{TFA}$  ( $\text{b}''''$  end group, \* cyclics) [1].

$^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3/\text{TFA}$ ,  $\delta$ ): 0.86 (t, 3H;  $\text{C}^{\text{p}}\text{H}_3$ ), 1.20-1.50 (m, 16H;  $\text{C}^{\text{f}}\text{H}_2$  and  $\text{C}^{\text{o}}\text{H}_2$ ), 1.54-1.69 (m, 2H;  $\text{C}^{\text{n}}\text{H}_2$ ), 1.68-1.82 (m, 2H;  $\text{C}^{\text{e}}\text{H}_2$ ), 1.94-2.10 (m, 2H;  $\text{C}^{\text{g}}\text{H}_2$ ), 2.14-2.26 (m, 4H;  $\text{C}^{\text{l}}\text{H}_2$ ), 2.26-2.45 (m, 2H;  $\text{C}^{\text{a}}\text{H}_2$ ), 2.45-2.56 (m, 4H;  $\text{C}^{\text{d}}\text{H}_2$ ), 4.30-4.40 (m, 2H;  $\text{C}^{\text{b}''''}\text{H}_2$ ), 4.45-4.52 (m, 2H;  $\text{C}^{\text{b}'}\text{H}_2$ ), 4.53-4.75 (m, 4H;  $\text{C}^{\text{b}''}\text{H}_2$ ), 5.09-5.25 (m, 1H;  $\text{C}^{\text{m}}\text{H}$ ), 5.25-5.44 and 5.45-5.60 (2m, 2H;  $\text{C}^{\text{i}}\text{H}$  and  $\text{C}^{\text{h}}\text{H}$ ), 7.93-8.29 (s, 4H;  $\text{C}^{\text{c}}\text{H}_2$ ).

**Table S1.** Average number of viable cells obtained after 24 h of contact with samples as described in section 2.2.7. The values regarding the bacterial cell suspensions used as positive controls have also been reported.

Sample	<i>E.coli</i>	<i>S.aureus</i>
PRA	0	0
P(PI- <i>co</i> -RA)-75/25	$5.0 \times 10^2$	$6.1 \times 10^4$
P(PI- <i>co</i> -RA)-82/18	$7.8 \times 10^4$	$1.7 \times 10^6$
P(PI- <i>co</i> -RA)-90/10	$1.9 \times 10^5$	$3.8 \times 10^6$
PPI	$1.0 \times 10^6$	$2.3 \times 10^6$
P(PT- <i>co</i> -RA)-75/25	$8.0 \times 10^4$	$6.4 \times 10^4$
P(PT- <i>co</i> -RA)-90/10	$1.4 \times 10^5$	$2.3 \times 10^6$
PPT	$2.2 \times 10^5$	$2.6 \times 10^6$
Bacterial cell suspension	$1.2 \times 10^5$	$3.0 \times 10^6$

## References

- [1] B. Min, B. Ho Lim, S. Yen Ko, Separation and Identification of Cyclic Oligomers in Poly (trimethylene terephthalate). *Journal of the Korean Magnetic Resonance Society* 10 (2006) 38 – 45.