

Alma Mater Studiorum Università di Bologna Archivio istituzionale della ricerca

Genomic Profiling of Primary Diffuse Large B-Cell Lymphoma of the Central Nervous System Suggests Novel Potential Therapeutic Targets

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Agostinelli, C., Morandi, L., Righi, S., Cirillo, L., Iommi, M., Tonon, C., et al. (2023). Genomic Profiling of Primary Diffuse Large B-Cell Lymphoma of the Central Nervous System Suggests Novel Potential Therapeutic Targets. MODERN PATHOLOGY, 36(12), 1-7 [10.1016/j.modpat.2023.100323].

Availability:

[This version is available at: https://hdl.handle.net/11585/955073 since: 2024-09-09](https://hdl.handle.net/11585/955073)

Published:

[DOI: http://doi.org/10.1016/j.modpat.2023.100323](http://doi.org/10.1016/j.modpat.2023.100323)

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

> This item was downloaded from IRIS Università di Bologna (https://cris.unibo.it/). When citing, please refer to the published version.

> > (Article begins on next page)

Genomic profiling of CNS-DLBC suggests novel potential therapeutic targets.

3 Claudio Agostinelli^{1,2}, Luca Morandi^{3,4}, Simona Righi², Luigi Cirillo⁴, Marica Iommi³, Caterina 4 Tonon^{3,4}, Diego Mazzatenta⁵, Matteo Zoli^{3,4}, Maura Rossi¹, Gianmarco Bagnato^{2,6}, Alessandro 5 Broccoli^{2,6}, Raffaele Lodi^{3,4}, Pier Luigi Zinzani^{2,6}, Elena Sabattini¹, Caterina Giannini^{3,7}, Sofia 6 $\text{Asioli}^{3,5}$.

8¹ Haematopathology Unit, IRCCS Azienda Ospedaliero-Universitaria of Bologna, Via Massarenti 9, 40138, Bologna, Italy.

² Department of Medical and Surgical Sciences, University of Bologna, 40138 Bologna, Italy

³Department of Biomedical and Neuromotor Sciences, University of Bologna, 40138 Bologna, Italy.

⁴ IRCCS Istituto delle Scienze Neurologiche di Bologna, Functional and Molecular Neuroimaging Unit, Bologna, Italy.

⁵ IRCCS Istituto delle Scienze Neurologiche di Bologna, Center for the Diagnosis and Treatment of Hypothalamic-Pituitary Diseases, Pituitary Unit

⁶IRCCS Azienda Ospedaliero-Universitaria di Bologna Istituto di Ematologia "Seràgnoli" Bologna Italy.

⁷ Department of Laboratory Medicine and Pathology, Mayo Clinic, Rochester, Minnesota, USA

Correspondence: Sofia Asioli, MD; Department of Biomedical and Neuromotor Sciences, Alma Mater Studiorum - University of Bologna; IRCCS Istituto delle Scienze Neurologiche di Bologna Via Altura No. 3, 40126 Bologna; Tel 0516225006; Fax 0516225759; Email: sofia.asioli3@unibo.it

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Author Contributions: All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by AS, CA, SR, LM; MI performed statistical analysis. The draft of the manuscript was written by CA, SA, CG. All authors read and approved the final manuscript. This work was partially presented as poster presentation (#1091) at the 111th Meeting of the United States and Canadian Academy of Pathology in Los Angeles , CA , USA, 19-24 March, 2022. **Acknowledgements and Funding**: The publication of this article was supported by the "Ricerca Corrente" funding from the Italian Ministry of Health to SA. This work was supported by funds for

selected research topics from the Fondazione CARISBO Project (#19085) to SA.

Data Availability Statement: The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request

-
-
-
-
-
-
-
-
-
-
-
-
-
-
-
-

ABSTRACT

Primary diffuse large B-cell lymphoma of the CNS (CNS-DLBCL) is an aggressive disease, with dismal prognosis despite the use of high dose methotrexate (MTX)-based polychemotherapy. Our study aimed to expand the biologic profiles of CNS-DLBCL and to correlate them with clinical/imaging findings to gain diagnostic insight and possibly identify new therapeutic targets. We selected 61 CNS-DLBCL whose FFPE samples at first diagnosis were available. These were investigated by immunohistochemistry, cMYC rearrangements were explored by fluorescence in situ hybridization and CNS-DLBCL mutated genes were evaluated by Next Generation Sequencing. CD10, BCL6 and IRF4 were observed in 16%, 83.6% and 93% of cases respectively. As typical of CNS-lymphoma 10/61 (16.4%) cases were classified as germinal center (GCB) and type and 51/61 (83.6%) as non-germinal center (non-GCB) type according to the Hans algorithm. Double expression (DE) status for BCL2 and cMYC was detected in 36/61 (59%) cases while 25/61 (41%) were non DE. Rearrangement of the *cMYC* gene was detected in 2 cases, associated with *BCL6* translocation only 1 case. *MYD88, PIM1*, *CD79B* and *TP53* were mutated in 54.5%, *53.5%*, 30.2% and 18.4% cases respectively. Novel mutations not previously reported in CNS-DLBCL were found: *AIP* in 23,1%, *PI3KCA* in 15%, *NOTCH1 in* 11,4%, *GNAS* 8.1%, *CASP8 in* 7.9%, *EGFR* in 6.4% *PTEN* in 5.1 and *KRAS* in 2.6%. Survival was significantly longer for patients with mutated *MYD88* (8.7 months vs. 1.7 months; $log-rank \text{ test} = 5.43$; $p = 0.020$) and for patients with mutated *CD79B* (10.8 months vs. 2.5 months; log-rank test = 4.64; p = 0.031).*MYD88* and *CD79B* predicted a longer survival in patients affected by CNS-DLBCL. Notably, we identified novel mutations that enrich mutational landscape of CNS-DLBCL, suggest a role of PTEN-PI3K-AKT and RTK-RAS-MAPK signalling in a subset of CNS-DLBCL and provide new potential therapeutic targets.

-
-
-

79 **Introduction**

80 Diffuse large B cell lymphoma of the primary central nervous system (CNS-DLBCL) accounts for 81 \leq <3% of brain tumor with a peak incidence between the 5th and 6th decade¹. The most frequent 82 (60%) localization is the supra-tentorial frontal region. Ocular lesions develop concurrently in 20% 83 of patients,^{1,2} while extra-neural dissemination is rare.^{1,3} CNS-DLBCL lesions may be single or 84 multiple, with distinct margins or poorly defined with diffuse parenchyma infiltration 1 . The neoplastic population consists of medium/large proliferating blasts, with a mature B phenotype.¹ 85 86 Most CNS-DLBCL are $BCL6⁺/IRF4⁺$ and^{1,4} approximately 80% of CNS-DLBCL are 87 BCL2⁺/cMYC⁺ (double expressors), a protein profile that confers poor clinical prognosis; double-88 hit or triple-hit high grade B-cell lymphomas, with MYC and BCL2 and/or BCL6 rearrangements, are rare.^{5,6} Despite high-dose polychemotherapy and methotrexate (MTX), the prognosis remains 90 poor, with a median overall survival (OS) of about 3 years and a median progression free survival 91 (PFS) of 12 months.⁷ At the bio-pathological level, proliferation and survival of the neoplastic 92 clones are supported by the constitutive activation of the B-cell receptor (BCR), Toll-like receptor 93 (TLR) and NF-kB signaling pathways caused by mutations of *MYD88* and/or *CD79B*.¹ Recently, 94 the landscape of recurrent genetic drivers in DLBCL has been expanded by Schmitz G et al and 95 Chapuy B et al. that uncovered genetic subtypes of DLBCL with distinct clinical characteristics, 96 providing a potential nosology for precision-medicine strategies in DLBCL. $8,9$ These subtypes 97 showed distinct outcomes after immunochemotherapy with the potential to affect the selection of 98 targeted therapies owing to their distinct oncogenic abnormalities. Wright et al revealed a high 99 prevalence of the sub-type termed MCD (based on the co-occurrence of *MYD88* p.L265P and 100 *CD79B* mutations) in primary CNS-DLBCL, defined by gene aberrations involving *MYD88*, 101 CD79B and PIM1.¹⁰ Their combined genetic, phenotypic, functional, and clinical data suggest that 102 MCD-subtype may be sensitive to BTK, PI3K, BET, BCL2, and JAK inhibitors.¹¹

103 Despite all attempts, the prognosis of CNS-DLBCL remains poor and new therapeutic approaches 104 are needed to improve patient survival. With this background, our aim was to characterize and correlate the biologic profiles CNS-DLBCL to clinical findings and explore new potential therapeutic targets.

Methods

Patients' selection and tumour samples collection. From the review of the medical records of the Department of Neurosurgery of IRCCS Istituto Delle Scienze Neurologiche, Bologna, we identified 107 pathologically confirmed cases of CNS-DLBCL diagnosed between 2005 and 2020. Of those, sixty-one cases with formalin fixed paraffin embedded (FFPE) adequate material were included in the study. The study was approved by local ethics committee (620/2020/OSS/AUSLBO) and it was performed in accordance with the Declaration of Helsinki. Thirty patients (49.2%) were male and 31 females (50.8%), with an age range between 32 and 82 years and a median age of 66 years (IQR: 57–72).

Immunohistochemistry. Paraffin-embedded sections were deparaffinized in HistoClear and dehydrated through graded ethanol. The antigen retrieval was performed in the PT-Link (Agilent Dako, Santa Clara, CA, USA, code PT100/ PT 101), for 5 min at 92°C in EnVision Flex Target retrieval solution High pH (Agilent Dako, code K 8004). Then, tissue samples were incubated at RT for 30 minutes with the following antibodies: CD20 (Agilent Dako 1:300, clone L26, code M0755), CD10 (Leica NewCastle, UK, 1:30, clone 56C6 code CD10-270-L), BCL2 (Abcam, Cambridge , UK, 1:100, clone E17, code Ab32124), BCL6 (kindly provided by Prof. Falini, indiluted, clone PG-B6p), cMYC (Epitomics, Burlingame, CA 1:80, clone Y69, code 1472-1,), IRF4 (kindly provided by Prof. Falini, 1:3, clone IRF4). Immunostaining has been completed using the Alkaline REAL Detection System Alkaline Phosphatase/RED Rabbit/Mouse (Agilent Dako, code K5005) and chromogen (Fast red), provided with the kit. Finally, slides were counterstained with Hematoxylin, mounted in Glycerine and observed and analysed by Olympus microscope. Slides were observed by 2 expert pathologists (CA, SA). Results were recorded as percentage of positive cells and graded as follows: + if >30% positive neoplastic cells, for CD10/BCL6/IRF4 according to Hans' algorithm. 130 [11] A case was defined as cMyc or BCL2 positive if $\geq 40\%$ and $\geq 50\%$ cells were stained 131 respectively. $6\overline{)}$

Genetic analysis.

Fluorescence in situ hybridization (FISH). Paraffin embedded tissue were deparaffinized and then pre-treated with two different antigen retrievals. The slides were before incubated in 1mM EDTA buffer (pH 8) in a pressure cooker (9 minutes) and after in Pepsin solution (Sigma Aldrich code 136 P7012) for 14 minutes at 38 $^{\circ}$ C¹². The probes and the samples were denatured at 80 $^{\circ}$ C for 22 min and then hybridizated at 38°C for 22 hours in a hybridizer (Agilent Dako, Santa Clara, CA USA). The experiments were conducted using the probes LSI MYC Dual Color Break Apart Rearrangement (Vysis, Abbott, Downers Grove, Illinois, USA, code 01N63-020, cut-off 3,8%), LSI BCL2 Dual Color Break Apart Rearrangement (Vysis Abbott, code 05N51-020, cut-off 4,7%) and LSI BCL6 dual color Break apart Rearrangement t(3q27) (Vysis Abbott, code 01N23-020, cut off 5,5%). Following a stringency washes (0,4XSSC/0,03%NP-40 pH 7,4 solutions at 73°C for 2 min and with 2XSSC/0,1% NP-40 pH 7 solutions at room temperature for 1 min), the slides were mounted and counterstained with DAPI I (Vysis Abbott, code 06J49-001). Microscopical analysis 145 were carried out with an Olympus BX61 microscope and images were recorded using the Cell^{\triangle F} program. *BCL6* and *BCL2* rearrangements were investigated only in cases with translocation of the *cMYC* gene.

Next Generation Sequencing. DNA from FFPE PCNSL tissue samples was purified by Quick Extract FFPE DNA Extraction Kit (Epicentre, Madison, WI) with some modifications described by 150 Ricci C et al.¹³ DNA mutations were detected using the protocol described previously¹⁴ analysing the following gene panel: *MYD88, CD79B, PIM1, GNAS, NOTCH1, KRAS, PIK3CA, EGFR, CASP8, AIP, PTEN*. In brief, after target enrichment by multiplex PCR, libraries with tagged primers were generated using Nextera adapters. Each run on MiSEQ platform (Illumina, Palo Alto, CA) was designed to allocate at least 2K reads/region aimed to have a depth of coverage of at least 2000×. FASTQ files were filtered with PHRED quality score > Q30 and length > 100 bp, and reads were mapped in a Galaxy Project environment to the hg38 human reference genome with BWA-157 MEM, GATK local realignment, HaplotypeCaller and Picard MarkDuplicates.¹⁵ The BAM files were visualized using the Integrative Genomic Viewer (IGV) to identify mutations with Variant Allele Frequency (VAF) of at least 10%; only bidirectional variant calls with more than 10 reads 160 were reported. ¹⁵ Next generation sequencing analysis was available in 46 cases, as fifteen FFPE specimens were over-fixed and not amplifiable. Protein sequence and functional information were obtained by Uniprot database.

Statistical analysis. Demographic and clinical features were described using absolute frequencies and percentages for categorical variables, mean and standard deviation for quantitative symmetrical variables or median and interquartile range (IQR) for quantitative asymmetrical variables. The associations between immunohistochemistry and radiology, between immunohistochemistry and mutated genes, and between radiology and mutated genes were investigated using the χ2 test, or Exact Fisher's test when expected cell count is less than 5. The Kaplan–Meier product limit was used to estimate the overall survival (OS) curve. Differences in survival between subgroups were assessed with the log-rank test. Statistical analysis was performed with IBM SPSS version 25.0 and 171 Stata 13. The significance level was set at $p \le 0.05$.

Results

Phenotypic profile. By immunohistochemistry, diffuse and strong expression of CD20 characterized all 61 (100%) samples. CD10, BCL6 and IRF4 were observed in 10 (16%), 51 (83.6%) and 57 (93%) of 61 cases respectively. As typical of CNS-lymphoma 10 of 61 (16.4%) cases were classified as GCB type and 51/61 (83.6%) as non-GCB type according to the Hans algorithm. BCL2 and cMYC proteins were expressed in 51 (83.6%) and 41 (67.2%) of 61 samples respectively. Double expression (DE) status for BCL2 and cMYC was detected in 36 (59%) cases while 25 (41%) were non DE. The immunohistochemical data is summarized in Table 1.

Fish analysis. FISH analysis was successful in 59 of 61 (96.7%) samples; *cMYC* rearrangements were present in 2 (3.4%) of 59 cases and in one was associated to *BCL6* translocation. This latter harboured *MYD88* mutated (p.L265P).

Next generation sequencing showed *MYD88* mutations in 24 (54.5%) of 44, all carrying p.L265P except one case with p.A260T. Both mutations are located in the Toll/Il-1 Receptor (TIR) domain of the protein.

Twenty-three cases carried a mutation in proto-oncogene with serine/threonine kinase *PIM1 (23/43, 53.5%)*, with a great variety of different mutations, mostly missense mutations, occurring at protein kinase domain, of which the p.E135K was the most common involving four cases. In 9 samples 2 or 3 different *PIM1* mutations were simultaneously present. *CD79B* missense mutations were found in 13 cases (of 43, 30.2%), located within immunoreceptor tyrosine-based activation motif (ITAM), 5 of which had p.Y196H, 2 cases p.Y196N, 2 p.Y196S, 2 p.Y196C, 1 p.Y196D, and one the rare p.E198G. Concurrent mutations were found: *MYD88*, *PIM1* and *CD79B* in 4 cases, *MYD88* and *PIM1* in 13 cases, *MYD88* and *CD79B* in 9 cases, and *CD79B* and *PIM1* in 8. Seven cases carried missense mutations in *TP53* gene (7 of 38, 18.4%). Additional mutations were

found: *NOTCH1 in* 4 of 35 (11,4%), *AIP in* 9 of 39 (23,1%), *GNAS in* 3 of 37 (8.1%) and *CASP8 in* 3 of 38 (7.9%).

PI3KCA missense mutations were found in 6 of 40 (15%); in 5 cases they produced an amino acid substitution in the catalytic domain (p.A1046V, p.D1045N, p.G1049S and p.G1049D in two cases). *PTEN and EGFR* genes mutations were found in 2 of 39 (5.1%) and 2 of 31 (6.4%) cases respectively, while p.G12D and in CIS p.G13D mutations of *KRAS* gene co-occurred in a single 201 patient (1/39). Table 2 summarizes all these sequencing data.

Clinical findings. The disease presented with a single lesion in 25 of 61 (41%) of patients and with multiple lesions in 36 of 61 (59%). On MRI, most patients showed deeply located lesions with homogeneous contrast-enhancement (36 of 50, 72%). Multiple enhancing was seen in 53%. Only one of 34 patients with DWI performed showed a high ADC value, all others presented hyperintense diffusion signal. Perfusion T2-w studies was available in 24 patients, 54% with 2 to 7 times higher value of rCBV. No significant statistical correlations were observed between biological and clinical parameters and neuroradiological features.

Gross total resection was achieved in 13 of 61 (21%), subtotal resection in 21 (35%) and biopsy in 27 (44%). Twenty-four patients, whose treatment was known, had undergone the matrix protocol MTX + cytarabine or MATRix regimen. Of the 61 patients included in the study, 57 died during the follow-up, with a median follow-up time of 3.3 months (95% CI: [2.1 - 5.0]). The longest follow-up was 69.2 months. Figure 1 shows the Kaplan-Meier survival curve. One-month, 3-month, 6-month overall survival rates were 88.5% (95% CI: [77.4% - 94.4%]), 52.5% (95% CI: [39.3% - 64.1%]), and 37.7% (95% CI: [25.7% - 49.6%]), respectively.

No statistically significant correlation was found between cell of origin GCB/non-GCB, BCL2 and cMYC expression and survival nor between DE or non-DE status and prognosis. Survival was 218 significantly longer for patients with mutated *MYD88* (8.7 months vs. 1.7 months; log-rank test = 5.43; p = 0.020) compared to those without *MYD88* mutation, and for patients with mutated *CD79B* (10.8 months vs. 2.5 months; log-rank test = 4.64; p = 0.031) compared to those without *CD79B* mutation.

Discussion

CNS-DLBCL, together with vitreoretinal and testicular DLBCL, is now grouped in a new category 224 called large B-cell lymphoma of the immune-privileged sites.¹⁶ The prognosis of CNS-DLBCL remains poor and new therapeutic approaches are needed to improve patient survival. OS is lower in our series than prior studies, this could be due to patient selection bias, which in our study had a median older age (56 vs 64) and a much higher frequency of multiple lesions (59% vs 30%).¹

Similar to previously reported, based on our data, single and double hit rearrangements of MYC 229 gene were rare 1,6 Differently from a recent report by Asano et al and Kim et al DE status for BCL2 230 and cMYC expression did not impact on prognosis in our cohort of patients.^{5,17}

Genomic studies suggest that cell proliferation and survival in CNS-DLBCL are driven by 232 deregulated TLR and BCR signaling pathways inducing constitutive NFKB activation, with a high 233 frequency of somatic non-synonymous mutations in *MYD88* and *CD79B* genes.¹⁸⁻²⁴ Therefore inhibitors of TLR/BCR signaling such as ibrutinib, blocking Bruton's tyrosine kinase (BTK), was 235 proposed as alternative therapeutic target and seem to be effective in CNS-DLBCL.²⁵⁻²⁸ We found *MYD88* mutations in 54.5% of the cases resulting the most frequently mutated gene in our series; *CD79B* mutations considered one of the hallmark of CNS-DLBCL mutational signature, were demonstrated in 30.2% of the cases. This gene encodes the Ig-beta protein of BCR multimeric complex and its activating mutations reinforce BCR signalling contributing to sensitivity to Ibrutinib. Interestingly, survival was significantly longer for patients with mutated *MYD88* and *CD79B* compared to those with wild type genes. Our findings confirm data reported by Curran et al and Zhou et al respectively, and are in contrast with two studies reporting a poor prognosis 243 associated with the MYD88 mutation. $29-32$ These conflicting results might reflect a selection bias among small study populations, given the rarity of CNS-DLBCL.

PIM1 was found to be the second most frequently mutated gene (53.5%), with a great variety of different mutations. PIM1 protein belongs to the Ser/Thr protein kinase family which is overexpressed in hematopoietic malignancies and in prostate and breast cancers where it was 248 proposed as new therapeutic target.³³⁻³⁵ Although *PIM1* mutation status was reported to impact the 249 outcome, we did not find any correlation with prognosis.³²

TP53 alterations seem to play a minor role in CNS-DLBCL. Zorofchian et al (26.7%) and other 251 authors have suggested that disruption of the p53-pathway may be associated with poor PFS . $37,38$

In our series *TP53* mutations were not significantly related to prognosis.

Notably, we detected for the first time *AIP* mutations in CNS-DLBCL (23,1% of the cases). Aryl hydrocarbon receptor-interacting protein (*AIP*) is a co-chaperone to heat shock proteins and nuclear receptors which behaves as tumor suppressor gene. In colorectal, pancreatic and gastric cancer high expression of AIP is associated with tumour development and more aggressive disease and 257 inhibiting aryl hydrocarbon receptor was proposed as potential therapeutic target.³⁹⁻⁴¹ Recently *AIP* was found to be a positive regulator of BCL6 expression in germinal centers cells, protecting BCL6 from ubiquitin-mediated proteasomal degradation, and deletion of *AIP* in B cells decrease BCL6 260 expression, reducing germinal center B cells and diminishing adaptive immune responses. ⁴² Furthermore AIP was required for optimal AKT signaling in response to BCR stimulation and seems to be highly expressed in primary DLBCL compared to healthy tissue with implications for 263 the pathobiology of this disease.⁴²

The NOTCH signalling pathway is widely involved in cellular proliferation, differentiation, and apoptosis. ⁴³We *NOTCH1* mutations in 11,4% of the samples: *NOTCH1* mutations are distinctive drivers of systemic DLBCL and were not previously described in CNS-DLBCL.⁸

GNAS and *CASP8* were mutated in 8.1% and 7.9% of the cases, respectively. *GNAS* is the most frequently mutated G-protein in human cancers and activating mutations in the gene GNAS have been found in pituitary, thyroid, pancreatic, biliary tract and intestine tumors as well as in Burkitt 270 lymphoma, systemic DLBCL and Hodgkin lymphoma.⁴⁴⁻⁴⁷ Zhou et al reported *GNA13* mutations to be associated with a shorter PFS and overall survival in primary central nervous system lymphoma 272 patients, however we did not find any correlation with prognosis.⁴⁸ We also identified somatic mutations *CASP8* for which a functional role has not been previously suspected in CNS-DLBCL, although p.R472* nonsense mutation detected in one of our samples was previously reported in 275 systemic DLBCL.⁴⁹ CASP8 encodes a member of the cysteine-aspartic acid protease (caspase) family and sequential activation of caspases plays a central role in the execution-phase of cell apoptosis. In addition, caspase-8 participates in maintenance of genomic integrity and loss of caspase-8 in B lymphocytes leads to B-cell malignancies; moreover, *CASP8* mutations were found 279 in mantle cell lymphoma.⁵⁰

PI3KCA missense mutations, previously described in several types of carcinomas,¹ were found in 6 of 40 (15%), while *PTEN* mutations were detected in 2 of 39 samples (5.1%). The PI3K/AKT/mTOR is an important pro-survival pathway which plays a pivotal role in the development of malignant tumours being often aberrantly activated in different types of cancer also including lymphomas. Zang et al recently showed that PI3K/AKT/mTOR signaling is aberrantly 285 activated in CNS-DLBCL and correlated with a poor prognosis.⁵¹ They found that p-mTOR expression was an independent risk factor in terms of PFS in patients with CNS-DLBCL. Moreover, Takashima et al found that copy number losses in the PTEN-PI3K-AKT proapoptotic pathway are associated with poor prognosis in CNS-DLBCL patients while PTEN mutation was 289 related to shorter OS in the study of Todorovic et al.^{52,53} PI3K/mTOR inhibitors has also become a potential therapeutic target in CNS-DLBCL. A phase II trial of 37 relapsed/refractory CNS-DLBCL 291 patients demonstrated that temsirolimus had a positive effect with 54% overall response rate, while lower response rate of 25% was seen in a clinical trial targeting using the pan-PI3K inhibitor 293 buparlisib.^{54,55} The use of dual pan-PI3K/mTOR inhibitor bimiralisib (PQR309) in a multicenter phase I/II trial (NCT02669511) as well as BAY80-6946 (copanlisib) in a phase Ib/II clinical trial (NCT03581942) are still under investigation in relapsed/refractory CNS-DLBCL. Moreover, 296 Inhibition of the PI3K isoforms p110 α /p110 δ or mTOR synergized with ibrutinib to induce cell death in CD79B-mutant CNS-DLBCL cells suggesting that combined inhibition of BTK and 298 PI3K/mTOR could overcome the resistance of lymphoma cells to ibrutinib.⁵⁶ So far, no PI3K mutations were described in CNS-DLBCL and our data provide a strong rational for a clinical application of inhibitors targeting the PI3K/ AKT/mTOR signalling pathway at least in a subset of patients.

Finally, mutations in the *EGFR* and *KRAS* genes were found in in 6.4% and 2.6% of the cases respectively that could be promising therapeutic targets. Although copy number alterations with

֬֒

- amplifications in RTK-RAS-MAPK signalling has been correlated to a poorer prognosis in CNS-305 DLBCL we didn't find any association with survival.
- **Conclusion**

MYD88 and *CD79B* predict a longer survival in patients affected by CNS-DLBCL. The *rearrangements* of the *MYC* gene are rare as well as double hit events involving BCL6 and/or BCL2 genes in the present series of CNS-DLBCL. COO and DE status does not affect the prognosis of the 61 CNS-DLBCL cases. Notably, we identified novel mutations that enrich the mutational landscape of CNS-DLBCL, suggests a role of PTEN-PI3K-AKT and RTK-RAS-MAPK signalling in a subset of CNS-DLBCL and that provides new potential therapeutic targets.

References

- 1. Kluin PM, et al. Primary diffuse large B-cell lymphoma of the CNS. In: Swerdlow SH, Campo E,
- Harris NL, et al. (eds). WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues.
- Lyon, France: IARC Press; 2017; 300-2.
- 2. Schlegel. Primary CNS lymphoma. *Ther Adv Neurol Disord 2*:93-104 U (2009).
- 3. Deckert M, Engert A, Brück W, Ferreri AJ, Finke J, Illerhaus G, et al. Modern concepts in the biology, diagnosis, differential diagnosis and treatmentof primary central nervous system
- lymphoma. *Leukemia* **25**:1797-807 (2011)
- 4. Marcus C, Maragkos GA, Alterman RL, Uhlmann E, Pihan G, Varma H. GCB-type is a
- favorable prognostic factor in primary CNS diffuse large B-cell lymphomas*. J Clin Neurosci* **83**: 49-55 (2021)
- 5. Kim S, Nam SJ, Kwon D, Kim H, Lee E, Kim TM, et al. MYC and BCL2 overexpression is associated with a higher class of Memorial Sloan-Kettering Cancer Center prognostic model and poor clinical outcome in primary diffuse large B cell lymphoma of the central nervous system. *BMC Cancer* **16**:363 (2016)
	-
- 6. Pina-Oviedo S, Bellamy WT, Gokden M. Analysis of primary central nervous system large B-
- cell lymphoma in the era of high-grade B-cell lymphoma: Detection of two cases with MYC and
- BCL6 rearrangements in a cohort of 12 cases. *Ann Diagn Pathol* **48**: 151610 (2020)
- 7. Abrey LE, DeAngelis LM, Yahalom J. Long-term survival in primary CNS lymphoma. *J Clin Oncol* **16**:859–63 (1998)
- 8. Schmitz R, Wright GW, Huang DW, Johnson CA, Phelan JD, Wang JQ, et al. Genetics and Pathogenesis of Diffuse Large B-Cell Lymphoma. *N Engl J Med* **378**, 1396–1407 (2018).
- 9. Chapuy B, Stewart C, Dunford AJ, Kim J, Kamburov A, Redd RA,, et al. Molecular subtypes of
- diffuse large B cell lymphoma are associated with distinct pathogenic mechanisms and outcomes.
- *Nat Med* **24**, 679–690 (2018)
- 10. Wright GW, Huang DW, Phelan JD, Coulibaly ZA, Roulland S, Young RM, et al. A
- Probabilistic Classification Tool for Genetic Subtypes of Diffuse Large B Cell Lymphoma with Therapeutic Implications. *Cancer Cell* **37**(4):551-568 (2020)
- 11. Hans CP, Weisenburger DD, Greiner TC, Gascoyne RD, Delabie J, Ott G, et al. Confirmation of the molecular classification of diffuse large B-cell lymphoma by immunohistochemistry using a
- tissue microarray. *Blood* **1**;103(1):275-82 (2004)
- 12. Ventura RA, Martin-Subero JI, Jones M, McParland J, Gesk S, Mason DY, et al. FISH analysis
- for the detection of lymphoma-associated chromosomal abnormalities in routine paraffin-embedded tissue. *J Mol Diagn* **8**(2):141-51 (2006)
- 13. Ricci C, Morandi L, Ambrosi F, Righi A, Gibertoni D, Maletta F, et al. Intron 4-5 hTERT DNA Hypermethylation in Merkel Cell Carcinoma: Frequency, Association with Other Clinico-pathological Features and Prognostic Relevance. Multicenter Study. *Endocr Pathol* **32**(3):385-395 (2021)
- 14. Gabusi A, Gissi DB, Montebugnoli L, Asioli S, Tarsitano A, Marchetti C, et al. Prognostic
- impact of intra-field heterogeneity in oral squamous cell carcinoma. *Virchows Arch* **476**(4):585-595
- (2020)
- 15. Afgan E, Baker D, van den Beek M, Blankenberg D, Bouvier D, Čech M, et al. The Galaxy platform for accessible, reproducible and collaborative biomedical analyses: 2016 update. *Nucleic Acids Res* **8**;44(W1):W3-W10 (2016)
- 16. Alaggio R, Amador C, Anagnostopoulos I, Attygalle AD, Araujo IBO, Berti E, et al Leukemia.
- The 5th edition of the World Health Organization Classification of Haematolymphoid Tumours:
- Lymphoid Neoplasms. 2022 Jul;36(7):1720-1748.
- 17. Asano K, Yamashita Y, Ono T, Natsumeda M, Beppu T, Matsuda K et al . Clinicopathological risk factors for a poor prognosis of primary central nervous system lymphoma in elderly patients in
- the Tohoku and Niigata area: a multicenter, retrospective, cohort study of the Tohoku Brain Tumor
- Study Group*. Brain Tumor Pathol* **39**(3):139-150 (2022)
- 18. Braggio E, McPhail ER, Macon W, Lopes MB, Schiff D, Law M, et al. Primary central nervous system lymphomas: a validation study of array-based comparative genomic hybridization in formalin-fixed paraffin-embedded tumor specimens. *Clin Cancer Res.***17**:4245–53 (2011)
- 19. Montesinos-Rongen, M., Schafer, E., Siebert, R. & Deckert, M. Genes regulating the B cell receptor pathway are recurrently mutated in primary central nervous system lymphoma. *Acta Neuropathol.* **124**, 905–906 (2012).
- 20. Bruno A, Boisselier B, Labreche K, Marie Y, Polivka M, Jouvet A, et al. Mutational analysis of primary central nervous system lymphoma. *Oncotarget* **5**:5065–75 (2014)
- 21. Vater I, Montesinos-Rongen M, Schlesner M, Haake A, Purschke F, Sprute R, et al. The
- mutational pattern of primary lymphoma of the central nervous system determined by whole-exome
- sequencing. *Leukemia* **29**:677–85 (2015)
- 22. Radke J, Ishaque N, Koll R, Gu Z, Schumann E, Sieverling L, et al. The genomic and transcriptional landscape of primary central nervous system lymphoma. *Nat Commun* **10**;13(1):2558 (2022)
- 23. Yamada, S., Ishida, Y., Matsuno, A, Yamazaki, K. Primary diffuse large B-cell lymphomas of central nervous system exhibit remarkably high prevalence of oncogenic MYD88 and CD79B mutations. *Leuk. Lymphoma* **56**, 2141–2145 (2015)
- 24. Nayyar N, White MD, Gill CM, Lastrapes M, Bertalan M, Kaplan A, et al. MYD88 L265P mutation and CDKN2A loss are early mutational events in primary central nervous system diffuse
- large B-cell lymphomas. *Blood Adv* **3,** 375–383 (2019)
- 25. Grommes, C., Nayak, L., Tun, H. W, Batchelor, T. T. Introduction of novel agents in the treatment of primary CNS lymphoma. *Neuro-Oncol* **21**, 306–313, (2019)
- 26. Soussain C, Choquet S, Blonski M, Leclercq D, Houillier C, Rezai K, et al. Ibrutinib monotherapy for relapse or refractory primary CNS lymphoma and primary vitreoretinal lymphoma: Final analysis of the phase II 'proof-of-concept' iLOC study by the Lymphoma study association (LYSA) and the French oculo-cerebral lymphoma (LOC) network. *Eur. J. Cancer* **117**,
- 121–130 (2019)
- 27. Lionakis MS, Dunleavy K, Roschewski M, Widemann BC, Butman JA, Schmitz R, et al. Inhibition of B cell receptor signaling by Ibrutinib in primary CNS lymphoma. *Cancer Cell* **31**, 833–843 e835 (2017)
- 28. Grommes C, Pastore A, Palaskas N, Tang SS, Campos C, Schartz D, et al. Ibrutinib unmasks critical role of Bruton tyrosine kinase in primary CNS lymphoma. *Cancer Discov* **7**, 1018–1029 (2017)
- 29. Curran OE, Poon MTC, Gilroy L, Torgersen A, Smith C, Al-Qsous W. MYD88 L265P mutation in primary central nervous system lymphoma is associated with better survival: A single-center experience. *Neurooncol Adv* **7**;3(1):vdab090 (2021)
- 30. Zhou J, Zuo M, Li L, Li F, Ke P, Zhou Y, Xu Y, et al. PIM1 and CD79B Mutation Status Impacts the Outcome of Primary Diffuse Large B-Cell Lymphoma of the CNS. *Front Oncol* **9**;12:824632 (2022)
- 31. Takano S, Hattori K, Ishikawa E, Narita Y, Iwadate Y, Yamaguchi F, et al. MyD88 Mutation in
- Elderly Predicts Poor Prognosis in Primary Central Nervous System Lymphoma: Multi-Institutional
- Analysis. *World Neurosurg* **112**:e69-e73 (2018)
- 32. Hattori K, Sakata-Yanagimoto M, Okoshi Y, Goshima Y, Yanagimoto S, Nakamoto-Matsubara
- R, et al. MYD88 (L265P) mutation is associated with an unfavorable outcome of primary central
- nervous system lymphoma*. Br J Haematol* **177**(3):492-494 (2017)
- 33. Mahata S, Sahoo PK, Pal R, Sarkar S, Mistry T, Ghosh S, et al. PIM1/STAT3 axis: a potential co-targeted therapeutic approach in triple-negative breast cancer. *Med Oncol* **15**;39(5):74 (2022)
- 34. Zhong S, Peng S, Chen Z, Chen Z, Luo JL. Choosing Kinase Inhibitors for Androgen
- Deprivation Therapy-Resistant Prostate Cancer. *Pharmaceutics* **24**;14(3):498 (2022)
- 35. Zhao Y, Aziz AUR, Zhang H, Zhang Z, Li N, Liu B. A systematic review on active sites and functions of PIM-1 protein. *Hum Cell* **35**(2):427-440 (2022)
-
- DLBCL, molecular subtypes, and outcomes: A Haematological Malignancy Research Network report. *Blood* **135**, 1759–1771 (2020).

36. Lacy SE, Barrans SL, Beer PA, Painter D, Smith AG, Roman E, et al. Targeted sequencing in

- 37. Zorofchian S, El-Achi H, Yan Y, Esquenazi Y, Ballester LY. Characterization of genomic alterations in primary central nervous system lymphomas. *J Neurooncol* **140**(3):509-517 (2018)
- 38. Munch-Petersen HD, Asmar F, Dimopoulos K, Areškevičiūtė A, Brown P, Girkov MS, et al.
- TP53 hotspot mutations are predictive of survival in primary central nervous system lymphoma
- patients treated with combination chemotherapy. *Acta Neuropathol Commun* **22**;4:40 (2016)
- 39. Solís-Fernández G, Montero-Calle A, Sánchez-Martínez M, Peláez-García A, Fernández-
- Aceñero MJ, Pallarés P, et al. Aryl-hydrocarbon receptor-interacting protein regulates tumorigenic and metastatic properties of colorectal cancer cells driving liver metastasis. *Br J Cancer*
- **126**(11):1604-1615 (2022)
- 40. Fernández-Aceñero MJ, Barderas R, Peláez García A, Martínez-Useros J, Díez-Valladares L, Pérez-Aguirre, et al. Aryl hydrocarbon receptor interacting protein (AIP) significantly influences prognosis of pancreatic carcinoma. *Ann Diagn Pathol* **53**:151742 (2021)
- 41. Díaz Del Arco C, Estrada Muñoz L, Barderas Manchado R, Peláez García A, Ortega Medina L,
- Molina Roldán E, et al. Prognostic Role of Aryl Hydrocarbon Receptor Interacting Protein (AIP)
- Immunohistochemical Expression in Patients with Resected Gastric Carcinomas. *Pathol Oncol Res*
- **26**(4):2641-2650 (2020)
- 42. Sun D, Stopka-Farooqui U, Barry S, Aksoy E, Parsonage G, Vossenkämper A, et al. Aryl
- Hydrocarbon Receptor Interacting Protein Maintains Germinal Center B Cells through Suppression
- of BCL6 Degradation. *Cell Rep* 30;27(5):1461-1471 (2019)
- 43. Artavanis-Tsakonas S, Rand MD, Lake RJ. Notch signaling: cell fate control and signal integration in development. *Science* **30**;284(5415):770-6 (1999)
- 44. O'Hayre M, Vazquez-Prado J, Kufareva I, Stawiski EW, Handel TM, Seshagiri S, et al. The
- emerging mutational landscape of G proteins and G-protein-coupled receptors in cancer. N*ature reviews Cancer* **13**(6):412–24 (2013)
- 45. Love C, Sun Z, Jima D, Li G, Zhang J, Miles R, et al. The genetic landscape of mutations in Burkitt lymphoma*. Nature genetics* **44**(12):1321–5 (2012)
- 46. Lohr JG, Stojanov P, Lawrence MS, Auclair D, Chapuy B, Sougnez C, et al. Discovery and prioritization of somatic mutations in diffuse large B-cell lymphoma (DLBCL) by whole-exome sequencing. *Proc Natl Acad Sci U S A* **6**;109(10):3879-84 (2012)
- 47. Wienand K, Chapuy B, Stewart C, Dunford AJ, Wu D, Kim J, et al. Genomic analyses of flow-
- sorted Hodgkin Reed-Sternberg cells reveal complementary mechanisms of immune evasion. *Blood*
- *Adv* **10**;3(23):4065-4080 (2019)
- 48. Zhou Y, Liu W, Xu Z, Zhu H, Xiao D, Su W, et al. Analysis of Genomic Alteration in Primary
- Central Nervous System Lymphoma and the Expression of Some Related Genes. *Neoplasia*
- **20**(10):1059-1069 (2018)
- 49. Hakem A, El Ghamrasni S, Maire G, Lemmers B, Karaskova J, Jurisicova A, et al. Caspase-8 is essential for maintaining chromosomal stability and suppressing B-cell lymphomagenesis. *Blood* **12**;119(15):3495-502 (2012)
- 50. Yang P, Zhang W, Wang J, Liu Y, An R, Jing H. Genomic landscape and prognostic analysis of mantle cell lymphoma. *Cancer Gene Ther* **25**(5-6):129-140 (2018)
- 51. Zhang X, Wu Y, Sun X, Cui Q, Bai X, Dong G, et al.The PI3K/AKT/mTOR signaling pathway
- is aberrantly activated in primary central nervous system lymphoma and correlated with a poor prognosis. *BMC Cancer* **20**;22(1):190 (2022)
- 52. Takashima Y, Sasaki Y, Hayano A, Homma J, Fukai J, Iwadate Y, et al. Target amplicon
- exome-sequencing identifies promising diagnosis and prognostic markers involved in RTK-RAS
- and PI3K-AKT signaling as central oncopathways in primary central nervous system lymphoma.
- *Oncotarget* **8**;9(44):27471-27486 (2018)
- 53. Todorovic BM, Jelicic J, Mihaljevic B, Kostic J, Stanic B, Balint B, et al. Gene mutation profles in primary difuse large B cell lymphoma of central nervous system: next generation sequencing analyses*. Int J Mol Sci* **17**(5):683 (2016)
- 54. Korfel A, Schlegel U, Herrlinger U, Dreyling M, Schmidt C, von Baumgarten L,, et al. Phase II
- trial of temsirolimus for relapsed/refractory primary CNS lymphoma. *J Clin Oncol* **34**(15):1757–
- 1763 (2016)
- 55. Grommes C, Pentsova E, Nolan C, Wolfe J, Mellinghoff IK, Deangelis L. Phase II study of single agent buparlisib in recurrent/refractory primary (PCNSL) and secondary CNS lymphoma (SCNSL). *Ann Oncol* **27**(suppl 6):335 (2016)
- 56. Grommes C, Pastore A, Palaskas N, Tang SS, Campos C, Schartz D,, et al. Ibrutinib unmasks critical role of bruton tyrosine kinase in primary CNS lymphoma. *Cancer Discov* **7**: 1018–1029 (2017)
-
-