

Stereoselective Solvolysis in the Synthesis of Dorzolamide Intermediates

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ABSTRACT: The key intermediate in the synthesis of dorzolamide, (4S,6S)-methyl-5,6-dihydro-4*H*-thieno[2,3-*b*]thiopyran-4-ol-7,7-dioxide, can be obtained in the diastereoisomerically pure form in two straightforward steps starting from diastereoisomeric mixtures of *cis/trans*-(6*S*)-6-methyl-5,6-dihydro-4*H*-thieno[2,3-*b*]thiopyran-4-yl acetate, regardless of their ratio. The reaction of crucial importance in this scheme is a remarkably stereoselective solvolysis of the acetate ester in an acetone/phosphate buffer mixture as the solvent system. Investigation of this so far unrecognized stereoselective reaction reveals that it proceeds via an S_N1-like pathway as indicated by the correlation of the solvolysis rate constants with the Y_{OTs} values of different solvent mixtures and by trapping of the reaction intermediate with sodium azide. The structure of (4*S*,6*S*)-methyl-5,6-dihydro-4*H*-thieno[2,3-*b*]thiopyran-4ol-7,7-dioxide was confirmed by single-crystal X-ray analysis.

■ INTRODUCTION

Dorzolamide, (4S,6S)-2-(aminosulfonyl)-4-(ethylamino)-5,6dihydro-6-methyl-4*H*-thieno[2,3-b]thiopyran-7,7-dioxide (1) (Scheme 1), is an inhibitor of carbonic anhydrase (CA), which displays activity in the low-nanomolar range, especially against human CA isoenzyme II.^{1,2} Aqueous solutions of its hydrochloride salt are used topically as ophthalmic drops (Trusopt), often in association with the β blocker timolol,³ for the reduction of elevated intraocular pressure (IOP) in glaucoma patients.⁴ Dorzolamide was also proved to act as an inhibitor of bacterial carbonic anhydrases such as those derived from Helicobacter pylori⁵ and Mycobacterium tuberculosis.⁶ More recently, dorzolamide was reported as an inhibitor of oseltamivir-resistant H1N1 influenza viruses with the H275Y mutation at the neuraminidase gene.⁷ These findings suggest the possibility of repositioning this already approved drug as an antibacterial or antiviral, with significant reduction of the cost and time associated with the introduction of new drugs into clinical use.^{8–10} In this context, it is worth mentioning that the inhibition of bacterial CA is emerging as an approach to target potentially drug-resistant microorgan-isms¹¹ or alternative to antibiotics.^{12–15} Besides the activity of dorzolamide itself, the functionalization of its molecular framework is presently pursued as a strategy to achieve new chemical entities with increased activity for IOP management. Indeed, in seeking improved IOP-reducing agents, dorzolamide was used as a scaffold for decoration with nitric-oxide-releasing moieties, providing compounds with promising activity in an *in vivo* IOP-lowering model.¹⁶

The synthesis of dorzolamide was first achieved in Merck's laboratories in 1989.^{17–19} Afterward, some attention was paid to increasing the overall yield of 1 by improving the preparation of the key intermediate *trans*-2 or its precursor *trans*-3 (Scheme 1).

These efforts were led by the observation that achieving diastereoisomerically enriched mixtures of *trans-2* is pivotal for the efficacy of the subsequent Ritter reaction required in the

 Received:
 June 5, 2023

 Accepted:
 July 6, 2023

 Published:
 July 21, 2023





© 2023 The Authors. Published by American Chemical Society Scheme 1. Molecular Structures of Dorzolamide 1 and Its Synthetic Intermediates^a



^aNumbering scheme of the 5,6-dihydro-4*H*-thieno[2,3-*b*]thiopyran moiety is displayed on *trans*-3.

synthetic route to 1, as outlined by Blacklock and Sohar.²⁰ Among the strategies deployed to this end, the bioreduction of the 4-ketosulfone 4 promoted by *Neurospora crassa* whole cells^{21–23} or *Pichia halophila*^{21,22} (Zeneca) allowed the preparation of *trans*-2 with a selectivity of 95% and conversions of 100%. In addition, the kinetic resolution of *rac-cis*-3 and *rac-trans*-3 by acylation and epimerization were described, in a detailed study, using lipase in organic solvents.²⁴

As a part of our studies aimed to develop a new synthetic approach to 1, we took into consideration the kinetic resolution of diastereoisomeric mixtures of esters 5 or 6 by exploiting Candida antarctica lipase B (CALB) in hydrolytic reactions. However, these attempts were unsuccessful as a parallel, uncatalyzed reaction led to diastereoisomeric mixtures of alcohols 3 largely unbalanced toward the desired transisomer with a cis/trans ratio of up to 7:93. Control experiments confirmed that the enzymatic reaction was essentially abrogated under these conditions. We found that this remarkably stereoselective process is entirely consistent with an S_N 1-like solvolytic reaction of 5, and we report here some mechanistic observations on this, so far unrecognized, stereoselective process. Moreover, we propose the oxidation of 3 (cis/trans 9:91) to the corresponding sulfone trans-2 that can be obtained diastereomerically pure by crystallization.

RESULTS AND DISCUSSION

To study the experimental conditions for the kinetic resolution of **5** via hydrolysis with the enzyme CALB (Table 1), we prepared a *cis/trans* diastereoisomeric mixture of **5** with a ratio close to 50:50. To this end, alcohol **3** was obtained in 98% yield by the reduction of the corresponding 4-ketosulfide 7 with LiAlH₄ in toluene.²⁰ The reaction yields a diasteroisomeric mixture of **3** with a *cis/trans* ratio of 95:5, in agreement with literature reports. Afterward, the *cis/trans*-**3** ratio was changed to 52:48 by a controlled acid-catalyzed epimerization according to the procedure reported by Blacklock.²⁰

The ester 5 (*cis/trans* = 52:48) was obtained by the acylation of 3 (*cis/trans* = 52:48) with acetic anhydride in the presence of excess DMAP. Preliminary kinetic resolutions of the ester 5 (*cis/trans* 52:48) at 0.1 mmol concentration were run at 25 °C using 250 U of CALB supported on an acrylic resin suspended in 5 mL of phosphate buffer (PB; 0.1 M, pH

Table 1. Experiments for the Screening^a of Solvent Systemsfor the Solvolysis of 5



no.	solvent system	conv., % ^b	dr cis/trans-3 ^b	dr cis/trans-5 ^b
1	acetone/water 80:20	6	12:88	54:46
2	acetone/water 50:50	27	14:86	56:44
3	acetone/PB (0.1 M) 50:50	68	10:90	53:47
4	acetone/PB (0.3 M) 50:50	61	9:91	54:46
5	TFE/water 40:60	100	24:76	0
6	TFE/PB (0.1 M) 40:60	100	6:94	0
7	TFE/PB (0.1 M) 40:60 ^c	99	5:95	N.D. ^d
8	TFE/PB (0.3 M) 40:60	100	6:94	0
9	NaNO ₃ (aq., 0.1 M) ^e	100	25:75	0
10	Na ₂ SO ₄ (aq., 0.1 M) ^e	100	27:73	0
11	water ^d	100	30:70	0
12	PB $(0.01 \text{ M})^d$	81	20:80	56:44
13	PB $(0.1 \text{ M})^d$	81	7:93	60:40
14	PB (0.1 M) ^{<i>c,d</i>}	37	8:92	56:44
15	PB $(0.3 \text{ M})^d$	100	6:94	0
16	acetic acid	2	N.D.	16:84
17	HFIP/H ₂ O 90:10	N.D.	degradation	degradation

^{*a*}Reaction conditions: 5 mg of 5 (*cis/trans* = 52:48); 1.1 mL of the solvent system; 25 °C. ^{*b*}Determined by integration of the appropriate ¹H NMR signals as reported in the Supporting Information. ^{*c*}Experiment run at 10 °C. ^{*d*}N.D. = not determined. ^{*e*}Reaction mixture is heterogeneous.

7.4). The reaction was monitored with time by ¹H NMR (see details in the Supporting Information), observing the complete conversion of the starting material to the alcohols that were obtained with a *cis/trans* ratio of up to 7:93, which remained constant throughout the reaction. This result is not compatible with either ester hydrolysis or the known enzymatic activity of CALB because it necessarily requires the loss of stereochemical integrity at carbon 4. In order to assess whether the uncatalyzed formation of alcohols accompanied the enzymatic reaction or was the only active process in the solution, the ester



Figure 1. (A) Comparison of the conversion of isomeric mixture 5 in different solvent systems; the conversion is expressed as the residual concentration of 5 determined at a given time; legend: solid circles: acetone/water 80:20; open circles: acetone/water 50:50; crossed circles: acetone/water 45:55. (B) Typical reaction kinetic profile for the solvolysis of 5 in acetone/water 45:55; legend: open squares: *cis*-5; open circles: *trans*-5; solid squares: *cis*-3; solid circles: *trans*-3.

5 was suspended in PB (0.1 M, pH 7.4) at 25 °C in the absence of the enzyme. Under these conditions, alcohol 3 was obtained with the same diastereoisomeric ratio observed in the attempts of kinetic resolution, confirming that the enzymatic reaction was essentially inoperative. Furthermore, in control experiments, we established that the alcohols 3 are stereo-chemically stable in PB (0.1 M, pH 7.4) at 25 °C (see Figure S1), a further indication that the reaction occurred on esters 5. Based on this experimental evidence, we put forth the hypothesis that this reaction could proceed through a solvolytic pathway with $S_{\rm N}1$ characteristics.

This assumption seemed reasonable because the structure of the reagent allows the formation of a secondary carbenium ion stabilized by conjugation with the thiophene ring and by the sulfur atom in position 7 through the thiophene ring. Control experiments performed on sulfone esters 6 (cis/trans = 50:50), which are incapable of such a stabilization, either because of the less electron-rich thiophene moiety or because of the oxidized sulfur atom at position 7, did not yield solvolysis products after the reaction at 25 °C (Figure S2). Accordingly, to validate this working hypothesis and to preliminarily analyze the effect of the solvent on the conversion and diastereoisomeric ratio of the products, we performed a small-scale screening of reaction conditions by using different solvent systems including solutions of electrolytes (Table 1). In this screening, the reactions were performed at 25 $^\circ C$ and arbitrarily stopped after 66 h. After solvent removal and work-up, the composition of the crude materials was assessed by ¹H NMR. The conversion of the reagents was determined from the integral ratio of the signals at 6.02 ppm pertaining to 4-H of 5 and at 4.89 ppm pertaining to 4-H of 3. The diastereoisomeric ratio of 5 was determined as the ratio between the integrals of the signals at 6.88 and 7.02 ppm pertaining to 2-H of cis-5 and 3-H of trans-5, respectively. The diastereoisomeric ratio of isomers 3 was determined from the integrals of the signal at 7.10 ppm pertaining to 3-H of cis-3 and by the integration of the signals between 7.08 and 7.04 ppm due to protons 3-H of trans-3 and 2-H of cis-3.25 In all of the cases, a significant amount of trans-3 in excess to the amount of trans-5 originally present in the reaction mixture could be observed. Moreover, an increase in the polarity of the

medium leads to higher conversions over 66 h (Table 1, entries 1, 2, and 5). In addition, under the conditions explored, increased conversion to trans-3 was obtained by using either PB (pH 7.4) as a cosolvent (entries 3-4 and 6, 7, 8) or as the sole solvent (entries 12-15); however, in the latter case, the reaction system was heterogeneous. The effect of phosphate concentration was briefly explored by running the reactions in 0.01 and 0.1 M PB (pH 7.4) (entries 12-14), and a monotonic increase in the trans-selectivity with respect to water was observed (entry 11); further increasing the PB concentration to 0.3 M (entry 15) had no effects. Other electrolyte solutions devoid of buffer properties such as 0.1 M $NaNO_3$ or 0.1 M Na_2SO_4 (entries 9, 10) were ineffective in increasing the selectivity toward the *trans* isomer, producing 3 with the same diastereoisomeric composition observed in water (entry 11). Upon acetolysis of 5 with a cis/trans ratio of 52:48 (entry 16), the diastereoisomeric composition of the recovered ester changed to cis/trans = 16:84, further confirming the solvolytic nature of the process under analysis.²⁶ Higher proportions of trans-3 were obtained by using a 40:60 solvent mixture of 2,2,2-trifluoromethylethanol (TFE):0.1 M PB at either 25 or 10 °C; in both conditions, the product was obtained with an identical diastereoisomeric composition: *cis/trans* ratios = 6:94 (entry 6) and 5:95 (entry 7), respectively. The limited effect of the temperature on the selectivity was also observed for the reactions carried out in 0.1 M PB at 25 or 10 °C (entries 13 and 14, respectively). Attempts to explore other fluorinated alcohols such as hexafluoro-2-propanol (HFIP) led to extensive degradation of the reagents (entry 17).²⁷

To derive a quantitative insight into the mechanism of the reaction, the solvolysis of esters **5** was performed in a series of binary solvent mixtures of different ionizing power, defined according to the Y_{OTs} scale.^{28,29} The reactions were run at 30 °C, and the kinetic profiles were obtained by ¹H NMR analysis by sampling at selected reaction times. The conversions of **5** to the corresponding alcohols become faster in solvent systems with a higher ionizing power. Figure 1A shows the overall conversion of the isomeric mixtures of esters **5** for some selected acetone:water mixtures, while a typical reaction kinetic profile is displayed in Figure 1B. The formations of *cis*-**5** and

solvent system ^b	Y _{OTs} ^c	$10^6 k_{\rm C}, {\rm s}^{-1}$	$\log k_{\rm C}^{d}$	$10^6 k_{\rm T}$, s ⁻¹	$\log k_{\mathrm{T}}^{d}$	dr cis/trans-3 ^e
acetone/H ₂ O 80:20	-0.94	0.2 ± 0.5	-6.70 ± 2.50	0.2 ± 0.3	-6.70 ± 1.50	13:87 ^f
acetone/H ₂ O 50:50	1.26	2.9 ± 0.2	-5.54 ± 0.07	3.0 ± 0.1	-5.52 ± 0.03	13:87
acetone/H ₂ O 45:55	1.56 ^g	13.9 ± 0.2	-4.86 ± 0.01	18.0 ± 0.7	-4.74 ± 0.04	12:88
acetone/H ₂ O 60:40	0.66	2.04 ± 0.1	-5.69 ± 0.05	1.75 ± 0.1	-5.76 ± 0.06	13:87
ethanol/H ₂ O 80:20	0.00	1.3 ± 0.1	-5.89 ± 0.08	0.57 ± 0.06	-6.24 ± 0.11	10:90 ^h
ethanol/H ₂ O 67:33	0.57 ^g	4.4 ± 0.3	-5.36 ± 0.07	2.76 ± 0.08	-5.56 ± 0.03	8:92
TFE/H ₂ O 60:40	2.02 ^g	123 ± 7	-3.91 ± 0.06	246 ± 8	-3.61 ± 0.03	23:77
TFE/H ₂ O 40:60	2.14 ^g	144 ± 6	-3.81 ± 0.04	221 ± 30	-3.66 ± 0.14	20:80

 ${}^{a}k_{C}$ is referred to as cis-5; k_{T} is referred to trans-5. b Solvent composition: refer to v/v percentage before mixing. c From ref 5. d Errors are calculated by the uncertainty propagation method. e At complete conversion, unless otherwise stated. f At 80% conversion. g Extrapolated from data in ref 23. h At 70% conversion.



Figure 2. (A) Log $k_{\rm C}$ as a function of $Y_{\rm OTs}$ values of the solvent systems employed. Abbreviations: A, acetone; E, ethanol; T, 2,2,2-trifluoroethanol; the regression line has $R^2 = 0.86$. (B) Log $k_{\rm T}$ as a function of $Y_{\rm OTs}$ values of the solvent systems employed; the regression line has $R^2 = 0.89$. Abbreviations of the solvent systems as in (A).

*trans-***5** are clearly first-order processes; the observed first-order rate constants for the solvolysis of *cis-***5** ($k_{\rm C}$) and *trans-***5** ($k_{\rm T}$) were calculated by the nonlinear fitting of the experimental data and are reported in Table 2.

Despite some scattering, not infrequent in these types of analyses, the logarithms of $k_{\rm C}$ and $k_{\rm T}$ yield acceptable linear correlations with the $Y_{\rm OTs}$ pertinent to the solvent mixtures used (Figure 2). The slopes of the fitting curves are 0.9 ± 0.1 for *cis*-**5** and 1.0 ± 0.1 for *trans*-**5**, in line with an S_N1 reaction.

If the hypothesis of an S_N 1-like solvolysis holds true, the final composition of the products should not depend on the composition of the starting material; indeed, when the solvolysis experiments were performed on a mixture of ester 5 with a *cis/trans* ratio of 95:5,³⁰ the composition of the alcohol 3 obtained was the same as that observed when starting from 5 with a *cis/trans* ratio equal to 52:48. The intervention of a cationic intermediate was also confirmed by running the solvolysis of 5 in the presence of 50 molar equiv of sodium azide in acetone/water (45:55) (Table 3).

As in the preceding cases, the reaction kinetics was monitored by 1 H NMR analysis of aliquots of the reaction mixture withdrawn at selected reaction times; a typical kinetic profile is reported in Figure 3A. Under the reaction conditions employed, we observed the almost exclusive formation of azides 8, while only traces of alcohol 3 were detected.

A comparison of the ¹H NMR spectra of mixtures of 3, 5, and azides 8 is reported in Figure S3. Azides 8 were obtained Table 3. First-Order Rate Constants for the Consumption of Esters 5 in the Presence and Absence of 50 Equiv of NaN_3 in Acetone/Water $(45:55)^{a}$

QAc		N ₃	N ₃	
	NaN ₃ (50 eq)		+ SSS	
s	Acetone: H ₂ O 45:55 30 °C	s		
5 , <i>cis:trans</i> =52:4	8	cis -8	trans- 8	
NaN ₃ (equiv)	$10^6 k_{\rm C}, {\rm s}^{-1}$	$10^6 k_{\rm T}$, s ⁻¹	dr <i>cis/trans</i> -8 ^b	
0	12.7 ± 0.6	16.3 ± 0.8	N/A	
0	13.9 ± 0.2	18.0 ± 0.7	N/A	
50	13.9 ± 0.4	19.5 ± 0.3	36:64	
50	13.7 ± 1.8	19.8 ± 1.5	36:64	
$^{a}k_{C}$ refers to cis-	5; $k_{\rm T}$ refers to trans-	5. ^b Determined	by ¹ H NMR.	

with a *cis/trans* ratio of 36:64, closer to the original diastereoisomeric ratio of the reactants and to the *cis/trans* ratio of products 3 observed for the solvolyses in the absence of azides. The chemical nature and the stereochemistry of the azides were confirmed by mass spectrometry and NOE experiments (see Supporting Information page S15 and Figures S3 and S40).

In solvolysis experiments performed in the presence of NaN₃, the first-order rate constants (average of the two last entries in Table 3) for the consumption of *cis*-**5** and *trans*-**5** were $(13.8 \pm 0.1) \times 10^{-6}$ and $(19.7 \pm 0.2) \times 10^{-6}$ s⁻¹,



Figure 3. (A) Typical reaction kinetic profile for the solvolysis of 5 in acetone/ H_2O (45:55) in the presence of 50 equiv of NaN₃. Legend: open squares, *cis*-5; open circles, *trans*-5; solid squares, *cis*-8; solid circles, *trans*-8. (B) Formation of products 3 and 8 in the reactions run in acetone/ H_2O (45:55) in the absence and presence of NaN₃, respectively. Legend: solid circles, sum of *cis* and *trans*-3 for the solvolysis run in the absence of NaN₃; open circles and open triangles: sum of the products *cis*-8 and *trans*-8 obtained in the presence of 50 equiv of NaN₃ in the first run and second run, respectively.





^aThe thiophene ring has been omitted for clarity in the side view structures of *cis*-5 and *trans*-5.

respectively. These values are in keeping with the rate constants observed in the same solvent system but in the absence of added azide anions (Table 3). The overall rate of formation of the azides 8 was the same as that observed for the overall formation of alcohols 3 in the same solvent system without the addition of NaN₃ (Figure 3B). This confirms that the azide anion does not participate in the rate-determining step of the reaction, but rather, it likely traps the cationic species formed from the starting material. Changing the solvent system to acetone:PB (45:55) had no influence on the diastereoisomeric composition of azide 8 (Figure S4). Hence, in this case, PB did not shift the formation of the products toward the *trans* isomer of the azide 8 at variance with that

observed in the solvolysis experiments. All of these data may be accounted for by an $S_N 1$ mechanism in which the first ionization products are in equilibrium with each other, as summarized in Scheme 2. Ionization, with the initial formation of two diastereoisomeric ion pairs Ia and IIa,³¹ is likely to occur on the half chair conformations of *cis*-5 and *trans*-5 with the leaving group in the pseudo-axial position because, only in this case, a favorable overlap between the developing, vacant, p orbital, and the π system of the thiophene ring is possible.^{32,33}

Equilibration of the two diastereoisomeric ion pairs via migration of the leaving group from one face of the carbenium ion to another is unlikely. Indeed, this is a relatively slow process compared to the dissociation of the ion pair, as

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Scheme 3. Implementation of the Solvolysis of 5 in the Synthesis of Trans-2



Figure 4. Stick representations of the structures of the two independent molecules of trans-2 from crystallographic data.

reported by Mayr for the solvolysis of 1,3-diarylallyl 4nitrobenzoates,³⁴ and repeatedly shown to be undetectable in the S_N1 solvolysis reactions of 5-methyl-2-cyclohexenyl acid phthalates,³⁵ 4-nitrobenzoates,^{36,37} and chlorides³⁸ studied by Goering. It is worth emphasizing here that the substrates studied by Goering are structurally similar to esters 5. In addition, migration of the leaving group from one face of the carbenium ion to the other could have led to the geometric isomerization of cis-5 to trans-5 and vice versa, a process that we did not observe. Hence, most likely, each of the initial products of heterolysis evolves toward the reasonably free carbenium ion Ib or IIb. These carbenium ions are conformers that can easily interconvert into one another by the flipping of the 6-C from one face of the plane to another. It is reasonable that an incoming nucleophile acting on Ib or IIb would form, preferentially, a pseudo-axial bond, as suggested by Goering, and this will give the products the same stereochemistry as the starting materials.³³ However, the attack of nucleophiles on the re face of **Ib** (upper panel in Scheme 2) will be hindered by the presence of the methyl group at 6-C, while an attack on the si face (lower panel in Scheme 2) will not form a pseudo-axial bond; both of these pathways are unfavorable. In contrast, an attack on the *si* face of **IIb** is the least hindered approach of the nucleophile and produces a pseudo-axial bond, resulting in the preferential formation of trans products, as observed experimentally. When water is used as a nucleophile, the reaction proceeds with high selectivity; instead, in the case of the azide ion, which reacts with the cationic species with a rate constant close to the diffusion limit,³⁹ a lower selectivity is obtained. This is in line with an equilibration of the intermediates Ib and IIb, which remains likely unattained in the case of the azide nucleophile.

The role of phosphate buffer in this reaction is probably to trap the equivalence of acid formed upon solvolysis, as can be inferred from an inspection of the data in Table 1. Indeed, in the absence of phosphate buffer, the diastereoisomeric compositions of the alcohols 3 obtained upon solvolysis in the various solvent mixtures are very similar, and, on average, the *cis/trans* ratio is 22:78 (entries 1, 2, 5, 9, 10, and 11 in Table 1). This value is very close to the equilibrium *cis/trans* ratio of 24:76 obtained in the acid-catalyzed epimerization of *cis/trans*-3 by Blacklock and Sohar.²⁰ In the reactions where

acid is not formed, such as in the formation of the azides 8, the presence of PB does not produce any increase in the *cis/trans* ratio, vide supra.

To assess the viability of this stereoselective process in the synthesis of *trans*-2 (Scheme 3), the reaction was performed on 0.50 g of 5 using acetone/PB (45:55) as the solvent system at a 20 mM concentration of 5; the alcohol 3 (*cis/trans* 9:91) was obtained in 93% isolated yield.

Oxidation of 3 in the presence of hydrogen peroxide and Na₂WO₄·2H₂O in methanol afforded hydroxysulfone 2 (cis/ trans 9:91) quantitatively. Slow crystallization by vapor diffusion of *n*-hexane into an ethyl acetate solution of the crude trans-2 afforded the spectroscopically pure compound in 89% yield over these last two steps based on the amount of trans-2 present in the starting material before crystallization. This represents a significant improvement over the procedure developed by Blacklock and Sohar,²⁰ in which 3 could be obtained with a cis/trans ratio of 24:76 and, upon oxidation, afforded 2 with the same cis/trans ratio. In addition, the reactions of Scheme 2 are straightforward and much simpler with respect to the bioreduction of ketone 4 by N. crassa whole cells.²¹ Most notably, the concentration of 4 described in the Zeneca process $(30 \text{ mM})^{21}$ is similar to that employed in the solvolytic process described here, which also has the benefit of having a lower impact from the point of view of waste management.

The structure of *trans*-**2** was confirmed by single-crystal Xray analysis. Indeed, the crystals of *trans*-**2** (see the Supporting Information for further details) were of suitable quality for structure determination. *Trans*-**2** crystallizes in the orthorhombic form in the chiral space group $P2_12_12_1$ with cell parameters a = 7.8834(2) Å, b = 9.2510(3) Å, c = 26.4745(9)Å, and volume = 1930 Å³. The asymmetric unit consists of two independent molecules (Figures 4 and S5). After gently grinding the bulk material, the X-ray powder diffraction data of *trans*-**2** (Figure S6) were collected and compared to the expected pattern calculated for *trans*-**2**, on the basis of singlecrystal data (Figure S6). The structures of the two molecules of *trans*-**2** displayed in Figure 4 enable one to appreciate the planarity of 4-C and 7-S, substituents of the thiophene ring, with the 5-membered ring plane.

CONCLUSIONS

Given the continuing interest in dorzolamide as a therapeutic agent itself or as a scaffold for the development of new chemical entities targeting carbonic anhydrase, either human or bacterial, improved synthetic routes to this species are highly desirable. 40 In this paper, we described the preparation of diastereoisomerically pure (4S,6S)-methyl-5,6-dihydro-4Hthieno[2,3-b]thiopyran-4-ol-7,7-dioxide, trans-2, a key intermediate in the synthesis of dorzolamide, by a straightforward and cost-effective route exploiting previously unrecognized and remarkably stereoselective solvolysis of esters 5 to trans-3. By this reaction, trans-3 was obtained with a diastereomeric ratio of 91:9 starting from 5, regardless of the diasteroisomeric ratio of the reagent. It was successfully demonstrated that the reaction proceeds via an S_N1-like mechanism through a detailed kinetic analysis and trapping of the cationic intermediate by the very efficient nucleophilic azide anion. Oxidation of diastereoisomerically enriched trans-3 affords trans-2, which, upon crystallization, is obtained in the diastereoisomerically pure form in 87% yield starting from diastereoisomeric mixtures of 3 largely unbalanced toward the cis isomer. The yield and stereoselectivity allowed by the process described are comparable, if not superior, to some of the methods previously described for the synthesis of dorzolamide.

MATERIALS AND METHODS

All commercially available reagents and solvents were purchased from Aldrich and Fluka and used as received, with the exception of enantiomerically pure ketosulfone 4 and ketosulfide 7 provided by ZaCh system S.p.A.; anhydrous solvents were obtained by distilling commercial solvents over sodium-benzophenone ketyl under an argon atmosphere. The chlorinated solvents (CH2Cl2, CHCl3, and CDCl3) were treated with K₂CO₃ for at least 24 h before use. Purifications by flash column chromatography were performed using Merck silica gel 60 (230 ÷ 400 mesh). TLC analyses were performed on Merck TLC silica gel 60 F254 plates. ¹H NMR, ¹³C NMR, H-H COSY, and HSQC spectra were recorded in CDCl₃ at 298 K on a Varian 500 MHz spectrometer (operating at 500 MHz for proton and at 125 MHz for carbon), ¹H NMR spectra are referenced to signals of the residual nondeuterated solvent. Coupling constants are reported in Hz, and the multiplicity is described as follows: s (singlet), d (doublet), t (triplet), q (quartet), qt (quintet), m (multiplet), and br (broad signal). The melting points were determined by using a Buchi SMP-20 apparatus. Electrospray ionization (ESI) mass analyses were performed on a Perkin Elmer APII at 5600 eV by Dr. Fabio Hollan of the Department of Chemical and Pharmaceutical Sciences of the University of Trieste.

Detailed synthetic methods and characterization data are provided in the Supporting Information.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsomega.3c03959.

Preparation procedures of compounds 2-6 and characterization data, kinetic analyses, and solid-state characterization of *trans-2*; NMR spectra (PDF)

trans-2 data (CIF)

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Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

A.M. is grateful for the continuous support from the ZaCh System received during his Ph.D.

REFERENCES

 Supuran, C. T. Structure-based drug discovery of carbonic anhydrase inhibitors. *J. Enzyme Inhib. Med. Chem.* 2012, 27, 759–772.
 Sugrue, M. F.; Mallorga, P.; Schwam, H.; Baldwin, J. J.; Ponticello, G. S. A comparison of L-671,152 and MK-927, two topically effective ocular hypotensive carbonic anhydrase inhibitors, in experimental animals. *Curr. Eye Res.* 1990, *9*, 607–615.

(3) Supuran, C. T. Carbonic anhydrase inhibitors. *Bioorg. Med. Chem. Lett.* 2010, 20, 3467–3474.

(4) Supuran, C. T. Carbonic anhydrases: novel therapeutic applications for inhibitors and activators. *Nat. Rev. Drug Discovery* **2008**, *7*, 168–181.

(5) Nishimori, I.; Minakuchi, T. T.; Morimoto, K.; Sano, S.; Onishi, S.; Takeuchi, H.; Vullo, D.; Scozzafava, A.; Supuran, C. T. Carbonic Anhydrase Inhibitors: DNA Cloning and Inhibition Studies of the α -Carbonic Anhydrase from Helicobacter pylori, A New Target for Developing Sulfonamide and Sulfamate Gastric Drugs. *J. Med. Chem.* **2006**, *49*, 2117–2126.

(6) Carta, F.; Maresca, A.; Covarrubias, A. S.; Mowbray, S. L.; T Jones, A.; Supuran, C. T. Carbonic anhydrase inhibitors. Characterization and inhibition studies of the most active β -carbonic anhydrase from Mycobacterium tuberculosis, Rv3588c. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 6649–6654. (7) Bao, J.; Marathe, B.; Govorkova, E. A.; Zheng, J. J. Asymmetric Induction by a Nitrogen $^{14}N/^{15}N$ Isotopomer in Conjunction with Asymmetric Autocatalysis. *Angew. Chem., Int. Ed.* **2016**, *55*, 3438–3441.

(8) Chong, C. R.; Sullivan, D. J., Jr. New uses for old drugs. *Nature* **2007**, *448*, 645–646.

(9) Aronson, J. K. Old drugs – new uses. Br. J. Clin. Pharmacol. 2007, 64, 563–565.

(10) Würth, R.; Thellung, S.; Bajetto, A.; Mazzanti, M.; Florio, T.; Barbieri, F. Drug-repositioning opportunities for cancer therapy: novel molecular targets for known compounds. *Drug Discovery Today* **2016**, *21*, 190–199.

(11) Capasso, C.; Supuran, C. T. Inhibition of Bacterial Carbonic Anhydrases as a Novel Approach to Escape Drug Resistance. *Curr. Top. Med. Chem.* **2017**, *17*, 1237–1248.

(12) Supuran, C. T. Bacterial carbonic anhydrases as drug targets: toward novel antibiotics? *Front. Pharmacol.* **2011**, *2*, 34.

(13) Supuran, C. T.; Capasso, C. Carbonic Anhydrase from Porphyromonas Gingivalis as a Drug Target. *Pathogens* **2017**, *6*, 30. (14) Köhler, S.; Ouahrani-Bettache, S.; Winum, J.-Y. Brucella suis carbonic anhydrases and their inhibitors: Towards alternative antibiotics? *J. Enzyme Inhib. Med. Chem.* **2017**, *32*, 683–687.

(15) Supuran, C. T.; Capasso, C. An Overview of the Bacterial Carbonic Anhydrases. *Metabolites* **2017**, *7*, 56.

(16) Steele, R. M.; Benedini, F.; Biondi, S.; Borghi, V.; Carzaniga, L.; Impagnatiello, F.; Miglietta, D.; Chong, W. K. M.; Rajapakse, R.; Cecchi, A.; Temperini, C.; Supuran, C. T. Nitric oxide-donating carbonic anhydrase inhibitors for the treatment of open-angle glaucoma. *Bioorg. Med. Chem. Lett.* **2009**, *19*, 6565–6570.

(17) Baldwin, J. J.; Ponticello, G. S.; Christy, M. E. Thieno Thiopyran Sulfonamide Derivatives, Pharmaceutical Compositions and Use. U.S. Patent US4797413, 1989.

(18) Blacklock, T. J.; Grabowsky, E. J. J.; Sohar, P. (S)-Alkyl 3-(thien-2-ylthio)butyrate and Analogs and Synthesis Thereof. U.S. Patent US4968814, 1990.

(19) Blacklock, T. J.; Shinkai, I. Synthesis of (S)-3-(thien-2-ylthio)butyric Acid Analogs. U.S. Patent US4968815, 1990.

(20) Blacklock, T. J.; Sohar, P.; Butcher, J. W.; Lamanec, T.; Grabowski, E. J. J. An Enantioselective Synthesis of the Topically-Active Carbonic Anhydrase Inhibitor MK-0507: 5,6-Dihydro-(S)-4-(ethylamino)-(S)-6-methyl-4H-thieno[2,3-b]thiopyran-2-sulfonamide 7,7-Dioxide Hydrochloride. J. Org. Chem. **1993**, 58, 1672–1676.

(21) Holt, R. A.; Rigby, S. R. Process for Microbial Reduction Producing 4(S)-Hydroxy-6(S)methyl-tHienopyran Derivatives. U.S. Patent US5580764, 1996.

(22) Chirality in Industry II: Developments in the Commercial Manufacture and Applications of Optically Active Compounds; Collins, A. N.; Sheldrake, G. N.; Crosby, J., Eds.; Wiley: New York, 1997.

(23) Goldberg, K.; Schroer, K.; Lütz, S.; Liese, A. Biocatalytic ketone reduction—a powerful tool for the production of chiral alcohols—part II: whole-cell reductions. *Appl. Microbiol. Biotechnol.* **2007**, *76*, 249–255.

(24) Turcu, M. C.; Rantapaju, M.; Kanerva, L. T. Applying Lipase Catalysis to Access the Enantiomers of Dorzolamide Intermediates. *Eur. J. Org. Chem.* **2009**, 2009, 5594–5600.

(25) This group of signals account for the 3-H protons of *cis*-5 and *trans*-3 and 2-H of *cis*-3. The contribution by 3-H *trans*-3 was obtained by subtracting from the overall integral the values of the integrals of 3-H for *cis*-3 and the value of the integral of 2-H for *cis*-5.

(26) Little hydrolysis to alcohols *cis/trans-3*, was observed and quantified as the 2% by ¹H NMR

(27) Decomposition was evident in the ¹H NMR spectrum of the crude product; the reason for this decomposition was not further analyzed.

(28) Schadt, F. L.; Bentley, T. W.; von R Schleyer, P. The $S_N 2-S_N 1$ spectrum. 2. Quantitative treatments of nucleophilic solvent assistance. A scale of solvent nucleophilicities. *J. Am. Chem. Soc.* **1976**, *98*, 7667–7674.

(29) Bentley, T. W.; Llewellyn, G. Progress in Physical Organic Chemistry; John Wiley & Sons, Inc., 1990; Vol. 17, p 121.

(30) Prepared by acylation of the product obtained after the reduction of the 4-keto sulfide 6 without any epimerization steps.

(31) Winstein, S.; Robinson, G. C. Salt Effects and Ion Pairs in Solvolysis and Related Reactions. IX. The threo-3-*p*-Anisyl-2-butyl System. *J. Am. Chem. Soc.* **1958**, *80*, 169–181.

(32) Corey, E. J.; Sneen, R. A. Stereoelectronic Control in Enolization-Ketonization Reactions. J. Am. Chem. Soc. **1956**, 78, 6269–6278.

(33) Goering, H. L.; Josephson, R. R. Stereochemistry of Allylic Rearrangements. XII. Oxygen Exchange Associated with the Acidcatalyzed Rearrangement of *cis*- and *trans*-5-Methyl-2-cyclohexenol. *J. Am. Chem. Soc.* **1962**, *84*, 2779–2785.

(34) Troshin, K.; Mayr, H. Ion Pair Dynamics: Solvolyses of Chiral 1,3-Diarylallyl Carboxylates as a Case Study. J. Am. Chem. Soc. 2013, 135, 252–265.

(35) Goering, H. L.; Silversmith, E. F. Stereochemistry of Allylic Rearrangements. III. The Solvolysis of cis- and trans-5-Methyl-2-cyclohexenyl Acid Phthalate in Aqueous Acetone. *J. Am. Chem. Soc.* **1955**, 77, 1129–1133.

(36) Goering, H. L.; Silversmith, E. F. Stereochemistry of Allylic Rearrangements. VII. The Acid-catalyzed Hydrolysis of *cis* and *trans*-5-Methyl-2-cyclohexenyl *p*-Nitrobenzoate in Aqueous Acetone. *J. Am. Chem. Soc.* **1955**, 77, 6249–6253.

(37) Goering, H. L.; Takahashi Doi, J. Stereochemistry of Allylic Rearrangements. XI. The Isomeric Rearrangement of *trans*-5-Methyl-2-cyclohexenyl *p*-Nitrobenzoate-carbonyl-O¹⁸ in Aqueous Acetone. *J. Am. Chem. Soc.* **1960**, *82*, 5850–5854.

(38) Goering, H. L.; Nevitt, T. D.; Silversmith, E. F. Stereochemistry of Allylic Rearrangements. VI. The Ethanolysis and Acetolysis of cisand trans-5-Methyl-2-cyclohexenyl Chloride. J. Am. Chem. Soc. **1955**, 77, 5026–5032.

(39) Richard, J. P.; Yeary, P. E. Stereochemistry of Allylic Rearrangements. VI. The Ethanolysis and Acetolysis of *cis*- and *trans*-5-Methyl-2-cyclohexenyl Chloride. *J. Am. Chem. Soc.* **1993**, *115*, 1739–1744.

(40) Fei, W.; Xu, P.; Hou, J.; Yao, W. Phosphine catalyzed [3+2] cyclization/Michael addition of allenoate with CS₂ to form 2-thineyl vinyl sulfide. *Chem. Commun.* **2020**, *56*, 11669–11672.

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