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Field Competitiveness of *Aedes albopictus* (Diptera: Culicidae) Irradiated Males in Pilot Sterile Insect Technique Trials in Northern Italy

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1 **Field competitiveness of *Aedes albopictus* [Diptera: Culicidae] irradiated males in pilot Sterile**
2 **Insect Technique trials in Northern Italy**

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11

12 **Abstract**

13 Vector-borne diseases account for 17% of infectious diseases, leading to more than one million
14 deaths each year. Mosquitoes are responsible for 90% of the casualties and alternative control
15 methods to insecticides are urgently needed, especially against *Aedes* vectors. *Aedes albopictus* is
16 a particularly important species, causing major public health problems because it is a vector of
17 several arboviruses and has a strong invasive behaviour. Various genetic control methods have
18 been proposed to be integrated into the management strategies of *Aedes* species, among which
19 the sterile insect technique (SIT), which proved efficient against various insect pests and vectors.
20 However, the ability of released irradiated sterile male mosquitoes to compete with their wild
21 counterparts and induce sterility in wild females, which is critical to the success of this strategy,
22 remained poorly defined. Here, we assessed the field competitiveness of *Ae. albopictus* irradiated
23 male using data from six release trials implemented in Northern Italy for three years. Sterile males
24 were capable of inducing a good level of sterility in the wild female population, however with high

25 variability in time and space. The field competitiveness of the released males was strongly
26 negatively correlated with the ratio of sterile to wild males. This should be taken into
27 consideration when designing future programmes to suppress field populations of *Aedes*
28 mosquitoes.

29 **Keywords**

30 Gamma-ray, suppression, competitiveness, genetic, control.

31 **Introduction**

32 In the recent document on the global vector control response 2017-2030, the World Health
33 Organization (WHO) pointed out the urgent need for alternative mosquito control methods,
34 particularly against *Aedes* vectors (WHO 2017). The main reason behind this need is that the
35 integrated control of these vectors as proposed until now is challenging in many countries and in
36 different climatic conditions without guarantying satisfactory levels of population reduction
37 (Reiter 2016). Moreover, the EU Biocide directive is progressively restricting many insecticides in
38 Europe, reducing *de facto* the vector control options and increasing the probability for the
39 development of resistance against the remaining insecticides (Grigoraki et al. 2017, Pichler et al.
40 2018). New techniques to control urban mosquitoes are therefore under development and field
41 evaluation, such as genetic control strategies targeting the reproductive capacity of disease
42 transmitting mosquitoes (Flores and O'Neill 2018, McGraw and O'Neill 2013). Amongst those, the
43 sterile insect technique (SIT) shows great promise but requires the production of large quantities
44 of mosquitoes in adequate facilities. The male mosquitoes are sterilized using ionizing radiation
45 that generates random dominant lethal mutations in the germinal cells. When released in large
46 numbers to outcompete their wild counterparts, sterile males will mate with wild females and
47 sterility is induced due to embryonic arrest. There will be no offspring, thus reducing the
48 population growth rate in the next generation (Dyck et al. 2005). This technique has been used

49 very successfully against various agricultural pests (Wyss 2006, Enkerlin et al. 2015) and vectors
50 (Dicko et al. 2014, Vreysen et al. 2000), and has been under development for several years against
51 disease transmitting mosquitoes with major progresses reported recently (Lees et al. 2020). Thirty
52 four pilot SIT trials against mosquitoes are reported as presently ongoing worldwide (Bouyer et al.
53 2020a).

54 *Aedes albopictus* is a major public health concern as it is a very good vector of several arboviruses
55 such as dengue, chikungunya and Zika (Mitchell 1995, Wong et al. 2013). The potential epidemiologic
56 consequences of adaptation to *Ae. albopictus* mosquitoes are well-documented for chikungunya
57 virus, for which some mutations of the virus increased vector competence (Tsetsarkin et al. 2007).

58 The epidemiological risk related to *Ae. albopictus* has become a reality in temperate Europe,
59 where a number of disease outbreaks have occurred in recent years (Gossner et al. 2018).

60 Feasibility studies on the use of the SIT for targeting the invasive *Ae. albopictus* mosquito were
61 started in Italy in 2000 (Bellini et al. 2007), including several release field trials with irradiated
62 males that produced large data sets (Bellini et al. 2013a).

63

64 **Materials and Methods**

65 **Study area**

66 Between 2008-2018, eight release trials with irradiated *Aedes albopictus* males were carried out in
67 several localities in Northern Italy to test their efficacy in population suppression. The selected
68 urban localities were representative of the urban conditions in Northern Italy. They were also well-
69 isolated from other urban areas by agricultural land and with size adequate to fit the local sterile
70 male production capacity.

71 The main characteristics of these trials are presented in the Tab. 1.

72

73

74 Tab. 1 Main descriptive data from the eight SIT field release trials on *Aedes albopictus*

Locality and release year	Sterile male strain	*Larval Diet	Dose GY	Stage released	Release period (from-to)	N. release station/ha	N. releases	Release area (ha)
Boschi 2008	Rimini F26	CAA	30/40	pupae	Apr 29-Oct 02	0.62	29	16
Budrio 2008	Pinerolo F25	CAA	40	pupae	Apr 30-Aug 08	0.70	18	17
Boschi 2009	Rimini F35	CAA	30	pupae	May 14-Sep 10	0.62	14	16
Caselline 2009	Pinerolo F35	CAA	30	pupae	May 21-Sep 25	0.61	19	18
Boschi 2010	Rimini F41	IAEA	30	pupae	May 14-Sep 14	0.25	19	16
Gherghenzano 2012	RER F7	IAEA-BY	30	pupae & adults	May 15-Sept 25	1.00	20	10
Caselle 2018	Rimini F68-70	IAEA-BY	35	adults	June 15-Aug 31	1.00	14	16
Guisa Pepoli 2018	Rimini F68-70	IAEA-BY	35	adults	June 15-Aug 31	1.00	12	7

75 *CAA diet—larval diet consisting of 80% Friskies dry adult cat food, 14% brewer's yeast, and 6% Tetramin fish food.

76 IAEA diet—larval diet consisting of 50% bovine liver powder, 50% tuna meal, and 0.4% w/v of Vitamin Mix.

77 IAEA-BY diet— larval diet consisting of 50% bovine liver powder, 36% tuna meal, 14% brewer's yeast and 0.2% w/v of Vitamin Mix.

78

79 **Mass rearing and strains**

80 Different mosquito strains and generations were used in the pilot trials, but in all cases, colonies
81 were started from eggs collected in Northern Italy (Tab. 1). The mosquitoes were reared under
82 standard holding conditions: 27±2°C, 85% RH, and a photoperiod of 14:10 (L:D) h, as described in
83 Bellini et al. (2013a).

84

85 **Male sorting, sterilization and release protocol**

86 Separation of the sexes was carried out on the pupae in water by means of metal sieves with a
87 mesh size of 1,400 µm. The male insects were sterilized by exposing 24-36 h old male pupae to 30-
88 40 Gy in an IBL 437 Cobalt irradiator (CIS Bio International, Bagnols-sur-Ceze, France) (Bellini et al.
89 2013a). The male pupae were released 1-2 h after irradiation into plastic containers positioned on
90 the ground in permanent shaded sites. In Gherghenzano, each batch of irradiated pupae was

91 divided into two and one half was released as pupae; the other half was taken to the laboratory
92 and emerged adults were released three days later. In Caselle and Guisa Pepoli, adults were
93 ground released in fixed stations distanced about 100 m from each other.

94

95 **Field data collection**

96 In the period from 2008-2012, ovitraps CAA7 consisting of black plastic pots of 400 ml (upper
97 diameter 8 cm, lower diameter 6 cm) holding about 250 ml dechlorinated water and one 12.5 x
98 2.5 cm Masonite strip were used for egg oviposition. In 2018, ovitraps CAA14GR consisting of
99 black plastic cylindrical pots of 1,400 ml holding 800 ml of dechlorinated water, with a grill screen
100 to prevent access to the animals and a strip of masonite (15x2.5 cm) as egg deposition substrate.
101 The traps were deployed in the release as well as in the control areas and checked weekly (Tab. 2).
102 Eggs were counted under a stereomicroscope and hatched using standard procedures (Bellini et al.
103 2007) to check for their fertility rate (% egg hatch).

104

105 Tab. 2 Number of ovitraps used in the release trials

Release locality and year	No. of ovitraps in the release area	No. of ovitraps in the control area
Boschi 2008-'09	15	10
Budrio 2008	15	10
Caselline 2009	15	10
Boschi 2010	15	15
Gherghenzano 2012	10	10
Caselle 2018	14	7
Guisa Pepoli 2018	7	7

106

107

108 **Statistical analysis**

109 The percentage of egg-induced sterility (S) was calculated in relation to the number of hatched
110 eggs in the control area using Abbott's equation:

111

$$S = \{1 - [PS / PW]\}$$

112 where PS and PW are the percentages of hatched egg in the release and in the control area,
113 respectively.

114 The percentage of decrease of egg collection (D) in the release area in relation to the control area
115 was calculated by the following equation:

$$116 \quad D = (E_{\text{SIT}} - E_{\text{Control}}) / E_{\text{Control}}$$

117 where E_{SIT} and E_{Control} are the mean number of eggs per ovitrap per week in the release and in the
118 control area, respectively (see also supplementary data).

119 The t-test for dependent samples was performed to compare the results obtained by the methods
120 of estimation of the sterile/wild ratio with the observed data.

121 The linear regression analysis was used to determine the relationship between the adult caught by
122 HLC and the weekly number of eggs collected by ovitraps.

123

124 **Estimation of wild and sterile male densities**

125 The estimation of wild and sterile male densities required to calculate the field competitiveness
126 was conducted with three methods.

127 *Method 1* – Wild male population density was estimated based on the relation between the mean
128 number of eggs collected by ovitraps CAA14GR and the male density estimated in three urbanized
129 areas in Bologna city (Italy) in 2011. In the three areas (total 841 hectares), all breeding sites
130 present in 610 premises (equaling 9.82% of the total premises present in the area) were sampled
131 and 15 ovitraps were activated. Each urban area was divided into 13-16 zones and 15 premises per
132 zone were randomly selected for thorough inspection. During the inspections, all *Ae. albopictus* L4
133 larvae and pupae were collected in all breeding sites, counted and classified. The male density was
134 estimated based on the number of sampled L4 larvae and pupae using the model developed by
135 Vallorani et al. (2013) and considering a sex ratio (F:M) of 1.08 (0.6 SD) which was calculated in a

136 control area during a mark-release-recapture (MRR) study conducted in 2018 (supplementary
137 data). The relationship between the wild male/ha (M_w) and the number of eggs/ovitrap/week
138 (E_{CAA}) is given by:

$$139 \quad M_w = 1.61 (\pm) E_{CAA14GR} = 3.45 (\pm) E_{CAA7}$$

140 where $E_{CAA14GR}$ and E_{CAA7} are the number of eggs per model ovitrap CAA14GR (biweekly collection)
141 and CAA7 (weekly collection), respectively (the rate $E_{CAA14GR}/E_{CAA7} = 2.14 (\pm 0.31 \text{ SD})$) (Carrieri et al.
142 2017).

143 The daily sterile male population density was estimated, taking into account the number of sterile
144 males released and their daily survival rate (SR) that was estimated by the mean daily relative
145 humidity (RH) as described in Bellini et al. (2010) using the equation:

$$146 \quad [1] \quad SR = 0.02 \text{ RH} - 0.48$$

147 This equation is considered valid in the RH range 48-72.5 %; above RH=72.5%, SR is assumed to be
148 equal to 97%, and below RH=48%, SR is assumed to be equal to 52% (data set supporting this
149 approach is available in Supplementary data).

150 To estimate more precisely the S / W males ratio, the number of sterile males that survived in the
151 previous release (M_{ss}) were considered by adding their number (estimated by the equation 1) to
152 the wild males (M_w) in order to be comparable to the values observed in MRR studies (marked
153 males/wild males + sterile males from the previous week).

154 *Method 2 – Ae. albopictus* wild male and female population densities were estimated by human
155 landing collection (HLC) using manual aspirators in the release and control areas in MRR studies. In
156 2018 two sampling sessions were conducted in parallel in the release (Guisa Pepoli and Caselle)
157 and in the control areas (Bolognina). The male to female ratios (M / F) in the release and in the
158 control areas were calculated and compared to determine the sterile to wild males ratio in the
159 release area for each egg monitoring week using the equation:

160
$$R_{S/W} = [(M_s + M_w)/F_w] * (F_c/M_c) - 1$$

161 where $R_{S/W}$ is the ratio of sterile/wild males; M_s , M_w and F_w are the numbers of sterile males, wild
162 males and wild females, respectively, collected in the release area; F_c and M_c are the numbers of
163 females and males, respectively, collected in the control area.

164 Method 2 has also been corrected to compare it with the rate observed in MRR studies; the rate
165 $R_{S/W}$ calculated were multiplied by $(1 - M_{ss})/M_w$.

166 Method 3 - The ratio of sterile (colored) to wild males were calculated in MRR studies to verify the
167 outcome of methods 1 and 2 (Supplementary data).

168 Two MRR sessions were undertaken in 2018: the first in the period July 06-13 and the second
169 between August 03-10 (supplementary data). Marked sterile males were released in the SIT
170 treatment areas only (Guisa and Caselle) while the recapture sessions were performed in parallel
171 either in the release or in the control localities (Bolognina).

172 Twenty-four sampling stations were randomly selected in each locality. The field sample
173 collections, using a manual aspirator for 5 minutes in each sampling station, were conducted daily
174 from 5:00 PM to 7:00 PM, starting from the first day after release and continued for seven
175 consecutive days.

176

177 **Field competitiveness of irradiated males**

178 Field competitiveness was estimated through the weekly capacity to induce sterility (CIS) index, a
179 simplified version of Fried's competitiveness index (Bellini et al. 2013b). CIS was calculated using
180 the following equation:

181
$$\text{CIS Index} = W/S * [(PW - PS) / PS]$$

182 where W and S are the numbers of wild and sterile males, respectively; PW is the percentage egg
183 hatch in the control area and PS is the percentage egg hatch in the release area.

184 Dependent-samples T-test was used to compare the two methods for estimating the sterile to wild
 185 male ratios. For describing the central tendency of the sterile/wild ratio, we also used the median
 186 because it was less affected by outliers.

187 Method 1 was used to estimate the sterile to wild males ratio for the trials carried out in all
 188 periods (2009-2018).

189

190 **Results**

191 Results showed that sterile males released weekly at the dose range of 900-1500 males/ha/week
 192 induced sterility levels from 15 to 70% in the local egg population. When induced egg sterility
 193 reached 70%, a similar reduction was found in egg numbers in the ovitraps.

194 The field data were analyzed to estimate the field competitiveness of the released *Ae. albopictus*
 195 sterile males.

196 The sterile to wild male ratios observed during the field trials using the two methods applied are
 197 shown in Table 3.

198

199 Tab. 3 Estimation of the weekly sterile to wild male ratios using the two applied methods and data
 200 observed in MRR. Field data (mean \pm SD) were collected in Guisa and Caselle (SIT area) and
 201 Bolognina (control area) in 2018
 202

Ratio Sterile/wild	Mean	SD	N	Diff.	S.D. Diff.	T-test for dependent samples			Product Moment Correlation	
						t	Df	p	R	P
Observed	0.65	0.82								
Method 1	0.69	0.53	24	-0.04	0.45	-0.42	23	0.68	0.87	<0.001
Method 2	1.07	1.52	24	-0.42	1.14	-1.79	23	0.09	0.68	<0.001

203

204

205 The mean values of the weekly sterile to wild male ratios in 2018 were similar for the two

206 methods used and no significant difference was observed when the two methods were compared

207 with observed MRR data. The sterile to wild male ratio S/W calculated with method 1 was
 208 thereafter used to assess the competitiveness of the sterile males in all the other pilot trials.
 209 The mean CIS index strongly varied in space and time ranging from 0.02 to 0.37, which indicates
 210 that the sterile males were 3 to 100 times less competitive than the wild males (Table 4).

211

212 Tab. 4 Data from eight sterile male release trials carried out in Northern Italy. The capacity to
 213 induce sterility (CIS) was estimated weekly.

214

Area_year	No. sterile males released/ha		D (% egg reduction)		S (% induced egg sterility)		Weekly CIS index		Rate sterile/wild	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Boschi_2008	1475.1	1005.2	-47.38	25.20	56.02	28.42	0.29	0.33	51.55	89.92
Boschi_2009	592.0	295.5	-73.03	12.39	64.75	18.24	0.67	1.02	35.36	109.14
Boschi_2010	857.8	236.3	6.84	61.24	16.00	31.60	0.11	0.15	5.69	5.78
Budrio_2008	1632.0	959.4	20.04	81.43	48.08	22.56	0.67	1.18	72.92	195.06
Casellina_2009	2800.3	9288.0			48.65	25.51	0.89	1.00	37.43	100.53
Gherghenzano_'12	10659.9	6842.8			50.98	19.18	0.04	0.04	105.17	135.74
Guisa_2018	783.1	545.1	-52.61	17.77	22.23	23.44	0.19	0.43	4.27	3.95
Caselle_2018	226.2	97.0	-29.28	23.69	5.02	14.46	0.16	0.32	0.72	0.40
All Grps	2870.8	5732.9	-33.33	52.92	42.42	29.42	0.39	0.75	44.82	111.02

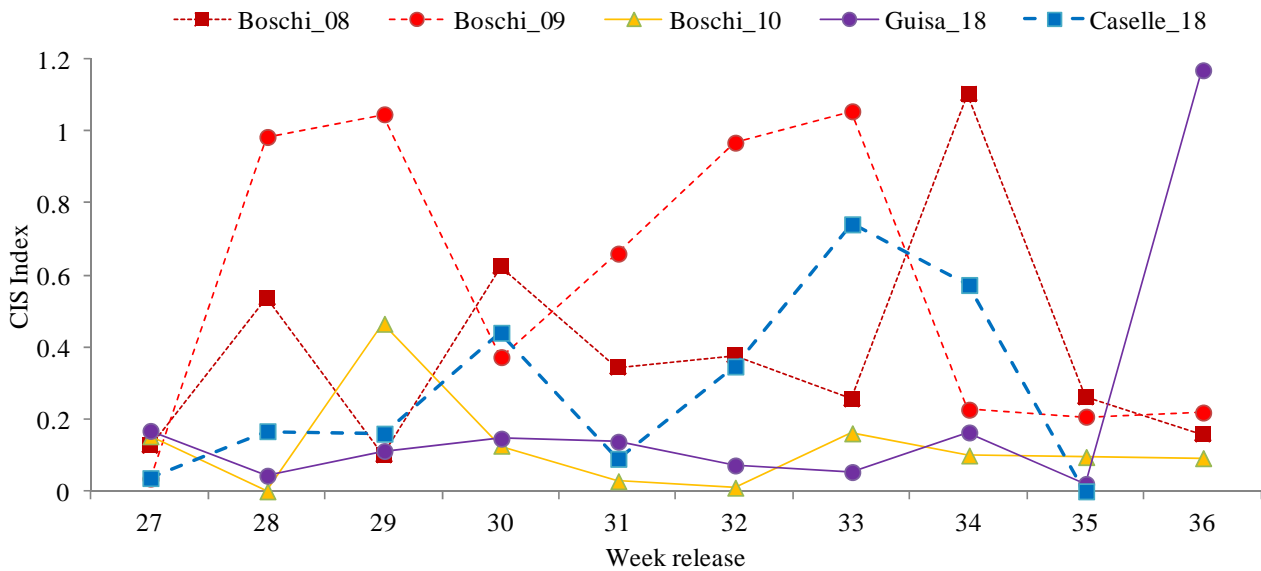
215 * The percentage decrease of egg collection (D) was calculated using the following equation: $D = (E_{Control} - E_{SIT}) / E_{Control}$ were E_{SIT} and $E_{Control}$ are the mean number of eggs per ovitrap per week in the release and in the
 216 control area, respectively. The percentage of egg sterility in SIT and control areas was estimated by
 217 calculating egg hatch using standard procedures, as reported in Bellini et al. (2013b).
 218
 219

220

221 The capacity to induce sterility was highly variable in the eight field pilot trials analysed, with a
 222 mean value of CIS 0.39 (SD±0.75) and a median value of 0.13 (0.02-0.39 Q₂₅₋₇₅). A strong temporal
 223 variability was observed when plotting the CIS index against the seasonal period, with lower CIS
 224 values observed at the beginning and at the end of the season, when the wild population density
 225 is usually lower (Albieri et al. 2010, Carrieri et al. 2011a, 2011b) and therefore the ratio of sterile
 226 to wild males tends to be higher (Fig. 1).

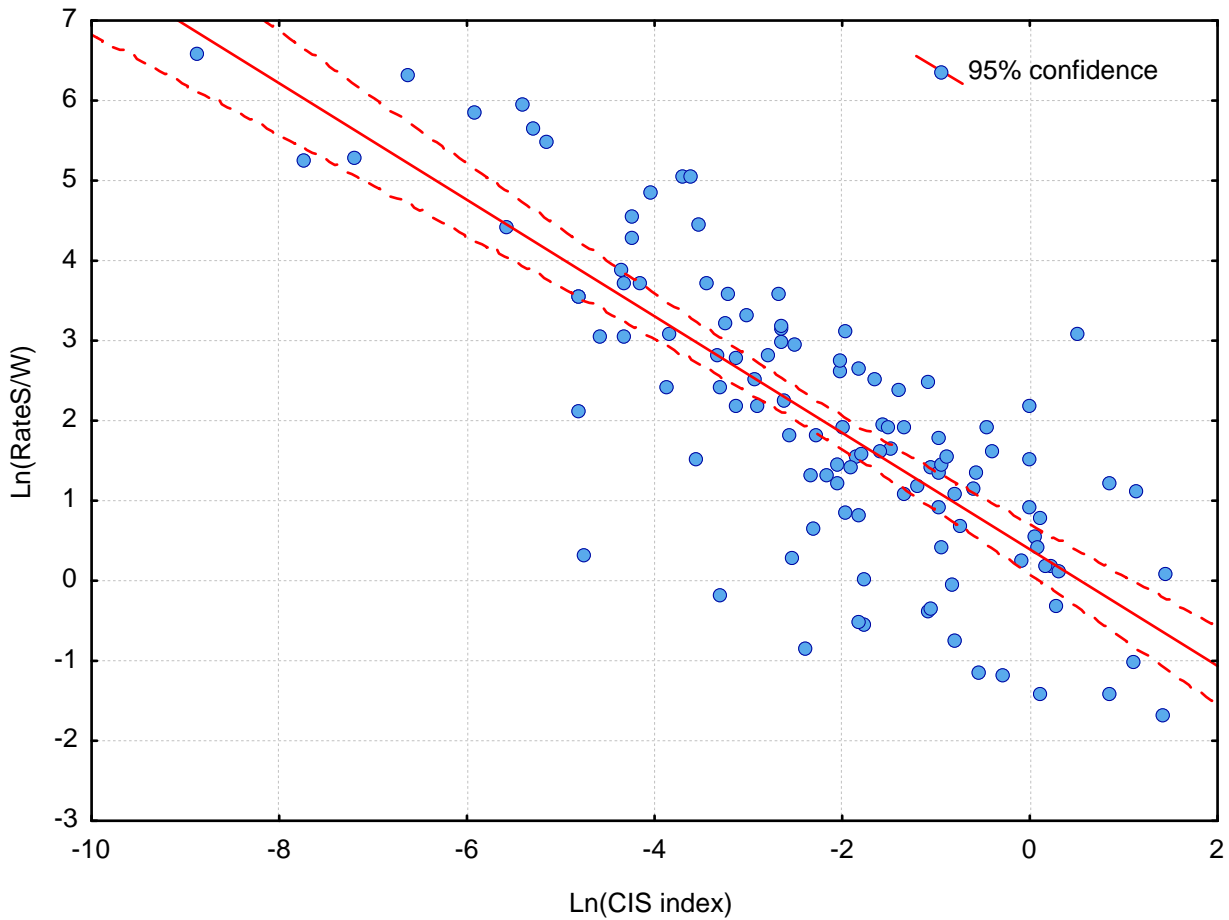
227

228 Fig. 1 Seasonal dynamics of the weekly CIS index during field pilot trials conducted in Boschi (2008,
 229 2009 and 2010) and in Guisa and Caselle in 2018.
 230



231
 232
 233 These low values of the CIS index observed at the beginning and at the end of the season may
 234 indicate that the released males were less prone to disperse and mate due to lower temperatures
 235 during these periods in comparison with the higher temperatures in the middle of the summer,
 236 thus reducing the actual sterile to wild males ratio with distance from the release points.
 237 Competitiveness of the sterile males might also be density-dependent as indicated by the strong
 238 negative correlation between CIS values and ratios of sterile/wild males as expressed by the
 239 equation: $\ln R_{S/W} = 0.39 - 0.73 \ln (\text{CIS index})$ ($R^2 = 0.62$, $F_{(1,113)} = 185.26$, $p < 0.0001$).
 240
 241

242 Fig. 2 Linear regression between Ln (S/W) males and Ln (CIS values). This analysis included all
243 weekly results and all sites.



244

245

246 Discussion

247 The competitiveness of the sterile males used in the field trials analysed in the present study was
248 tested in previous large semi-field cages showing a CIS index of 0.96 ± 0.62 and 0.71 ± 0.36 at 30
249 and 40 Gy, respectively (Bellini et al. 2013b), indicating the good quality of the sterile males
250 released.

251 A competitiveness index of at least 0.2, measured in semi-field cages with a ratio between sterile
252 to fertile males of 1: 1, is considered acceptable in Tephritid flies SIT programs (FAO/IAEA/USDA,
253 2019).

254 When moving from semi-field setting to the field, a reduction in the competitiveness index is
255 expected for several reasons. These include: the pulsing release of sterile males (mainly weekly

256 releases in this case) while wild males emerge every day; the daily mortality of sterile males
257 causing their density to decline during the intra-release time; the capillary unknown distribution of
258 breeding sites matched with the point site release of sterile males; the obstacles to the dispersal
259 of sterile males caused by artificial barriers in urban areas; and the immigration of already mated
260 females from outside the SIT pilot area.

261 The median CIS index obtained in the eight SIT field trials conducted in Northern Italy in the period
262 2008-2018 was 0.12 (range 0.03 - 0.38), which is considered good when compared with
263 competitiveness indexes observed in other genetic control pilot field trials.

264 In pilot trials conducted in Brazil and the Cayman Islands by releasing the transgenic *Ae. aegypti*
265 strain OX513A, field competitiveness values of 0.031 (95% CI: 0.025-0.036) and 0.059 (95% CI:
266 0.011 – 0.210), respectively, were obtained (Harris et al. 2011, Carvalho et al. 2015). The field
267 competitiveness of *Ae. albopictus* irradiated mosquitoes assessed in our study was thus overall
268 much higher than that of *Ae. aegypti* transgenic strains.

269 The large variability observed in the CIS index among releases and among weeks of the same trial
270 is difficult to explain and may be attributable to the impact of climatic condition.

271 We hypothesized that part of the wild females might be unattainable by the sterile males
272 whatever the ratio imposed because they are located in cryptic habitats difficult for the sterile
273 males to access. This might be particularly relevant in urban areas because of the numerous
274 artificial obstacles such as perimetric hedges and walls. It will probably be possible to reduce this
275 gap by air releasing sterile males in order to achieve a more homogeneous covering of the target
276 area (Bouyer et al. 2020b).

277 In natural environments, the field competitiveness of *Glossina palpalis gambiensis* sterile males
278 was estimated 0.07 in Burkina Faso (Sow et al. 2012), while it resulted 0.14 (SD 0.10) and 0.76 (SD
279 0.11) for two different strains in Senegal (Bassène et al. 2017).

280 In the case of the New World screwworm (*Cochliomyia hominivorax*), field competitiveness was
281 estimated at 0.29–0.43 at smaller scales, decreasing to 0.1 for larger programs (Mayer et al. 1998).
282 An estimated $C = 0.17$ has been reported for a small-scale trial of irradiated Mediterranean fruit fly
283 (*Ceratitis capitata*), increasing to 0.42 if the males were exposed to ginger root oil (Shelly et al.
284 2007). The authors also observed that the competitiveness values varied inversely with the S / W
285 ratio. In a large trial included in a successful Medfly control program, no significant induced
286 sterility was observed until sterile/wild males ratios reached 100:1 or higher with an estimated
287 competitiveness in the range 0.0001–0.001 (Rendón et al. 2004).

288 A negative correlation between the S/W males ratio and the CIS index value similar to our finding
289 was observed in previous trials conducted under semi-field and field conditions using irradiated
290 and transgenic sterile males (Harris et al. 2011, Damiens et al. 2016). Understanding the reasons
291 for this negative correlation will be instrumental in the proper planning of future mosquito control
292 programs that rely on genetic control. Actually, there is probably an optimal range of S/W males
293 ratio, beyond which the increase in the dose of sterile males released does not add a strong
294 benefit in the induced sterility. This is in line with the marginal decreasing productivity law and
295 supports the empirical observation about the difficulty to achieve eradication. It is important to
296 identify this optimal S/W ratio as this will allow accurate estimates of sterile male release densities
297 to maximize the cost-efficiency of SIT campaigns aimed at suppression.

298 Finally, the mean CIS field index observed in our studies resulted in the range of the CIS values
299 obtained in successful programs against other insect species, indicating that the SIT technology
300 applied to *Aedes* mosquito control could achieve satisfactory results. However, the strong
301 variability shows that the various processes of the SIT package, namely mass-rearing, handling,
302 irradiation and release of the sterile males should be mastered to reach the highest values
303 obtained in our study, in order to optimize cost-benefit in the field.

304

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312

313 **Author contributions**

314 RB, MC, FB and JB conceptualized the study. MC conducted the statistical analysis. RB, MC, FB, AP,
315 MM and JB contributed to the writing of the original draft and to the review & editing process.

316

317 **References**

- 318 Albieri, A., M. Carrieri, P. Angelini, F. Baldacchini, C. Venturelli, S. Mascali Zeo, et al. 2010. Quantitative
319 monitoring of *Aedes albopictus* in Emilia-Romagna, Northern Italy: cluster investigation and
320 geostatistical analysis. *Bull. Insectol.* 63: 209-216.
- 321 Bassène, M. D., M. T. Seck, S. Pagabeleguem, A. G. Fall, M. J. B. Vreysen, G. Gimonneau, and J. Bouyer.
322 2017. Competitiveness and survival of two strains of *Glossina palpalis gambiensis* in an urban area
323 of Senegal. *PLoS Negl. Trop. Dis.* 11: e0006172.
- 324 Bellini, R., M. Calvitti, A. Medici, M. Carrieri, G. Celli, and S. Maini. 2007. Use of the Sterile Insect Technique
325 against *Aedes albopictus* in Italy: First Results of a Pilot Trial, pp. 505-515. In M. J. B. Vreysen, A. S.
326 Robinson and J. Hendrichs (eds.), *Area-Wide Control of Insect Pests: From Research to Field*
327 *Implementation*. Springer, Dordrecht, The Netherlands.
- 328 Bellini, R., A. Albieri, F. Balestrino, M. Carrieri, D. Porretta, S. Urbanelli, et al. 2010. Dispersal and survival of
329 *Aedes albopictus* (Diptera: Culicidae) males in Italian urban areas and significance for sterile insect
330 technique application. *J. Med. Entomol.* 47(6): 1082-1091. doi: 10.1603/ME09154.
- 331 Bellini, R., A. Medici, A. Puggioli, F. Balestrino, and M. Carrieri. 2013a. Pilot field trials with *Aedes albopictus*
332 irradiated sterile males in Italian urban areas. *J. Med. Entomol.* 50: 317-325.
- 333 Bellini, R., F. Balestrino, A. Medici, G. Gentile, R. Veronesi, and M. Carrieri. 2013b. Mating competitiveness
334 of *Aedes albopictus* radio-sterilized males in large enclosures exposed to natural conditions. *J. Med.*
335 *Entomol.* 50(1): 94-102. doi: <http://dx.doi.org/10.1603/ME1105850>, 94-102.
- 336 Bouyer, J., H. Yamada, R. Pereira, K. Bourtzis, and M. J. B. Vreysen. 2020a. Phased Conditional Approach for
337 Mosquito Management using the Sterile Insect Technique. *Trends Parasitol.* 36: 325-336.
- 338 Bouyer, J., N. Culbert, A. H. Dicko, A. Klaptocz, J. Germann, T. Wallner, G. S. Herranz, R. Argiles Herrero, J.
339 Virgilio, M. Gomez, H. Yamada, F. Balestrino, and M. Vreysen. 2020b. Field performance of sterile
340 male mosquitoes released from a drone. *Science Robotics* (in press).
- 341 Carrieri, M., P. Angelini, C. Venturelli, B. Maccagnani, and R. Bellini. 2011a. *Aedes albopictus* (Diptera:
342 Culicidae) population size survey in the 2007 Chikungunya outbreak area in Italy. I. Characterization
343 of breeding sites and evaluation of sampling methodologies. *J. Med. Entomol.* 48: 1214-1225.

344 Carrieri, M., A. Albieri, P. Angelini, F. Baldacchini, C. Venturelli, S. Mascali Zeo, et al. 2011b. Surveillance of
345 the chikungunya vector *Aedes albopictus* (Skuse) in Emilia-Romagna (northern Italy): organizational
346 and technical aspects of a large scale monitoring system. *J. Vector Ecol.* 36: 108-116. doi:
347 10.1111/j.1948-7134.2011.00147.x.

348 Carrieri, M., A. Alessandro, S. Urbanelli, P. Angelini, C. Venturelli, C. Matrangolo, and Romeo Bellini. 2017.
349 Quality control and data validation procedure in large-scale quantitative monitoring of mosquito
350 density: the case of *Aedes albopictus* in Emilia-Romagna region, Italy. *Pathogens and Global Health*
351 111(2): 83-90. doi: 10.1080/20477724.2017.1292992.

352 Carvalho, D. O., A. R. McKemey, L. Garziera, R. Lacroix, C. A. Donnelly, L. Alphey, et al. 2015. Suppression of
353 a field population of *Aedes aegypti* in Brazil by sustained release of transgenic male mosquitoes.
354 *PLoS Negl Trop Dis* 9(7): e0003864. doi:10.1371/journal.pntd.000386.

355 Damiens, D., P. O. Tjeck, C. Lebon, G. Le Goff, and L. C. Gouagna. 2016. The effects of age at first mating and
356 release ratios on the mating competitiveness of gamma-sterilised *Aedes albopictus* males under
357 semi field conditions. *Vector Biol. J.* 1: 29-31. doi:10.4172/2473-4810.1000107.

358 Dicko, A. H., R. Lancelot, M. T. Seck, L. Guerrini, B. Sall, M. Lo, et al. 2014. Using species distribution models
359 to optimize vector control: the tsetse eradication campaign in Senegal. *Proc. Natl. Acad. Sci. U. S. A.*
360 111: 10149-10154, doi:10.1073/pnas.1407773111.

361 Dyck, V. A., J. Hendrichs, and A. S. Robinson (eds). 2005. *Sterile Insect Technique: principles and practice in*
362 *area-wide integrated pest management.* Springer Netherlands.

363 Enkerlin, W., J. M Gutiérrez-Ruelas, A. V. Cortes, E. C., Roldan, D. Midgarden, E. Lira, et al. 2015. Area
364 freedom in Mexico from Mediterranean fruit fly (Diptera: Tephritidae): a review of over 30 years of
365 a successful containment program using an integrated Area-Wide SIT approach. *Fla. Entomol.* 98(2):
366 665-681.

367 FAO/IAEA/USDA. 2019. *Product Quality Control for Sterile Mass-Reared and Released Tephritid Fruit Flies.*
368 *Version 7.0.* International Atomic Energy Agency, Vienna, Austria. 148 pp.

369 Flores, H. A., and S. L. O'Neill. 2018. Controlling vector-borne diseases by releasing modified mosquitoes.
370 *Nat Rev Microbiol.* 16(8): 508-518. doi: 10.1038/s41579-018-0025-0.

371 Gossner, C. M., E. Ducheyne, and F. Schaffner. 2018. Increased risk for autochthonous vector-borne
372 infections transmitted by *Aedes albopictus* in continental Europe. Euro Surveill. 23(24). doi:
373 10.2807/1560-7917.ES.2018.23.24.1800268.

374 Grigoraki, L., D. Pipini, P. Labbe, A. Chaskopoulou, M. Weill, and J. Vontas. 2017. Carboxylesterase gene
375 amplifications associated with insecticide resistance in *Aedes albopictus*: Geographical distribution
376 and evolutionary origin. PloS Negl. Trop. Dis. 11, e0005533.

377 Harris, A. F., D. Nimmo, A. R. McKemey, N. Kelly, S. Scaife, C. A. Donnelly, et al. 2011. Field performance of
378 engineered male mosquitoes. Nat. Biotechnol. 29(11): 1034-1037. doi: 10.1038/nbt.2019.

379 Lees, R. S., D. O. Carvalho, and J. Bouyer. 2020. Potential impact of integrating the sterile insect technique
380 into the fight against disease-transmitting mosquitoes. In A. V. Dyck, J. Hendrichs and A. S.
381 Robinson (eds.), Sterile Insect Technique. Principles and Practice in Area-Wide Integrated Pest
382 Management, vol. in press, Second edition ed. CRC Press, Vienna.

383 McGraw, E. A. and S. L. O'Neill. 2013. Beyond insecticides: new thinking on an ancient problem. Nat Rev
384 Microbiol. 11: 181-193.

385 Mayer, D. G, M. G. Atzeni, M. A. Stuart, K. A. Anaman, and D. G. Butler. 1998. Mating competitiveness of
386 irradiated flies for screwworm fly eradication campaigns. Preventive Veterinary Medicine 36: 1-9.

387 Mitchell, C. J. 1995. The role of *Aedes albopictus* as an arbovirus vector. Parassitologia 37: 109-113.

388 Pichler, V., R. Bellini, R. Veronesi, D. Arnoldi, A. Rizzoli, R. Paolo Lia, et al. 2018. First evidence of resistance
389 to pyrethroid insecticides in Italian *Aedes albopictus* populations 26 years after invasion. Pest
390 manag. sci. 74: 1319-1327.

391 Reiter, P. 2016. Control of urban Zika vectors: should we return to the successful PAHO/WHO strategy? PloS
392 Negl. Trop. Dis. 10, e0004769.

393 Rendòn, P., D. McInnis, D. Lance, and J. Stewart. 2004. Medfly (Diptera: Tephritidae) genetic sexing: large-
394 scale field comparison of males-only and bisexual sterile fly releases in Guatemala. J. Econ.
395 Entomol. 97: 1547-1553.

396 Sow, A., I. Sidibè, Z. Bengaly, A. Z. Bancè, G. J. Sawadogo, P. Solano, et al. 2012. Irradiated male tsetse from
397 a 40-year-old colony are still competitive in a riparian Fforest in Burkina Faso. PLoS ONE 7(5):
398 e37124. doi:10.1371/journal.pone.0037124.

399 Tsetsarkin, K. A., D. L. Vanlandingham, C. E. McGee, and S. A Higgs. 2007. Single mutation in chikungunya
400 virus affects vector specificity and epidemic potential. Plos Pathog. 3, e201.

401 Vreysen, M. J. B., K. M. Saleh, M. Y. Ali, A. M. Abdulla, Z.-R. Zhu, K. G. Juma, et al. 2000. *Glossina austeni*
402 (Diptera: Glossinidae) eradicated on the Island of Unguja, Zanzibar, using the Sterile Insect
403 Technique. J. Econ. Entomol. 93: 123-135.

404 Wong, P.-S. J., M.-Z. I. Li, C.-S. Chong, and L.-C. Ng Cheong-Huat Tan. 2013. *Aedes (Stegomyia) albopictus*
405 (Skuse): a potential vector of Zika virus in Singapore. PloS Negl. Trop. Dis. 7, e2348.

406 Wyss, J. H. 2006. Screwworm Eradication in the Americas. Ann. N. Y. Acad. Sci. 916: 186-193.