

Electronic Supplementary Information (ESI) for

Photochemical generation of acyl and carbamoyl radicals using a nucleophilic organic catalyst: applications and mechanism thereof

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A. General Information

The NMR spectra were recorded at 400 MHz and 500 MHz for ^1H and 100 or 125 MHz for ^{13}C . The chemical shift (δ) for ^1H and ^{13}C are given in ppm relative to residual signals of the solvents (CHCl_3 @ 7.26 ppm ^1H NMR and 77.16 ppm ^{13}C NMR, and tetramethylsilane @ 0 ppm). Coupling constants are given in Hertz. The following abbreviations are used to indicate the multiplicity: s, singlet; d, doublet; q, quartet; m, multiplet; bs, broad signal; app, apparent.

High resolution mass spectra (HRMS) were obtained from the ICIQ HRMS unit on MicroTOF Focus and Maxis Impact (Bruker Daltonics) with electrospray ionization. (ESI).

UV-vis measurements were carried out on a Shimadzu UV-2401PC spectrophotometer equipped with photomultiplier detector, double beam optics and D₂ and W light sources or an Agilent Cary60 spectrophotometer.

Emission spectra of light sources were recorded on Ocean Optics USB4000 fiber optic spectrometer.

Isolated yields refer to materials of >95% purity as determined by ^1H NMR.

The authors are indebted to the team of the Research Support Area at ICIQ, particularly to the NMR and the High-Resolution Mass Spectrometry Units. Grace Fox is thanked for proofreading the manuscript.

General Procedures. All reactions were set up under an argon atmosphere in oven-dried glassware. Synthesis grade solvents were used as purchased, anhydrous solvents were taken from a commercial SPS solvent dispenser. Chromatographic purification of products was accomplished using forced-flow chromatography (FC) on silica gel (35-70 mesh). For thin layer chromatography (TLC) analysis throughout this work, Merck pre-coated TLC plates (silica gel 60 GF₂₅₄, 0.25 mm) were employed, using UV light as the visualizing agent and an acidic mixture of vanillin or basic aqueous potassium permanganate (KMnO_4) stain solutions, and heat as developing agents. Organic solutions were concentrated under reduced pressure on a Büchi rotatory evaporator.

Determination of Enantiomeric Purity. UPC² analysis on chiral stationary phase was performed on a Waters Acquity instrument using an ID3 chiral column. The exact conditions for the analyses are specified within the characterization section.

Materials. Most of the starting materials used in this study are commercial and were purchased in the highest purity available from Sigma-Aldrich, Fluka, Alfa Aesar, Fluorochem, and used as received, without further purifications.

The following substrates were synthesized according to reported procedures (Figure S1).¹⁻⁸

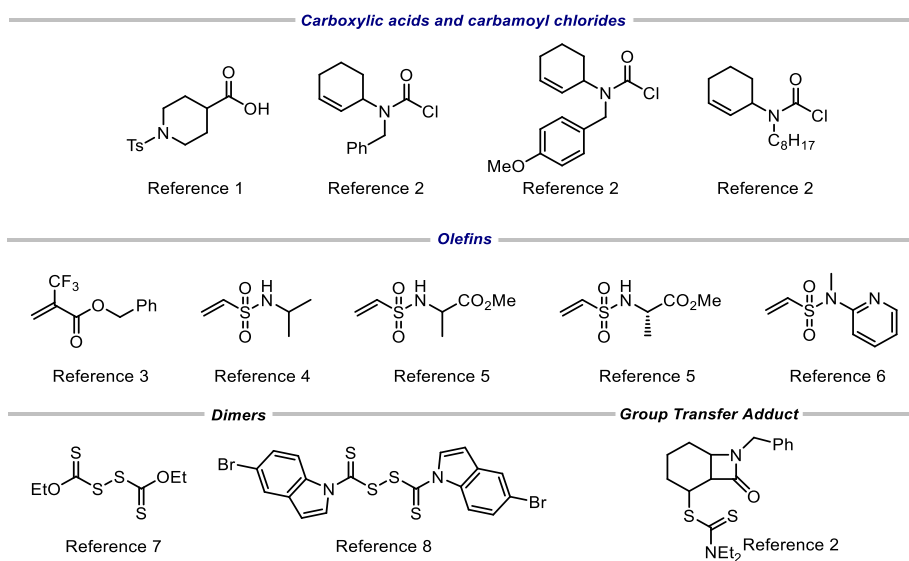
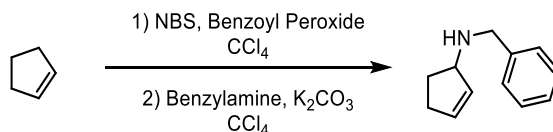


Figure S1: Starting materials synthesised according to known procedures.

B. Substrate Synthesis

Synthesis of *N*-benzylcyclopent-2-en-1-amine:

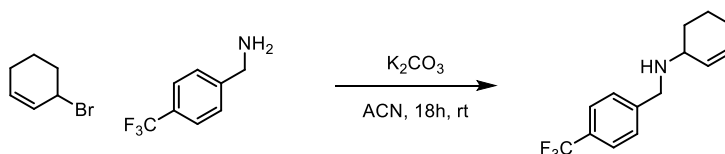


A 2 M solution of cyclopentene (0.973 mL, 11 mmol) in CCl₄ (5.4 mL) was prepared and then NBS (2.26 g, 10 mmol) and benzoic peroxyanhydride (36 mg, 0.15 mmol) were sequentially added. The solution was stirred at 40 °C for 4 hours, after which the reaction was left at ambient temperature without stirring. After 2 hours, the floating materials were filtered off and washed with CCl₄. The organic phase was then placed in a separatory funnel and washed with distilled water. The organic phase was collected, treated with MgSO₄ and subsequently filtered. At this stage, benzylamine (3.28 mL, 30 mmol) and K₂CO₃ (1.38 g, 10 mmol) were directly added to the solution. The reaction was stirred overnight at r.t and the resulting crude mixture was purified by silica gel chromatography (eluent: 9:1 hexane/AcOEt) to afford 234 mg of *N*-benzylcyclopent-2-en-1-amine (14% yield over 2 steps) as a yellow-orange oil.

¹H NMR (400 MHz, CDCl₃) δ 7.39 – 7.24 (m, 5H), 5.89 (m, 2H), 3.93 (m, 1H), 3.84 (dd, *J* = 16.6, 12.9 Hz, 2H), 2.48 (m, 1H), 2.36 – 2.18 (m, 2H), 1.63 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 140.5, 133, 132.8, 128.4, 128.3, 126.9, 63.9, 51.8, 31.3, 30.8

Synthesis of *N*-(4-(trifluoromethyl)benzyl)cyclohex-2-en-1-amine:



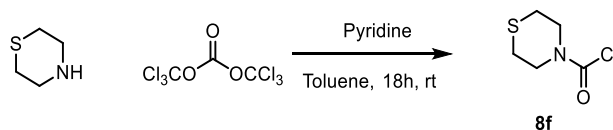
N-(4-(trifluoromethyl)benzyl)cyclohex-2-en-1-amine was prepared according to a reported procedure.² A solution of 4-(trifluoromethyl)phenylmethanamine (1.314 g, 7.5 mmol) in CH₃CN (1.7 mL) was treated with 3-bromocyclohexene (0.288 mL, 2.5 mmol) and K₂CO₃ (346 mg, 2.5 mmol). After 2 hours at ambient temperature, the reaction mixture was quenched with H₂O (20 mL) and extracted with EtOAc (2 x 25 mL). The combined organic extracts were washed with brine (20 mL), dried over MgSO₄, filtered, evaporated under reduced pressure, and purified by column chromatography (eluent: CH₂Cl₂ to CH₂Cl₂/EtOH 9:1) to give the product (607 mg, 95 % yield) as a light-yellow oil.

¹H NMR (400 MHz, CDCl₃) δ 7.57 (d, *J* = 8.1 Hz, 2H), 7.47 (d, *J* = 8.0 Hz, 2H), 5.82 – 5.76 (m, 1H), 5.75 – 5.62 (m, 1H), 3.90 (dd, *J* = 13.6, 3.3 Hz, 2H), 3.23 – 3.16 (m, 1H), 2.10 – 1.97 (m, 2H), 1.93 – 1.85 (m, 1H), 1.80 – 1.70 (m, 1H), 1.61 – 1.42 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 145.1, 129.8, 129.4, 129.3 (q, *J* = 32.7 Hz), 128.4, 125.4 (q, *J* = 3.7 Hz), 124.4 (q, *J* = 271.9 Hz), 52.6, 50.6, 29.6, 25.4, 20.3.

¹⁹F NMR (376 MHz, CDCl₃) δ -62.47.

Synthesis of compound **8f**:



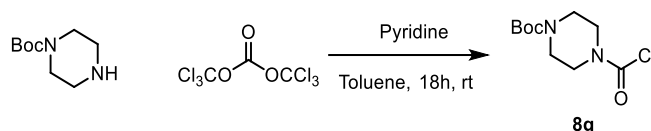
Compound **8f** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (294 mg, 0.99 mmol) in toluene (15 mL) under N₂, pyridine (0.290 mL, 3.60 mmol)

and subsequently a solution of thiomorpholine (310 mg, 3 mmol) in toluene (5 mL) were added. The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH₄Cl (20 mL of a saturated aq. solution) and extracted with Et₂O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H₂O (40 mL) and brine (40 mL), dried over MgSO₄, filtered and evaporated under reduced pressure to afford carbamoyl chloride **8f** (310 mg, 75% yield) as a colorless oil.

¹H NMR (400 MHz, CDCl₃) δ 3.94 (dt, *J* = 38.9, 5.1 Hz, 1H), 2.72 – 2.64 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 148.5, 51.7, 49.3, 27.7, 27.3

Synthesis of **8g**:



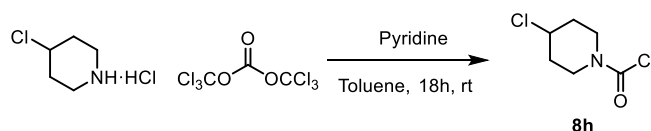
Compound **8g** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (294 mg, 0.99 mmol) in toluene (15 mL) under N₂, pyridine (0.290 mL, 3.60 mmol) and subsequently a solution of *N*-Boc Piperazine (559 mg, 3 mmol) in toluene (5 mL) were sequentially added. The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH₄Cl (20 mL of a saturated aq. solution) and extracted with Et₂O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H₂O (40 mL) and brine (40 mL), dried over MgSO₄, filtered and evaporated under reduced pressure to give carbamoyl chloride **8g** (597 mg, 82% yield) as a colorless oil.

¹H NMR (300 MHz, CDCl₃) δ 3.64 (m, 4H), 3.48 (d, *J* = 4.8 Hz, 4H), 1.46 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 154.3, 148.5, 80.7, 48.5, 46, 28.3.

HRMS (ESI pos): calculated for C₁₀H₁₇ClNaN₂O₃ (M+Na⁺): 271.0800, found: 271.0818.

Synthesis of compound **8h**:

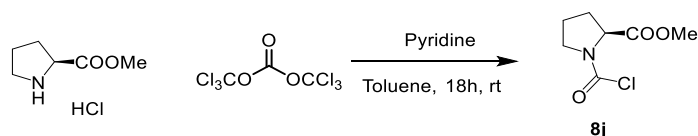


Compound **8h** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (259 mg, 0.872 mmol) in toluene (15 mL) under N₂, pyridine (0.498 mL, 6.15 mmol) and subsequently a solution of 4-chloropiperidine hydrochloride (400 mg, 2.56 mmol) in toluene (5 mL) were sequentially added. The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH₄Cl (20 mL of a saturated aq. solution) and extracted with Et₂O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H₂O (40 mL) and brine (40 mL), dried over MgSO₄, filtered and evaporated under reduced pressure to give carbamoyl chloride **8h** (420 mg, 90% yield) as a colorless oil.

¹H NMR (400 MHz, CDCl₃) δ 4.34 (tt, *J* = 6.5, 3.6 Hz, 1H), 4.01 – 3.64 (m, 4H), 2.18 – 2.03 (m, 2H), 2.00 – 1.86 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 148.4, 55.6, 45.6, 43.1, 34.7, 34.2.

Synthesis of compound 8j:

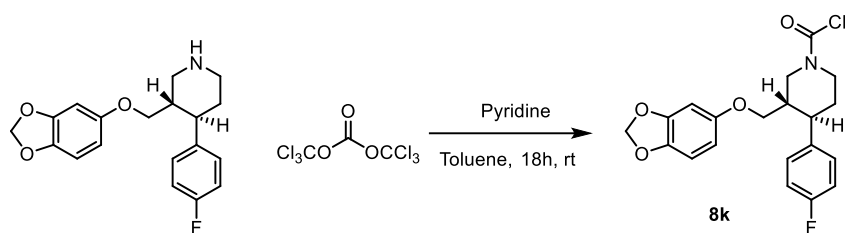


Compound **8j** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (207 mg, 0.68 mmol) in toluene (15 mL) under N₂, pyridine (0.204 mL, 2.54 mmol) and subsequently a solution of L-methyl proline hydrochloride (350 mg, 2.13 mmol). The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH₄Cl (20 mL of a saturated aq. solution), and extracted with Et₂O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H₂O (40 mL) and brine (40 mL), dried over MgSO₄, filtered and evaporated under reduced pressure to give carbamoyl chloride **8j** (275 mg, 68% yield) as a colorless oil.

¹H NMR (400 MHz, CDCl₃, 1:1 mixture of rotamers) δ : 4.57 – 4.46 (m, 1H), 3.85 - 3.55 (m, 2H), 3.77 (d, *J* = 12.4 Hz, 3H), 2.35 – 2.25 (m, 1H), 2.17 – 1.93 (m, 3H).

¹³C NMR (124 MHz, CDCl₃, 1:1 mixture of rotamers) δ : 171.8, 171.3, 147.8, 146.9, 62.4, 60.7, 52.9, 50.7, 49.2, 30.4, 30.3, 23.7, 23.6

Synthesis of compound 8k:



Compound **8k** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (89 mg, 0.30 mmol) in toluene (3 mL) under N₂, pyridine (88 μ L, 1 mmol) and subsequently a solution of Paroxetine (300 mg, 0.91 mmol) in toluene (6 mL), were added. The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH₄Cl (20 mL of a saturated aq. solution) and extracted with Et₂O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H₂O (40 mL) and brine (40 mL), dried over MgSO₄, filtered and evaporated under reduced pressure to afford carbamoyl chloride **8k** (130 mg, 36% yield) as a white crystal.

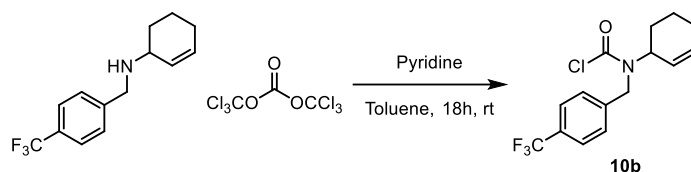
¹H NMR (500 MHz, CDCl₃) δ 7.14 (m, 2H), 7.00 (m, 2H), 6.64 (m, 1H), 6.36 (s, 1H), 6.14 (m, 1H), 5.89 (s, 2H), 4.63 (m, 1H), 4.49 (m, 1H), 3.63 (t, *J* = 8.7 Hz, 1H), 3.48 (dd, *J* = 9.6, 6.0 Hz, 1H), 3.20 (t, *J* = 12.9 Hz, 1H), 3.02 (m, 1H), 2.82 (m, 1H), 2.10 (m, 1H), 1.93 (m, 1H), 1.82 (qd, *J* = 12.9, 4.4 Hz, 1H)

¹³C NMR (126 MHz, CDCl₃) δ 162.9, 160.9, 154.0, 148.4, 142.0, 128.9, 115.8, 108.0, 105.8, 101.3, 98.1, 68.4, 52.2, 49.6, 47.0, 43.8, 42.3, 33.7

¹⁹F NMR (376 MHz, CDCl₃, proton decoupled) δ -115.51

HRMS (ESI pos): calculated for C₂₀H₁₉ClFNaNO₄ (M+Na⁺): 414.09, found 414.0884

Synthesis of compound 10b:



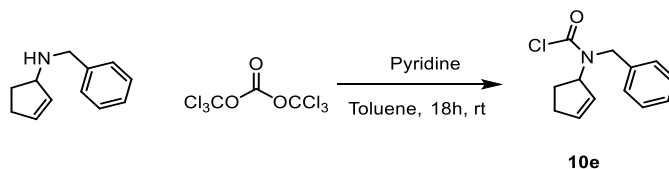
Compound **10b** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (252 mg, 0.848 mmol) in toluene (15 mL) under N_2 , pyridine (0.242 mL, 2.99 mmol) and subsequently a solution of *N*-(4-(trifluoromethyl)benzyl)cyclohex-2-en-1-amine (637 mg, 2.5 mmol) in toluene (4 mL), were added. The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH_4Cl (20 mL of a saturated aq. solution) and extracted with Et_2O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H_2O (40 mL) and brine (40 mL), dried over MgSO_4 , filtered and evaporated under reduced pressure to give carbamoyl chloride **10b** (722 mg, 91% yield) as a yellow oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3 mixture of rotamers) δ 7.60 (dd, $J = 16.7, 8.1$ Hz, 2H), 7.36 (d, $J = 7.9$ Hz, 2H), 5.85 – 6.00 (m, 1H), 5.55 – 5.41 (m, 1H), 5.10 – 4.90 (m, 1H), 4.82 – 4.49 (m, 2H), 2.08 – 1.94 (m, 3H), 1.85 – 1.40 (m, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3 mixture of rotamers) δ 150.4, 150.3, 141.8, 141.4, 133.5, 133.4, 127.3, 126.5, 126.1, 126.1, 125.84 – 125.46 (m), 124.1 (q, $J = 271.9$ Hz) 58.7, 57.0, 50.4, 49.1, 28.5, 27.6, 24.5, 24.4, 21.2, 21.1.

$^{19}\text{F NMR}$ (376 MHz, CDCl_3 mixture of rotamers) δ -62.59, -62.63.

Synthesis of compound 10e:



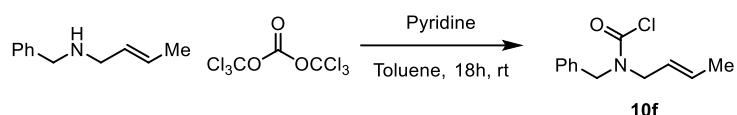
Compound **10e** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (134 mg, 0.44 mmol) in toluene (8 mL) under N_2 , pyridine (0.129 mL, 1.6 mmol) and subsequently a solution of *N*-benzylcyclopent-2-en-1-amine (231 mg, 1.33 mmol) in toluene (2 mL). The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH_4Cl (20 mL of a saturated aq. solution) and extracted with Et_2O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H_2O (40 mL) and brine (40 mL), dried over MgSO_4 , filtered and evaporated under reduced pressure to give carbamoyl chloride **10e** (106 mg, 34% yield) as a yellowish oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3 , mixture of rotamers) δ . 7.40 – 7.20 (m, 5H), 6.02 - 5.95 (m, 1H), 5.58 – 5.46 (m, 2H), 4.66 – 4.37 (m, 1H), 2.44 – 2.20 (m, 3H), 1.75 – 1.59 (m, 1H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3 , mixture of rotamers) δ 150.1, 150, 137.3, 136.9, 136.4, 128.8, 128.7, 128.7, 128.6, 128.5, 127.3, 127.2, 126.2, 67.6, 66, 50.6, 48.9, 31.4, 31.3, 28.9, 28.4.

HRMS (ESI pos): calculated for $\text{C}_{13}\text{H}_{14}\text{ClNaNO}$ ($\text{M}+\text{Na}^+$) 258.07, found 258.0653.

Synthesis of compound 10f:

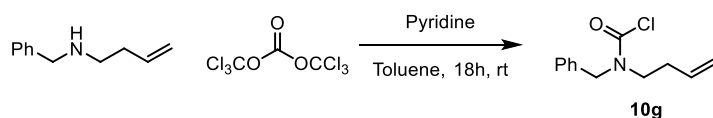


Compound **10f** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (250 mg, 0.843 mmol) in toluene (15 mL) under N₂, pyridine (0.241 mL, 2.98 mmol) and subsequently a solution of *N*-benzylbut-2-en-1-amine (400 mg, 2.48 mmol) in toluene (4 mL), were added. The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH₄Cl (20 mL of a saturated aq. solution) and extracted with Et₂O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H₂O (40 mL) and brine (40 mL), dried over MgSO₄, filtered and evaporated under reduced pressure to give carbamoyl chloride **10f** (508 mg, 92% yield) as a yellow oil.

¹H NMR (400 MHz, CDCl₃, 1:1 mixture of rotamers) δ 7.42 – 7.30 (m, 3H), 7.29 – 7.24 (m, 2H), 5.75 – 5.54 (m, 1H), 5.53 – 5.34 (m, 1H), 4.68 (s, 1H), 4.55 (s, 1H), 3.90 (dd, *J* = 15.6, 6.3 Hz, 2H), 1.78 – 1.66 (m, 3H).

¹³C NMR (101 MHz, CDCl₃, 1:1 mixture of rotamers) δ 150.2, 149.6, 135.8, 135.6, 131.3, 130.7, 129.0, 128.9, 128.4, 128.3, 128.1, 128.1, 127.3, 124.3, 124.0, 53.2, 51.9, 51.5, 50.5, 17.8.

Synthesis of compound 10g:

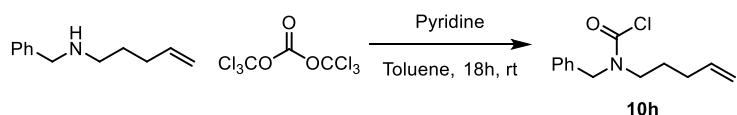


Compound **10g** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (252 mg, 0.85 mmol) in toluene (15 mL) under N₂, pyridine (0.243 mL, 3.0 mmol) and subsequently a solution of *N*-benzylbut-3-en-1-amine (403 mg, 2.5 mmol) in toluene (4 mL), were added. The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH₄Cl (20 mL of a saturated aq. solution) and extracted with Et₂O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H₂O (40 mL) and brine (40 mL), dried over MgSO₄, filtered and evaporated under reduced pressure to give carbamoyl chloride **10g** (475 mg, 85% yield) as a yellow oil.

¹H NMR (400 MHz, CDCl₃, 1:1 mixture of rotamers) δ 7.41 – 7.31 (m, 3H), 7.30 – 7.24 (m, 2H), 5.80 – 5.67 (m, 1H), 5.16 – 5.04 (m, 2H), 4.72 (s, 1H), 4.59 (s, 1H), 3.53 – 3.35 (m, 2H), 2.42 – 2.30 (m, 2H).

¹³C NMR (101 MHz, CDCl₃, 1:1 mixture of rotamers) δ 150.3, 149.6, 135.8, 135.6, 134.3, 134.0, 129.1, 129.0, 128.3, 128.2, 128.2, 127.2, 118.0, 117.8, 54.7, 52.7, 49.8, 48.9, 32.6, 31.7.

Synthesis of compound 10h:



Compound **10h** was prepared according to a modification of a reported procedure.² To a solution of triphosgene (252 mg, 0.85 mmol) in toluene (15 mL) under N₂, pyridine (0.243 mL, 3.0 mmol) and subsequently a solution of *N*-benzylpent-4-en-1-amine (438 mg, 2.5 mmol) in toluene (4 mL), were added. The reaction mixture was stirred for 18 hours at ambient temperature, quenched with NH₄Cl (20 mL of a saturated aq. solution) and extracted with Et₂O (2 x 30 mL). The combined organic extracts were washed sequentially with HCl (40 mL of a 0.25 M aq. solution), H₂O (40

mL) and brine (40 mL), dried over MgSO₄, filtered and evaporated under reduced pressure to give carbamoyl chloride **10h** (553 mg, 93% yield) as a yellow oil.

¹H NMR (400 MHz, CDCl₃, 1:1 mixture of rotamers) δ 7.41 – 7.30 (m, 3H), 7.29 – 7.24 (m, 2H), 5.81 – 5.68 (m, 2H), 5.08 – 4.92 (m, 2H), 4.71 (s, 1H), 4.58 (s, 1H), 3.36 (dt, *J* = 13.6, 7.8 Hz, 2H), 2.05 (q, *J* = 7.5 Hz, 2H), 1.70 (app h, *J* = 7.6 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃, 1:1 mixture of rotamers) δ 150.3, 149.6, 137.3, 137.1, 135.9, 135.7, 129.0, 129.0, 128.2, 127.2, 115.8, 115.6, 54.5, 52.6, 50.0, 49.1, 30.8, 30.8, 27.1, 26.3.

C. Experimental Procedures

C1. Reaction of Aromatic Acyl Chlorides

C1.1 Experimental Setup

Our photoreactor consisted of a 12.5 cm diameter jar, fitted with 4 standard 29 sized ground glass joints arranged in a square and a central 29 sized joint. A commercial 1-meter LED strip was wrapped around the jar, followed by a layer of aluminium foil and cotton for insulation (Figure S2).

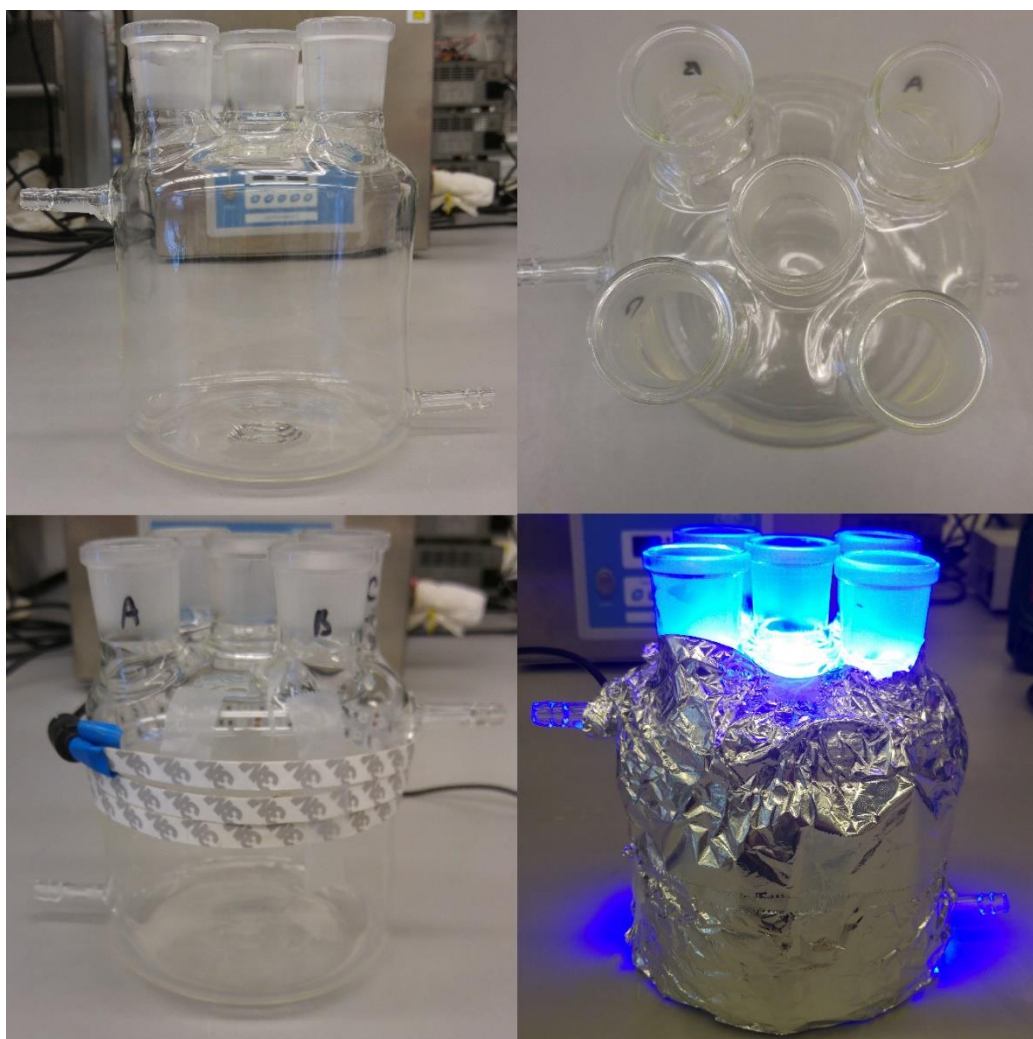


Figure S2: Photoreactor used for temperature-controlled reactions - pictures taken at different stages of the set-up assembly.

Each of the joints could be used to fit a standard 16 mm or 25 mm diameter Schlenk tube with a Teflon adaptor (Figure S3).



Figure S3: Teflon adaptors to use Schlenk tubes in the photoreactor.

An inlet and an outlet allow the circulation of liquid from a Huber Minichiller 300 inside the jar. This setup allows to perform reactions at temperatures ranging from -20 °C to 80 °C with accurate control of the reaction temperature ($\pm 1^\circ\text{C}$, Figure S4).

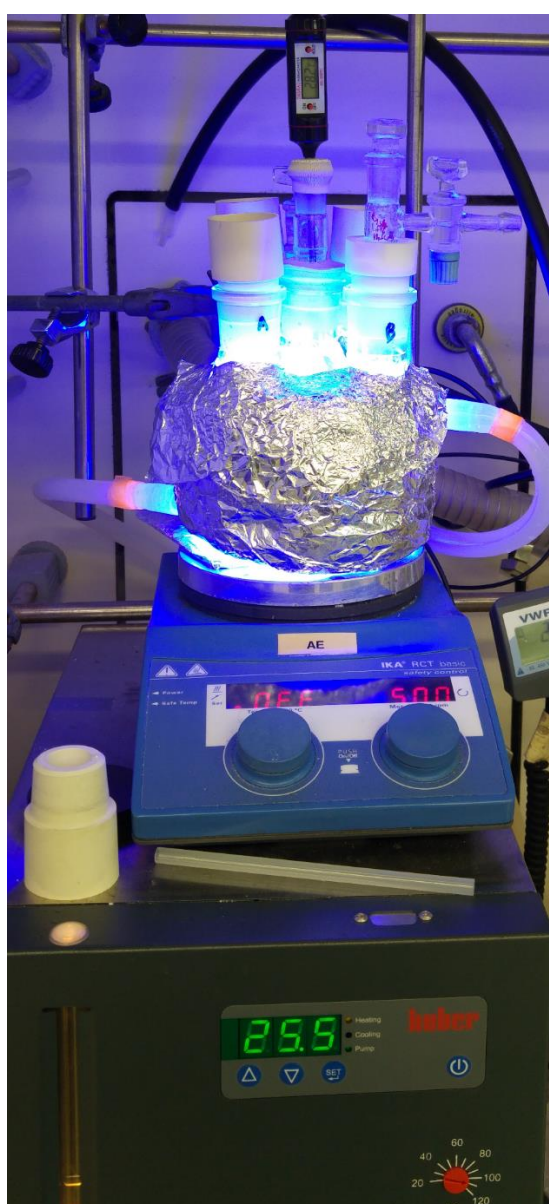


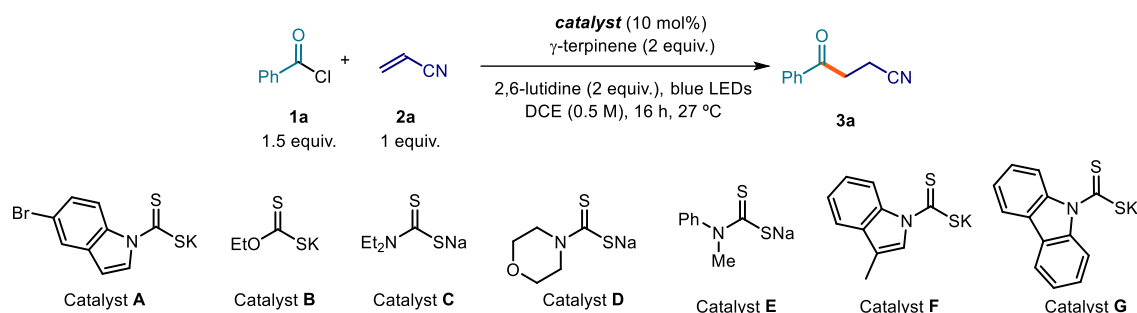
Figure S4: Fully assembled controlled temperature photoreactor in operation.

In order to maintain consistent illumination between different experiments, only the four external positions were used to perform reactions. The central position was used to monitor the

temperature inside a Schlenk tube identical to those used to perform reactions, ensuring that the reaction mixtures are at the desired temperature.

C1.2 Optimization Studies

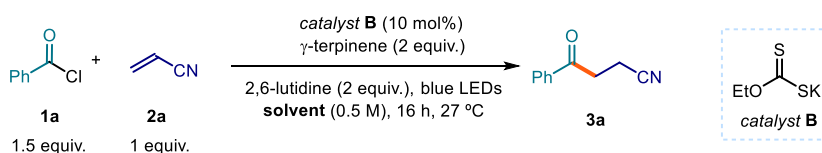
Table S1. Screening of the catalysts



entry	catalyst	wavelength (nm)	NMR yield (%)
1	A	465	<5
2	B	465	30
3	B	400	18
4	C	465	23
5	C	400	20
6	D	465	18
7	E	465	<5
8	F	465	<5
9	G	465	<5

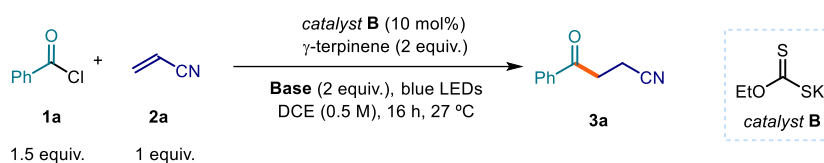
All reaction performed on 0.5 mmol scale; yield determined by ^1H NMR analysis of the crude reaction mixture using trichloroethylene as the internal standard.

Table S2. Screening of the solvents



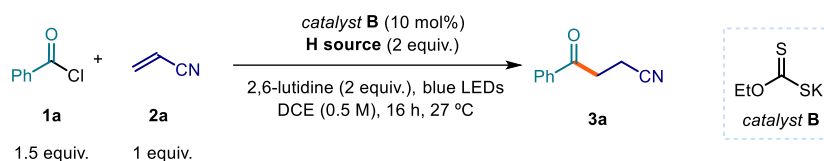
entry	solvent	NMR yield (%)
1	MeCN	20
2	AcOEt	20
3	DCE	30
4	MTBE	13
5	Toluene	20
6	Dioxane	20
7	DMF	0
8	Acetone	16
9	DCM	25
10	PhCl	16

All reaction performed on 0.5 mmol scale; yield determined by ^1H NMR analysis of the crude reaction mixture using trichloroethylene as the internal standard.

Table S3. Screening of the bases

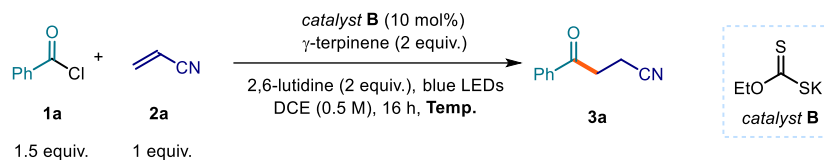
entry	base	NMR yield (%)
1	2,6-lutidine	30
2	2,4,6-collidine	16
3	N-Methylmorpholine	0
4	N-Methylimidazole	0
5	K ₂ CO ₃	24
6	NaOAc	19
7	KH ₂ PO ₄	16
8	NaHCO ₃	16

All reaction performed on 0.5 mmol scale; yield determined by ¹H NMR analysis of the crude reaction mixture using trichloroethylene as the internal standard.

Table S4. Screening of the H-atom source

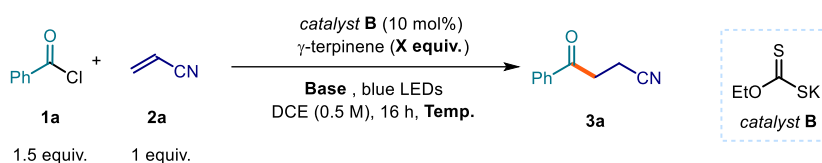
entry	H source	NMR yield (%)
1	γ -terpinene	30
2	Hantzsch ester	25
3	1,4-Cyclohexadiene	9
10	2,5-Dihydrofuran	5

All reaction performed on 0.5 mmol scale; yield determined by ¹H NMR analysis of the crude reaction mixture using trichloroethylene as the internal standard.

Table S5. Screening of the temperature

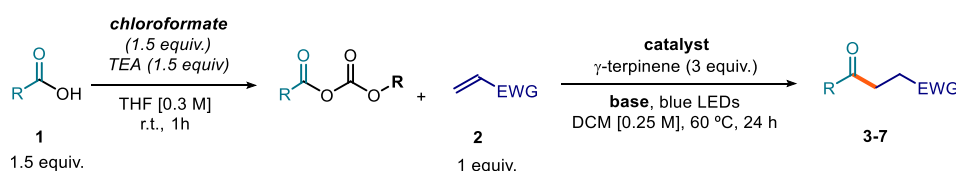
entry	temperature	NMR yield (%)
1	27 °C	30
2	40 °C	35
3	60 °C	50

All reaction performed on 0.5 mmol scale; yield determined by ¹H NMR analysis of the crude reaction mixture using trichloroethylene as the internal standard.

Table S6. Final cycle of optimization

entry	Base	Base (equiv.)	γ -terpinene (equiv.)	Other variations	yield (%)
1	2,6-lutidine	2	2	-	50
2	2,6-lutidine	1.2	2	-	50
3	Na ₂ CO ₃	1.2	2	-	66
4	Na ₂ HPO ₄	1.2	2	-	20
5	Na ₃ PO ₄	1.2	2	-	66
6	Na ₃ PO ₄	1.2	2	20 mol% catalyst	45
7	Na ₃ PO ₄	1.2	3	-	55
8	Na ₃ PO ₄	2	2	-	84
9	Na ₃ PO ₄	2	2	5 mol% catalyst	62
10	Na ₃ PO ₄	2	2	DCM [0.25 M]	85
11	Na ₃ PO ₄	2	2	Vinyl sulfone as acceptor & DCM [0.25 M]	57
12	Na ₃ PO ₄	2	3	Vinyl sulfone as acceptor & DCM [0.25 M]	85

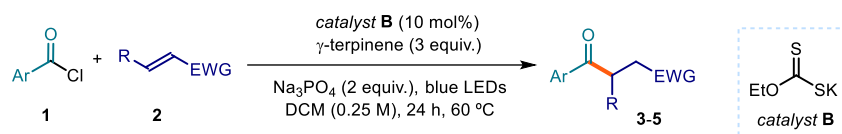
All reaction performed on 0.5 mmol scale; yield determined by ¹H NMR analysis of the crude reaction mixture using trichloroethylene as the internal standard.

Table S7. Anhydride method - optimization

entry	Acid	Chloroformate	Base	Catalyst	Acceptor	NMR yield (%)
1	Benzoic	Ethyl	Na ₃ PO ₄	10 mol% B	Acrylonitrile	50
2	Benzoic	Ethyl	Na ₃ PO ₄	10 mol% C	Acrylonitrile	55
3	Benzoic	Ethyl	Na ₃ PO ₄	20 mol% B	Acrylonitrile	60
4	Benzoic	Ethyl	Na ₃ PO ₄ (1 equiv.)	20 mol% B	Acrylonitrile	69
5	Benzoic	Ethyl	-	20 mol% B	Acrylonitrile	71
6	Benzoic	Methyl	-	20 mol% B	Acrylonitrile	61
7	Benzoic	Isobutyl	-	20 mol% B	Acrylonitrile	15
8	Cyclohexyl	Ethyl	-	20 mol% B	Vinyl sulfone	64
9	Cyclohexyl	Ethyl	-	20 mol% C	Vinyl sulfone	42
10	Cyclohexyl	Ethyl	-	30 mol% B	Vinyl sulfone	83
11	Cyclohexyl	Ethyl	-	50 mol% B	Vinyl sulfone	82

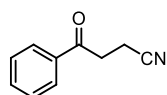
All reaction performed on 0.5 mmol scale; yield determined by ¹H NMR analysis of the crude reaction mixture using trichloroethylene as the internal standard.

C1.3 General Procedure A



In an oven dried tube of 15 mL (16 mm \times 125 mm) with a Teflon septum screw cap, potassium ethyl xanthogenate **B** (8 mg, 0.05 mmol, 0.1 equiv.), sodium phosphate (164 mg, 1.00 mmol, 2 equiv.), acyl chloride **1** (0.75 mmol, 1.5 equiv.) and the electron-poor olefin **2** (0.5 mmol, 1 equiv., *if solid*), were dissolved in DCM (2 mL, HPLC grade). Then, γ -terpinene (240 μL , 1.5 mmol, 3 equiv.) was added. The resulting yellow mixture was degassed with argon sparging for 60 seconds. When the electron-poor olefin **2** is *liquid*, it was added via syringe after the argon sparging. The reaction vessel was then placed in the temperature-controlled photoreactor (Figures S2-4) set at 60 °C (60–61 °C measured in the central well) and irradiated for 16 hours upon stirring, if not otherwise specified. Then, the solvent was evaporated and the residue purified by column chromatography to afford the corresponding product in the stated yield with >95% purity according to ^1H NMR analysis.

C1.4 Characterization of Products

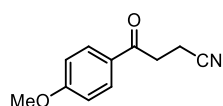


4-Oxo-4-phenylbutanenitrile (3a): Synthesized according to the general procedure A using benzoyl chloride (87 μL , 0.75 mmol, 1.5 equiv.) and acrylonitrile (33 μL , 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **3a** (65 mg, 82% yield) as a white solid.

^1H NMR (500 MHz, CDCl_3) δ 7.98 – 7.93 (m, 2H), 7.64 – 7.59 (m, 1H), 7.53 – 7.47 (m, 2H), 3.41 – 3.35 (m, 2H), 2.80 – 2.75 (m, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 195.4, 135.7, 134.0, 129.0, 128.1, 119.3, 34.4, 11.9

Matching reported literature data.⁹

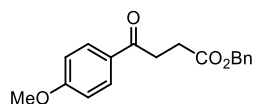


4-(4-methoxyphenyl)-4-oxobutanenitrile (3b): Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μL , 0.75 mmol, 1.5 equiv.) and acrylonitrile (33 μL , 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (15% AcOEt in hexanes as eluent) to afford **3b** (80 mg, 84% yield) as a white solid.

^1H NMR (500 MHz, CDCl_3) δ 7.92 (app d, J = 8.9 Hz, 2H), 6.95 (app d, J = 9.0 Hz, 2H), 3.87 (s, 3H), 3.32 (t, J = 7.3 Hz, 2H), 2.75 (t, J = 7.3 Hz, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 193.9, 164.2, 130.4, 128.8, 119.5, 114.1, 55.7, 34.0, 12.0

Matching reported literature data.⁹

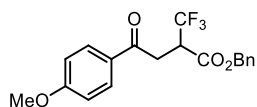


Benzyl 4-(4-methoxyphenyl)-4-oxobutanoate (3c): Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μL , 0.75 mmol, 1.5 equiv.) and benzyl acrylate (77 μL , 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (15% AcOEt in hexanes as eluent), followed by a second purification (AcOEt/hexanes/toluene 1:6:6 as eluent) to afford **3c** (96 mg, 64% yield) as a yellow solid.

^1H NMR (500 MHz, CDCl_3) δ 7.99 – 7.94 (m, 2H), 7.38 7.29 (m, 5H), 6.96 – 6.91 (m, 2H), 5.15 (s, 2H), 3.87 (s, 3H), 3.28 (t, J = 6.7 Hz, 2H), 2.81 (t, J = 6.7 Hz, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 196.6, 173.0, 163.7, 136.1, 130.4, 129.8, 128.6, 128.3 (2C overlapping), 113.9, 66.6, 55.6, 33.1, 28.5.

HRMS (ESI pos): calculated for $\text{C}_{18}\text{H}_{18}\text{NaO}_4$ ($\text{M}+\text{Na}^+$): 321.1097, found: 321.1091.



Benzyl 4-(4-methoxyphenyl)-4-oxo-2-(trifluoromethyl)butanoate (3d):

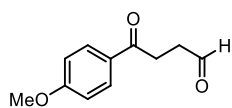
Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μL , 0.75 mmol, 1.5 equiv.) and benzyl 2-(trifluoromethyl)acrylate (115 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (10% AcOEt in hexanes as eluent). In order to remove traces of *p*-anisaldehyde formed as a byproduct during the reaction, after the chromatographic purification and solvent removal, the mixture was dissolved in 1.5 mL of MeOH, and 7.5 mL of saturated NaHSO_3 (aq) were added. Subsequently, the mixture was stirred for 30 s, diluted with 7.5 mL of H_2O , and extracted with 7.5 mL of 10% AcOEt in hexanes. The organic layer was dried with MgSO_4 , filtered, and concentrated in vacuo¹⁰ to afford **3d** (150 mg, 82% yield) as a colorless oil.

^1H NMR (400 MHz, CDCl_3) δ 7.99 – 7.91 (m, 2H), 7.42 – 7.29 (m, 5H), 6.99 – 6.91 (m, 2H), 5.24 (dd, J = 18.3; 12.3 Hz, 2H), 4.01 – 3.89 (m, 1H), 3.87 (s, 3H), 3.78 (dd, J = 17.7, 10.8 Hz, 1H), 3.32 (dd, J = 17.7, 3.1 Hz, 1H).

^{13}C NMR (100 MHz, CDCl_3) δ 193.8, 166.9 (q, J = 2.9 Hz), 164.2, 135.2, 130.6 (2CH overlapping), 128.9, 128.7, 128.5, 128.1, 125.0 (q, J = 280.4 Hz), 114.0, 67.9, 55.6, 45.9 (q, J = 27.9 Hz), 34.9 (d, J = 1.4 Hz).

^{19}F NMR (376 MHz, CDCl_3 , proton decoupled) δ -67.51 (s, 3F).

HRMS (ESI pos): calculated for $\text{C}_{19}\text{H}_{17}\text{F}_3\text{NaO}_4$ ($\text{M}+\text{Na}^+$): 389.0971, found: 389.0978.

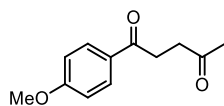


4-(4-Methoxyphenyl)-4-oxobutanal (3e): Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μL , 0.75 mmol, 1.5 equiv.) and acrolein (33 μL , 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent), followed by a second purification (5:47:48 of $\text{Et}_2\text{O}/\text{DCM}/\text{hexanes}$ as eluent) to afford **3e** (50 mg, 52% yield) as a yellow oil.

^1H NMR (400 MHz, CDCl_3) δ 9.89 (s, 1H), 7.99 – 7.91 (m, 2H), 6.96 – 6.89 (m, 2H), 3.85 (s, 3H), 3.27 (app t, J = 6.4 Hz, 2H), 2.89 (app t, J = 6.5 Hz, 2H).

^{13}C NMR (100 MHz, CDCl_3) δ 201.0, 196.4, 163.7, 130.4, 129.6, 113.9, 55.6, 37.8, 30.8

Matching reported literature data.¹¹



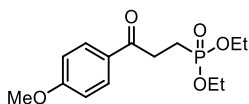
1-(4-Methoxyphenyl)pentane-1,4-dione (3f): Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μL , 0.75 mmol, 1.5 equiv.) and methyl vinyl ketone (41 μL , 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent), followed by a second one (10:45:45 of $\text{Et}_2\text{O}/\text{DCM}/\text{Hexanes}$ as eluent) to afford **3f** (49 mg, 48% yield) white solid.

^1H NMR (400 MHz, CDCl_3) δ 7.98 – 7.91 (m, 2H), 6.95–6.89 (m, 2H), 3.85 (s, 3H), 3.25 – 3.18 (m, 2H), 2.88 – 2.82 (m, 2H), 2.24 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3) δ 207.6, 197.1, 164.6, 130.4, 129.9, 113.8, 55.6, 37.2, 32.2, 30.2.

Matching reported literature data.⁹

Matching reported literature data.⁹



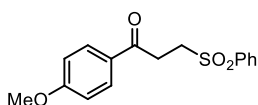
Diethyl (3-(4-methoxyphenyl)-3-oxopropyl)phosphonate (3g):

Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μL , 0.75 mmol, 1.5 equiv.) and diethyl vinylphosphonate (77 μL , 0.5 mmol, 1 equiv.). Irradiations time: 24 hours. The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **3g** (81 mg, 54% yield) as a white solid.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.91 (app d, $J = 8.8$ Hz, 2H), 6.89 (app d, $J = 8.9$ Hz, 2H), 4.20 – 3.96 (m, 4H), 3.82 (s, 3H), 3.28 – 3.12 (m, 2H), 2.22 – 2.04 (m, 2H), 1.28 (t, $J = 7.0$ Hz, 6H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 196.0 (d, $J = 16.2$ Hz), 163.7, 130.3, 129.4, 113.8, 61.7 (d, $J = 6.6$ Hz), 55.5, 31.3 (d, $J = 2.9$ Hz), 19.9 (d, $J = 144.3$ Hz), 6.5 (d, $J = 6.5$ Hz).

Matching reported literature data.¹²



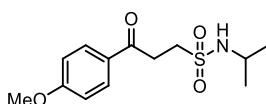
1-(4-Methoxyphenyl)-3-(phenylsulfonyl)propan-1-one (3h):

Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μL , 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). Reaction time: 24 hours. The crude mixture was purified by flash column chromatography on silica gel (25% AcOEt in hexanes as eluent), followed by a second one (20% AcOEt in hexanes as eluent) to afford **3h** (128 mg, 84% yield) as a white solid.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.96 – 7.93 (m, 2H), 7.91 – 7.87 (m, 2H), 7.69 – 7.63 (m, 1H), 7.60 – 7.54 (m, 2H), 6.95 – 6.90 (m, 2H), 3.86 (s, 3H), 3.57 – 3.51 (m, 2H), 3.46 – 3.40 (m, 2H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 193.9, 164.1, 139.3, 134.0, 130.5, 129.5, 129.0, 128.1, 114.1, 55.7, 51.3, 31.0.

Matching reported literature data.¹³



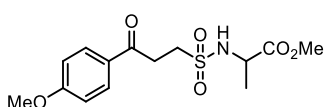
N-Isopropyl-3-(4-methoxyphenyl)-3-oxopropane-1-sulfonamide (3i):

Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μL , 0.75 mmol, 1.5 equiv.) and N-isopropylethanesulfonamide (75 mg, 0.5 mmol, 1 equiv.). Reaction time: 24 hours. The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **3i** (98 mg, 69% yield) as a white solid.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.98 – 7.91 (m, 2H), 6.97 – 6.90 (m, 2H), 4.35 (bs, 1H, NH), 3.87 (s, 3H), 3.74 – 3.58 (m, 1H), 3.54 – 3.38 (m, 4H), 1.24 (d, $J = 6.4$ Hz, 6H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 194.7, 164.1, 130.6, 129.2, 114.1, 55.7, 48.7, 46.5, 32.7, 24.4.

HRMS (ESI neg): calculated for $\text{C}_{13}\text{H}_{18}\text{NO}_4\text{S}$ (M⁻): 284.0962, found 284.0974.

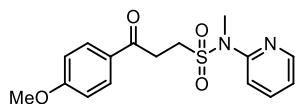


Methyl ((3-(4-methoxyphenyl)-3-oxopropyl)sulfonyl)alaninate (3j):

Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μL , 0.75 mmol, 1.5 equiv.) and methyl (vinylsulfonyl)alaninate (97 mg, 0.5 mmol, 1 equiv.). Reaction time: 24 hours. The crude mixture was purified by flash column chromatography on silica gel (35% AcOEt in hexanes as eluent), followed by a second one (20:40:40 of AcOEt/DCM/Hexanes as eluent) to afford **3j** (83 mg, 50% yield) as a colorless oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.97 – 7.89 (m, 2H), 6.96 – 6.89 (m, 2H), 5.29 (d, $J = 8.4$ Hz, 2H), 4.25 – 4.14 (m, 1H), 3.85 (s, 3H), 3.73 (s, 3H), 3.53 – 3.40 (m, 4H), 1.45 (d, $J = 7.15$ Hz, 3H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 194.5, 173.2, 164.0, 130.5, 129.1, 114.0, 55.6, 52.9, 51.7, 48.4, 32.2, 19.9. **HRMS (ESI pos):** calculated for $\text{C}_{14}\text{H}_{19}\text{NNaO}_6\text{S}$ (M+Na⁺): 352.0825, found 352.0814.



3-(4-Methoxyphenyl)-N-methyl-3-oxo-N-(pyridin-2-yl)propane-

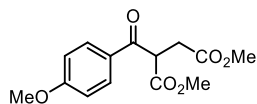
1-sulfonamide (3k): Synthesized according to the general procedure

A using 4-methoxybenzoyl chloride (102 μ L, 0.75 mmol, 1.5 equiv.) and N-methyl-N-(pyridin-2-yl)ethanesulfonamide (99 mg, 0.5 mmol, 1 equiv.). Reaction time: 24 hours. The crude mixture was purified by flash column chromatography on silica gel (20:20:60 of AcOEt/DCM/Hexanes as eluent), followed by a second one (20:20:60 of Et₂O/DCM/Hexanes as eluent) to afford **3k** (75 mg, 45% yield) as a yellowish oil.

¹H NMR (400 MHz, CDCl₃) δ 8.41 – 8.34 (m, 1H), 7.92 – 7.84 (m, 2H), 7.72 – 7.65 (m, 1H), 7.42 (app d, J = 8.3 Hz, 1H), 7.15 – 7.08 (m, 1H), 6.95 – 6.87 (m, 2H), 3.85 (s, 3H), 3.72 – 3.64 (m, 2H), 3.45 (s, 3H), 3.46 – 3.39 (m, 2H).

¹³C NMR (100 MHz, CDCl₃) δ 194.2, 164.0, 154.0, 148.3, 138.2, 130.5, 129.1, 121.0, 118.6, 114.0, 55.6, 46.3, 35.9, 31.8.

HRMS (ESI pos): calculated for C₁₆H₁₉N₂O₄S (M+H⁺): 335.1060, found 335.1043.



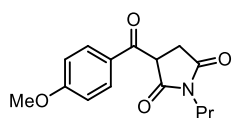
Dimethyl 2-(4-methoxybenzoyl)succinate (3l): Synthesized according

to the general procedure A using 4-methoxybenzoyl chloride (102 μ L, 0.75 mmol, 1.5 equiv.) and dimethyl fumarate (72 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **3l** (122 mg, 87% yield) as a colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 8.03 – 7.98 (m, 2H), 6.96 – 6.92 (m, 2H), 4.82 (t, J = 7.1 Hz, 1H), 3.85 (s, 3H), 3.66 (s, 3H), 3.65 (s, 3H), 3.09 – 2.97 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 192.3, 171.9, 169.5, 164.2, 131.4, 128.8, 114.0, 129.0, 128.1, 114.1, 55.6, 52.8, 52.1, 33.2.

Matching reported literature data.¹⁴



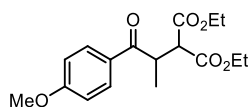
3-(4-Methoxybenzoyl)-1-propylpyrrolidine-2,5-dione (3m):

Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μ L, 0.75 mmol, 1.5 equiv.) and 1-propyl-1H-pyrrole-2,5-dione (69.6 μ L, 0.5 mmol, 1 equiv.). Chromatography on silica gel (20% AcOEt in hexanes as eluent) could not remove byproduct completely. Therefore, a further purification by semipreparative HPLC (Column SunFire C18, 60:40 Methanol/Water 6 min, up to 100% Methanol 1 min, 100% Methanol 4 min, 1 mL/min) was performed to obtain an analytical amount of the isolated product as a white solid. NMR yield (Trichloroethylene was used as internal standard): 53%.

¹H NMR (500 MHz, CDCl₃) δ 8.12-8.07 (m, 2H), 7.02 – 6.97 (m, 2H), 4.78 (dd, J = 8.3, 3.9 Hz, 1H), 3.90 (s, 3H), 3.47 (t, J = 7.3 Hz, 2H), 3.37 (dd, J = 18.1, 3.8 Hz, 1H), 2.82 (dd, J = 18.1, 8.8 Hz, 1H), 1.64 – 1.54 (m, 2H, overlapping with water peak), 0.87 (t, J = 7.4 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 190.9, 176.1, 173.4, 164.7, 132.4, 128.6, 114.2, 55.7, 48.2, 41.0, 31.8, 21.0, 11.3

HRMS (ESI pos): calculated for C₁₅H₁₇NNaO₄ (M+Na⁺): 298.1050, found 298.1056.



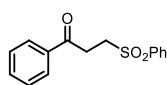
Diethyl 2-(1-(4-methoxyphenyl)-1-oxopropan-2-yl)malonate (3n):

Synthesized according to the general procedure A using 4-methoxybenzoyl chloride (102 μ L, 0.75 mmol, 1.5 equiv.) and diethyl 2-ethylidenemalonate (93 μ L, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (10% AcOEt in hexanes as eluent) to afford **3n** (60 mg, 37% yield) as a colorless oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.03 – 7.96 (m, 2H), 6.98 – 6.93 (m, 2H), 4.31 – 4.21 (m, 2H) 4.20 – 4.02 (m, 3H), 3.97 (d, J = 10.8 Hz, 1H), 3.87 (s, 3H), 1.31 (t, J = 7.0 Hz, 3H), 1.19 (d, J = 7.0 Hz, 3H), 1.16 (t, J = 7.1 Hz, 3H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 200.2, 169.0, 168.5, 163.8, 131.0, 128.6, 114.0, 61.7, 55.6, 55.1, 40.3, 16.25, 14.3, 14.0.

HRMS (ESI pos): calculated for $\text{C}_{17}\text{H}_{22}\text{NaO}_6$ ($\text{M}+\text{Na}^+$): 345.1309, found 345.1306.



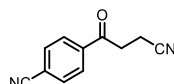
1-Phenyl-3-(phenylsulfonyl)propan-1-one (4a): Synthesized according to the general procedure A using benzoyl chloride (87 μ L, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified

by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **4a** (112 mg, 82% yield) as a white solid.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.99 – 7.88 (m, 4H), 7.70 – 7.63 (m, 1H), 7.63 – 7.54 (m, 3H), 7.51 – 7.43 (m, 2H), 3.60 – 3.53 (m, 2H), 3.53 – 3.46 (m, 2H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 195.5, 139.2, 135.9, 134.1, 133.9, 129.6, 128.9, 128.2, 128.1, 51.1, 31.5.

Matching reported literature data.¹³



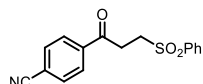
4-(3-Cyanopropanoyl)benzotrile (4b): Synthesized according to the general procedure A using 4-cyanobenzoyl chloride (124 mg, 0.75 mmol, 1.5 equiv.) and acrylonitrile (33 μ L, 0.5 mmol, 1 equiv.). In this case the reaction

was irradiated for 60 hours. The crude mixture was purified by flash column chromatography on silica gel (15% AcOEt in hexanes as eluent) to afford **4b** (92 mg, 45% yield) as a white solid.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.04 (app d, J = 8.2 Hz, 2H), 7.80 (app d, J = 8.2 Hz, 2H), 3.39 (t, J = 6.9 Hz, 2H), 2.79 (t, J = 7.0 Hz, 2H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 194.3, 138.5, 132.8, 128.6, 118.8, 117.7, 117.3, 34.7, 11.8.

Matching reported literature data.⁹



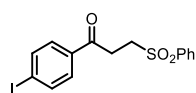
4-(3-(Phenylsulfonyl)propanoyl)benzotrile (4c): Synthesized according to the general procedure A using 4-cyanobenzoyl chloride (124 mg, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). In

this case the reaction was irradiated for 60 hours. The crude mixture was purified by flash column chromatography on silica gel (25% AcOEt in hexanes as eluent) to afford **4c** (79 mg, 53% yield) as a white solid.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.02 (app d, J = 8.5 Hz, 2H), 7.95 (app d, J = 7.3 Hz, 2H), 7.78 (app d, J = 8.4 Hz, 2H), 7.69 (app t, J = 7.4 Hz, 1H), 7.59 (app t, J = 7.7 Hz, 2H), 3.60 – 3.48 (m, 4H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 194.4, 139.1, 138.8, 134.2, 132.8, 129.6, 128.6, 128.1, 117.8, 117.2, 50.9, 31.8.

HRMS (ESI pos): calculated for $\text{C}_{16}\text{H}_{13}\text{NNaO}_3\text{S}$ ($\text{M}+\text{Na}^+$): 322.0508, found 322.0505.



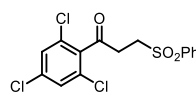
1-(4-Iodophenyl)-3-(phenylsulfonyl)propan-1-one (4d): Synthesized according to the general procedure A using 4-iodobenzoyl chloride (200 mg, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.).

In this case the reaction was irradiated for 60 hours. The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **4d** (130 mg, 65% yield) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 7.99 – 7.92 (m, 2H), 7.87 – 7.82 (m, 2H), 7.70 – 7.65 (m, 1H), 7.65 – 7.61 (m, 2H), 7.61 – 7.56 (m, 2H), 3.58 – 3.51 (m, 2H) 3.49 – 3.42 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 194.9, 139.2, 138.3, 135.2, 134.1, 129.6, 129.5, 128.1, 102.1, 51.0, 31.4

HRMS (ESI pos): calculated for C₁₅H₁₃INaO₃S (M+Na⁺): 422.9522, found 422.9519.



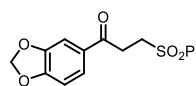
3-(Phenylsulfonyl)-1-(2,4,6-trichlorophenyl)propan-1-one (4e):

Synthesized according to the general procedure A using 2,4,6-trichlorobenzoyl chloride (183 mg, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). In this case the reaction was irradiated for 60 hours. The crude mixture was purified by flash column chromatography on silica gel (25% AcOEt in hexanes as eluent) followed by a second one (5:25:70 of AcOEt/DCM/Hexanes as eluent) to afford **4e** (95 mg, 50% yield) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 7.98 – 7.92 (m, 2H), 7.72 – 7.65 (m, 1H), 7.63 – 7.56 (m, 2H), 7.36 (s, 2H), 3.58 – 3.50 (m, 2H), 3.33 – 3.25 (m, 2H).

¹³C NMR (100 MHz, CDCl₃) δ 197.2, 138.9, 137.0, 136.6, 134.2, 131.2, 129.6, 128.5, 128.1, 50.1, 36.5

HRMS (ESI pos): calculated for C₁₅H₁₁Cl₃NaO₃S (M+Na⁺): 398.9387, found 398.9390.



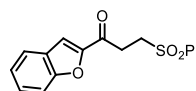
1-(Benzo[d][1,3]dioxol-5-yl)-3-(phenylsulfonyl)propan-1-one (4f):

Synthesized according to the general procedure A using benzo[d][1,3]dioxole-5-carbonyl chloride (138 mg, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). Reaction time: 24 hours. The crude mixture was purified by flash column chromatography on silica gel (25% AcOEt in hexanes as eluent) to afford **4f** (90 mg, 57% yield) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 7.96 – 7.92 (m, 2H), 7.67 (tt, *J* = 7.3, 1.8 Hz, 1H), 7.60 – 7.55 (m, 2H), 7.52 (dd, *J* = 8.1, 1.8 Hz, 1H), 7.36 (d, *J* = 1.7 Hz, 1H), 6.84 (d, *J* = 8.1 Hz, 1H), 6.04 (s, 2H), 3.57 – 3.50 (m, 2H), 3.44 – 3.37 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 193.5, 152.5, 148.5, 139.2, 134.0, 130.8, 129.5, 128.1, 124.7, 108.2, 107.8, 102.2, 51.3, 31.2.

HRMS (ESI pos): calculated for C₁₆H₁₄NaO₅S (M+Na⁺): 341.0454, found 341.0452.



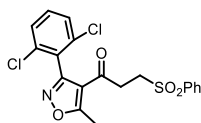
1-(Benzofuran-2-yl)-3-(phenylsulfonyl)propan-1-one (4g): Synthesized according to the general procedure A using benzofuran-2-carbonyl chloride (135 mg, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol,

1 equiv.). Reaction time: 24 hours. The crude mixture was purified by flash column chromatography on silica gel (25% AcOEt in hexanes as eluent) followed by a second one (20% AcOEt in hexanes as eluent) to afford **4g** (77 mg, 49% yield) as a yellowish solid.

¹H NMR (500 MHz, CDCl₃) δ 7.99 – 7.94 (m, 2H), 7.71 (app d, *J* = 7.8 Hz, 1H), 7.70 – 7.65 (m, 1H), 7.61 – 7.55 (app t, *J* = 8.0 Hz, 3H), 7.55 (s, 1H), 7.53 – 7.48 (m, 1H), 7.35 – 7.30 (m, 1H), 3.61 – 3.55 (m, 2H), 3.53 – 3.47 (m, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 186.6, 155.9, 151.6, 139.0, 134.2, 129.6, 128.9, 128.2, 126.9, 124.3, 123.6, 113.7, 112.6, 50.6, 31.8

HRMS (ESI pos): calculated for $\text{C}_{17}\text{H}_{14}\text{NaO}_4\text{S}$ ($\text{M}+\text{Na}^+$): 337.0505, found 337.0504.

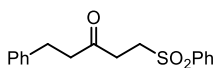


1-(3-(2,6-Dichlorophenyl)-5-methylisoxazol-4-yl)-3-(phenylsulfonyl)propan-1-one (4h): Synthesized according to the general procedure A using 3-(2,6-dichlorophenyl)-5-methylisoxazole-4-carbonyl chloride (218 mg, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). Reaction time: 24 hours. The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **4h** (75 mg, 35% yield) as a white solid.

^1H NMR (400 MHz, CDCl_3) δ 7.75 – 7.67 (m, 2H), 7.67 – 7.60 (m, 1H), 7.57 – 7.43 (m, 5H), 3.41 – 3.34 (m, 2H), 2.74 (s, 3H), 3.68 – 3.60 (m, 2H).

^{13}C NMR (126 MHz, CDCl_3) δ 189.0, 176.5, 157.2, 138.6, 135.6, 134.0, 132.2, 129.5, 128.6, 128.0, 127.9, 116.1, 50.1, 34.3, 14.2.

HRMS (ESI pos): calculated for $\text{C}_{19}\text{H}_{15}\text{Cl}_2\text{NNaO}_4\text{S}$ ($\text{M}+\text{Na}^+$): 445.9991, found 445.9992.

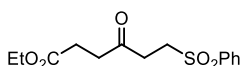


1-Phenyl-5-(phenylsulfonyl)pentan-3-one (5b): Synthesized according to the general procedure A using hydrocinnamoyl chloride (111 μL , 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). In this case the reaction was irradiated for 60 hours. The crude mixture was purified by two rounds of flash column chromatography on silica gel (25% AcOEt in hexanes as eluent): to afford **5b** (113 mg, 75% yield) as a white solid.

^1H NMR (400 MHz, CDCl_3) δ 7.92 – 7.87 (m, 2H), 7.71 – 7.63 (m, 1H), 7.61 – 7.53 (m, 2H), 7.31 – 7.23 (m, 2H), 7.22 – 7.16 (m, 1H), 7.16 – 7.10 (m, 2H), 3.42 – 3.33 (m, 2H), 2.92 – 2.82 (m, 4H), 2.81 – 2.72 (m, 2H).

^{13}C NMR (100MHz, CDCl_3) δ 205.3, 140.5, 139.1, 134.1, 129.5, 128.7, 128.4, 128.1, 126.5, 50.6, 44.4, 35.3, 29.7.

Matching reported literature data.¹⁵



Ethyl 4-oxo-6-(phenylsulfonyl)hexanoate (5c): Synthesized according to the general procedure A using ethyl 4-chloro-4-oxobutanoate (107 μL , 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). Reaction time: 24 hours. The crude mixture was purified by flash column chromatography on silica gel (33% AcOEt in hexanes as eluent). Product was then dissolved in DCM, washed 3 times with a solution of CuSO_4 (5% in water), dried with MgSO_4 and evaporated under reduced pressure to afford **5c** (112 mg, 75% yield) as a white solid.

^1H NMR (400 MHz, CDCl_3) δ 7.92 – 7.87 (m, 2H), 7.69 – 7.63 (m, 1H), 7.61 – 7.53 (m, 2H), 4.09 (q, $J = 7.1$ Hz, 2H), 3.42 – 3.35 (m, 2H), 2.98 – 2.91 (m, 2H), 2.75 – 2.69 (m, 2H), 2.59 – 2.52 (m, 2H), 1.22 (t, $J = 7.1$ Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 204.5, 172.5, 139.1, 134.1, 129.5, 128.1, 60.9, 50.6, 37.2, 35.3, 28.0, 14.2

HRMS (ESI pos): calculated for $\text{C}_{14}\text{H}_{18}\text{NaO}_5\text{S}$ ($\text{M}+\text{H}^+$): 321.0767, found 321.0765.

C2. Reaction of Aliphatic Acyl Chlorides

C2.1 Experimental Setup

Our 3D printed photoreactor consisted of a 9 cm diameter crystallizing dish with a 3D printed support of 6 positions, and a hole of 22 mm in the middle to allow ventilation. A commercial 1-meter LED strip was wrapped around the crystallizing dish. In order to control the temperature, a fan was used to cool down the reactor. Reaction temperature was measured, through a vial containing a thermometer, and it stayed between 35-40 °C (Figure S5). Each of the positions could be used to fit a standard 16 mm diameter vial with a Teflon screw cap.

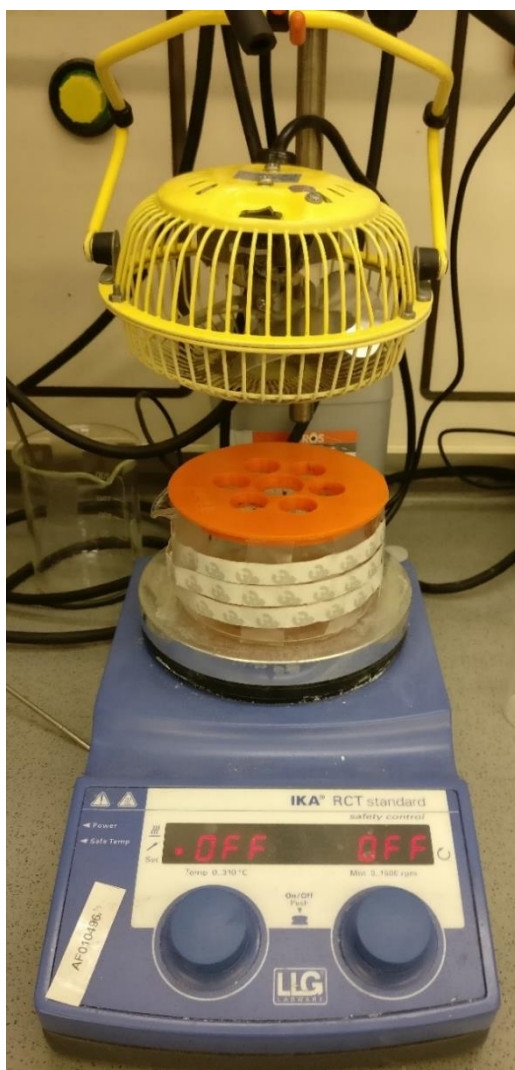


Figure S5: Photoreactor used for the reactions of aliphatic acyl chlorides.

Experiments at 465 nm were conducted using a 1m strip, 14.4W “LEDXON MODULAR 9009083 LED, SINGLE 5050” purchased from Farnell, catalog number 9009083. The emission spectrum of these LEDs was recorded (Figure S6).

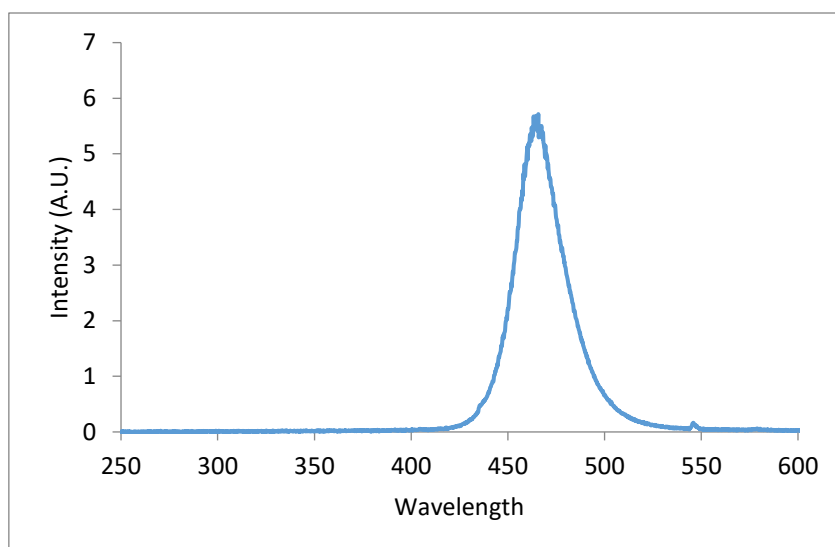
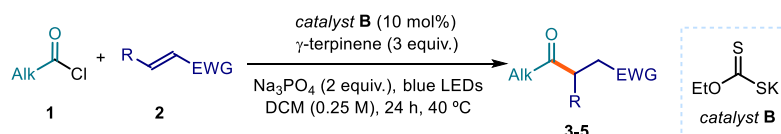


Figure S6: Emission spectrum of the 465 nm LED strip used in this study.

The emission maximum was determined as 465 nm with a spectral width of 30 nm (450-480 nm) at half peak intensity and a total spectral width of 120 nm (420-540 nm).

C2.2 General Procedure B



In an oven dried tube of 15 mL (16 mm × 125 mm) with a Teflon septum screw cap, potassium ethyl xanthogenate **B** (8 mg, 0.05 mmol, 0.1 equiv.), sodium phosphate (164 mg, 1.00 mmol, 2 equiv.), acyl chloride **1** (0.75 mmol, 1.5 equiv.) and the electron-poor olefin **2** (0.5 mmol, 1 equiv., *if solid*), were dissolved in DCM (2 mL, HPLC grade). Then, γ -terpinene (240 μ L, 1.5 mmol, 3 equiv.) was added. The resulting yellow mixture was degassed with argon sparging for 60 seconds. When the electron-poor olefin **2** is *liquid*, it was added via syringe after the argon sparging. The vial was then placed in the 3D printed support photoreactor (Figure S6) and irradiated under stirring for 24 hours, if not otherwise specified. After this, the solvent was evaporated and the residue purified by column chromatography to afford the corresponding product in the stated yield with >95% purity according to ^1H NMR analysis.

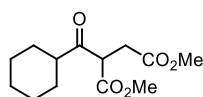
C2.3 Characterization of Products

4-(Phenylsulfonyl)butan-2-one (5a): Synthesized according to the general procedure B using acetyl chloride (53 μ L, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (gradient from 25% to 100% AcOEt in hexanes as eluent): to afford product **5a** (70 mg 66% yield) as a colorless oil.

^1H NMR (400 MHz, CDCl_3) δ 7.94 – 7.86 (m, 2H), 7.70 – 7.62 (m, 1H), 7.61 – 7.53 (m, 2H), 3.40 – 3.33 (m, 2H), 2.96 – 2.88 (m, 2H), 2.17 (s, 3H).

^{13}C NMR (100 MHz, CDCl_3) δ 203.8, 139.1, 134.0, 129.6, 128.1, 50.6, 36.0, 30.0

Matching reported literature data.¹⁶

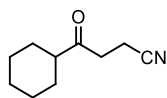


Dimethyl 2-(cyclohexanecarbonyl)succinate (5d): Synthesized according to general procedure B using cyclohexanecarbonyl chloride (100 μ L, 0.75 mmol, 1.5 equiv.) and dimethyl fumarate (72 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (10% AcOEt in hexanes as eluent) to afford **5d** (107 mg, 83% yield) as a white solid.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 4.14 (dd, $J = 8.1, 6.2$ Hz, 1H), 3.72 (s, 3H), 3.66 (s, 3H), 3.82 (s, 3H), 2.93 (dd, $J = 17.5, 8.1$ Hz, 1H), 2.81 (dd, $J = 17.6, 6.5$ Hz, 1H), 2.69 – 2.61 (m, 1H), 2.00 – 1.93 (m, 1H), 1.84 – 1.74 (m, 3H), 1.70 – 1.62 (m, 1H), 1.46 – 1.36 (m, 1H), 1.33 – 1.13 (m, 4H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 206.7, 171.9, 169.2, 52.8, 52.2, 52.1, 50.72, 32.4, 29.0, 28.1, 25.9, 25.8, 25.5.

Matching reported literature data.¹⁷



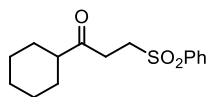
4-Cyclohexyl-4-oxobutanenitrile (5e): Synthesized according to the general procedure B using cyclohexanecarbonyl chloride (100 μ L, 0.75 mmol, 1.5 equiv.) and acrylonitrile (33 μ L, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% Et_2O in pentane as eluent) to afford **5e** (52 mg, 63% yield) as a colorless oil.

(For the gram scale procedure see paragraph C5, general procedure E)

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 2.85-2.80 (m, 2H); 2.60-2.54 (m, 2H); 2.36 (tt, $J = 11.3, 3.5$ Hz, 1H); 1.89-1.82 (m, 2H); 1.82-1.75 (m, 2H); 1.71-1.64 (m, 1H); 1.40-1.15 (m, 5H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 209.4, 119.3, 50.6, 35.9, 28.5, 25.8, 25.6, 11.6.

Matching reported literature data.¹⁷



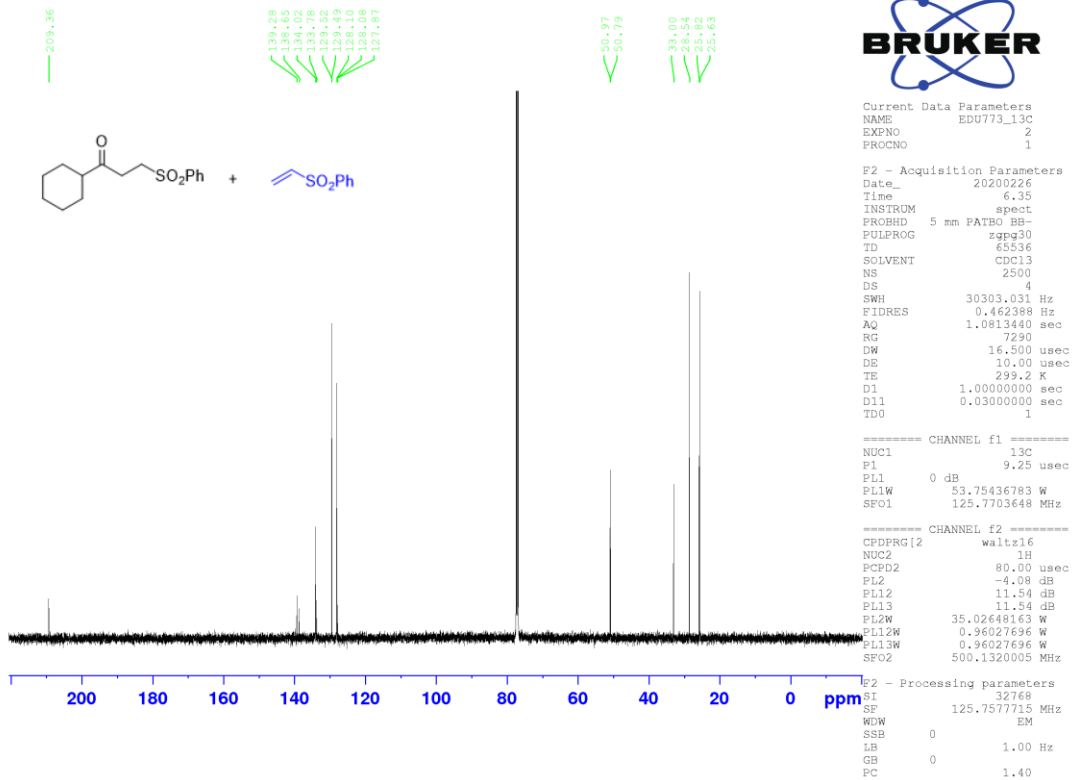
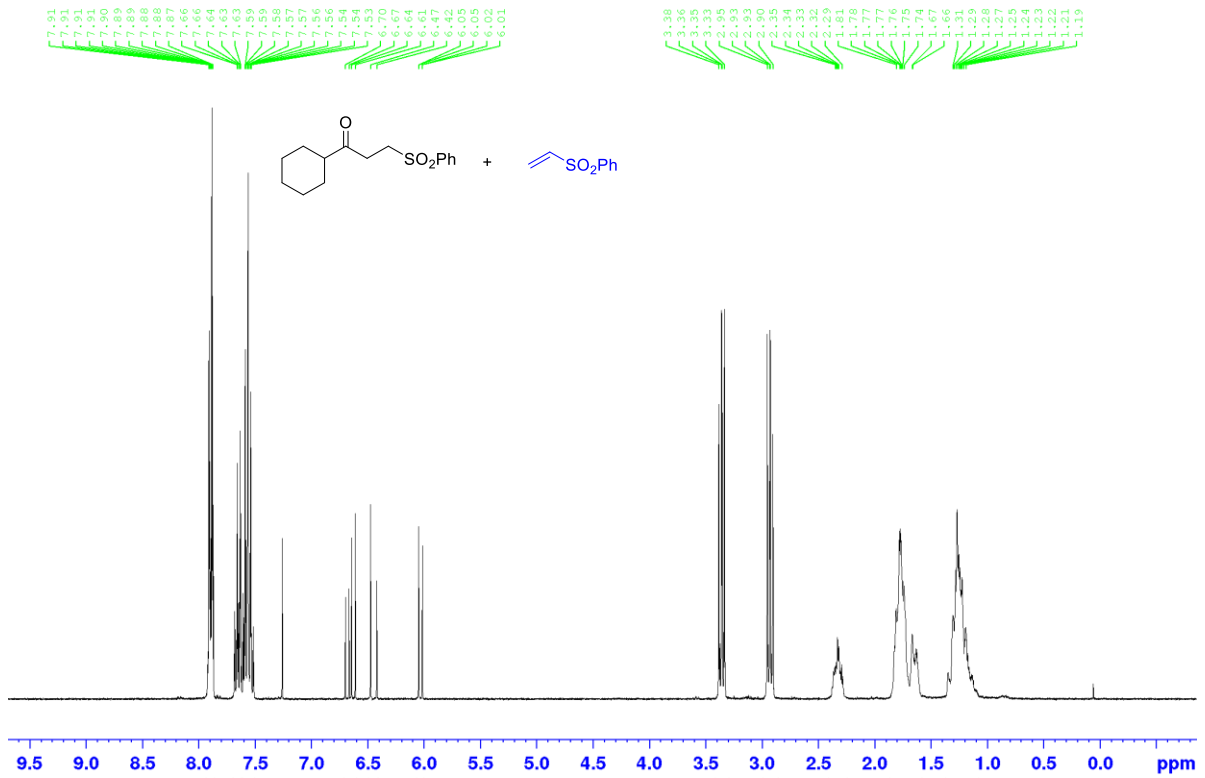
1-Cyclohexyl-3-(phenylsulfonyl)propan-1-one (5f): Synthesized according to the general procedure B using cyclohexanecarbonyl chloride (100 μ L, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone **2** (84 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **5f** (112 mg, 76% yield) as colorless oil.

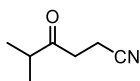
A purity of 85% weight was determined $^1\text{H NMR}$ analysis (mixture with phenyl vinyl sulfone, see Figure below). Corrected yield: 68%.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.93 – 7.86 (m, 2H), 7.69 – 7.62 (m, 1H), 7.62 – 7.50 (m, 2H), 3.39 – 3.32 (m, 2H), 2.97 – 2.89 (m, 2H), 2.39 – 2.25 (m, 1H), 1.88 – 1.58 (m, 5H), 1.38 – 1.08 (m, 5H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 193.9, 164.2, 130.4, 128.8, 119.5, 114.1, 55.7, 34.0, 12.0.

Matching reported literature data.¹⁸



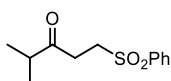


4-Cyclohexyl-4-oxobutanenitrile (5g): Synthesized according to the general procedure B using isobutanoyl chloride (100 μ L, 0.75 mmol, 1.5 equiv.) and acrylonitrile (33 μ L, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% Et₂O in pentane as eluent) to afford **5g** (39 mg, 62% yield) as a colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 2.83 (t, J = 7.1 Hz, 2H), 2.66 – 2.56 (m, 1H), 2.57 (t, J = 7.2 Hz, 2H), 1.12 (d, J = 7.0 Hz, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 210.0, 119.2, 40.7, 35.6, 18.2, 11.6

Matching reported literature data.¹⁹

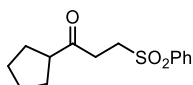


4-Methyl-1-(phenylsulfonyl)pentan-3-one (5h): Synthesized according to the general procedure B using isobutanoyl chloride (100 μ L, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent), followed by a second one (15% AcOEt in hexanes as eluent) to afford **5h** (61 mg, 51% yield) as a colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.94 – 7.88 (m, 2H), 7.69 – 7.64 (m, 1H), 7.61 – 7.54 (m, 2H), 3.40 – 3.35 (m, 2H), 3.00 – 2.93 (m, 2H), 2.66 – 2.55 (m, 1H), 1.08 (d, J = 7.0 Hz, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 210.0, 139.2, 134.0, 129.5, 128.1, 50.8, 41.1, 32.7, 18.3.

HRMS (ESI pos): calculated for C₁₂H₁₆NaO₃S (M+Na⁺): 263.0712, found 263.0714.

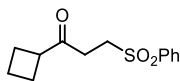


1-Cyclopentyl-3-(phenylsulfonyl)propan-1-one (5i): Synthesized according to the general procedure B using cyclopentanecarbonyl chloride (91 μ L, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **5i** (102 mg, 77% yield) colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.93 – 7.87 (m, 2H), 7.69 – 7.62 (m, 1H), 7.60 – 7.53 (m, 2H), 3.42 – 3.34 (m, 2H), 2.99 – 2.91 (m, 2H), 2.90 – 2.79 (m, 1H), 1.88 – 1.73 (m, 2H), 1.73 – 1.49 (m, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 208.5, 139.2, 134.0, 129.5, 128.1, 51.5, 50.8, 34.0, 29.0, 26.0.

HRMS (ESI pos): calculated for C₁₄H₁₈NaO₃S (M+Na⁺): 289.0869, found 289.0873.

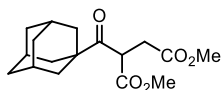


1-Cyclobutyl-3-(phenylsulfonyl)propan-1-one (5j): Synthesized according to the general procedure B using cyclobutanecarbonyl chloride (86 μ L, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **5j** (116 mg, 92% yield) as a colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.92 – 7.88 (m, 2H), 7.68 – 7.63 (m, 1H), 7.60 – 7.54 (m, 2H), 3.40 – 3.35 (m, 2H), 3.29 – 3.20 (m, 1H), 2.87 – 2.81 (m, 2H), 2.24 – 2.08 (m, 4H), 2.02 – 1.90 (m, 1H), 1.84 – 1.75 (m, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 207.1, 139.2, 134.1, 129.5, 128.1, 50.6, 45.4, 32.3, 24.5, 17.9

HRMS (ESI pos): calculated for C₁₃H₁₅NaO₃S (M+Na⁺): 275.0712, found 275.0716.



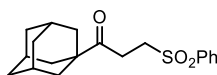
Dimethyl 2-((3r,5r,7r)-adamantane-1-carbonyl)succinate (5k):

Synthesized according to the general procedure B using 1-adamantanecarbonyl chloride (149 mg, 0.75 mmol, 1.5 equiv.) and dimethyl fumarate (72 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (5% AcOEt in hexanes as eluent) to afford **5k** (85 mg, 55% yield) as a colorless oil.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 4.37 (app t, $J = 7.2$ Hz, 1H), 3.67 (s, 3H), 3.64 (s, 3H), 2.79 (app d, $J = 7.2$ Hz, 2H), 2.03 (bs, 3H), 1.89 – 1.79 (m, 6H), 1.76 – 1.63 (m, 6H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 208.8, 171.7, 169.5, 52.7, 52.1, 47.7, 47.4, 38.0, 36.4, 33.5, 27.9

HRMS (ESI pos): calculated for $\text{C}_{17}\text{H}_{24}\text{NaO}_5$ ($\text{M}+\text{Na}^+$): 331.1516, found 331.1523.



1-((3r,5r,7r)-Adamantan-1-yl)-3-(phenylsulfonyl)propan-1-one (5l):

Synthesized according to the general procedure B using 1-adamantanecarbonyl chloride (149 mg, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (15% AcOEt in hexanes as eluent) to afford **5l** (165 mg, 95% yield) as a colorless oil.

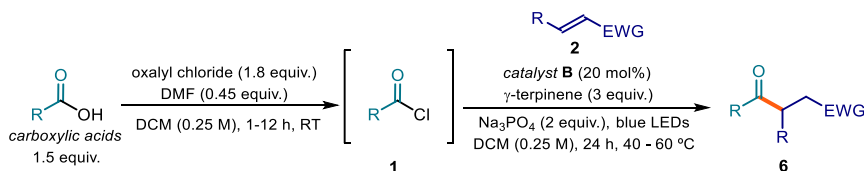
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.93 – 7.88 (m, 2H), 7.68 – 7.63 (m, 1H), 7.60 – 7.54 (m, 2H), 3.37 – 3.29 (m, 2H), 3.00-2.93 (m, 2H), 2.03 (bs, 3H), 1.79 – 1.70 (m, 9H), 1.66 (app d, $J = 12.2$ Hz, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 211.2, 139.3, 133.9, 129.4, 128.0, 50.9, 46.5, 38.3, 36.5, 29.1, 27.9.

HRMS (ESI pos): calculated for $\text{C}_{19}\text{H}_{25}\text{O}_3\text{S}$ ($\text{M}+\text{H}^+$): 333.1519, found 333.1519.

C3. Reaction of Carboxylic Acids through Acyl Chloride Formation

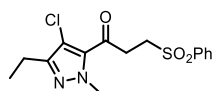
C3.1 General Procedure C:



In a round bottom flask, the carboxylic acid (0.75 mmol, 1.5 equiv) was dissolved in DCM (3 mL, HPLC grade). Then, oxalyl chloride (0.79 μL , 0.90 mmol, 1.8 equiv.) and DMF (17 μL , 0.22 mmol, 0.45 equiv.) were added at ambient temperature. The reaction was stirred at ambient temperature until complete consumption of the carboxylic acid was observed by TLC. After that, the solvent was evaporated to dryness under vacuum at ambient temperature to obtain the crude acyl chloride, which was used without further purification in the next step.

In an oven dried vial, with a Teflon septum screw cap, potassium ethyl xanthogenate **B** (16 mg, 0.10 mmol, 0.2 equiv.), sodium phosphate (164 mg, 1.00 mmol, 2 equiv.), and the electron-poor olefin **2** (0.5 mmol, 1 equiv.), were added. The crude acyl chloride was dissolved in DCM (2 mL, HPLC grade) and the solution was added to the vial, followed by γ -terpinene (240 μL , 1.5 mmol, 3 equiv.). The resulting yellow mixture was degassed via argon sparging for 60 seconds. If the electron-poor olefin **2** was *liquid*, it was added via syringe after the argon sparging. The vial was then placed in the correspondent reactor (depending on the temperature used for the reaction) and irradiated for 24 hours. The solvent was evaporated and the residue purified by column chromatography to afford the corresponding product in the stated yield with >95% purity according to $^1\text{H NMR}$ analysis.

C3.2 Characterization of Products

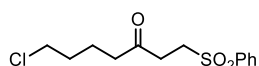


1-(4-Chloro-3-ethyl-1-methyl-1H-pyrazol-5-yl)-3-(phenylsulfonyl)propan-1-one (6a): Synthesized according to the general procedure C using 4-chloro-3-ethyl-1-methyl-1H-pyrazole-5-carboxylic acid (141 mg, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). Acyl chloride formation was complete after 2 hours. The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent) to afford **6a** (52 mg, 31% yield) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 7.98 – 7.92 (m, 2H), 7.71 – 7.64 (m, 1H), 7.62 – 7.54 (m, 2H), 3.98 (s, 3H), 3.57 – 3.51 (m, 2H), 3.49 – 3.42 (m, 2H), 2.63 (q, *J* = 7.6, 2H), 1.23 (t, *J* = 7.5, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 186.6, 150.4, 139.0, 134.6, 134.1, 129.5, 128.3, 112.7, 50.6, 41.6, 35.5, 19.2, 12.9.

HRMS (ESI pos): calculated for C₁₅H₁₈ClN₂O₃S (M+H⁺): 341.0721, found 341.0709.

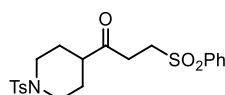


7-chloro-1-(phenylsulfonyl)heptan-3-one (6b): Synthesized according to the general procedure C using 5-chloropentanoic acid (72 μL, 0.75 mmol, 1.5 equiv.) and dimethyl fumarate (72 mg, 0.5 mmol, 1 equiv.). Acyl chloride formation was complete after 3 hours. The crude mixture was purified by flash column chromatography on silica gel (20% AcOEt in hexanes as eluent). The product was then dissolved in DCM, washed 3 times with a solution of CuSO₄ (5% in water), dried with MgSO₄ and evaporated under reduced pressure to afford **6b** (108 mg, 70% yield) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 7.92 – 7.87 (m, 2H), 7.69 – 7.63 (m, 1H), 7.60 – 7.54 (m, 2H), 3.53 – 3.47 (m, 2H), 3.41 – 3.35 (m, 2H), 2.92 – 2.86 (m, 2H), 2.50 – 2.44 (m, 2H), 1.78 – 1.64 (m, 4H).

¹³C NMR (100 MHz, CDCl₃) δ 205.5, 139.1, 134.1, 129.5, 128.1, 50.6, 44.6, 41.9, 35.0, 31.8, 20.9.

HRMS (ESI pos): calculated for C₁₃H₁₇ClNaO₃S (M+Na⁺): 311.0479, found 311.0477.

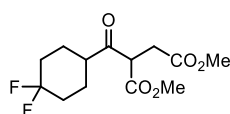


3-(Phenylsulfonyl)-1-(1-tosylpiperidin-4-yl)propan-1-one (6c): Synthesized according to the general procedure C using 1-tosylpiperidine-4-carboxylic acid (213 mg, 0.75 mmol, 1.5 equiv.) and dimethyl fumarate (72 mg, 0.5 mmol, 1 equiv.). Acyl chloride formation was complete after 3 hours. In this case the reaction was irradiated for 36 hours. The crude mixture was purified by flash column chromatography on silica gel (40% AcOEt in hexanes as eluent) to afford **6c** (164 mg, 75% yield) as a white solid.

¹H NMR (500 MHz, CDCl₃) δ 7.90 – 7.85 (m, 2H), 7.69 – 7.64 (m, 1H), 7.64 – 7.60 (m, 2H), 7.60 – 7.54 (m, 2H), 7.32 (app d, *J* = 7.9 Hz, 2H), 3.71 (dt, *J* = 12.2, 3.5 Hz, 2H), 3.35 (app t, *J* = 7.4 Hz, 2H), 2.91 (app t, *J* = 7.4 Hz, 2H), 2.43 (s, 3H), 2.38 (td, *J* = 11.7, 2.5 Hz, 2H), 2.33 – 2.24 (m, 1H), 1.93 – 1.84 (m, 2H), 1.75 – 1.63 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 206.9, 143.8, 139.1, 134.2, 133.2, 129.9, 129.6, 128.0, 127.8, 50.6, 47.6, 45.5, 33.0, 27.0, 21.7.

HRMS (ESI pos): calculated for C₂₁H₂₆NO₅S₂ (M+H⁺): 436.1247, found 436.1251.



dimethyl 2-(4,4-difluorocyclohexane-1-carbonyl)succinate (6d): Synthesized according to the general procedure C using 4,4-difluorocyclohexane-1-carboxylic acid (123 mg, 0.75 mmol, 1.5 equiv.) and dimethyl fumarate (72 mg, 0.5 mmol, 1 equiv.). Acyl chloride formation was complete after 1 hour. The crude mixture was purified by

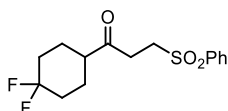
flash column chromatography on silica gel (20% Et₂O in hexanes as eluent) to afford **6d** (128 mg, 88% yield) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 4.16 (dd, *J* = 9.1, 5.3 Hz, 1H), 3.73 (s, 3H), 3.67 (s, 3H), 3.03 (dd, *J* = 17.6, 9.3 Hz, 1H), 2.82 (dd, *J* = 17.5, 5.3 Hz, 1H), 2.83 – 2.74 (m, 1H), 2.22 – 2.02 (m, 3H), 1.98 – 1.65 (m, 5H).

¹³C NMR (100 MHz, CDCl₃) δ 205.6, 171.9, 168.8, 122.7 (dd, *J* = 241.6, 240.9 Hz), 53.0, 52.2, 52.1, 48.0, 32.7 (dd, *J* = 25.0 Hz), 25.2 (d, *J* = 9.0 Hz) 24.4 (d, *J* = 8.6 Hz)

¹⁹F NMR (376 MHz, CDCl₃, proton decoupled) δ -93.75 (d, *J* = 237.2 Hz, 1F); -100.48 (d, *J* = 238.2 Hz, 1F).

HRMS (ESI pos): calculated for C₁₃H₁₈F₂NaO₅ (M+Na⁺): 315.1015, found 315.1017.



1-(4,4-Difluorocyclohexyl)-3-(phenylsulfonyl)propan-1-one (6e):

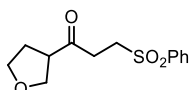
Synthesized according to the general procedure C using 4,4-difluorocyclohexane-1-carboxylic acid (123 mg, 0.75 mmol, 1.5 equiv.) and dimethyl fumarate (72 mg, 0.5 mmol, 1 equiv.). Acyl chloride formation was complete after 1 hour. The crude mixture was purified by flash column chromatography on silica gel (30% AcOEt in hexanes as eluent) to afford **6e** (112 mg, 71% yield) as a white solid.

¹H NMR (400 MHz, CDCl₃) δ 7.93 – 7.88 (m, 2H), 7.70 – 7.64 (m, 1H), 7.61 – 7.55 (m, 2H), 3.41 – 3.35 (m, 2H), 3.01 – 2.94 (m, 2H), 2.52 – 2.39 (m, 1H), 2.18 – 2.04 (m, 2H), 1.99 – 1.86 (m, 2H), 1.85 – 1.64 (m, 4H).

¹³C NMR (100 MHz, CDCl₃) δ 207.4, 139.2, 134.1, 129.6, 128.0, 122.5 (dd, *J* = 241.6, 240.7 Hz) 50.6, 48.1, 33.2, 32.7 (dd, *J* = 24.4, 24.2 Hz), 24.7 (d, *J* = 9.5 Hz).

¹⁹F NMR (376 MHz, CDCl₃, proton decoupled) δ -93.72 (dd, *J* = 237.6 Hz); -100.82 (d, *J* = 237.7 Hz).

HRMS (ESI pos): calculated for C₁₅H₁₈F₂NaO₃S (M+Na⁺): 339.0837, found 339.0840.



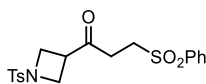
3-(Phenylsulfonyl)-1-(tetrahydrofuran-3-yl)propan-1-one (6f):

Synthesized according to the general procedure C using tetrahydrofuran-3-carboxylic acid (72 μL, 0.75 mmol, 1.5 equiv.) and dimethyl fumarate (72 mg, 0.5 mmol, 1 equiv.). Acyl chloride formation was complete after 1 hour. The crude mixture was purified by flash column chromatography on silica gel (50% AcOEt in hexanes as eluent). The product was then dissolved in DCM, washed 3 times with a solution of CuSO₄ (5% in water), dried with MgSO₄ and evaporated under reduced pressure to afford **6f** (78 mg, 58% yield) as a colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.94 – 7.87 (m, 2H), 7.71 – 7.64 (m, 1H), 7.62 – 7.54 (m, 2H), 3.94 – 3.73 (m, 4H), 3.47 – 3.34 (m, 2H), 3.26 – 3.17 (m, 1H), 3.07 – 2.89 (m, 2H), 3.16 – 2.01 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 205.6, 139.1, 134.1, 129.6, 128.1, 69.3, 68.4, 51.1, 50.6, 34.4, 29.1.

HRMS (ESI pos): calculated for C₁₃H₁₆NaO₄S (M+Na⁺): 291.0662, found 291.0665.



3-(Phenylsulfonyl)-1-(1-tosylazetididin-3-yl)propan-1-one (6g):

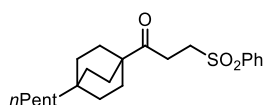
Synthesized according to the general procedure C using 1-tosylazetididine-3-carboxylic acid (191 mg, 0.75 mmol, 1.5 equiv.) and dimethyl fumarate (72 mg, 0.5 mmol, 1 equiv.). Acyl chloride formation was complete after 2 hours. In this case the reaction was irradiated for 36 hours. The crude mixture was purified by flash column

chromatography on silica gel (40% AcOEt in hexanes as eluent) to afford **6g** (63 mg, 31% yield) as a white solid.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.88 – 7.82 (m, 2H), 7.74 – 7.63 (m, 3H), 7.60 – 7.53 (m, 2H), 7.40 – 7.33 (m, 2H), 3.92 – 3.88 (m, 2H), 3.88 – 3.81 (m, 2H), 3.41 – 3.28 (m, 3H), 2.79 (app t, $J=7.3$ Hz, 2H), 2.44 (s, 3H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 202.1, 144.6, 138.9, 134.2, 131.3, 130.0, 129.6, 128.5, 128.0, 51.8, 50.3, 38.1, 33.1, 21.7.

HRMS (ESI pos): calculated for $\text{C}_{19}\text{H}_{21}\text{NNaO}_5\text{S}_2$ ($\text{M}+\text{Na}^+$): 430.0753, found 430.0748.



1-(4-Pentylbicyclo[2.2.2]octan-1-yl)-3-(phenylsulfonyl)propan-1-one (6h): Synthesized according to the general procedure C using 4-pentylbicyclo[2.2.2]octane-1-carboxylic acid (168 mg, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). Acyl chloride formation was complete after 2 hours. Chromatography on silica gel (10% AcOEt in hexanes as eluent) could not remove byproducts completely. Purification by semipreparative HPLC (IC column, 60:40 Hexane/Ethanol, 1 mL/min) was performed to obtain an analytical amount of product **6h** as a white solid. NMR yield (Trichloroethylene was used as internal standard): 80%.

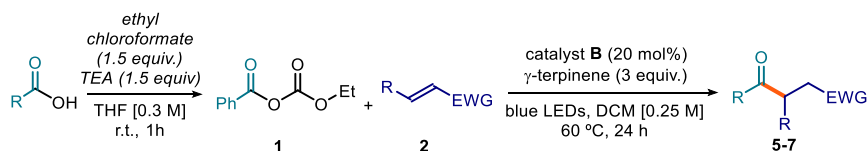
$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.93 – 7.87 (m, 2H), 7.69 – 7.63 (m, 1H), 7.61 – 7.54 (m, 2H), 3.36 – 3.29 (m, 2H), 2.97 – 2.90 (m, 2H), 1.69 – 1.61 (m, 6H), 1.43 – 1.34 (m, 6H), 1.31 – 1.24 (m, 2H), 1.24 – 1.12 (m, 4H), 1.11 – 1.04 (m, 2H), 0.87 (t, $J=7.3$ Hz, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 211.6, 139.4, 134.0, 129.5, 128.1, 51.0, 45.3, 41.3, 32.9, 30.8, 30.5, 30.1, 28.2, 23.5, 22.8, 14.2.

HRMS (ESI pos): calculated for $\text{C}_{22}\text{H}_{32}\text{NaO}_3\text{S}$ ($\text{M}+\text{Na}^+$): 399.1964, found 399.1951.

C4. Reaction of Carboxylic Acids through Anhydride Formation

C4.1 General Procedure D1



In a round bottom flask or vial, the carboxylic acid (0.75 mmol, 1.5 equiv) was dissolved in THF (2.5 mL, HPLC grade). Then, triethylamine (TEA, 105 μL , 0.75 mmol, 1.5 equiv.) and ethyl chloroformate (72 μL , 0.75 mmol, 1.5 equiv.) were added at ambient temperature. The reaction was stirred at ambient temperature for 1 hour. After that, it was filtered and the remaining solid was washed with diethyl ether. The organic layers were concentrated to dryness under reduced pressure to obtain the crude anhydride, which was used without further purification in the next step. (When the carboxylic acid is not completely soluble in 2.5 mL of THF, follow the general procedure D2 detailed below).

In an oven dried tube or a vial, with a Teflon septum screw cap, potassium ethyl xanthogenate **B** (16 mg, 0.10 mmol, 0.2 equiv.) and the electron-poor olefin **2** (0.5 mmol, 1 equiv., *if solid*), were added. The crude anhydride was dissolved in DCM (2 mL, HPLC grade) and the solution was added to the vial, followed by γ -terpinene (240 μL , 1.5 mmol, 3 equiv.). The resulting yellow mixture was degassed with argon sparging for 60 seconds. If the electron-poor olefin **2** was *liquid*, it was added via syringe after the argon sparging. The vial was then placed in the correspondent photoreactor (Figure S4 or S5, depending on the temperature used for the reaction) and irradiated for 24 hours. The solvent was evaporated and the residue purified by column chromatography to

afford the corresponding product in the stated yield with >95% purity, according to ¹H NMR analysis.

This procedure was employed for the optimization of the catalytic reaction with anhydride as radical precursor and for the scale-up process, as indicated below.

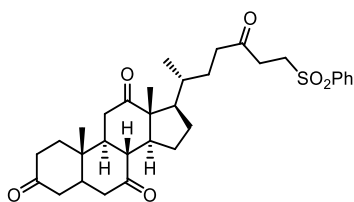
Note: Filtration of the triethylammonium salt is crucial for the reaction to work. Therefore, attempts to perform the reaction one-pot without removal of the ammonium salt were unsuccessful.

C4.2 General Procedure D2

In a round bottom flask or vial, the carboxylic acid (0.75 mmol, 1.5 equiv.) was dissolved in THF (10 mL, HPLC grade). Then, triethylamine (105 μL, 0.75 mmol, 1.5 equiv.) and ethyl chloroformate (72 μL, 0.75 mmol, 1.5 equiv.) were added at ambient temperature. The reaction was stirred for 1 hour. The reaction crude was washed with water and NaHCO₃ saturated solution, dried over MgSO₄, filtered and the organic layers concentrated to dryness under vacuum. The crude carbonate was used without further purification in the next step.

In an oven dried tube of 15 mL (16 mm × 12.5 mm) or a vial, with a Teflon septum screw cap, potassium ethyl xanthogenate (16.02 mg, 0.10 mmol, 0.2 equiv.) and the electron-poor olefin **2** (0.5 mmol, 1 equiv. *if solid*), were added. The crude carbonate was dissolved in DCM (2 mL, HPLC grade) and the solution was added to the vial, followed by γ-terpinene (240 μL, 1.5 mmol, 3 equiv.). The resulting yellow mixture was degassed with argon sparging for 60 seconds. If the electron-poor olefin **2** was *liquid*, it was added via syringe after the argon sparging. The vial was then placed in the correspondent photoreactor (Figure S4 or S5, depending on the temperature used for the reaction) and irradiated for 24 hours. After cooling to ambient temperature, the solvent was evaporated and the residue purified by column chromatography to afford the corresponding product in the stated yield with >95% purity according to ¹H NMR analysis.

C4.3 Characterization of Products



(8R,9S,10S,13R,14S,17R)-10,13-dimethyl-17-((R)-5-oxo-7-(phenylsulfonyl)heptan-2-yl)dodecahydro-3H-cyclopenta[a]phenanthrene-3,7,12(2H,4H)-trione (7):

Synthesized according to the general procedure **D2** using dehydrocholic acid (302 μL, 0.75 mmol, 1.5 equiv.) and phenyl vinyl sulfone (84 mg, 0.5 mmol, 1 equiv.). The crude mixture was purified by flash column chromatography on silica gel (33% acetone in hexanes as eluent), followed by a second purification (20% AcOEt in DCM as eluent) to afford **7** (145 mg, 52% yield) as a colorless oil.

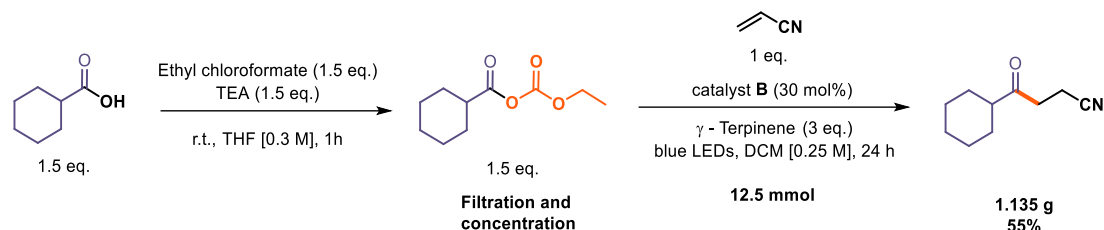
¹H NMR (500 MHz, CDCl₃) δ 7.93 – 7.88 (m, 2H), 7.70 – 7.64 (m, 1H), 7.61 – 7.55 (m, 2H), 3.44 – 3.32 (m, 2H), 2.95 – 2.79 (m, 5H), 2.54 – 2.44 (m, 1H), 2.44 – 2.10 (m, 9H), 2.07 – 1.90 (m, 4H), 1.89 – 1.80 (m, 1H), 1.80 – 1.70 (m, 1H), 1.67 – 1.56 (m, 1H), 1.39 (s, 3H), 1.36 – 1.20 (m, 4H), 1.05 (s, 3H), 0.81 (d, *J* = 6.5 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 212.0, 209.1, 208.8, 206.5, 139.2, 134.1, 129.6, 128.1, 57.0, 51.9, 50.6, 49.1, 47.0, 45.7, 45.7, 45.1, 42.9, 40.0, 38.8, 36.6, 36.1, 35.4, 35.4, 35.1, 29.1, 27.8, 25.2, 22.0, 18.9, 12.0.

HRMS (ESI pos): calculated for C₃₂H₄₂NaO₆S (M+Na⁺): 577.2594, found 577.2607.

C5 Scaled-Up Reaction

C5.1 Experimental Setup and Procedure



In a 100 mL round bottom flask, cyclohexanecarboxylic acid (2.34 mL, 18.75 mmol, 1.5 equiv.) was dissolved in THF (60 mL, HPLC grade). Then, triethylamine (2.61 mL, 18.75 mmol, 1.5 equiv.) and ethyl chloroformate (1.80 mL, 18.75 mmol, 1.5 equiv.) were added at ambient temperature. The reaction was stirred at ambient temperature for 1 hour. Then, it was filtered into a 100 mL reaction flask that will be used as reaction vessel for the next step (the remaining solid was carefully washed with diethyl ether). The organic phase was concentrated to dryness under vacuum to obtain the crude carbonate, which was used without further purification in the next step.

In the same 100 mL round bottom flask with a Teflon septum, the crude carbonate was dissolved in DCM (50 mL, HPLC grade). Potassium ethyl xanthogenate **B** (601 mg, 3.75 mmol, 0.3 equiv.) and γ -terpinene (6.01 mL, 37.5 mmol, 3 equiv.) were added to the solution. The resulting yellow mixture was degassed with Nitrogen sparging for 2 minutes. Finally, acrylonitrile (0.819 mL, 12.5 mmol, 1 equiv.) was added via syringe. The round bottom flask was then irradiated for 20 hours with a one meter 14W blue LED strip and cooled with a fan to keep the temperature between 30 and 35 °C (see Figure S7). After 24 hours, complete conversion of acrylonitrile was inferred by ^1H NMR analysis. The mixture was transferred to an extraction funnel, water was added and the organic layer was extracted with DCM. The organic layer was dried (MgSO_4) and concentrated to dryness. The product was then purified by chromatography on silica gel (10% AcOEt in hexanes) to afford 1.130 g of product **5e** (6.87 mmol, 55% yield) as a yellowish oil. NMR analysis was consistent with product synthesized in the small scale process.

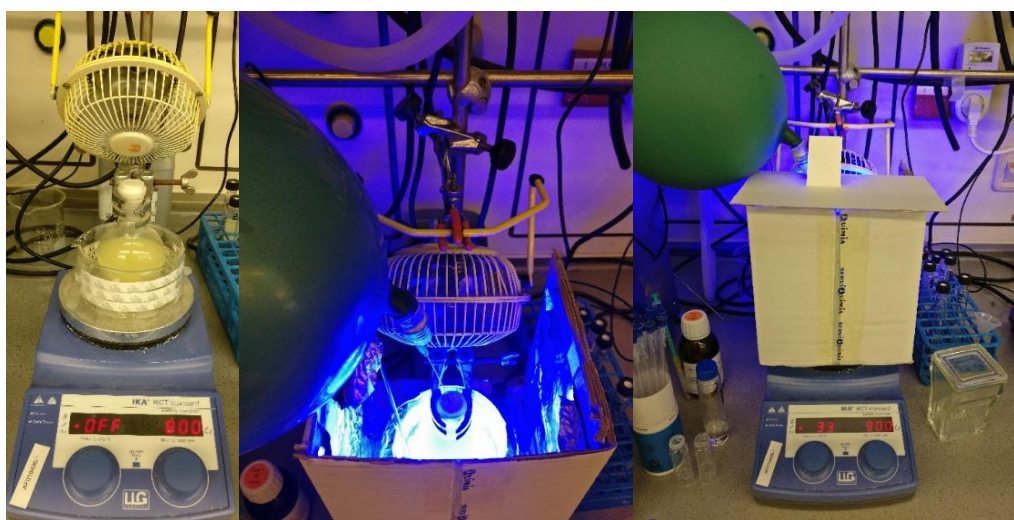


Figure S7: Experimental setup used for the large scale set up. (Left) Before irradiation. (Middle) Reaction set up from above. (Right) Reaction set up from the front.

C6. Reaction of Carbamoyl Chlorides

C6.1 Experimental Setup

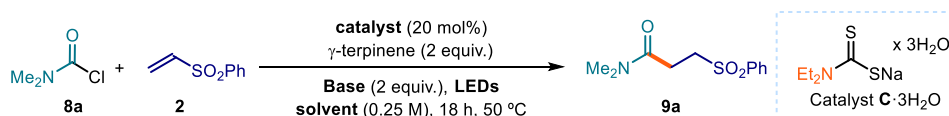
For the generation of carbamoyl radicals, we used the Hepatochem PhotoRedOx Box equipped with an EvoluChem LED 18 W light source at 405 nm, supplied by Hepatochem. The reactor was connected to a Huber Minichiller 300 in order to perform reactions at 50 °C with accurate control of the reaction temperature ($\pm 1^\circ\text{C}$, Figure S8).



Figure S8: Photoreactor used for the reaction with the carbamoyl chlorides

C6.2 Optimization Studies

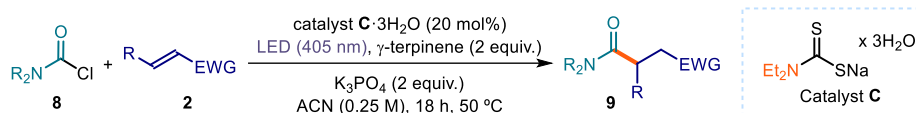
Table S8. Screening of the Catalysts



entry	catalyst	wavelength (nm)	Base	Solvent	NMR yield (%)
1	B	460	Na ₃ PO ₄	DCM	<5
2	B	405	Na ₃ PO ₄	DCM	10%
3	C·3H ₂ O	405	Na ₃ PO ₄	DCM	11%
4	C·3H ₂ O	405	Na ₃ PO ₄	MeCN	20%
5	C·3H ₂ O	405	K ₃ PO ₄	MeCN	60%

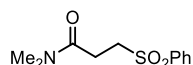
All reaction performed on 0.2 mmol scale, yield determined by ¹H NMR analysis of the crude reaction mixture by comparison with trichloroethylene as internal standard.

C6.3 General Procedure for the intermolecular Giese addition (General Procedure E)



An oven-dried 15 mL Schlenk tube was charged with a mixture of carbamoyl chloride **8** (0.4 mmol, 2 equiv.), catalyst **C** trihydrate (9 mg, 0.04 mmol, 0.2 equiv.), alkene **2** (0.2 mmol, 1 equiv.), γ -terpinene (64 μ L, 0.4 mmol, 2 equiv.) and K_3PO_4 (85 mg, 0.4 mmol, 2 equiv.) in acetonitrile (0.8 mL, 0.25 M). The reaction mixture was placed under an atmosphere of argon, cooled to -78 $^{\circ}C$, degassed *via* vacuum evacuation (5 minutes), backfilled with argon and, ultimately, warmed to ambient temperature. This freeze-pump-thaw cycle was repeated four times, and then the Schlenk tube was sealed with Parafilm and put into the Hepatochem PhotoRedOx Box equipped with a 405 nm EvoluChem LED 18 W light source at 50 $^{\circ}C$ (Figure S8). After 18 hours stirring, the reaction was cooled down to ambient temperature, water was added and the mixture was extracted with ethyl acetate (2x15 mL). The combined layers were dried over magnesium sulfate, filtered, and concentrated. The resulting crude mixture was purified by column chromatography on silica gel to give the corresponding product **9** in the stated yield.

C6.4 Characterization of Products



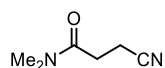
***N,N*-Dimethyl-3-(phenylsulfonyl)propanamide (9a)**: Synthesized according to general procedure E using dimethylcarbamic chloride (37 μ L, 0.4 mmol, 2.0 equiv.) and phenyl vinyl sulfone (34 mg, 0.2 mmol). The crude mixture was

purified by flash column chromatography on silica gel (gradient from hexane 100% to ethyl acetate 100%) to afford product **9a** (29 mg, 60% yield) as a pale-yellow oil.

1H NMR (400 MHz, $CDCl_3$) δ 7.95 – 7.88 (m, 2H), 7.69 – 7.62 (m, 1H), 7.61 – 7.52 (m, 2H), 3.53 – 3.33 (m, 2H), 2.99 (s, 3H), 2.89 (s, 3H), 2.85 – 2.73 (m, 2H).

^{13}C NMR (100 MHz, $CDCl_3$) δ 168.8, 139.2, 134.0, 129.5, 128.1, 52.2, 37.2, 35.7, 26.2.

HRMS (ESI pos): calculated for $C_{11}H_{15}NnaO_3S$ ($M+Na^+$): 264.0665, found 264.0653.

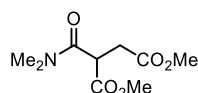


3-Cyano-*N,N*-dimethylpropanamide (9b): Synthesized according to general procedure E using dimethylcarbamic chloride (37 μ L, 0.4 mmol, 2.0 equiv.) and acrylonitrile (13 μ L, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (gradient from hexane 100% to ethyl acetate 100%) to afford product **9b** (16 mg, 63% yield) as a pale-yellow oil.

1H NMR (400 MHz, $CDCl_3$) δ 3.01 (s, 3H), 2.97 (s, 3H), 2.68 (s, 4H).

^{13}C NMR (100 MHz, $CDCl_3$) δ 168.8, 119.6, 37.0, 35.7, 29.5, 13.1.

HRMS (ESI pos): calculated for $C_6H_{11}N_2O$ ($M+H^+$): 127.0866, found: 127.0869.



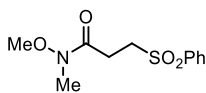
dimethyl 2-(dimethylcarbamoyl)succinate (9c): Synthesized according to general procedure E using dimethylcarbamic chloride (37 μ L, 0.4 mmol, 2.0 equiv.) and dimethyl fumarate (29 mg, 0.2 mmol). The crude mixture was

purified by flash column chromatography on silica gel (gradient from hexane 100% to ethyl acetate 100%) to afford product **9c** (27 mg, 62% yield) as a pale-yellow oil.

1H NMR (400 MHz, $CDCl_3$) δ 4.14 (dd, J = 8.3, 6.0 Hz, 1H), 3.72 (s, 3H), 3.67 (s, 3H), 3.16 (s, 3H), 3.11 – 2.80 (m, 6H).

^{13}C NMR (100 MHz, $CDCl_3$) δ 172.4, 169.5, 167.9, 53.0, 52.2, 44.6, 38.0, 36.4, 33.6.

HRMS (ESI pos): calculated for $C_9H_{15}NNaO_5$ ($M+Na^+$): 240.0842, found: 240.0844.

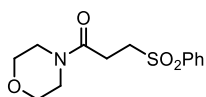


N-Methoxy-N-methyl-3-(phenylsulfonyl)propanamide (9d): Synthesized according to general procedure E using methoxy(methyl)carbamic chloride (41 μ L, 0.4 mmol, 2.0 equiv.) and phenyl vinyl sulfone (34 mg, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (gradient from hexane 100% to hexane 50% : 50% ethyl acetate) to afford product **9d** (16 mg, 30% yield) as a pale yellow oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.97 – 7.90 (m, 2H), 7.75 – 7.63 (m, 1H), 7.63 – 7.54 (m, 2H), 3.69 (s, 3H), 3.52 – 3.37 (m, 2H), 3.14 (s, 3H), 2.97 – 2.92 (m, 2H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) 170.4, 139.2, 134.0, 129.5, 128.2, 61.6, 51.5, 32.3, 25.4.

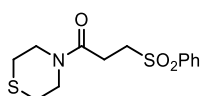
HRMS (ESI pos): calculated for $\text{C}_9\text{H}_{15}\text{NNaO}_5$ ($\text{M}+\text{Na}^+$): 240.0842, found: 240.0844.



1-Morpholino-3-(phenylsulfonyl)propan-1-one (9e): Synthesized according to general procedure E using morpholine-4-carbonyl chloride (47 μ L, 0.4 mmol, 2.0 equiv.) and phenyl vinyl sulfone (34 mg, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (gradient from hexane 100% to ethyl acetate 100%) to afford product **9e** (42 mg, 74% yield) as a pale-yellow oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 8.00 – 7.85 (m, 2H), 7.71 – 7.62 (m, 1H), 7.62 – 7.51 (m, 1H), 3.76 – 3.58 (m, 5H), 3.55 – 3.49 (m, 2H), 3.47 – 3.42 (m, 3H), 2.94 – 2.74 (m, 2H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 167.5, 139.1, 134.0, 129.5, 128.0, 66.7, 66.5, 52.0, 45.8, 42.3, 25.9. **HRMS (ESI pos):** calculated for $\text{C}_{13}\text{H}_{17}\text{NNaO}_4\text{S}$ ($\text{M}+\text{Na}^+$): 306.0770, found: 306.0762.

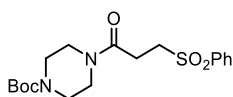


3-(Phenylsulfonyl)-1-thiomorpholinopropan-1-one (9f): synthesized according the general procedure E using thiomorpholine-4-carbonyl chloride **8f** (99 mg, 0.4 mmol, 2 eq.) and phenyl vinyl sulfone (34 mg, 0.2 mmol, 1 eq.). The crude mixture was purified by flash column chromatography on silica gel (5% AcOEt in hexane), followed by a second one (50% AcOEt in Hexane as eluent) to afford **9f** (37.8 mg, 63% yield) as a colorless oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.96 (m, 2H) 7.70 (m, 1H) 7.61 (m, 2H) 3.85 (m, 2H), 3.76 (m, 1H), 3.65 (m, 1H), 3.50 (m, 1H), 2.86 (m, 2H), 2.64 (m, 6H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 167.3, 139.3, 134.1, 129.5, 128.1, 52.2, 48.3, 44.8, 27.9, 27.4, 26.2

HRMS (ESI pos): calculated for $\text{C}_{13}\text{H}_{17}\text{NaNO}_3\text{S}_2$ ($\text{M}+\text{Na}^+$): 322.05, found: 322.0533.

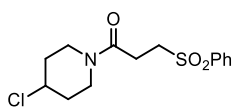


tert-Butyl 4-(3-(phenylsulfonyl)propanoyl)piperazine-1-carboxylate (9g): synthesized according general procedure E using tert-butyl 4-(chlorocarbonyl)piperazine-1-carboxylate **8g** (99 mg, 0.4 mmol, 2 eq.) and phenyl vinyl sulfone (34 mg, 0.2 mmol, 1 eq.). The crude mixture was purified by flash column chromatography on silica gel (5% AcOEt in hexane), followed by a second purification (50% AcOEt in hexane as eluent) to afford **9g** (55 mg, 73% yield) as a white solid.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.91 (m, 2H), 7.66 (m, 1H), 7.57 (m, 2H), 3.42 (m, 4H) 3.21 (m, 6H), 2.83 (t, $J = 7.8$ Hz, 2H), 1.45 (s, 9H)

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 167.6, 154.6, 139.3, 134.1, 129.6, 128.1, 80.7, 52.1, 45.4, 41.9, 28.5, 26.1

HRMS (ESI pos): calculated for $\text{C}_{18}\text{H}_{26}\text{NaN}_2\text{O}_5\text{S}$ ($\text{M}+\text{Na}^+$): 405.15, found 405.1455.



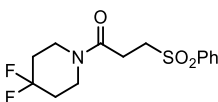
1-(4-Chloropiperidin-1-yl)-3-(phenylsulfonyl)propan-1-one (9h):

Synthesized according to general procedure E using 4-chloropiperidine-1-carbonyl chloride (73 mg, 0.4 mmol, 2 equiv.) and phenyl vinyl sulfone (34 mg, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (gradient from hexane 100% to ethyl acetate 100%) to afford product **9h** (41 mg, 65% yield) as a pale-yellow oil.

¹H NMR (400 MHz, CDCl₃) δ 7.96 – 7.87 (m, 2H), 7.72 – 7.62 (m, 1H), 7.60 – 7.53 (m, 2H), 4.27 (tt, *J* = 7.0, 3.6 Hz, 1H), 3.68 (tdt, *J* = 11.7, 8.2, 3.5 Hz, 2H), 3.58 (ddd, *J* = 13.6, 6.9, 4.0 Hz, 1H), 3.49 – 3.42 (m, 2H), 3.38 (ddd, *J* = 13.9, 7.0, 3.7 Hz, 1H), 2.83 (td, *J* = 7.1, 1.9 Hz, 2H), 2.16 – 1.93 (m, 2H), 1.90 – 1.72 (m, 2H).

¹³C NMR (100 MHz, CDCl₃) δ 167.2, 139.2, 134.0, 129.5, 128.0, 56.1, 52.1, 42.5, 39.1, 35.1, 34.4, 25.9.

HRMS (ESI pos): calculated for C₁₄H₁₉ClNO₃S (M+H⁺): 316.0769, found: 316.0773.



1-(4,4-Difluoropiperidin-1-yl)-3-(phenylsulfonyl)propan-1-one (9i):

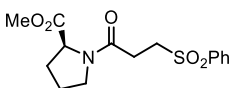
Synthesized according to general procedure E using 4,4-difluoropiperidine-1-carbonyl chloride (73 mg, 0.4 mmol, 2 equiv.) and phenyl vinyl sulfone (34 mg, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (gradient from hexane 100% to 1:1 hexane/ethyl acetate) to afford product **9i** (43 mg, 68% yield) as an off-white solid.

¹H NMR (400 MHz, CDCl₃) δ 7.96 – 7.88 (m, 2H), 7.71 – 7.63 (m, 1H), 7.61 – 7.54 (m, 2H), 3.61 (dt, *J* = 43.9, 6.0 Hz, 4H), 3.51 – 3.43 (m, 2H), 2.92 – 2.83 (m, 2H), 2.13 – 1.84 (m, 4H).

¹³C NMR (100 MHz, CDCl₃) δ 167.4, 139.2, 134.1, 129.5, 128.0, 121.3 (t, *J* = 242.4 Hz), 52.1, 40.7 (dt, *J* = 330.8, 5.4 Hz), 34.1 (dt, *J* = 78.0, 23.6 Hz), 25.9.

¹⁹F NMR (376 MHz, CDCl₃) δ -98.22 (p, *J* = 13.5 Hz).

HRMS (ESI pos): calculated for C₁₈H₁₈NaO₄ (M+Na⁺): 340.0789, found: 340.0785.



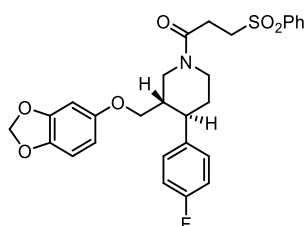
Methyl (3-(phenylsulfonyl)propanoyl)-L-prolinate (9j):

synthesized according the general procedure E using methyl (chlorocarbonyl)-L-prolinate (77 mg, 0.4 mmol, 2 eq.) and phenyl vinyl sulfone (34 mg, 0.2 mmol, 1 eq.). The crude mixture was purified by flash column chromatography on silica gel (5% AcOEt in hexane) to afford **9j** (46 mg, 58% yield) as a yellowish oil.

¹H NMR (400 MHz, CDCl₃) δ 7.95 – 7.89 (m, 2H), 7.69 – 7.64 (m, 1H), 7.61 – 7.53 (m, 2H), 4.42 (dd, *J* = 8.4, 3.5 Hz, 1H), 3.70 (s, 3H), 3.68 – 3.59 (m, 1H), 3.56 – 3.38 (m, 3H), 2.83 (dt, *J* = 9.7, 5.6 Hz, 2H), 2.11 – 1.94 (m, 4H).

¹³C NMR (126 MHz, CDCl₃) δ 172.6, 167.8, 139.2, 134, 129.5, 128.1, 58.9, 52.4, 51.7, 47.1, 29.3, 27.4, 24.8.

HRMS (ESI pos): calculated for C₁₅H₁₉NaNO₅S (M+Na⁺): 326.10, found: 326.1063.



1-(3-(((Benzo[d][1,3]dioxol-5-yl)oxy)methyl)-4-(4-

fluorophenyl)piperidin-1-yl)-3-(phenylsulfonyl)propan-1-one

(9k): synthesized according the general procedure E using 3-

((benzo[d][1,3]dioxol-5-yl)oxy)methyl)-4-(4-

fluorophenyl)piperidine-1-carbonyl chloride **8k** (157 mg, 0.4 mmol,

2 eq.) and phenyl vinyl sulfone (34 mg, 0.2 mmol, 1 eq.). The crude

mixture was purified by flash column chromatography on silica gel

(gradient from hexane 100% to hexane 50% : 50% ethyl acetate) to afford product **9k** (46 mg, 29% yield) as a yellowish oil.

¹H NMR (400 MHz, CDCl₃) δ 7.98 (m, 2H), 7.71 (m, 1H), 7.65 – 7.58 (m, 2H), 7.17 – 7.11 (m, 2H), 7.01 (m, 2H), 6.66 (dd, *J* = 17.8, 8.4 Hz, 1H), 6.38 (dd, *J* = 11.9, 2.5 Hz, 1H), 6.17 (ddd, *J* = 15.1, 8.5, 2.5 Hz, 1H), 5.92 (d, *J* = 13.5 Hz, 2H), 4.79 (dd, *J* = 55.2, 13.1 Hz, 1H), 4.07 (dd, *J* = 69.6, 13.5 Hz, 1H), 3.68 – 3.60 (m, 1H), 3.59 – 3.45 (m, 3H), 3.17 (m, 1H), 3.03 – 2.80 (m, 2H), 2.80 – 2.62 (m, 1H), 1.92 (m, 1H), 1.81 – 1.53 (m, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 167.5, 154.5, 148.7, 142.3, 139.6, 138.4, 134.3, 129.7, 129.1, 128.3, 115.7, 115.8, 108.2, 106.0, 101.5, 98.4, 68.9, 52.5, 49.2, 46.4, 44.5, 42.2, 34.6, 33.8, 30.1, 26.3.

¹⁹F NMR (376 MHz, CDCl₃, proton decoupled) δ – 115.58

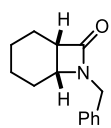
HRMS (ESI pos): calculated for C₂₈H₂₉FNO₆S (M+H⁺): 526.16, found: 526.1702.

C6.5 General Procedure for the intramolecular Giese type addition (General Procedure F)



An oven-dried 15 mL Schlenk tube was charged with a mixture of carbamoyl chloride **10** (0.2 mmol, 1 equiv.), catalyst **C** trihydrate (9 mg, 0.04 mmol, 0.2 equiv.), γ -terpinene (48 μ L, 0.3 mmol, 1.5 equiv.) and K₃PO₄ (85 mg, 0.4 mmol, 2 equiv.) in acetonitrile (0.8 mL, 0.25 M). The reaction mixture was placed under an atmosphere of argon, cooled to –78 °C, degassed *via* vacuum evacuation (5 minutes), backfilled with argon and, ultimately, warmed to ambient temperature. This freeze-pump-thaw cycle was repeated four times, and then the Schlenk tube was sealed with Parafilm and put into the Hepatochem PhotoRedOx Box equipped with a 405 nm EvoluChem LED 18 W light source at 50 °C (Figure S8). After 18 hours, the reaction vessel was cooled down to ambient temperature, water was added and the mixture was extracted with ethyl acetate (2x15 mL). The combined layers were dried over magnesium sulfate, filtered, and concentrated. The resulting crude mixture was purified by column chromatography on silica gel to give the corresponding product **3** in the stated yield.

C6.6 Characterization of Products

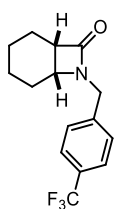


7-Benzyl-7-azabicyclo[4.2.0]octan-8-one (11a): Synthesized according to the general procedure F using benzyl(cyclohex-2-en-1-yl)carbamoyl chloride (50 mg, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (10% AcOEt in hexanes as eluent) to afford product **11a** (26 mg, 60% yield) as a pale-yellow oil.

¹H NMR (500 MHz, CDCl₃) δ 7.35 – 7.29 (m, 2H), 7.29 – 7.22 (m, 3H), 4.57 (d, *J* = 15.1 Hz, 1H), 4.08 (d, *J* = 15.1 Hz, 1H), 3.62 (ddd, *J* = 5.4, 4.2, 3.1 Hz, 1H), 3.20 – 3.15 (m, 1H), 1.89 – 1.80 (m, 1H), 1.74 – 1.49 (m, 5H), 1.49 – 1.33 (m, 2H).

¹³C NMR (126 MHz, CDCl₃) δ 171.1, 136.4, 128.9, 128.5, 127.8, 50.3, 47.1, 44.6, 23.0, 19.8, 19.1, 17.0.

HRMS (ESI pos): calculated for C₁₈H₁₈NaO₄ (M+H⁺): 216.1383, found: 216.1374.



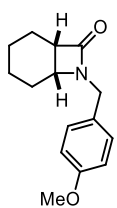
7-(4-(Trifluoromethyl)benzyl)-7-azabicyclo[4.2.0]octan-8-one (11b):

Synthesized according to general procedure F using cyclohex-2-en-1-yl(4-(trifluoromethyl)benzyl)carbamic chloride (64 mg, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (5% AcOEt in hexanes as eluent) to afford product **11b** (31 mg, 55% yield) as a yellow oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.59 (d, $J = 8.0$ Hz, 2H), 7.39 (d, $J = 8.0$ Hz, 2H), 4.62 (d, $J = 15.4$ Hz, 1H), 4.16 (d, $J = 15.4$ Hz, 1H), 3.66 (dt, $J = 5.1, 3.6$ Hz, 1H), 3.23 (dt, $J = 6.8, 4.6$ Hz, 1H), 1.92 – 1.83 (m, 1H), 1.77 – 1.55 (m, 5H), 1.50 – 1.34 (m, 2H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 171.1, 140.5, 130.1 (q, $J = 32.6$ Hz), 128.6, 125.9 (q, $J = 3.8$ Hz), 124.2 (q, $J = 272.1$ Hz), 50.5, 47.3, 44.1, 23.0, 19.7, 19.0, 16.9.

HRMS (ESI pos): calculated for $\text{C}_{15}\text{H}_{17}\text{F}_3\text{NO}$ ($\text{M}+\text{H}^+$): 284.1257, found: 284.1258.

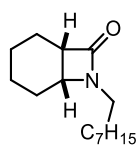


7-(4-Methoxybenzyl)-7-azabicyclo[4.2.0]octan-8-one (11c): Synthesized according to general procedure F using cyclohex-2-en-1-yl(4-methoxybenzyl)carbamic chloride (56 mg, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (15% AcOEt in hexanes as eluent) to afford product **11c** (30 mg, 61% yield) as a yellow oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.23 – 7.14 (m, 2H), 6.91 – 6.80 (m, 2H), 4.51 (d, $J = 14.9$ Hz, 1H), 4.04 (d, $J = 14.9$ Hz, 1H), 3.80 (s, 3H), 3.61 (ddd, $J = 5.4, 4.1, 3.2$ Hz, 1H), 3.17 (dt, $J = 6.9, 4.6$ Hz, 1H), 1.93 – 1.79 (m, 1H), 1.74 – 1.50 (m, 4H), 1.49 – 1.31 (m, 3H).

$^{13}\text{C NMR}$ (100 MHz, CDCl_3) δ 170.9, 159.2, 129.7, 128.4, 114.2, 55.4, 50.0, 46.9, 43.9, 23.0, 19.7, 19.0, 16.9.

HRMS (ESI pos): calculated for $\text{C}_{15}\text{H}_{19}\text{NaO}_2$ ($\text{M}+\text{Na}^+$): 268.1308, found: 268.1304.

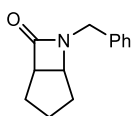


7-Octyl-7-azabicyclo[4.2.0]octan-8-one (11d): synthesized according the general procedure F using cyclohex-1-en-1-yl(octyl)carbamic chloride **10d** (54 mg, 0.2 mmol, 1 eq.). The crude mixture was purified by flash column chromatography on silica gel (5% AcOEt in Hexane) to afford **11d** (13 mg, 28% yield) as a colorless oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ : 3.72 (ddd, $J = 5.3, 4.2, 3.2$ Hz, 1H), 3.33 (dt, $J = 13.9, 7.6$ Hz, 1H), 3.20 – 3.11 (m, 1H), 2.94 (ddd, $J = 14.0, 7.9, 6.2$ Hz, 1H), 1.94 – 1.39 (m, 10H), 1.35 – 1.20 (m, 9H), 1.00 – 0.81 (m, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 171, 50.2, 46.6, 40.3, 31.9, 29.4, 29.3, 28.2, 27.3, 23.3, 22.8, 19.7, 19, 17.1, 14.2.

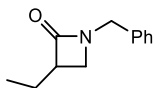
HRMS (ESI pos): calculated for $\text{C}_{15}\text{H}_{27}\text{NO}$ ($\text{M}+\text{H}^+$): 238,21 found: 238,2157



6-Benzyl-6-azabicyclo[3.2.0]heptan-7-one (11e): synthesized according the general procedure F using benzyl(cyclopent-2-en-1-yl)carbamic chloride **10e** (47 mg, 0.2 mmol, 1 eq.). The crude mixture was purified by flash column chromatography on silica gel (5% AcOEt in hexane) to afford **11e** (11 mg, 30% yield) as a yellowish oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.54 – 7.08 (m, 5H), 4.50 (d, $J = 15.1$ Hz, 1H), 4.08 (d, $J = 15.1$ Hz, 1H), 3.91 (t, $J = 4.1$ Hz, 1H), 3.47 (dd, $J = 8.0, 3.6$ Hz, 1H), 2.05 (dd, $J = 13.2, 6.3$ Hz, 1H), 1.83 – 1.70 (m, 2H), 1.58 (m, 1H), 1.47 – 1.30 (m, 1H), 1.18 (m, 1H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 169.6, 136.3, 128.8, 128.4, 127.8, 57.6, 55.1, 44.2, 27, 25.1, 22.8. **HRMS (ESI pos):** calculated for $\text{C}_{13}\text{H}_{15}\text{NaNO}$ ($\text{M}+\text{Na}^+$): 224.11 found: 224.1041

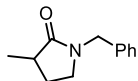


1-Benzyl-3-ethylazetidin-2-one (11f): synthesized according the general procedure F using benzyl(but-2-en-1-yl)carbamic chloride **10f** (47 mg, 0.2 mmol, 1 eq.). The crude mixture was purified by flash column chromatography on silica gel (5% AcOEt in hexane) to afford **11d** (11 mg, 29% yield) as a yellowish oil.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.37 – 7.26 (m, 5H), 4.46 (s, 2H), 3.60 – 3.52 (m, 2H), 3.18 – 3.10 (m, 1H), 1.72 – 1.65 (m, 2H), 0.93 (t, $J = 7.3$ Hz, 3H)

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ : 171.9 (C), 136.3 (C), 128.9 (CH), 128.3 (CH), 127.9 (CH), 53.6 (CH_2), 48.7 (CH_2), 44 (CH), 29 (CH_2), 11.9 (CH_3).

Matching reported literature data.²⁰

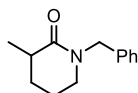


1-Benzyl-3-methylpyrrolidin-2-one (11g): Synthesized according to the general procedure F using benzyl(but-3-en-1-yl)carbamic chloride **10g** (48 mg, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (10% AcOEt in hexanes as eluent) to afford product **11g** (23 mg, 60% yield) as a yellow oil.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 7.37 – 7.30 (m, 2H), 7.29 – 7.24 (m, 1H), 7.24 – 7.20 (m, 2H), 4.49 – 4.40 (m, 2H), 3.22 – 3.09 (m, 2H), 2.51 (ddt, $J = 15.8, 8.7, 7.2$ Hz, 1H), 2.21 (dddd, $J = 12.9, 8.7, 6.5, 4.4$ Hz, 1H), 1.59 (dq, $J = 12.6, 8.5$ Hz, 1H), 1.24 (d, $J = 7.1$ Hz, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 177.5, 136.8, 128.8, 128.2, 127.6, 46.9, 44.8, 36.9, 27.2, 16.5.

HRMS (ESI pos): calculated for $\text{C}_{12}\text{H}_{16}\text{NO}$ ($\text{M}+\text{H}^+$): 190.1226, found: 190.1228.



1-Benzyl-3-methylpiperidin-2-one (11h): Synthesized according to the general procedure F using benzyl(pent-4-en-1-yl)carbamic chloride **10h** (48 mg, 0.2 mmol). The crude mixture was purified by flash column chromatography on silica gel (10% AcOEt in hexanes as eluent) to afford product **11h** (21 mg, 52% yield) as a yellow oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.36 – 7.28 (m, 2H), 7.28 – 7.21 (m, 3H), 4.66 (d, $J = 14.6$ Hz, 1H), 4.50 (d, $J = 14.6$ Hz, 1H), 3.20 (dd, $J = 7.2, 5.1$ Hz, 2H), 2.56 – 2.42 (m, 1H), 1.96 (dtd, $J = 12.8, 6.2, 3.4$ Hz, 1H), 1.84 (ddtd, $J = 13.5, 6.6, 5.0, 3.4$ Hz, 1H), 1.78 – 1.66 (m, 1H), 1.53 (dddd, $J = 12.8, 10.4, 9.1, 3.4$ Hz, 1H), 1.30 (d, $J = 7.2$ Hz, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 173.5, 137.7, 128.7, 128.1, 127.4, 50.4, 47.7, 36.8, 29.7, 21.8, 18.2.

HRMS (ESI pos): calculated for $\text{C}_{13}\text{H}_{18}\text{NO}$ ($\text{M}+\text{H}^+$): 204.1383, found: 204.1376.

C7. Unsuccessful Substrates

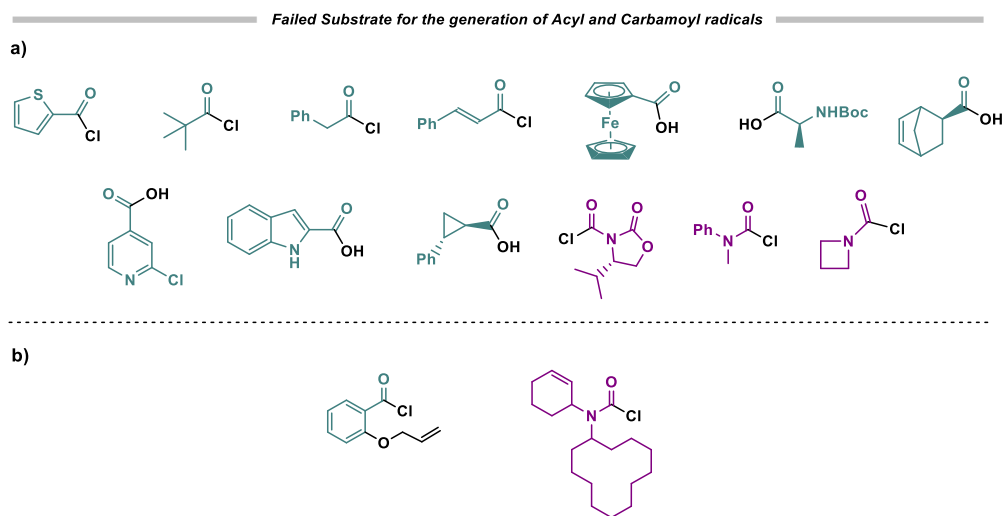
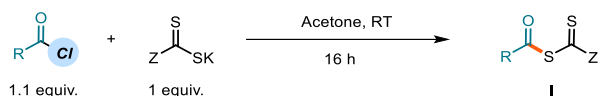


Figure S9: Unsuccessful substrates: (a) Intermolecular reaction using acrylonitrile as the electron-poor olefin. (b) Intramolecular reaction.

D. Mechanistic Studies

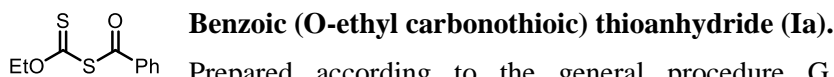
D1. Characterisation of Acylxanthate and Carbamoyldithiocarbamate Intermediates:

D1.1 General procedure for the synthesis of acyl xanthate intermediates (general procedure G)



In a round bottom flask, acyl chloride (1.1 equiv.) was dissolved in acetone (0.1 M) and cooled to 0 °C. The dithiocarbamate anion or the xanthate salt **1a-c** (1 equiv.) was then added and the resulting reaction mixture was stirred for 1 hour at 0 °C. The solvent was removed under reduced pressure at ambient temperature. The residue was then dissolved in DCM and washed with distilled water, NaHCO₃ solution and brine. The combined organic fractions were dried over MgSO₄ and concentrated to dryness to obtain the desired product.

D1.2 Characterization of the Intermediates

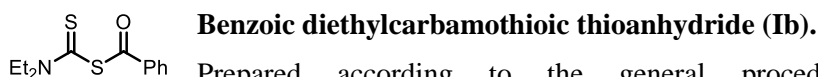


Prepared according to the general procedure G using potassium ethyl xanthogenate **B** (1 mmol, 160 mg) and benzoyl chloride (1.1 mmol, 128 μ L) in 10 mL of acetone. After work-up, the product **Ia** (226 mg, 99% yield) was obtained as a yellow oil.

¹H NMR (300 MHz, CDCl₃) δ 7.87 (app d, $J = 7.7$ Hz, 2 H); 7.64-7.54 (m, 1H); 7.49-7.39 (m, 2H); 4.69 (q, $J = 7.1$ Hz, 2H); 1.45 (t, $J = 7.1$ Hz, 3H).

¹³C NMR (75 MHz, CDCl₃) δ 203.4, 185.0, 135.7, 134.4, 129.0, 127.9, 71.1, 13.5.

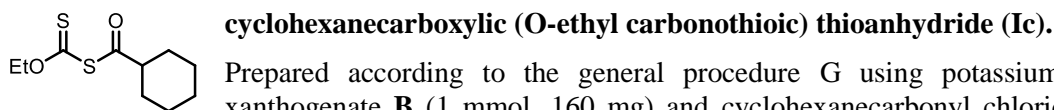
Matching reported literature data.²¹



Prepared according to the general procedure G using sodium diethylcarbamodithioate trihydrate **C** (1.05 mmol, 237 mg) and benzoyl chloride (1 mmol, 116 μ L) in 10 mL of acetone. After work-up, the product **Ib** was obtained as a yellow oil (220 mg, 87% yield).

¹H NMR (300 MHz, CDCl₃) δ 7.95-7.88 (m, 2 H); 7.64-7.56 (m, 1H); 7.52-7.42 (m, 2H); 4.07-3.86 (m, 2H); 1.36 (t, $J = 7.1$ Hz, 3H).

Matching reported literature data.²²



Prepared according to the general procedure G using potassium ethyl xanthogenate **B** (1 mmol, 160 mg) and cyclohexanecarbonyl chloride (1.1 mmol, 128 μ L) in 10 mL of acetone. After work-up, the product **Ic** was obtained as a yellow oil (207 mg, 89% yield).

¹H NMR (300 MHz, CDCl₃) δ 4.65 (q, $J = 7.1$ Hz, 2H); 2.53-2.39 (m, 1H); 1.98-1.85 (m, 2H); 1.83-1.70 (m, 2H); 1.68-1.53 (m, 1H); 1.55-1.35 (m, 5H); 1.34-1.10 (m, 3 H).

¹³C NMR (75 MHz, CDCl₃) δ 204.6 (C); 194.9 (C); 70.8 (CH₂); 52.8 (CH); 29.1 (CH₂); 25.5 (CH₂); 25.32 (CH₂); 13.6 (CH₃).

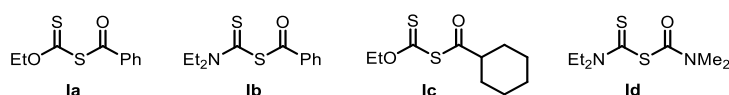
HRMS (ESI pos): calculated for C₁₀H₁₆NaO₂S₂ (M+Na⁺): 255.0484, found: 255.0482.

***N*-Diethyl, *N'*-Dimethyl-Thiodicarbamic diamide (**Id**).**
CCN(C)C(=O)S(=O)C(=O)NCC Prepared according to the general procedure G using potassium diethylcarbamodithioate trihydrate **C** (3 mmol, 724 mg) and dimethylcarbonyl chloride (1.1 mmol, 128 μ L) in 10 mL of acetone. After work-up and chromatography on silica gel (8:2 hexane/AcOEt), the product **Id** was obtained as a yellow oil (177 mg, 80% yield).

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 4.02 (q, $J = 7.1$ Hz, 1H), 3.79 (q, $J = 7.2$ Hz, 1H), 3.06 (s, 3H), 1.32 (dt, $J = 9.7, 7.1$ Hz, 3H)

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 185.2 (C), 162 (C), 50.1 (CH_3), 49 (CH_3), 38.5 (CH_2), 37.3 (CH_2), 13.5 (CH_3), 11.3 (CH_3).

D1.3 UV-Vis Characterization of the Intermediates.



Intermediate	λ_{max}	tail of absorption
la	397 nm	490 nm
lb	400 nm	490 nm
lc	400 nm	490 nm
ld	343 nm	500 nm

Table S9. Spectroscopic characteristics of **la-d**

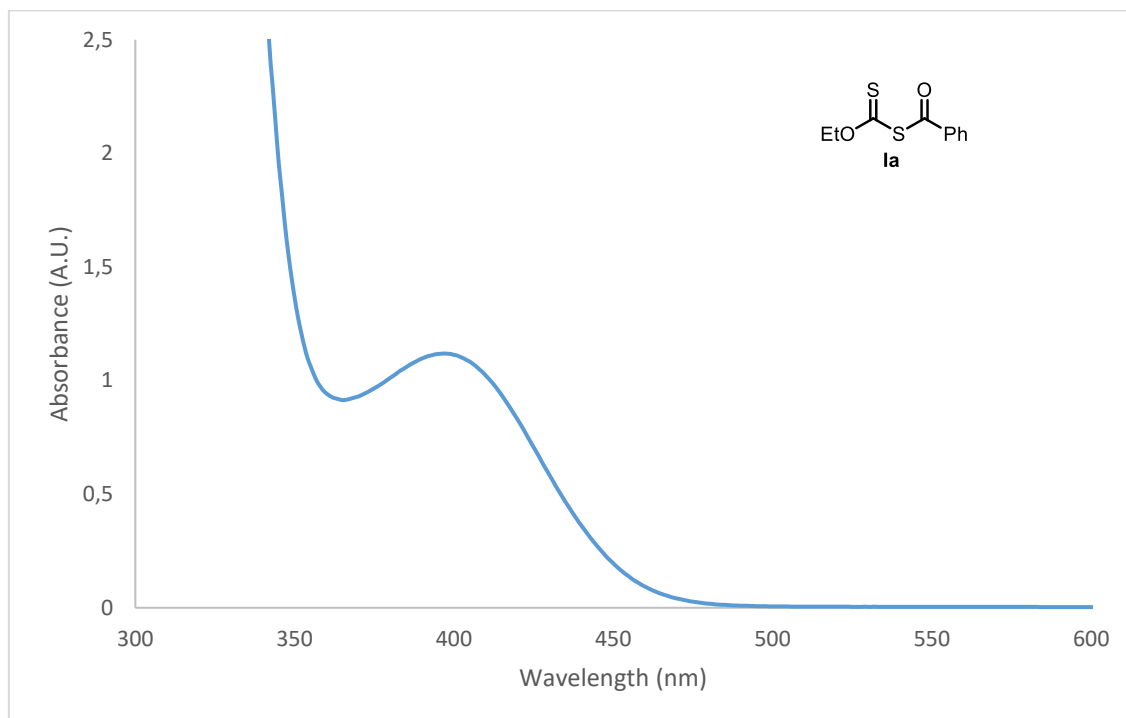


Figure S10: UV-Vis absorption spectrum of **la** recorded at $1 \cdot 10^{-2}$ M concentration in acetonitrile

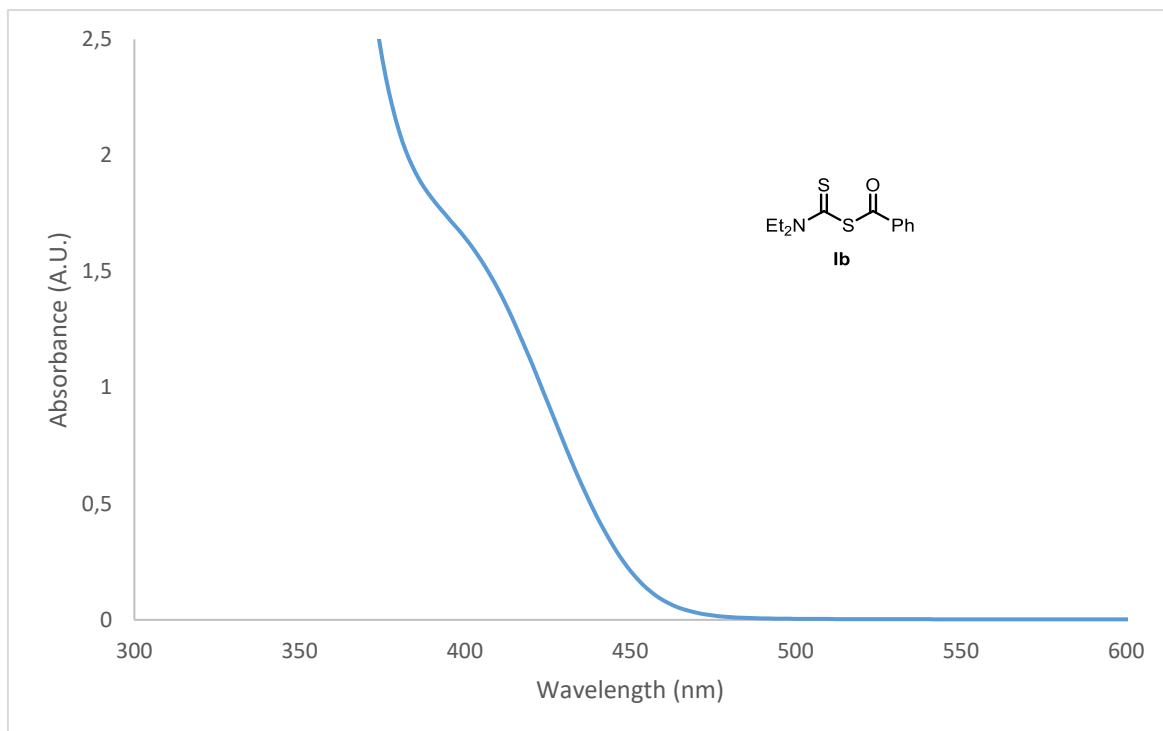


Figure S11: UV-Vis absorption spectrum of **Ib** recorded at $1 \cdot 10^{-2}$ M concentration in acetonitrile.

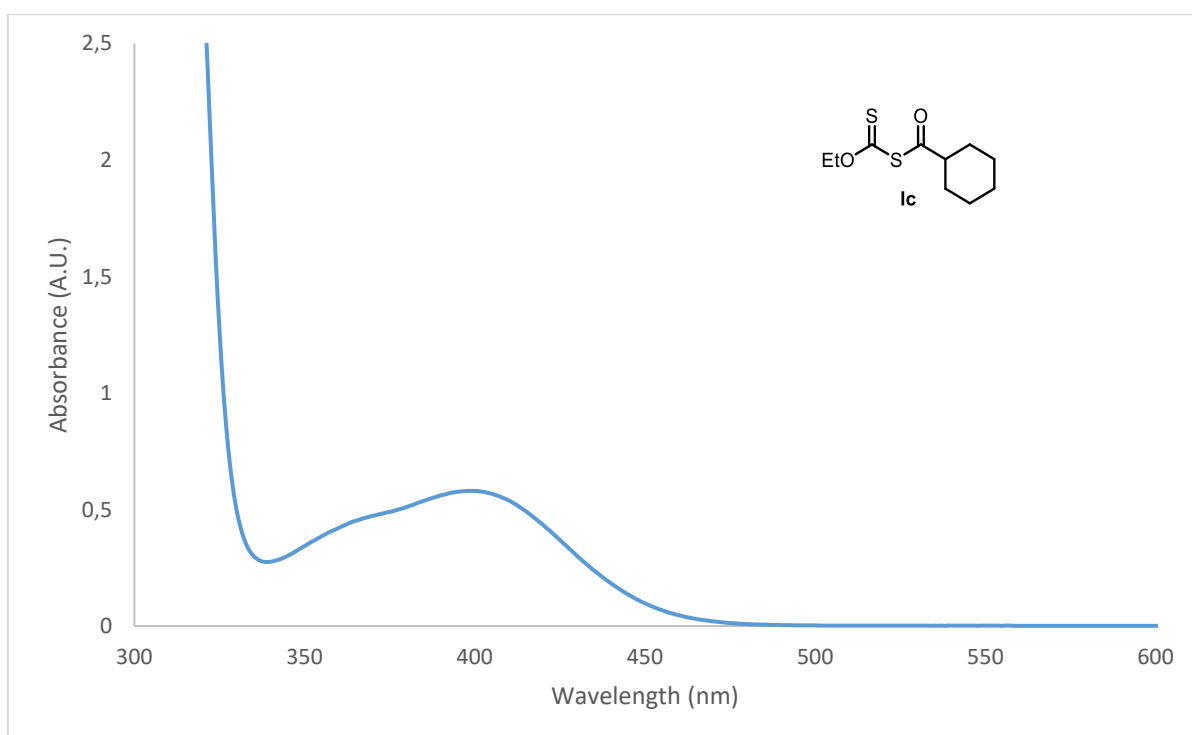


Figure S12: UV-Vis absorption spectrum of **Ic** recorded at $2 \cdot 10^{-2}$ M concentration in acetonitrile.

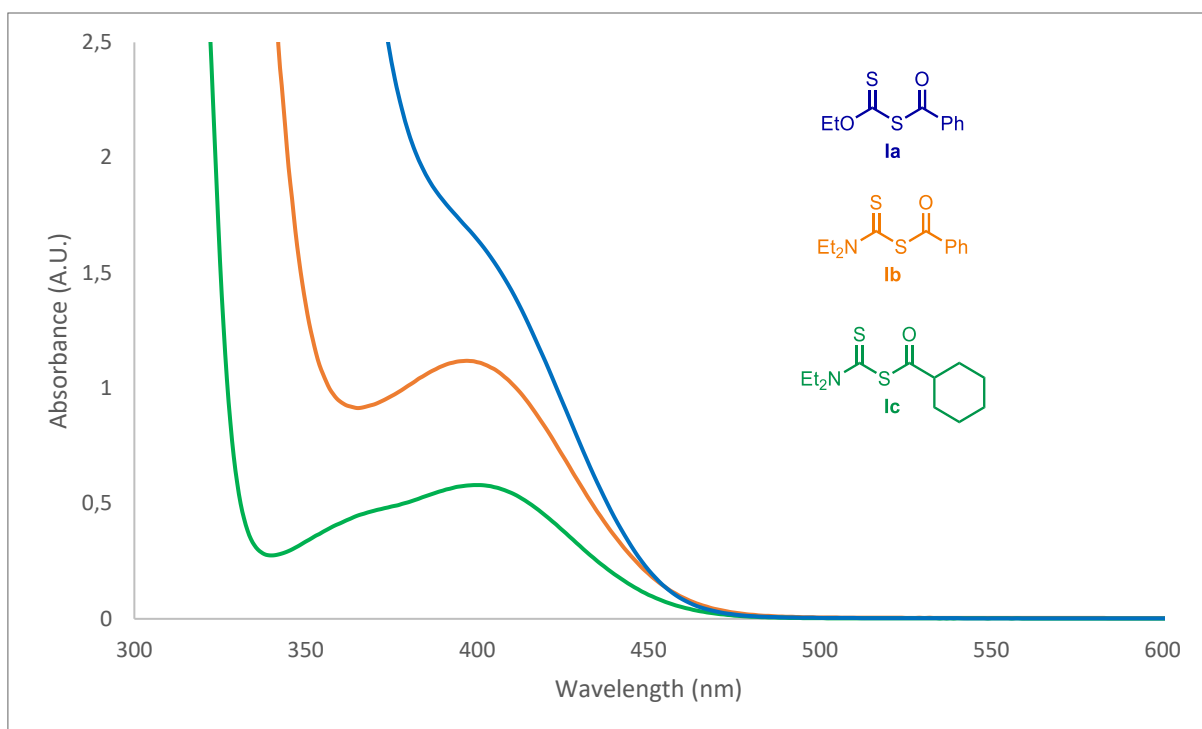


Figure S13: Superposition of the absorption spectra of the different acyl xanthates and acyl dithiocarbamate intermediates at the same concentration ($2 \cdot 10^{-2}$ M in acetonitrile).

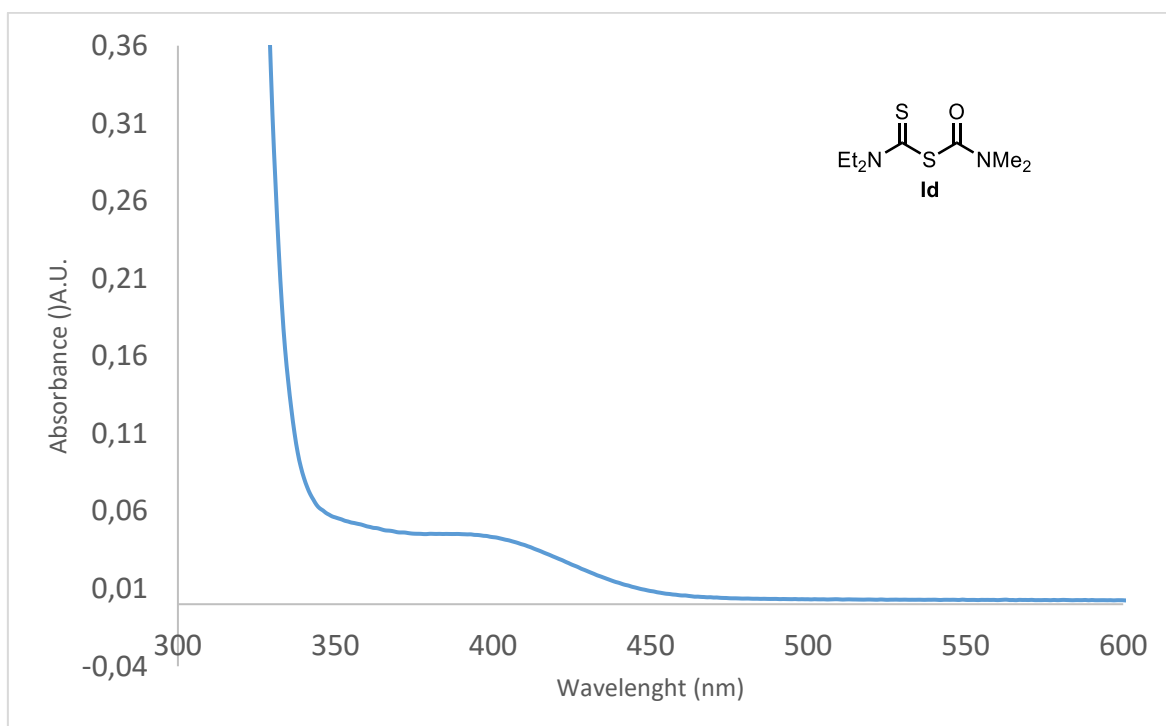


Figure S14: UV-Vis absorption spectrum of intermediate **Id** recorded at $2 \cdot 10^{-3}$ M concentration in acetonitrile.

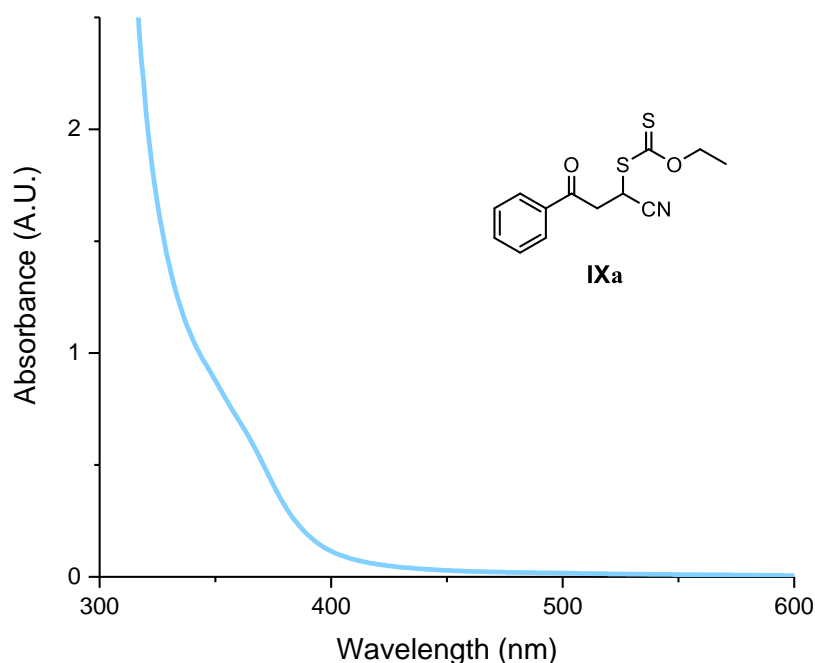
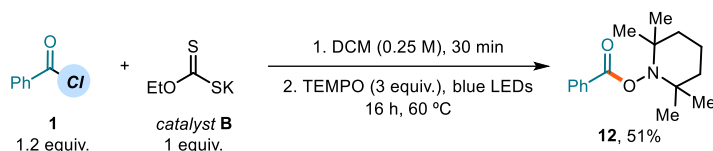


Figure S15: UV-Vis absorption spectrum of **IXa** recorded at $1 \cdot 10^{-2}$ M in acetonitrile

D1.4 TEMPO Trapping Experiments.

Stoichiometric reaction between TEMPO and in-situ acyl xanthate intermediate



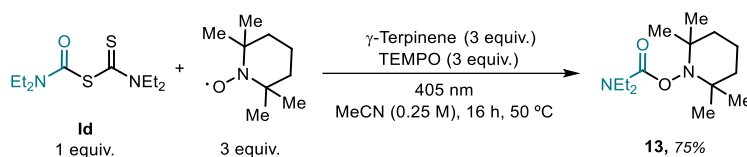
In an oven dried tube of 15 mL (16 mm × 125 mm) with a Teflon septum screw cap, potassium ethyl xanthogenate **B** (80.1 mg, 0.5 mmol, 1 equiv.) was suspended in DCM (2 mL, HPLC grade). Then, benzoyl chloride **1** (69.6 μ L, 0.6 mmol, 1.2 equiv.) was added and the mixture was stirred at ambient temperature for 30 minutes. Then, TEMPO (234.4 mg, 1.5 mmol, 3 equiv.) was added and the resulting yellow solution was degassed with argon sparging for 60 seconds. The tube was then placed in the temperature controlled photoreactor (Figure S4) set at a temperature of 60 °C (60–61°C measured in the central well) and irradiated for 16 hours. Chromatography on silica gel (5% AcOEt in hexanes as eluent) afforded adduct **12** (66 mg, yellow oil, 51% yield).

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 8.35 (d, $J = 6.9$ Hz, 2H), 7.83 (m, 1H), 7.74 (m, 2H), 2.12 – 1.70 (m, 6H), 1.55 (s, 6H), 1.40 (s, 6H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 161.45, 128.17, 124.89, 124.74, 123.78, 55.52, 34.28, 27.19, 16.09, 12.26.

Matching reported literature data.²³

Stoichiometric reaction between TEMPO and dithiocarbamate intermediate **Id**

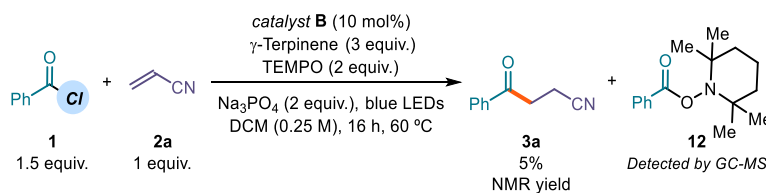


In an oven-dried 15 mL Schlenk tube was added a 0.25 M solution of the dithiocarbamate intermediate **Id** (28 mg, 127 μ mol, 1 equiv.) and TEMPO (60 mg, 0.38 mmol, 3 equiv.) in acetonitrile (0.5 mL). The reaction mixture was placed under an atmosphere of argon, cooled to -78 °C, degassed *via* vacuum evacuation (5 minutes), backfilled with argon and, ultimately, warmed to ambient temperature. This freeze-pump-thaw cycle was repeated four times, and then the Schlenk tube was sealed with Parafilm and put into the Hepatochem PhotoRedOx Box (Figure S8) equipped with a 405 nm EvoluChem LED 18 W light source at 50 °C. After 18 hours, the reaction vessel was cooled down to ambient temperature, water was added and the mixture was extracted with ethyl acetate (2x15 mL). The combined layers were dried over MgSO_4 , filtered, and concentrated. The resulting crude mixture was purified by column chromatography on silica gel (5% to 30% AcOEt in hexane) to give the corresponding product **11** (21 mg, 75% yield).

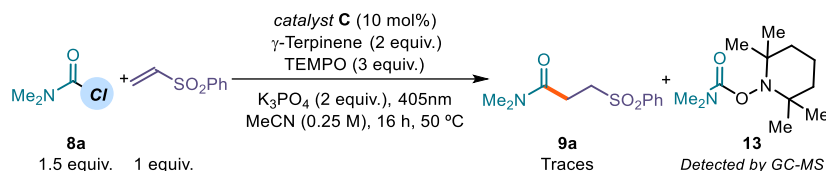
$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 2.96 (s, 6H), 1.73 – 1.48 (m, 6H), 3.06 (s, 3H), 1.13 (d, $J = 17.6$ Hz, 1H)

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 157.8, 60.2, 50.1, 39.1, 31.9, 21.1, 17.2.

TEMPO inhibition of the model reactions



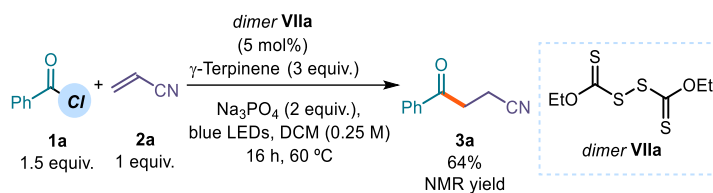
Reaction performed according to the general procedure A using benzoyl chloride (87 μ L, 0.75 mmol, 1.5 equiv.) and acrylonitrile (33 μ L, 0.5 mmol, 1 equiv.) and adding TEMPO (156.3 mg, 1 mmol, 2 equiv.) before the degassing step. The crude mixture was analyzed by $^1\text{H NMR}$ analysis after 16 hours using trichloroethylene (45 μ L, 0.5 mmol, 1 equiv.) as internal standard, and by GC-MS. NMR yield: 5%. Traces of the TEMPO adduct **12** were detected by GC-MS analysis.



Reaction performed according to the general procedure E using dimethylcarbamyl chloride (37 μ L, 0.4 mmol, 2 equiv.) and phenyl vinyl sulfone (34 mg, 0.2 mmol, 1 equiv.) and adding TEMPO (62 mg, 0.4 mmol, 2 equiv.) before the degassing step.

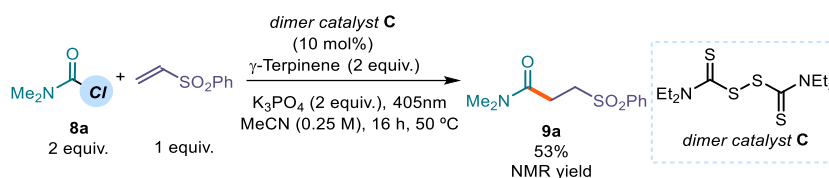
D1.5 Experiments with the Dimeric Catalysts.

Model reaction with catalyst dimer VIIa – Acylation



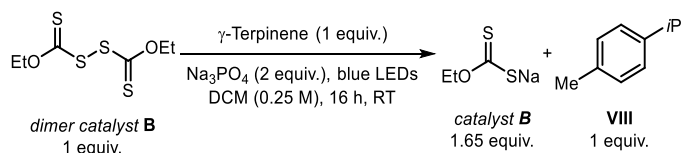
Reaction performed according to the general procedure A using benzoyl chloride (87 μL , 0.75 mmol, 1.5 equiv.) and acrylonitrile (33 μL , 0.5 mmol, 1 equiv.) while replacing *catalyst B* with dimer **VIIa** (6 mg, 0.025 mmol, 0.05 equiv.). The crude reaction mixture was analyzed by ^1H NMR analysis using trichloroethylene as internal standard. NMR yield: 64%.

Model reaction with dimer catalyst C - Carbamoylation



Reaction performed according to general procedure E using dimethylcarbamoyl chloride (37 μL , 0.4 mmol, 2.0 equiv.) and phenyl vinyl sulfone (34 mg, 0.2 mmol, 1 equiv.) while replacing *catalyst C* with dimer catalyst **C** (6 mg, 0.02 mmol, 0.1 equiv.). The crude reaction mixture was analyzed by ^1H NMR analysis using trichloroethylene as internal standard. NMR yield: 53%.

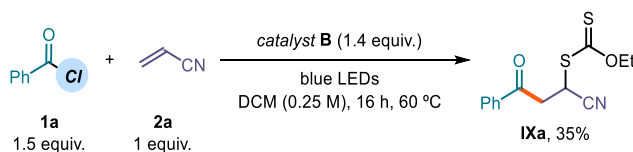
Turn-over experiment with dimer catalyst B and terpinene



In an oven dried vial (16 mm \times 50 mm) with a Teflon septum screw cap, dimer catalyst **B** (60.6 mg, 0.25 mmol, 1 equiv.) and sodium phosphate (82 mg, 0.5 mmol, 2 equiv.) were dissolved in DCM (2 mL, HPLC grade). Then, γ -terpinene (40 μL , 0.25 mmol, 1 equiv.) was added. The resulting yellow mixture was degassed with argon, sparging for 60 seconds. The vial was then placed in the 3D-printed support photoreactor (Figure S6) and irradiated for 24 hours. Trichloroethylene was added as internal standard and a sample of the crude mixture was diluted in d_6 -DMSO to record the NMR yield.

D1.6 Group Transfer Experiments.

Stoichiometric group transfer reaction with in-situ acyl xanthate intermediate



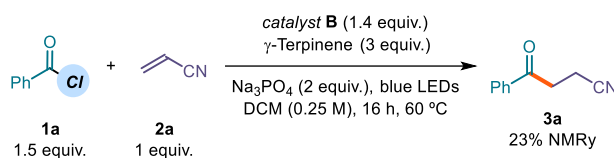
In an oven dried tube of 15 mL (16 mm \times 125 mm) with a Teflon septum screw cap, potassium ethyl xanthogenate **B** (112 mg, 0.7 mmol, 1.4 equiv.) was suspended in DCM (2 mL, HPLC

grade). Then, benzoyl chloride (87 μL , 0.75 mmol, 1.5 equiv.) was added and the mixture was stirred at ambient temperature for 30 min. The reaction mixture was then degassed with argon, sparging for 60 seconds. Finally, acrylonitrile (33 μL , 0.5 mmol, 1 equiv.) was added via syringe. The tube was then placed in the temperature controlled photoreactor (Figure S4) set at a temperature of 60 $^{\circ}\text{C}$ (60-61 $^{\circ}\text{C}$ measured in the central well) and irradiated for 16 hours. The crude mixture was purified by flash column chromatography on silica gel (5% to 10% AcOEt in hexanes as eluent) to afford **IXa** (50 mg, 35% yield) as a yellow oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.97 – 7.89 (m, 2H), 7.69 – 7.58 (m, 1H), 7.50 (app t, $J = 7.5$ Hz, 2H), 5.08 (t, $J = 6.2$ Hz, 1H), 4.72 (q, $J = 7.1$ Hz, 2H), 3.70 (d, $J = 6.3$ Hz, 2H), 1.47 (t, $J = 7.1$ Hz, 3H).

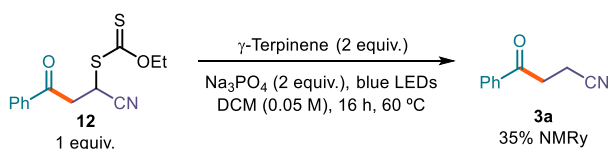
$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 209.05, 193.54, 135.35, 134.39, 129.09, 128.30, 117.96, 71.54, 40.67, 32.49, 13.84.

Stoichiometric group transfer reaction with in-situ acyl xanthate intermediate in the presence of γ -terpinene



In an oven dried tube of 15 mL (16 mm \times 125 mm) with a Teflon septum screw cap, potassium ethyl xanthogenate (112 mg, 0.7 mmol, 1.4 equiv.) and sodium phosphate (164 mg, 1.0 mmol, 2 equiv) were dissolved in DCM (2 mL, HPLC grade). Benzoyl chloride (87 μL , 0.75 mmol, 1.5 equiv.) was added and the mixture was stirred at ambient temperature for 30 min. Then, γ -terpinene (240 μL , 1.5 mmol, 3 equiv.) was added. The mixture was degassed via argon sparging for 60 seconds. Finally, acrylonitrile (33 μL , 0.5 mmol, 1 equiv.) was added via syringe. The tube was then placed in the temperature controlled photoreactor (Figure S4) set at a temperature of 60 $^{\circ}\text{C}$ (60-61 $^{\circ}\text{C}$ measured in the central well) and irradiated for 16 hours. Trichloroethylene was added as internal standard, and a sample of the crude mixture was diluted in CDCl_3 to record the NMR yield. *No group transfer product 12 was observed.*

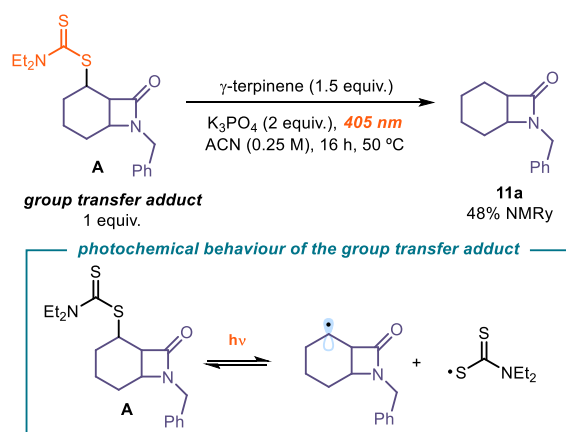
Direct photolysis of the group transfer product under reaction conditions



In an oven dried tube of 15 mL (16 mm \times 125 mm) with a Teflon septum screw cap, the group transfer product **12** (28.01 mg, 0.1 mmol, 1 equiv.) and sodium phosphate (33 mg, 0.2 mmol, 2 equiv) were dissolved in DCM (2 mL, HPLC grade). Then, γ -terpinene (33 μL , 0.2 mmol, 2 equiv.) was added. The reaction mixture was degassed with Argon sparging for 60 seconds. The tube was then placed in the temperature controlled photoreactor (Figure S4) set at a temperature of 60 $^{\circ}\text{C}$ (60-61 $^{\circ}\text{C}$ measured in the central well) and irradiated for 16 hours.

Trichloroethylene was added as internal standard and a sample of the crude mixture was diluted in CDCl_3 to record the NMR yield – product **3a** was formed in 35%.

Direct photolysis of the group transfer product for the intramolecular reaction



An oven-dried 15 mL Schlenk tube was charged with an authentic sample of the group transfer adduct **A**, prepared according to Ref. 2, γ -terpinene (48 μ L, 0.3 mmol, 1.5 equiv.) and K_3PO_4 (85 mg, 0.4 mmol, 2 equiv.) in acetonitrile (0.8 mL, 0.25 M). The reaction mixture was placed under an atmosphere of argon, cooled to -78 °C, degassed *via* vacuum evacuation (5 minutes), backfilled with argon and, ultimately, warmed to ambient temperature. This freeze-pump-thaw cycle was repeated four times, and then the Schlenk tube was sealed with Parafilm and put into the Hepatochem PhotoRedOx Box equipped with a 405 nm EvoluChem LED 18 W light source at 50 °C (Figure S8). After 18 hours stirring, the reaction was cooled down to ambient temperature, trichloroethylene was added as internal standard and a sample of the crude mixture was diluted in $CDCl_3$ to record the NMR of the crude - product **11a** was formed in 48%.

D2. Cyclic Voltammetry Measurements

For the cyclic voltammetry (CV) measurements, a glassy carbon disk electrode (diameter: 3 mm) was used as a working electrode. A silver wire coated with AgCl immersed in a 3.5 M aqueous solution of KCl and separated from the analyte by a fritted glass disk was employed as the reference electrode. A Pt wire counter-electrode completed the electrochemical setup. The scan rate of used in each CV experiment is indicated case by case.

Potentials are quoted with the following notation: E_p^C refers to the cathodic peak potential, E_p^A refers to the anodic peak potential, while the E^{red} value describes the electrochemical properties of the referred compound.

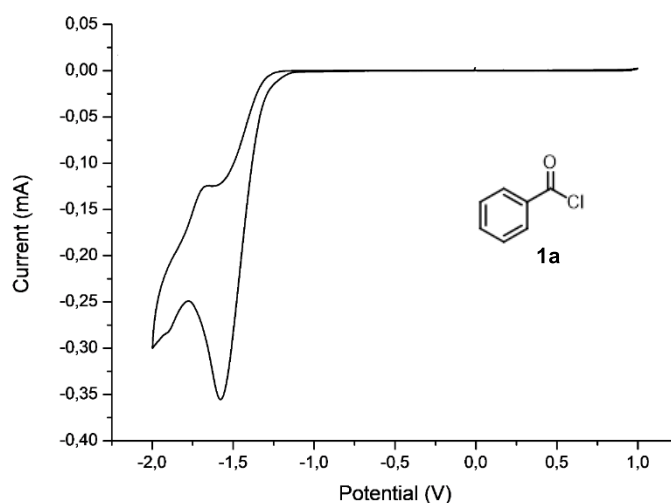


Figure S16: Cyclic voltammogram of benzoyl chloride **1a** [0.02 M] in [0.1 M] TBAPF₆ in CH₃CN. Sweep rate: 100 mV/s. Glassy carbon electrode working electrode, Ag/AgCl (KCl 3.5 M) reference electrode, Pt wire auxiliary electrode. Irreversible reduction, $E_p^C = E^{red}(\mathbf{1a}/\mathbf{1a}^{\cdot-}) = -1.57$ V.

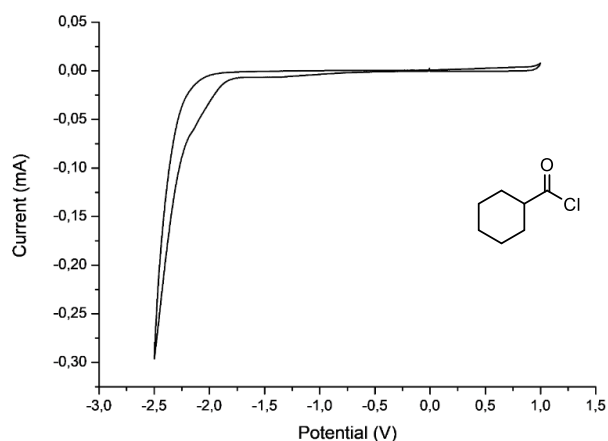


Figure S17: Cyclic voltammogram of cyclohexanecarbonyl chloride [0.02 M] in [0.1 M] TBAPF₆ in CH₃CN. Sweep rate: 100 mV/s. Glassy carbon electrode working electrode, Ag/AgCl (KCl 3.5 M) reference electrode, Pt wire auxiliary electrode. Reduction of cyclohexanecarbonyl chloride was not observed in the registered potential window (from 0 to -2.50 V).

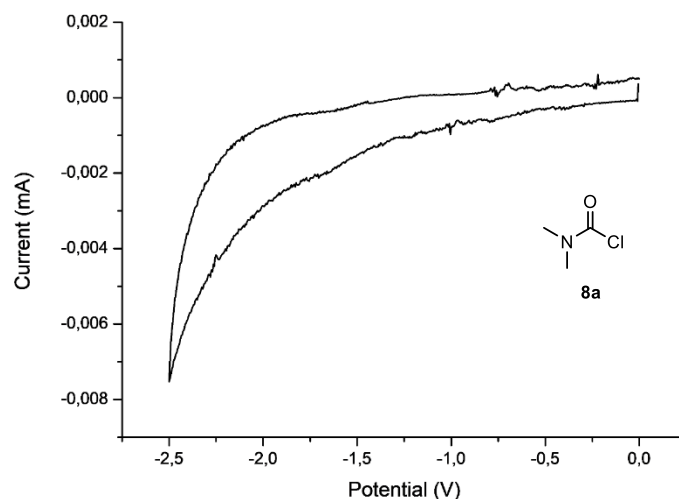


Figure S18: Cyclic voltammogram for **8a** [0.02M] in [0.1 M] TBAPF₆ in CH₃CN. Sweep rate: 50 mV/s. Glassy carbon electrode working electrode, Ag/AgCl (KCl 3.5 M) reference electrode, Pt wire auxiliary electrode. Reduction of **8a** was not observed in the registered potential window (from 0 to -2.50 V).

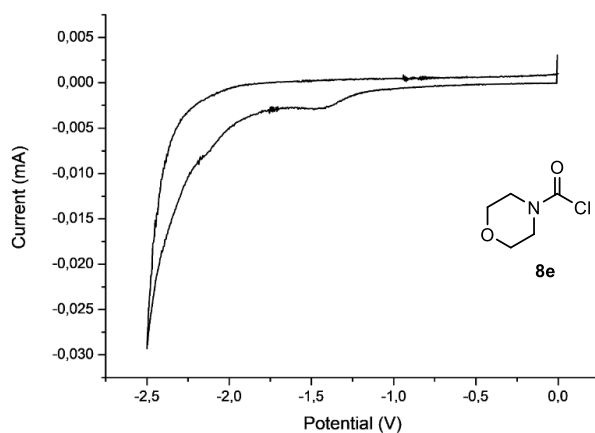


Figure S19: Cyclic voltammogram for **8e** [0.02M] in [0.1 M] TBAPF₆ in CH₃CN. Sweep rate: 100 mV/s. Glassy carbon electrode working electrode, Ag/AgCl (KCl 3.5 M) reference electrode, Pt wire auxiliary electrode. Reduction of **8e** was not observed in the registered potential window (from 0 to -2.50 V).

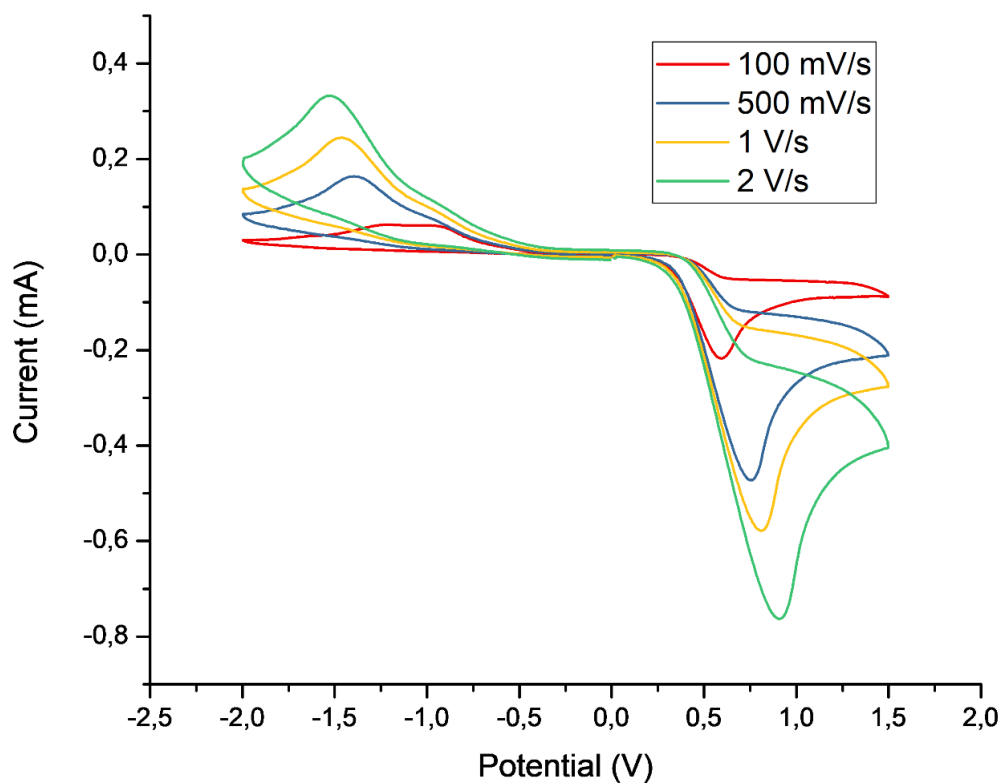


Figure S20: Cyclic voltammogram for catalyst **B** [0.02 M] in [0.1 M] TBAPF₆ in CH₃CN. Measurement started by oxidation from 0 to +1.5 V, followed by reduction from +1.5 V to -2.0 V, and finishing at 0 V. Glassy carbon electrode working electrode, Ag/AgCl (KCl 3.5 M) reference electrode, Pt wire auxiliary electrode. Two irreversible peaks observed increasing with sweep rate.

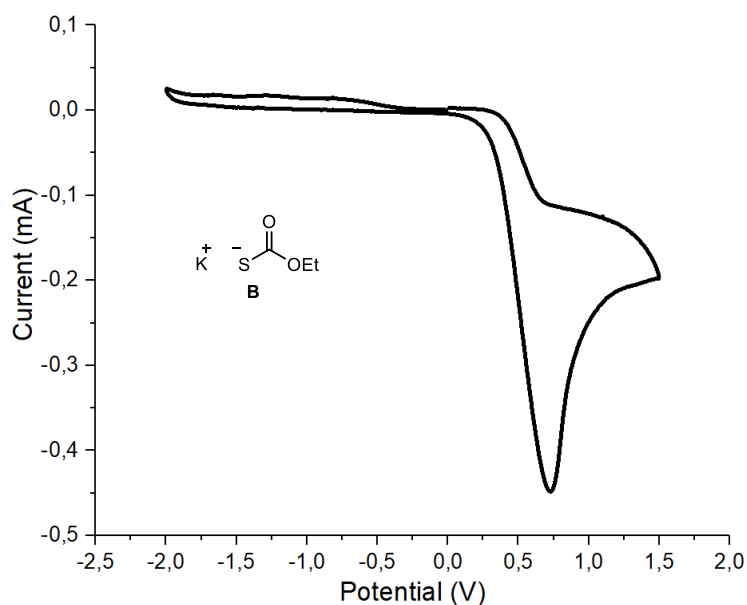


Figure S21: Cyclic voltammogram for catalyst **B** [0.02M] in [0.1 M] TBAPF₆ in CH₃CN. Measurement started by reduction from 0 to -2.0 V, followed by oxidation from -2.0 V to +1.5 V, and finishing at 0 V. Glassy carbon electrode working electrode, Ag/AgCl (KCl 3.5 M) reference electrode, Pt wire auxiliary electrode. Only one irreversible peak observed. Sweep rate: 500 mV/s.

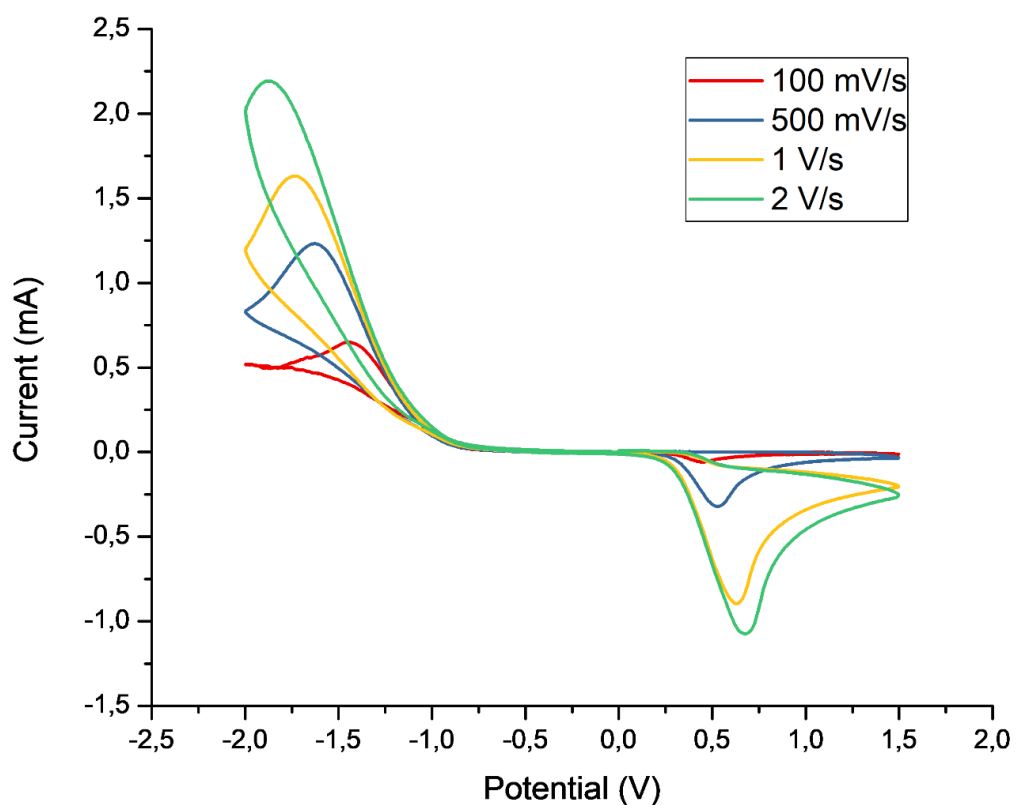


Figure S22: Cyclic voltammogram for dimer **VIIa** [0.02 M] in [0.1 M] TBAPF₆ in CH₃CN. Measurement started by reduction from 0 to -2.0 V, followed by oxidation from -2.0 V to +1.5 V, and finishing at 0 V. Glassy carbon electrode working electrode, Ag/AgCl (KCl 3.5 M) reference electrode, Pt wire auxiliary electrode. Two irreversible peaks observed increasing with sweep rate.

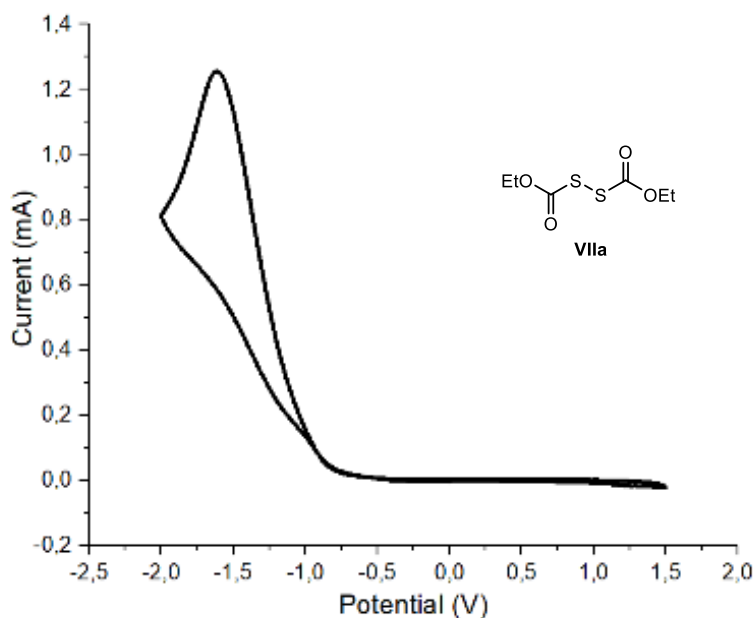


Figure S23: Cyclic voltammogram for dimer **VIIa** [0.02M] in [0.1 M] TBAPF₆ in CH₃CN. Measurement started by oxidation from 0 to +1.5 V, followed by reduction from +1.5 V to -2.0 V, and finishing at 0 V. Glassy carbon electrode working electrode, Ag/AgCl (KCl 3.5 M) reference electrode, Pt wire auxiliary electrode. Two irreversible peaks observed increasing with sweep rate.

D3. Transient Absorption Spectroscopy (TAS).

Studies with microsecond transient absorption spectroscopy (TAS) were performed using an excitation source of NdYAG (neodymium-doped yttrium aluminium garnet) Opolette laser with an optical parametric oscillator (OPO) system that allows variable wavelength excitation from 400 -1800 nm, pulse width of 6 ns, up to 2 mJ of energy from OPO output with fiber optic coupled, and high energy output from direct NdYAG harmonics 355 (20 mJ, 5 ns) and 532 (45mJ, 6 ns). The system is completed with 150 W tungsten lamp as probe; 2 monochromators Minuteman MM151; Si amplified photodetector module for VIS; DSPDAU high speed data rate recorder and interface software from RAMDSP. Laser intensities for each wavelength were the following: 355 nm – 1.30 mJ; 420 nm– 1.20 mJ; 460 nm – 1.95 mJ.

Several studies with different wavelengths and laser intensities were carried out, each of the conditions are indicated in a case by case bases. We selected a logarithmic time scale suitable for clearly showing the decay of the transient species in the samples. The characteristics of the detected transient species match literature data.²⁴

In a typical transient absorption spectroscopy experiment, solutions in acetonitrile of each of the substrates were prepared under an argon atmosphere and transferred into a screw-top 3.0 mL quartz cuvette for measurement. Upon irradiation with the appropriated wavelength, the decay of absorption at 620 nm of the transient xanthyl radical **IIIa** was recorded.

Acylxanthate **Ia**

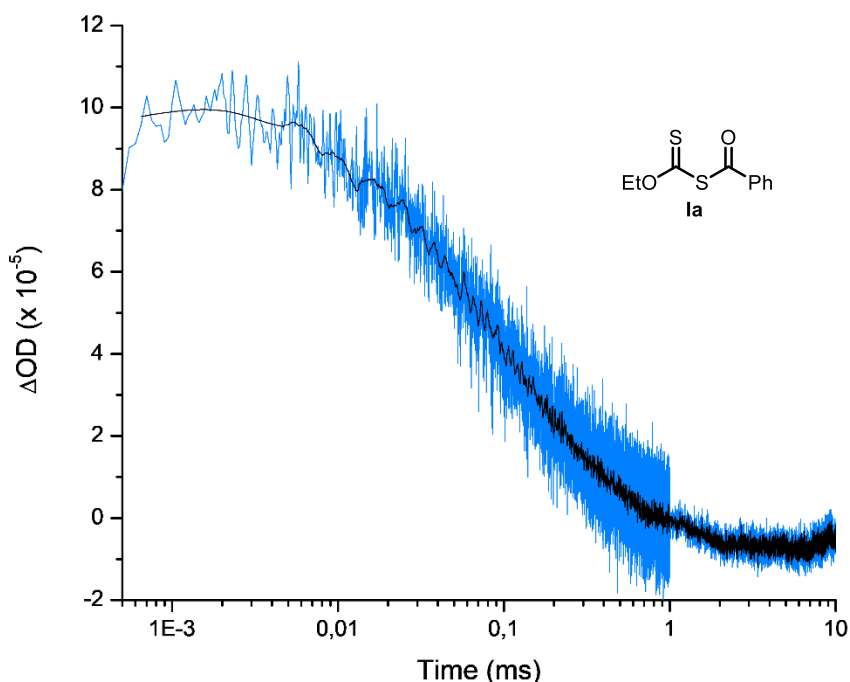


Figure S24: Absorption at 620 nm of the transient xanthyl radical **IIIa** (blue line) generated upon 355 nm laser excitation of Acylxanthate **Ia** ($[Ia]_0 = 3.00$ mM in acetonitrile). Note logarithmic scale for time. Absorption decay (black line) processed through Savinsky Golay filter to facilitate lifetime measurement. ΔOD : optical density variation.

Compound **Ia** was also measured upon 460 nm excitation in order to mimic the conditions of photolysis under catalytic conditions. Since photolysis of **Ia** is less efficient at longer wavelengths, a higher concentration of **Ia** was needed to obtain a comparable scale signal. Note that changes in concentration of both **Ia** and transient **IIIa** generated upon photolytic cleavage directly affects the lifetime of the detected species.

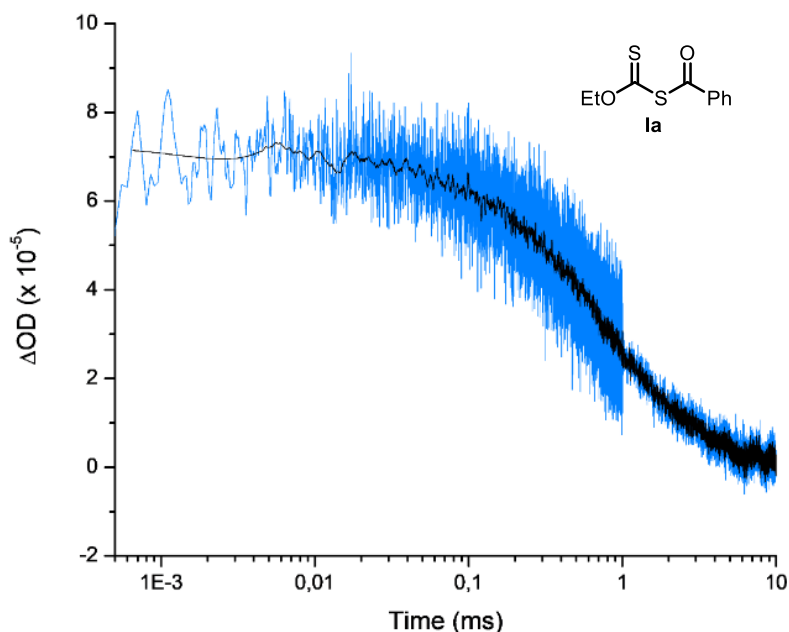


Figure S25: Absorption at 620 nm of the transient xanthy radical **IIIa** (blue line) generated upon 460 nm laser excitation of acylxanthate **Ia** ($[\mathbf{Ia}]_0 = 300$ mM in acetonitrile). Note logarithmic scale for time. Absorption decay (black line) processed through Savinsky Golay filter to facilitate lifetime measurement. ΔOD : optical density variation.

Dimer **VIIa**

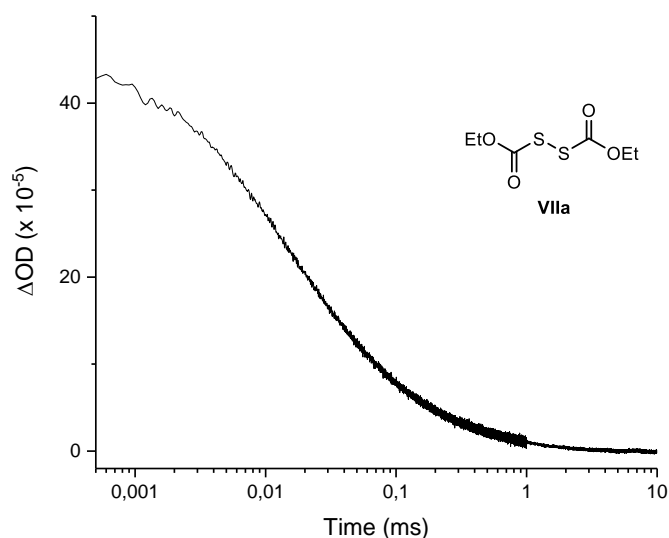


Figure S26: Absorption at 620 nm of the transient xanthy radical **IIIa** (black line) generated upon 355 nm laser excitation of dimer **VIIa** ($[\mathbf{VIIa}]_0 = 3.00$ mM in acetonitrile). Note logarithmic scale for time. ΔOD : optical density variation.

Dimer **VIIa** was also measured upon 420 nm and 460 nm irradiation in order to support photolysis under the reaction conditions. A higher concentration of **VIIa** was used to ensure comparable scale signal.

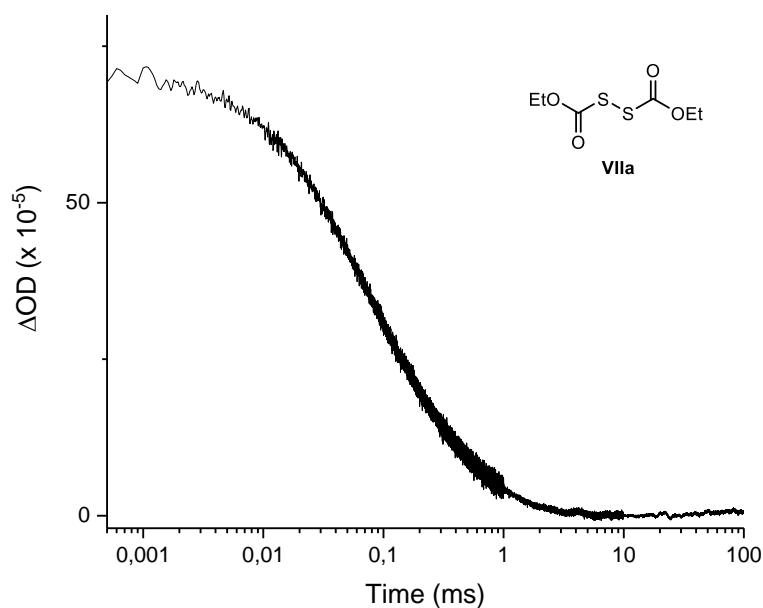


Figure S27: Absorption at 620 nm of the transient xanthyl radical **IIIa** (black line) generated upon 420 nm laser excitation of dimer **VIIa** ($[\text{VIIa}]_0 = 300 \text{ mM}$ in acetonitrile). Note logarithmic scale for time. ΔOD : optical density variation.

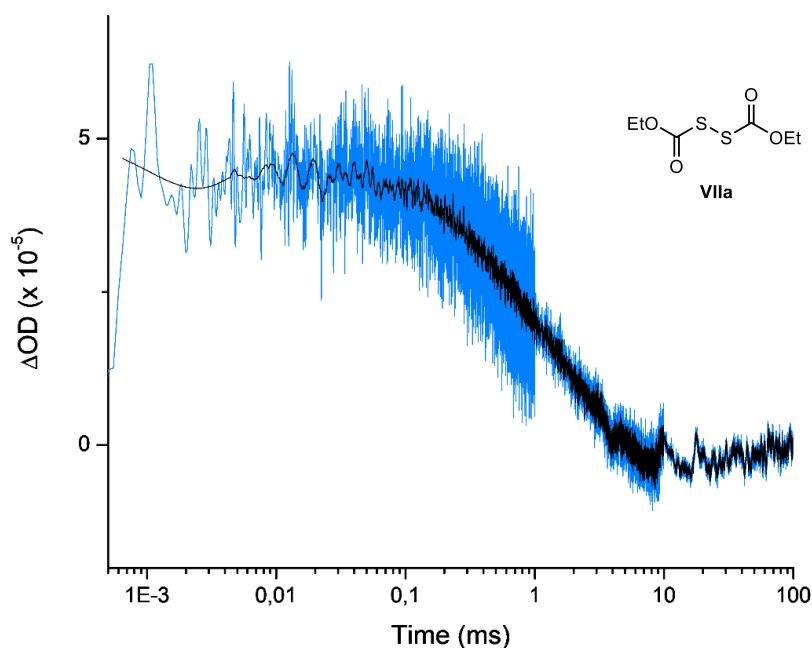


Figure S28: Absorption at 620 nm of the transient xanthyl radical **IIIa** (blue line) generated upon 460 nm laser excitation of dimer **VIIa** ($[\text{VIIa}]_0 = 300 \text{ mM}$ in acetonitrile). Note logarithmic scale for time. Absorption decay (black line) processed through Savitsky Golay filter to facilitate lifetime measurement. ΔOD : optical density variation.

In order to perform the quenching experiment, increasing amounts of pure γ -terpinene was added while observing the effect on the absorption at 620 nm of the transient **IIIa**, which was recorded after every addition. Increasing amounts of pure γ -terpinene (up to 60 equivalents, 60 μL) were added sequentially to a 2 mL solution of **VIIa** (3 mM in acetonitrile) in a screw-top quartz cuvette, providing a final concentration of 2.91 mM. A decay of absorption of the transient xanthyl radical,

and therefore a shorter lifetime, was observed upon addition of γ -terpinene (Figure S29), which is consonant with a reaction between the two species. Turquoise line: ratio **VIIa**/ γ -terpinene mimics the reaction conditions.

Note that precipitation of a solid, associated to ethyl xanthogenate, was observed upon addition and irradiation of the sample.

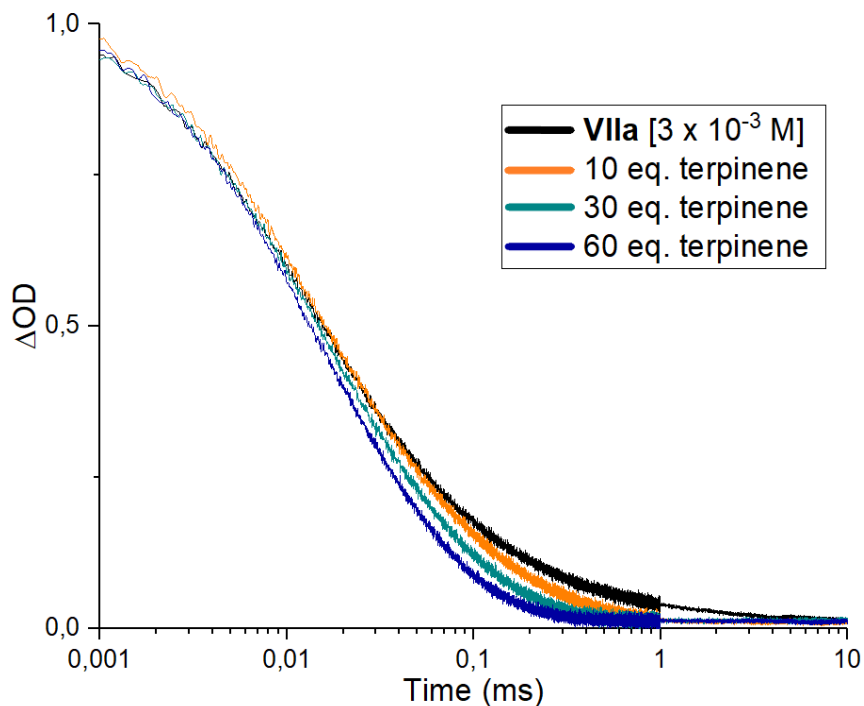


Figure S29: Absorption at 620 nm of the transient xanthyl radical **IIIa** (black line) generated upon 355 nm laser excitation of dimer **VIIa** ($[\text{VIIa}]_0 = 3 \text{ mM}$ in acetonitrile) and subsequent decay of the absorption upon addition of 10 (orange line), 30 (green line, mimics proportions under reaction conditions) and 60 (blue line) equivalents of γ -terpinene, respectively. Note logarithmic scale for time. Absorption decay was normalized to 1. Δ OD: normalized optical density variation.

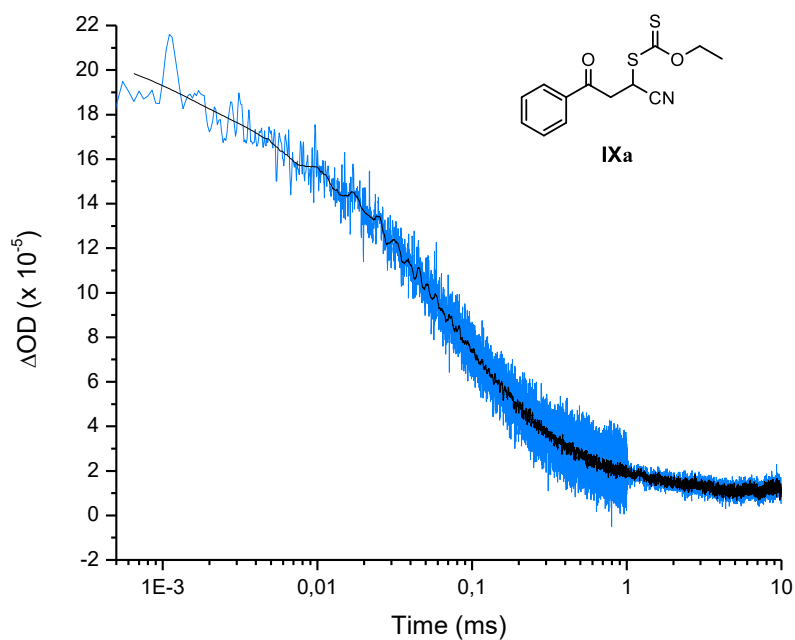


Figure S30: Absorption at 620 nm of the transient xanthyl radical **IIIa** (blue line) generated upon 355 nm laser excitation of **IXa** ($[\text{IXa}]_0 = 3 \text{ mM}$ in acetonitrile). Note logarithmic scale for time. Absorption decay (black line) processed through Savinsky Golay filter to facilitate lifetime measurement. ΔOD : optical density variation.

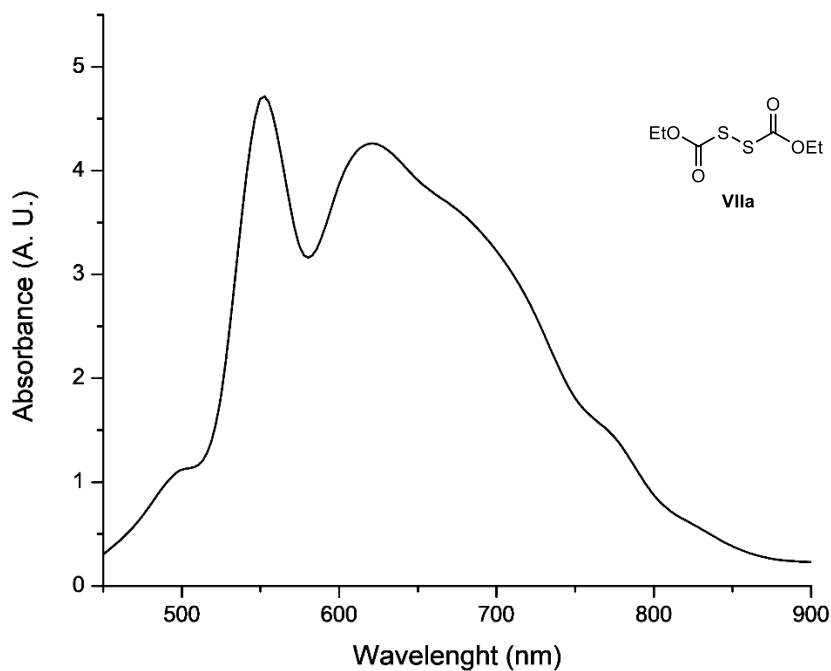


Figure S31: Absorption spectra of the transient xanthyl radical **IIIa** generated upon 355 nm laser excitation of Dimer **VIIa** ($[\text{VIIa}]_0 = 3 \text{ mM}$ in acetonitrile) at $1 \mu\text{s}$ time of irradiation. Maximum characteristic from xanthyl radical can be observed around 625 nm.

D4. Electron paramagnetic resonance (EPR)

EPR spectra were acquired on a Bruker EMX X-band EPR spectrometer with an ER 4116 HS cavity (9.86 GHz at room temperature) using 100 kHz field modulation (modulation amplitude: 1 G). Individual EPR tubes were filled with ~0.7 mL of the solution and were placed at the same position of the resonant cavity for EPR spectral acquisition. The spectral data were collected at 298 K with the following spectrometer settings: microwave power = 2.020 mW; center field = 3518 G, sweep width = 200 G, sweep time = 30 s, modulation frequency = 100 KHz, modulation amplitude = 1 G, power attenuation = 20 dB, time constant = 0.01 ms.

A fresh solution of acylxanthate **Ia** 0.10 M in Toluene was prepared under air and measured without further precautions to remove oxygen from the solution. As expected, no signal was observed before of irradiation (note that **Ia** decomposes rapidly, and a sample older than one day did show signals appearing before irradiation, due to decomposition); on the other hand, upon irradiation of the sample, appearance of a triplet at 3505 G was observed with a g-value of 2.00272 and a hyperfine splitting value α_H (2.6, 2H, γ -H). This signal reaches a maximum of intensity after 12.5 minutes of irradiation. The calculated EPR spectrum for the carbon radical of type **VI**, which lies in proximity of two sulfur atoms and an ethoxy moiety, is shown in the right panel of Figure S32.

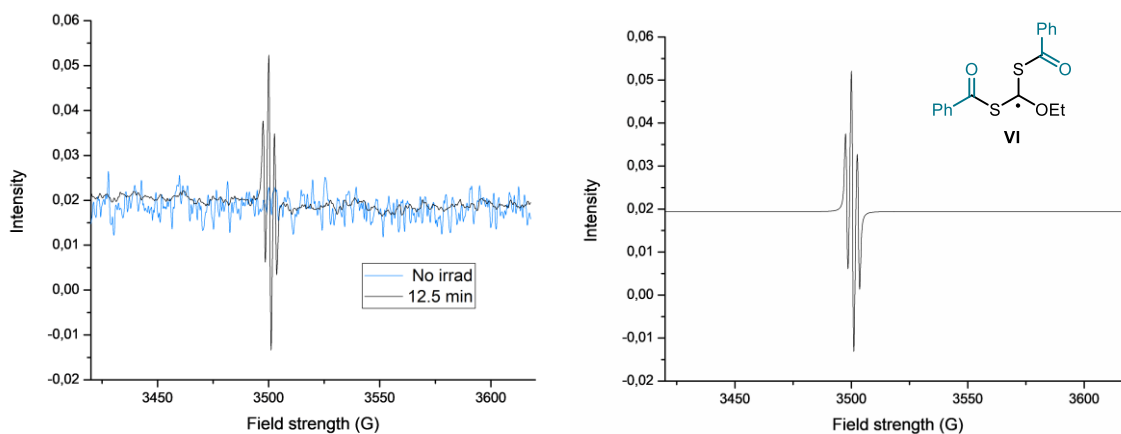


Figure S32: Comparison between (left) EPR spectra of acylxanthate **Ia** (0.1 M in toluene) before irradiation (blue line) and after irradiation with a LSB610 100W mercury lamp during 12.5 min (black line). Open-shell specie was detected by appearance of a new signal centered at 3505 G (triplet); and (right) calculated EPR spectrum for intermediate **VI** for a hyperfine coupling with two equivalent nuclei of spin $\frac{1}{2}$.

D5. Quantum Yield Determination

A ferrioxalate actinometer solution was prepared by following the Hammond variation of the Hatchard and Parker procedure outlined in the Handbook of Photochemistry.²⁵ The ferrioxalate actinometer solution measures the decomposition of ferric ions to ferrous ions, which are complexed by 1,10-phenanthroline and monitored by UV/Vis absorbance at 510 nm. The moles of iron-phenanthroline complex formed are related to moles of photons absorbed. The following solutions were prepared and stored in a dark laboratory (red light):

1. Potassium ferrioxalate solution: 294.8 mg of potassium ferrioxalate (commercially available from Alfa Aesar) and 139 μ L of sulfuric acid (96%) were added to a 50 mL volumetric flask, and filled to the mark with water (HPLC grade).

2. Phenanthroline solution: 0.2% by weight of 1,10-phenanthroline in water (100 mg in 50 mL volumetric flask).

3. Buffer solution: 2.47 g of NaOAc and 0.5 mL of sulfuric acid (96%) were added to a 50 mL volumetric flask, and filled to the mark with water (HPLC grade).

The actinometry measurements were done as follows:

1. 1 mL of the actinometer solution was added to a screw-cap vial and placed on a single HP LED 1.5 cm away from the light source. The solution was irradiated at 460 nm (irradiance 40 mW/cm²). This procedure was repeated 4 times, quenching the solutions after different time intervals: 10 sec, 15 sec, 20 sec, and 25 sec.

2. After irradiation, the actinometer solutions were removed and placed in a 10 mL volumetric flask containing 0.5 mL of 1,10-phenanthroline solution and 2 mL of buffer solution. These flasks were filled to the mark with water (HPLC grade).

3. The UV-Vis spectra of the complexed actinometer samples were recorded for each time interval. The absorbance of the complexed actinometer solution was monitored at 510 nm.

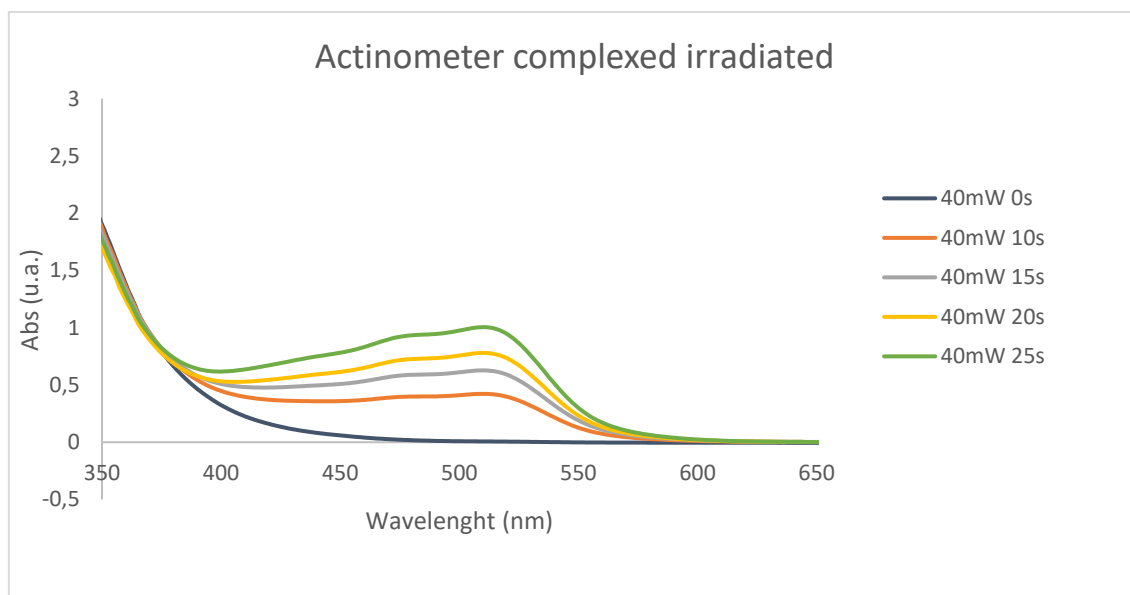


Figure S33: Absorbance of the complexed actinometer solutions.

The moles of Fe²⁺ formed for each sample is determined using Beers' Law (Eq. 1) :

$$\text{Mols of Fe(II)} = V_1 \times V_3 \times \Delta A(510 \text{ nm}) / 10^3 \times V_2 \times l \times \varepsilon(510 \text{ nm}) \quad (\text{Eq. 1})$$

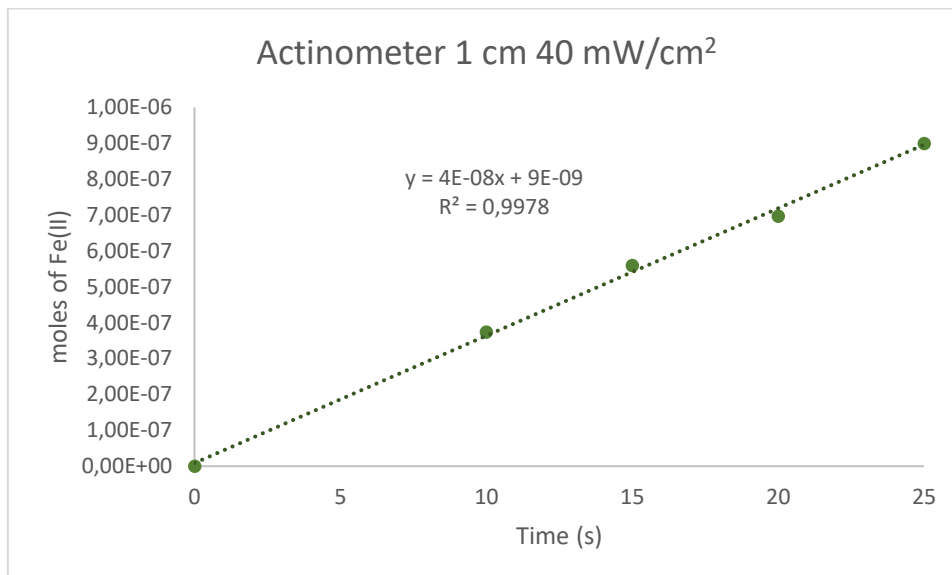
where V_1 is the irradiated volume (1 mL), V_2 is the aliquot of the irradiated solution taken for the determination of the ferrous ions (1 mL), V_3 is the final volume after complexation with phenanthroline (10 mL), l is the optical path-length of the irradiation cell (1 cm), $\Delta A(510 \text{ nm})$ is the optical difference in absorbance between the irradiated solution and the one stored in the dark, $\varepsilon(510 \text{ nm})$ is the extinction coefficient the complex Fe(phen)₃²⁺ at 510 nm (11100 L mol⁻¹ cm⁻¹). The moles of Fe²⁺ formed (x) are plotted as a function of time (t). The slope of this line was correlated to the moles of incident photons by unit of time ($q_{n,p}^0$) by the use of the following Equation 2:

$$\Phi(\lambda) = dx/dt q_{n,p}^0 [1 - 10^{-A(\lambda)}] \quad (\text{Eq. 2})$$

where dx/dt is the rate of change of a measurable quantity (spectral or any other property), the quantum yield (Φ) for Fe²⁺ at 458 nm is 1.1⁴⁵, $[1 - 10^{-A(\lambda)}]$ is the ratio of absorbed photons by the

solution, and $A(\lambda)$ is the absorbance of the actinometer at the wavelength used to carry out the experiments (460 nm). The absorbance at 460 nm $A(460)$ was measured using a Shimadzu 2401PC UV-Vis spectrophotometer in a 1 cm path quartz cuvette, obtaining an absorbance of 0.158.

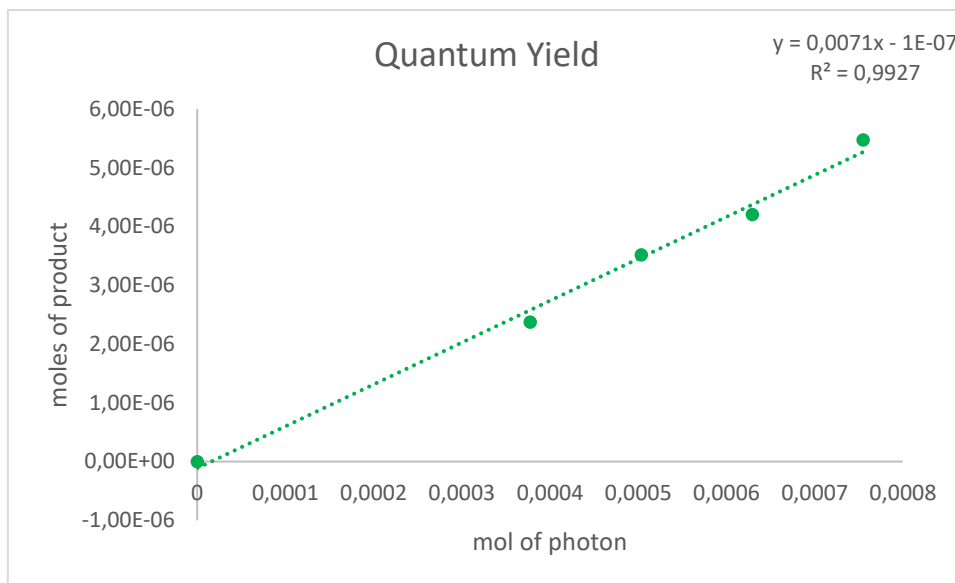
q_n^0 , which is the photon flux, was determined to be 1.048×10^{-7} einstein s^{-1} .



Consequently, the model reactions were performed.

The reactions were prepared on a screw-cap vial with stir bar. Cyclohexanecarbonyl chloride (50 μ L, 0.375 mmol, 1.5 equiv.), γ -terpinene (120 μ L, 0.75 mmol, 3 equiv.), and lutidine (58 μ L, 0.5 mmol, 2 equiv.), were added to a solution of catalyst **B** (4 mg, 0.1 equiv.) in acetonitrile (1 mL). After degassing by bubbling Argon for 30 s, acrylonitrile **2a** (16 μ L, 0.25 mmol) was added and the tube was sealed with parafilm and put in the HP-LED 460 nm at 1 cm distance at ambient temperature (reaction reaches around 35 $^{\circ}$ C) with irradiance of 40 mW/cm^2 . Four different reactions were setup and irradiated for different times: 60 min, 80 min, 100 min and 120 min.

The moles of product **5e** formed for the model reaction were determined by GC measurement (FID detector) using 1,3,5-trimethoxybenzene as internal standard. The moles of product per unit of time are related to the number of photons absorbed. The photons absorbed are correlated to the number of incident photons by the use of Equation 1. According to this, plotting the moles of product (x) versus the moles of incident photons ($q_n^0 \cdot dt$), the slope is equal to: $\Phi \cdot (1 - 10^{-A(\lambda)(460 \text{ nm})})$, where Φ is the quantum yield to be determined and $A(460 \text{ nm})$ is the absorption of the reaction under study. $A(460 \text{ nm})$ was measured using a Shimadzu 2401PC UV-Vis spectrophotometer in 10 mm path quartz. An absorbance of 0.103 was determined for the model reaction mixture. The quantum yield (Φ) of the photochemical transformation was measured to be 0.0338. The procedure was repeated a second time to provide a similar value: quantum yield (Φ) at 460 nm of 0.0332.

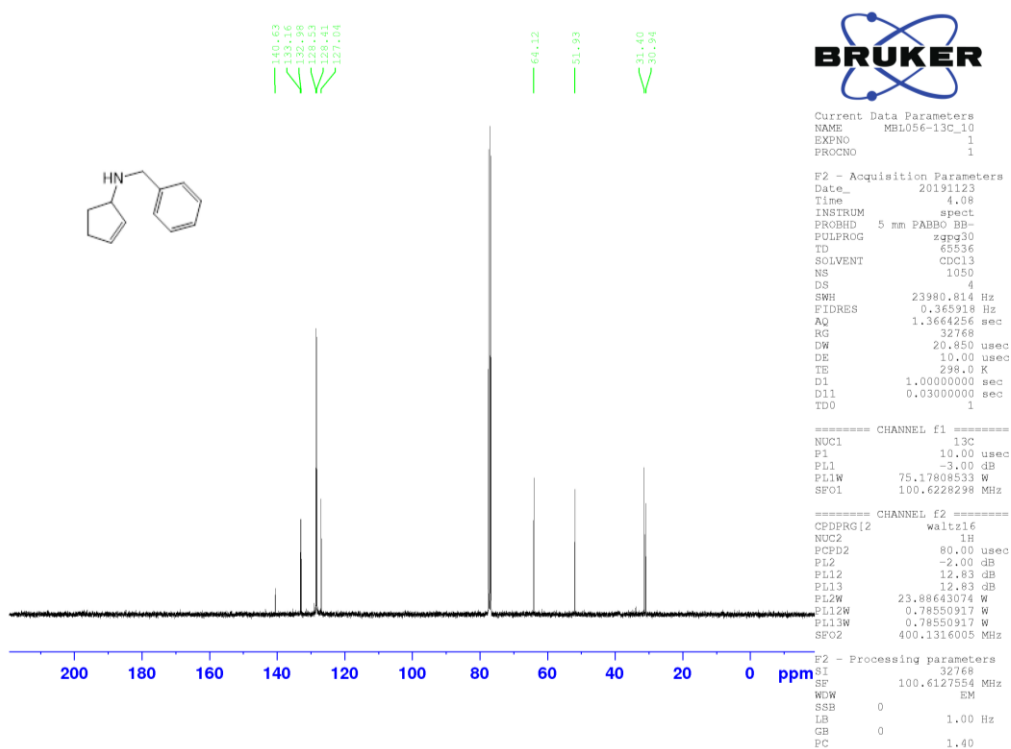
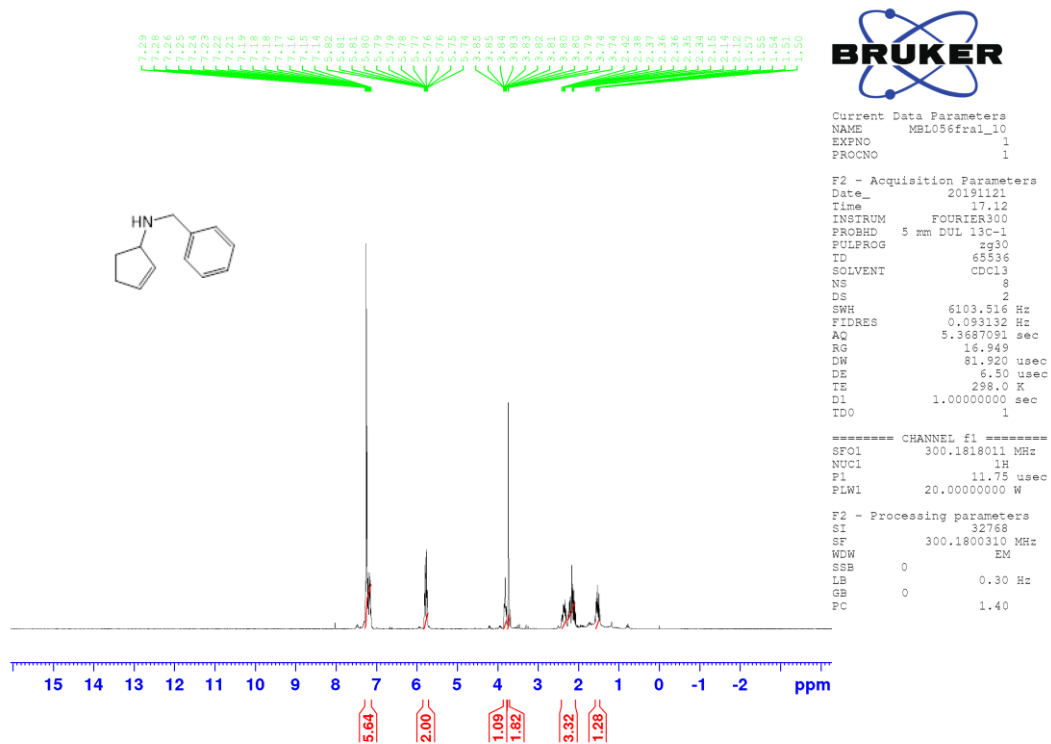


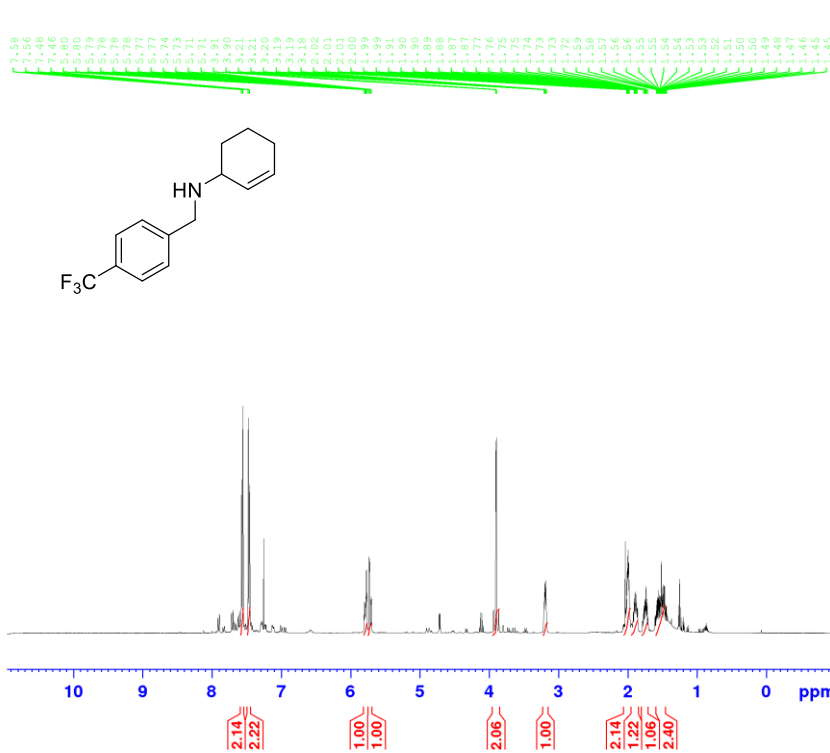
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F. NMR Spectra

F1. Starting Materials



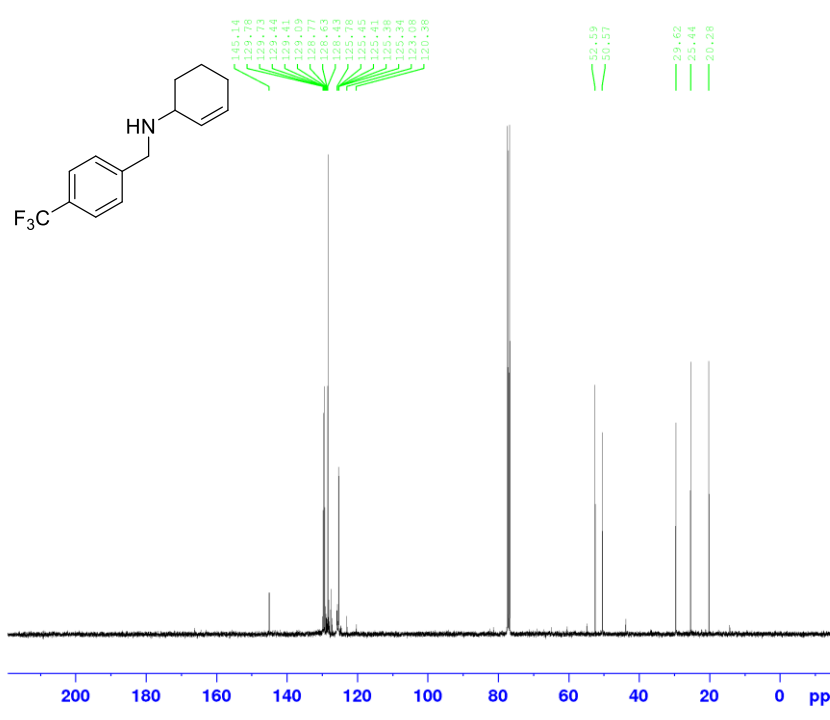


Current Data Parameters
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 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190921
 Time 13.25
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 128
 DW 104.400 usec
 DE 5.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SF01 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300095 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



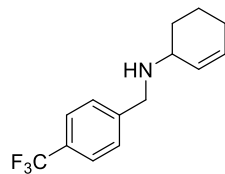
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 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1024
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 32768
 DW 20.850 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SF01 100.6228298 MHz

----- CHANNEL f2 -----
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SF02 400.1316005 MHz

F2 - Processing parameters
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 SF 100.6127549 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

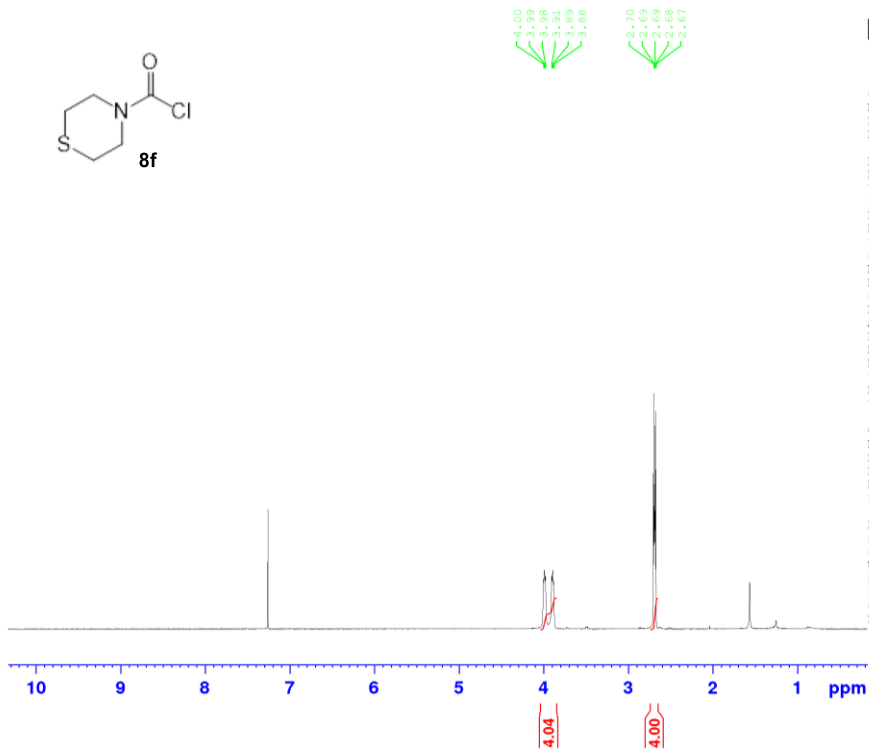
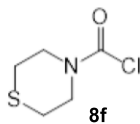


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 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190921
 Time 13.28
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 131072
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 75187.969 Hz
 FIDRES 0.573639 Hz
 AQ 0.8716288 sec
 RG 2048
 DW 6.650 usec
 DE 7.14 usec
 TE 298.1 K
 D1 5.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 19F
 P1 12.00 usec
 PL1 -3.00 dB
 SF01 376.4607040 MHz

F2 - Processing parameters
 SI 65536
 SF 376.4984036 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.00

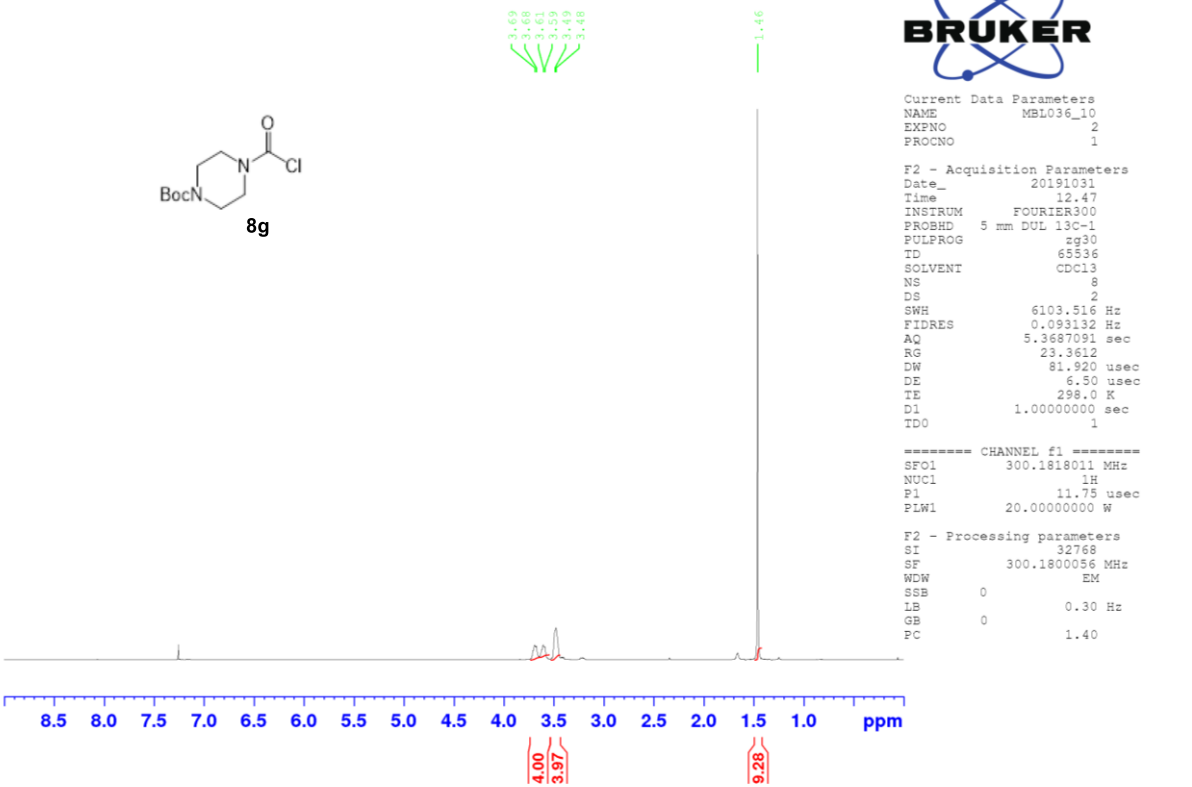
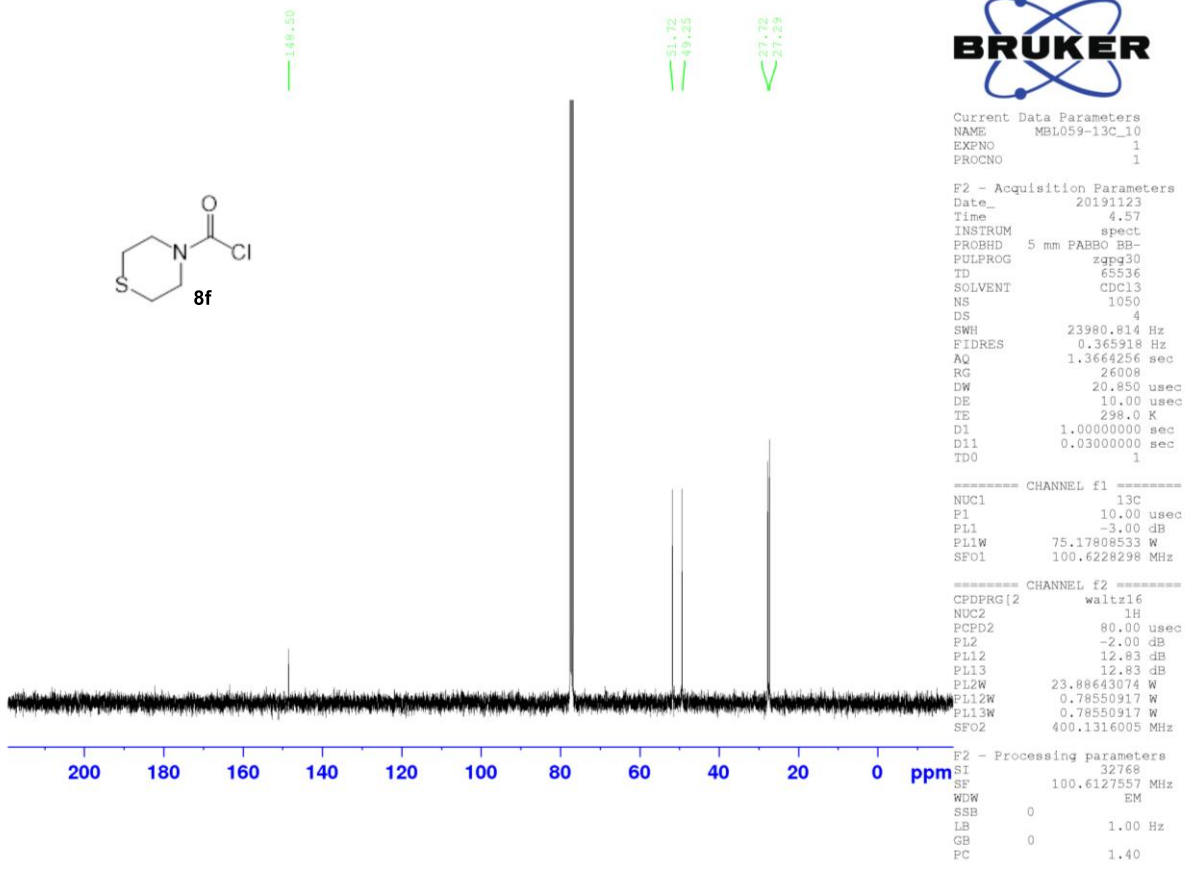


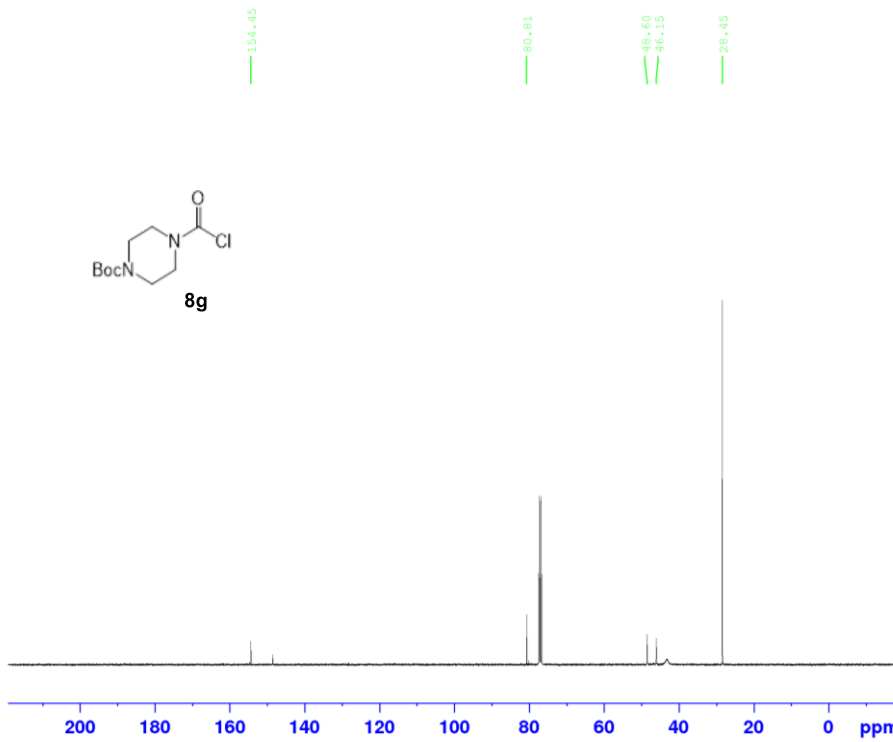
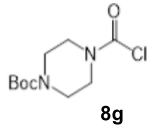
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 EXPNO 2
 PROCNO 1

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 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 256
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SF01 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300097 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





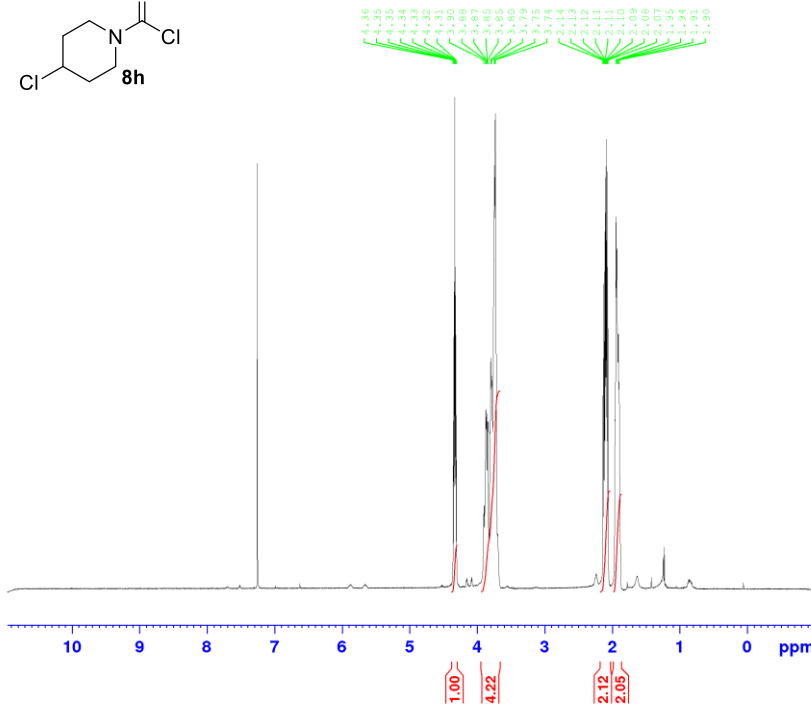
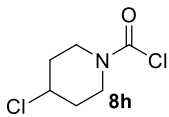
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 PROCNO 1

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 Time 11.33
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1050
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 26008
 DW 20.850 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
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 SF 100.6127564 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

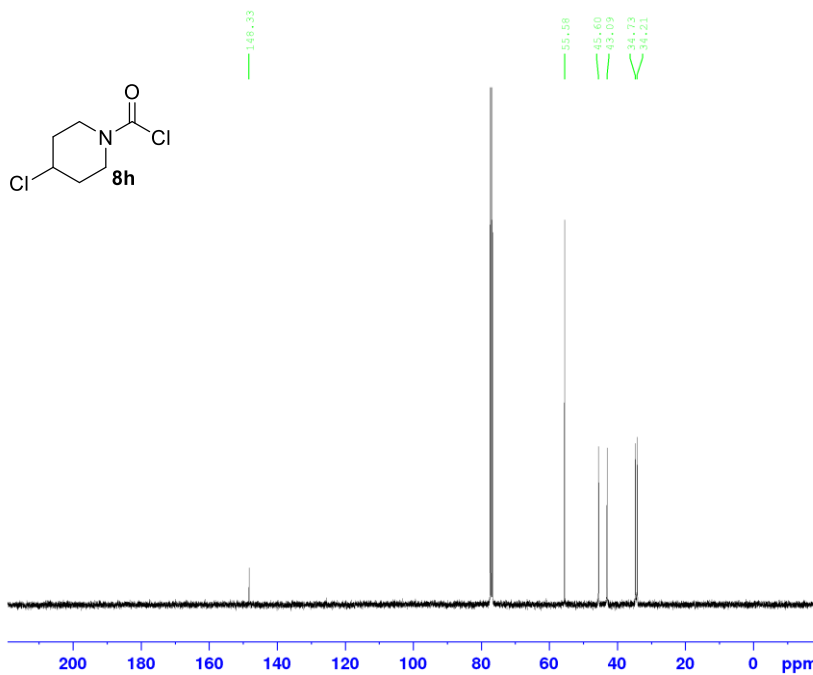
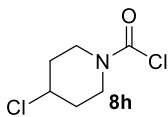


Current Data Parameters
 NAME DMZ-1-218dry_10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191217
 Time 12.19
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 181
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300095 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



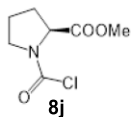
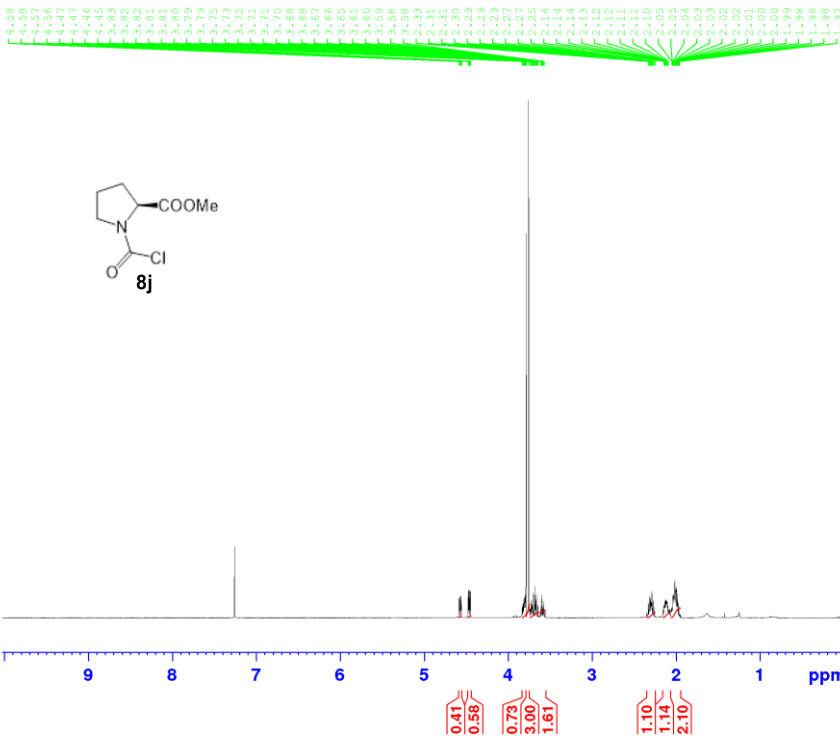
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EXPNO 1
PROCNO 1

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PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 1548
DS 4
SWH 23980.814 Hz
FIDRES 0.365918 Hz
AQ 1.3664256 sec
RG 32768
DW 20.850 usec
DE 10.00 usec
TE 298.1 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

==== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -3.00 dB
PL1W 75.17808533 W
SFO1 100.6228298 MHz

==== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL2 12.83 dB
PL13 12.83 dB
PL2W 23.8643074 W
PL12W 0.78550917 W
PL13W 0.78550917 W
SFO2 400.1316005 MHz

F2 - Processing parameters
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SF 100.6127581 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

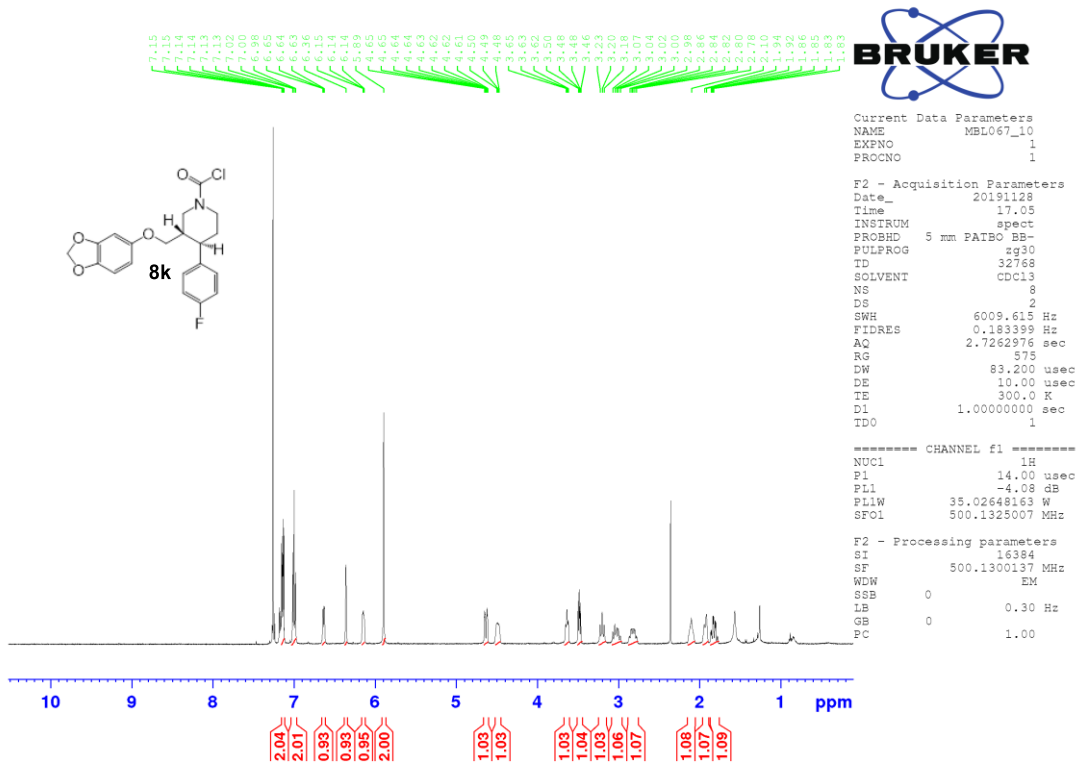
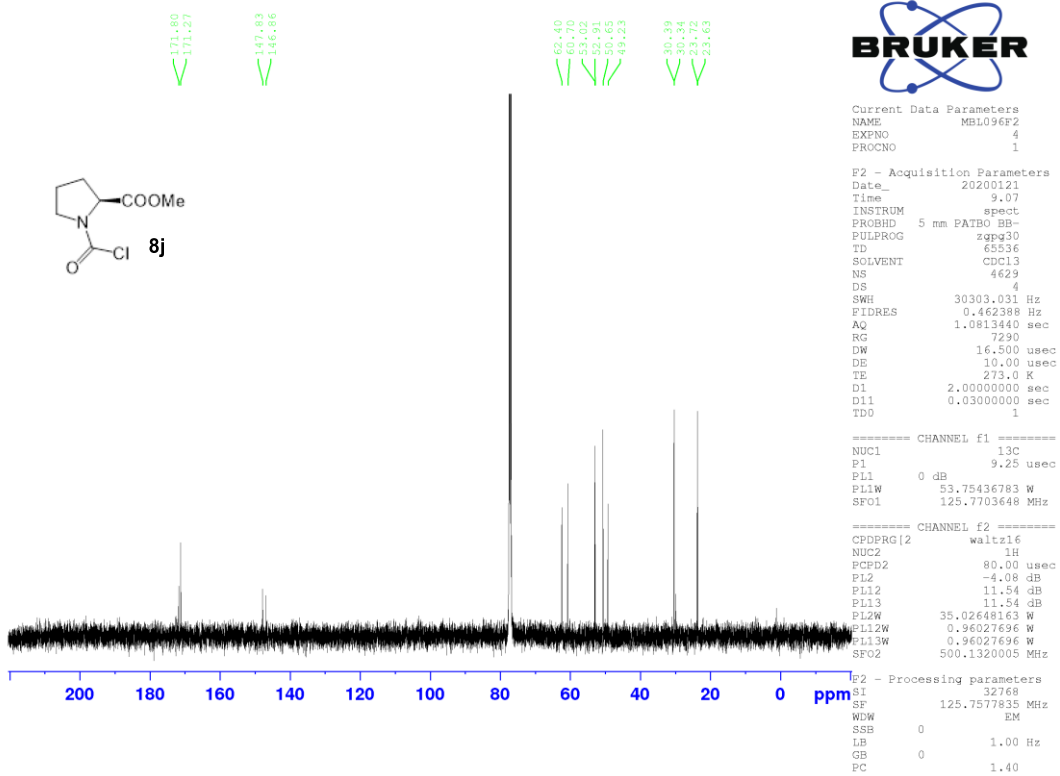


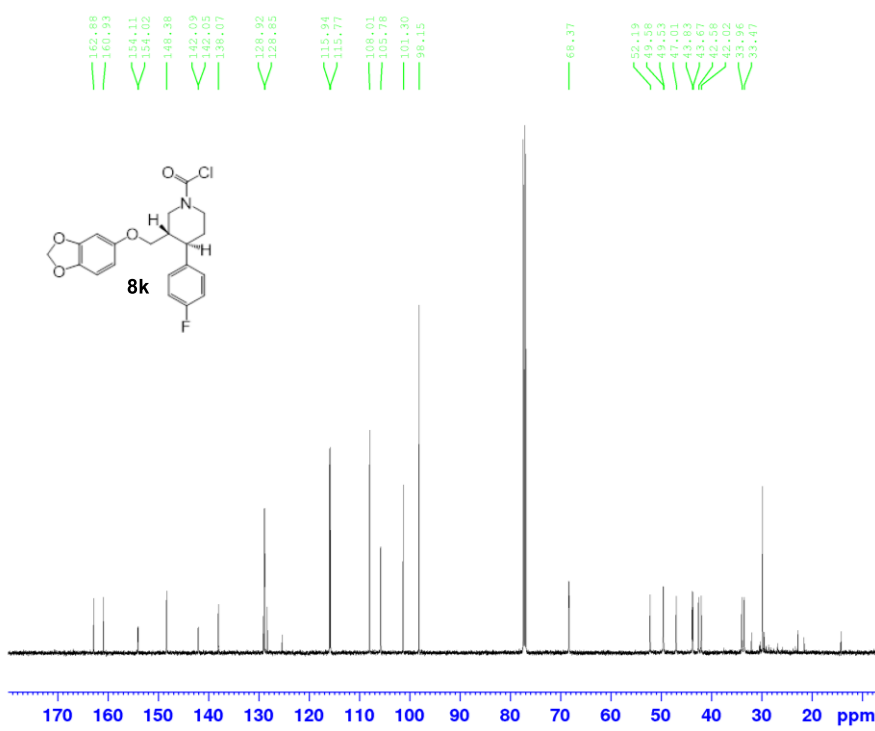
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EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20200116
Time 17.13
INSTRUM spect
PROBHD 5 mm PATBO BB-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 6009.615 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 322
DW 83.200 usec
DE 10.00 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1

==== CHANNEL f1 =====
NUC1 1H
P1 14.00 usec
PL1 -4.08 dB
PL1W 35.02648163 W
SFO1 500.1325007 MHz

F2 - Processing parameters
SI 16384
SF 500.1300137 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00





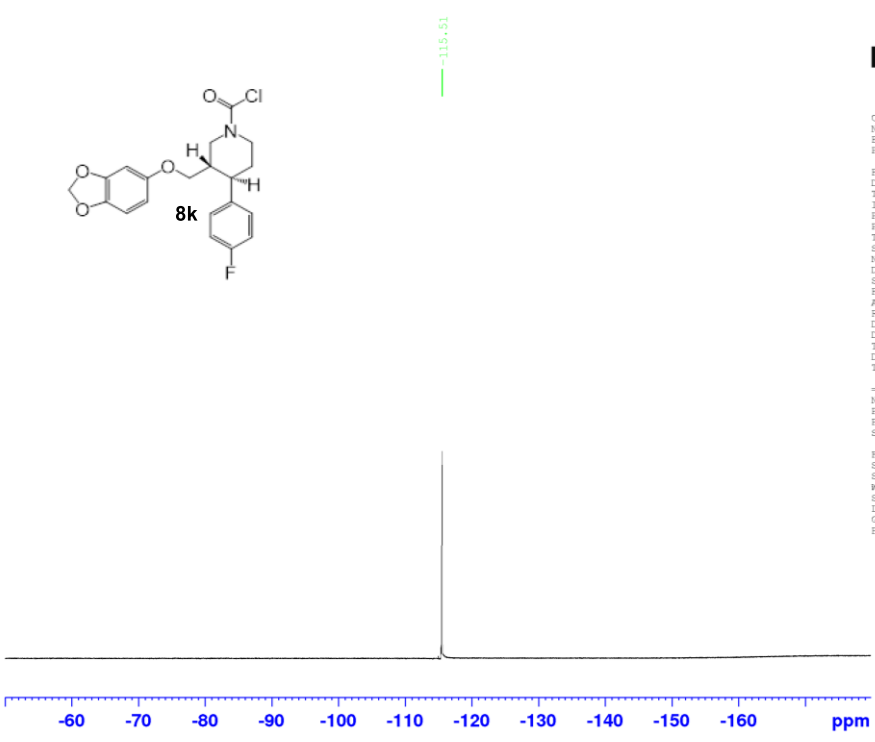
Current Data Parameters
 NAME MEL067-13C_10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20191218
 Time 17.03
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1179
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 10300
 DW 16.500 usec
 DE 10.00 usec
 TE 299.4 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

==== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.08 dB
 PL12 11.54 dB
 PL13 11.54 dB
 PL2W 35.02648163 W
 PL12W 0.96027696 W
 PL13W 0.96027696 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577759 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

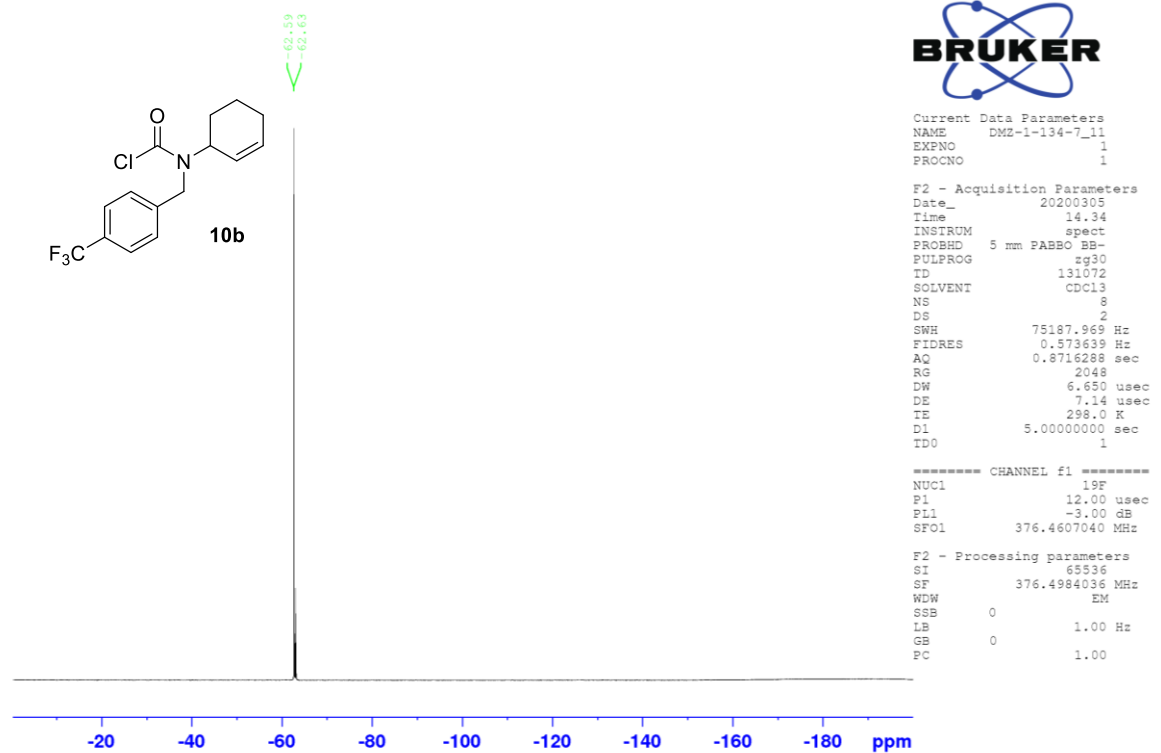
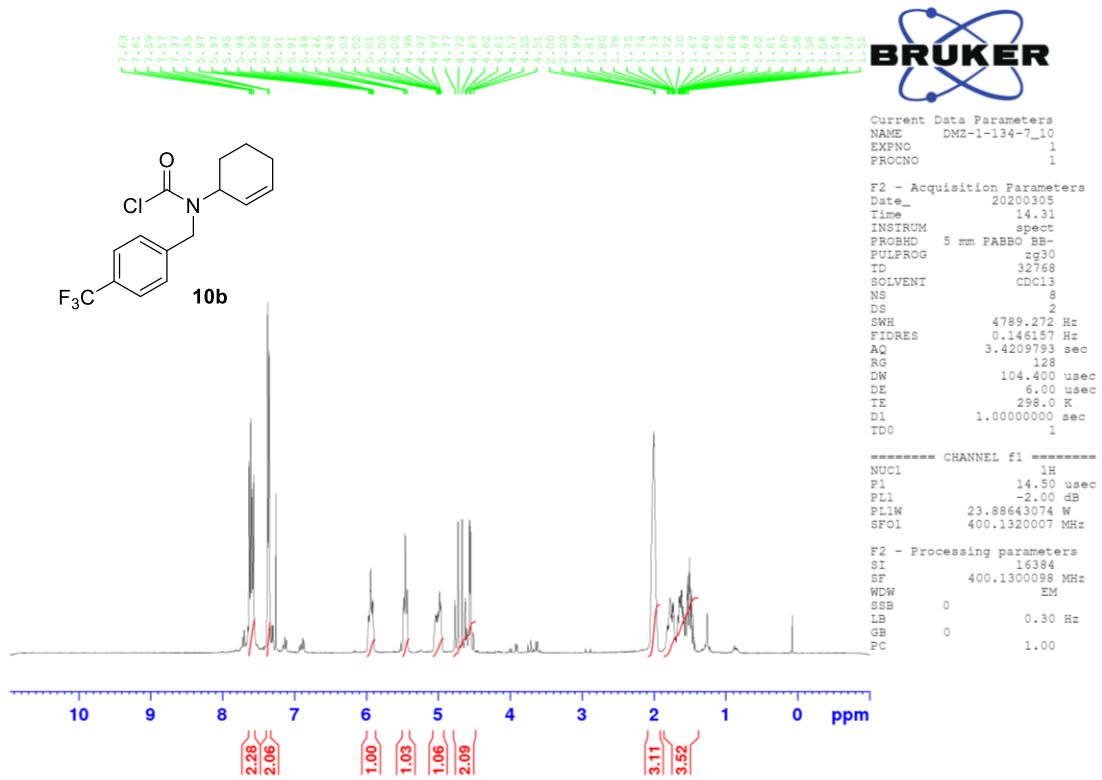


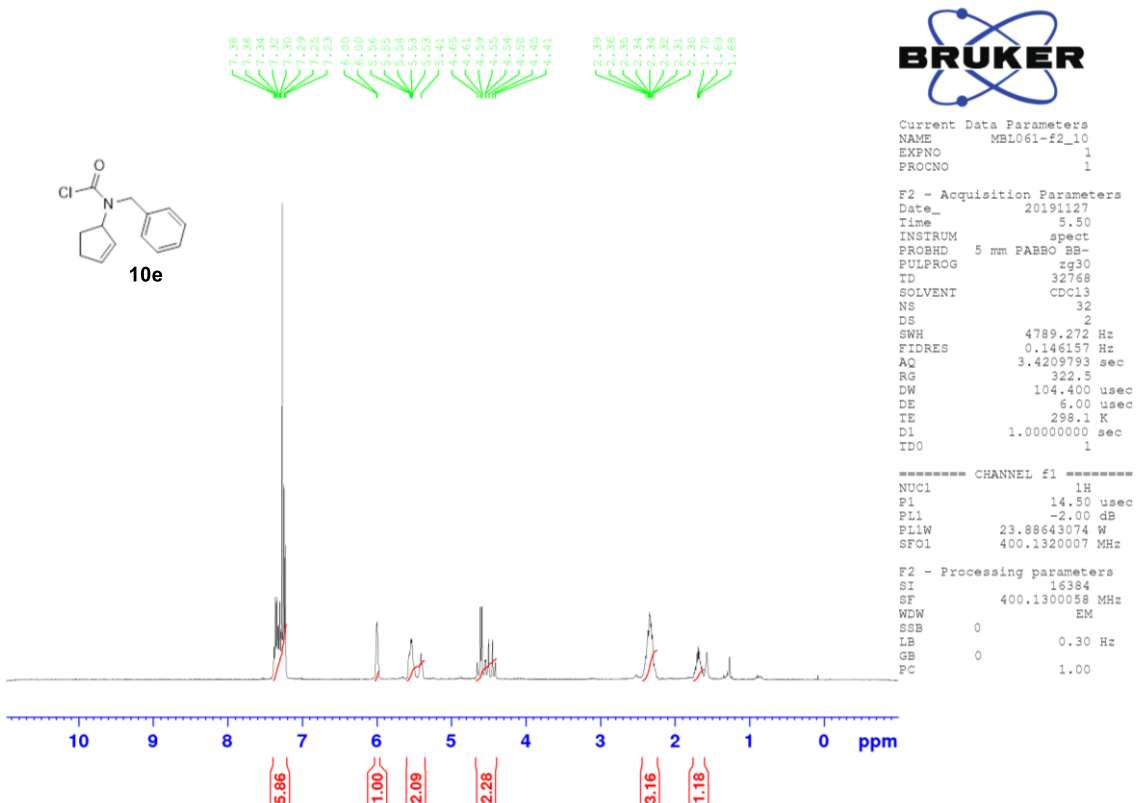
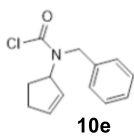
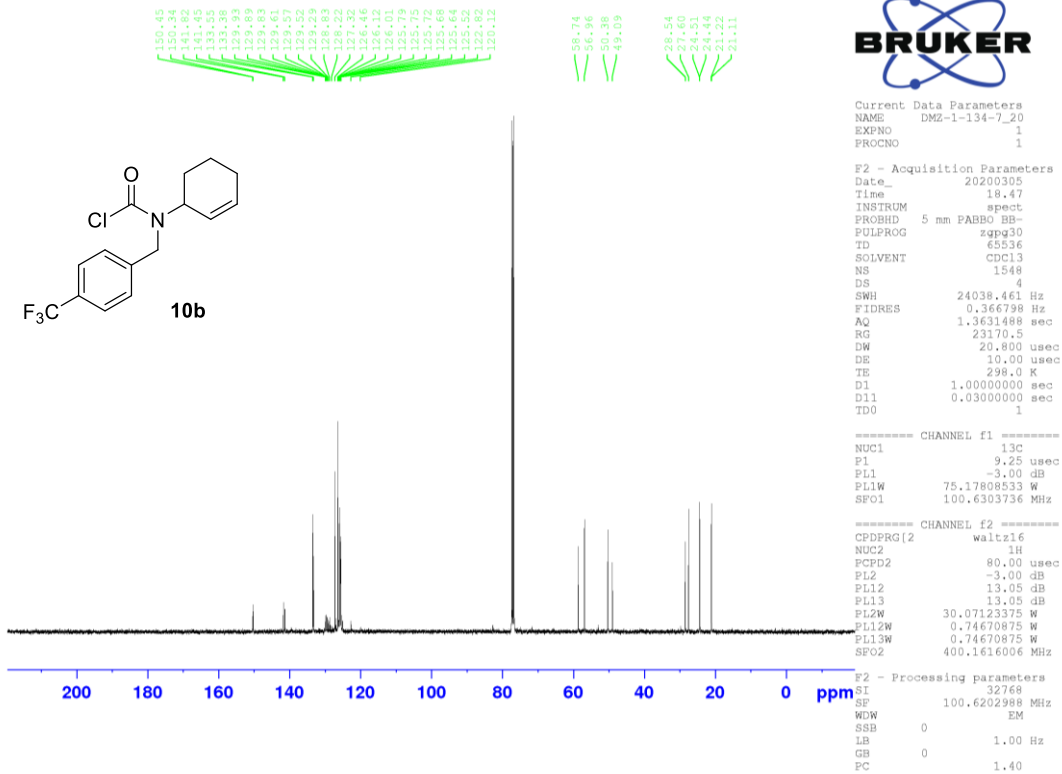
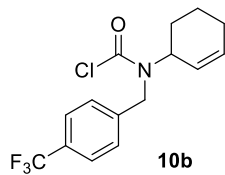
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 PROCNO 1

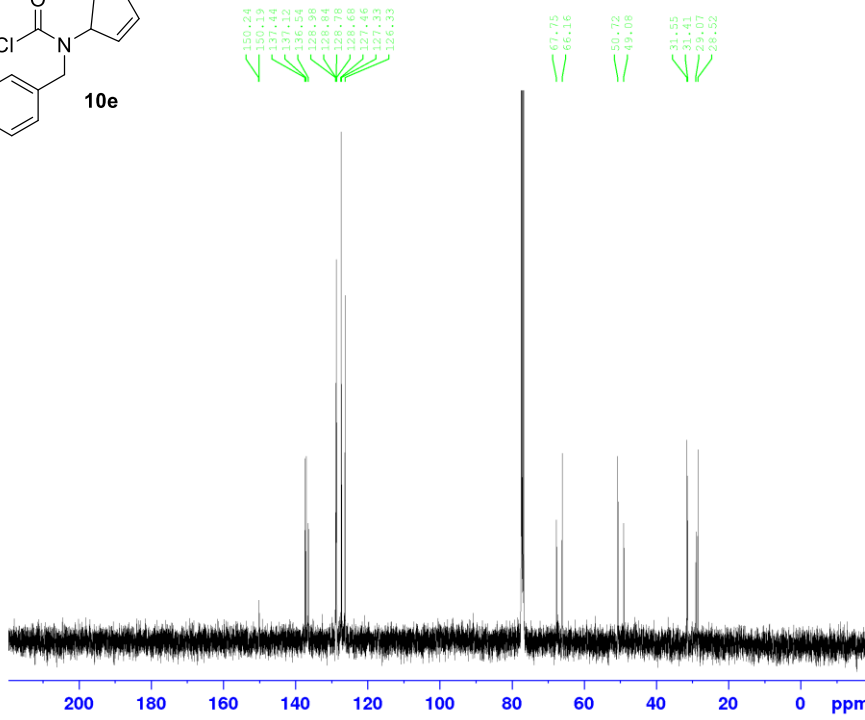
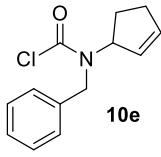
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 Time 11.09
 INSTRUM spect
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 PULPROG zg30
 TD 131072
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 75187.969 Hz
 FIDRES 0.573639 Hz
 AQ 0.8712888 sec
 RG 2048
 DW 6.650 usec
 DE 7.14 usec
 TE 298.1 K
 D1 5.00000000 sec
 TDO 1

==== CHANNEL f1 =====
 NUC1 13F
 P1 12.00 usec
 PL1 -3.00 dB
 SFO1 376.4607040 MHz

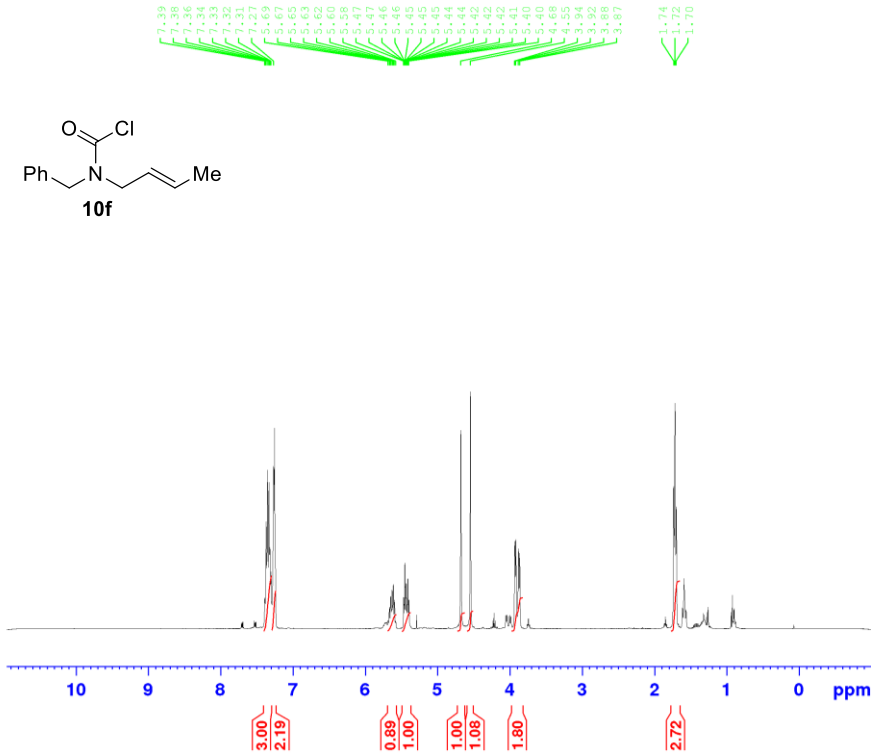
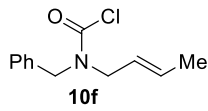
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 SF 376.4394036 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.00



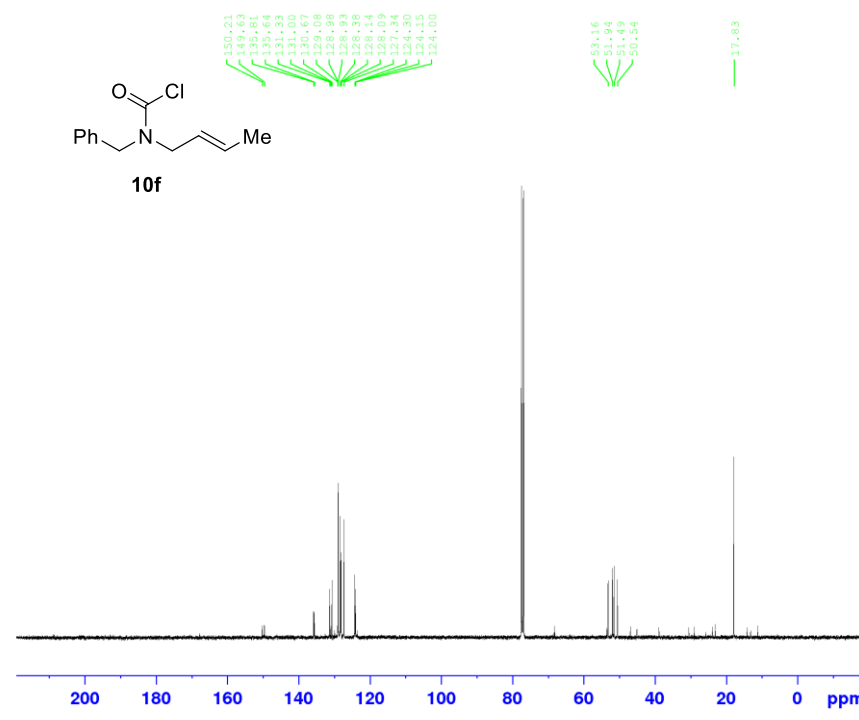
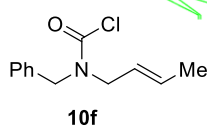




Current Data Parameters
 NAME MEL061-13C (PRODOTTO)
 EXPNO 1
 PROCNO 1
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 Date_ 20191127
 Time 22.15
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 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 ID 65536
 SOLVENT CDCl3
 NS 2200
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 26008
 DW 20.800 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1
 ===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6303736 MHz
 ===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -3.00 dB
 PL12 13.05 dB
 PL13 13.05 dB
 PL1W 30.07123375 W
 PL12W 0.74670875 W
 PL13W 0.74670875 W
 SFO2 400.1616006 MHz
 F2 - Processing parameters
 SI 32768
 SF 100.6202979 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



Current Data Parameters
 NAME DMZ-1-114-2_10
 EXPNO 1
 PROCNO 1
 F2 - Acquisition Parameters
 Date_ 20200302
 Time 16.36
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 ID 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 128
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TDO 1
 ===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz
 F2 - Processing parameters
 SI 16384
 SF 400.1300097 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



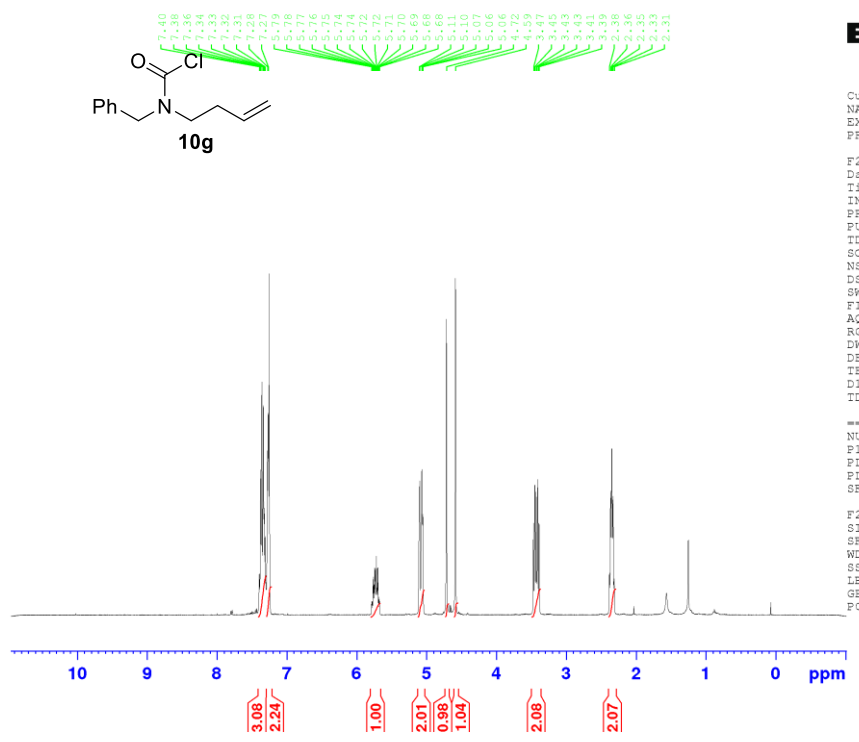
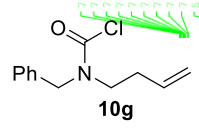
Current Data Parameters
 NAME DMZ-1-114-2_12
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200303
 Time 1.51
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1548
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 20642.5
 DW 20.850 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SF01 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SF02 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127571 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

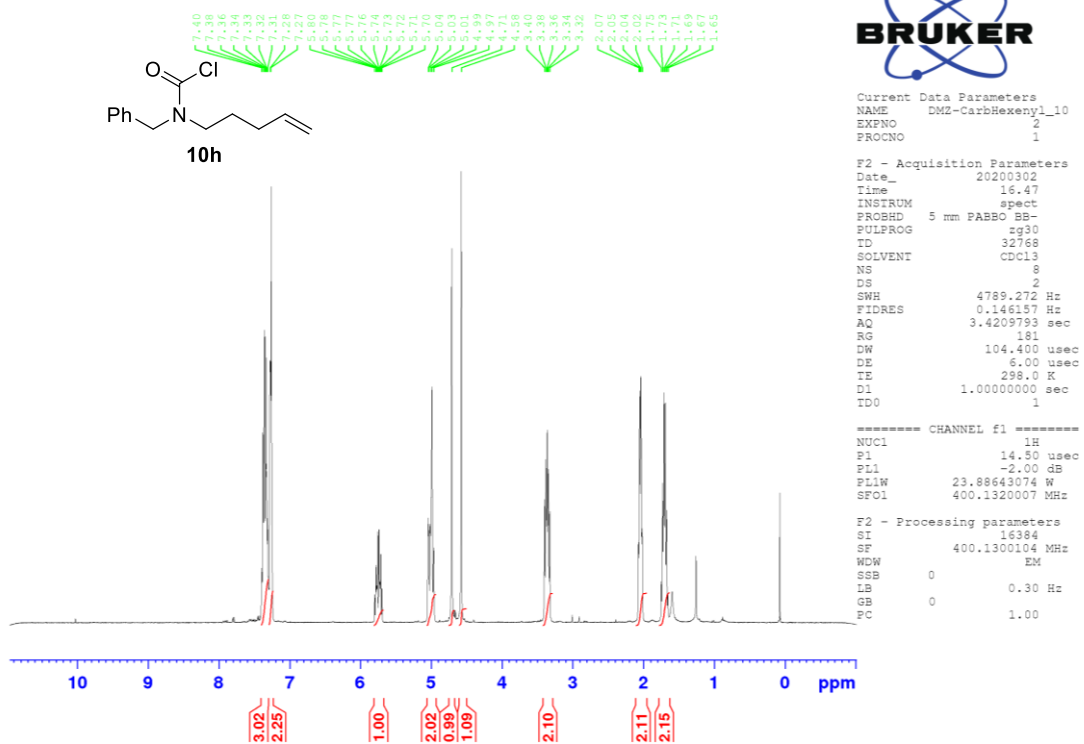
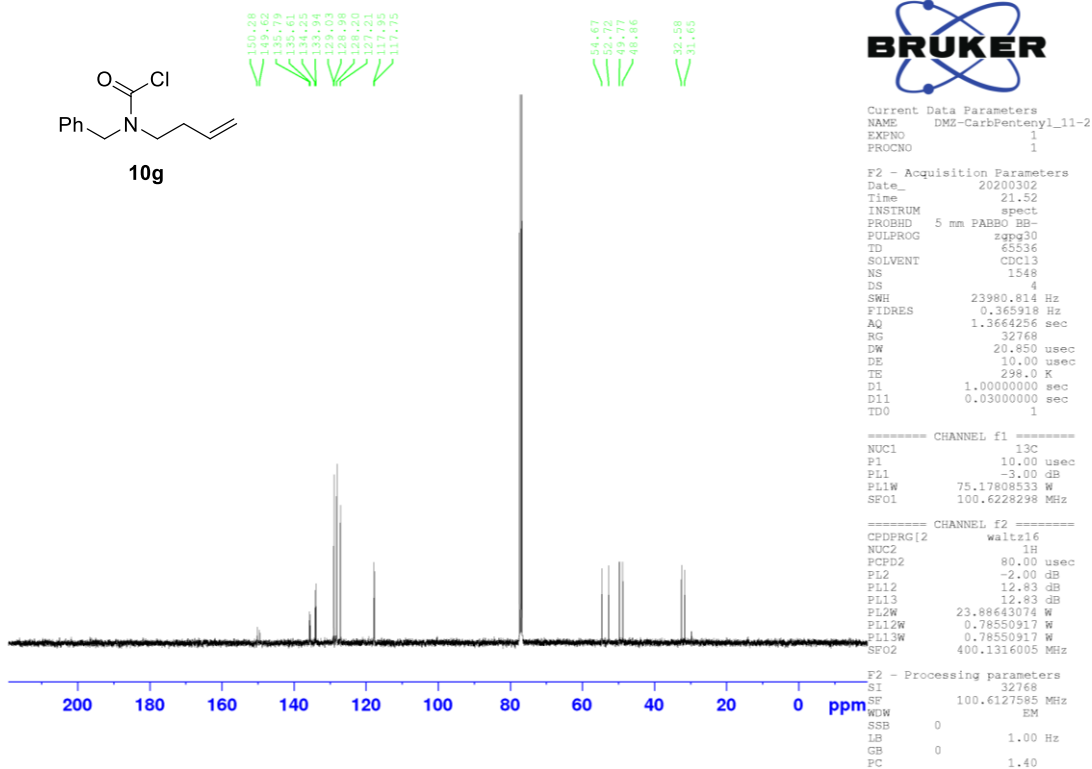


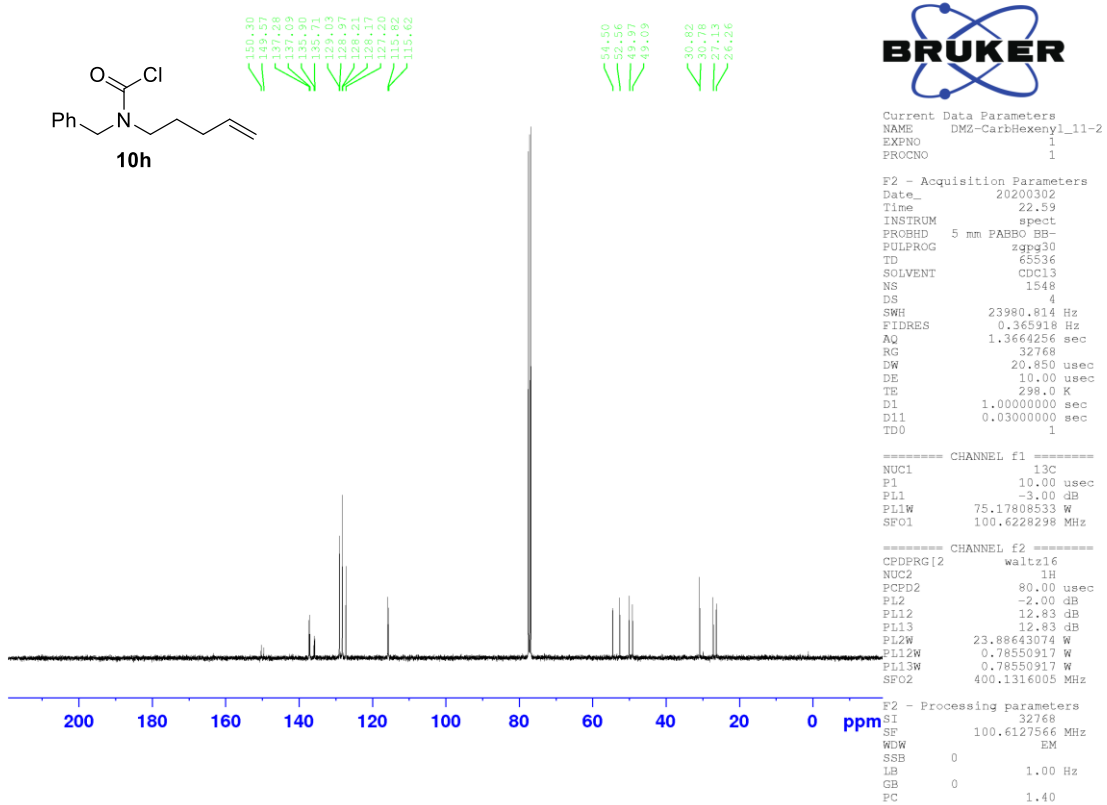
Current Data Parameters
 NAME DMZ-CarbPentery1_10
 EXPNO 4
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200302
 Time 16.42
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT cdcl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 181
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TDO 1

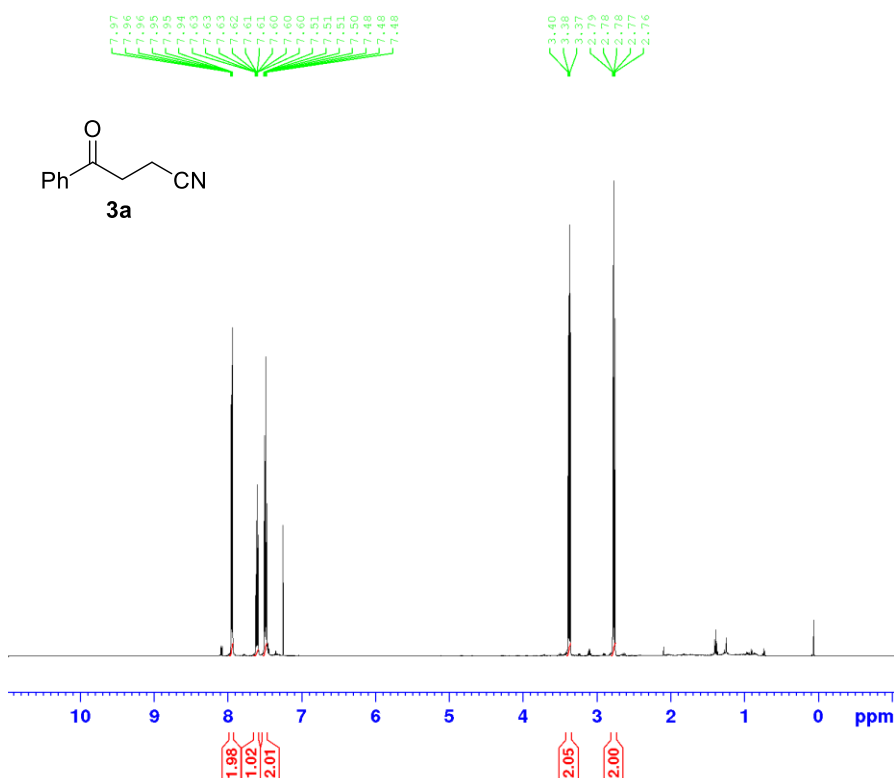
===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SF01 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300117 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00





F2. Products

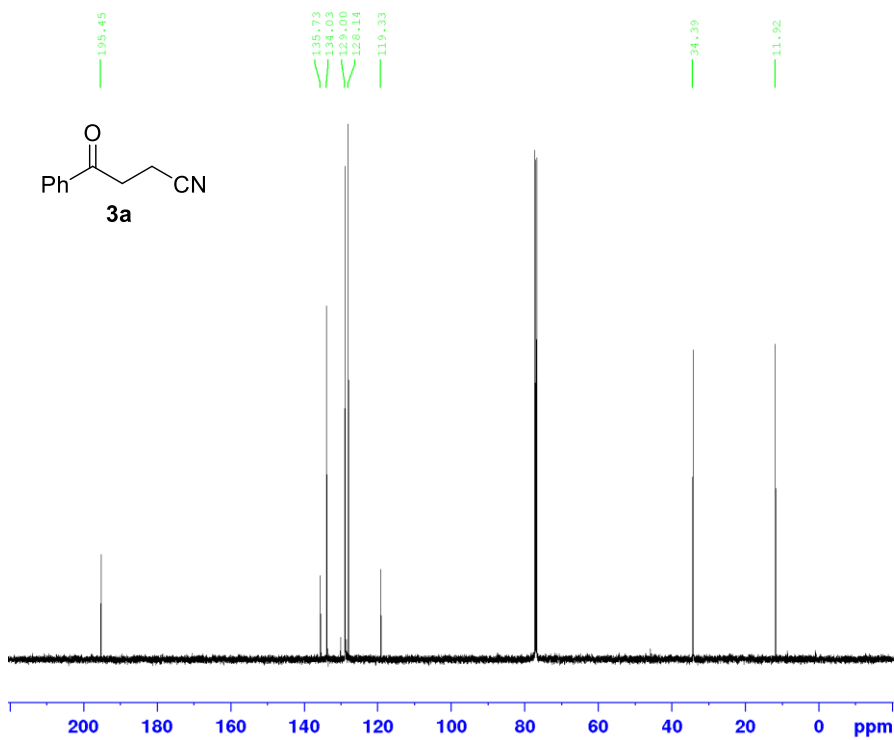


Current Data Parameters
 NAME epb713-prod-1H
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190520
 Time 17.58
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 203
 DW 83.200 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300137 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



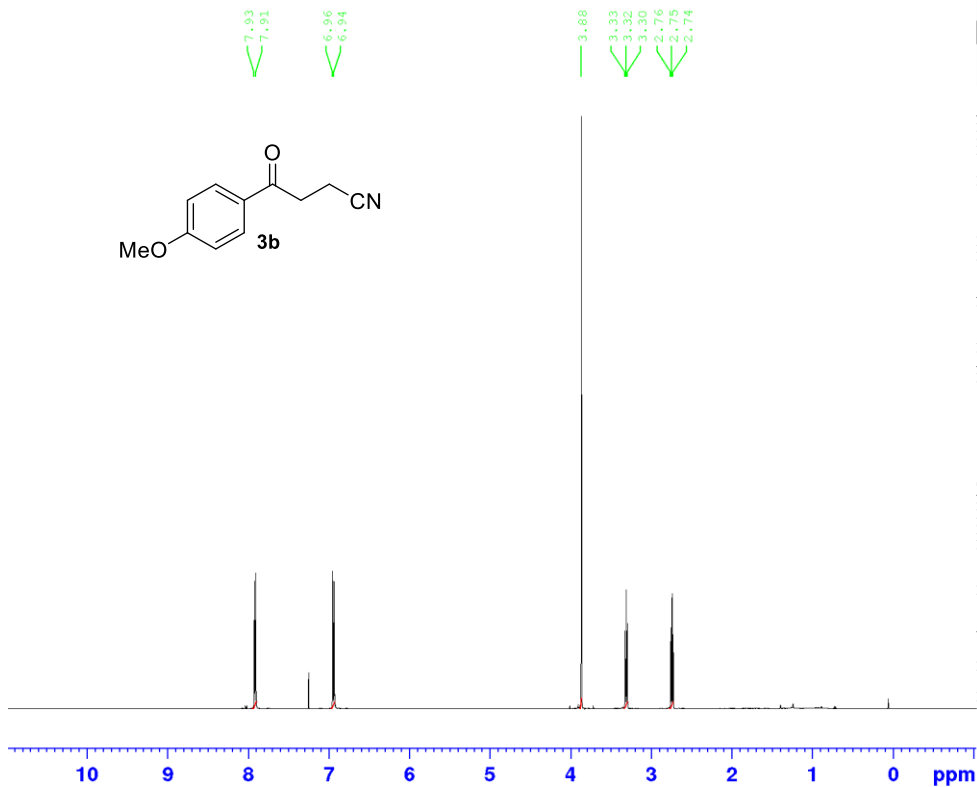
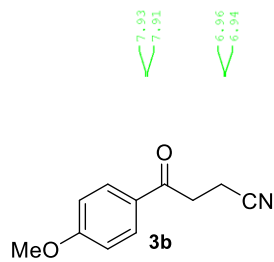
Current Data Parameters
 NAME epb713-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190520
 Time 23.07
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 14600
 DW 16.500 usec
 DE 10.00 usec
 TE 298.3 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577751 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

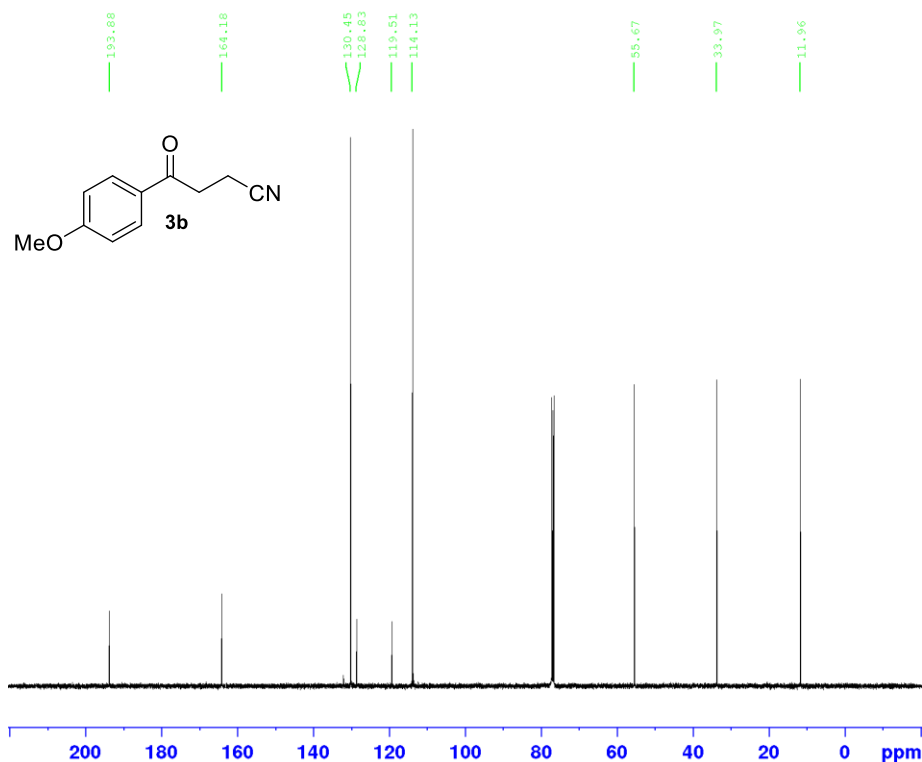
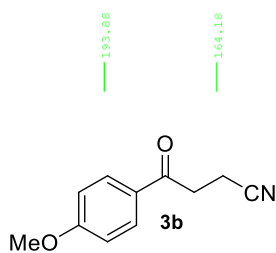


Current Data Parameters
 NAME epb717-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190318
 Time 18.19
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 144
 DW 83.200 usec
 DE 10.00 usec
 TE 297.8 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300130 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



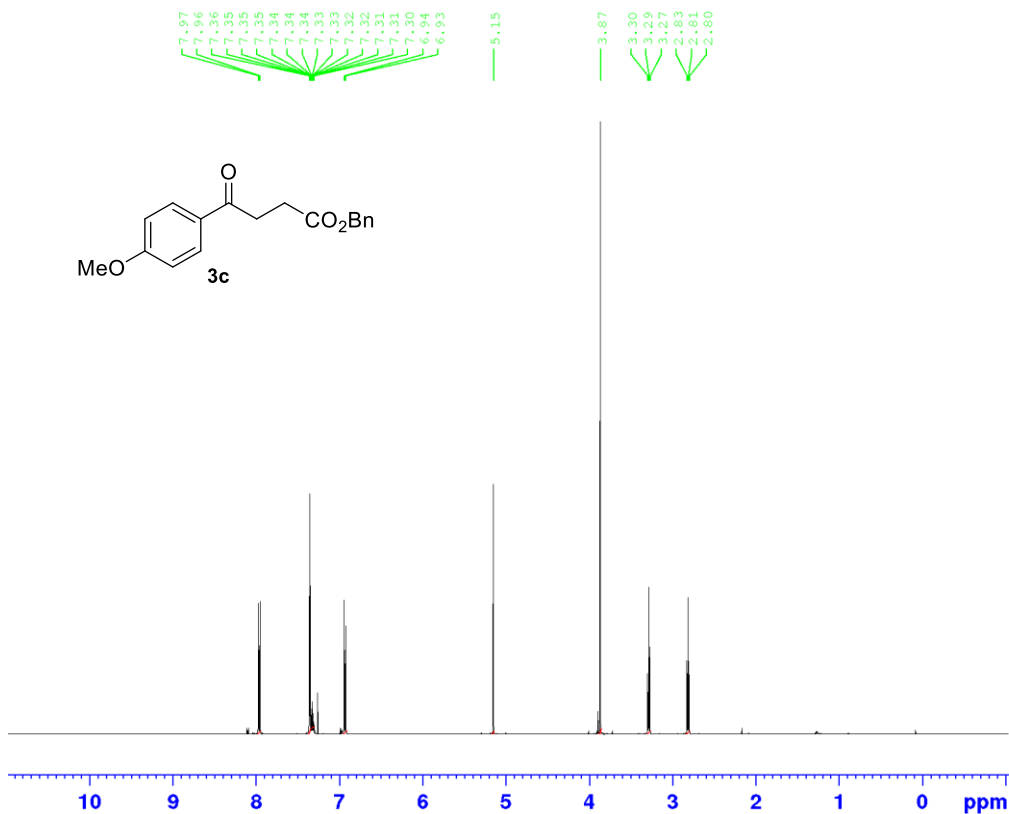
Current Data Parameters
 NAME epb717-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190318
 Time 21.57
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 11500
 DW 16.500 usec
 DE 10.00 usec
 TE 298.7 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPM2PRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577764 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

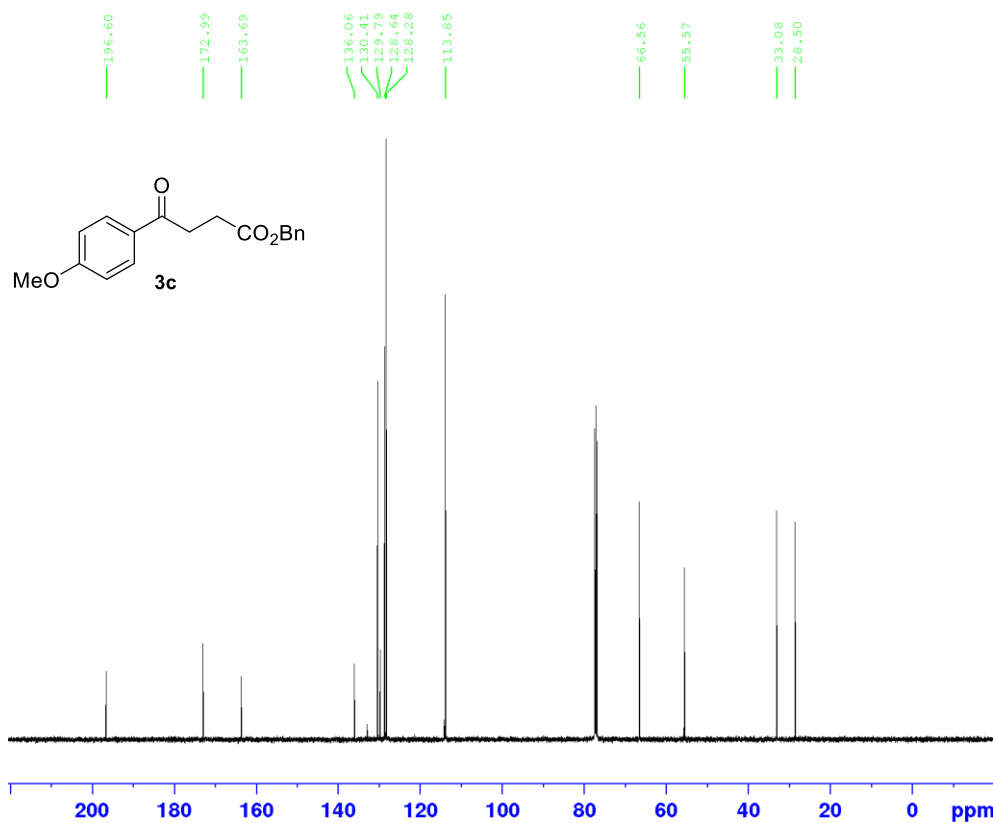


Current Data Parameters
NAME epb743-prod
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190318
Time 16.17
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 6009.615 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 80.6
DW 83.200 usec
DE 10.00 usec
TE 300.0 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 12.50 usec
PL1 0 dB
PL1W 13.68978119 W
SFO1 500.1325007 MHz

F2 - Processing parameters
SI 16364
SF 500.1300134 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 1.00



Current Data Parameters
NAME epb743-prod-13C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190318
Time 19.15
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 512
DS 4
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813440 sec
RG 10300
DW 16.500 usec
DE 10.00 usec
TE 300.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 8.60 usec
PL1 0 dB
PL1W 53.75436783 W
SFO1 125.7703648 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 0 dB
PL12 16.12 dB
PL13 16.12 dB
PL2W 13.68978119 W
PL12W 0.33450022 W
PL13W 0.33450022 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577774 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

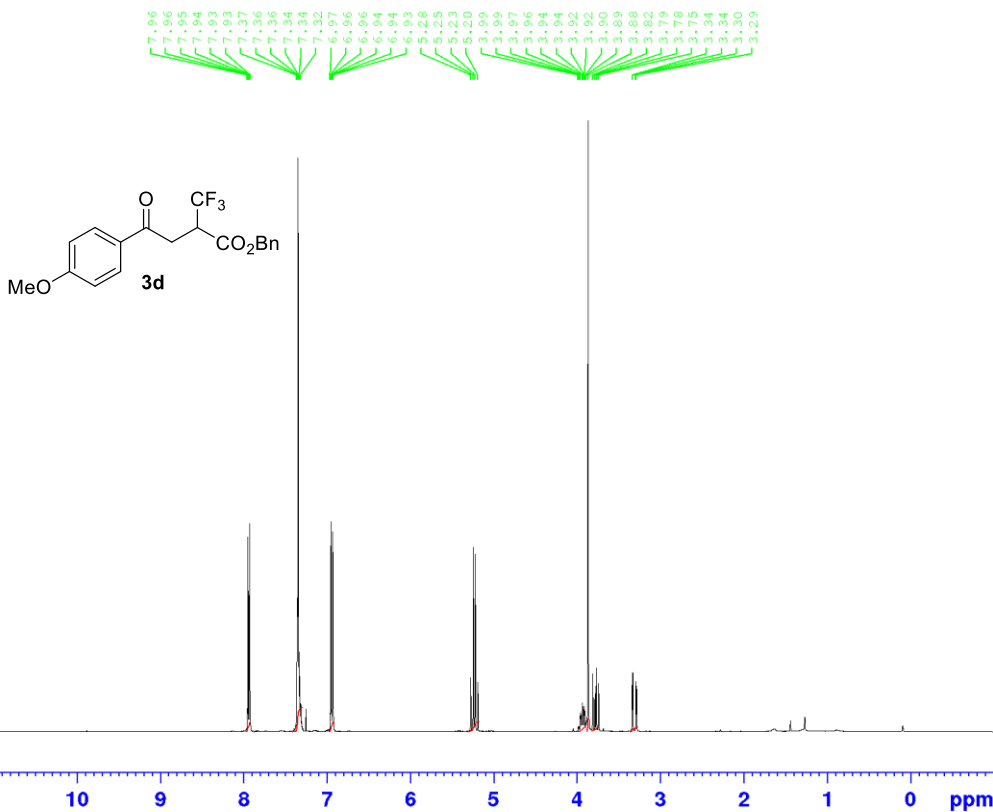


Current Data Parameters
NAME epb802-prod
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190531
Time 12.04
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4209793 sec
RG 71.8
DW 104.400 usec
DE 6.00 usec
TE 298.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.50 usec
PL1 -2.00 dB
PL1W 23.88643074 W
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300095 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



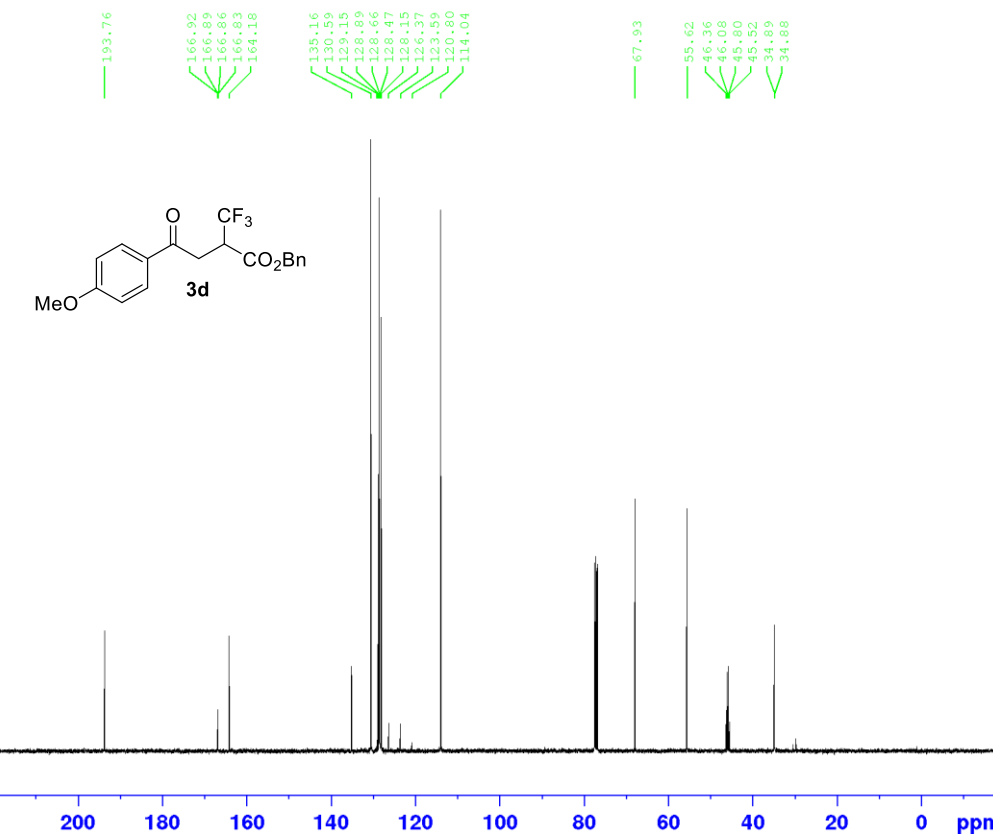
Current Data Parameters
NAME epb802-prod-13C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190531
Time 14.07
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 512
DS 4
SWH 23980.814 Hz
FIDRES 0.365918 Hz
AQ 1.3664256 sec
RG 32768
DW 20.850 usec
DE 10.00 usec
TE 298.3 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -3.00 dB
PL1W 75.17808533 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL12 12.83 dB
PL13 12.83 dB
PL2W 23.88643074 W
PL12W 0.78550917 W
PL13W 0.78550917 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127590 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



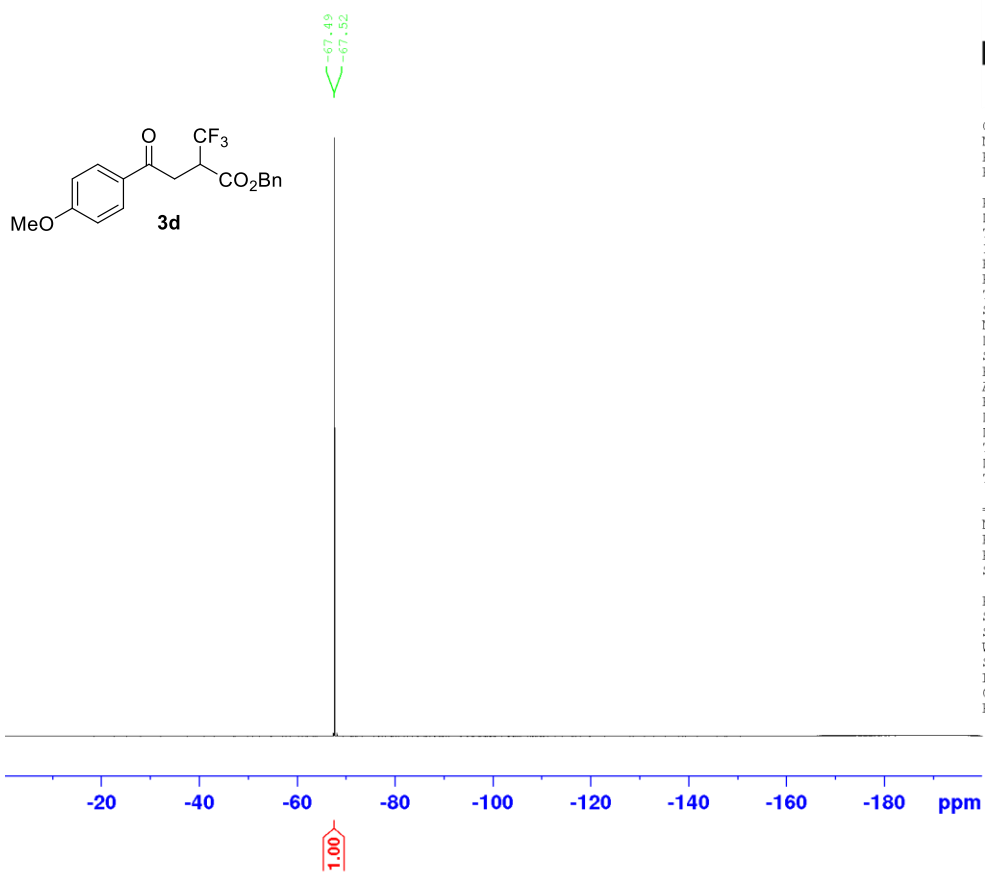


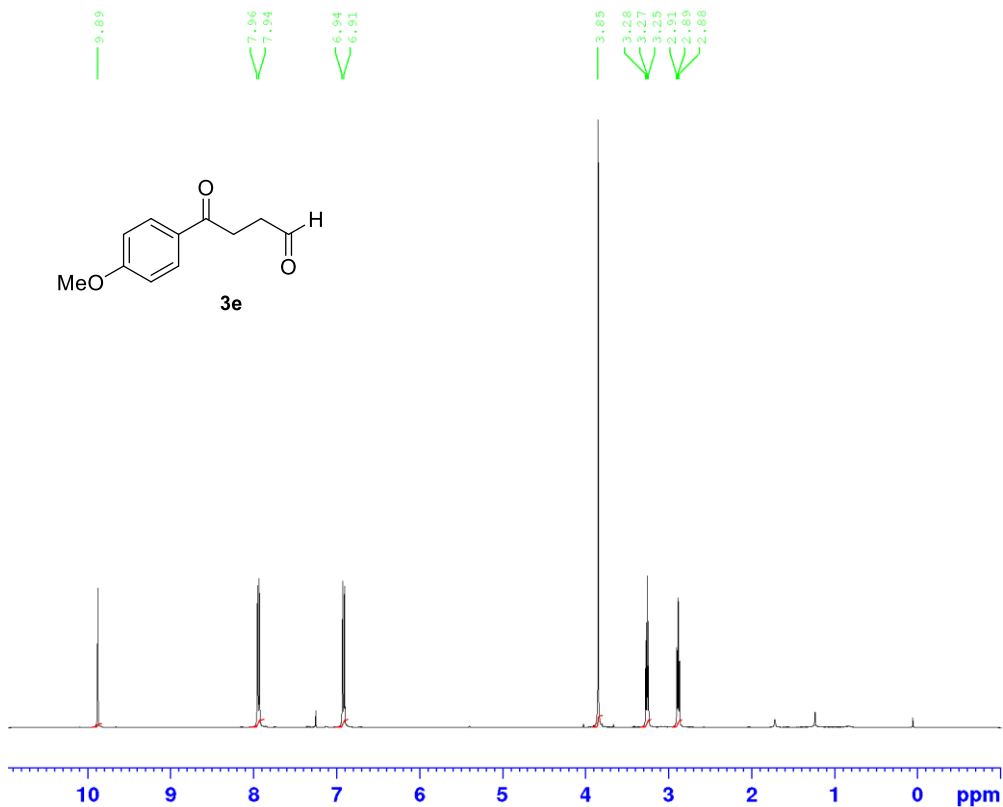
Current Data Parameters
NAME epb802-prod-19F
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190531
Time 12.06
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 131072
SOLVENT CDC13
NS 8
DS 2
SWH 75187.969 Hz
FIDRES 0.573639 Hz
AQ 0.8716288 sec
RG 1625.5
DW 6.650 usec
DE 7.14 usec
TE 298.1 K
D1 5.0000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 19F
P1 12.00 usec
PL1 -3.00 dB
SFO1 376.4607040 MHz

F2 - Processing parameters
SI 65536
SF 376.4984036 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00



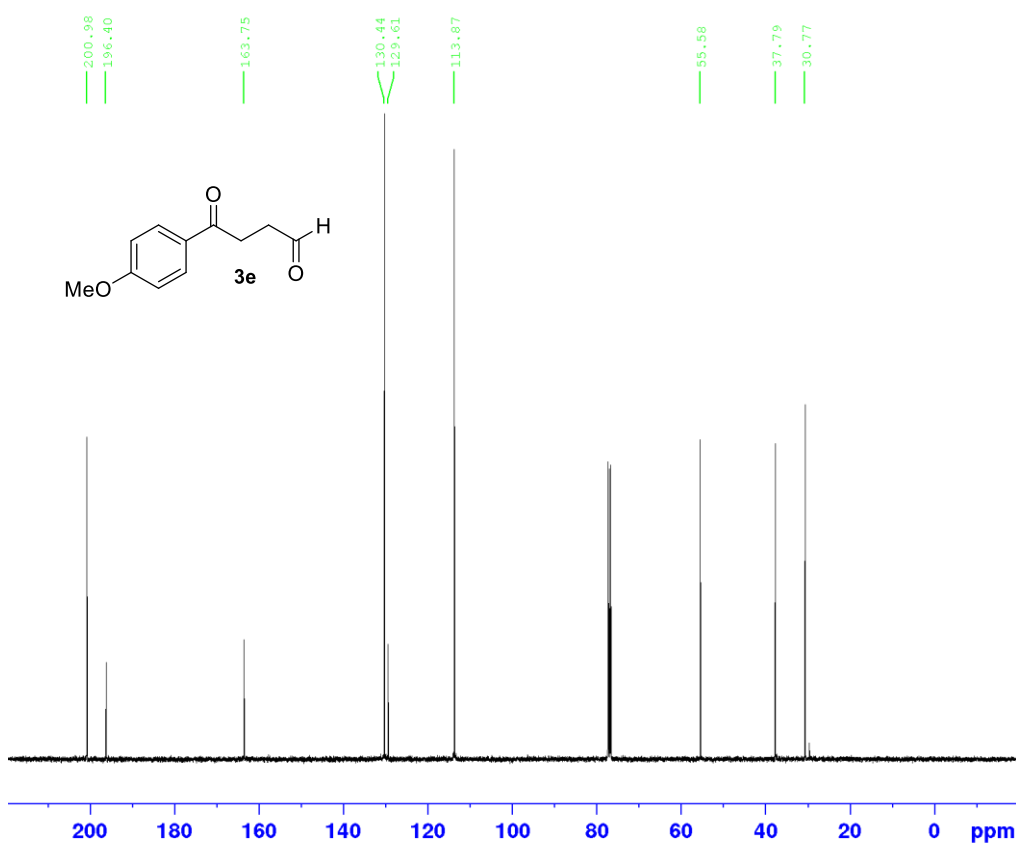


Current Data Parameters
 NAME epb997-prod-1H
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190617
 Time 9.38
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 114
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300098 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



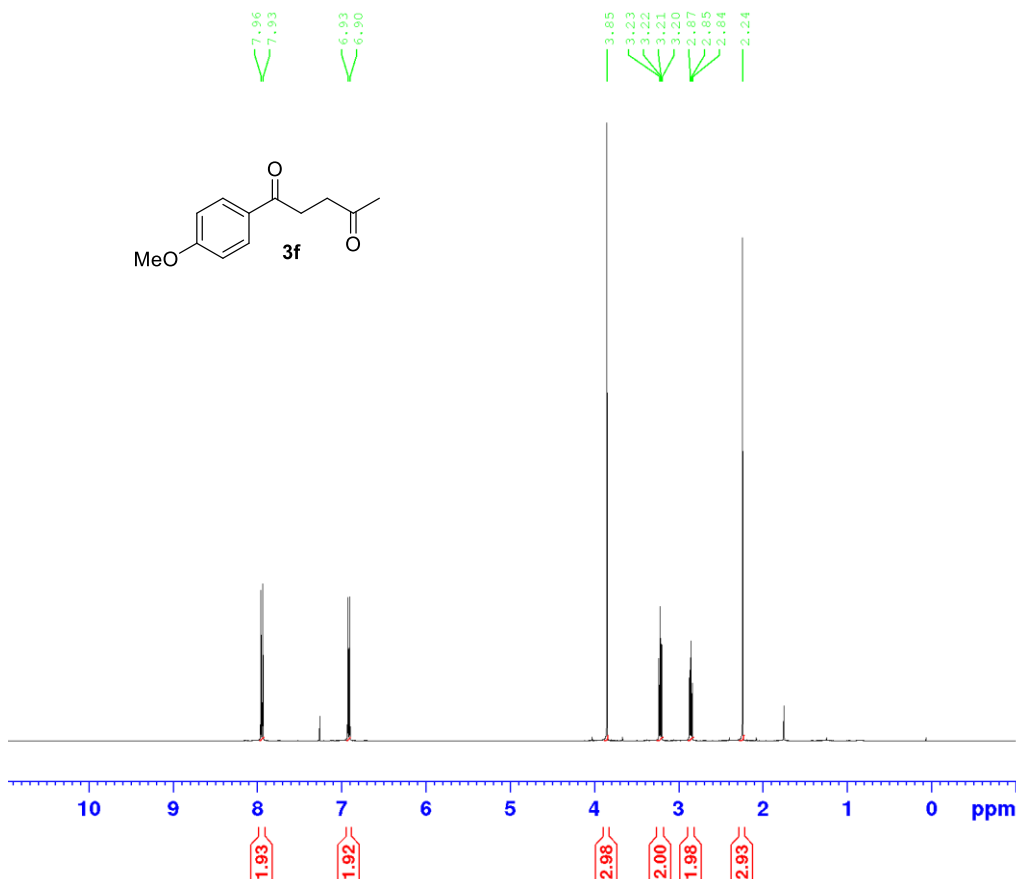
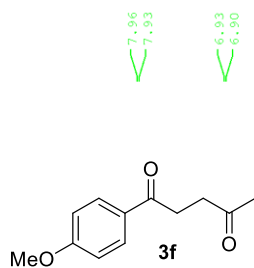
Current Data Parameters
 NAME epb997-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190617
 Time 14.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 23170.5
 DW 20.800 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6303736 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -3.00 dB
 PL12 13.05 dB
 PL13 13.05 dB
 PL2W 30.07123375 W
 PL12W 0.74670875 W
 PL13W 0.74670875 W
 SFO2 400.1616006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6203027 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

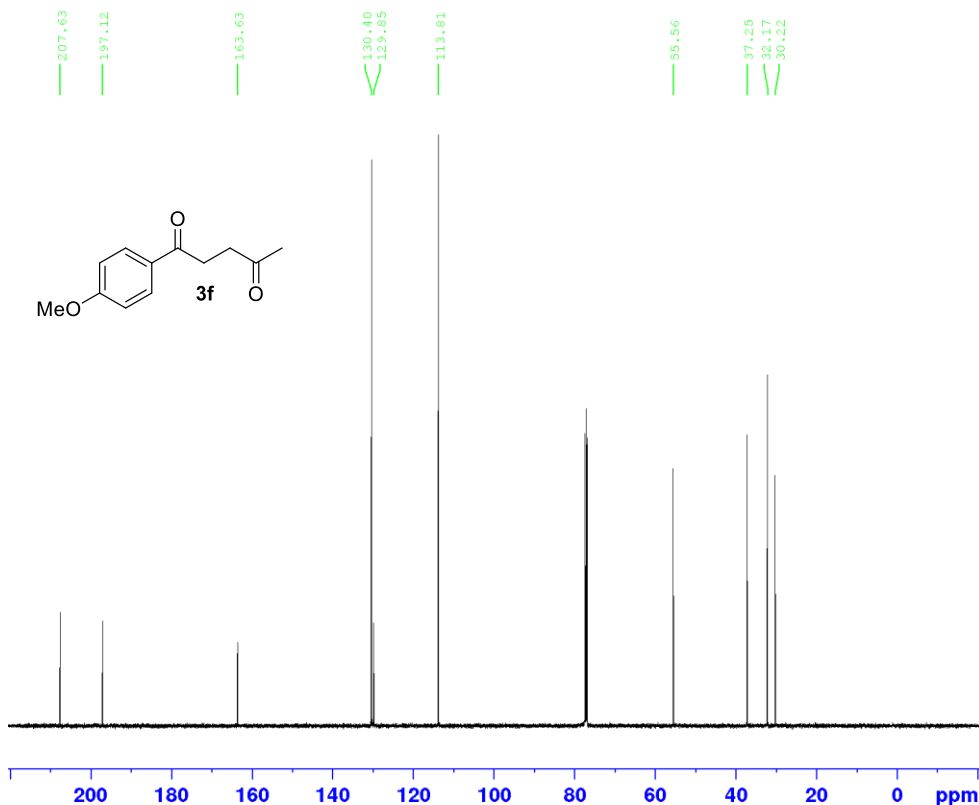
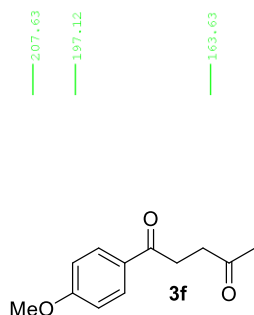


Current Data Parameters
NAME epb995-prod-1H
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190617
Time 9.33
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT CDC13
NS 8
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4209793 sec
RG 114
DW 104.400 usec
DE 6.00 usec
TE 298.0 K
D1 1.00000000 sec
TD0 1

----- CHANNEL f1 -----
NUC1 1H
P1 14.50 usec
PL1 -2.00 dB
PL1W 23.88643074 W
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300095 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 1.00



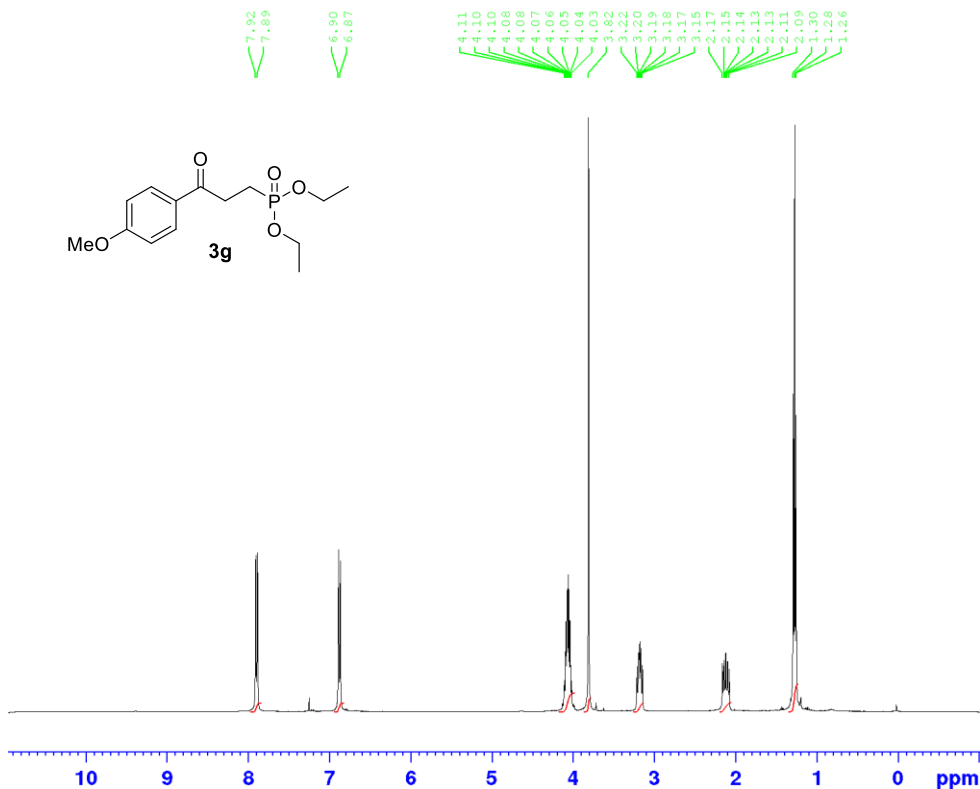
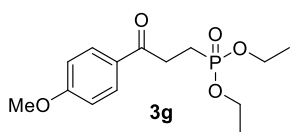
Current Data Parameters
NAME epb995-prod-13C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190617
Time 10.40
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 512
DS 4
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813440 sec
RG 10300
DW 16.500 usec
DE 10.00 usec
TE 298.1 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

----- CHANNEL f1 -----
NUC1 13C
P1 8.60 usec
PL1 0 dB
PL1W 53.75436783 W
SFO1 125.7703648 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 0 dB
PL12 16.12 dB
PL13 16.12 dB
PL2W 13.68978119 W
PL12W 0.33450022 W
PL13W 0.33450022 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577769 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

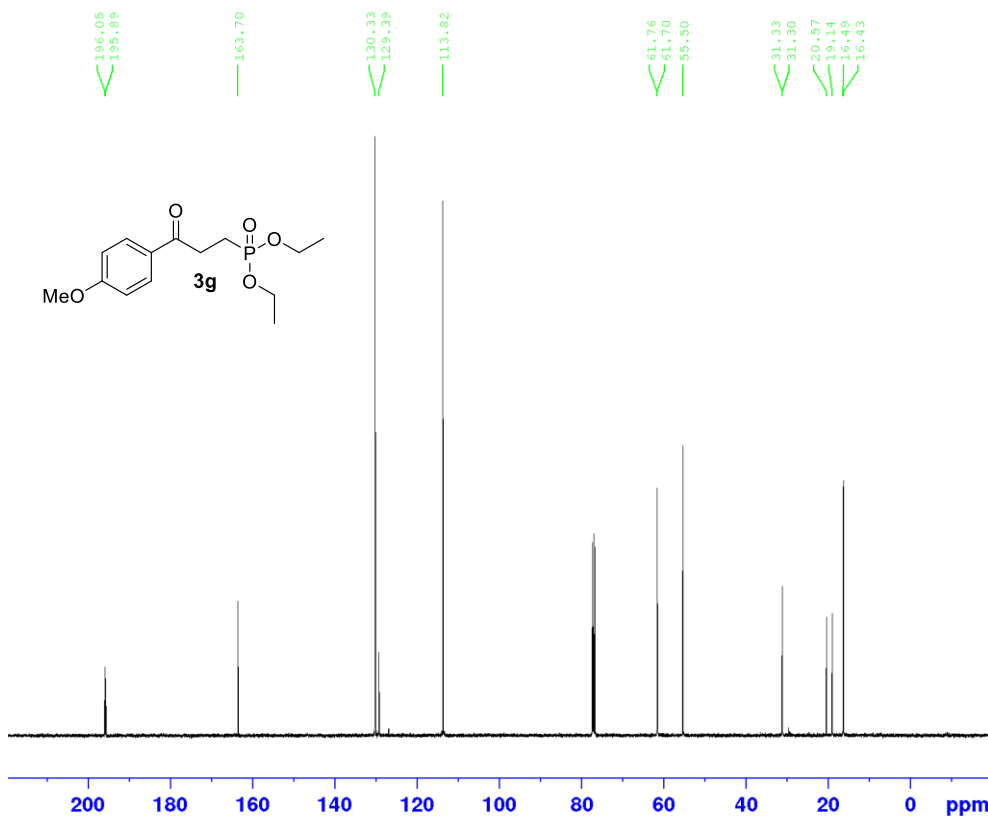
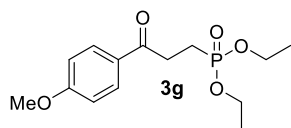


Current Data Parameters
NAME epb849-prod
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190314
Time 10.49
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4209793 sec
RG 57
DW 104.400 usec
DE 6.00 usec
TE 297.9 K
D1 1.00000000 sec
TDO 1

----- CHANNEL f1 -----
NUC1 1H
P1 14.50 usec
PL1 -2.00 dB
PL1W 23.88643074 W
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300096 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



Current Data Parameters
NAME epb849-prod-13C
EXPNO 10
PROCNO 1

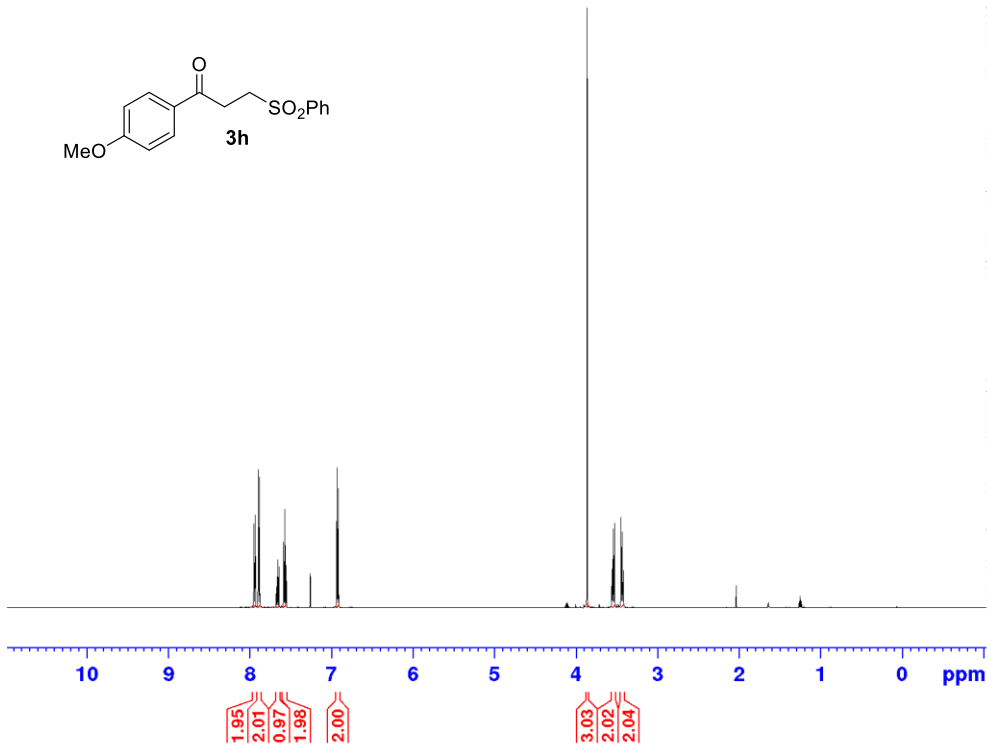
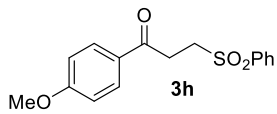
F2 - Acquisition Parameters
Date_ 20190314
Time 19.15
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 512
DS 4
SWH 24038.461 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 26008
DW 20.800 usec
DE 10.00 usec
TE 298.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TDO 1

----- CHANNEL f1 -----
NUC1 13C
P1 9.25 usec
PL1 -3.00 dB
PL1W 75.17808533 W
SFO1 100.6303736 MHz

----- CHANNEL f2 -----
CPDPRG[2] waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -3.00 dB
PL12 13.05 dB
PL13 13.05 dB
PL2W 30.07123375 W
PL12W 0.74670875 W
PL13W 0.74670875 W
SFO2 400.1616006 MHz

F2 - Processing parameters
SI 32768
SF 100.6203070 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

7.95
7.95
7.94
7.94
7.91
7.91
7.90
7.89
7.88
7.87
7.66
7.66
7.65
7.65
7.64
7.59
7.58
7.57
7.56
6.94
6.93
6.92
6.91
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3.45
3.44
3.44
3.43
3.42
3.42



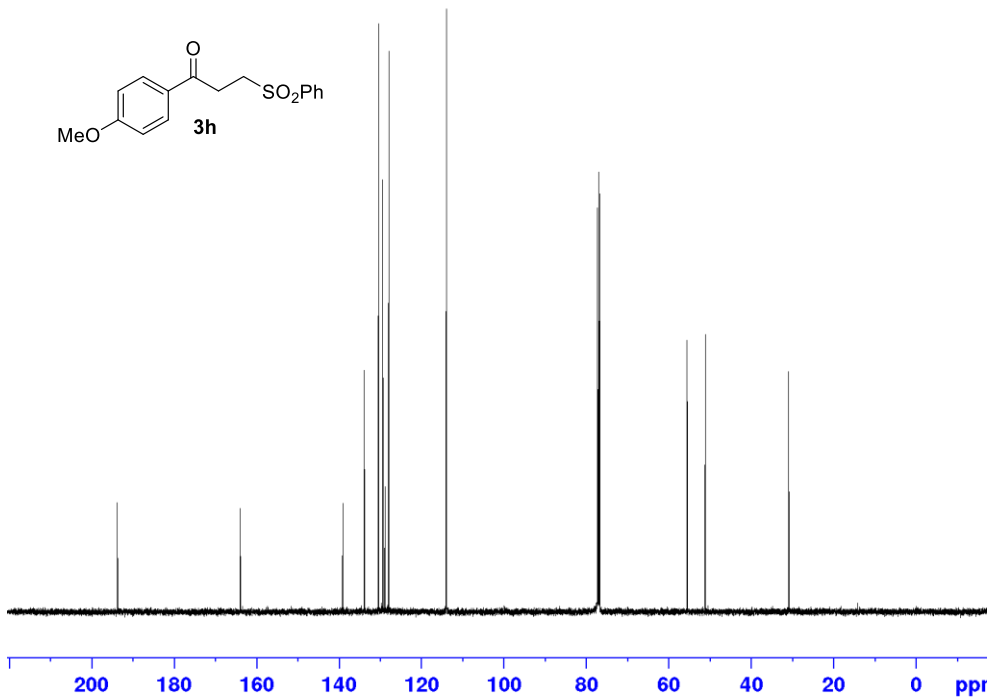
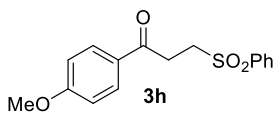
Current Data Parameters
 NAME epb737-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190318
 Time 18.26
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 90.5
 DW 83.200 usec
 DE 10.00 usec
 TE 297.9 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300133 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

193.95
164.12
139.25
136.91
129.52
129.01
128.10
114.06
55.65
51.29
31.04



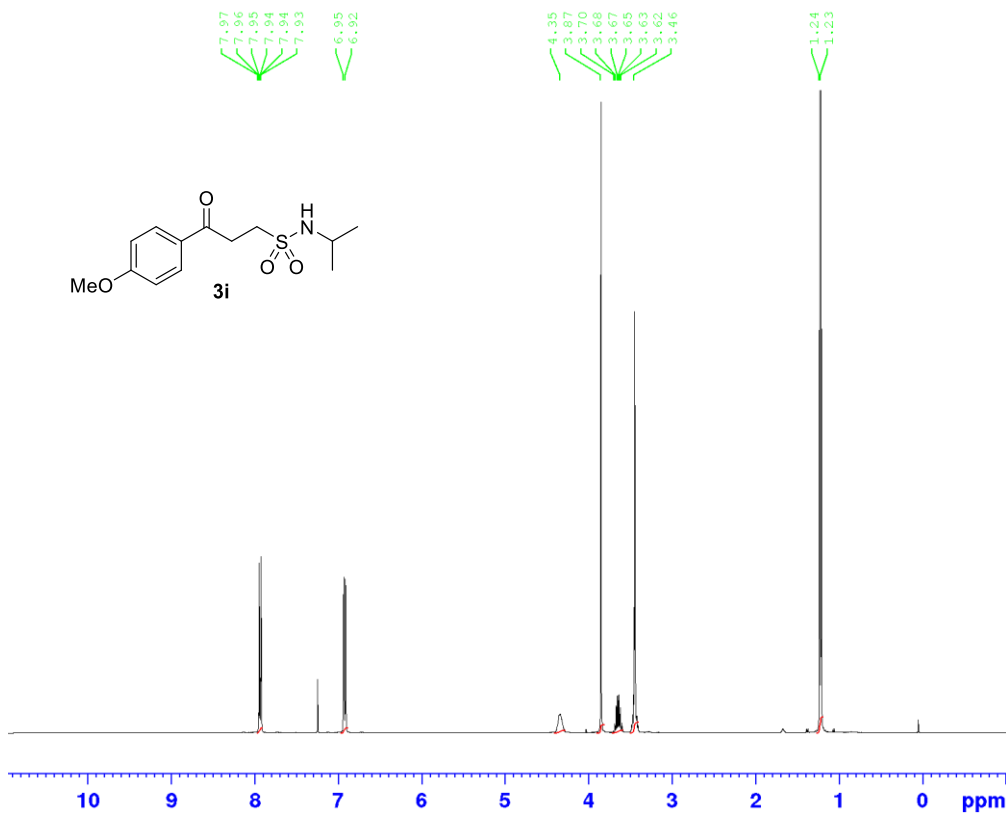
Current Data Parameters
 NAME epb737-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190318
 Time 22.27
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 11500
 DW 16.500 usec
 DE 10.00 usec
 TE 298.3 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577759 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

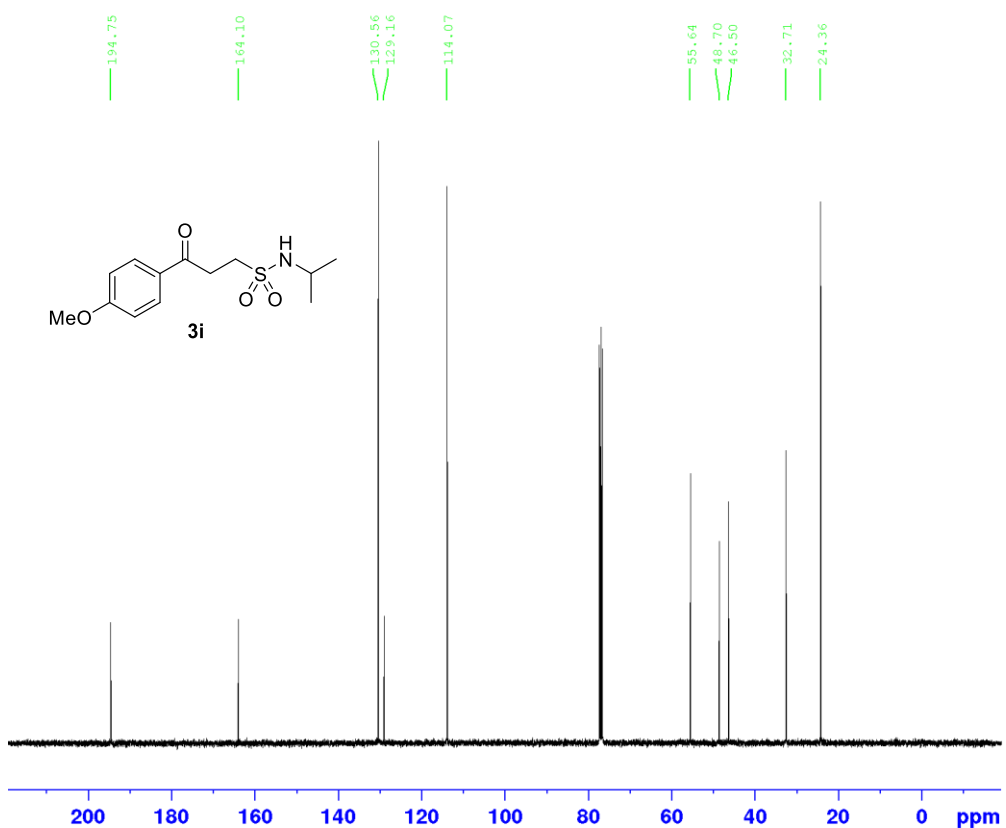


Current Data Parameters
 NAME epb801-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190319
 Time 18.31
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 114
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300097 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



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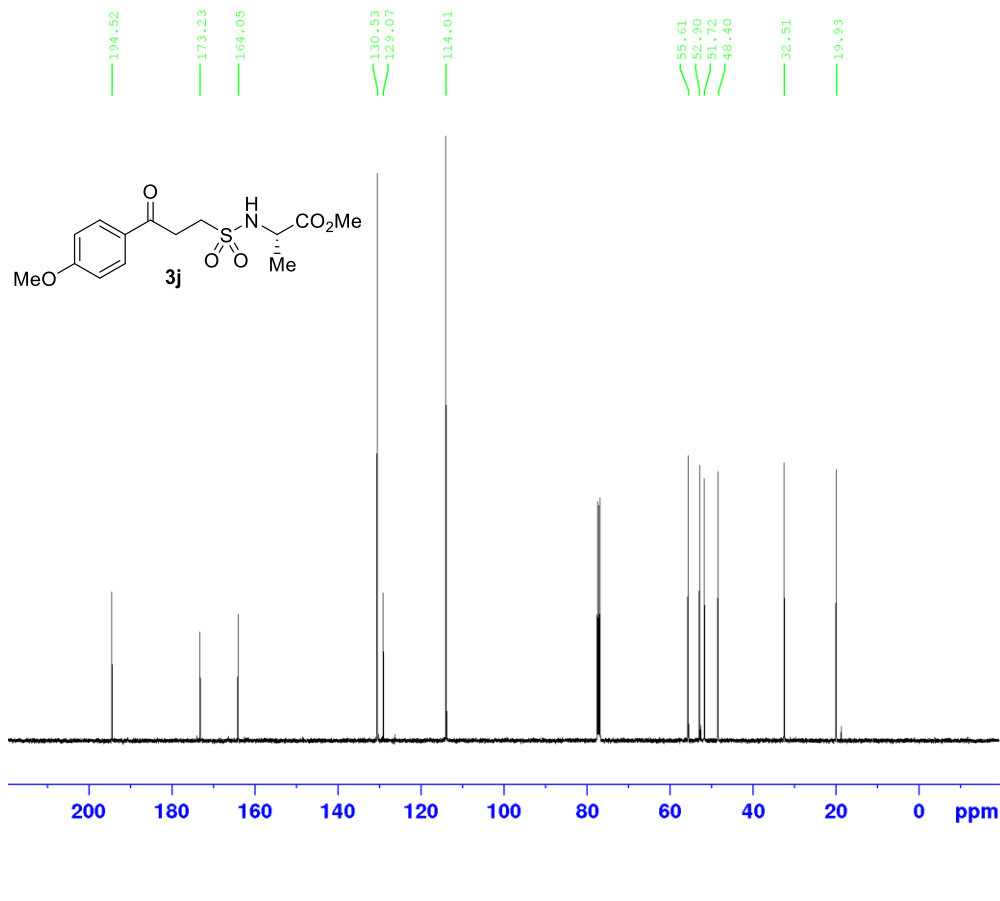
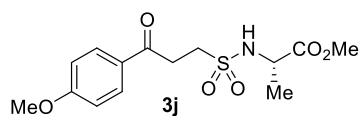
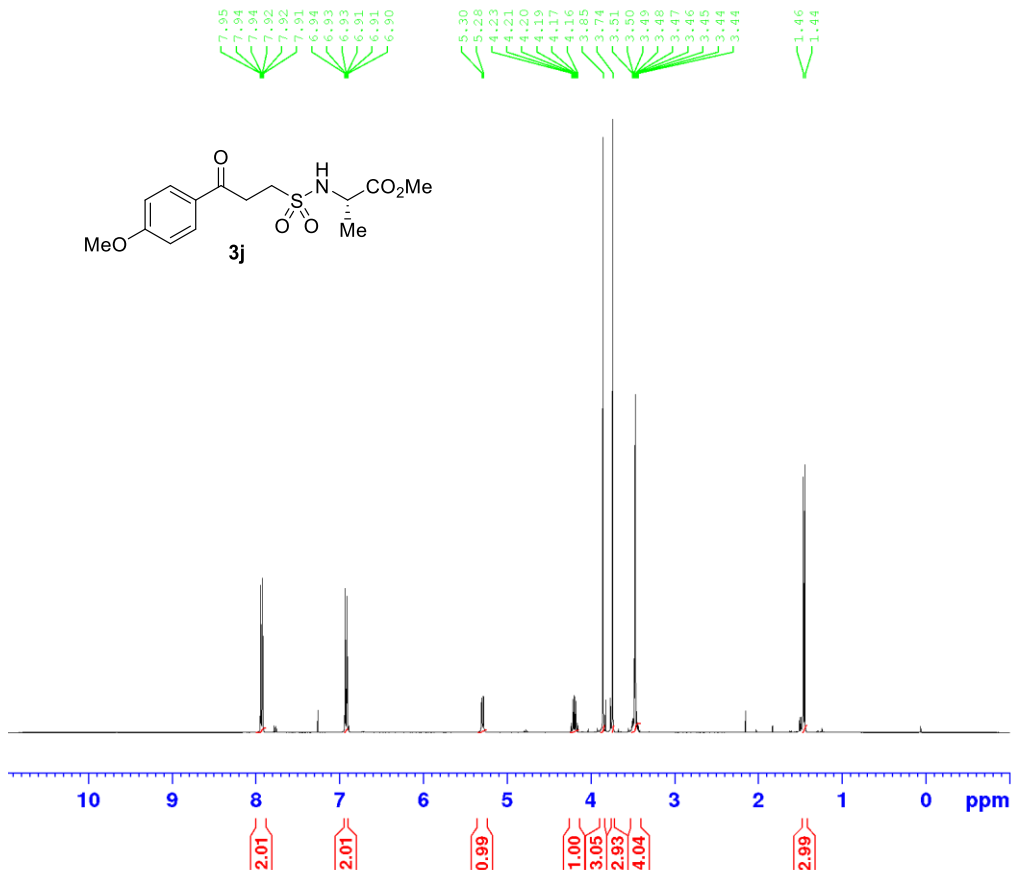
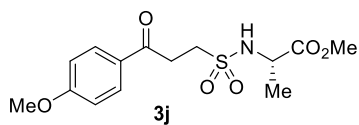
Current Data Parameters
 NAME epb801-prod-13C
 EXPNO 10
 PROCNO 1

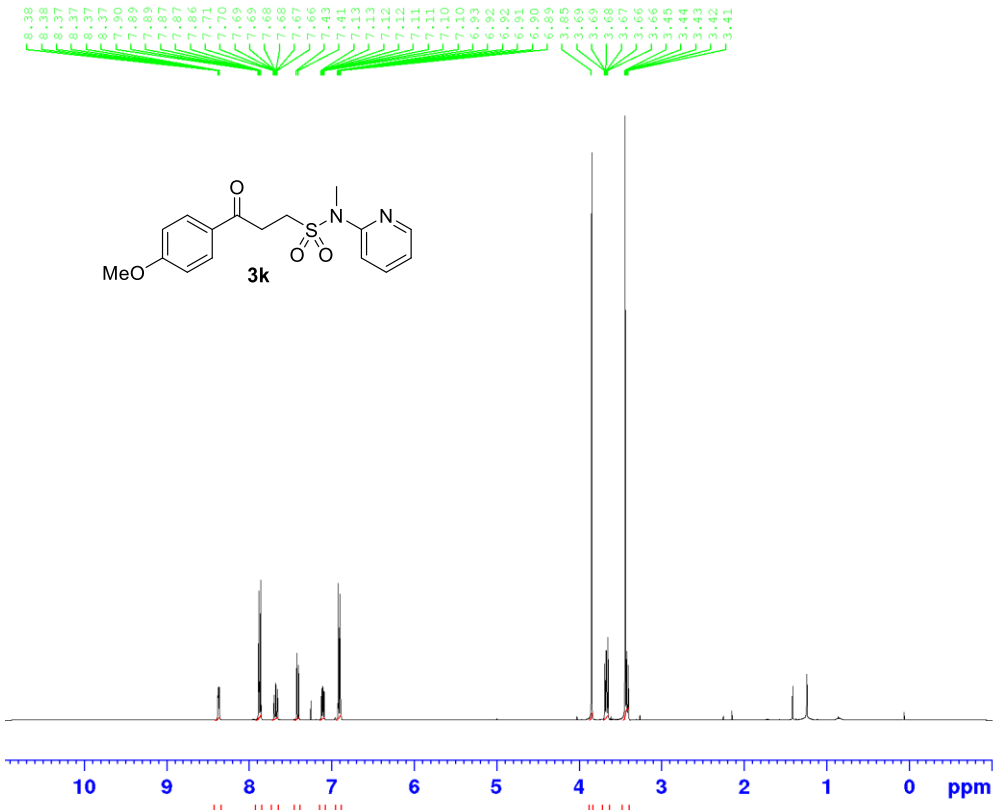
F2 - Acquisition Parameters
 Date_ 20190319
 Time 22.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 26008
 DW 20.850 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127583 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



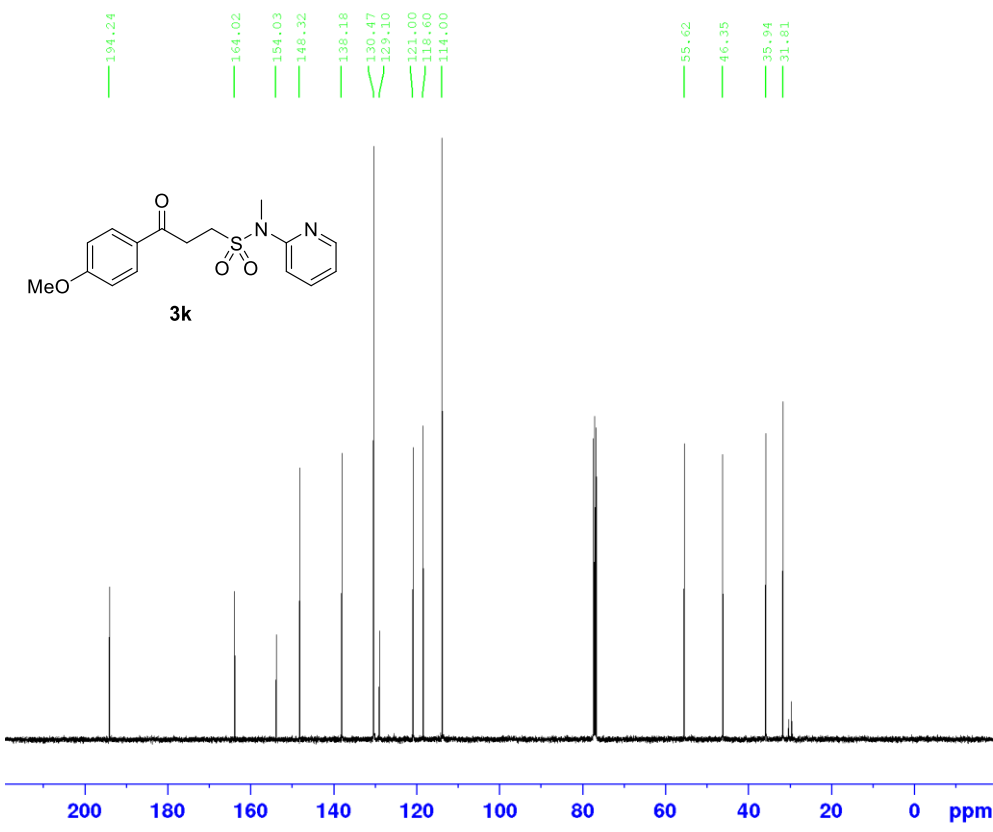


Current Data Parameters
 NAME epb818-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190611
 Time 8.37
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 90.5
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300095 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



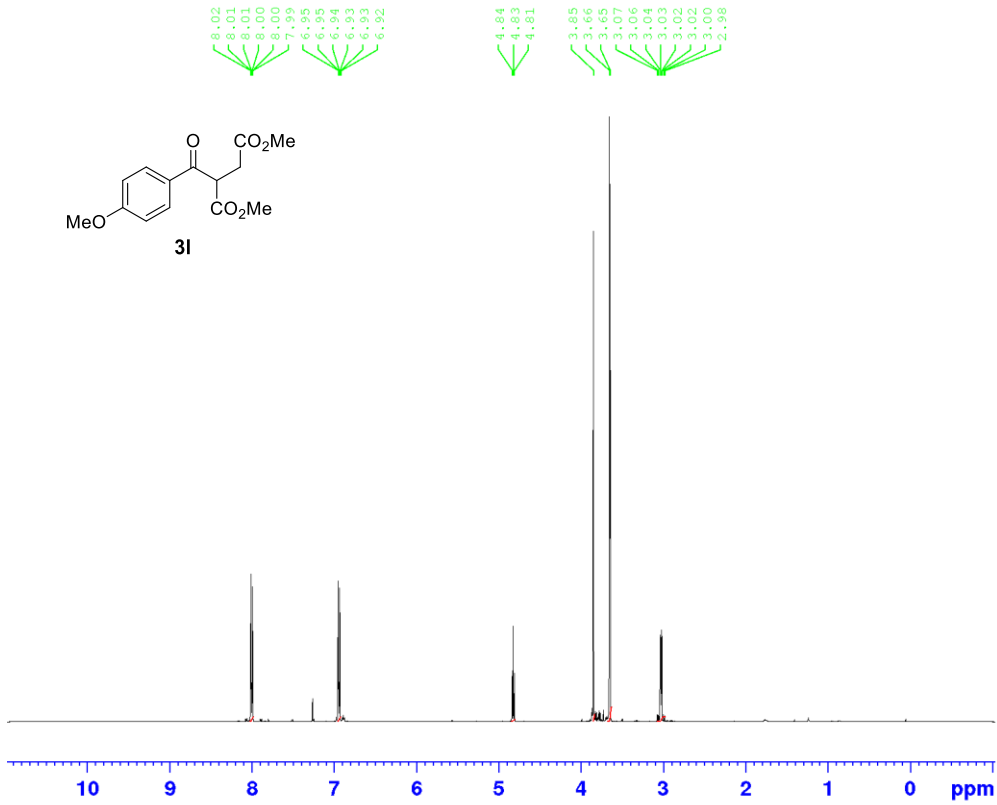
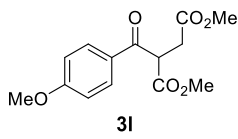
Current Data Parameters
 NAME epb818-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190611
 Time 9.00
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 26008
 DW 20.850 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127596 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

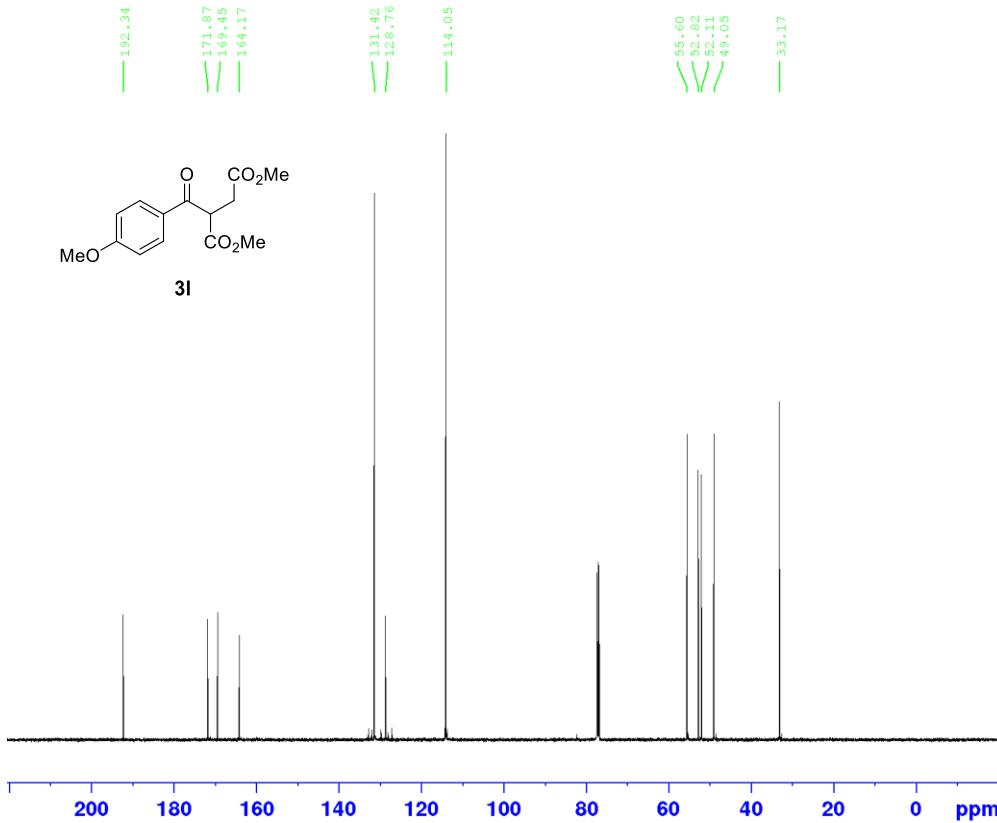
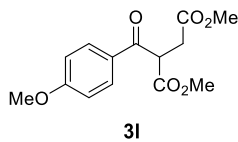


Current Data Parameters
NAME epb742-prod-1H
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190520
Time 18.04
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 6009.615 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 50.8
DW 83.200 usec
DE 10.00 usec
TE 298.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 12.50 usec
PL1 0 dB
PL1W 13.68978119 W
SFO1 500.1325007 MHz

F2 - Processing parameters
SI 16384
SF 500.1300130 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 1.00



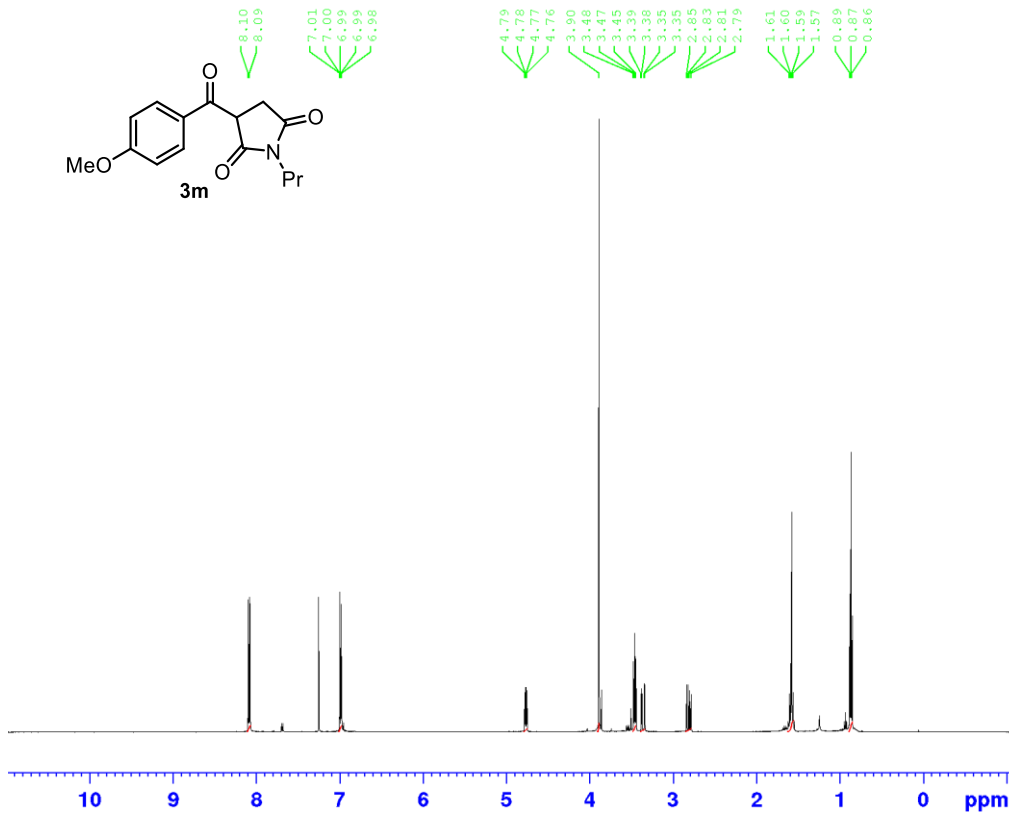
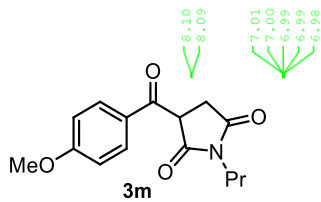
Current Data Parameters
NAME epb742-prod-13C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190520
Time 23.37
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 512
DS 4
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813440 sec
RG 9200
DW 16.500 usec
DE 10.00 usec
TE 298.2 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 8.60 usec
PL1 0 dB
PL1W 53.75436783 W
SFO1 125.7703648 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 0 dB
PL12 16.12 dB
PL13 16.12 dB
PL2W 13.68978119 W
PL12W 0.33450022 W
PL13W 0.33450022 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577808 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

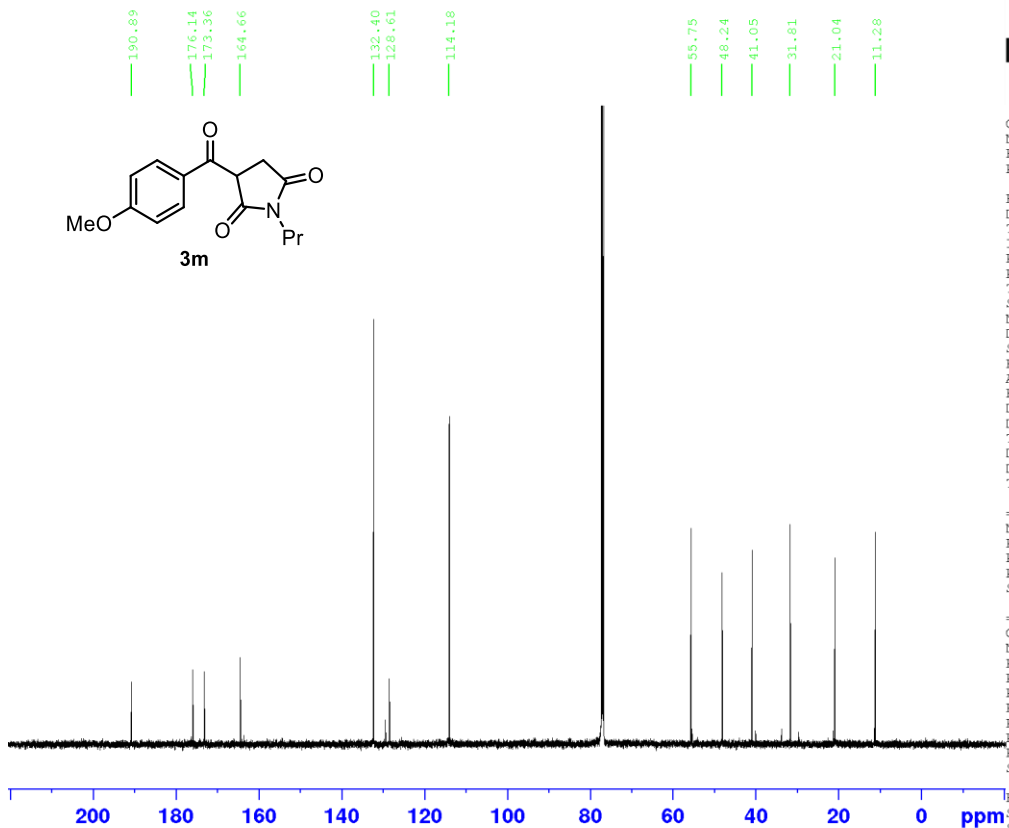
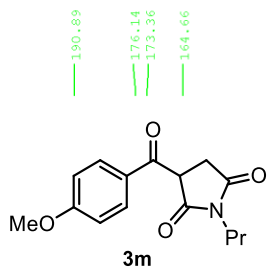


Current Data Parameters
 NAME epb-2-2-f1-HPLC
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190724
 Time 15.01
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.163399 Hz
 AQ 2.7262976 sec
 RG 406
 DW 83.200 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.00 usec
 PL1 -4.08 dB
 PL1W 35.02648163 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300138 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



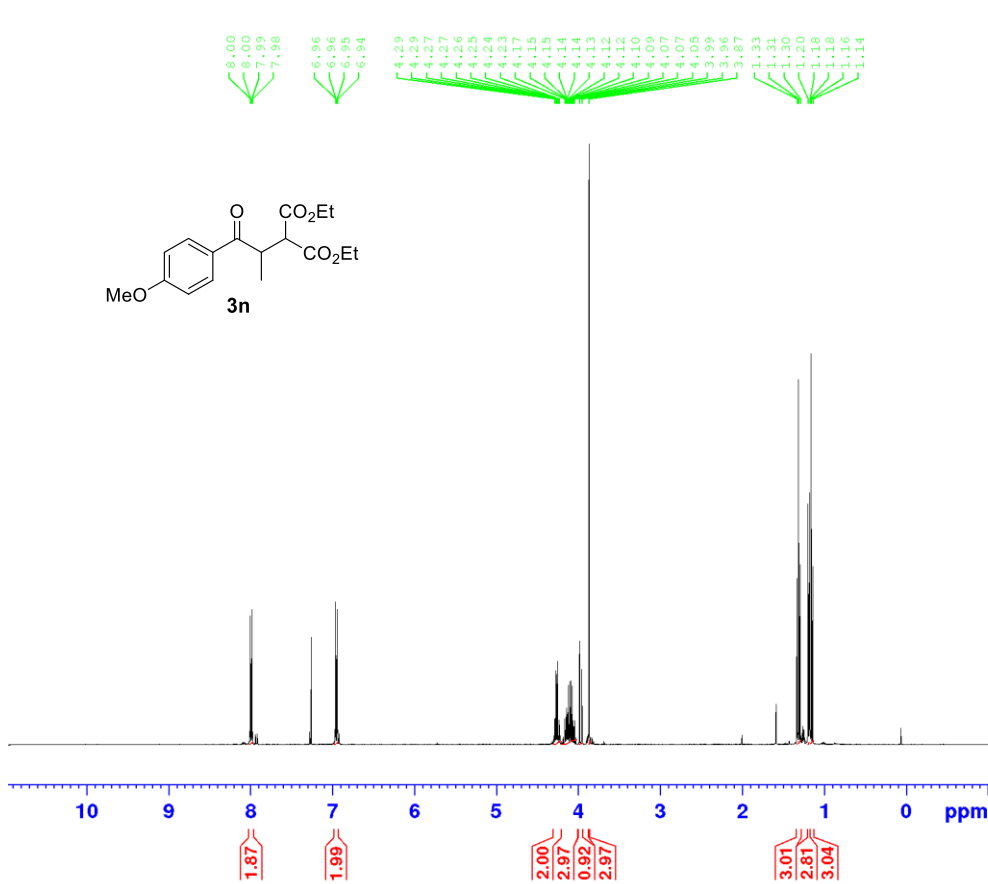
Current Data Parameters
 NAME epb-2-2-f1-HPLC-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190724
 Time 22.08
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 2048
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 13000
 DW 16.500 usec
 DE 10.00 usec
 TE 299.4 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.08 dB
 PL12 11.54 dB
 PL13 11.54 dB
 PL2W 35.02648163 W
 PL12W 0.96027696 W
 PL13W 0.96027696 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577723 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

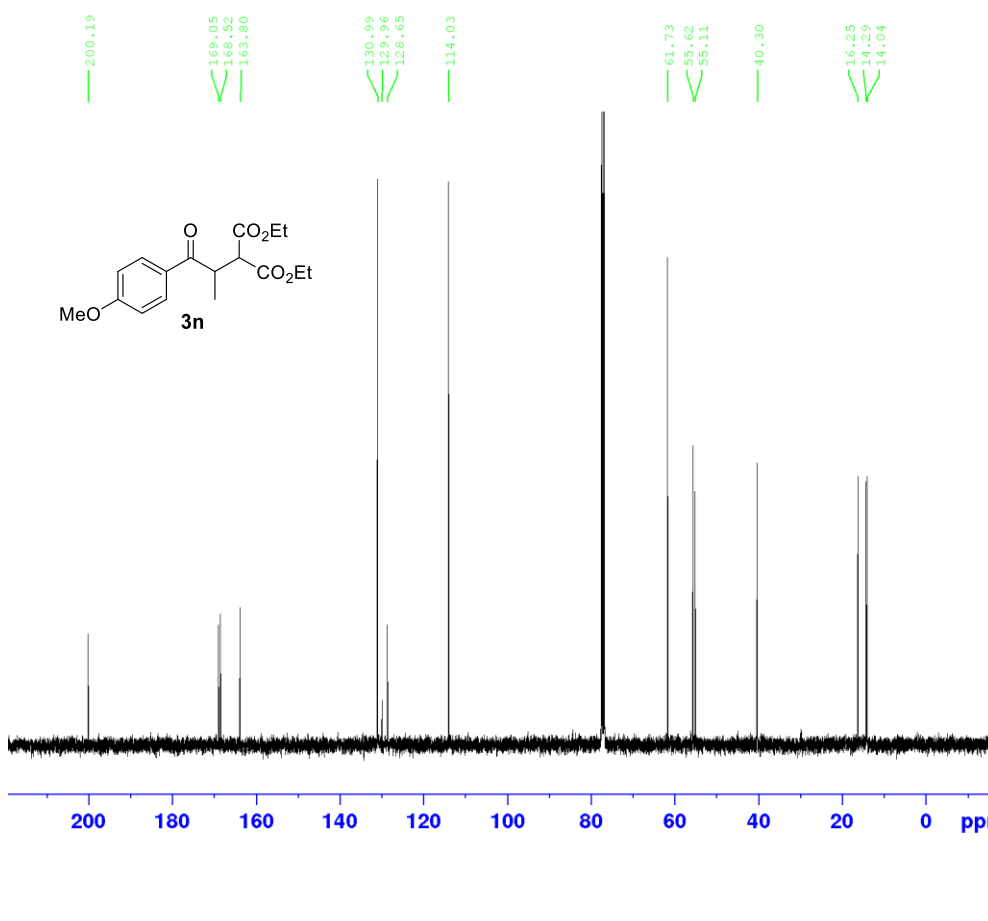


Current Data Parameters
 NAME epb750-f1-1
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190609
 Time 15.50
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 181
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300095 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



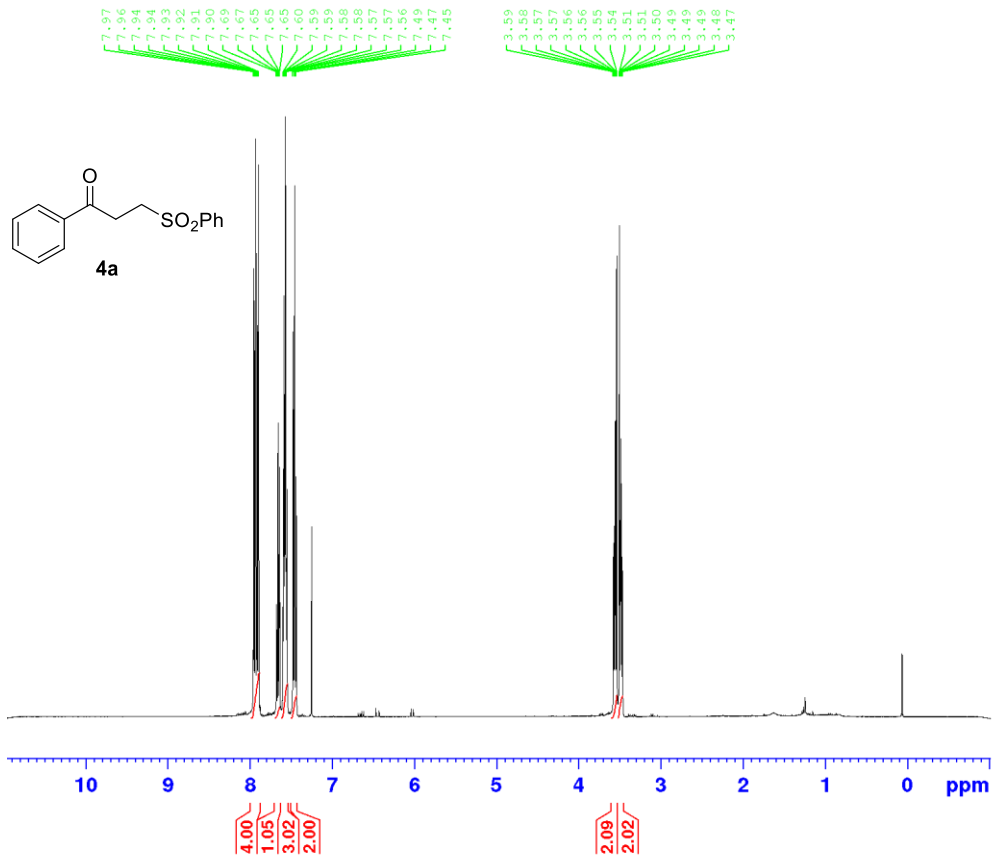
Current Data Parameters
 NAME epb750-f1-1-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190609
 Time 16.12
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 26008
 DW 20.850 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127555 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

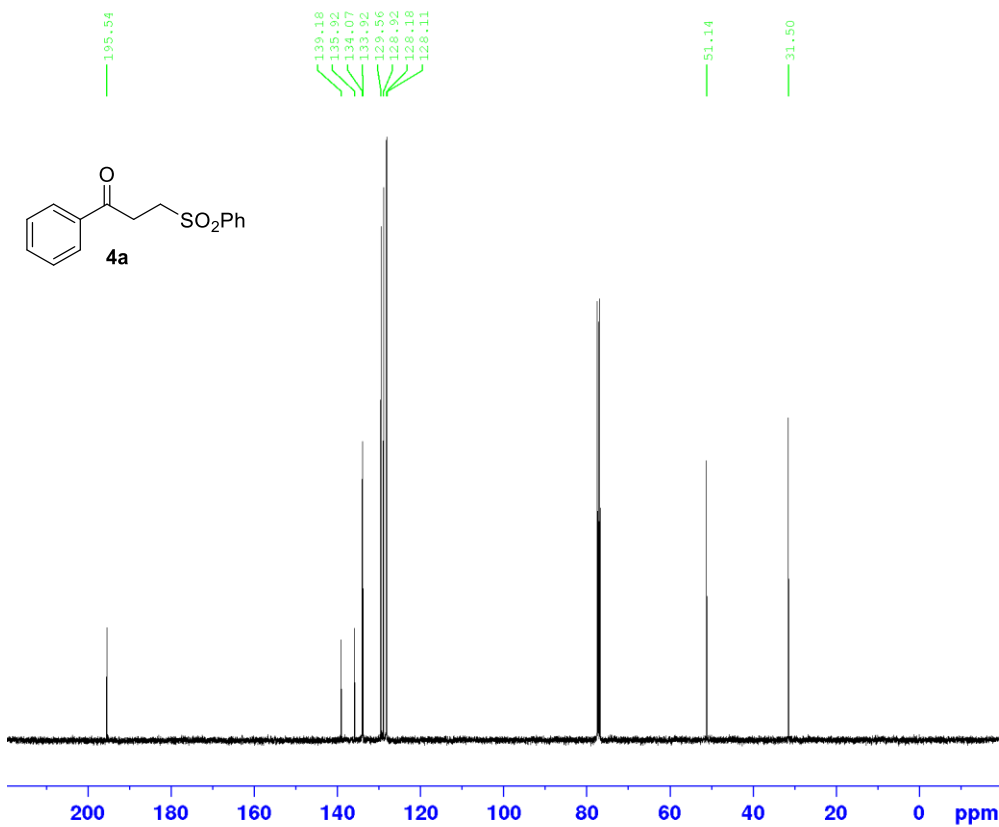


Current Data Parameters
 NAME epb696-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190611
 Time 9.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 161.3
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300097 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



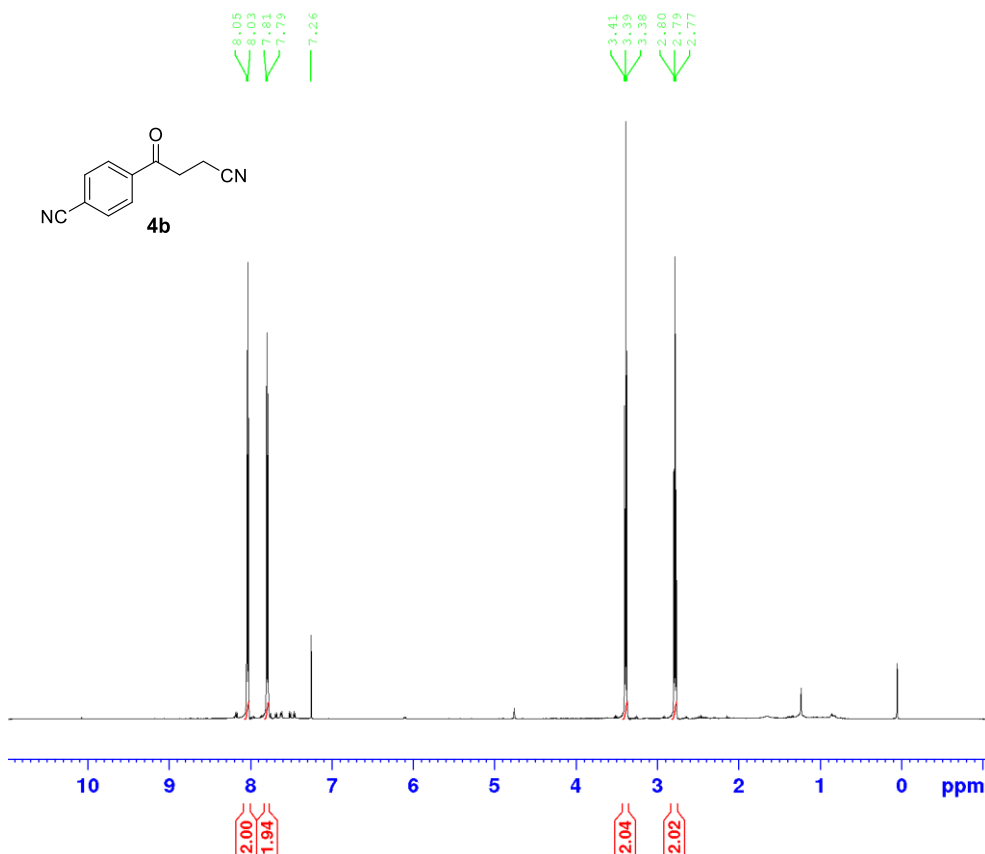
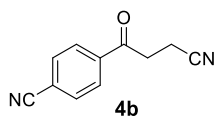
Current Data Parameters
 NAME epb696-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190611
 Time 18.27
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 20642.5
 DW 20.800 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6303736 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -3.00 dB
 PL12 13.05 dB
 PL13 13.05 dB
 PL2W 30.07123375 W
 PL12W 0.74670875 W
 PL13W 0.74670875 W
 SFO2 400.1616006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6203016 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

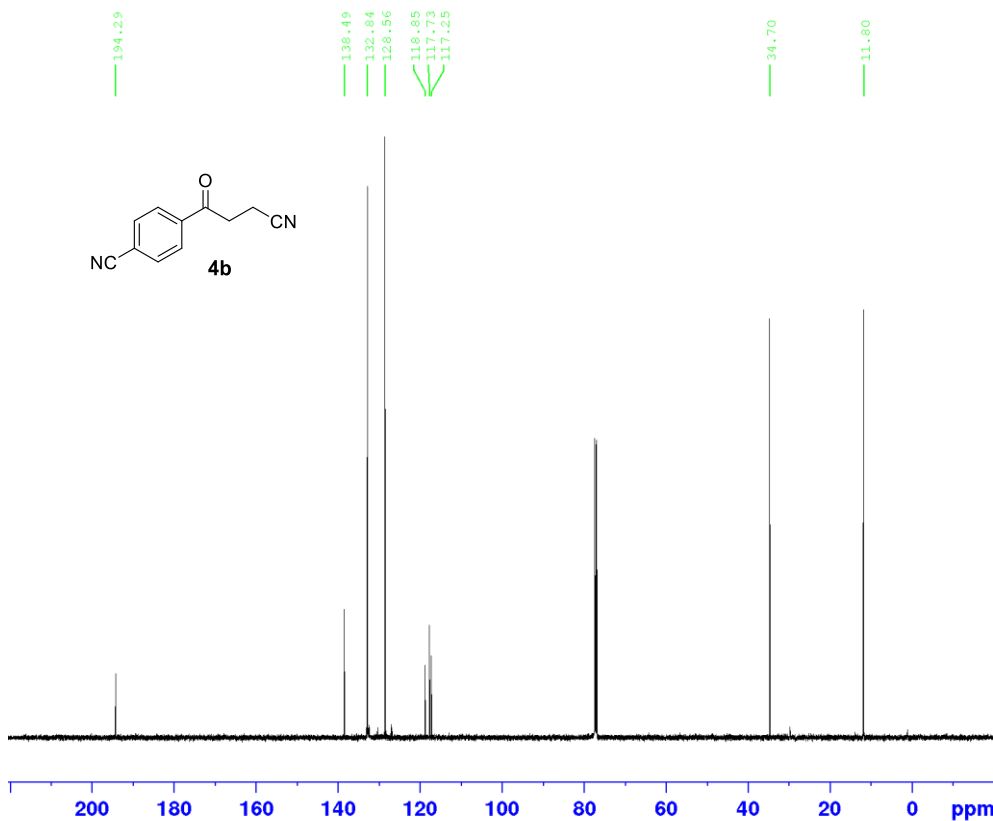
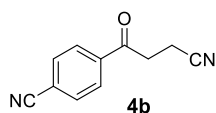


Current Data Parameters
NAME epb745-prod
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190315
Time 18.45
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 6009.615 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 144
DW 83.200 usec
DE 10.00 usec
TE 298.2 K
D1 1.0000000 sec
TDO 1

==== CHANNEL f1 =====
NUC1 1H
P1 12.50 usec
PL1 0 dB
PL1W 13.68978119 W
SFO1 500.1325007 MHz

F2 - Processing parameters
SI 16384
SF 500.1300134 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 1.00



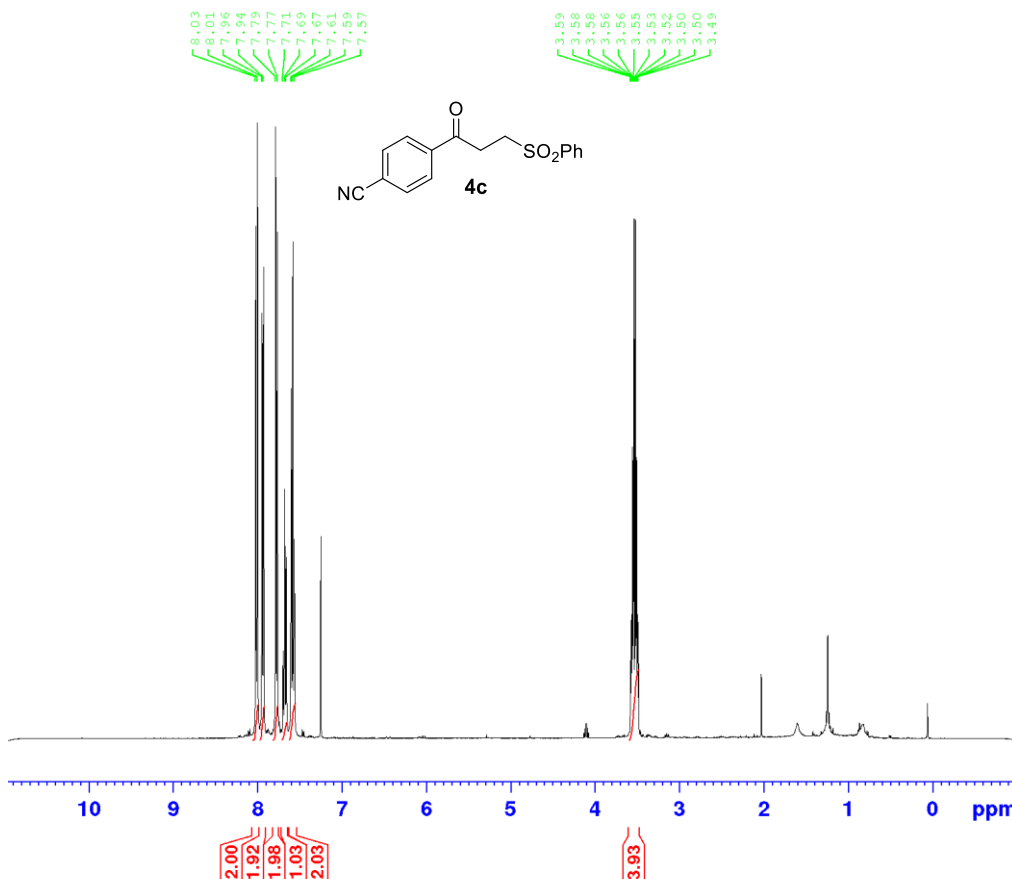
Current Data Parameters
NAME epb745-prod-13C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190315
Time 15.32
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 512
DS 4
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813440 sec
RG 14600
DW 16.500 usec
DE 10.00 usec
TE 298.3 K
D1 1.0000000 sec
D11 0.0300000 sec
TDO 1

==== CHANNEL f1 =====
NUC1 13C
P1 8.60 usec
PL1 0 dB
PL1W 53.75436783 W
SFO1 125.7703648 MHz

==== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 0 dB
PL12 16.12 dB
PL13 16.12 dB
PL2W 13.68978119 W
PL12W 0.33450022 W
PL13W 0.33450022 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577805 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

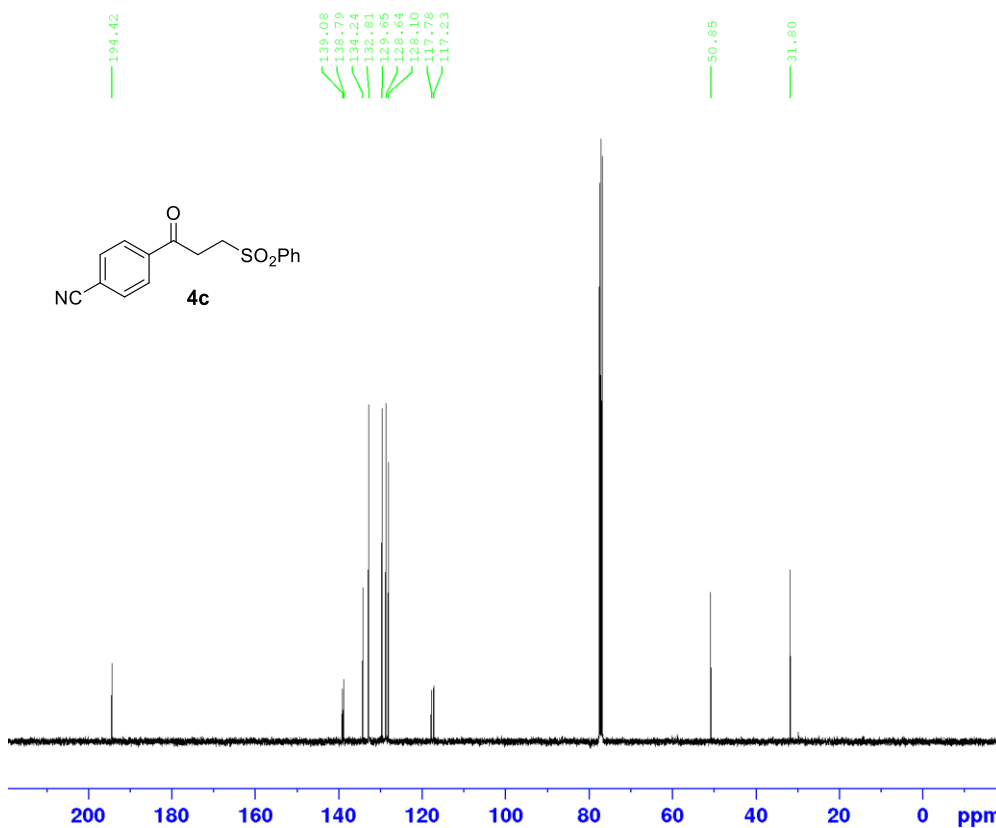


Current Data Parameters
NAME epb835-f1
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190305
Time 1.12
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 32
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4209793 sec
RG 181
DW 104.400 usec
DE 6.00 usec
TE 298.0 K
D1 1.00000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.50 usec
PL1 -2.00 dB
PL1W 23.88643074 W
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300098 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



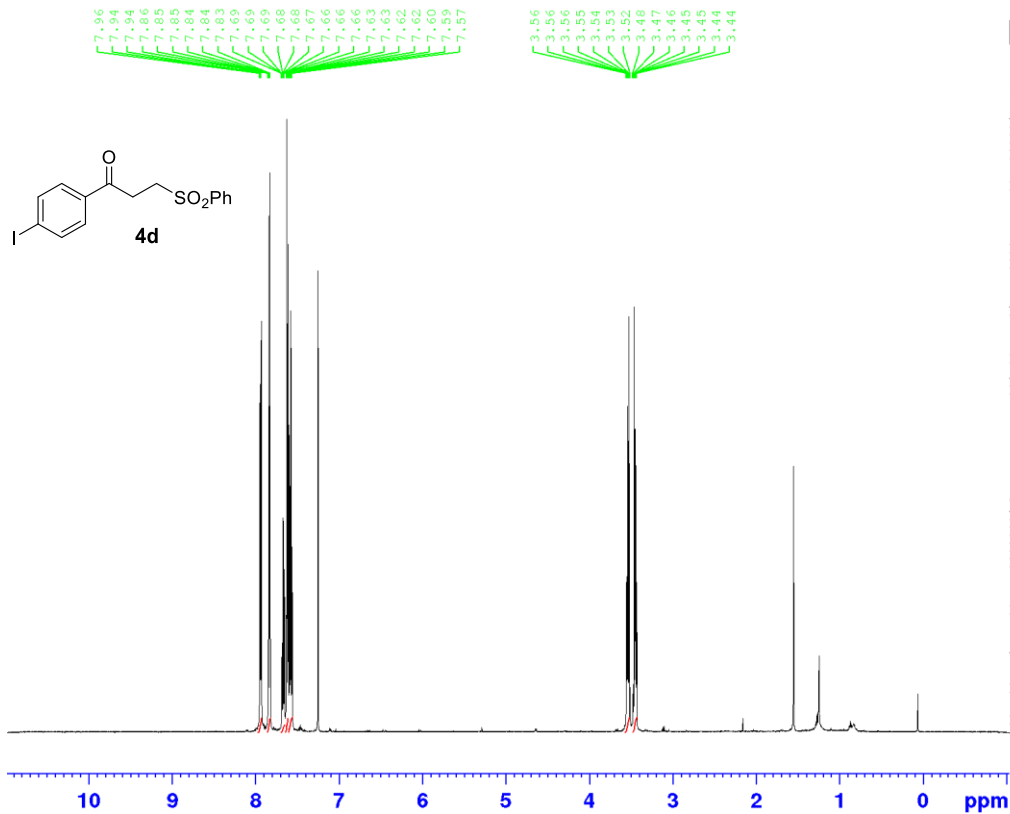
Current Data Parameters
NAME epb835-f1-13C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190305
Time 1.34
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 512
DS 4
SWH 23980.814 Hz
FIDRES 0.365918 Hz
AQ 1.3664256 sec
RG 32768
DW 20.850 usec
DE 10.00 usec
TE 298.1 K
D1 1.00000000 sec
D11 0.03000000 sec
TDO 1

===== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -3.00 dB
PL1W 75.17808533 W
SFO1 100.6228298 MHz

===== CHANNEL f2 =====
CPDPRG2 waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL12 12.83 dB
PL13 12.83 dB
PL2W 23.88643074 W
PL12W 0.78550917 W
PL13W 0.78550917 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127570 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

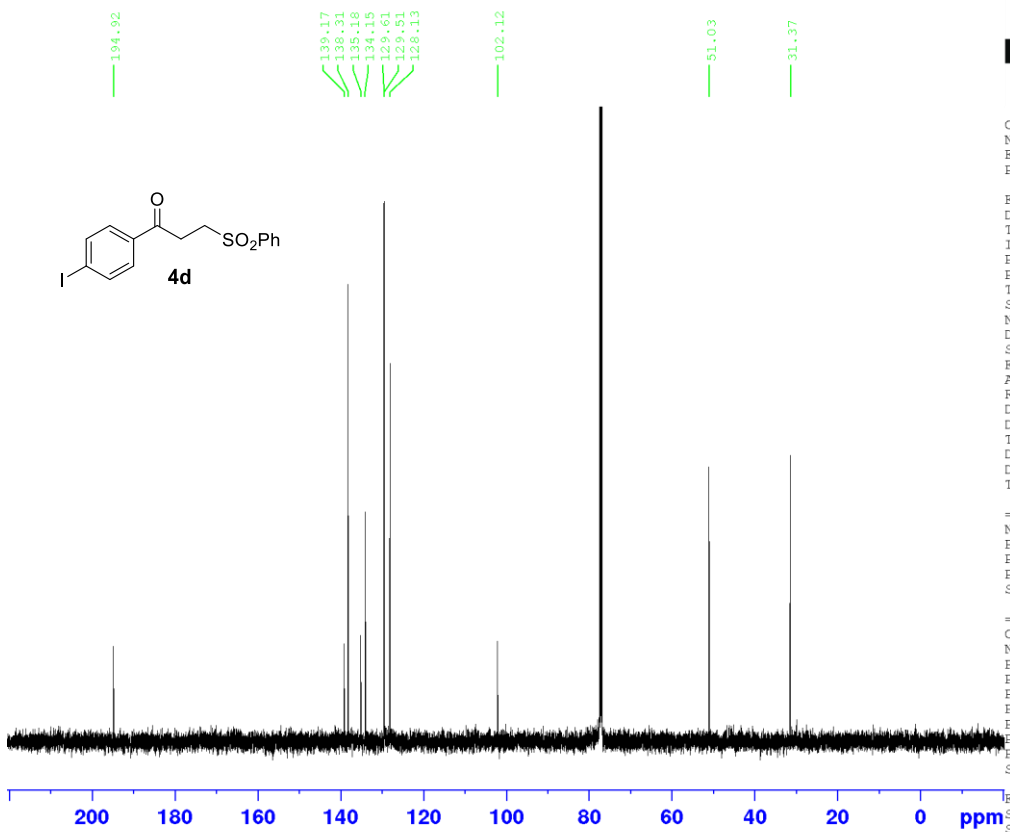


Current Data Parameters
 NAME epb836-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190315
 Time 19.04
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 362
 DW 83.200 usec
 DE 10.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300134 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



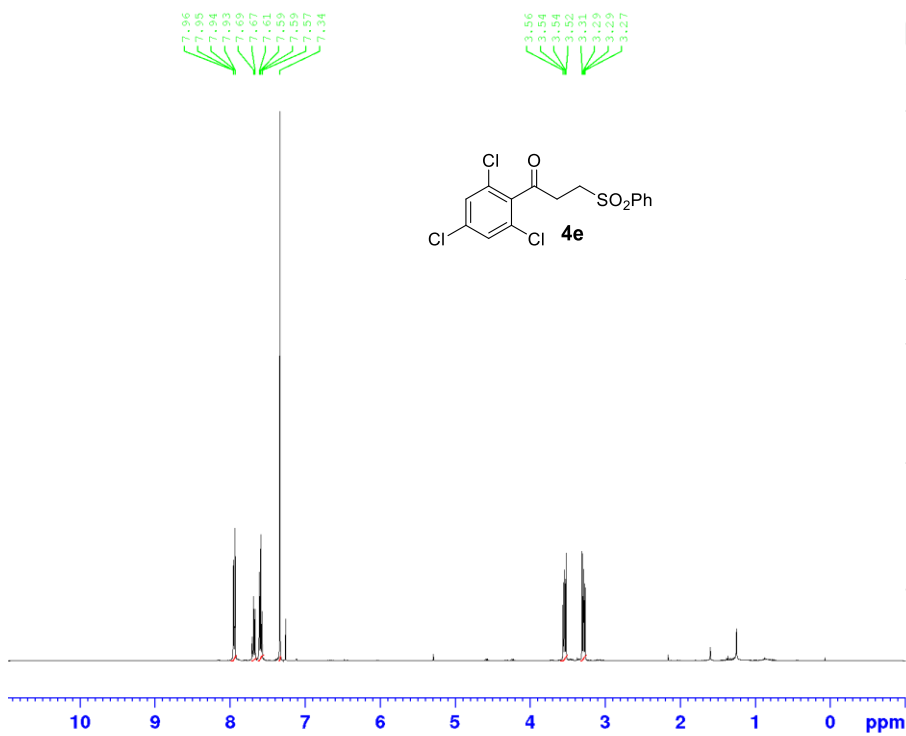
Current Data Parameters
 NAME epb836-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190316
 Time 17.40
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 13000
 DW 16.500 usec
 DE 10.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577727 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

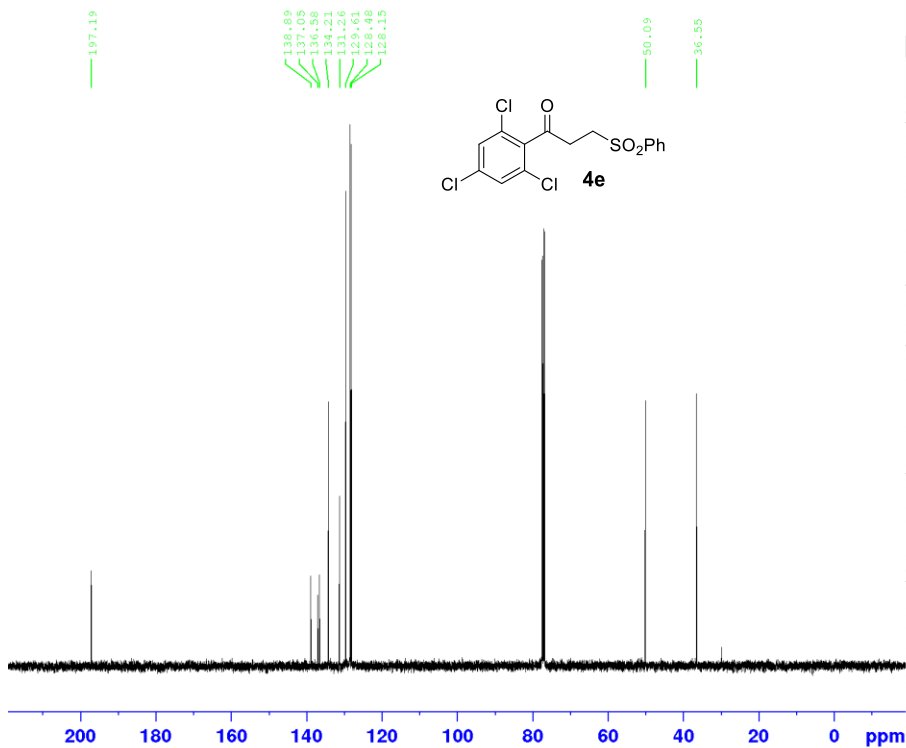


Current Data Parameters
 NAME epb837-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190610
 Time 8.29
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 181
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300098 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



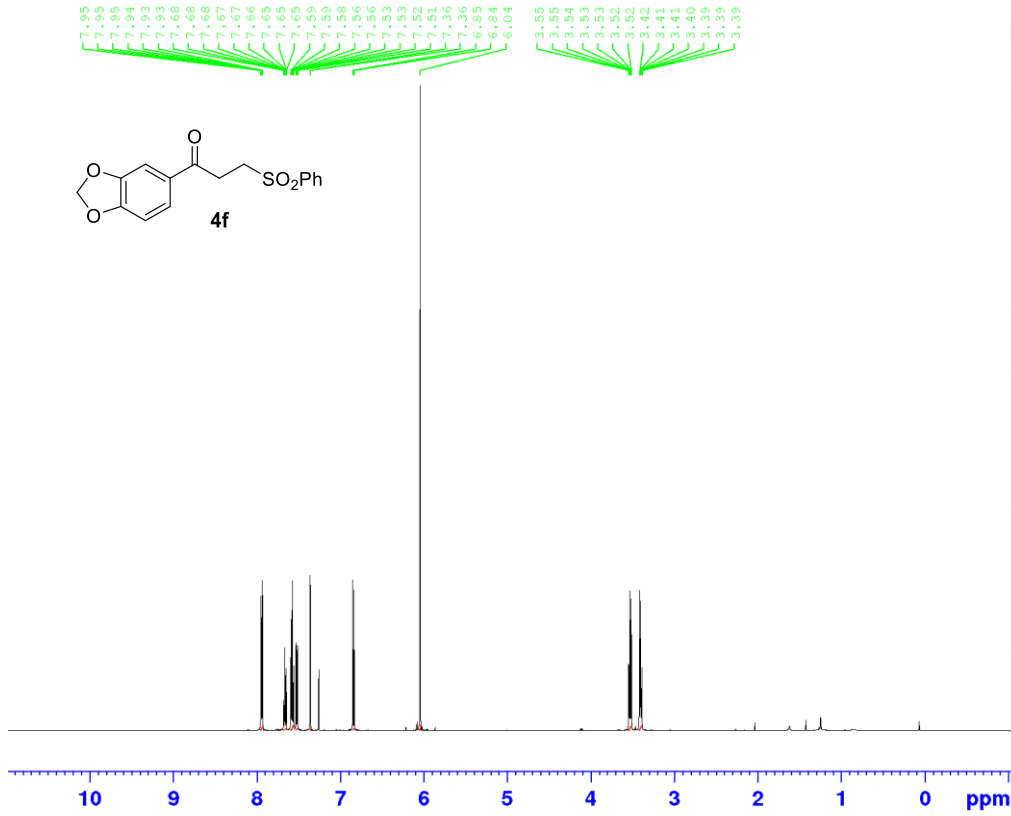
Current Data Parameters
 NAME epb837-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190610
 Time 8.41
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 267
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 23170.5
 DW 20.850 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CDPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127577 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

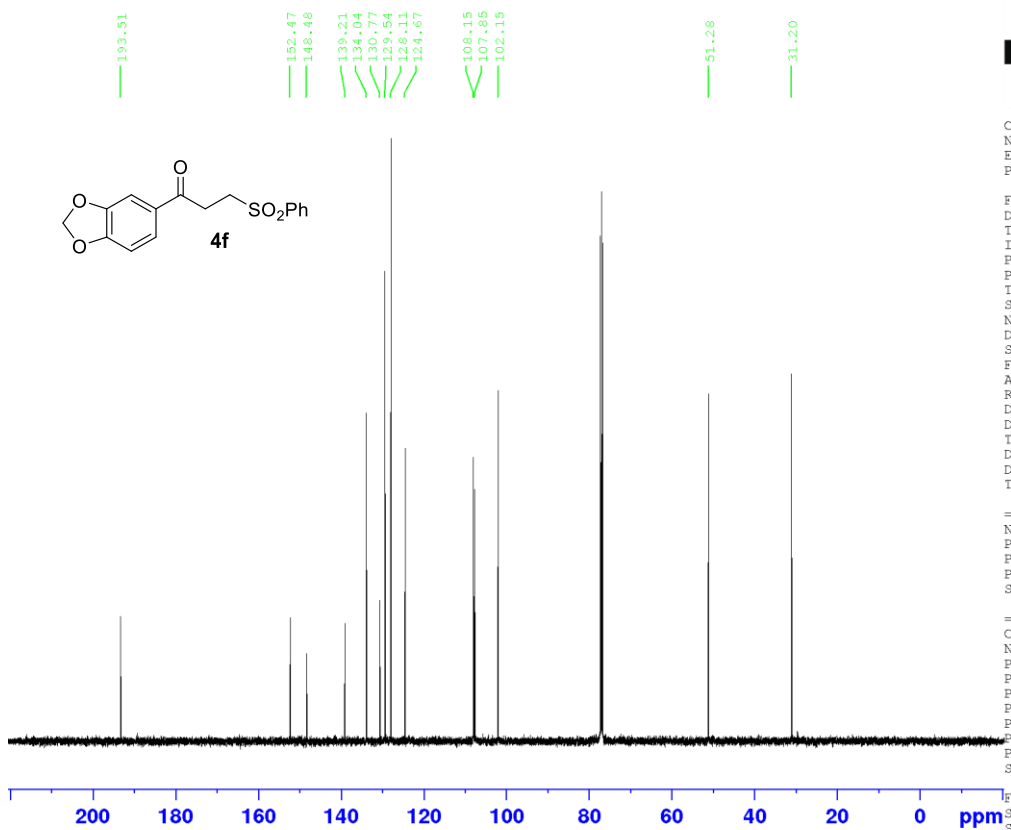


Current Data Parameters
 NAME epb827-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190315
 Time 18.58
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 181
 DW 83.200 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300130 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



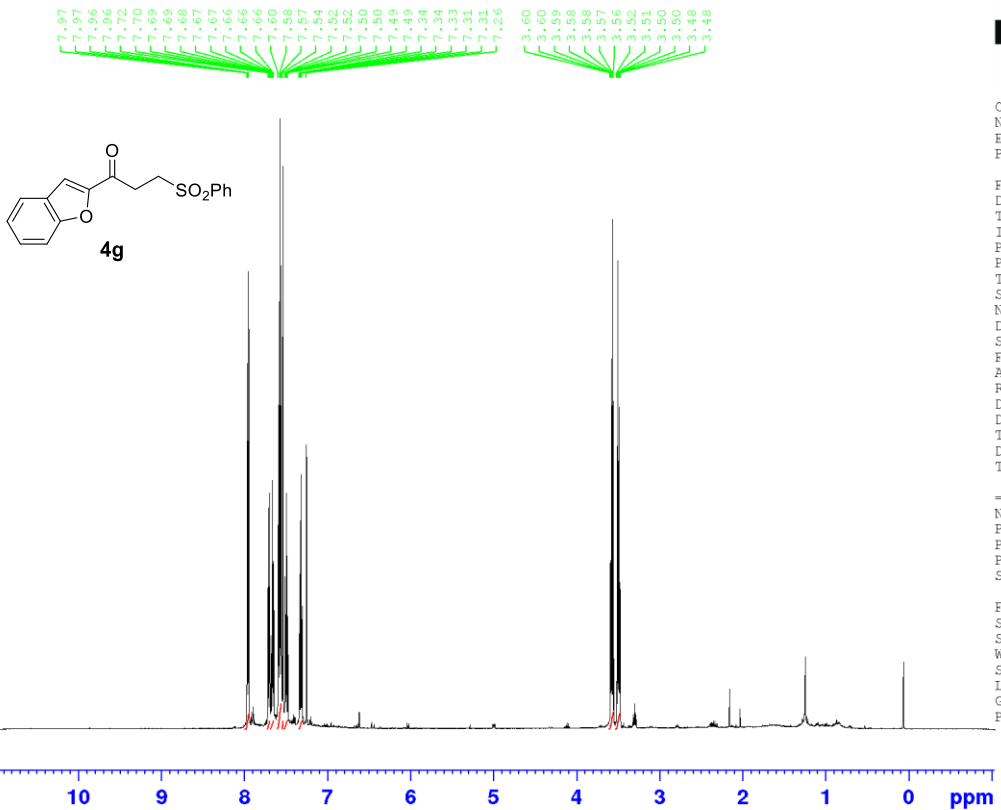
Current Data Parameters
 NAME epb827-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190316
 Time 17.12
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 13000
 DW 16.500 usec
 DE 10.00 usec
 TE 298.3 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL12W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577750 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

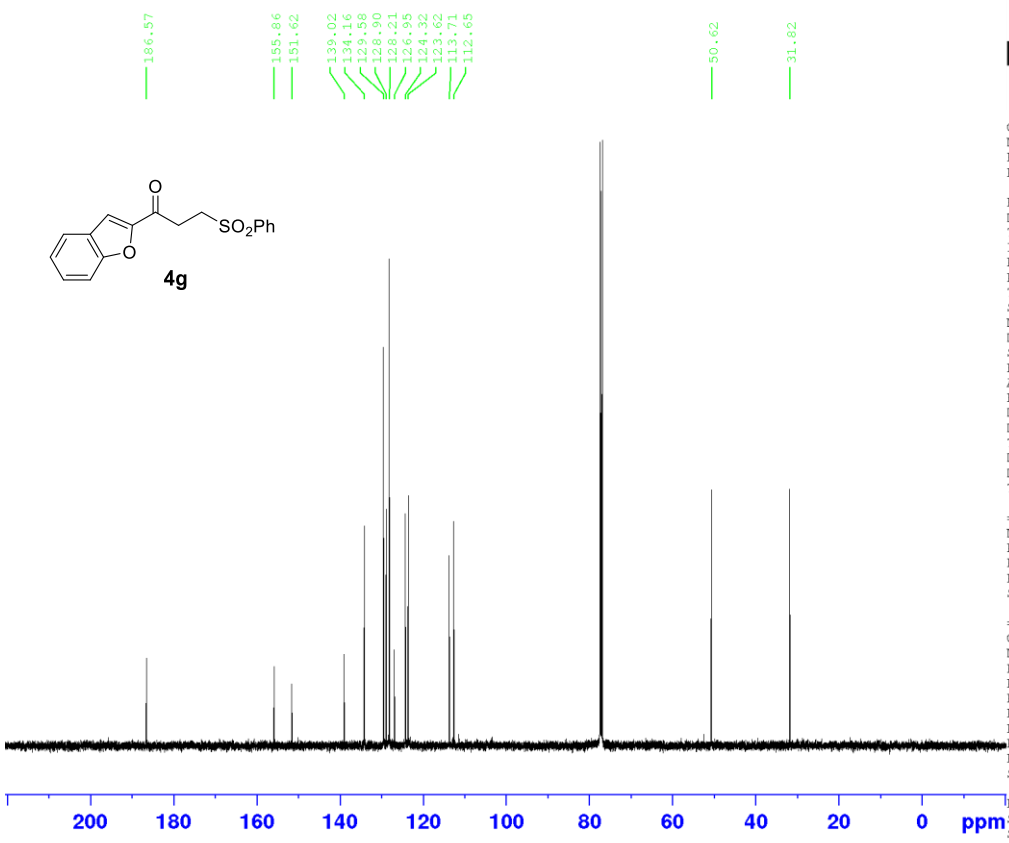


Current Data Parameters
 NAME epb840-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190315
 Time 19.10
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 203
 DW 83.200 usec
 DE 10.00 usec
 TE 297.8 K
 D1 1.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PLL 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300133 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



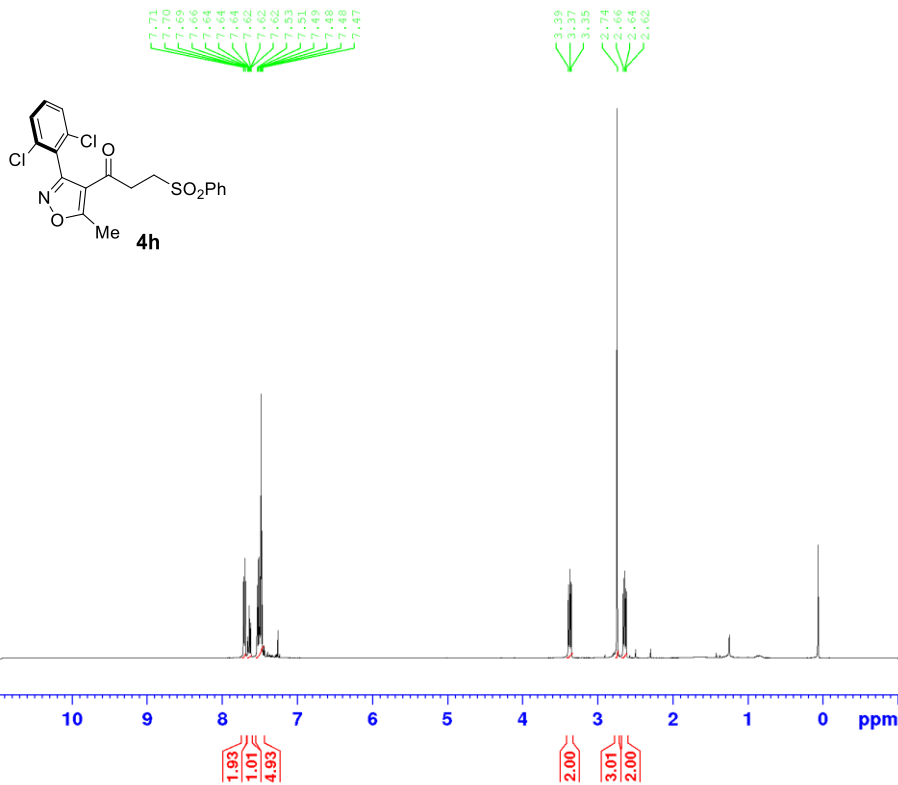
Current Data Parameters
 NAME epb840-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190316
 Time 18.09
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 13000
 DW 16.500 usec
 DE 10.00 usec
 TE 298.3 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PLL 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

==== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577740 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

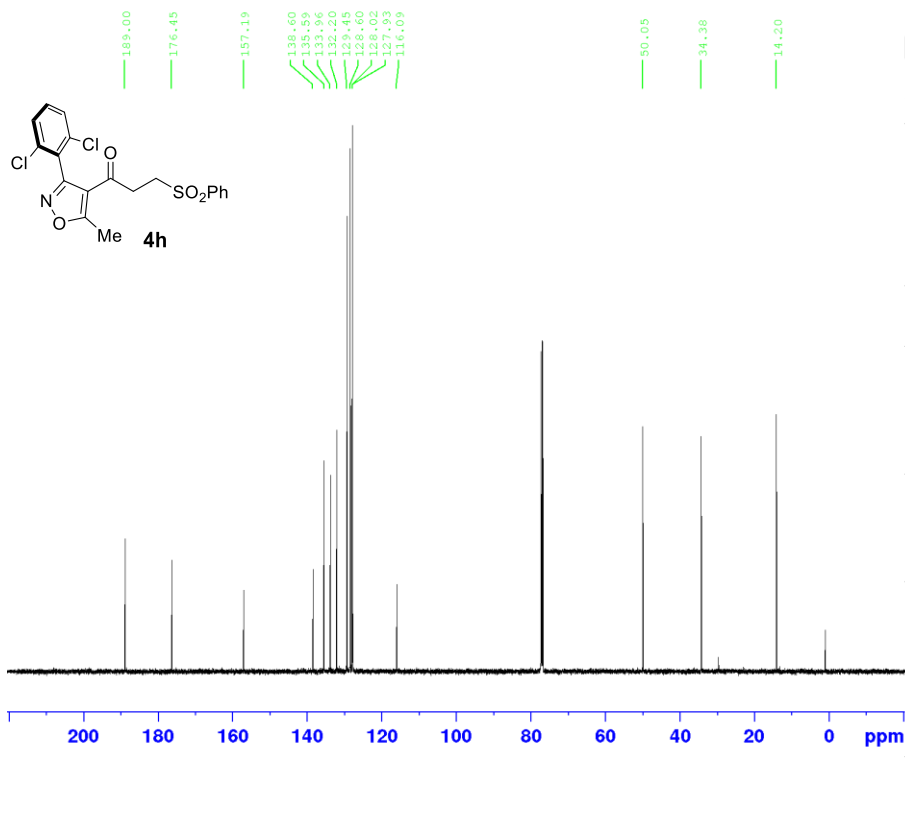


Current Data Parameters
 NAME epb-2-1-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190627
 Time 11.11
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 128
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300098 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



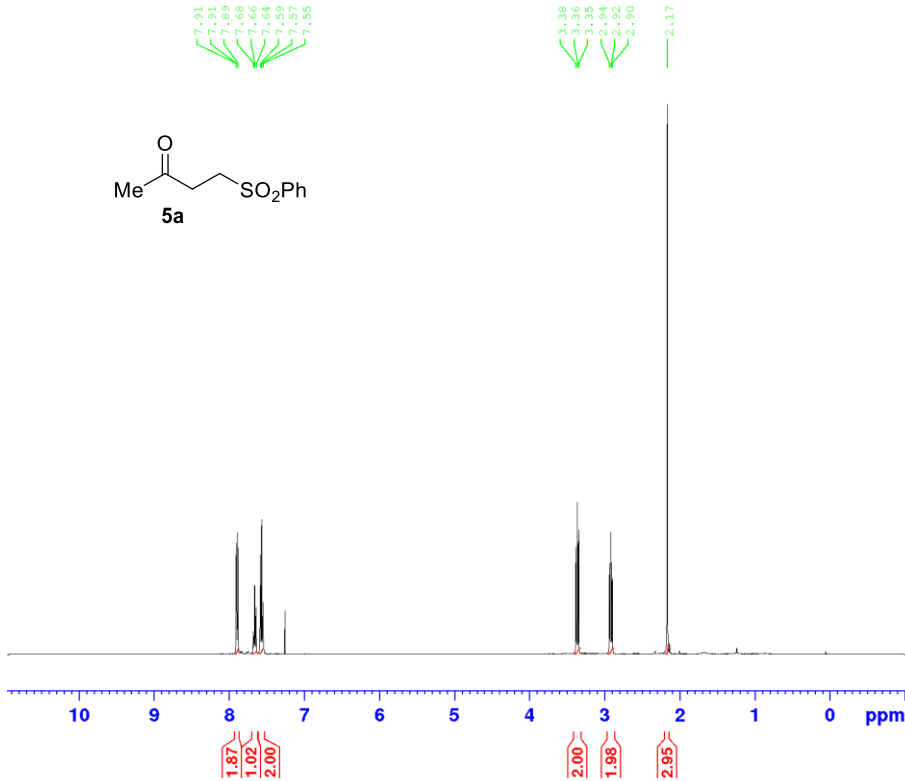
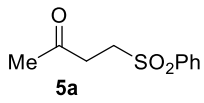
Current Data Parameters
 NAME epb-2-1-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190628
 Time 17.54
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 9200
 DW 16.500 usec
 DE 10.00 usec
 TE 298.6 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.08 dB
 PL12 11.54 dB
 PL13 11.54 dB
 PL2W 35.02648163 W
 PL12W 0.96027696 W
 PL13W 0.96027696 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577787 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

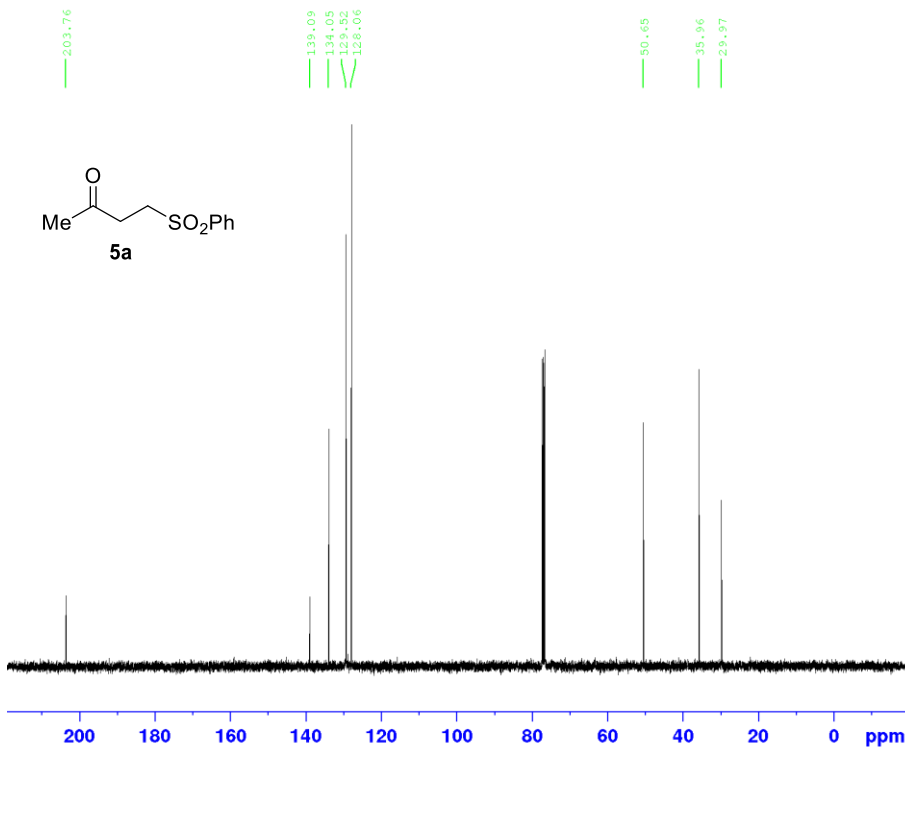
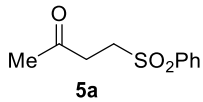


Current Data Parameters
 NAME epb791-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190319
 Time 18.26
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 128
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300097 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



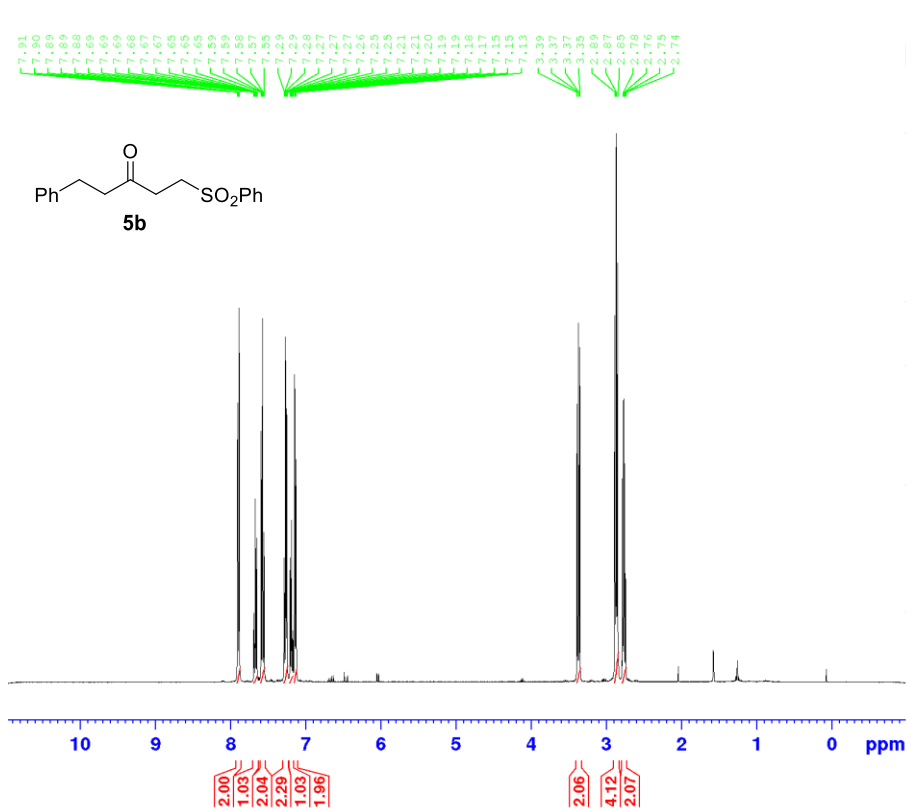
Current Data Parameters
 NAME epb791-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190319
 Time 21.39
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 128
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 32768
 DW 20.850 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127588 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

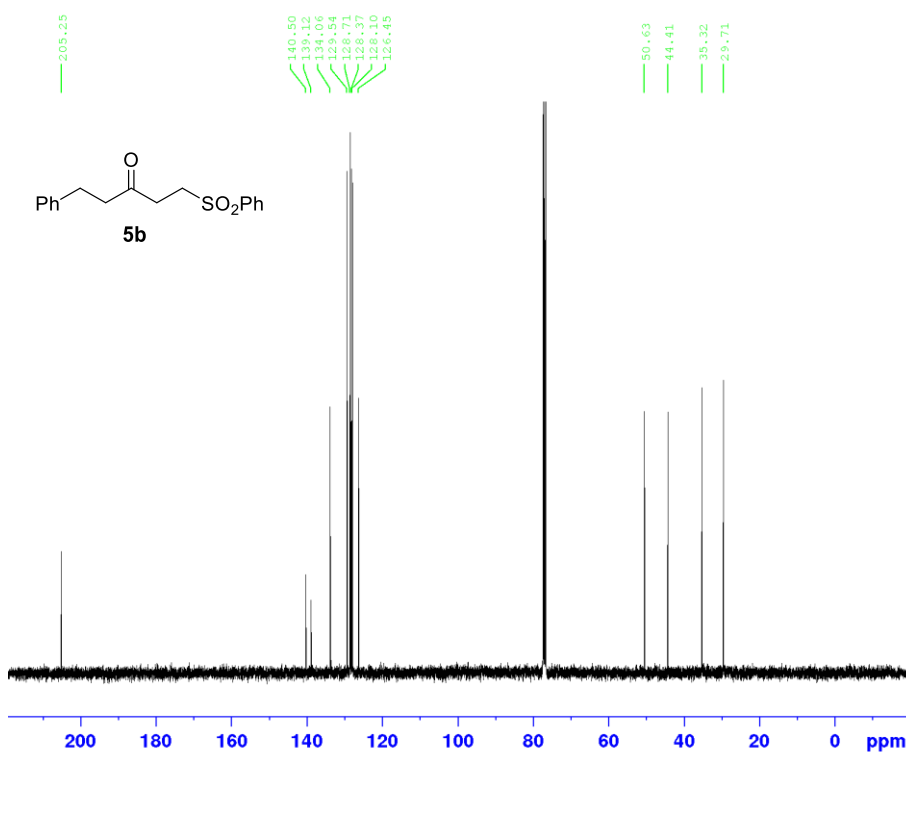


Current Data Parameters
 NAME epb838-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190319
 Time 18.21
 INSTRUM spect
 PROBHD 5 mm PABBO BE-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl₃
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 228.1
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300097 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



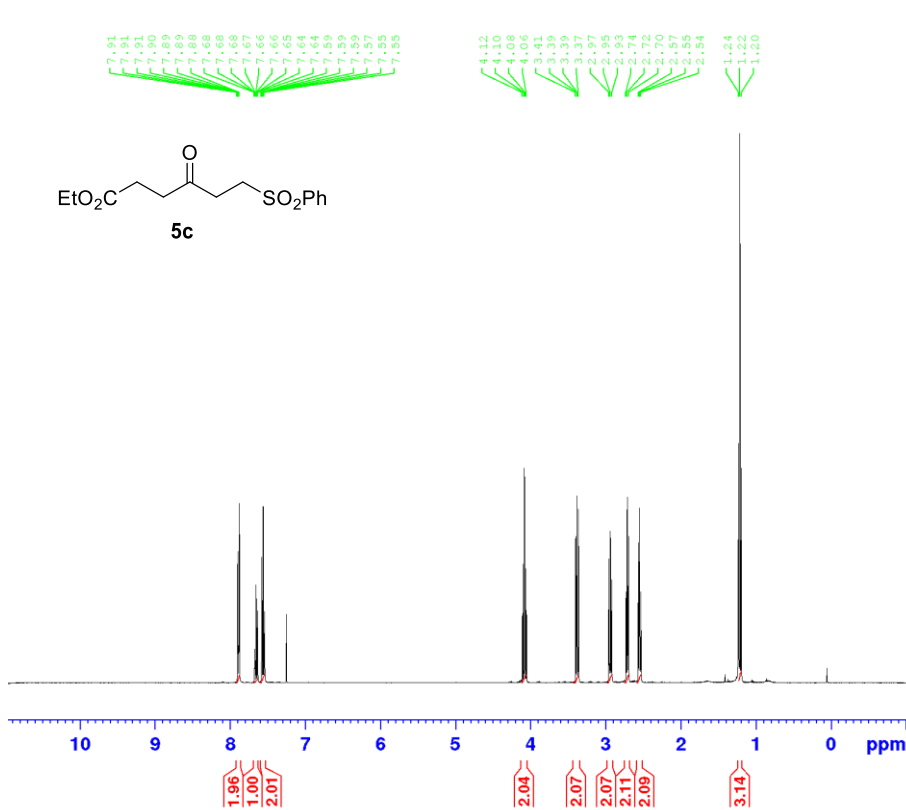
Current Data Parameters
 NAME epb838-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190319
 Time 21.22
 INSTRUM spect
 PROBHD 5 mm PABBO BE-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl₃
 NS 512
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 26008
 DW 20.850 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

==== CHANNEL f2 =====
 CDPGPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127562 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

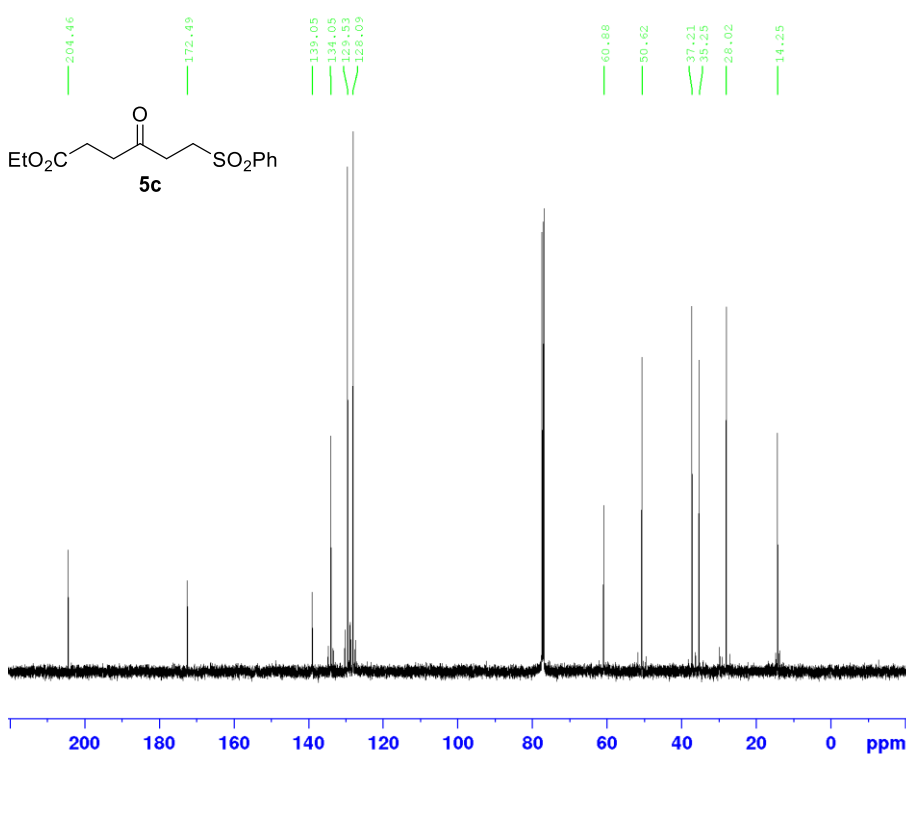


Current Data Parameters
 NAME epb814-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190416
 Time 15.11
 INSTRUM spect
 PROBHD 5 mm PABBO BE-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 4
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 128
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300097 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



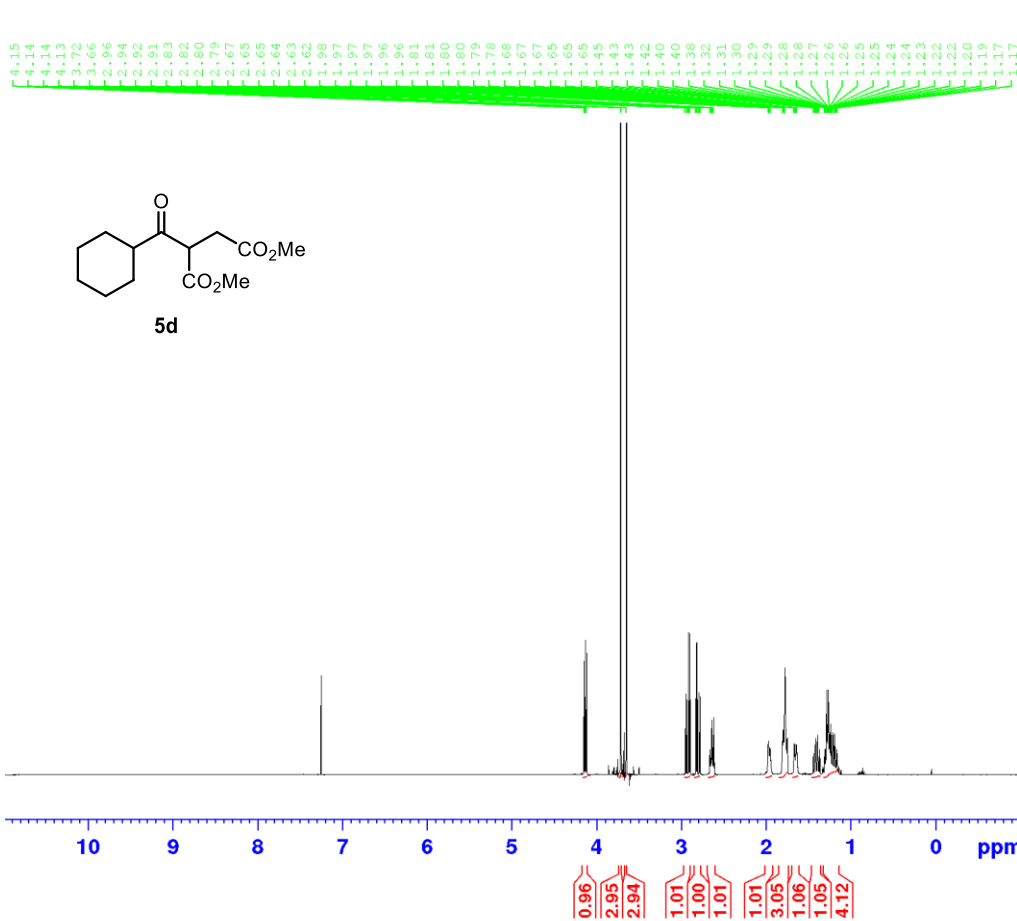
Current Data Parameters
 NAME epb814-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190416
 Time 16.26
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 10300
 DW 16.500 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 16.20 usec
 PL1 -0.50 dB
 PL1W 60.31339645 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -1.00 dB
 PL12 20.80 dB
 PL13 20.80 dB
 PL2W 17.23441315 W
 PL12W 0.11386666 W
 PL13W 0.11386666 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577761 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

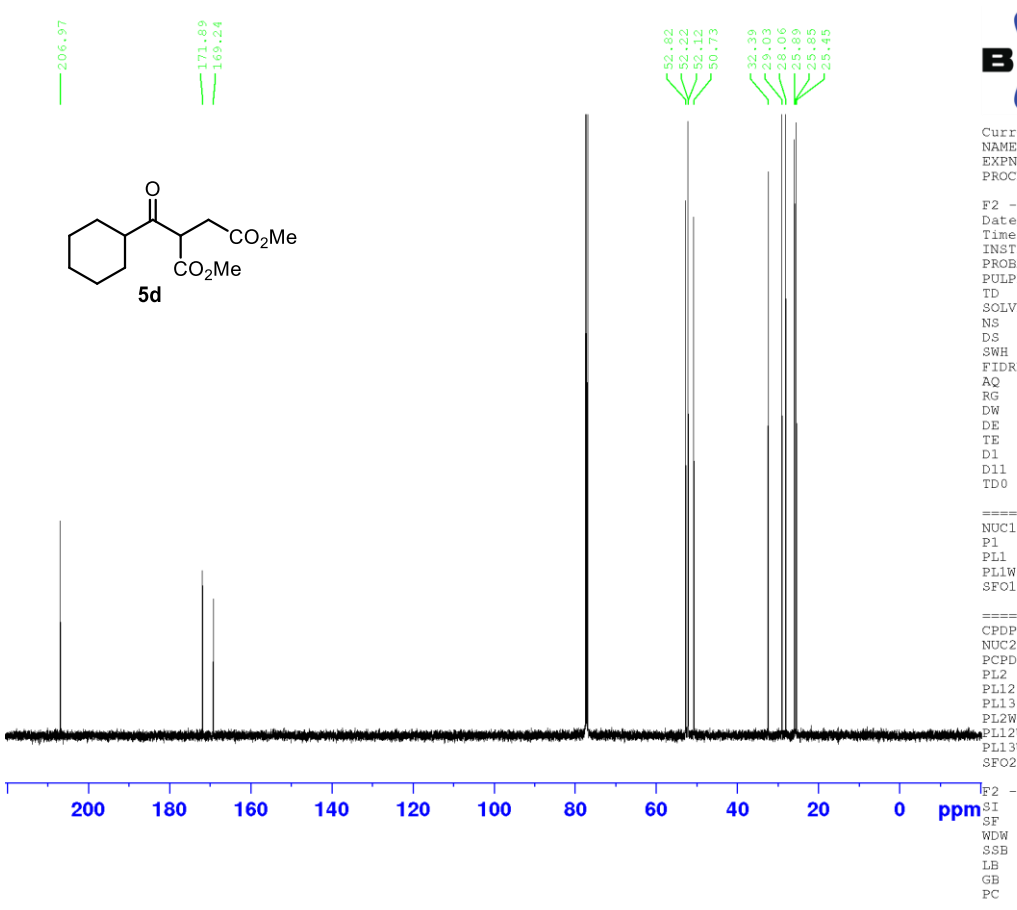
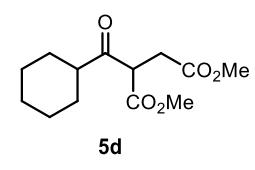


Current Data Parameters
 NAME epb766-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190318
 Time 18.32
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDC13
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 71.8
 DW 83.200 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300130 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



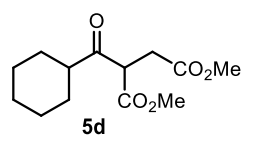
Current Data Parameters
 NAME epb766-prod-13C
 EXPNO 10
 PROCNO 1

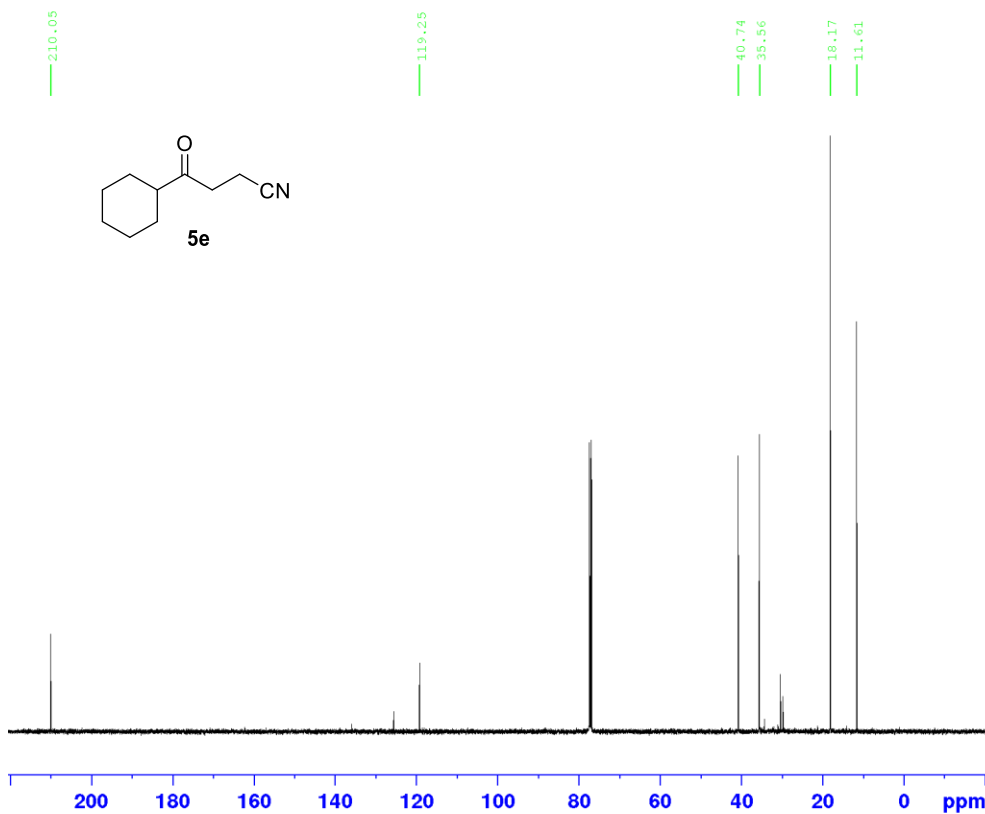
F2 - Acquisition Parameters
 Date_ 20190318
 Time 22.55
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 10300
 DW 16.500 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL12W 13.68978119 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577733 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40





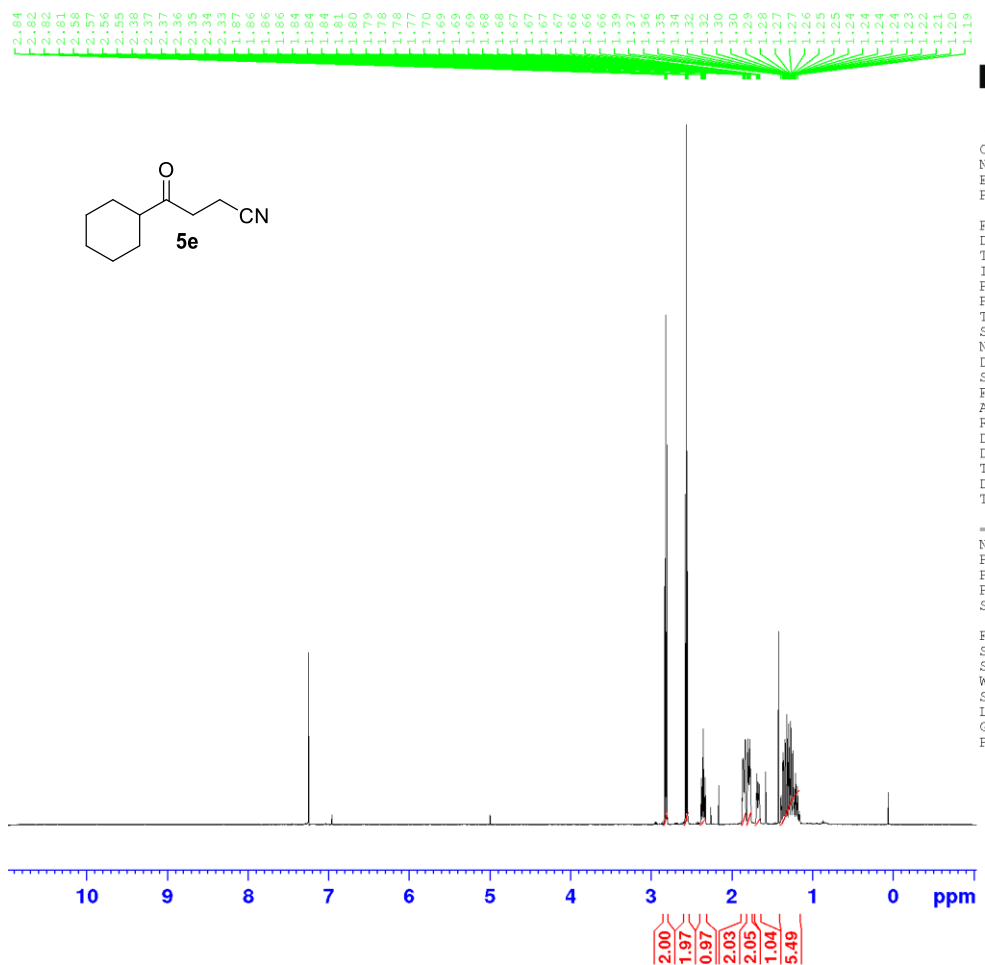
Current Data Parameters
 NAME epb778-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190318
 Time 23.24
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 10300
 DW 16.500 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577776 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

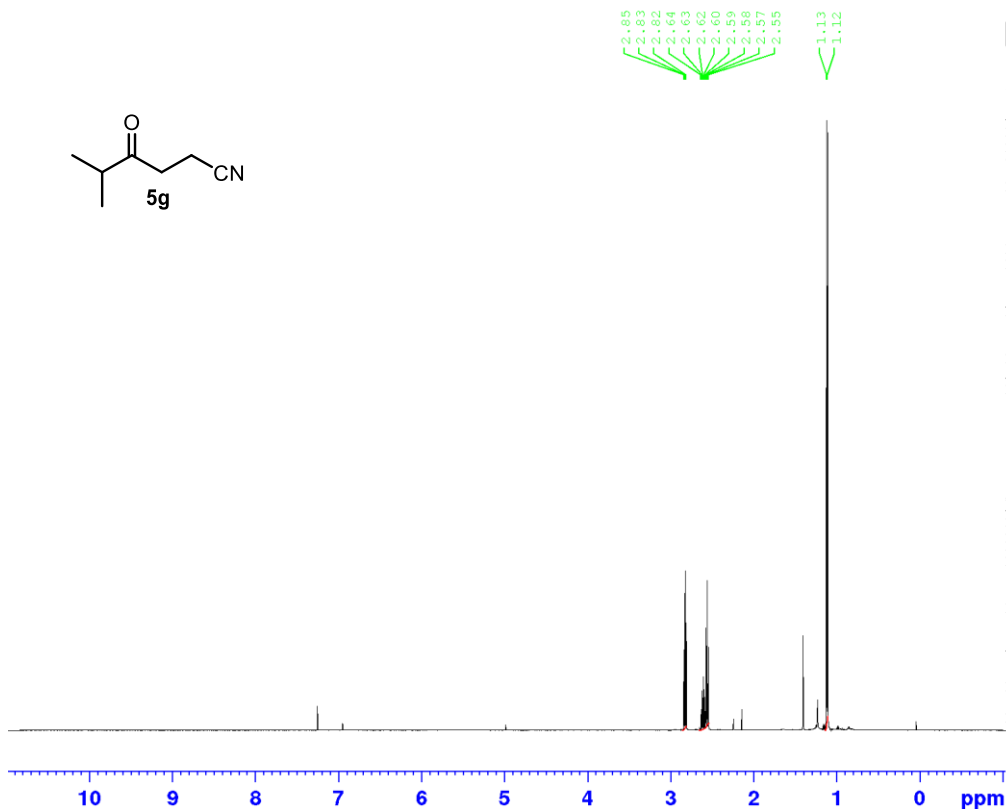
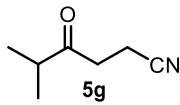


Current Data Parameters
 NAME epb776-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190318
 Time 16.23
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 181
 DW 83.200 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300138 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME epb778-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190318
 Time 18.39
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 64
 DW 83.200 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300134 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00

209.38

119.32

50.58

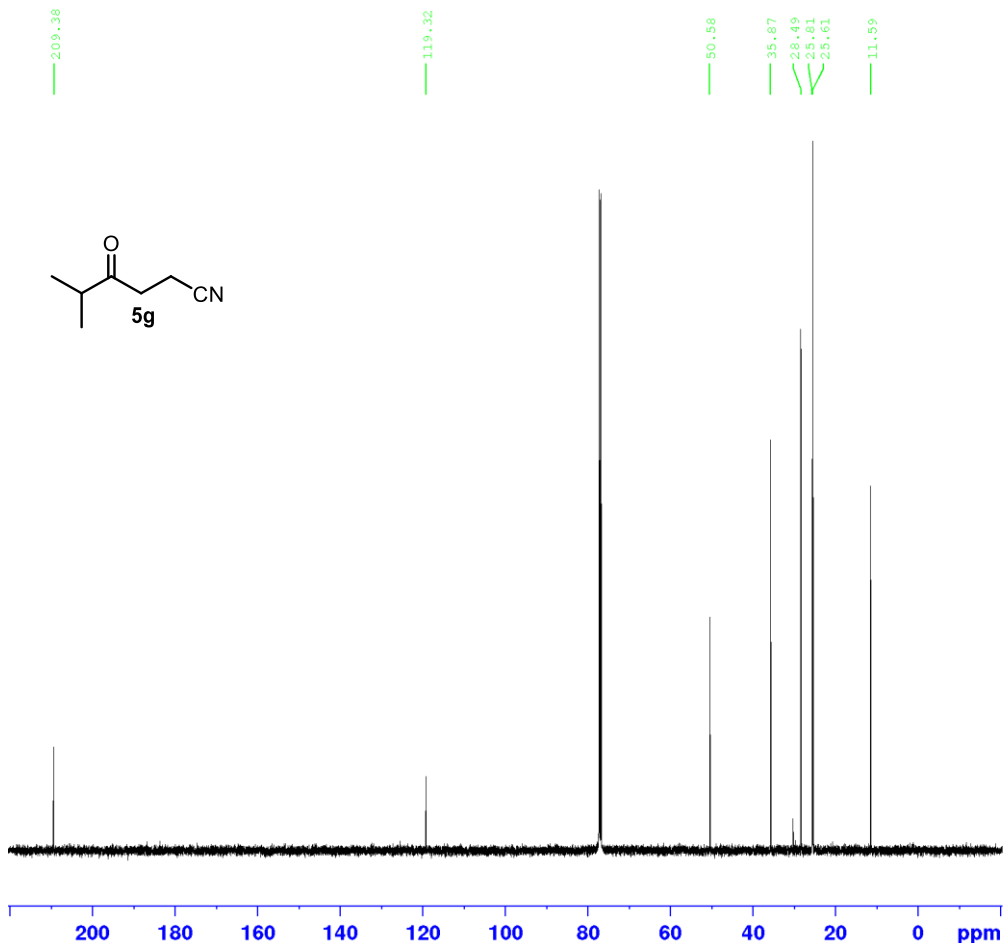
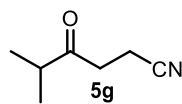
35.87

28.49

25.81

25.61

11.99



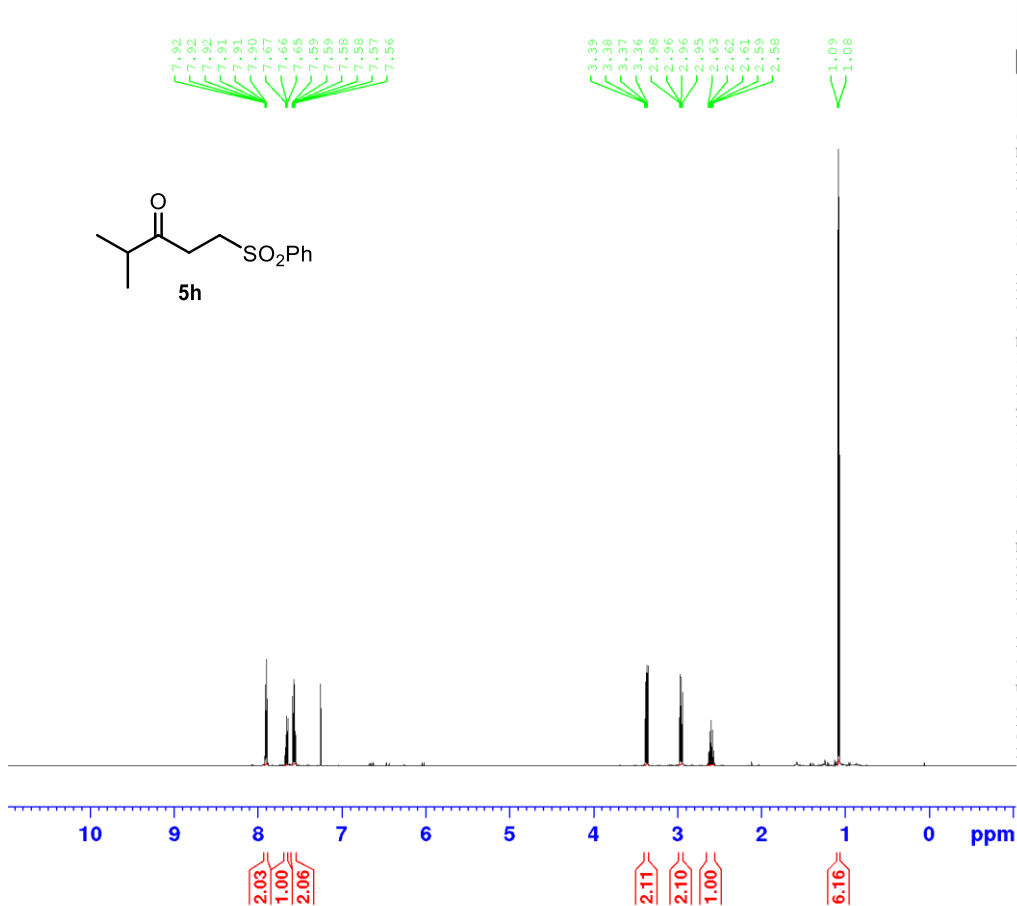
Current Data Parameters
 NAME epb776-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190318
 Time 20.12
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 10300
 DW 16.500 usec
 DE 10.00 usec
 TE 298.5 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

==== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577733 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

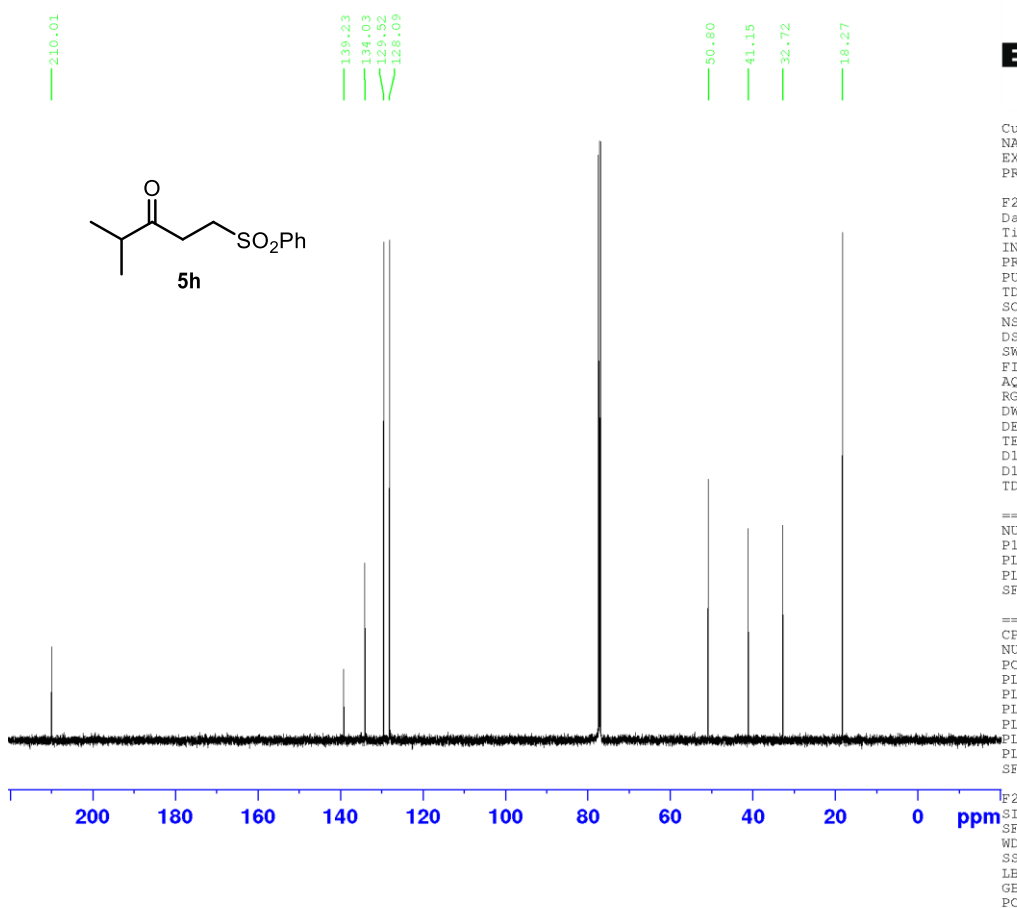


Current Data Parameters
 NAME epb780-f1-1
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190415
 Time_ 19.25
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 228
 DW 83.200 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300134 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



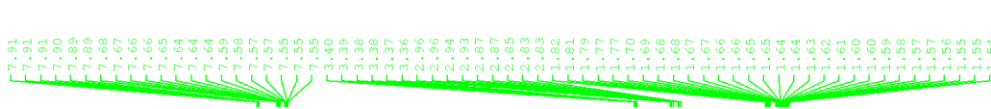
Current Data Parameters
 NAME epb780-f1-1-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190416
 Time_ 1.24
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 11500
 DW 16.500 usec
 DE 10.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

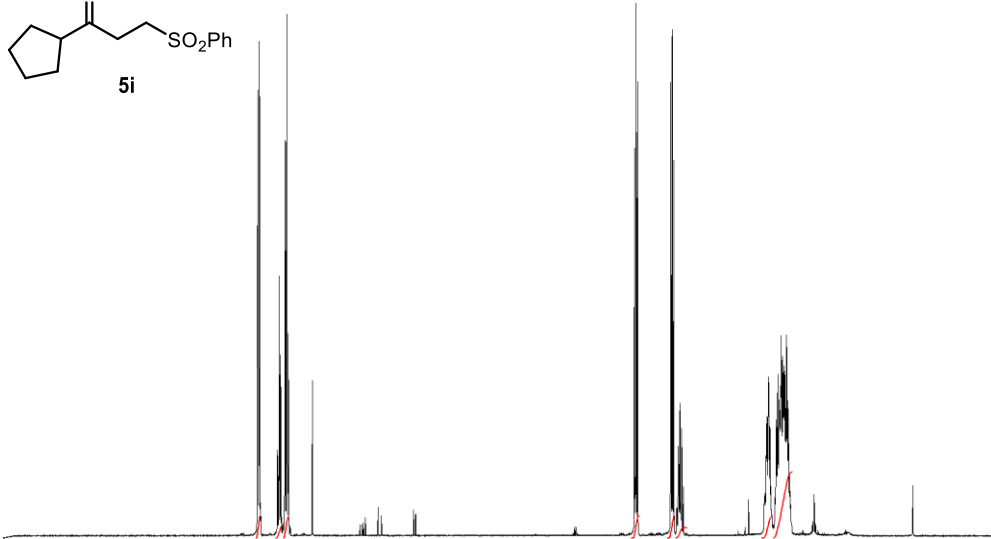
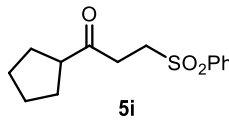
===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577733 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40



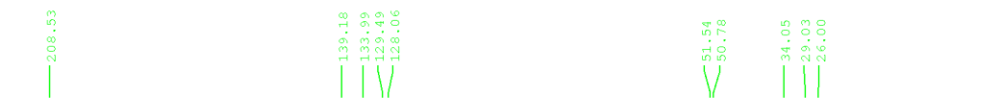
Current Data Parameters
 NAME epb789-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190319
 Time 12.22
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 4
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 128
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1



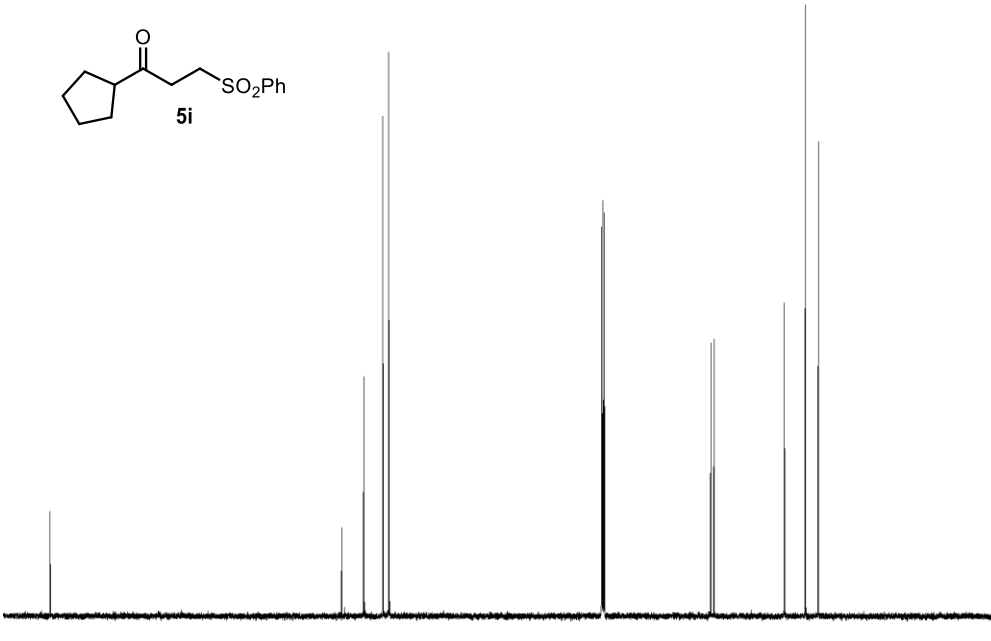
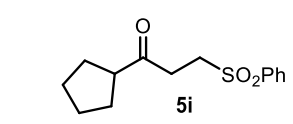
===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PLLW 23.86643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300098 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME epb789-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190319
 Time 16.30
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 4
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 32768
 DW 20.800 usec
 DE 10.00 usec
 TE 297.9 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

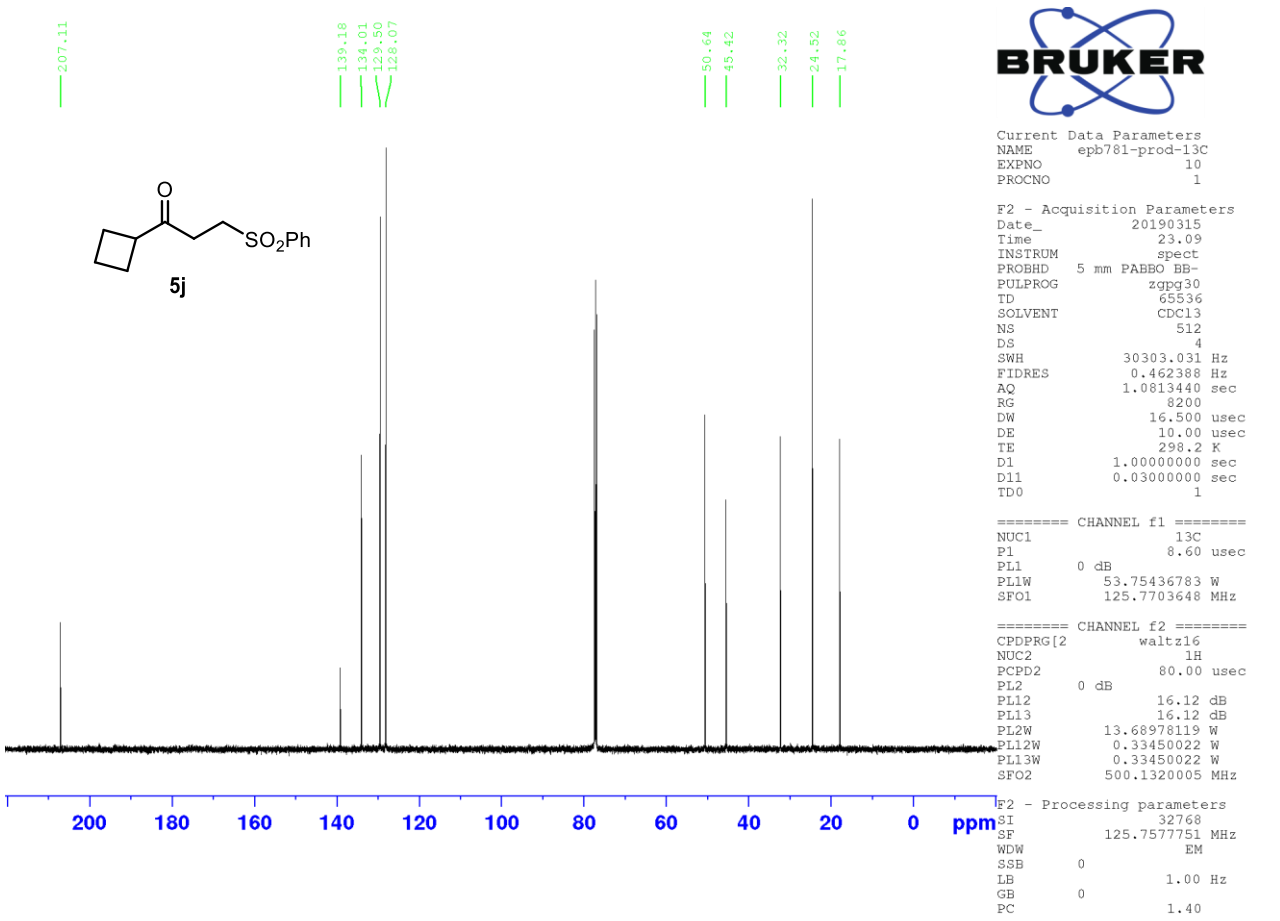
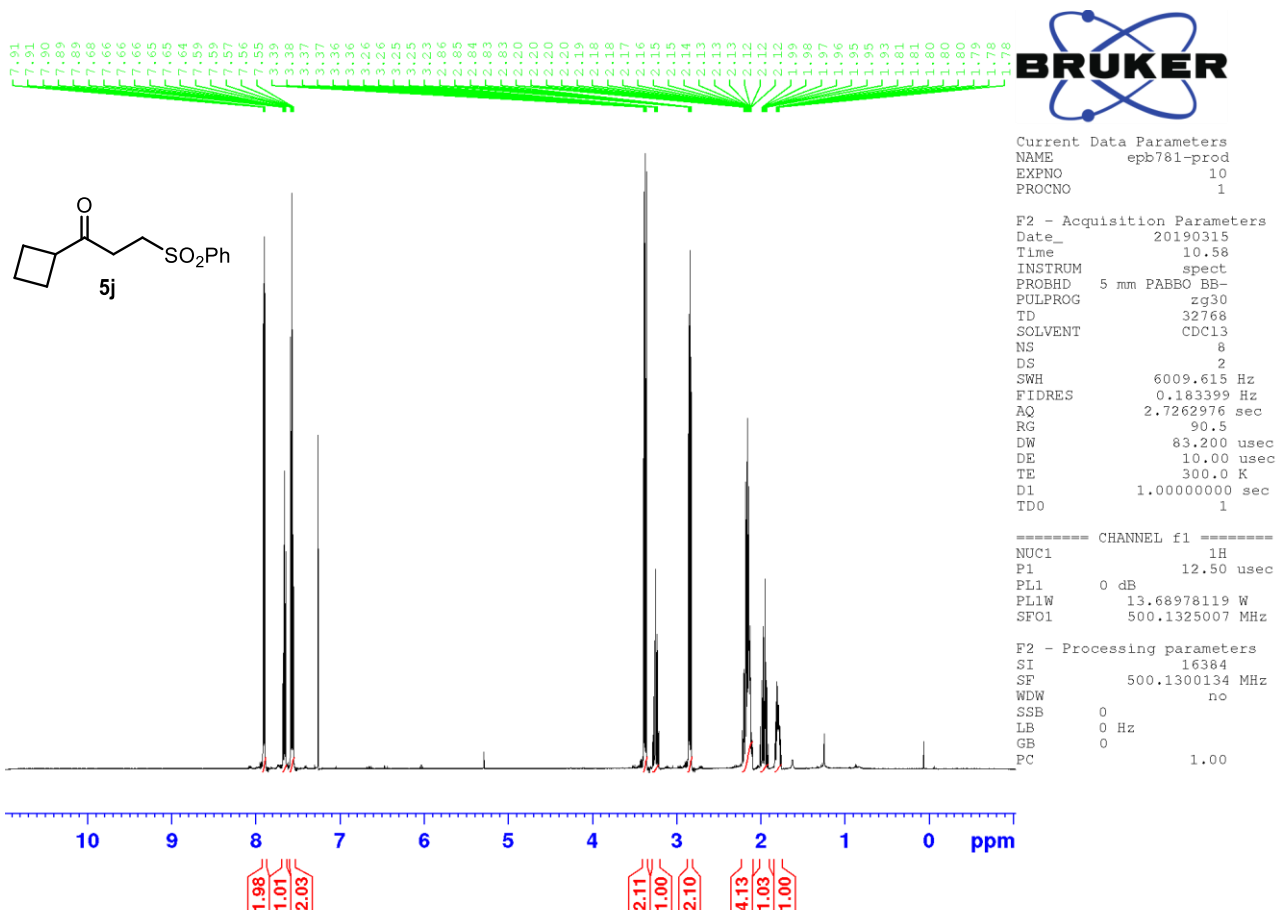


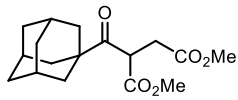
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 -3.00 dB
 PLLW 75.17808533 W
 SFO1 100.6303736 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -3.00 dB
 PL12 13.05 dB
 PL13 13.05 dB
 PL2W 30.07123375 W
 PL12W 0.74670875 W
 PL13W 0.74670875 W
 SFO2 400.1616006 MHz

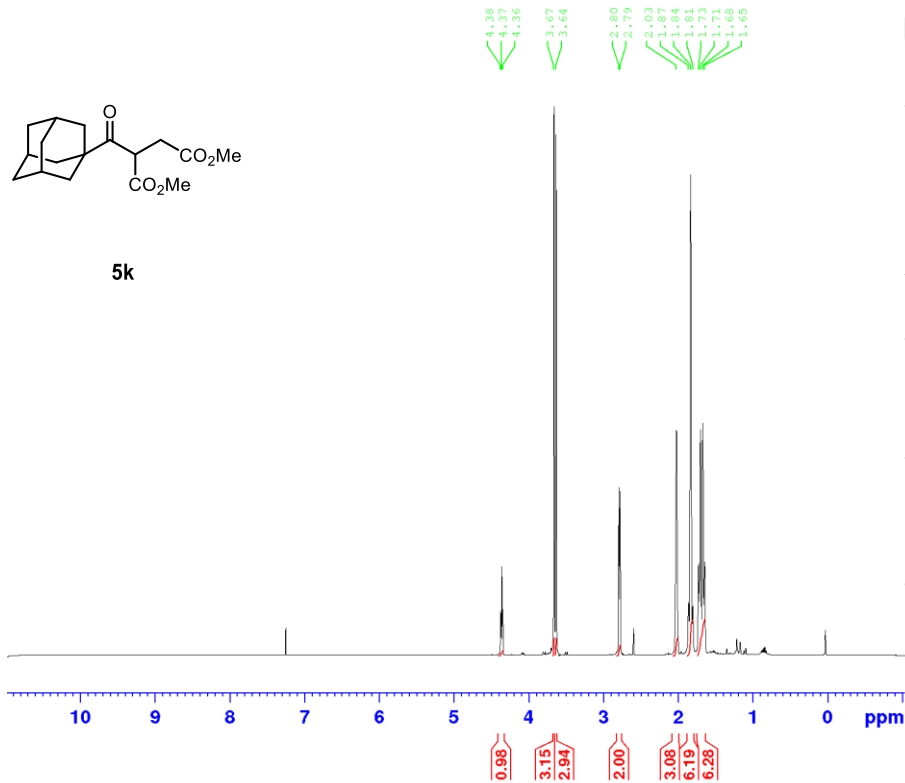


F2 - Processing parameters
 SI 32768
 SF 100.6203019 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40





5k

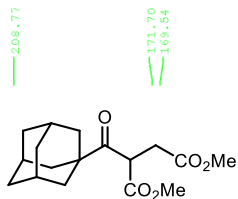


Current Data Parameters
 NAME epb764-prod-1H
 EXPNO 10
 PROCNO 1

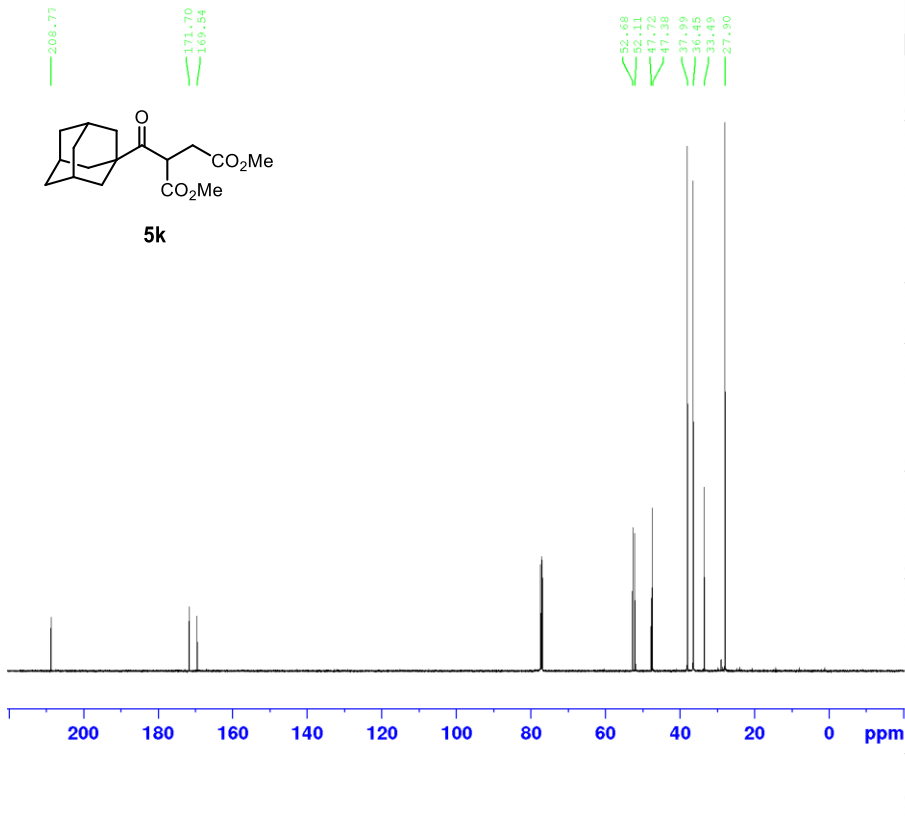
F2 - Acquisition Parameters
 Date_ 20190520
 Time 18.09
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 40.3
 DW 83.200 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.0000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300131 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



5k



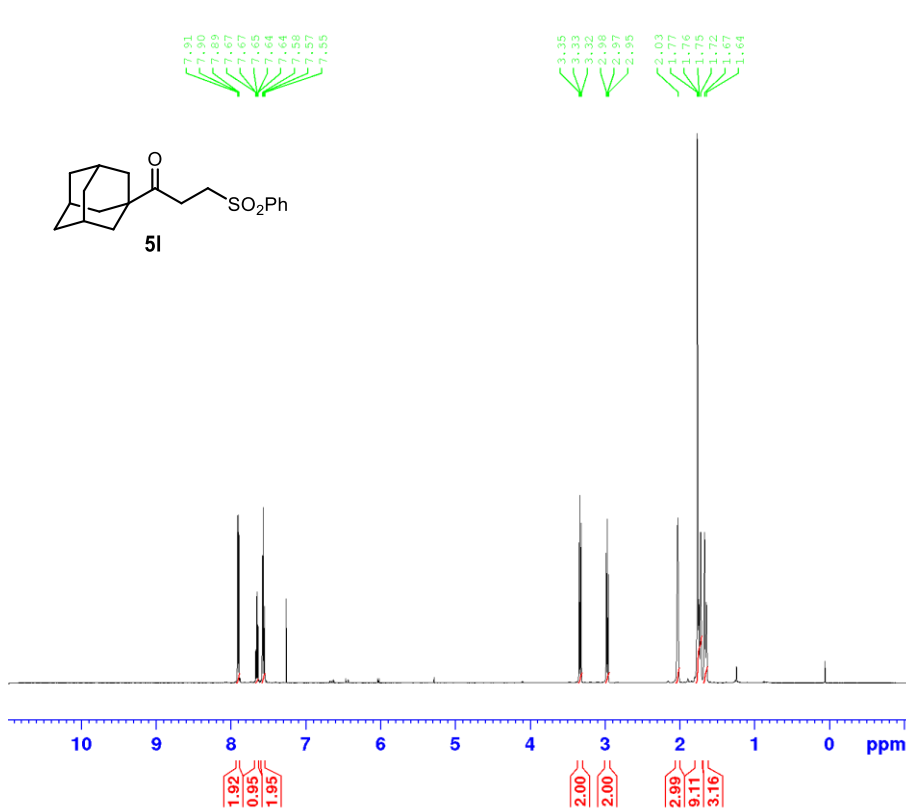
Current Data Parameters
 NAME epb764-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190521
 Time 0.06
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 9200
 DW 16.500 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.0000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577777 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

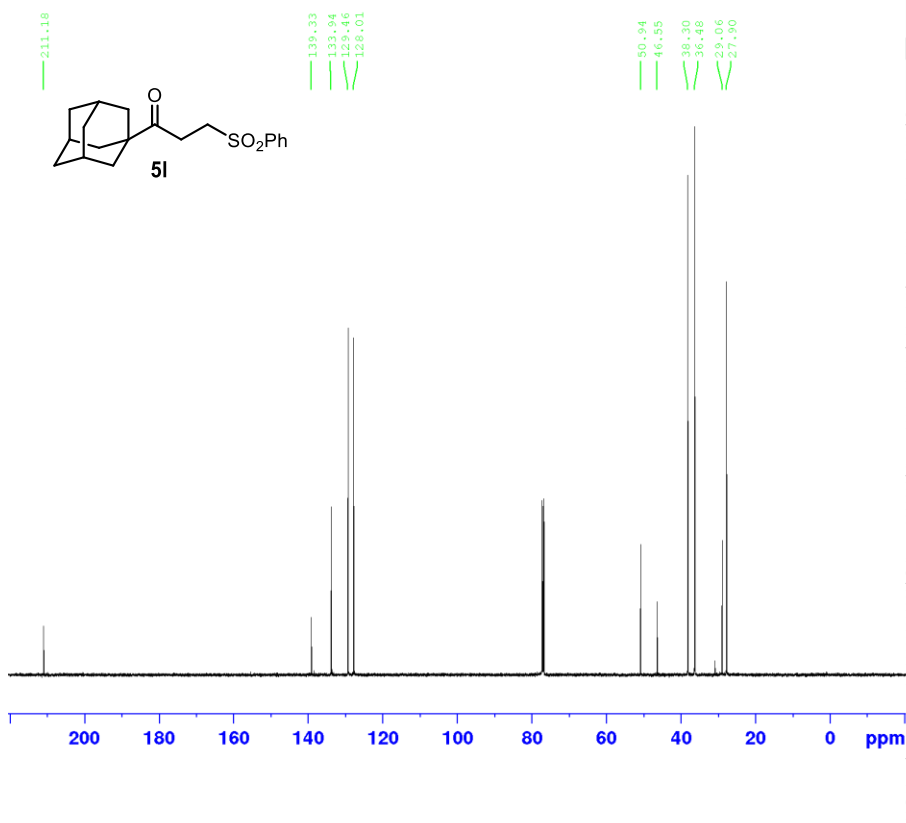


Current Data Parameters
 NAME epb783-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190315
 Time 11.04
 INSTRUM spect
 PROBH 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 57
 DW 83.200 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300134 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



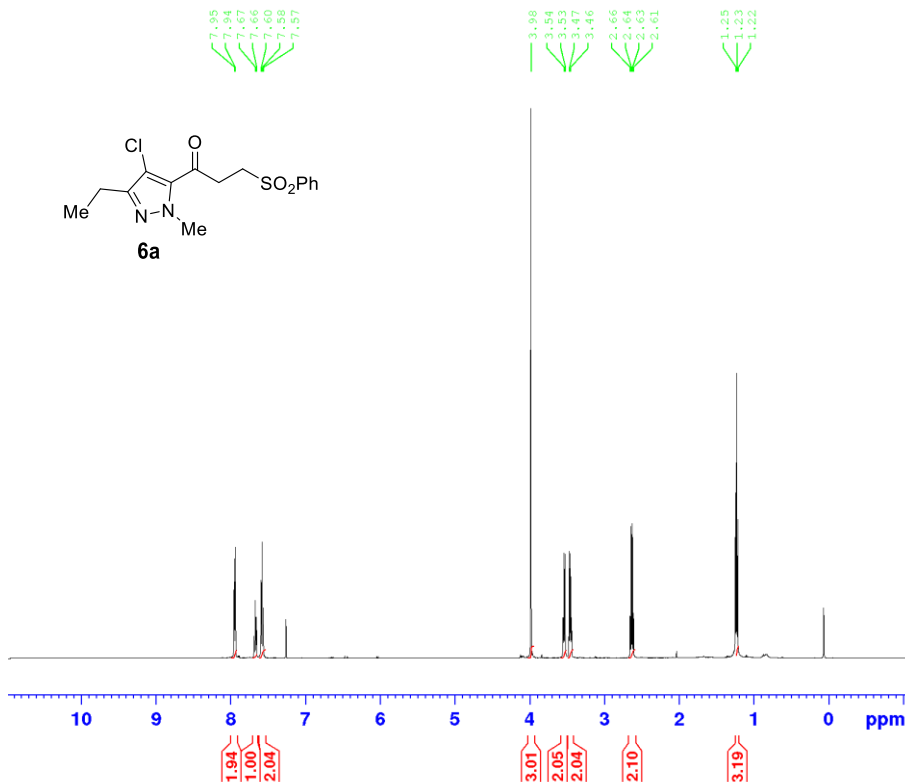
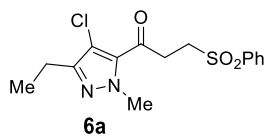
Current Data Parameters
 NAME epb783-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190315
 Time 23.40
 INSTRUM spect
 PROBH 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 14600
 DW 16.500 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CDPFRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PLW 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577777 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

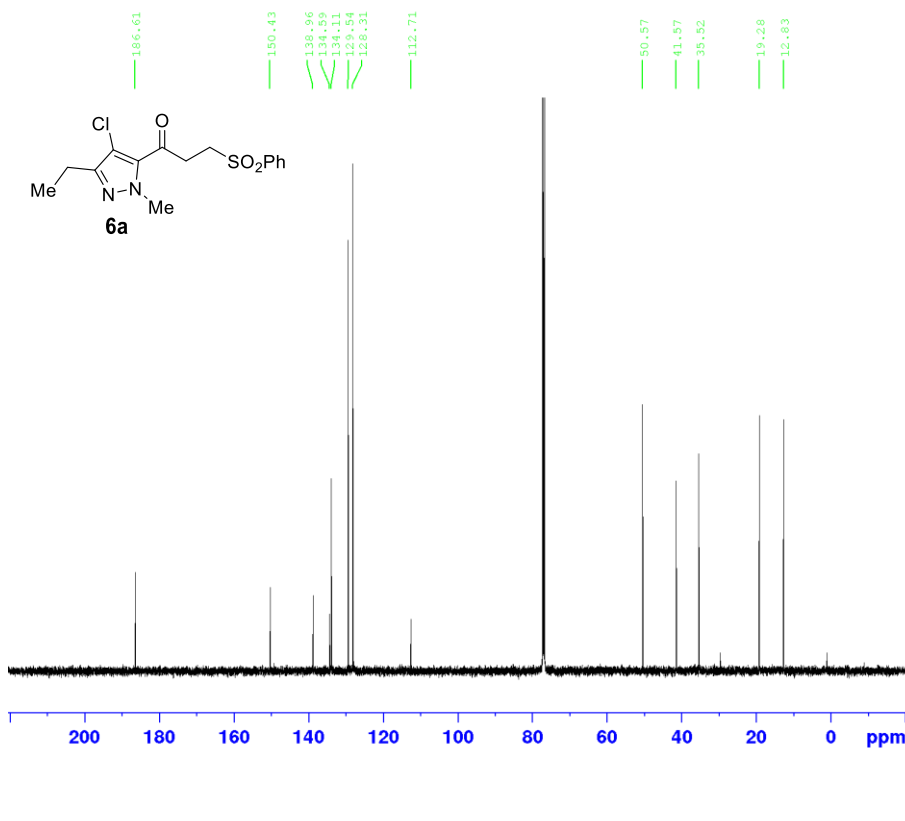
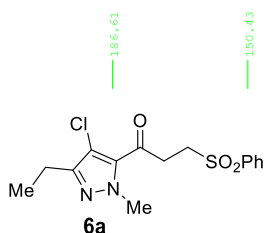


Current Data Parameters
 NAME epb989-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190609
 Time 17.49
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 181
 DW 83.200 usec
 DE 10.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300133 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



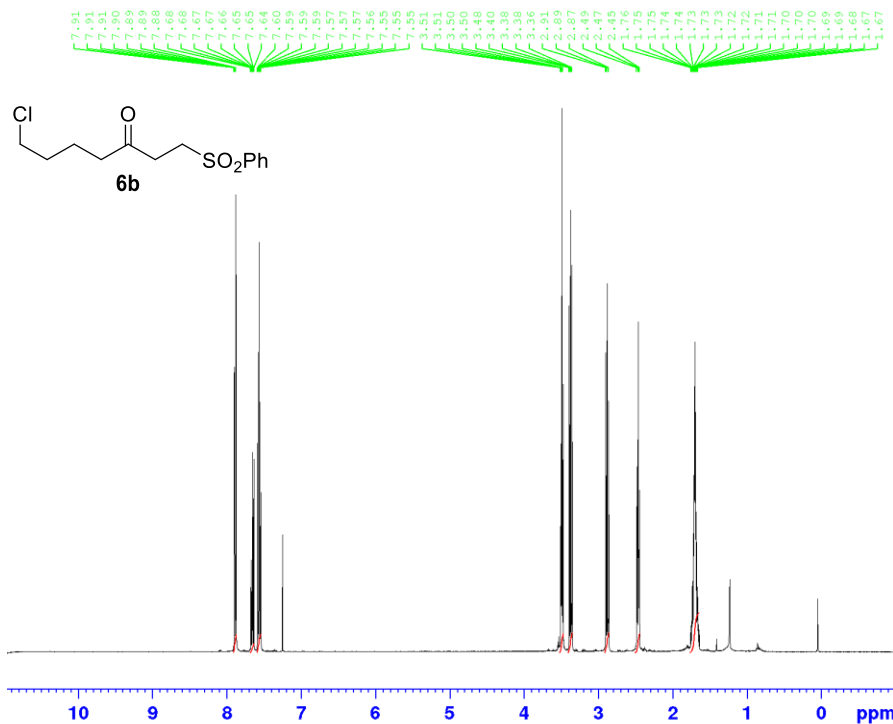
Current Data Parameters
 NAME epb989-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190609
 Time 18.08
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 13000
 DW 16.500 usec
 DE 10.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

----- CHANNEL f1 -----
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

----- CHANNEL f2 -----
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577733 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

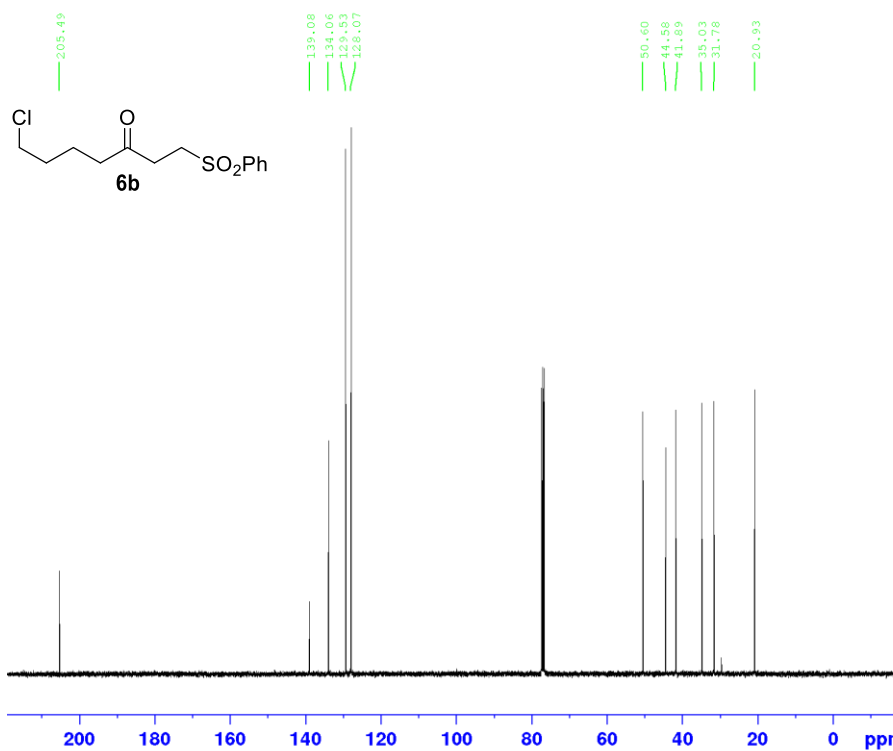


Current Data Parameters
 NAME epb857-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190423
 Time 9.05
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDC13
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 114
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TDO 1

----- CHANNEL f1 -----
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.132007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.130098 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



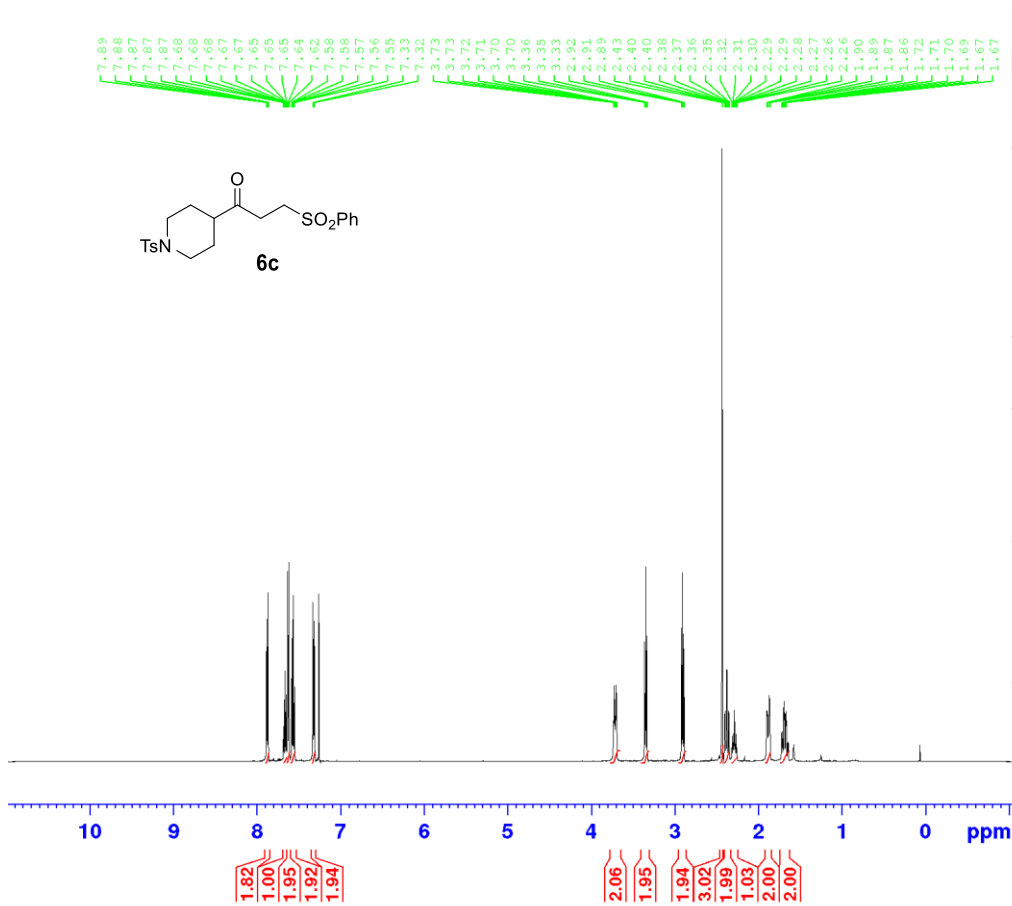
Current Data Parameters
 NAME epb857-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190423
 Time 10.18
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 512
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 26008
 DW 20.850 usec
 DE 10.00 usec
 TE 298.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

----- CHANNEL f1 -----
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

----- CHANNEL f2 -----
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127591 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

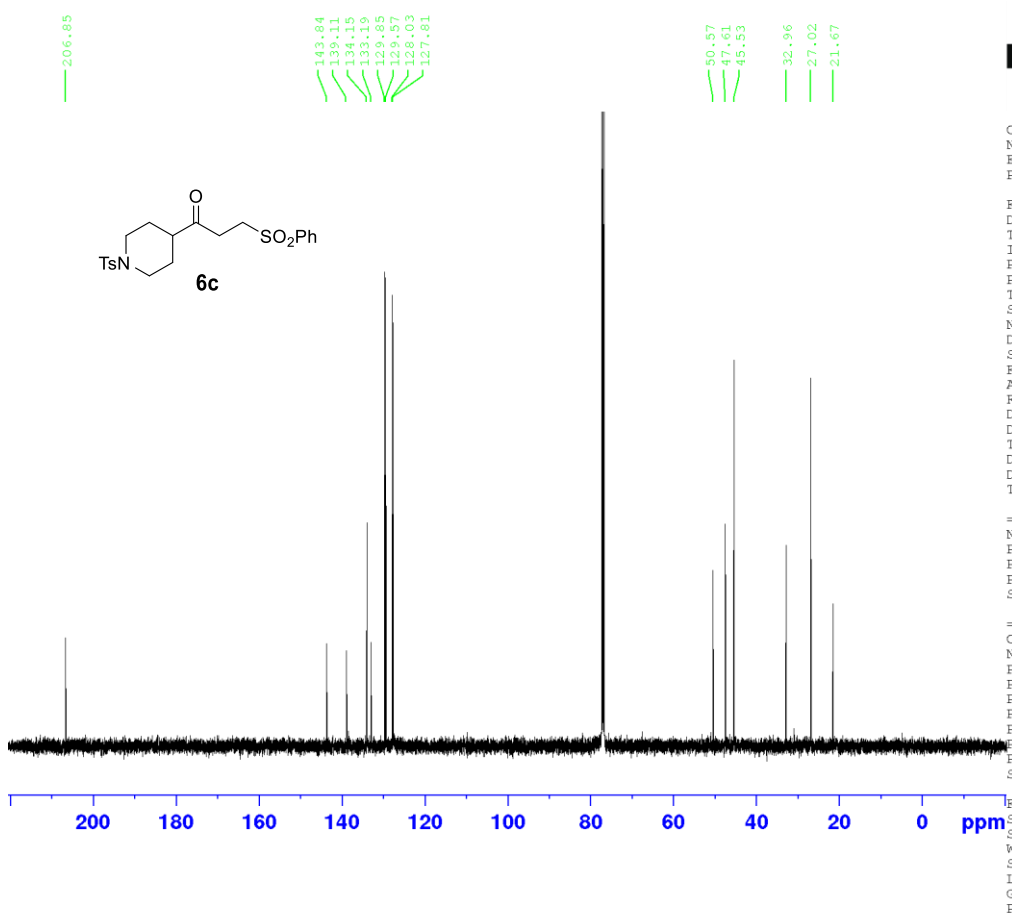


Current Data Parameters
 NAME epb823-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190315
 Time 18.50
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 228
 DW 83.200 usec
 DE 10.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 12.50 usec
 PL1 0 dB
 PL1W 13.68978119 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300134 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



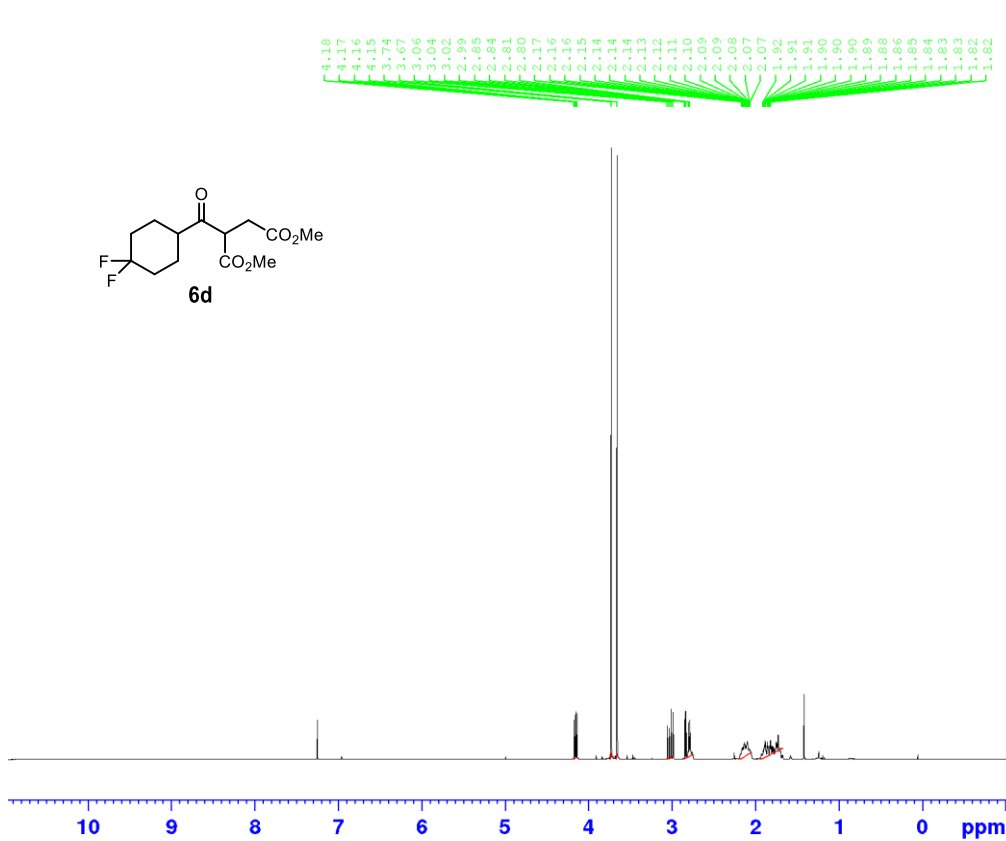
Current Data Parameters
 NAME epb823-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190316
 Time 16.42
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 13000
 DW 16.500 usec
 DE 10.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 8.60 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 0 dB
 PL12 16.12 dB
 PL13 16.12 dB
 PL2W 13.68978119 W
 PL12W 0.33450022 W
 PL13W 0.33450022 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577733 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

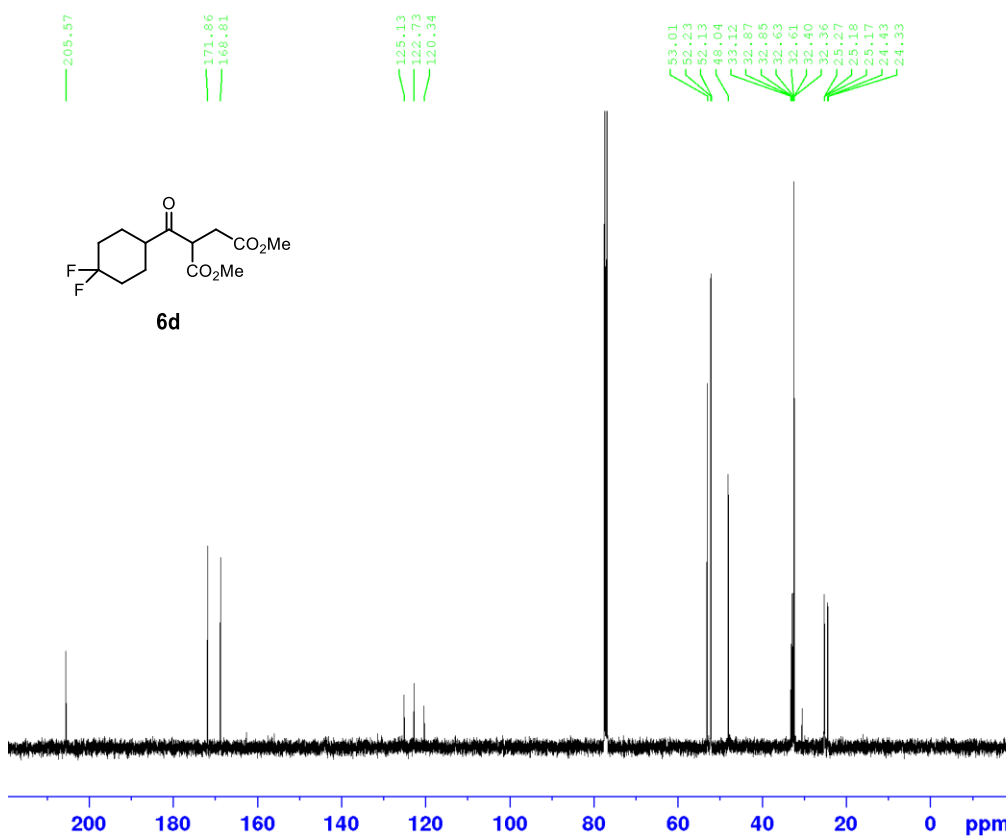


Current Data Parameters
 NAME epb784-f1
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190307
 Time 17.56
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 128
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.86643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300096 MHz
 WDW no
 SSB 0
 LB 0 Hz
 GB 0
 PC 1.00



Current Data Parameters
 NAME epb784-f1-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190307
 Time 21.59
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 512
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 26008
 DW 20.850 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.86643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127551 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

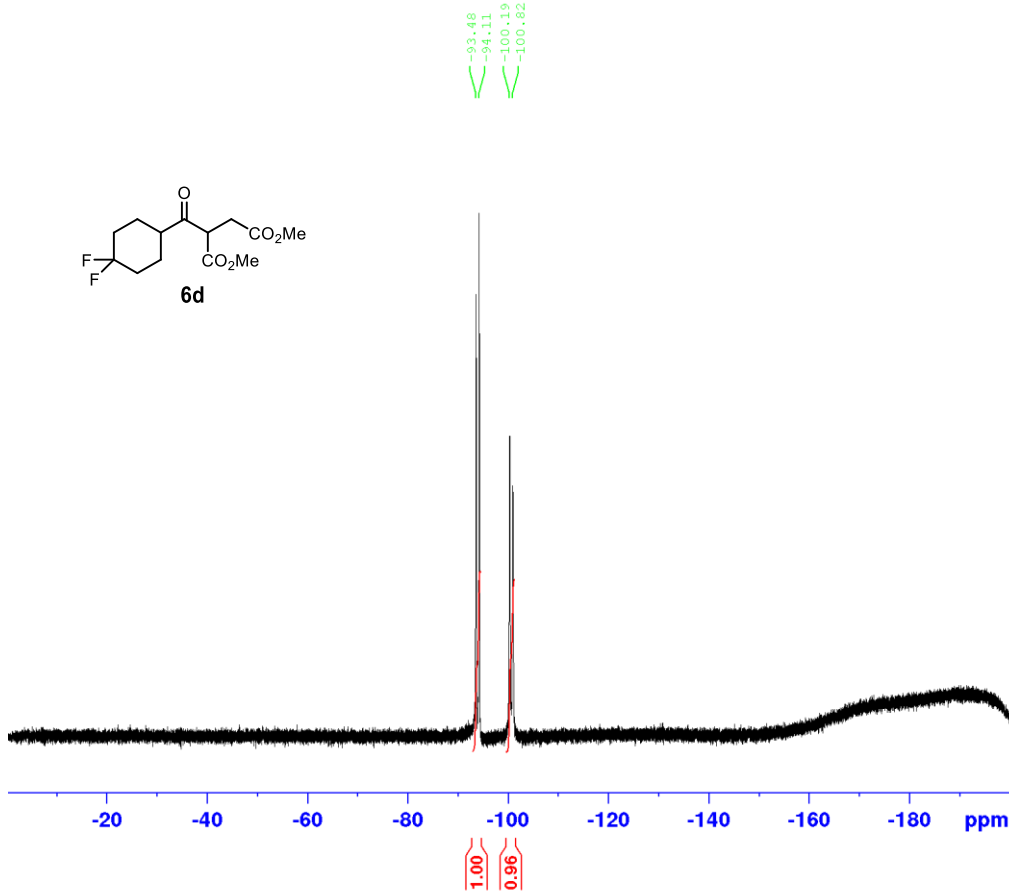
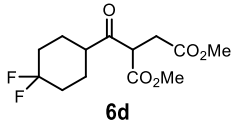


Current Data Parameters
NAME epb784-fl-19F
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190307
Time 17.59
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 131072
SOLVENT CDCl3
NS 8
DS 2
SWH 75187.969 Hz
FIDRES 0.573639 Hz
AQ 0.8716288 sec
RG 2048
DW 6.650 usec
DE 7.14 usec
TE 297.9 K
D1 5.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 19F
P1 12.00 usec
PL1 -3.00 dB
SFO1 376.4607040 MHz

F2 - Processing parameters
SI 65536
SF 376.4984036 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00

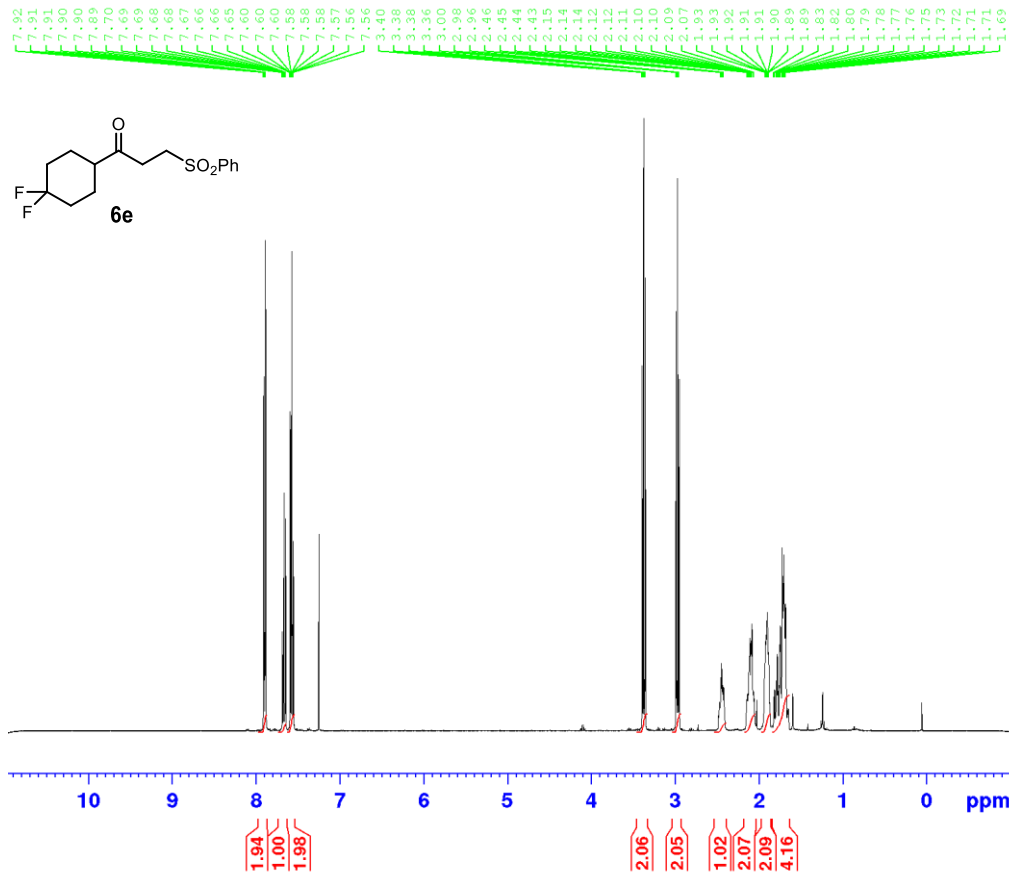
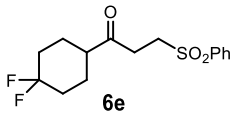


Current Data Parameters
NAME epb795-fl
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190307
Time 18.04
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4209793 sec
RG 128
DW 104.400 usec
DE 6.00 usec
TE 298.1 K
D1 1.00000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.50 usec
PL1 -2.00 dB
PL1W 23.88643074 W
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300098 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 1.00



207.37

139.17
134.12
129.57
128.04
124.93
122.53
120.13

50.61
48.07
33.18
32.79
32.47
24.73
24.63



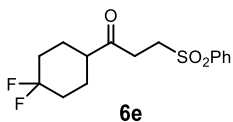
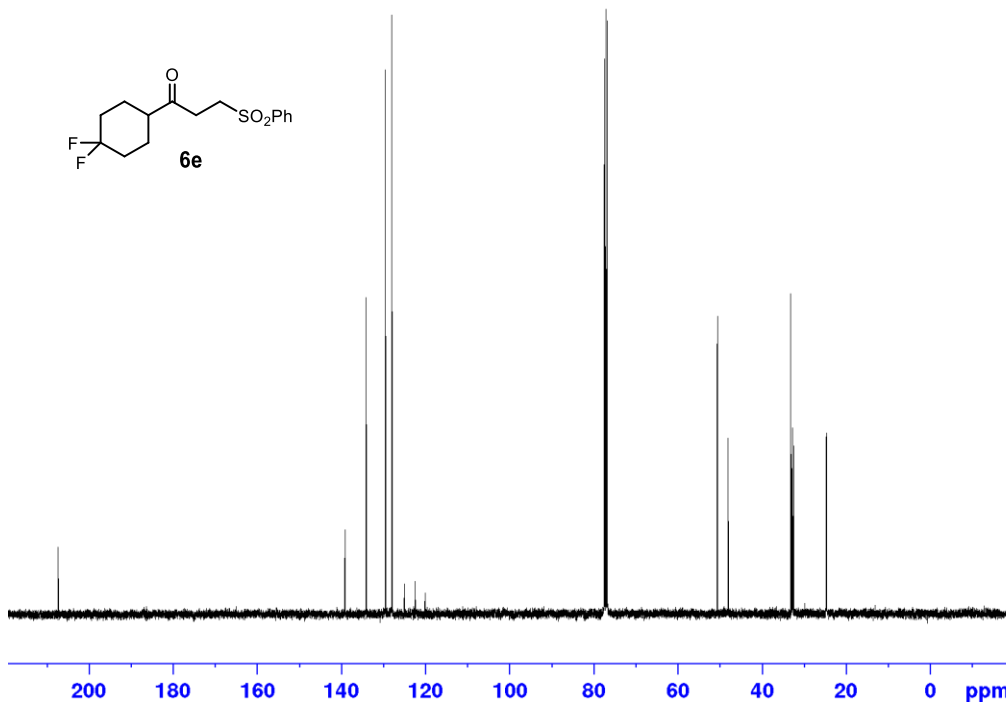
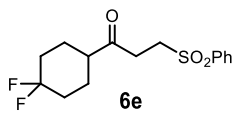
Current Data Parameters
NAME epb795-fl-13C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190307
Time 22.30
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 512
DS 4
SWH 23980.814 Hz
FIDRES 0.365918 Hz
AQ 1.3664256 sec
RG 32768
DW 20.850 usec
DE 10.00 usec
TE 298.0 K
D1 1.00000000 sec
D11 0.03000000 sec
TD0 1

==== CHANNEL f1 =====
NUC1 13C
P1 10.00 usec
PL1 -3.00 dB
PL1W 75.17808533 W
SFO1 100.6228298 MHz

==== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -2.00 dB
PL12 12.83 dB
PL13 12.83 dB
PL2W 23.88643074 W
PL12W 0.78550917 W
PL13W 0.78550917 W
SFO2 400.1316005 MHz

F2 - Processing parameters
SI 32768
SF 100.6127570 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40



-93.41
-94.03
-100.50
-101.14

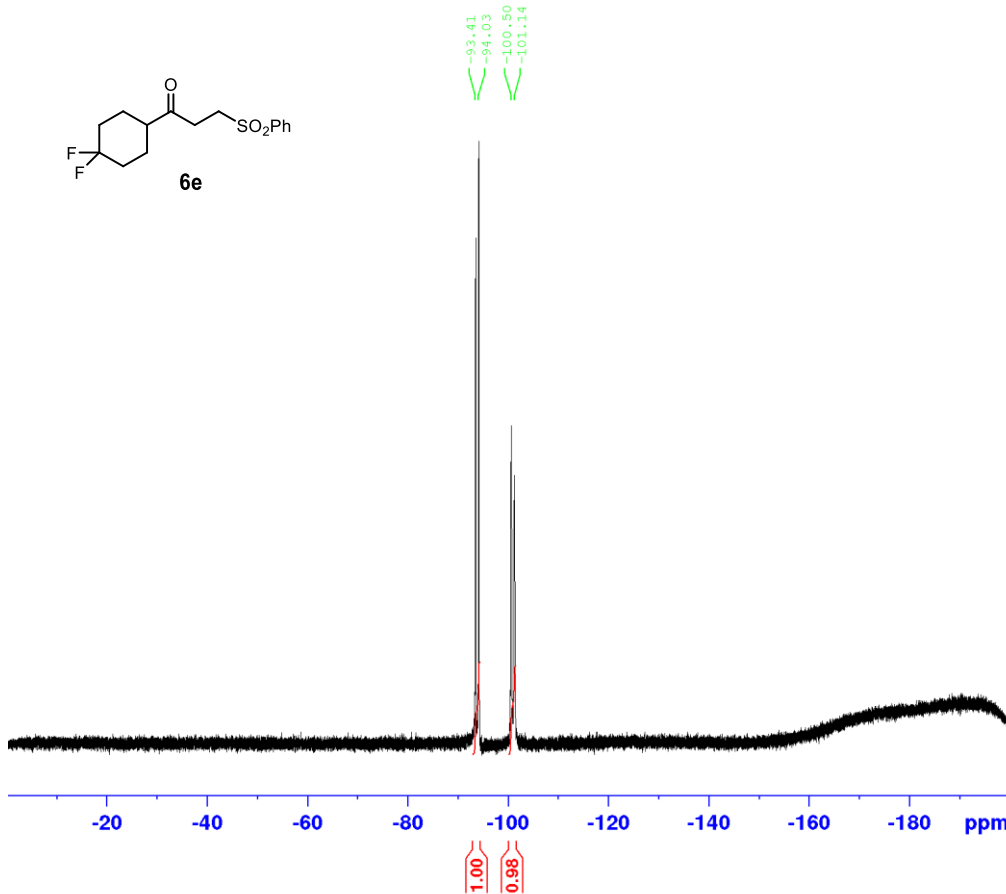


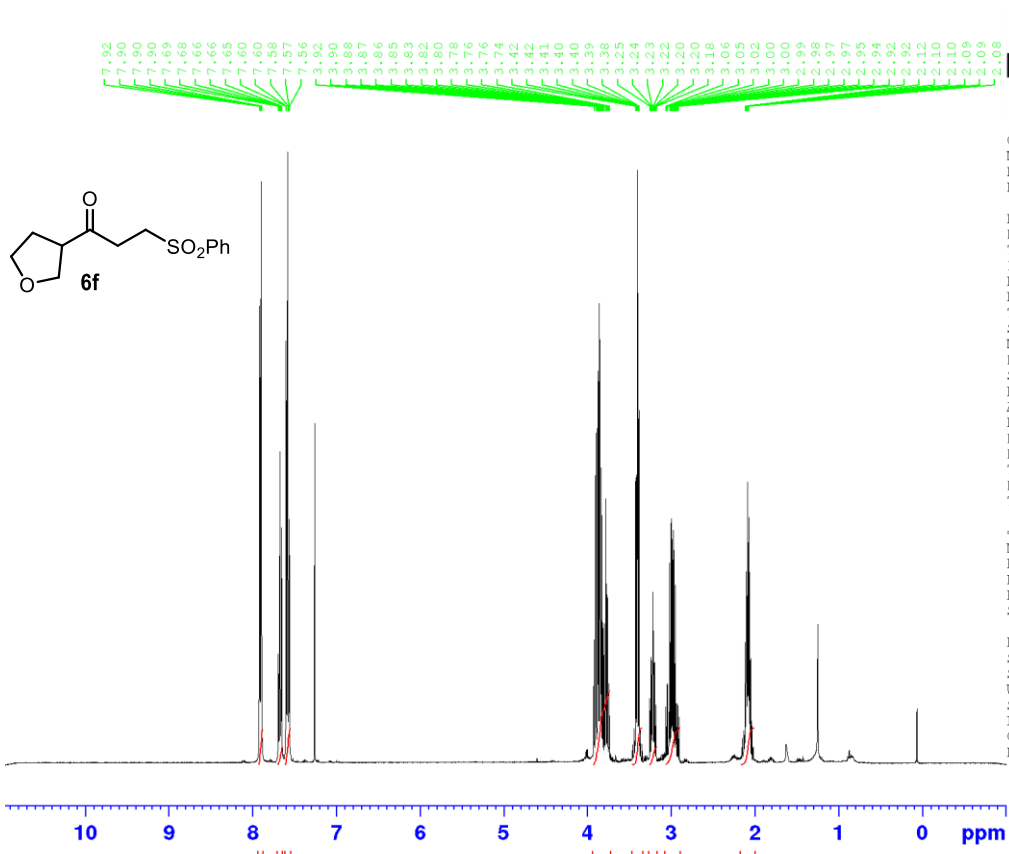
Current Data Parameters
NAME epb795-fl-19F
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190307
Time 18.05
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 131072
SOLVENT CDCl3
NS 8
DS 2
SWH 75187.969 Hz
FIDRES 0.573639 Hz
AQ 0.8716288 sec
RG 2048
DW 6.650 usec
DE 7.14 usec
TE 297.9 K
D1 5.00000000 sec
TD0 1

==== CHANNEL f1 =====
NUC1 19F
P1 12.00 usec
PL1 -3.00 dB
SFO1 376.4607040 MHz

F2 - Processing parameters
SI 65536
SF 376.4984036 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00



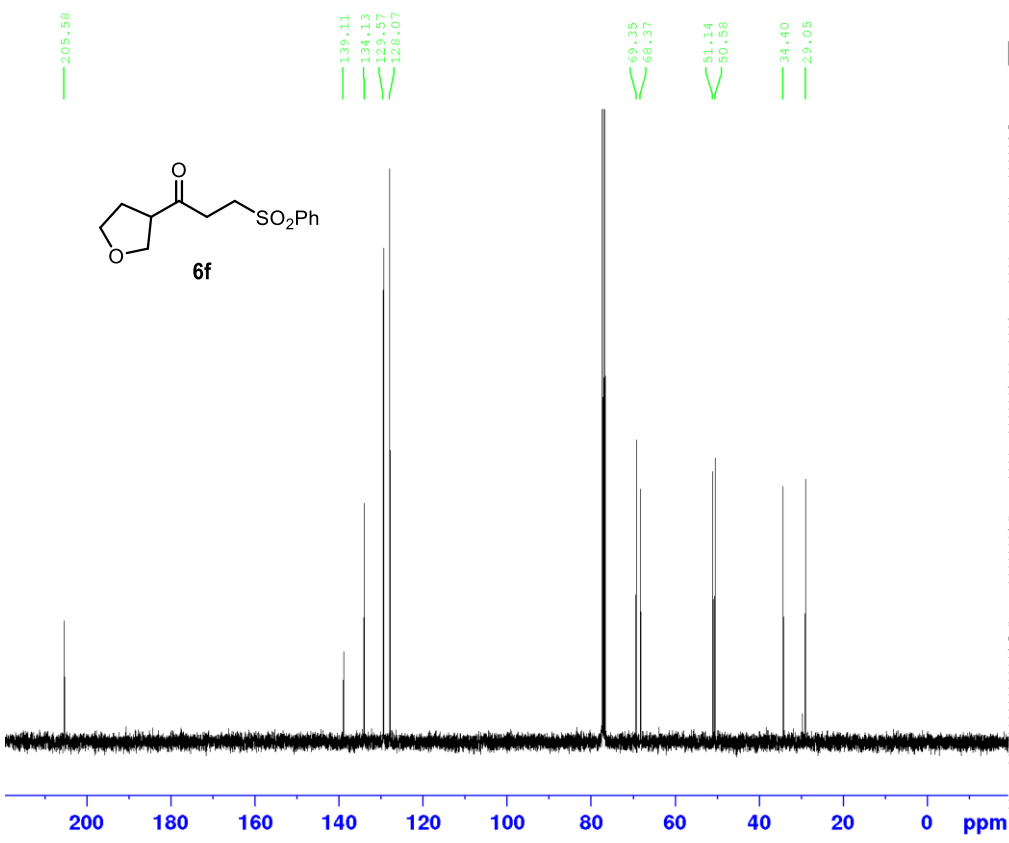


Current Data Parameters
 NAME epb807-prod
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190417
 Time 9.32
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDC13
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 181
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300096 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



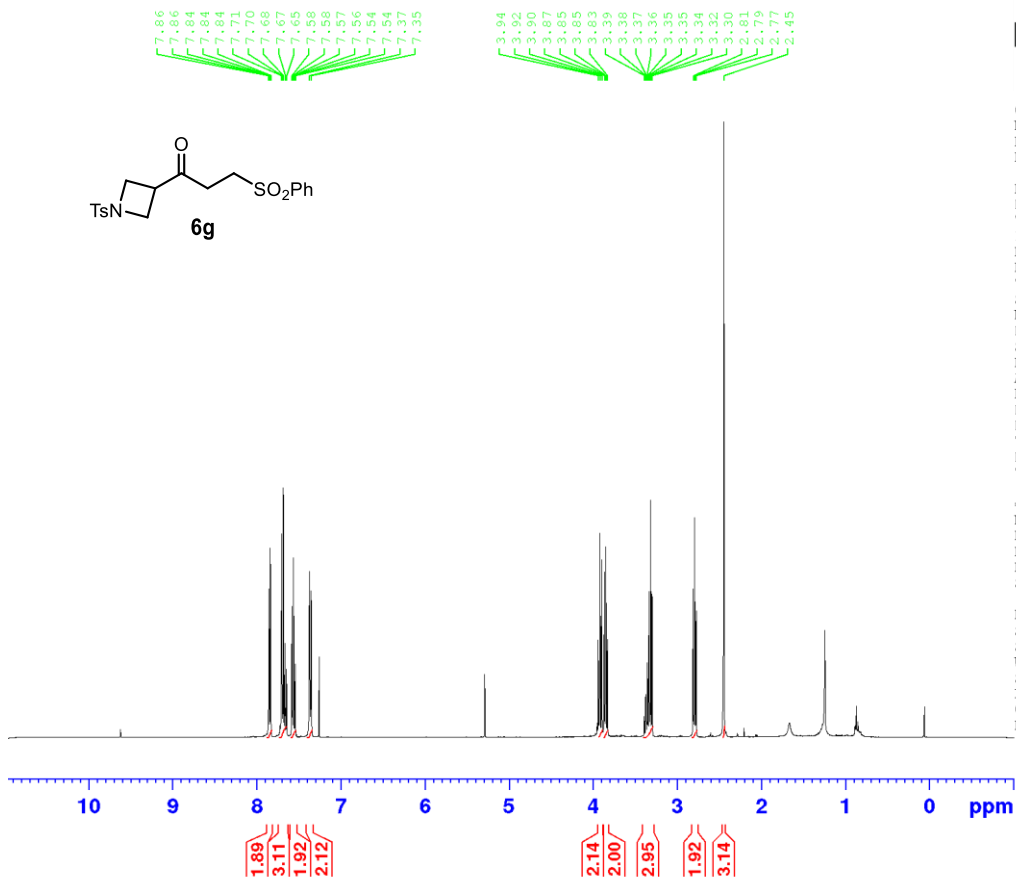
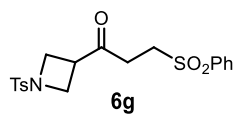
Current Data Parameters
 NAME epb807-prod-13C
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190417
 Time 16.38
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 4
 DS 188
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 32768
 DW 20.800 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

==== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6303736 MHz

==== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -3.00 dB
 PL12 13.05 dB
 PL13 13.05 dB
 PL2W 30.07123375 W
 PL12W 0.74670875 W
 PL13W 0.74670875 W
 SFO2 400.1616006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6203004 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

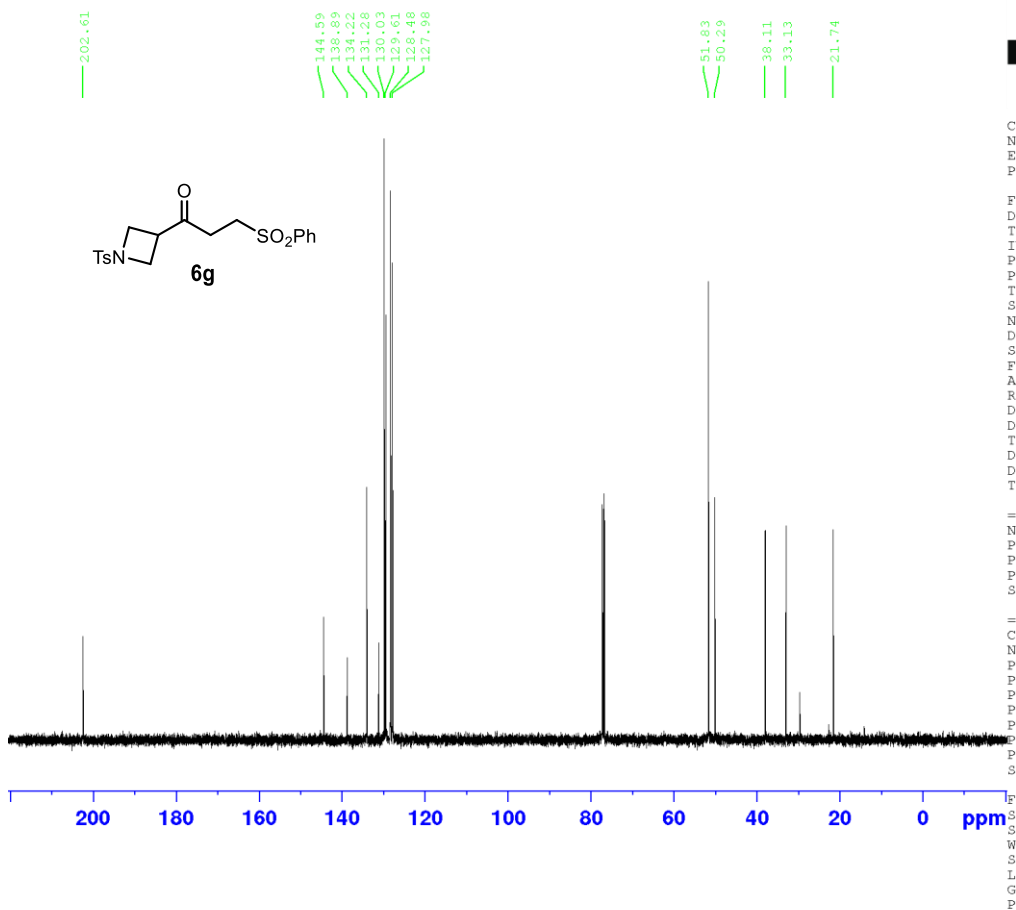
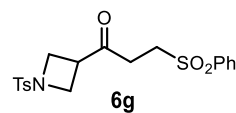


Current Data Parameters
NAME epb933-prod
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190425
Time 9.44
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zg30
TD 32768
SOLVENT CDC13
NS 8
DS 2
SWH 4789.272 Hz
FIDRES 0.146157 Hz
AQ 3.4209793 sec
RG 128
DW 104.400 usec
DE 6.00 usec
TE 298.1 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.50 usec
PL1 -2.00 dB
PL1W 23.88643074 W
SFO1 400.1320007 MHz

F2 - Processing parameters
SI 16384
SF 400.1300097 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



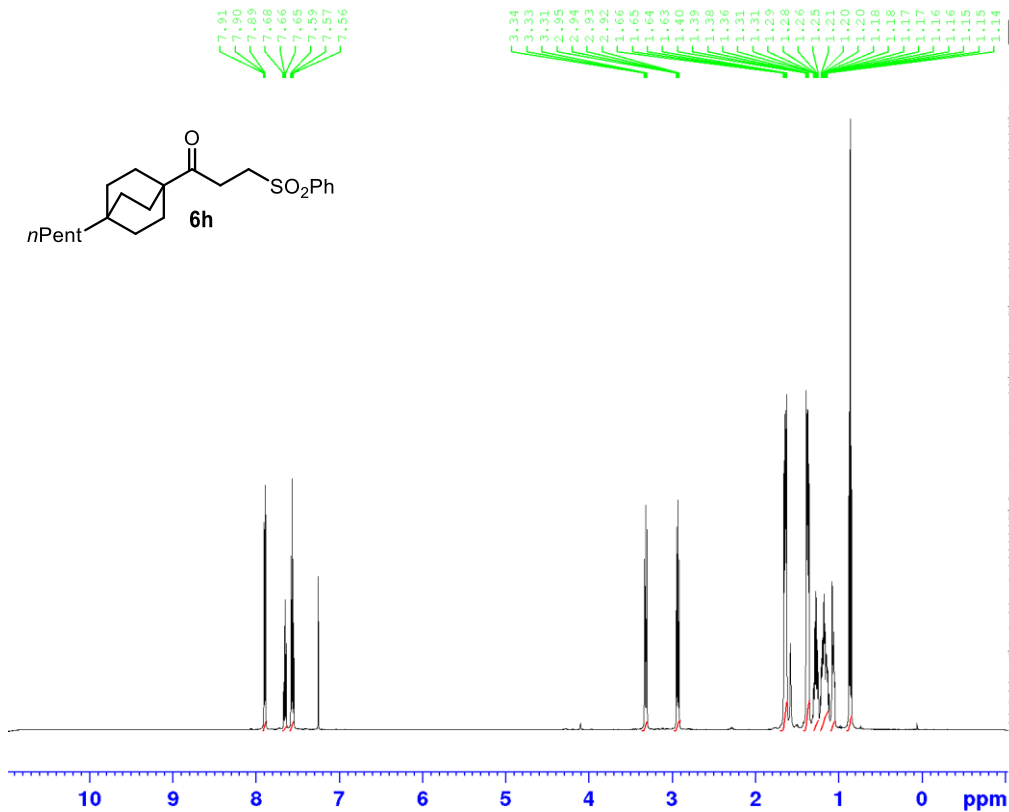
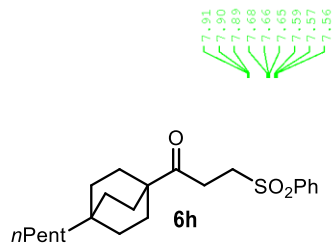
Current Data Parameters
NAME epb933-prod-13C
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20190425
Time 15.41
INSTRUM spect
PROBHD 5 mm PABBO BB-
PULPROG zgpg30
TD 65536
SOLVENT CDC13
NS 168
DS 4
SWH 30303.031 Hz
FIDRES 0.462388 Hz
AQ 1.0813440 sec
RG 10300
DW 16.500 usec
DE 10.00 usec
TE 298.0 K
D1 1.0000000 sec
D11 0.0300000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 13C
P1 16.20 usec
PL1 -0.50 dB
PL1W 60.31339645 W
SFO1 125.7703648 MHz

===== CHANNEL f2 =====
CPDPRG[2] waltz16
NUC2 1H
PCPD2 80.00 usec
PL2 -1.00 dB
PL12 20.80 dB
PL13 20.80 dB
PL2W 17.23441315 W
PL12W 0.11386666 W
PL13W 0.11386666 W
SFO2 500.1320005 MHz

F2 - Processing parameters
SI 32768
SF 125.7577771 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

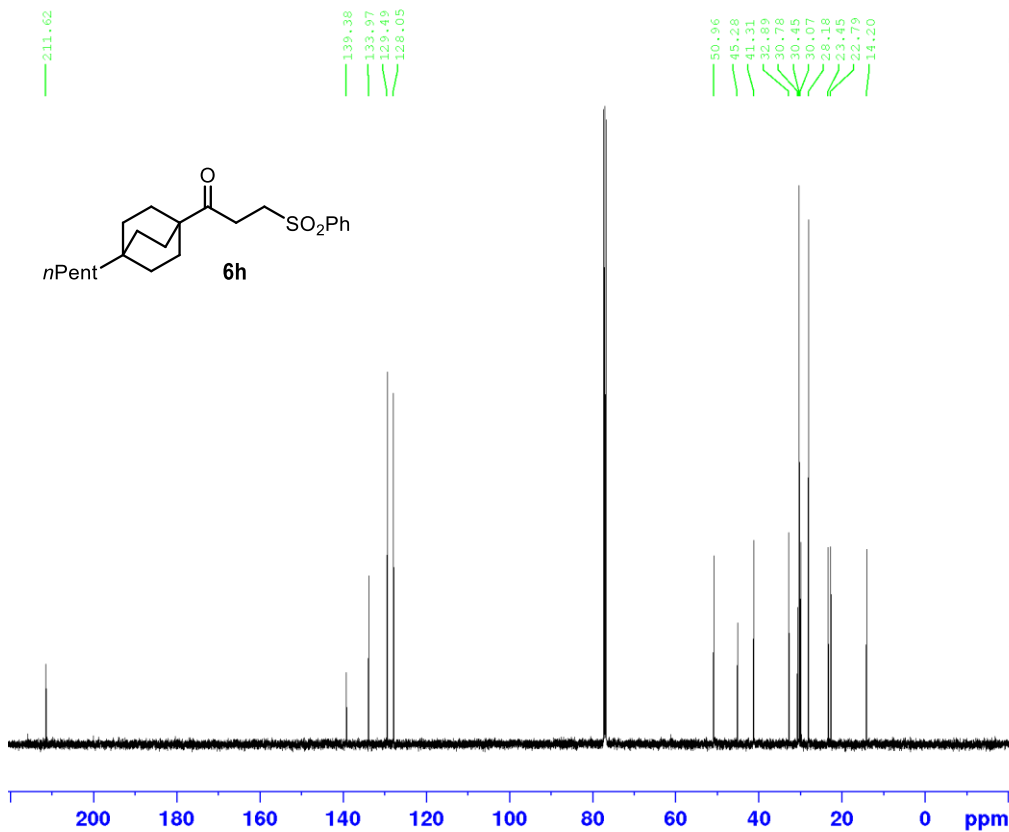
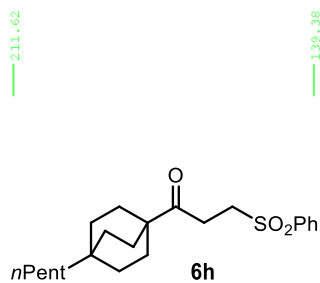


Current Data Parameters
 NAME epb856-prod1H
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190805
 Time 19.49
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDC13
 NS 32
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 228
 DW 83.200 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.00 usec
 PL1 -4.08 dB
 PL1W 35.02648163 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300137 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



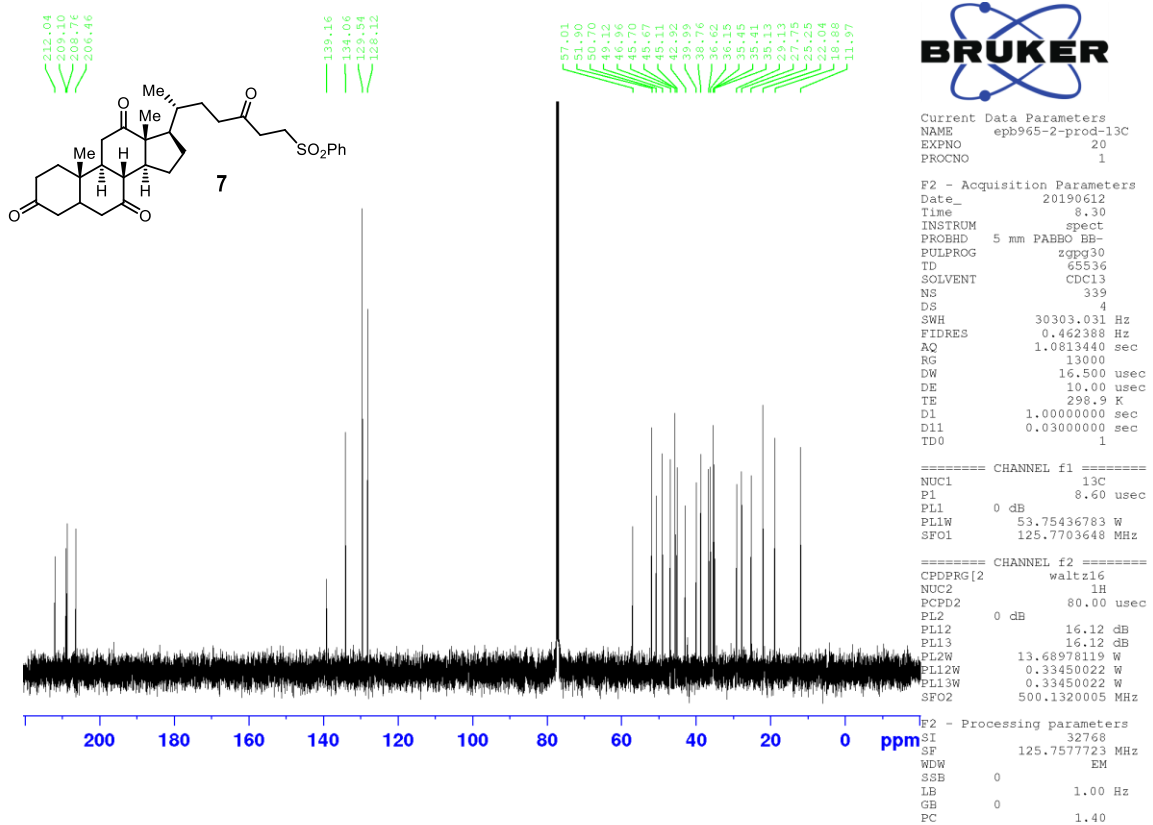
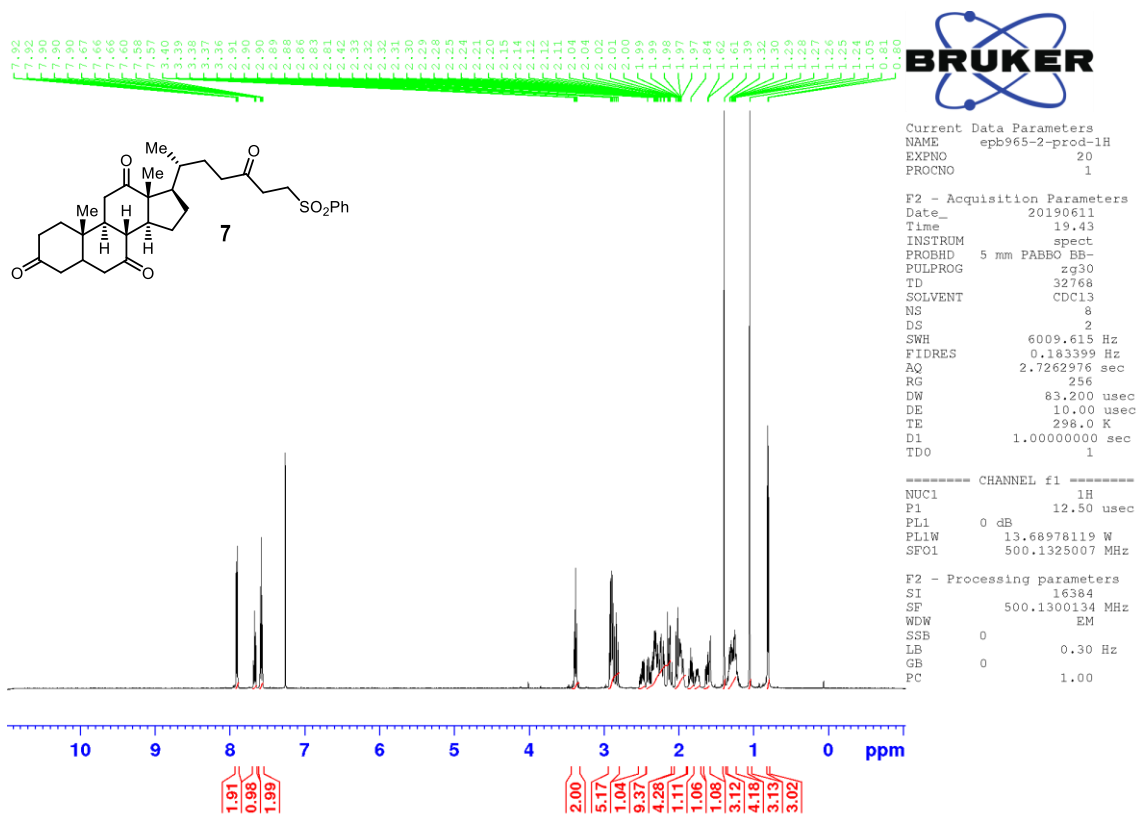
Current Data Parameters
 NAME epb856-prod-13C
 EXPNO 10
 PROCNO 1

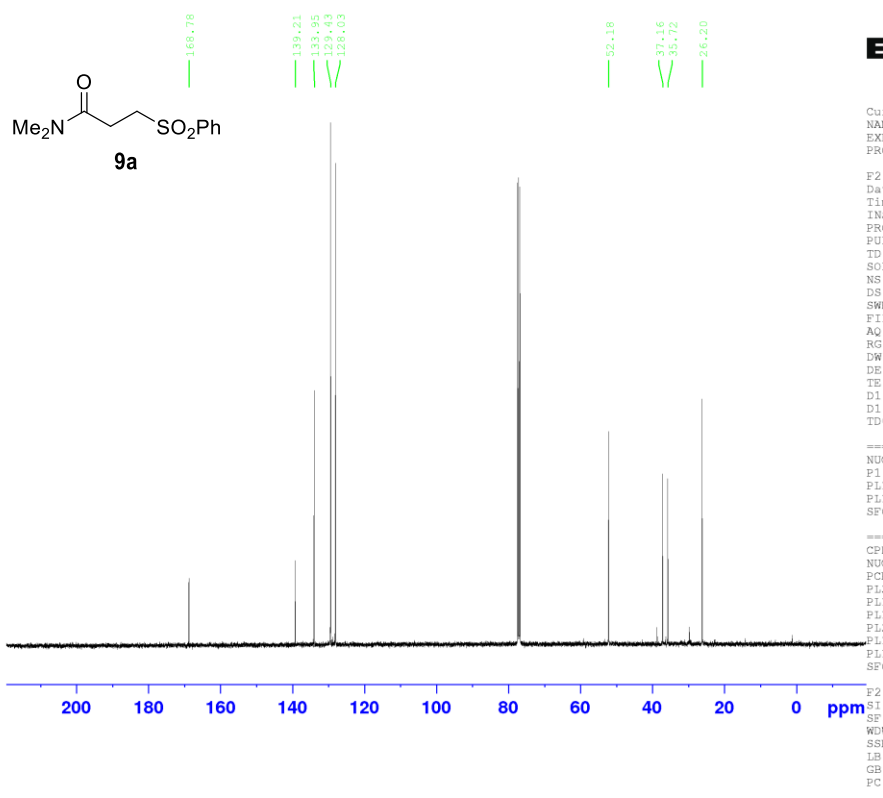
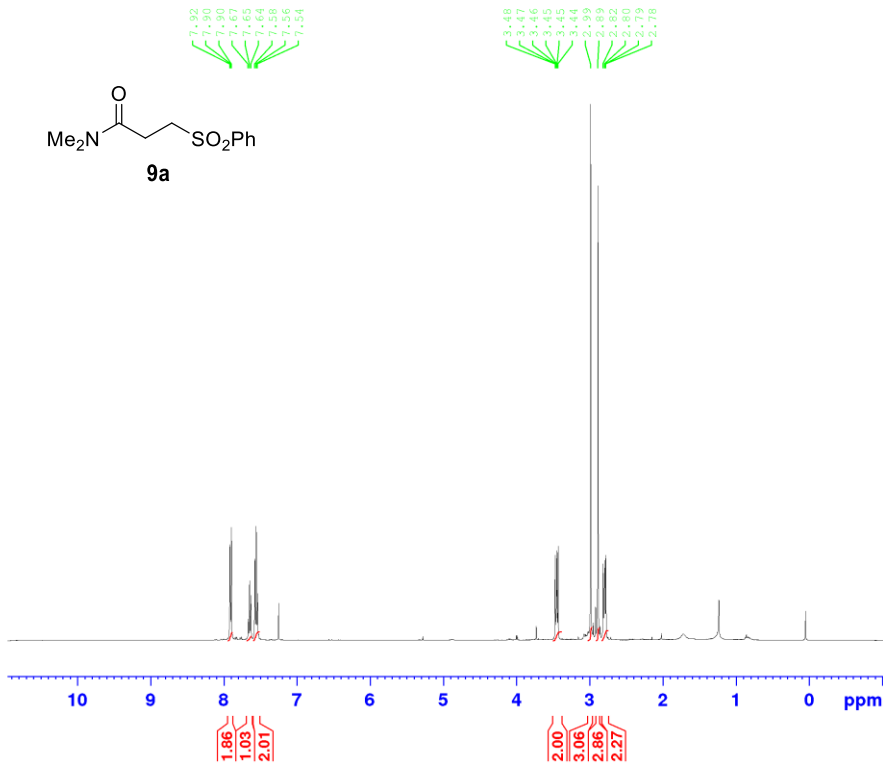
F2 - Acquisition Parameters
 Date_ 20190806
 Time 5.33
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDC13
 NS 512
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 8200
 DW 16.500 usec
 DE 10.00 usec
 TE 299.2 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

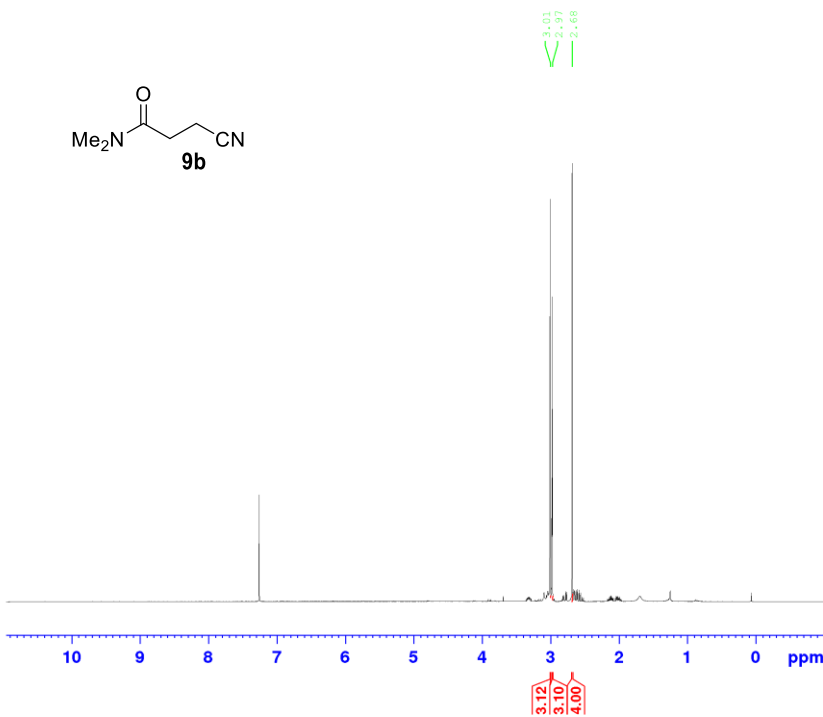
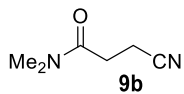
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG[2] waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.08 dB
 PL12 11.54 dB
 PL13 11.54 dB
 PL2W 35.02648163 W
 PL12W 0.96027696 W
 PL13W 0.96027696 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577723 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40





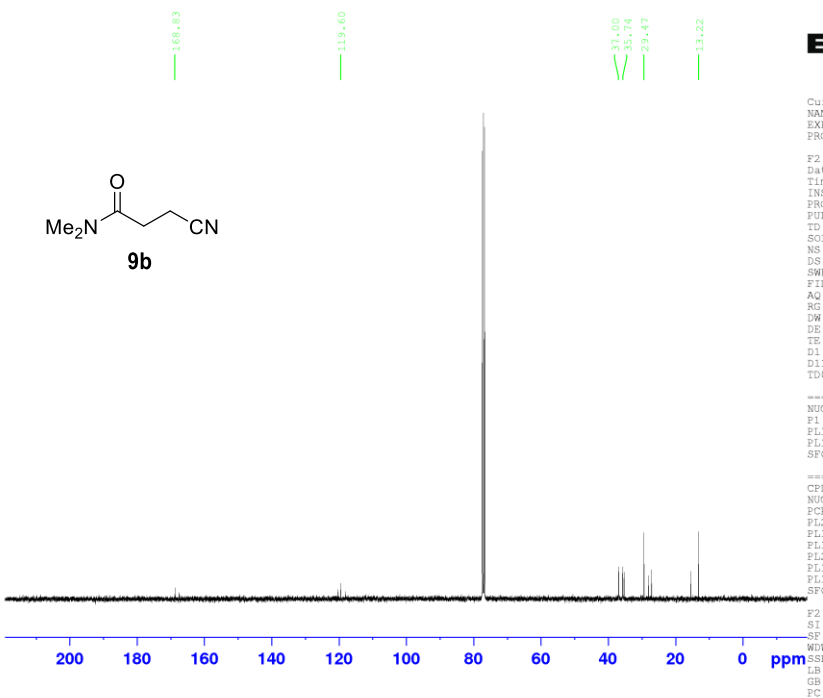
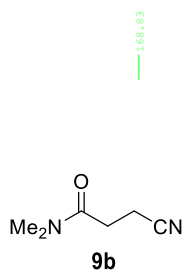


Current Data Parameters
 NAME DMZ-1-119-3-Fr23-27_10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200226
 Time 18.28
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 322.5
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300098 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



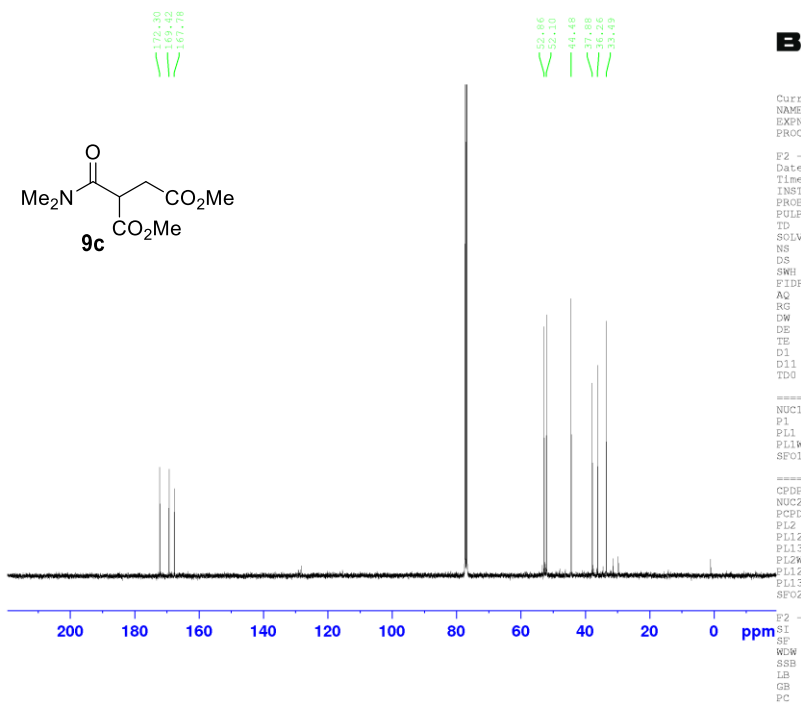
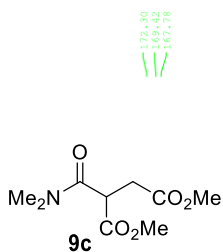
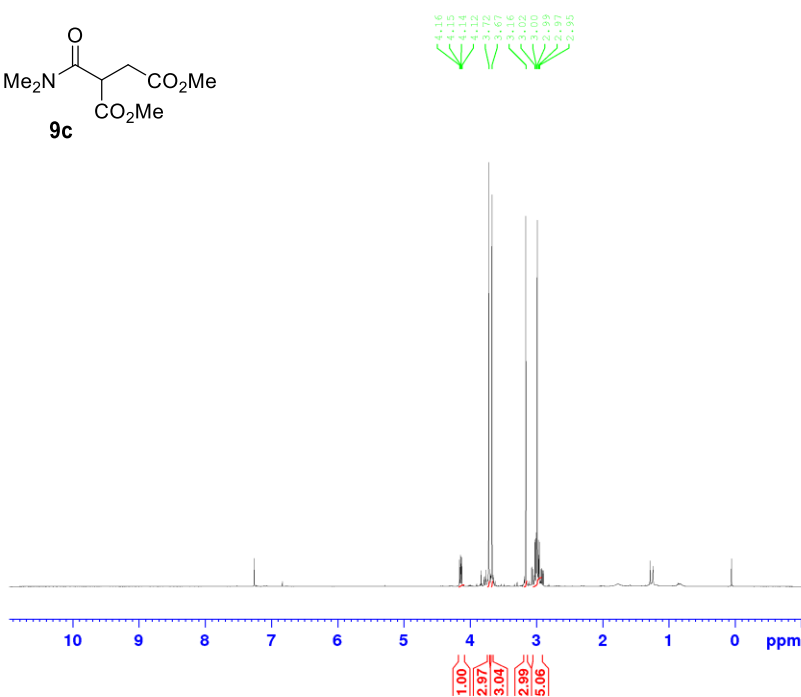
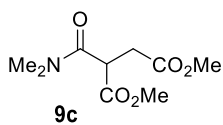
Current Data Parameters
 NAME DMZ-1-119-3-Fr23-27_20
 EXPNO 1
 PROCNO 1

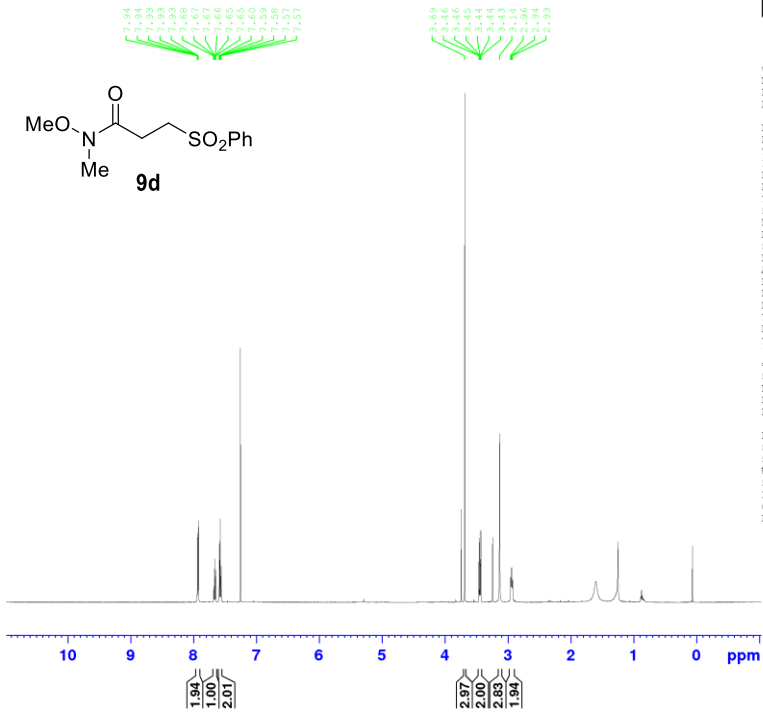
F2 - Acquisition Parameters
 Date_ 20200227
 Time 0.27
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1024
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 23170.5
 DW 20.850 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.0000000 sec
 D11 0.0300000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CDFPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127563 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40





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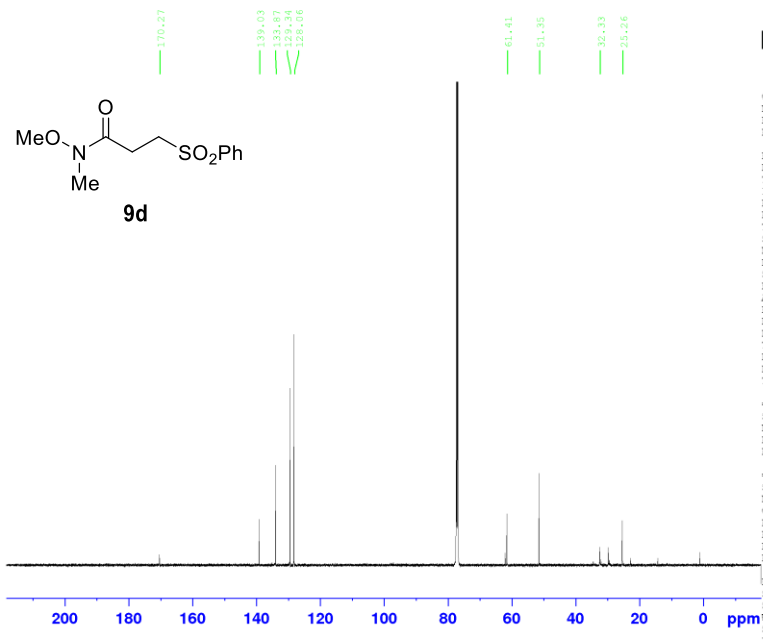
Current Data Parameters
NAME DME-1-167-2F12-19
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date_ 20200226
Time 9.35
INSTRUM spect
PROBHD 5 mm CPQNP 1H/
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 8
DS 2
SWH 6009.615 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 181
DN 83.200 usec
DE 40.00 usec
TE 298.0 K
D1 1.0000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 500.4925025 MHz
NUC1 1H
P1 15.00 usec
PLW1 7.49889994 W

F2 - Processing parameters
SI 32768
SF 500.4900124 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.40

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```

Current Data Parameters
NAME DME-1-167-2F12-19
EXPNO 2
PROCNO 1

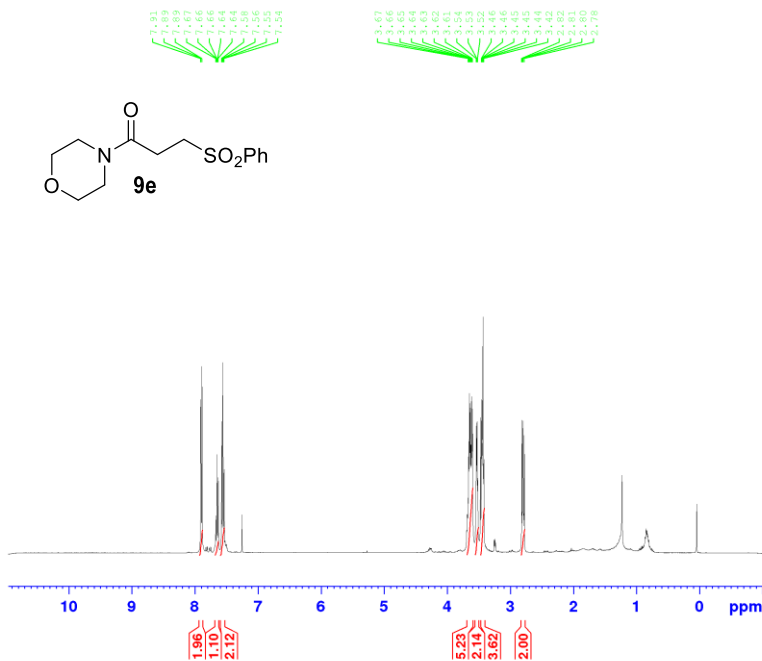
F2 - Acquisition Parameters
Date_ 20200226
Time 9.39
INSTRUM spect
PROBHD 5 mm CPQNP 1H/
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 3316
DS 2
SWH 29761.904 Hz
FIDRES 0.454131 Hz
AQ 1.1010048 sec
RG 1440
DN 16.800 usec
DE 18.00 usec
TE 298.0 K
D1 2.0000000 sec
D11 0.0300000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 125.8608968 MHz
NUC1 13C
P1 10.00 usec
PLW1 63.00000000 W

===== CHANNEL f2 =====
SFO2 500.4925025 MHz
NUC2 1H
CPDPRG2 waltz16
PCPD2 80.00 usec
PLW2 7.49889994 W
PLW12 0.26363000 W
PLW13 0.16873001 W

F2 - Processing parameters
SI 32768
SF 125.8482942 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

```

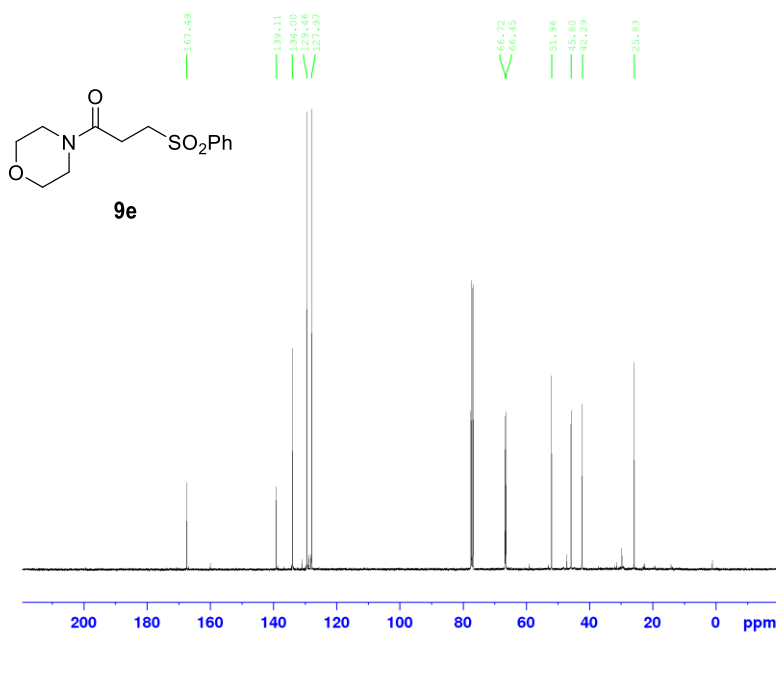


Current Data Parameters
 NAME DMZ-1-120Clean_10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190909
 Time 17.49
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 50.5
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 15394
 SF 400.1300098 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



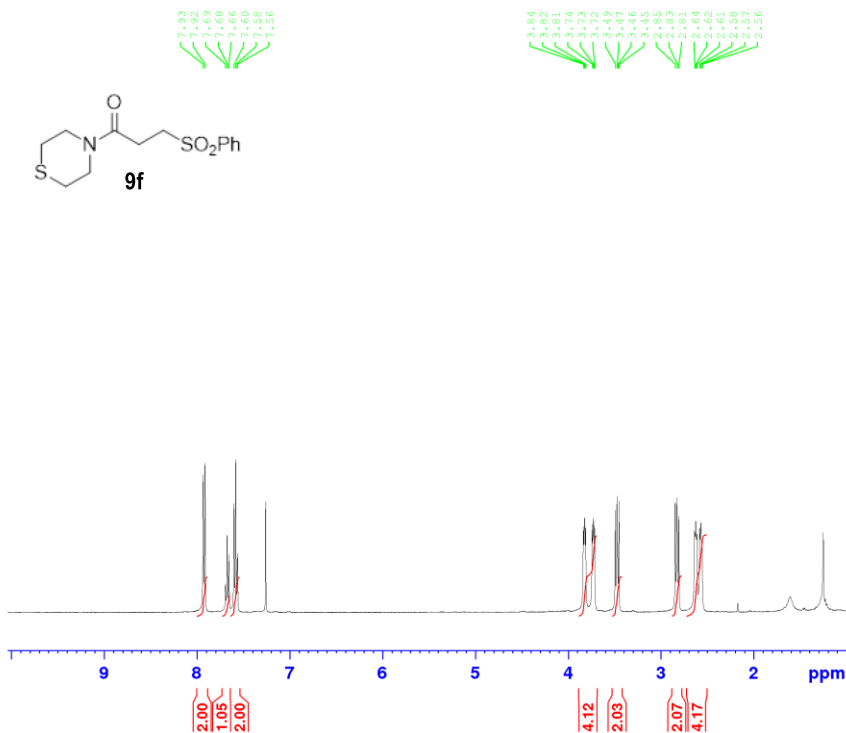
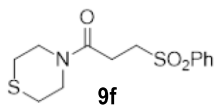
Current Data Parameters
 NAME DMZ-1-120Clean_12
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190910
 Time 0.56
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1508
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 26008
 DW 20.800 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.0000000 sec
 D11 0.0300000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 -3.00 dB
 PL1W 75.17808833 W
 SFO1 100.6303736 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 FL2 -3.00 dB
 PL12 13.05 dB
 PL13 13.05 dB
 PL2W 30.07123375 W
 PL12W 0.74670875 W
 PL13W 0.74670875 W
 SFO2 400.1616006 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6203074 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

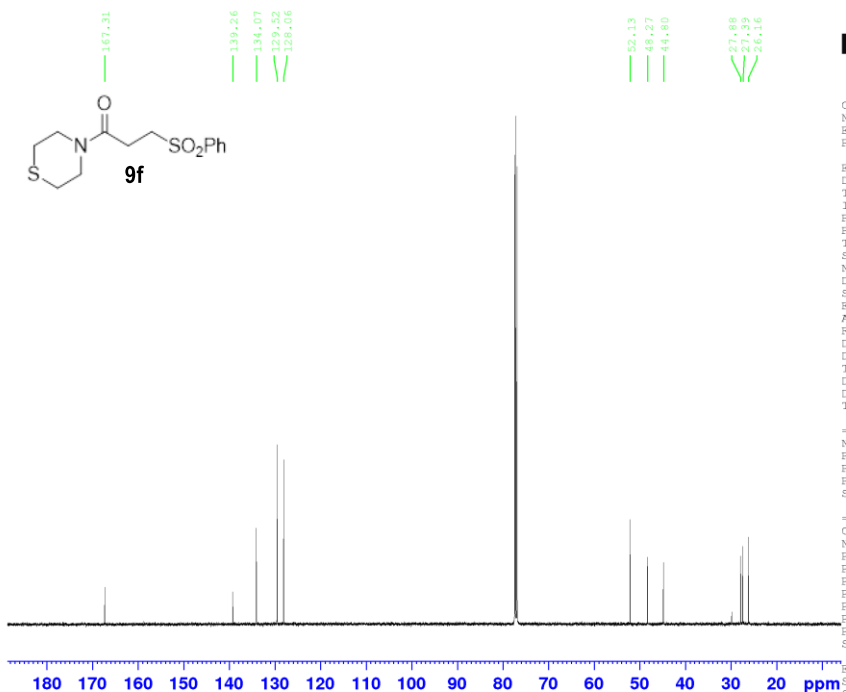
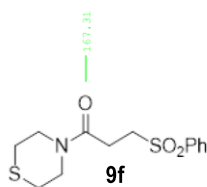


Current Data Parameters
 NAME MBL060-1H (Prodotto)
 EXPNO 3
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200225
 Time 8.32
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 ID 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 256
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SF01 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300098 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



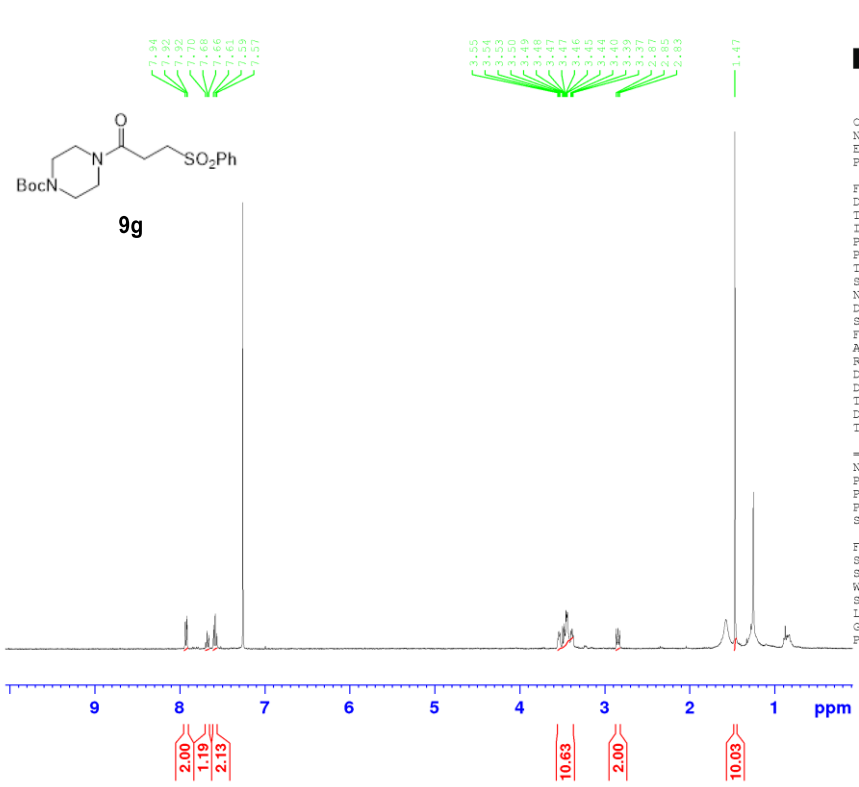
Current Data Parameters
 NAME MBL060-13C_OK
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200225
 Time 17.29
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 2100
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 7290
 DW 16.500 usec
 DE 10.00 usec
 TE 299.8 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SF01 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.08 dB
 PL12 11.54 dB
 PL13 11.54 dB
 PL2W 35.02648163 W
 PL12W 0.96027696 W
 PL13W 0.96027696 W
 SF02 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577740 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

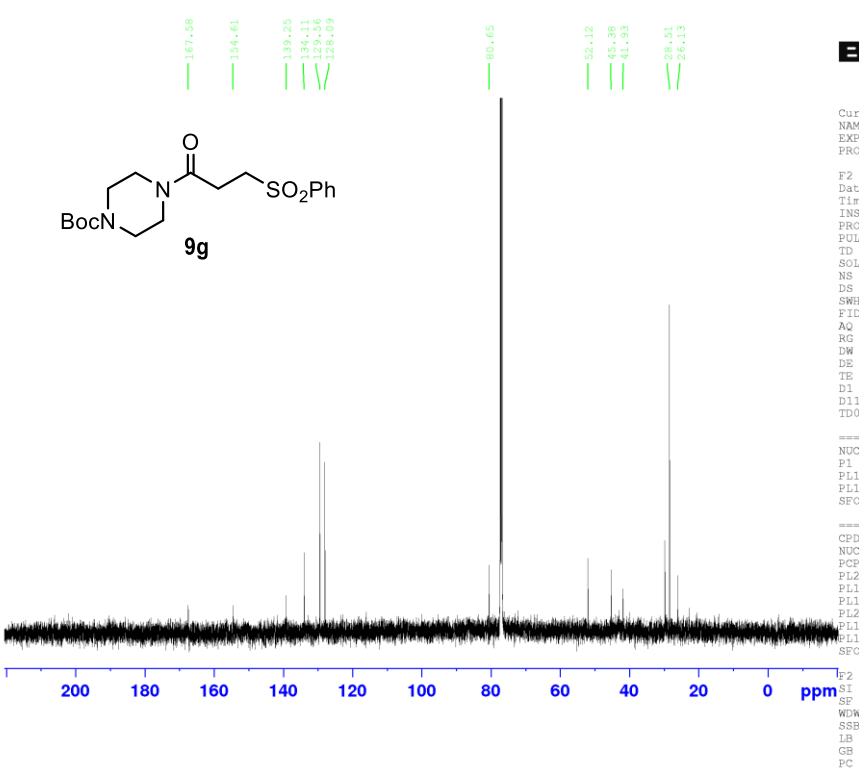


Current Data Parameters
 NAME MBL044_1H (PROC)
 EXPNO 7
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200225
 Time 8.27
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT cdcl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 362
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.0000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300098 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



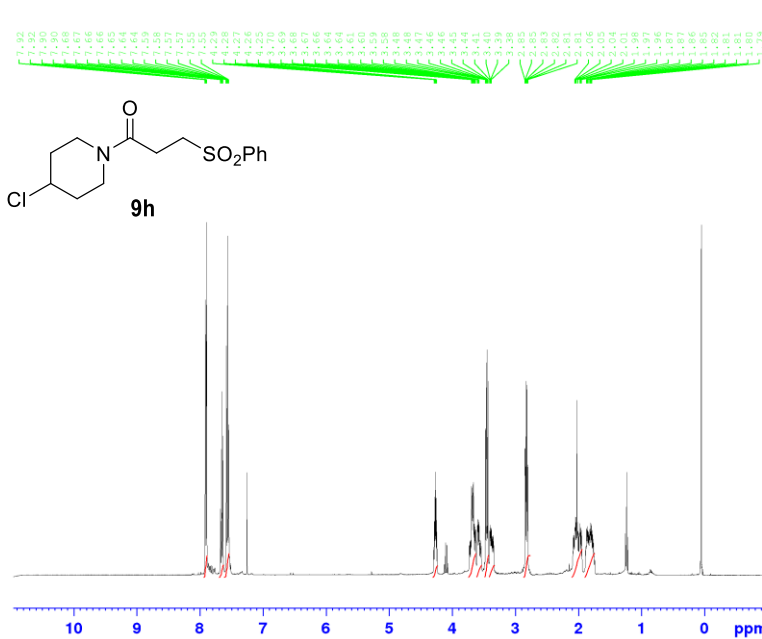
Current Data Parameters
 NAME MBL044-13C_20
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200228
 Time 1.37
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT cdcl3
 NS 4500
 DS 4
 SWH 30303.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 7290
 DW 16.500 usec
 DE 10.00 usec
 TE 298.4 K
 D1 1.0000000 sec
 D11 0.0300000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -4.08 dB
 PL12 11.54 dB
 PL13 11.54 dB
 PL2W 35.02648163 W
 PL12W 0.96027696 W
 PL13W 0.96027696 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577708 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

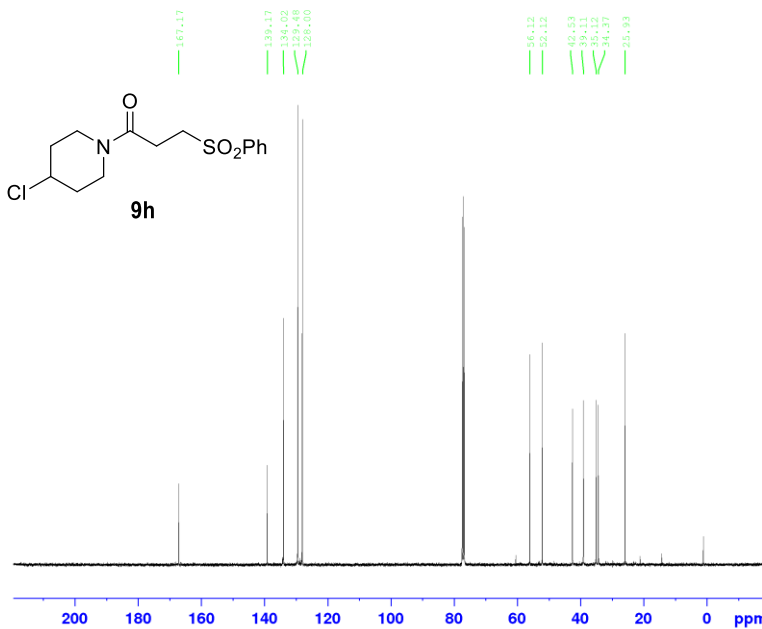


Current Data Parameters
 NAME DMZ-1-221PURE_10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200113
 Time 11.04
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 90.5
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing Parameters
 SI 16384
 SF 400.1300095 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

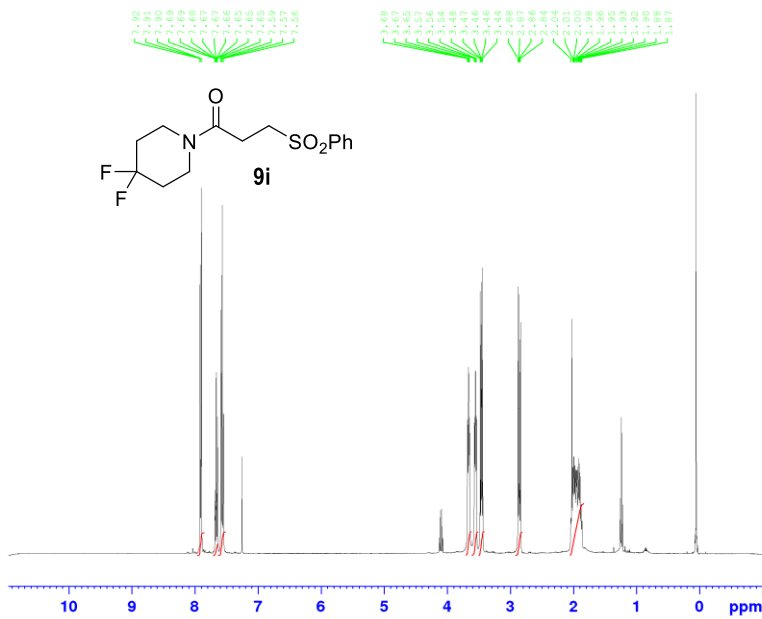


Current Data Parameters
 NAME DMZ-1-221PURE_11
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200113
 Time 21.08
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 1524
 DS 4
 SWH 24038.461 Hz
 FIDRES 0.366798 Hz
 AQ 1.3631488 sec
 RG 26008
 DW 20.800 usec
 DE 10.00 usec
 TE 298.0 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6303736 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 F2 2.00 dB
 PL12 13.05 dB
 PL13 13.05 dB
 PL1W 30.07123375 W
 PL12W 0.74670875 W
 PL13W 0.74670875 W
 SFO

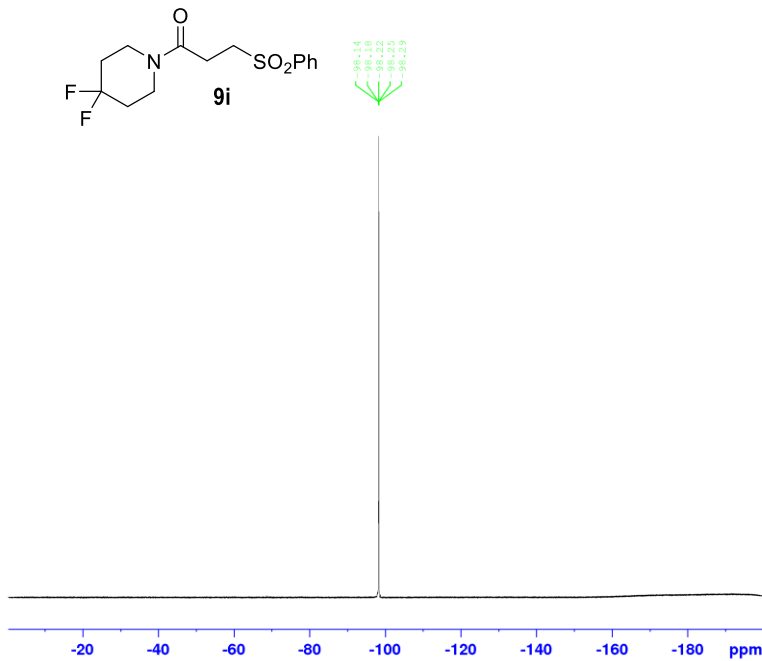


Current Data Parameters
 NAME DME-1-226Fr36-43_10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200116
 Time 17:33
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 ID 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 50.5
 DW 104.400 usec
 DE 6.00 usec
 TE 298.0 K
 D1 1.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16394
 SF 400.1300095 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

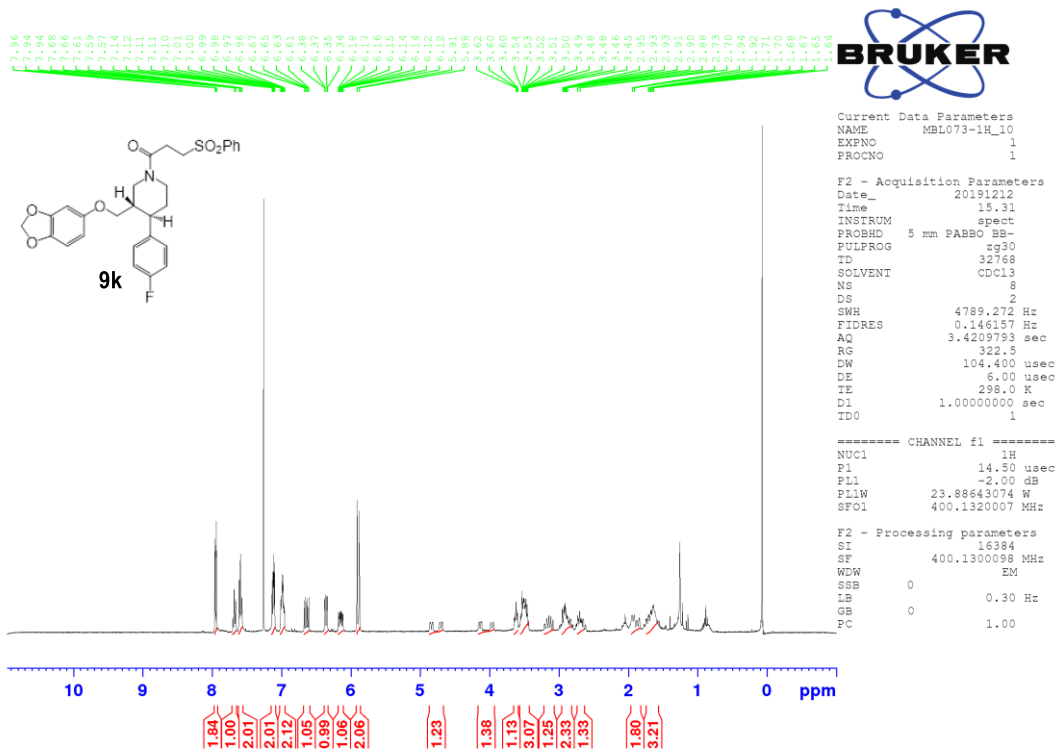
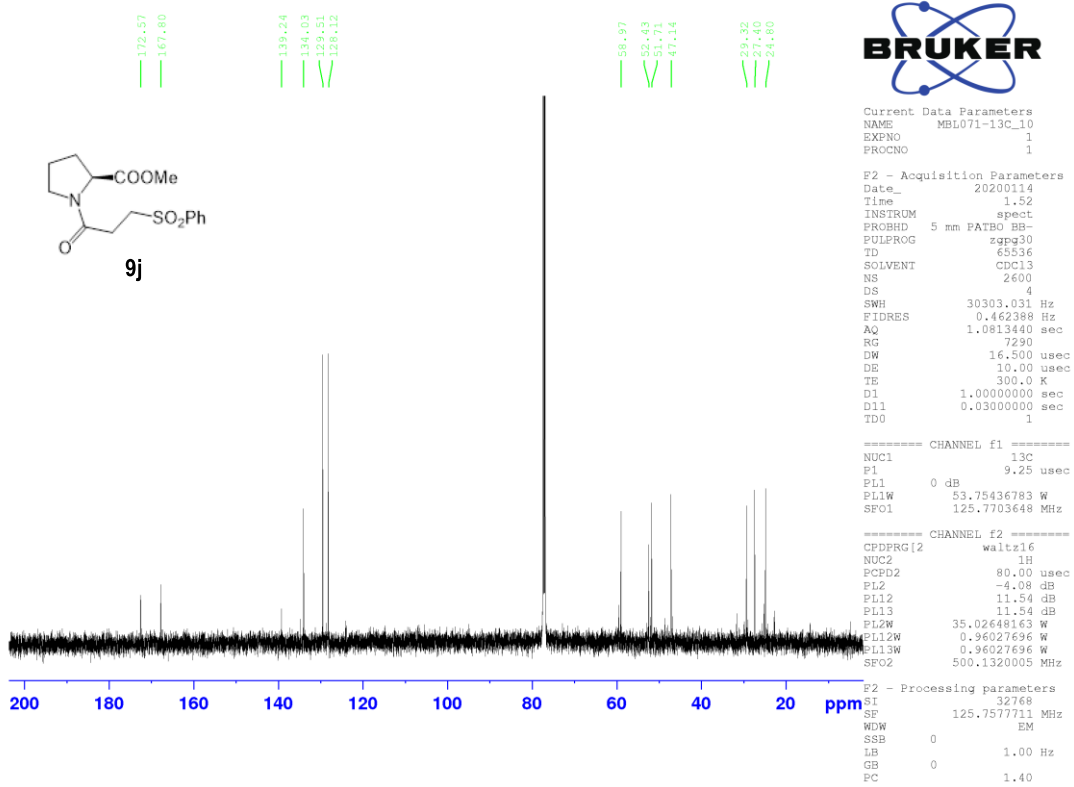


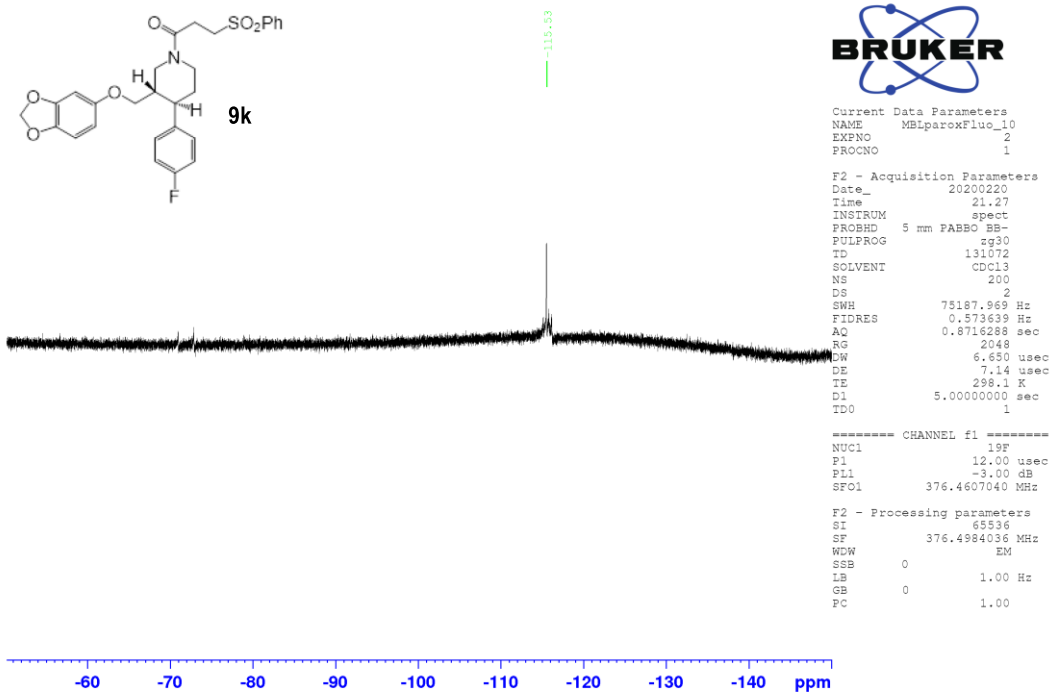
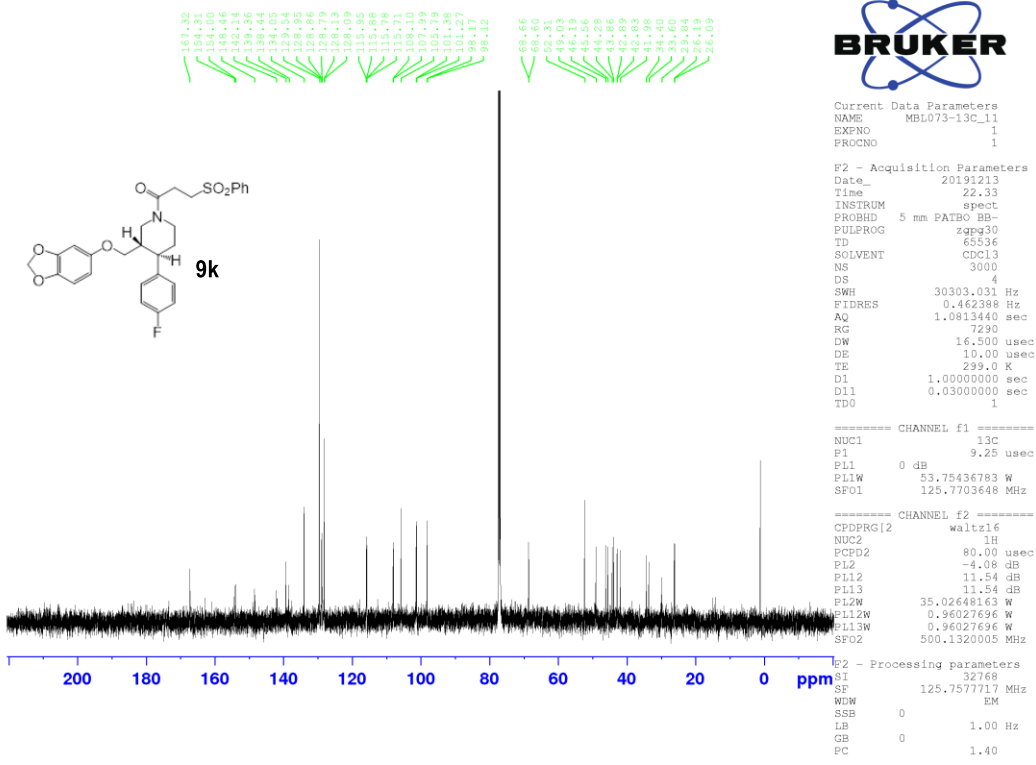
Current Data Parameters
 NAME DME-1-226Fr36-43_11
 EXPNO 1
 PROCNO 1

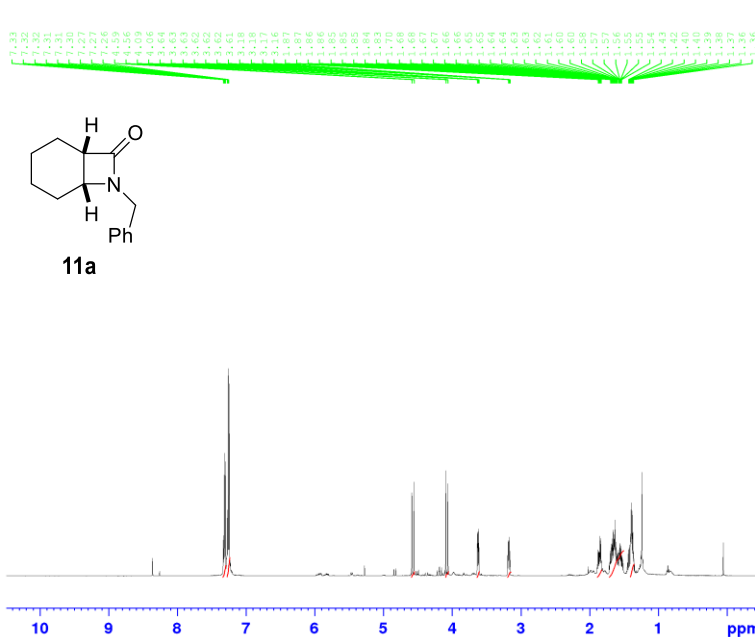
F2 - Acquisition Parameters
 Date_ 20200116
 Time 17:34
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 ID 131072
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 75187.969 Hz
 FIDRES 0.573639 Hz
 AQ 0.8716288 sec
 RG 2896.3
 DW 6.650 usec
 DE 7.14 usec
 TE 298.0 K
 D1 5.0000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13F
 P1 12.00 usec
 PL1 -3.00 dB
 SFO1 376.4607040 MHz

F2 - Processing parameters
 SI 65536
 SF 376.4984036 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.00





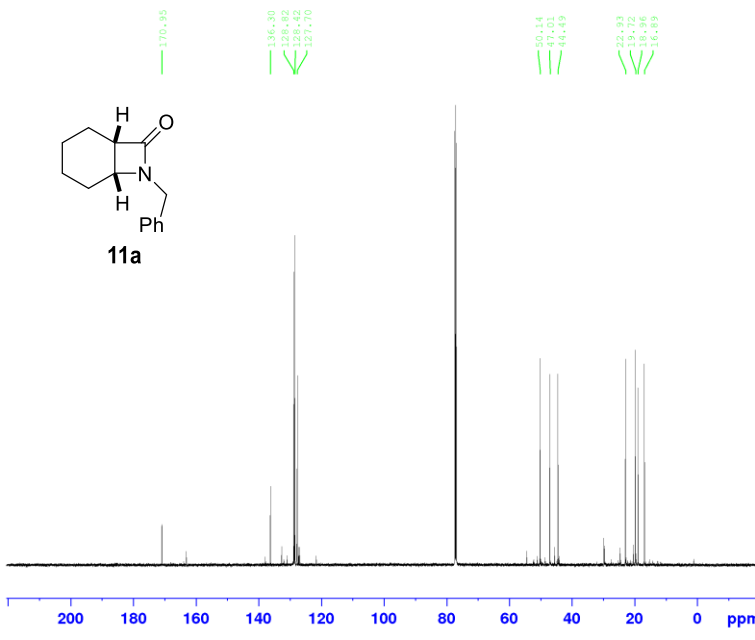
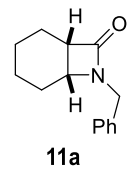


Current Data Parameters
 NAME DMZ-1-072PURE_10
 EXPNO 2
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190628
 Time 15.13
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6009.615 Hz
 FIDRES 0.183399 Hz
 AQ 2.7262976 sec
 RG 203
 DW 83.200 usec
 DE 10.00 usec
 TE 300.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 1H
 P1 14.00 usec
 PL1 -4.08 dB
 PL1W 35.02648163 W
 SFO1 500.1325007 MHz

F2 - Processing parameters
 SI 16384
 SF 500.1300210 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00



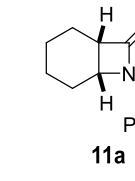
Current Data Parameters
 NAME DMZ-1-072PURE_11
 EXPNO 3
 PROCNO 1

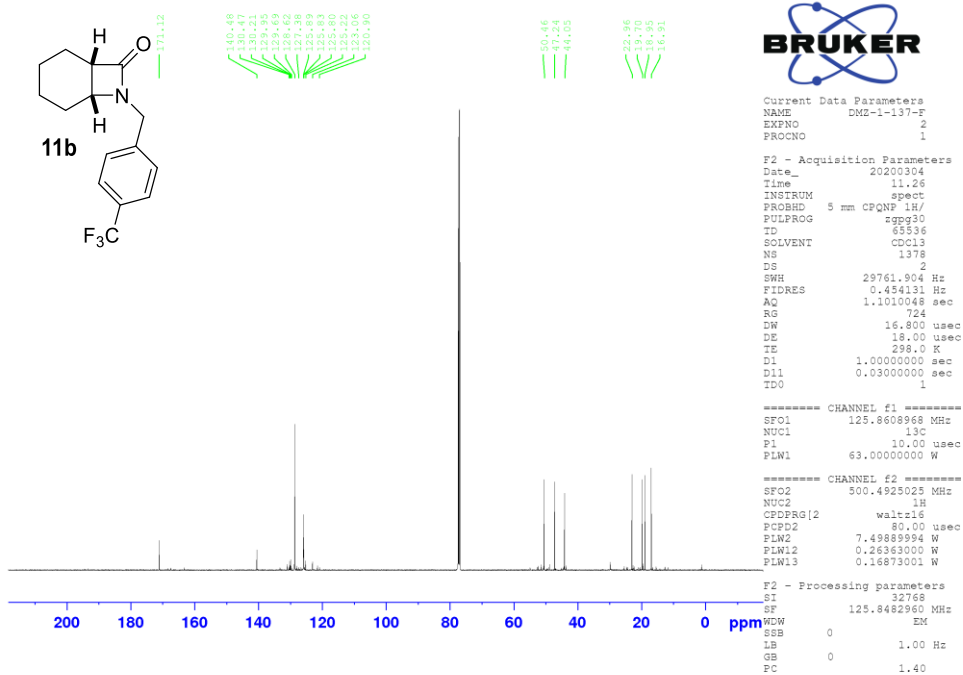
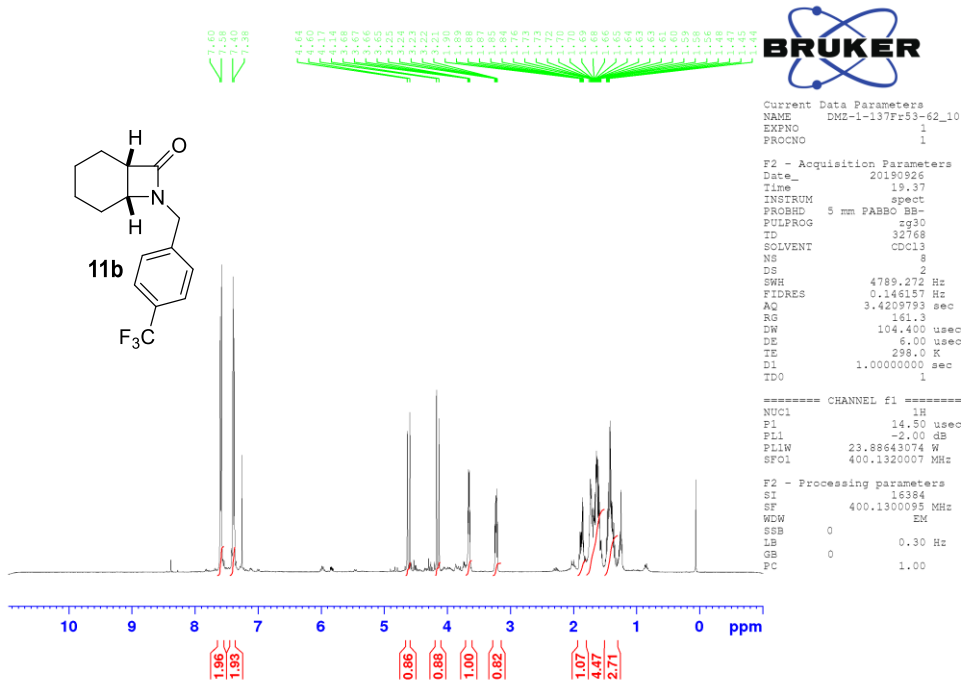
F2 - Acquisition Parameters
 Date_ 20190628
 Time 23.57
 INSTRUM spect
 PROBHD 5 mm PATBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 2048
 DS 4
 SWH 30305.031 Hz
 FIDRES 0.462388 Hz
 AQ 1.0813440 sec
 RG 9200
 DW 16.500 usec
 DE 10.00 usec
 TE 298.9 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TDO 1

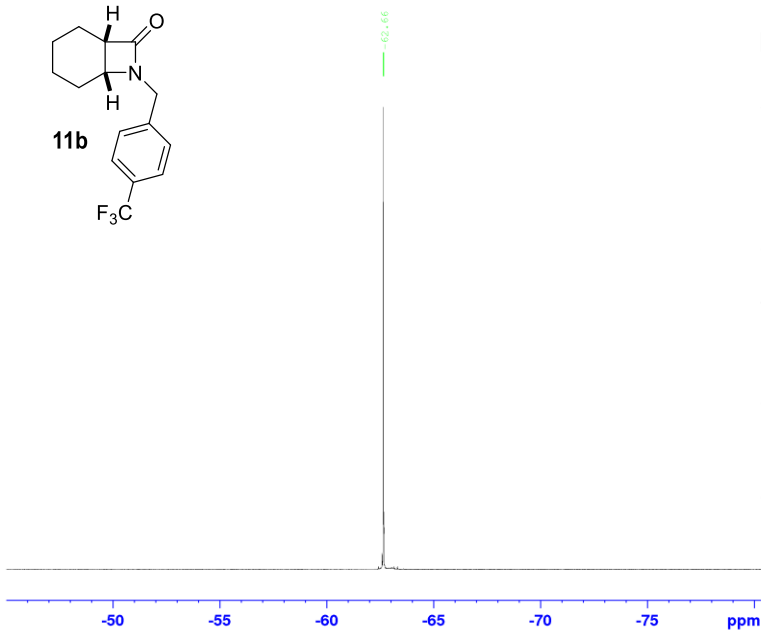
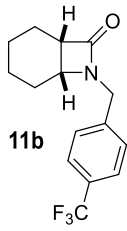
===== CHANNEL f1 =====
 NUC1 13C
 P1 9.25 usec
 PL1 0 dB
 PL1W 53.75436783 W
 SFO1 125.7703648 MHz

===== CHANNEL f2 =====
 CPDPRG2 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 FL2 -4.08 dB
 FL12 11.54 dB
 FL13 11.54 dB
 PL2W 35.02648163 W
 FL12W 0.96027696 W
 FL13W 0.96027696 W
 SFO2 500.1320005 MHz

F2 - Processing parameters
 SI 32768
 SF 125.7577758 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40





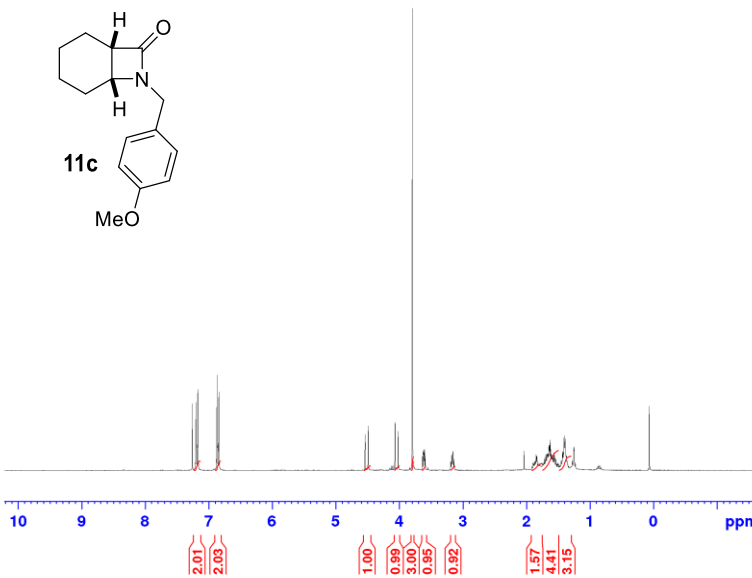
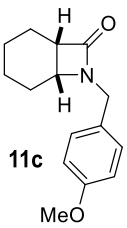


Current Data Parameters
 NAME DME-1-137-F_10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200302
 Time 16.27
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 ID 131072
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 75187.969 Hz
 FIDRES 0.573639 Hz
 AQ 0.8716288 sec
 RS 2298.8
 DW 6.650 usec
 DE 7.14 usec
 TE 298.0 K
 D1 5.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 NUC1 13C
 P1 12.00 usec
 PL1 -3.00 dB
 SFO1 376.4607040 MHz

F2 - Processing parameters
 SI 65536
 SF 376.4964036 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 FC 1.00

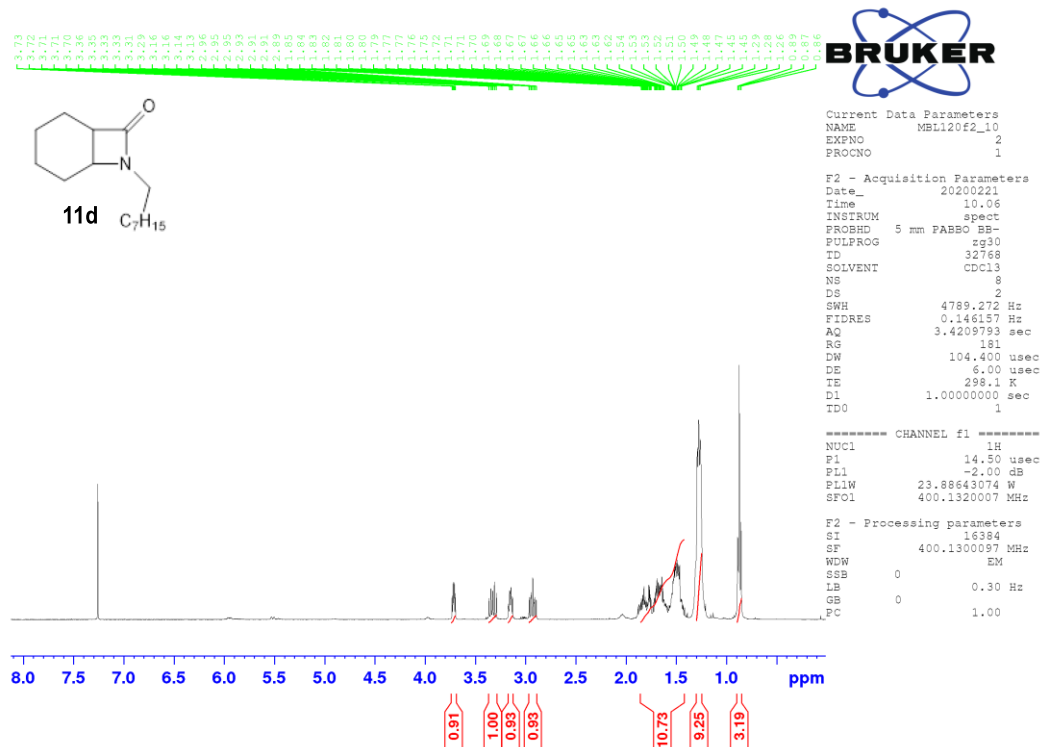
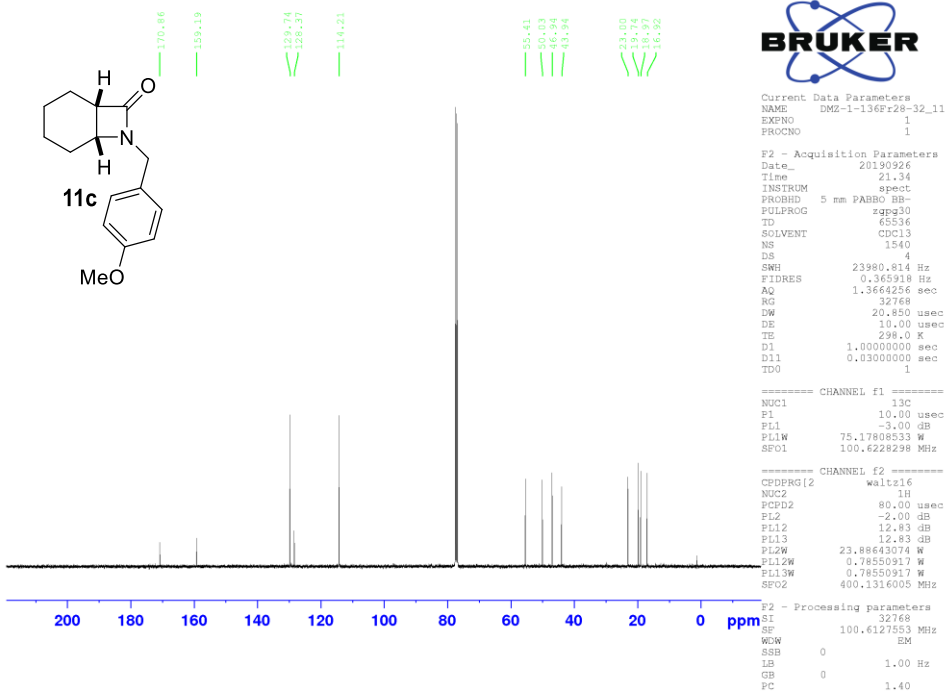


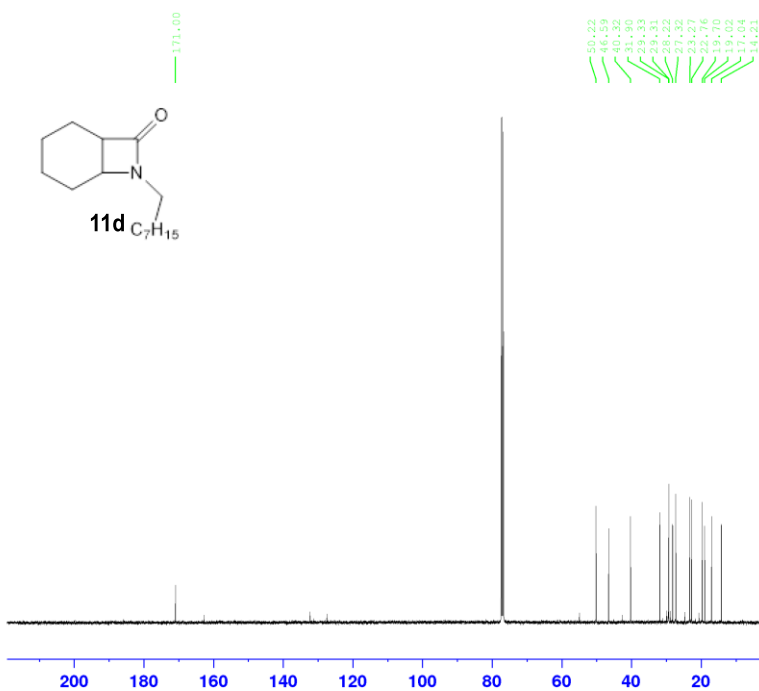
Current Data Parameters
 NAME DME-1-136Fz28-32_10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190926
 Time 14.54
 INSTRUM FOURIER300
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 ID 68836
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6103.516 Hz
 FIDRES 0.093132 Hz
 AQ 5.3687091 sec
 RS 48.5692
 DW 81.920 usec
 DE 6.50 usec
 TE 298.0 K
 D1 1.00000000 sec
 TDO 1

===== CHANNEL f1 =====
 SFO1 300.1818011 MHz
 NUC1 1H
 P1 11.75 usec
 PLW1 20.00000000 W

F2 - Processing parameters
 SI 32768
 SF 300.1800057 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 FC 1.40





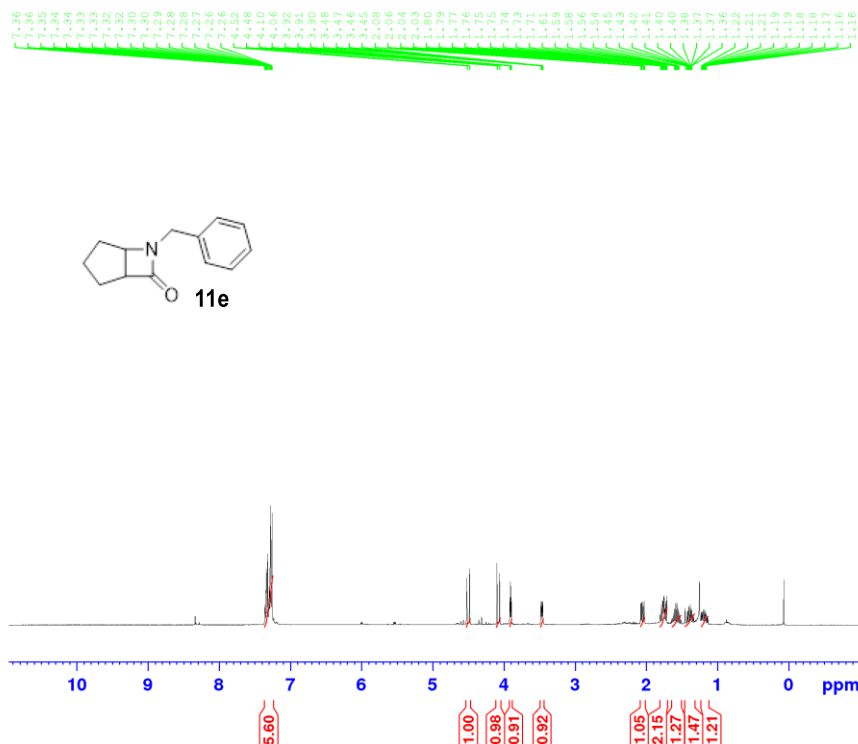
Current Data Parameters
 NAME MBL120-13C_10
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20200221
 Time 22.11
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 3000
 DS 4
 SWH 23980.814 Hz
 FIDRES 0.365918 Hz
 AQ 1.3664256 sec
 RG 23170.5
 DW 20.850 usec
 DE 10.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 TD0 1

===== CHANNEL f1 =====
 NUC1 13c
 P1 10.00 usec
 PL1 -3.00 dB
 PL1W 75.17808533 W
 SFO1 100.6228298 MHz

===== CHANNEL f2 =====
 CPDPRG12 waltz16
 NUC2 1H
 PCPD2 80.00 usec
 PL2 -2.00 dB
 PL12 12.83 dB
 PL13 12.83 dB
 PL2W 23.88643074 W
 PL12W 0.78550917 W
 PL13W 0.78550917 W
 SFO2 400.1316005 MHz

F2 - Processing parameters
 SI 32768
 SF 100.6127551 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

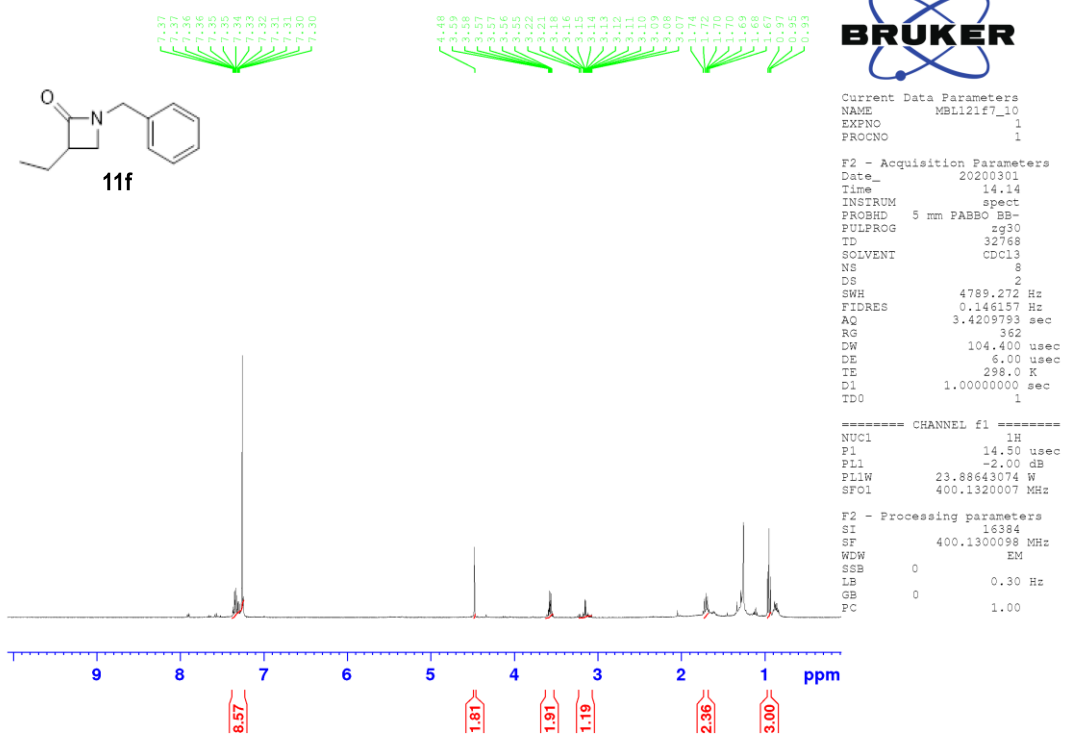
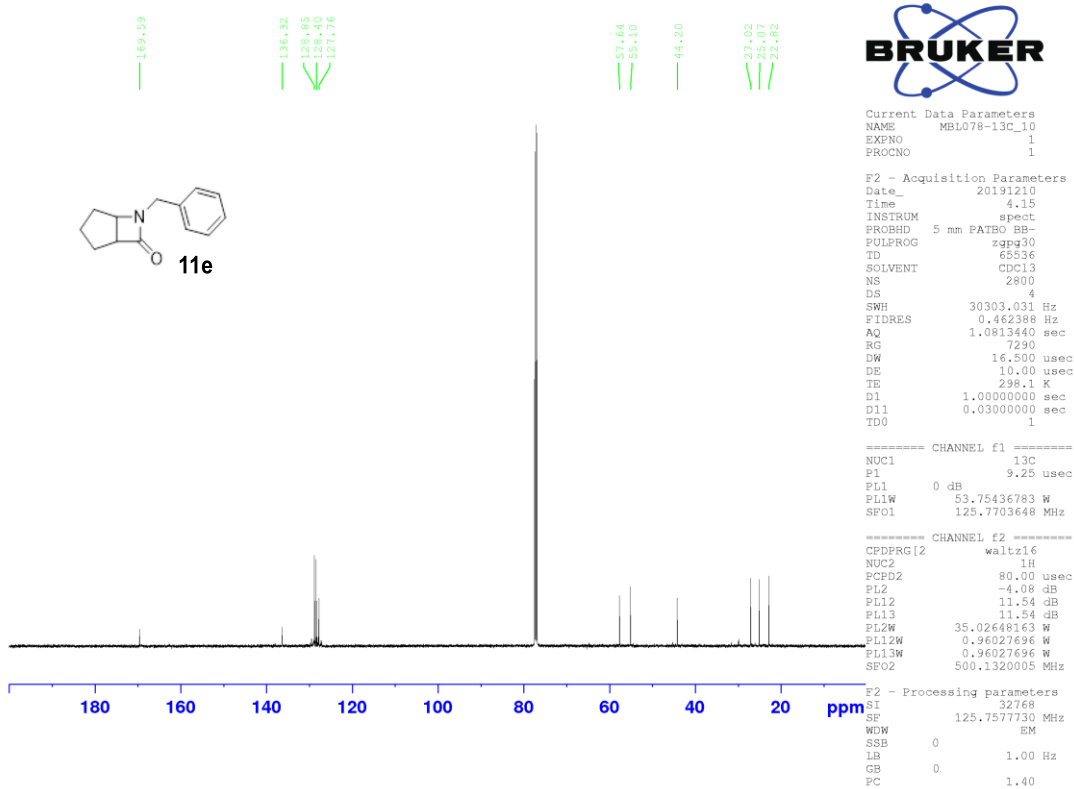


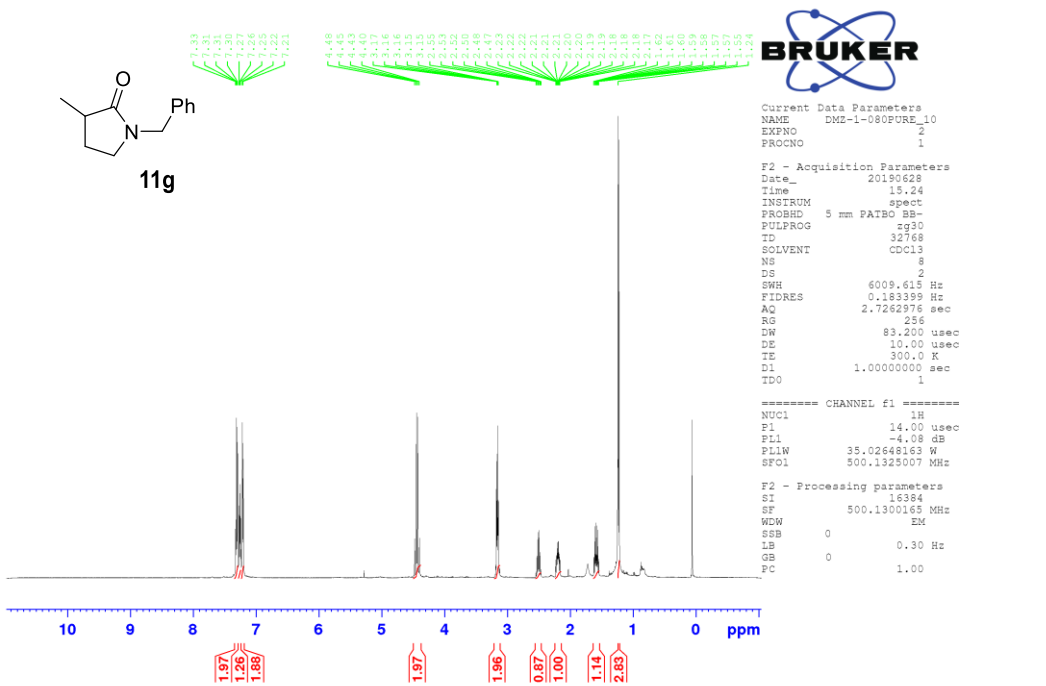
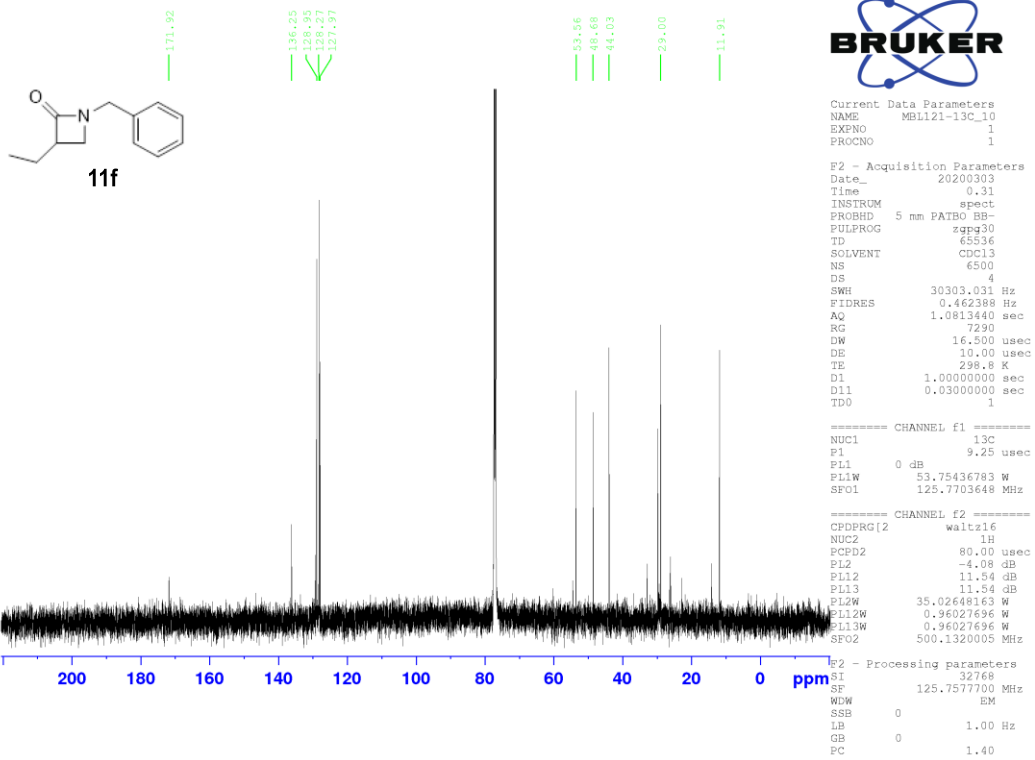
Current Data Parameters
 NAME MBL078-f2_ (PROD)
 EXPNO 2
 PROCNO 1

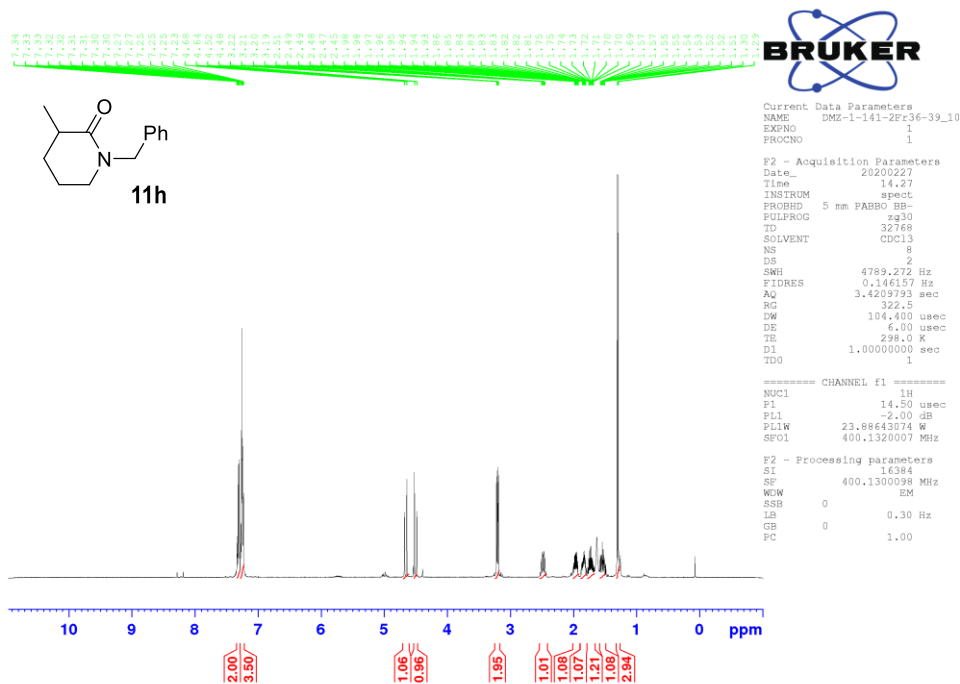
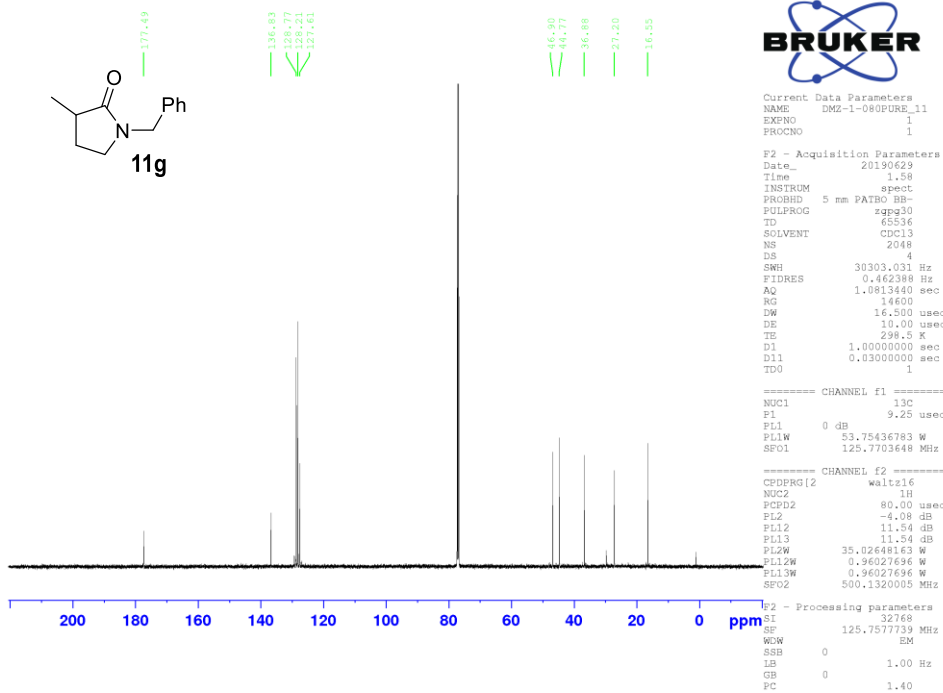
F2 - Acquisition Parameters
 Date_ 20191209
 Time 18.54
 INSTRUM spect
 PROBHD 5 mm PABBO BB-
 PULPROG zg30
 TD 32768
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 4789.272 Hz
 FIDRES 0.146157 Hz
 AQ 3.4209793 sec
 RG 181
 DW 104.400 usec
 DE 6.00 usec
 TE 298.1 K
 D1 1.00000000 sec
 TD0 1

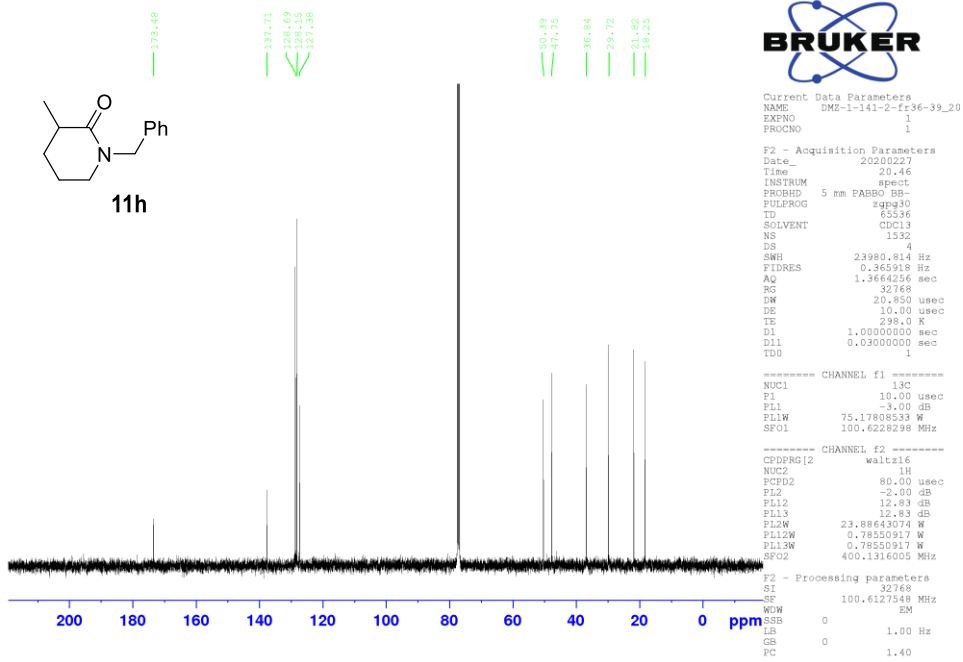
===== CHANNEL f1 =====
 NUC1 1H
 P1 14.50 usec
 PL1 -2.00 dB
 PL1W 23.88643074 W
 SFO1 400.1320007 MHz

F2 - Processing parameters
 SI 16384
 SF 400.1300101 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.00

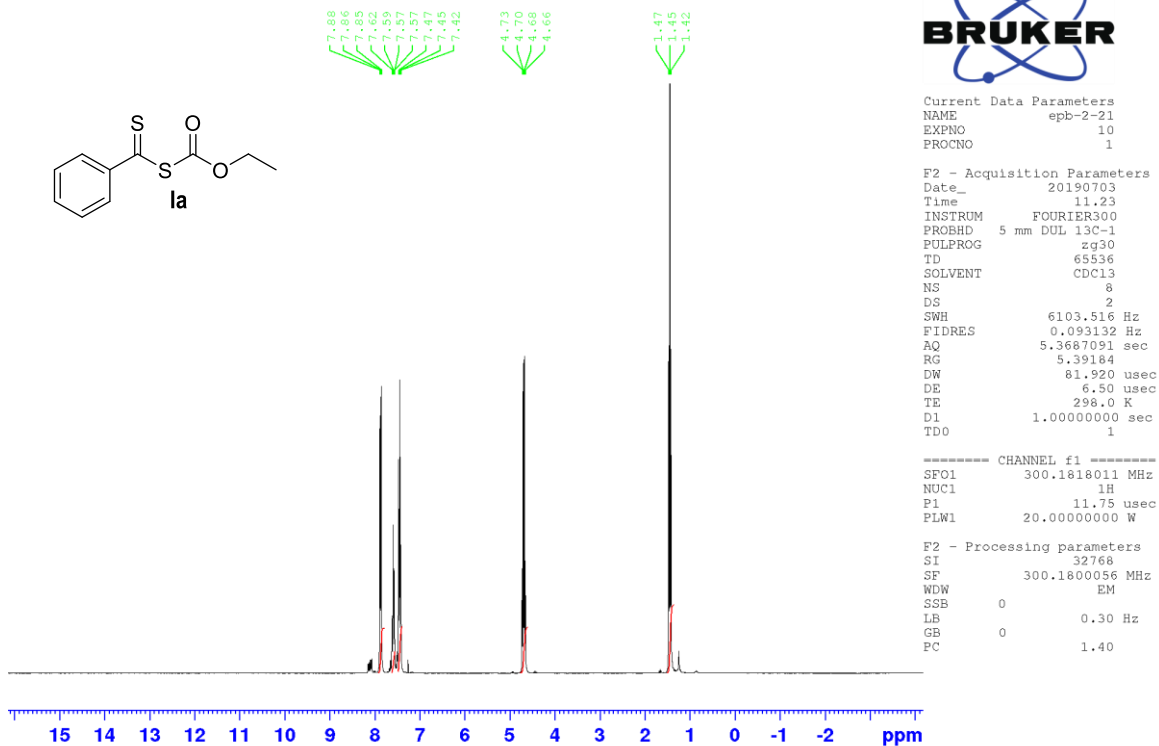








F3. Acyl Intermediates, TEMPO Trapping and Group Transfer Products

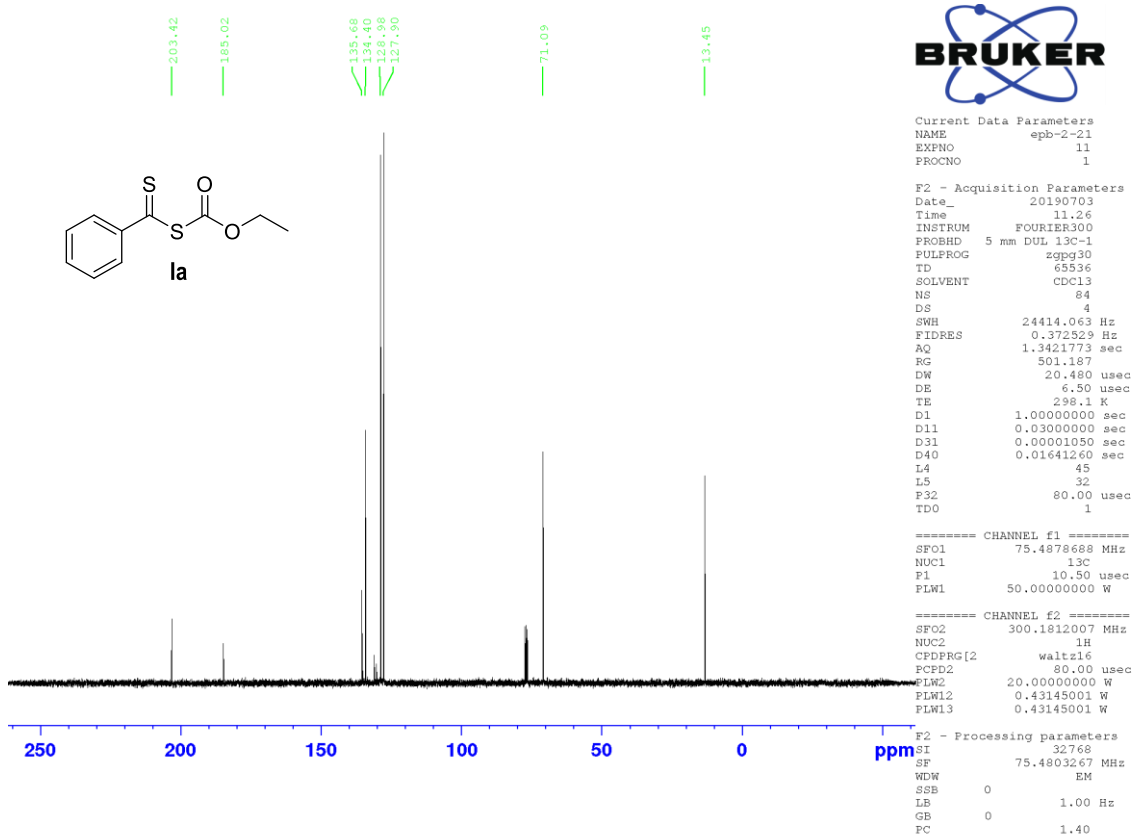


Current Data Parameters
 NAME epb-2-21
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190703
 Time 11.23
 INSTRUM FOURIER300
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6103.516 Hz
 FIDRES 0.093132 Hz
 AQ 5.3687091 sec
 RG 5.39184
 DW 81.920 usec
 DE 6.50 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

----- CHANNEL f1 -----
 SFO1 300.1818011 MHz
 NUC1 1H
 P1 11.75 usec
 PLW1 20.00000000 W

F2 - Processing parameters
 SI 32768
 SF 300.1800056 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40



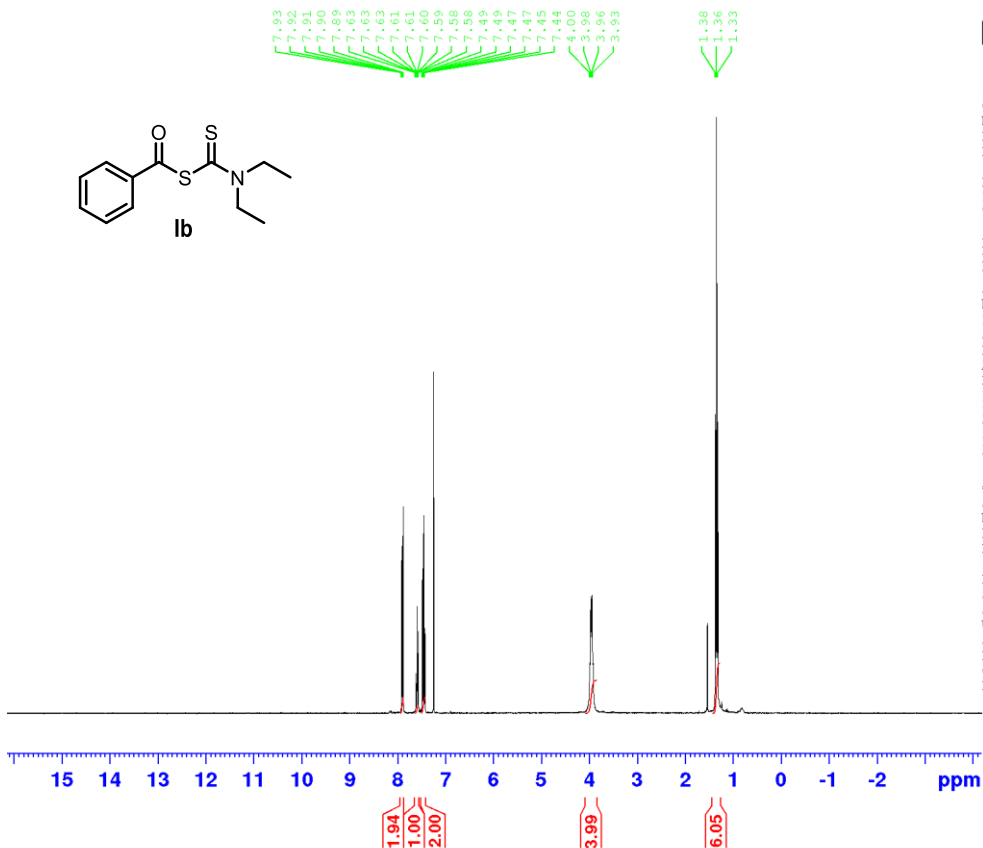
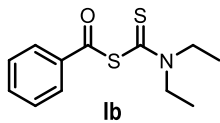
Current Data Parameters
 NAME epb-2-21
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190703
 Time 11.26
 INSTRUM FOURIER300
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 84
 DS 4
 SWH 24414.063 Hz
 FIDRES 0.372529 Hz
 AQ 1.3421773 sec
 RG 501.187
 DW 20.480 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D31 0.00001050 sec
 D40 0.01641260 sec
 L4 45
 L5 32
 F32 80.00 usec
 TD0 1

----- CHANNEL f1 -----
 SFO1 75.4878688 MHz
 NUC1 13C
 P1 10.50 usec
 PLW1 50.00000000 W

----- CHANNEL f2 -----
 SFO2 300.1812007 MHz
 NUC2 1H
 CDFPRG2 waltz16
 PCPD2 80.00 usec
 PLW2 20.00000000 W
 PLW12 0.43145001 W
 PLW13 0.43145001 W

F2 - Processing parameters
 SI 32768
 SF 75.4803267 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

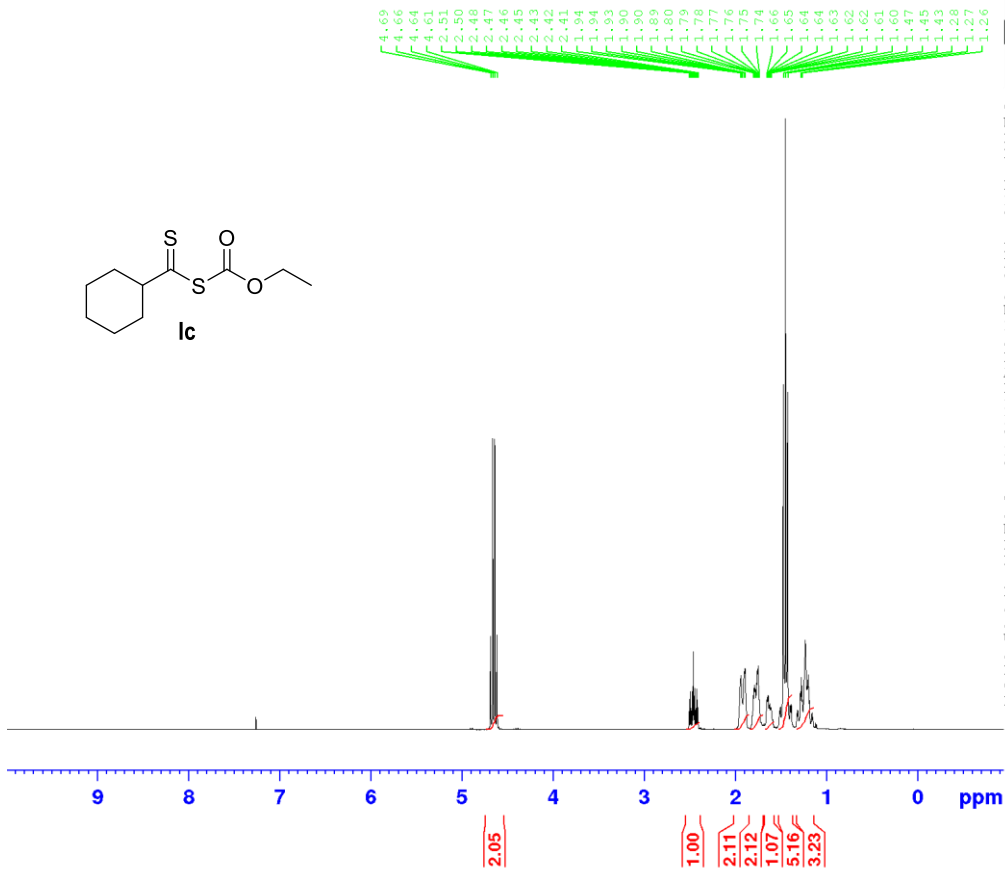
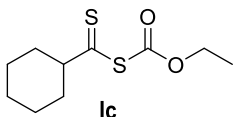


Current Data Parameters
 NAME epb-2-22-f1
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190724
 Time 10.45
 INSTRUM FOURIER300
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6103.516 Hz
 FIDRES 0.093132 Hz
 AQ 5.3687091 sec
 RG 97.5238
 DW 81.920 usec
 DE 6.50 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 300.1818011 MHz
 NUC1 1H
 P1 11.75 usec
 PLW1 20.00000000 W

F2 - Processing parameters
 SI 32768
 SF 300.1800055 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40

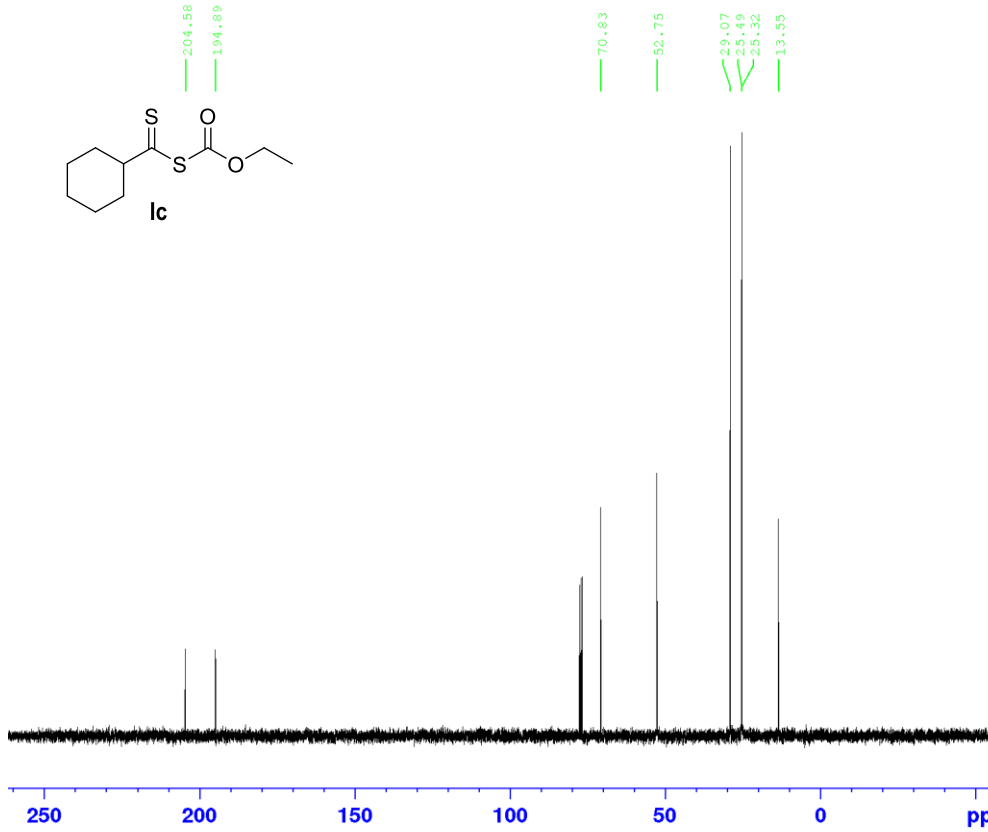
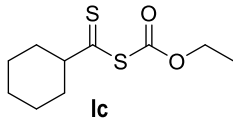


Current Data Parameters
 NAME epb-2-18
 EXPNO 10
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190703
 Time 11.13
 INSTRUM FOURIER300
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6103.516 Hz
 FIDRES 0.093132 Hz
 AQ 5.3687091 sec
 RG 4.72252
 DW 81.920 usec
 DE 6.50 usec
 TE 298.0 K
 D1 1.00000000 sec
 TD0 1

===== CHANNEL f1 =====
 SFO1 300.1818011 MHz
 NUC1 1H
 P1 11.75 usec
 PLW1 20.00000000 W

F2 - Processing parameters
 SI 32768
 SF 300.1800057 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40



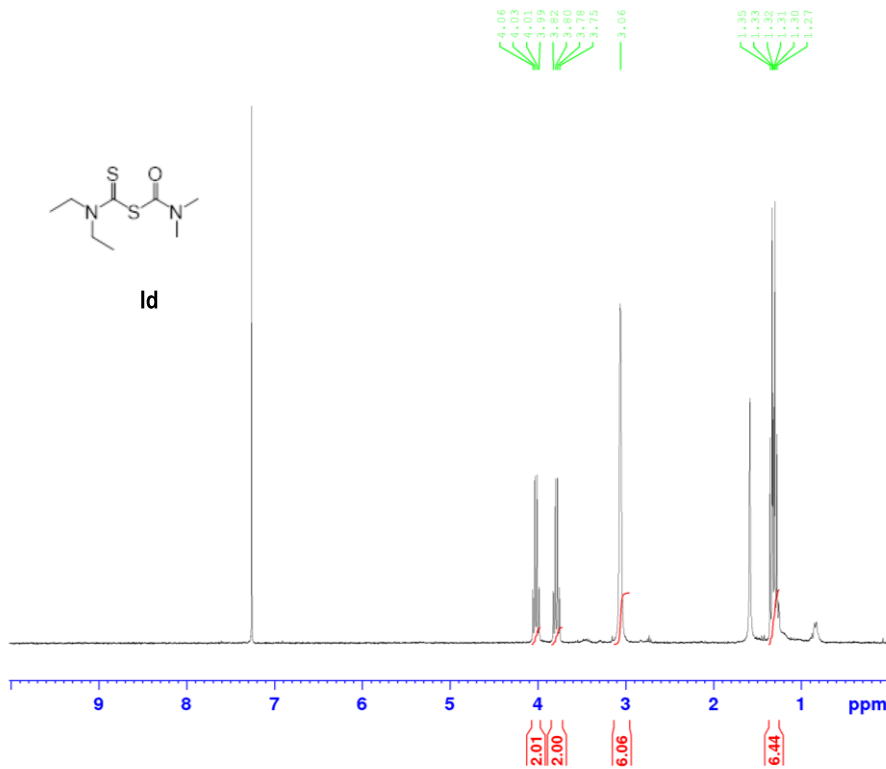
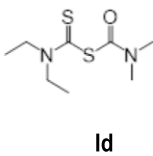
Current Data Parameters
 NAME epb-2-18
 EXPNO 11
 PROCNO 1

F2 - Acquisition Parameters
 Date_ 20190703
 Time 11.16
 INSTRUM FOURIER300
 PROBHD 5 mm DUL 13C-1
 PULPROG zgpg30
 TD 65536
 SOLVENT CDCl3
 NS 43
 DS 4
 SWH 24414.063 Hz
 FIDRES 0.372529 Hz
 AQ 1.3421773 sec
 RG 501.187
 DW 20.480 usec
 DE 6.50 usec
 TE 298.1 K
 D1 1.00000000 sec
 D11 0.03000000 sec
 D31 0.00001050 sec
 D40 0.01641260 sec
 L4 45
 L5 32
 P32 80.00 usec
 TDO 1

===== CHANNEL f1 =====
 SFO1 75.4878688 MHz
 NUC1 13C
 P1 10.50 usec
 PLW1 50.00000000 W

===== CHANNEL f2 =====
 SFO2 300.1812007 MHz
 NUC2 1H
 CPDPRG2 waltz16
 PCPD2 80.00 usec
 PLW2 20.00000000 W
 PLW12 0.43145001 W
 PLW13 0.43145001 W

F2 - Processing parameters
 SI 32768
 SF 75.4803190 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

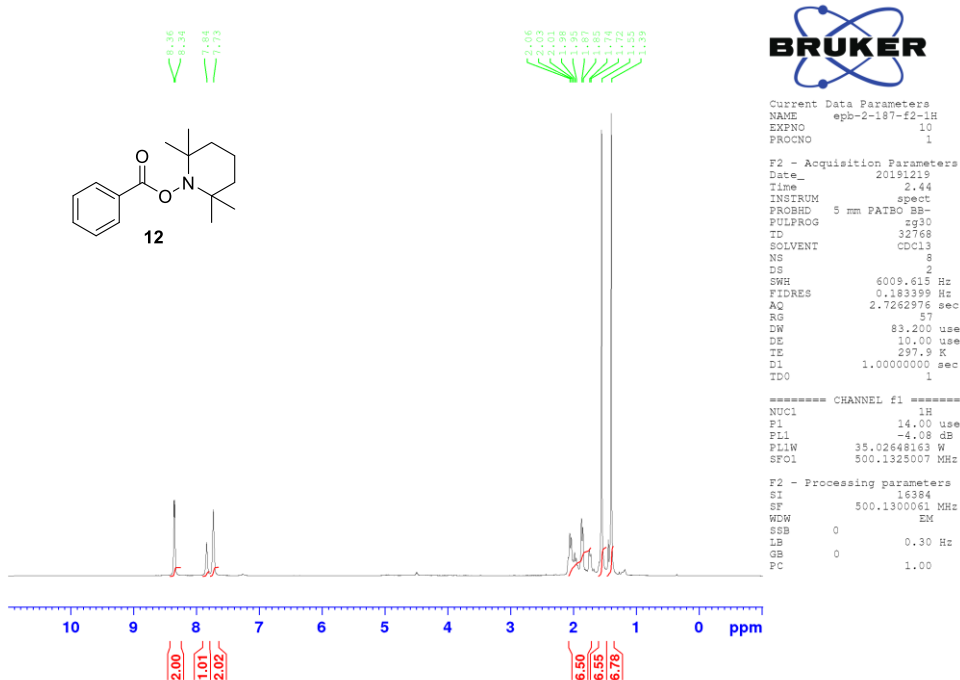
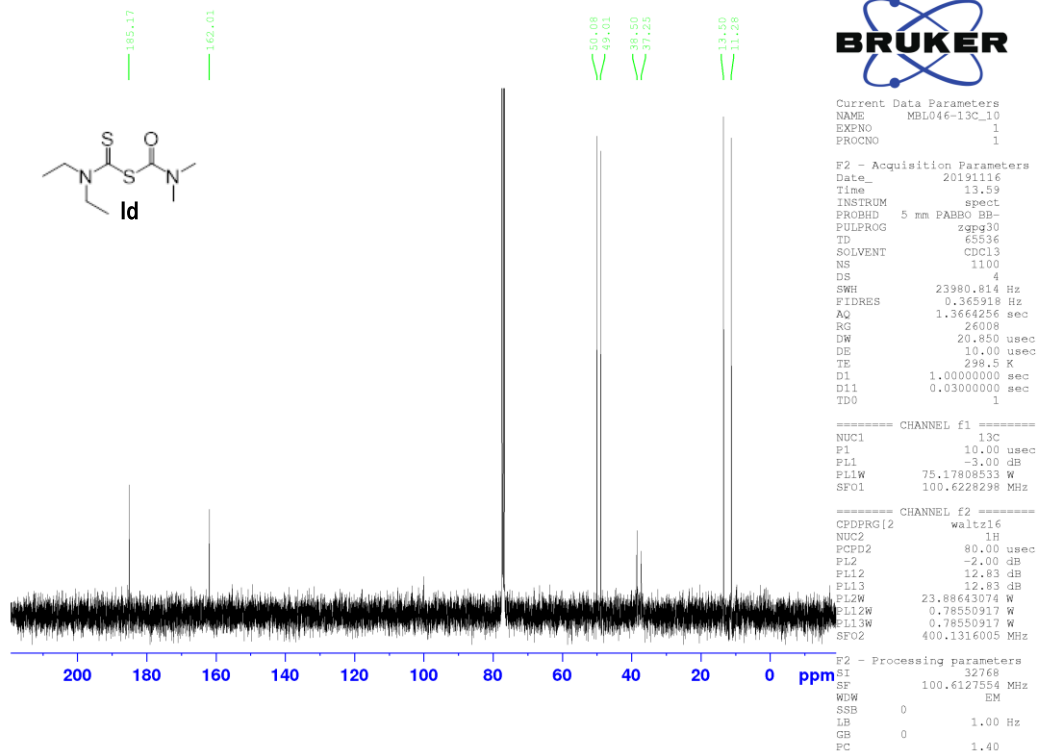


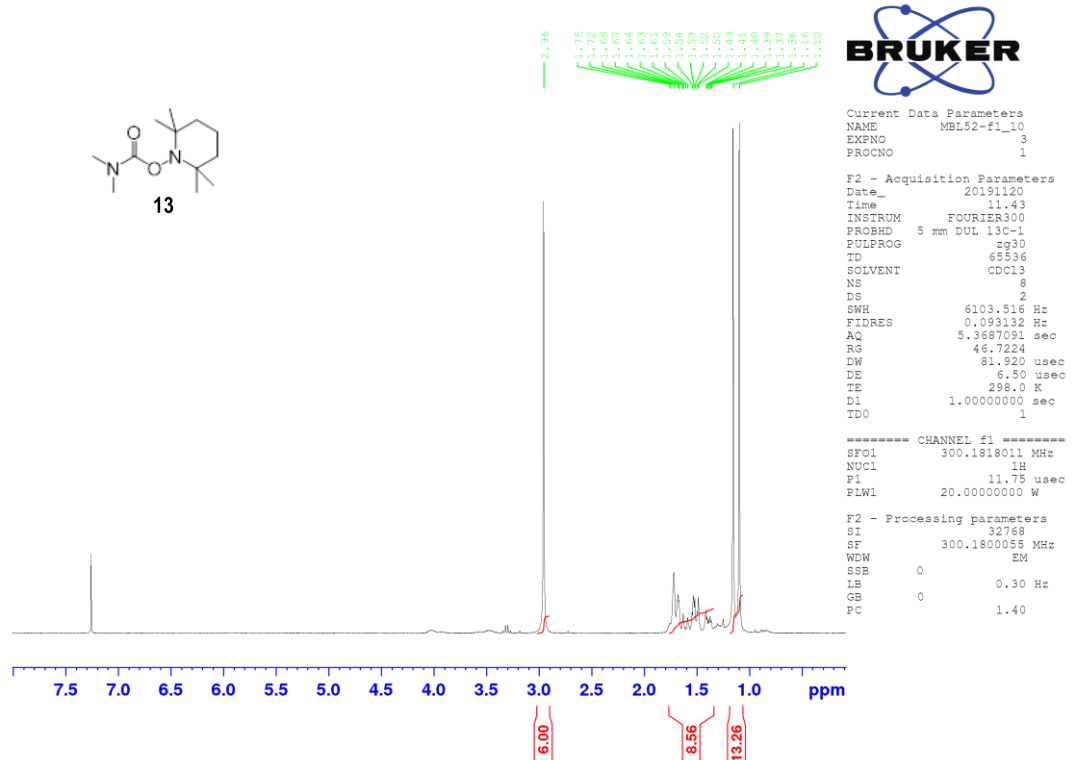
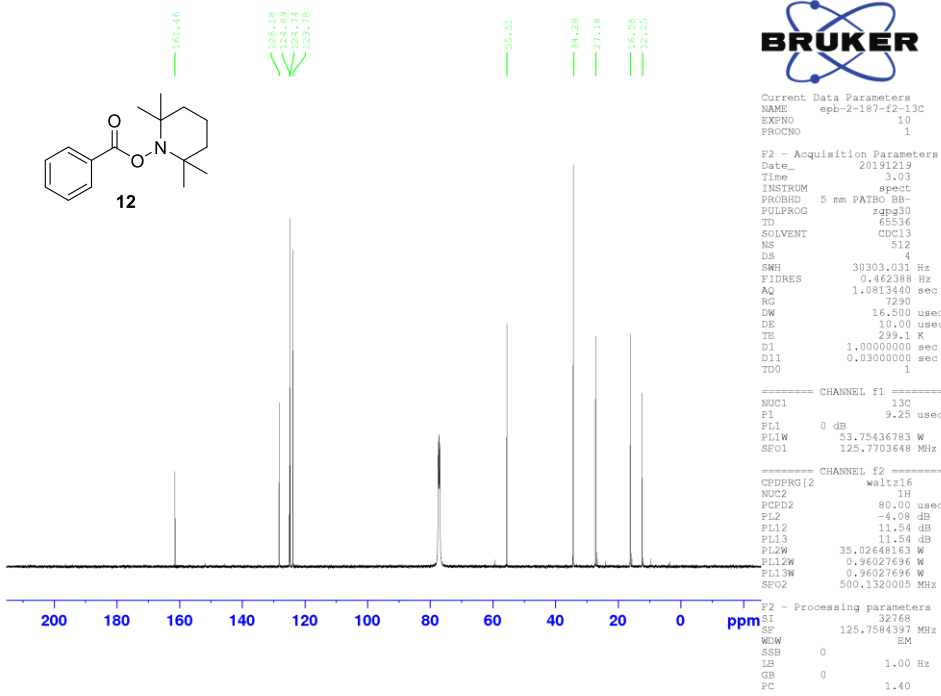
Current Data Parameters
 NAME MBL046_10
 EXPNO 1
 PROCNO 1

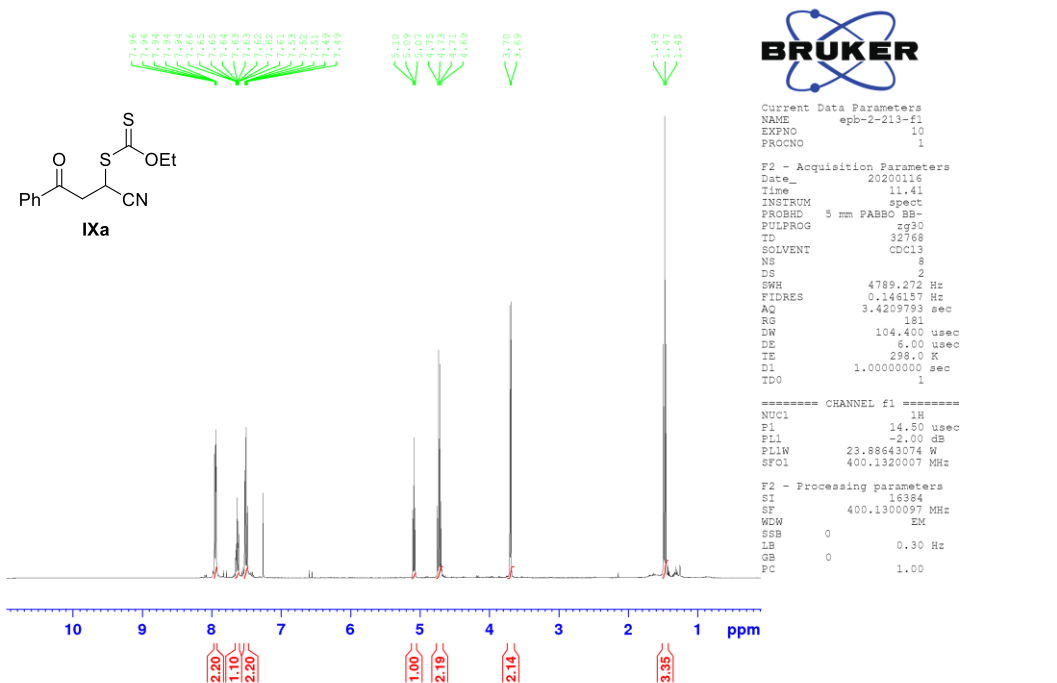
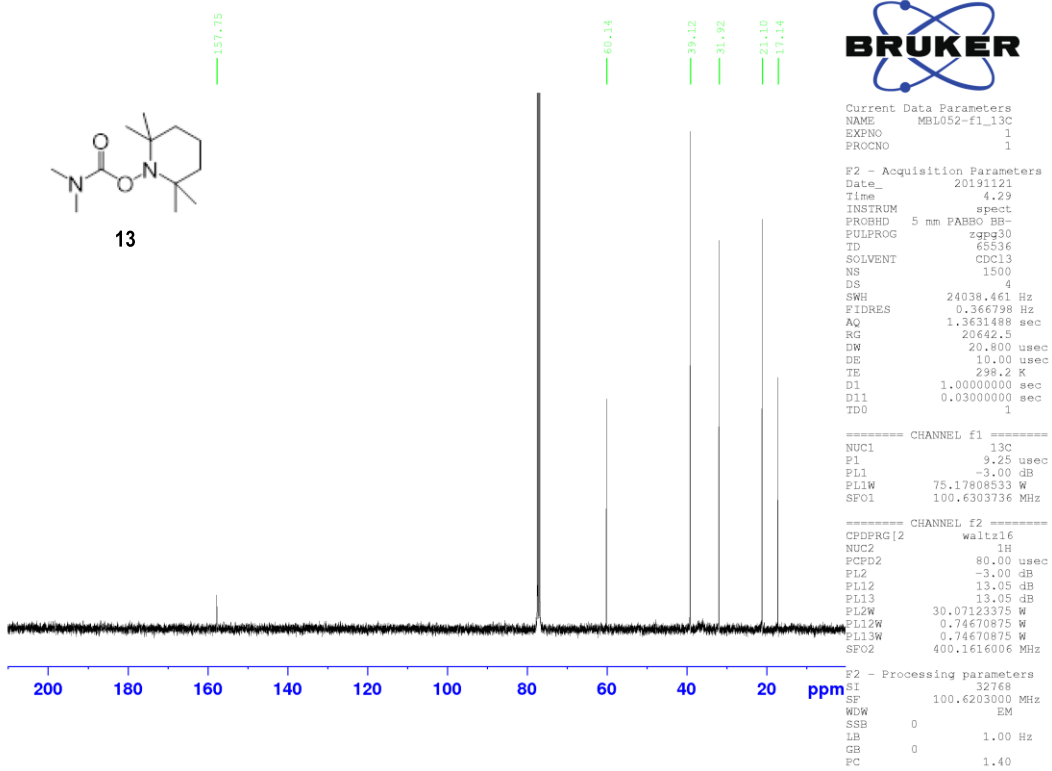
F2 - Acquisition Parameters
 Date_ 20191115
 Time 14.35
 INSTRUM FOURIER300
 PROBHD 5 mm DUL 13C-1
 PULPROG zg30
 TD 65536
 SOLVENT CDCl3
 NS 8
 DS 2
 SWH 6103.516 Hz
 FIDRES 0.099132 Hz
 AQ 5.3687091 sec
 RG 108.743
 DW 81.920 usec
 DE 6.50 usec
 TE 298.0 K
 D1 1.00000000 sec
 TDO 1

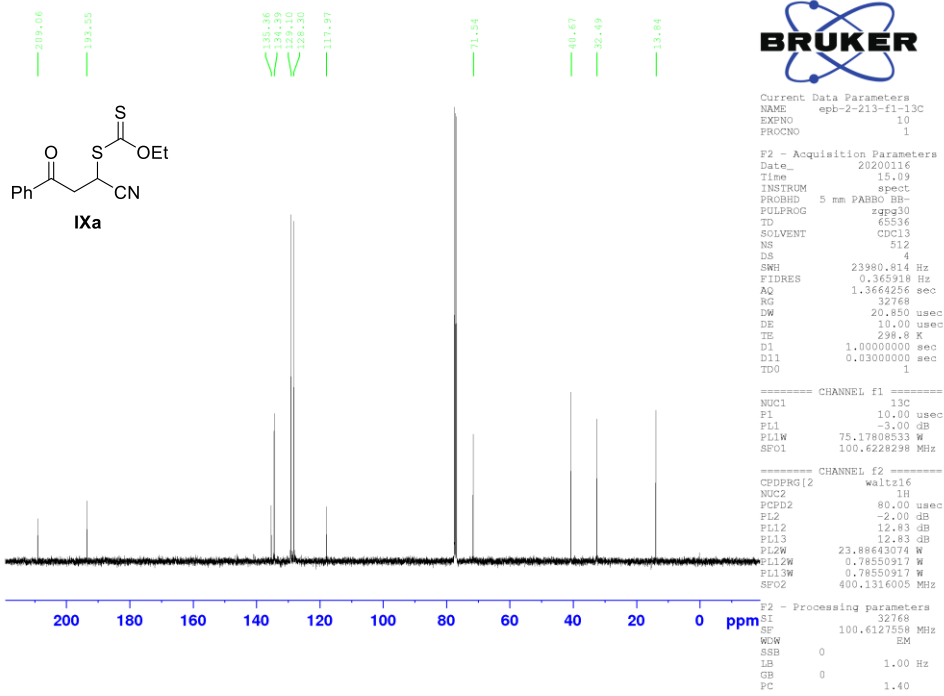
===== CHANNEL f1 =====
 SFO1 300.1818011 MHz
 NUC1 1H
 P1 11.75 usec
 PLW1 20.00000000 W

F2 - Processing parameters
 SI 32768
 SF 300.1800055 MHz
 WDW EM
 SSB 0
 LB 0.30 Hz
 GB 0
 PC 1.40



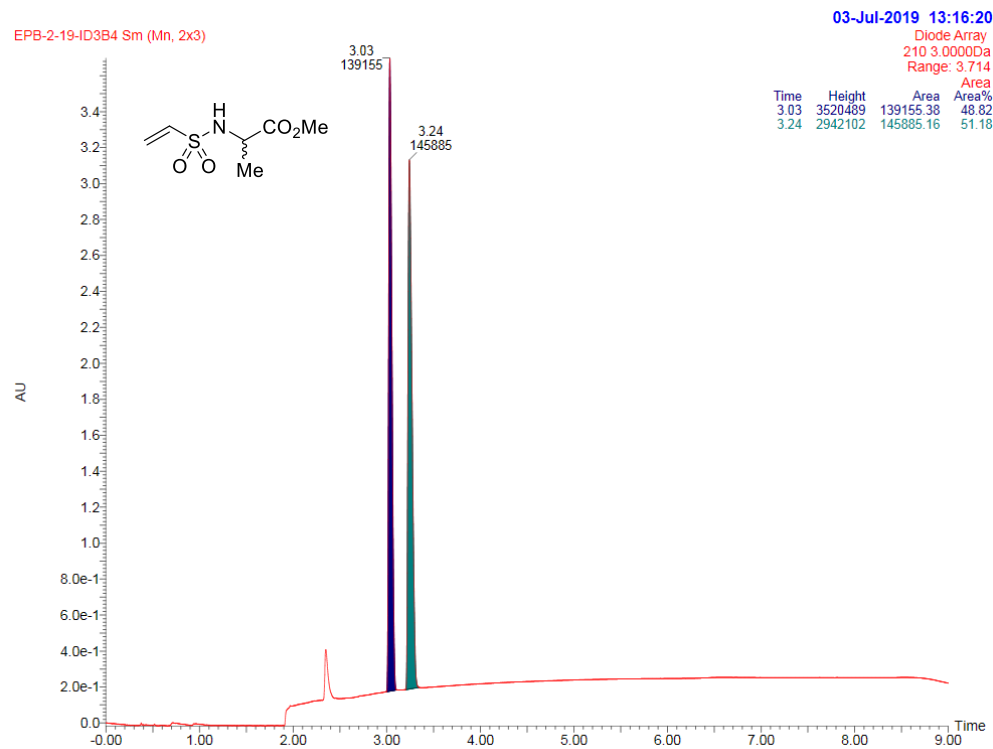




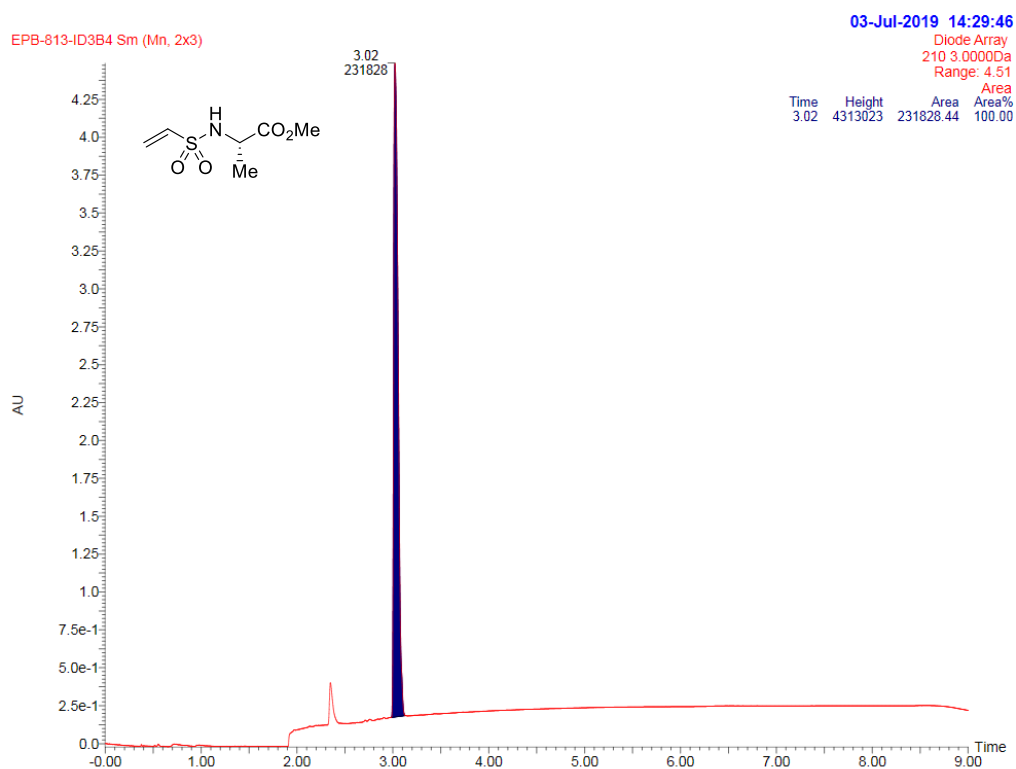


G. UPC² Traces

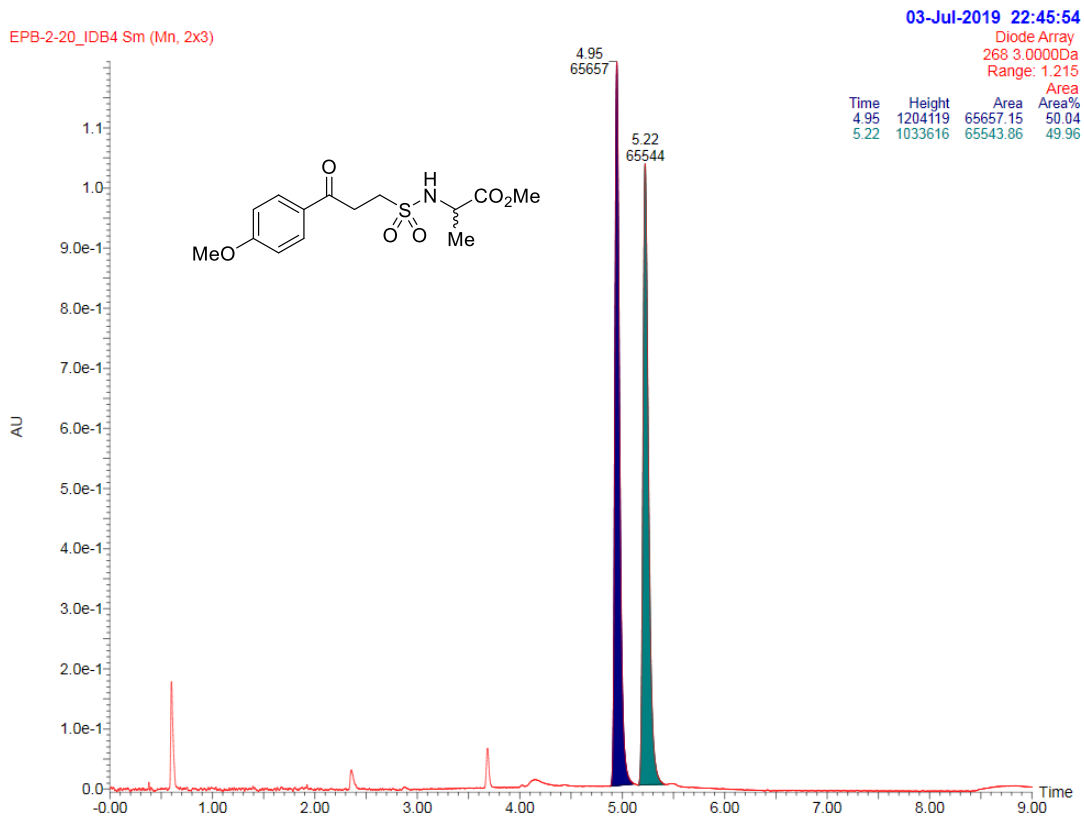
Conditions: UPC2 (Daicel Chiralpak ID-3 column, 20 °C, gradient CO₂/EtOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm).



Conditions: UPC2 (Daicel Chiralpak ID-3 column, 20 °C, gradient CO₂/EtOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, $\lambda = 220$ nm).



Conditions: UPC2 (Daicel Chiralpak ID-3 column, 20 °C, gradient CO₂/EtOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, λ = 268 nm).



Conditions: UPC2 (Daicel Chiralpak ID-3 column, 20 °C, gradient CO₂/EtOH from 100% CO₂ to 60:40 over 4 minutes, curve 6, flow rate 3 mL/min, λ = 268 nm).

