

Alma Mater Studiorum Università di Bologna
Archivio istituzionale della ricerca

Impact of Triflumuron on Halyomorpha halys (Hemiptera: Pentatomidae): laboratory and field studies

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Masetti A., Depalo L., Pasqualini E. (2021). Impact of Triflumuron on Halyomorpha halys (Hemiptera: Pentatomidae): laboratory and field studies. JOURNAL OF ECONOMIC ENTOMOLOGY, 114(4), 1709-1715 [10.1093/jee/toab102].

Availability:

This version is available at: <https://hdl.handle.net/11585/868626> since: 2022-02-25

Published:

DOI: <http://doi.org/10.1093/jee/toab102>

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>).
When citing, please refer to the published version.

(Article begins on next page)

Impact of triflumuron on Halyomorpha halys (Hemiptera: Pentatomidae): laboratory and field studies

Journal:	<i>Journal of Economic Entomology</i>
Manuscript ID	ECONENT-2020-1007.R2
Manuscript Type:	Research
Date Submitted by the Author:	n/a
Complete List of Authors:	Masetti, Antonio; Università of Bologna, Dipartimento di Scienze e Tecnologie Agro-Alimentari Depalo, Laura; Università di Bologna, Dipartimento di Scienze e Tecnologie Agro-Alimentari Pasqualini, Edison; Università di Bologna, Dipartimento di Scienze e Tecnologie Agro-Alimentari
Please choose a section from the list:	Horticultural Entomology
Field Keywords:	Chemical Control, Fruit Tree Entomology, Insecticide Testing, Invasive Species, IPM
Organism Keywords:	Pentatomidae

SCHOLARONE™
Manuscripts

**Impact of triflumuron on *Halyomorpha halys* (Hemiptera: Pentatomidae):
laboratory and field studies**

Antonio Masetti*, Laura Depalo and Edison Pasqualini

Dipartimento di Scienze e Tecnologie Agro-Alimentari, *Alma mater studiorum* - Università di
Bologna, v.le G. Fanin, 42 – 40127 Bologna, Italy

* Corresponding author, E-mail: antonio.masetti@unibo.it

10 **Abstract**

11 *Halyomorpha halys*, (the brown marmorated stink bug, BMSB), is a high-concern invasive species
12 causing severe damage to orchards in many countries outside its native Asian range. Control
13 options matching both effectiveness and sustainability are currently lacking. Inhibitors of chitin
14 biosynthesis might be exploited for integrated management programs because of the overall better
15 ecotoxicological profile in comparison with most neurotoxic insecticides used so far against BMSB.
16 In this study, the activity of triflumuron, a benzoylphenyl urea hampering chitin biosynthesis, was
17 tested on BMSB in laboratory and field conditions. In laboratory bioassays, the insecticide was
18 sprayed on potted peach plants (30 cm high) and residues were aged in a glasshouse for 0, 7, 14 and
19 21 days. Then 3rd instar bugs were placed on the plants and continuously exposed to residues.
20 Mortality was scored after 7, 14 and 21 day exposure. Triflumuron caused significantly higher
21 mortality on BMSB nymphs in comparison with water controls at all aging periods. Moreover,
22 aging of residues up to 21 days did not cause any significant reduction of activity. Field
23 experiments were also carried out in 2019 in eight pear orchards. Injuries to fruits at harvest were
24 compared between plots where triflumuron was added to insecticide sprays against BMSB and
25 control plots managed exactly in the same way but without any triflumuron treatment. An overall
26 mean of $9.99 \pm 1.98\%$ stink bug injured fruits was detected in plots managed with the strategy
27 including triflumuron, whereas $19.45 \pm 3.55\%$ of fruits were injured in plots assigned to controls.

28

29 **Keywords:** Brown marmorated stink bug; Invasive species; Inhibitors of chitin biosynthesis;
30 Integrated pest management; Reduced-risk insecticides.

31

32 **Introduction**

33 The brown marmorated stink bug (BSMB) - *Halyomorpha halys* (Stål) (Hemiptera: Pentatomidae) -
34 is a serious agricultural pest in most countries where it has been accidentally introduced (Bosco et
35 al. 2017, Leskey and Nielsen 2018). Current management strategies rely mostly on neurotoxic
36 broad-spectrum insecticides that are quite effective in preventing BMSB damages but have a short
37 residual activity (Kuhar and Kamminga 2017). This forces farmers to increase the frequency of
38 insecticide sprays (Leskey et al. 2012) causing the disruption of previously established integrated
39 pest management (IPM) programs and the resurgence of secondary pests (Leskey and Nielsen
40 2018). The identification of more selective insecticides targeting BMSB would significantly reduce
41 the environmental issues associated to its control and could contribute to IPM principles. Besides
42 efficacy on BMSB and less severe side effects, residual activity of insecticides could play a key role
43 in restoring sustainable management programs.

44 Benzoylphenyl ureas (BPUs) are classified by Insecticide Resistance Action Committee (IRAC,
45 <https://www.irac-online.org/modes-of-action>) into group 15 (Inhibitors of chitin biosynthesis, type
46 0). These compounds interfere with chitin biosynthesis, thus hindering insect molts. Detrimental
47 effects on fecundity and egg viability are reported as well (Pener and Dhadialla 2012). Although
48 BPUs have been used as pesticides since the 1970s (Retnakaran and Wright 1987, Spomer and
49 Sheets 2019), the exact mode of action is still quite obscure (Zhu et al. 2016). However, Douris et
50 al. (2016) shed some light on the molecular targets of BPUs by demonstrating a direct interaction
51 with the enzyme chitin synthase 1. These insecticides are persistent and do not show cross-
52 resistance with neurotoxic active ingredients (Doucet and Retnakaran 2012, Arruda et al. 2020).
53 Moreover, BPUs are considered less harmful to beneficial insects and non-target organisms than
54 most nerve poisons (Sun et al. 2015). For these reasons, BPUs are desirable for inclusion in IPM
55 programs in several cropping systems (Dhadialla et al. 2005).

56 Several studies focused on the effects of a number of insecticides on BMSB, but the activity of

BPU has not been investigated extensively. Kamminga et al. (2012) reported promising effects of diflubenzuron and novaluron on BMSB nymphs in laboratory assays carried out in Petri dishes, no more investigations were undertaken. The first objective of this study was to evaluate the activity of triflumuron (the only BPU currently allowed on orchards in EU) on BMSB nymphs by means of laboratory assays. The effects of aged residues were also investigated because triflumuron is known to be persistent on vegetation (Marx 1977, Aplada-Sarlis et al. 1999) and this could be a crucial factor to reduce the frequency of insecticide sprays (Kuhar and Kamminga 2017). Potted peach plants were used in the experiments because this is more representative of real conditions for residues aging. Moreover, the use of living plants allowed a prolonged exposure of insects, which is needed because of the mode action of BPUs (Dhadialla et al. 2005). A second aim of the study was to test, by mean of a field experiment set up in 2019 on pear orchards, if the addition of triflumuron to IPM strategies against BMSB could lead to a decrease in injuries to fruit.

Materials and methods

Insects

BMSB nymphs were obtained from a laboratory colony established in 2015 at the Department of Agricultural and Food Sciences (University of Bologna). Bugs were reared in a walk-in climatic chamber at $25^{\circ}\text{C} \pm 2^{\circ}\text{C}$, 50%–70% RH, with a 14 L:10 D photoperiod. Adults and nymphs were fed twice per week with green beans, carrots and soybean seeds; fresh fruits (pears, apples or kiwi fruits) were provided weekly to adults only.

Insecticide treatments

The insecticide Alsystin 48SC (triflumuron 39.34% = 480 g L⁻¹) was provided by Bayer Crop Sciences (Milan, Italy) and applied at 25 mL hL⁻¹, the maximum field recommended concentration for pome and stone fruits. Potted peach plants (GF-677 rootstock, approximately 30 cm high) were

sprayed using a hand sprayer up to dripping and allowed to dry completely before trials. Control plants were sprayed in the same way with tap water.

Laboratory bioassays

From July 2017 to November 2018, four aging periods (0, 7, 14 and 21 days) of insecticide residues were investigated, each in a separate experiment. Twelve experimental units, each consisting of a potted peach plant, were arranged for each aging period. Six plants were treated with triflumuron and the others, which were assigned to control, with tap water. Overall, 48 experimental units were set up and 480 BMSB nymphs were tested.

Once sprayed outdoor, plants were transferred into a glasshouse box where residues of the treatments aged. Here the plants were maintained for 7, 14 or 21 days at the following conditions:

T_{\max} 30° C, T_{\min} 15° C, 16 L:8 D photoperiod, minimum artificial lighting in photophase 70,000 lx.

In the experiment of 0-day aging of residues, the peach plants were used immediately after treatment droplets dried.

To set up experimental units, potted plants were placed in plexiglas cylinders (diameter 8 cm, height 30 cm), sealed with a fine net on the top and with a hole (diameter 8 cm) closed with a fine net on the base to allow air circulation and avoid mold growth. The soil surface on the pot was covered with nonwoven fabric, which was tightly wrapped to plant stem. Ten BMSB early 3rd instar nymphs were placed in each cylinder. Given that BMSB nymphs must feed on fruit or vegetables to complete development, carrots and green beans were provided ad libitum and changed twice per week. The green beans were hung on the plants to force the nymphs to climb on canopy in order to feed (Supplementary data Fig. SD1).

Experimental units were held at the same conditions reported for BMSB rearing and mortality was checked after 7, 14 and 21 days of continuous exposure of nymphs to residues. Besides the number of dead nymphs also their instars, and therefore the number of molts they successfully completed,

was recorded. Moribund nymphs (i.e. unable to upright themselves when flipped on their back) found at the last exposure interval were reared individually until they died or reached the adult stage (recorded as a recovery). Nymphs were held in ventilated 200-mL plastic jars at the same conditions and with the same food provisions as previously reported.

Field trials

From 21 June to 5 September 2019, field experiments were carried out in eight pear orchards located in Northern Italy, which suffered severe damages by BMSB in the previous years. Each orchard was considered as an experimental block and two contiguous plots (> 0.5 ha) were delimited. One plot was managed according to IPM strategies (here after referred as “farmer’s strategy”) recommended in Northern Italy. The other plot of each orchard underwent the same management practices as the former, but a treatment with Alsystin 48 SC at 25 mL hL^{-1} was also added (hereafter reported as triflumuron strategy). This treatment was carried out between 21 and 30 June targeting the first-generation nymphs of the BMSB. Of course, no triflumuron treatment was allowed in plots managed with farmer’s strategy. The sprays of broad-spectrum insecticides, which included chlorpyrifos-methyl, acetamiprid and pyrethroids (deltamethrin, lambda-cyhalothrin and tau-fluvalinate), varied slightly among orchards, as farmers were allowed to make minor changes in order to adjust for pest pressure and phenology. These sources of variation were accounted in the block factor.

Fruit sampling was carried out in a transect of 3 rows x 15 m. The transects were established in the middle of each plot at least 10 m away from plot edges to avoid border effect from the perimeter of the orchards and to accommodate for drift of triflumuron spray. External injuries were checked at harvesting on a sample of 400 fruits per plot. Two hundred fruits were sampled at eye-level and 200 were sampled above 2.5-m height using a fruit picking ladder, because most severe injuries by BMSB are usually recorded on fruit growing in the upper parts of the trees (Bariselli et al. 2016).

Pears were examined directly on the trees and scored as injured if at least one distinct depression or discoloration on fruit surface compatible with BMSB punctures could be detected (Acebes-Doria et al. 2016). Although the occasional occurrence of Heteropteran bugs other than BMSB could not be ruled out, the overwhelming majority of fruit injuries were likely due to feeding activity by BMSBs. Damages to fruit by Heteropteran bugs have been rarely reported in orchards of Northern Italy and the symptoms used to score injuries to pears are quite specific to BMSB.

Statistical analysis

To assess the activity of triflumuron on BMSB nymphs in laboratory experiments, insecticide-treated plants and relative water controls were compared within each aging period of residues. Data were analyzed by general linear mixed models (GLMM) with first-order autoregressive covariance structure, binomial distribution, probit link function and Kenward–Roger method for estimating degrees of freedom. The number of dead insects out of the total nymphs tested was considered as dependent variable. The treatments (triflumuron and water control) were used as factors, the exposure intervals (7, 14 and 21 days) were included as repeated measures and their interaction (treatment x exposure interval) were tested as well.

To study the possible decrease in the activity of triflumuron residues (i.e. to compare mortality only in insecticide-treated plants among different aging period), nymphs mortality recorded in triflumuron experimental units was corrected by Schneider-Orelli formula (Schneider-Orelli 1947) considering as natural the mean mortality recorded in water controls aged for the same period. Corrected percentages of mortality matched the assumptions of parametric tests and were analyzed by a mixed design ANOVA without any transformation. Aging periods (0, 7, 14 and 21 days) were considered as between-subject factor and exposure intervals (7, 14 and 21) as within-subject factor. The interaction aging period x exposure interval was tested as well.

Pearson χ^2 was used to test the association between the ages of residues and the number of molts that

dead nymphs exposed to triflumuron were able to successfully complete before dying.

A GLMM was carried out to analyze data from the field experiments. Binomial error distribution and log link function were selected, the number of injured fruits out of the total number of fruits checked was considered as dependent variable, treatments (farmer's strategy vs triflumuron strategy) and location of fruits (>2.5 m vs eye level) were considered as fixed factors, and orchards were included as random block factor.

All the analyses were performed using IBM SPSS Statistics (ver. 26).

Results

Laboratory bioassays

GLMMs detected significant effects on mortality by both main factors (i.e., treatment and exposure interval) for all aging periods of residues. Whereas the interaction treatment * exposure interval was significant only for 7- and 14 -day aged residues (Table 1). In other word, triflumuron residues caused overall higher mortality than water controls and longer exposure interval increased mortality (Fig. 1). Because of significant interactions, treatments were compared within each of the exposure intervals for 7-and 14-day old residues. For both aging periods, GLMMs did not detect significant difference between triflumuron and control at 7-day exposure but indicated higher mortality in triflumuron than in water at 14- and 21-day exposures (Table 1).

Mortality increased at longer exposure intervals also in the controls with a certain degree of variation among aging periods. Overall, the lowest number of dead nymphs was counted in the controls of residues tested immediately after drying. Whereas for other aging periods the natural mortality ranged from $16.06 \pm 3.99\%$ to $28.60 \pm 9.80\%$, $23.55 \pm 4.99\%$ to $38.14 \pm 7.30\%$ and $30.07 \pm 4.77\%$ to $47.79 \pm 6.61\%$ for 7-, 14- and 21-day exposure, respectively.

The comparison of corrected percentages of mortality did not show any significant effect of aging periods on the activity of triflumuron residues ($F_{(3; 20)} = 1.34$; $p = 0.29$). The interaction aging

period * exposure interval was not significant too ($F_{(6, 40)} = 1.32$; $p = 0.27$). Therefore, the aging of residues up to 21 days did not significantly decrease the activity of the insecticide.

The number of molts that nymphs exposed to triflumuron were able to complete before dying did not differ among aging periods ($\chi^2 = 6.34$; $df = 6$; $p = 0.39$). Therefore, the ages of residues did not significantly affect the insecticide speed of action. Pooling all aging periods, 29.70% of BMSB nymphs died without completing any molt, 57.43% of individuals successfully molted from 3rd to 4th instar but died before or during the next molt, 12.87% reached the 5th instar but failed to cast the exuviae and died during the molt to adult stage (Supplementary data Fig. SD2).

Field trials

The addition of triflumuron to insecticide treatments allowed in IPM strategy for BMSB led to a significant decrease in the percentage of injured fruits ($F_{(1; 28)} = 106.93$; $p < 0.001$). Fruit location had also a significant effect ($F_{(1; 28)} = 137.92$; $p < 0.001$), and the percentage of injured fruits was higher above 2.5 m than at eye level. The interaction strategy * fruit location was not significant ($F_{(1; 28)} = 0.81$; $p = 0.78$). A mean of $9.99 \pm 1.98\%$ injured fruits was detected in plots managed with triflumuron strategy, whereas $19.45 \pm 3.55\%$ of fruits were injured in plots assigned to farmer's strategy. Overall, $20.19 \pm 3.42\%$ of pears showed symptoms of feeding activity by BMSB above 2.5 m. This percentage dropped to 9.25 ± 1.95 at eye level (Fig. 2).

Discussion

Triflumuron residues on potted peach plants showed a significant activity on BMSB nymphs that died increasingly between progressive nymphal instars. These results agreed with laboratory assays by Kamminga et al. (2012) who reported 90% and 65% mortality of BMSB nymphs fed for 7 days with fresh beans dipped in solutions of novaluron and diflubenzuron, respectively. Significant reduction in the abundance of BMSB and other stink bugs was also described for diflubenzuron

207 applications to soybean in open field conditions (Herbert et al. 2013). On the other hand, Gradish et
208 al. (2019) found neither contact nor residual toxicity of novaluron on BMSB nymphs. However, in
209 this study, mortality was checked only in the short term, whereas prolonged exposure to BPU
210 residues is necessary to detect activity of this group of insecticide (Dhadialla et al. 2005).

211 The activity of triflumuron on BMSB nymphs persisted up to 21 days of residue aging in
212 glasshouse. Overall, the mortality of bugs exposed to the insecticide was higher than in controls for
213 all ages of residues. Moreover, no significant decrease in mortality could be detected on insecticide-
214 treated plants at increasing periods of residues aging. The environmental persistence of BPUs has
215 been well documented and the slow decrease in residue concentration of triflumuron over time
216 (Bayer Crop Sciences, unpublished data) is in line with the steady insecticidal activity of this active
217 ingredient up to the longest aging period. Given that treated plants were maintained in a glasshouse,
218 the only relevant insecticide degradation factor was photolysis. Rainfall, which was not considered
219 in this study, seems indeed a minor issue as BPUs are known to adhere to plant waxes and to
220 undergo to slight wash off (Retnakaran and Wright 1987, Doucet and Retnakaran 2012).

221 A relatively high mortality in the controls of some aging periods was related to the long-term
222 exposure used in the bioassays. Fluctuations in nymphal survival has been often observed in BMSB
223 laboratory colonies. Our findings were in line with Medal et al. (2013) who reported a juvenile
224 mortality from 23% to 50% in BMSB rearing and with Fisher et al. (2020) who recorded overall
225 low nymphal survival when testing the influence of temperature and humidity on BMSB.

226 Progressive poisoning through tarsal adsorption seemed the main way of insecticide contamination
227 because most of BMSB nymphs did not die at the first molt, but many insects completed one or
228 even two molts before succumbing. Ingestion has been usually considered as the major way of
229 uptake for BPUs, but slow penetration through cuticle was also described (Hammann and
230 Sirrenberg 1980, Retnakaran and Wright 1987, Sun et al. 2015). This second way of triflumuron
231 uptake was likely prevalent in our experiments, given that untreated food was used and the bug

232 probing on treated peach plants was negligible.

233 Given that intoxicated nymphs could survive for a couple of weeks, the damage they may cause

234 before succumbing needs to be taken into account. In this context, sub lethal effects may be worth

235 of consideration as secondary impacts on pest population (Cira et al. 2017, Depalo et al. 2017). In

236 particular, BPUs could reduce hatch rate, impair feeding activity and increase susceptibility to

237 pathogens on intoxicated BMSB (Catchot et al. 2021).

238 Early BMSB instars are more susceptible to insecticides than adults and several insecticides have

239 demonstrated a significant activity on nymphs (Kuhar and Kamminga 2017, Gradish et al. 2019).

240 Mostly broad-spectrum neurotoxic active ingredients have been so far investigated and, in

241 comparison with these insecticides, BPUs are more compatible with IPM (Dhadialla et al. 2005,

242 Doucet and Retnakaran 2012). Nevertheless, BPUs do not lack any impact on beneficials. Lethal

243 effects have been found on nymphs of predatory bugs (Soares et al. 2019) and on larvae of

244 ladybeetles (Cabrera et al 2017). Some sub-lethal effects have been reported on adult parasitoids

245 either after direct contact with residues (Matioli et al. 2019) or when parasitized eggs of the hosts

246 were treated (Goulart et al. 2012). Therefore, the possible detrimental effects of BPUs on natural

247 enemies of BMSB seem worth of investigations to draw a comprehensive picture of the possible

248 role of these insecticides in IPM scenario.

249 Besides the environmental side effects and the disruption of IPM practices, the overuse of a few

250 neurotoxic insecticides to control BMSB often impaired the rotation of insecticide chemistries upon

251 which insecticide resistance management strategy should rely (Hurley and Mitchell 2014). The

252 potential for insecticide-resistance development in BMSB populations has not been established yet,

253 but it would be wise to avoid over-reliance on neurotoxic active ingredients (Alford et al. 2020).

254 From this standpoint, the inclusion of BPU in IPM practices could help to restore active ingredient

255 rotation as a strategy to delay the insurgence of resistance (Spomer and Sheets 2019).

256 The long-lasting activity of triflumuron, and possibly other BPU, could be exploited to reduce

257 damages by BMSB nymphs without the need of frequent treatments. The short residual activity,
258 which lasts less than three days for most insecticides used against BMSB (Leskey et al. 2014), is
259 widely recognized as a major problem (Kuhar and Kamminga 2017). This pest is polyphagous and
260 very mobile; both nymphs and adults have big dispersal abilities and move across the landscape
261 following phenology of crops and uncropped plants (Martinson et al. 2015, Hamilton et al. 2018).
262 Repeated and unpredictable infestations of orchards are common and frequent sprays of short-
263 lasting insecticides are needed.

264 The field experiments, which showed a significant decrease in the percentages of injured pears
265 where triflumuron was added, reinforced the findings of laboratory assays. Field trials pointed out
266 also that in 2019 strategies mainly relying on neurotoxic active ingredients did not achieve a
267 satisfactory control of BMSB. The mean percentage of injured fruit recorded in plots managed by
268 farmer's strategy (19.45 ± 3.55) far exceeded the economic threshold. Although the percentage of
269 injured fruits dropped to 9.99 ± 1.98 in the plots where triflumuron was added, this was still not
270 completely acceptable from an economic standpoint. A refinement of management strategies for
271 BMSB seems therefore urgently needed and the development of treatment thresholds for this stink
272 bug would be a relevant step forward (Short et al. 2017).

273 BPU's lack any lethal effect on adult insects and have no chance to decrease the pressure exerted by
274 highly damaging adult bugs that migrate into crops from surrounding areas (Acebes-Doria et al.
275 2016). Therefore, triflumuron and other BPU's might be considered for field applications only as a
276 tool in a wider management strategy. Control methods aimed at preventing the invasion of crops by
277 adult bugs such as exclusion nets (Candian et al. 2018), insecticide netting screens and sprays at
278 orchard perimeter (Blaauw et al. 2015) might take major benefits from few and long-lasting BPU
279 treatments aimed decreasing nymphal populations and hampering reproduction cycle that can occur
280 within the orchards.

281

282 Acknowledgments

283 We are grateful to Elisa Marchetti and Andrea Angelici for technical assistance in BMSB rearing
284 and in bioassays. We also thank personnel from Bayer Crop Science for help in fruit samplings.
285 Part of this study was supported by Bayer Crop Science.

286

287

288 References Cited

- 289 **Acebes-Doria, A. L., T. C. Leskey, and Bergh, J. C. 2016.** Injury to apples and peaches at harvest
290 from feeding by *Halyomorpha halys* (Stål) (Hemiptera: Pentatomidae) nymphs early and
291 late in the season. Crop Prot. 89: 58-65.
- 292 **Alford A., T. P. Kuhar, G. C. Hamilton, P. Jentsch, G. Krawczyk, J. F. Walgenbach, and C.**
293 **Welty. 2020.** Baseline toxicity of the insecticides bifenthrin and thiamethoxam on
294 *Halyomorpha halys* (Hemiptera: Pentatomidae) collected from the eastern United States. J.
295 Econ. Entomol. 113: 1043-1046.
- 296 **Aplada-Sarlis, P. G., G. E. Miliadis, and N. G.Tsiropoulos. 1999.** Dissipation of teflubenzuron
297 and triflumuron residues in field-sprayed and cold-stored pears. J. Agric. Food. Chem. 47:
298 2926-2929.
- 299 **Arruda, L. S., A. R. Rodrigues, N. C. Bermudez, L. M. Ribeiro, J. E. L. Neto, and H. A.**
300 **Siqueira. 2020.** Field resistance of *Plutella xylostella* (Lepidoptera: Plutellidae) to
301 lufenuron: Inheritance and lack of cross-resistance to methoxyfenozide. Crop. Prot. 136:
302 article 105237.
- 303 **Bariselli, M., R. Bugiani, and L. Maistrello. 2016.** Distribution and damage caused by
304 *Halyomorpha halys* in Italy. EPPO Bull. 46: 332-334.
- 305 **Bosco, L., S. T. Moraglio, and L. Tavella. 2017.** *Halyomorpha halys*, a serious threat for hazelnut
306 in newly invaded areas. J. Pest. Sci. 91: 661–670.

- 307 **Blaauw, B. R., D. Polk, and A. L. Nielsen. 2015.** IPM-CPR for peaches: incorporating
308 behaviorally-based methods to manage *Halyomorpha halys* and key pests in peach. *Pest.*
309 *Man. Sci.* 71: 1513-1522.
- 310 **Cabrera, P., D. Cormier, and É. Lucas. 2017.** Differential sensitivity of an invasive and an
311 indigenous ladybeetle to two reduced-risk insecticides. *J. Appl. Entomol.* 141: 690–701.
- 312 **Candian, V., M. G. Pansa, R. Briano, C. Peano, R. Tedeschi, and L. Tavella. 2018.** Exclusion
313 nets: a promising tool to prevent *Halyomorpha halys* from damaging nectarines and apples
314 in NW Italy. *Bull. Insectol.* 71: 21-30.
- 315 **Catchot, B. D., F. R. Musser, J. Gore, N. Krishnan, D. R. Cook, S. D. Stewart, G. M. Lorenz,**
316 **S. Brown, N. Seiter, A. L. Catchot, D. L. Kerns, R. Jackson, and K. S. Knighten. 2021.**
317 Sublethal impacts of novaluron on Tarnished Plant Bug (Hemiptera: Miridae) adults. *J.*
318 *Econ. Entomol.* 114: 739-746.
- 319 **Cira, T. M., E. C. Burkness, R. L. Koch, and W. D. Hutchison. 2017.** *Halyomorpha halys*
320 mortality and sublethal feeding effects following insecticide exposure. *J. Pest. Sci.* 90:
321 1257–1268.
- 322 **Depalo, L., A. Lanzoni, A. Masetti, E. Pasqualini, and G. Burgio. 2017.** Lethal and sub-lethal
323 effects of four insecticides on the aphidophagous coccinellid *Adalia bipunctata* (Coleoptera:
324 Coccinellidae). *J. Econ. Entomol.* 110: 2662-2671.
- 325 **Dhadialla, T. S., A. Retnakaran, and G. Smagghe. 2005.** Insect growth and development
326 disrupting insecticides. *In* L. I. Gilbert, K. Iatrou, and S. S. Gill (eds.), *Comprehensive*
327 *insect molecular science*, vol 6. Pergamon Press, Oxford, pp 55–100
- 328 **Doucet, D., and A. Retnakaran. 2012.** Insect chitin: metabolism, genomics and pest management.
329 *In* T. S. Dhadialla (ed.), *Advances in insect physiology - Insect growth disruptors*, vol 43.
330 Elsevier, Oxford, pp 437-511.

- 331 **Douris, V., D. Steinbach, R. Panteleri, I. Livadaras, J. A. Pickett, T. Van Leeuwen, R. Nauen,**
332 **and J. Vontas. 2016.** Resistance mutation conserved between insects and mites unravels
333 the benzoylurea insecticide mode of action on chitin biosynthesis. *Proc. Natl. Acad. Sci.*
334 U.S.A. 113: 14692-14697.
- 335 **Fisher, J. J., R. P. Rijal, and F. G. Zalom. 2020** Temperature and humidity interact to influence
336 Brown Marmorated Stink Bug (Hemiptera: Pentatomidae), survival. *Environ. Entomol.* (IN
337 PRESS, <https://doi.org/10.1093/ee/nvaa146>).
- 338 **Goulart, R. M., H. X. Volpe, A. M. Vacari, R. T. Thuler, and S. A. De Bortoli. 2012.** Insecticide
339 selectivity to two species of *Trichogramma* in three different hosts, as determined by
340 IOBC/WPRS methodology. *Pest Man. Sci.* 68: 240-244.
- 341 **Gradish, A. E., H. Fraser and C. D. Scott-Dupree. 2019.** Direct and residual contact toxicity of
342 insecticides to *Halyomorpha halys* (Hemiptera: Pentatomidae). *Can. Entomol.* 151: 209-218.
- 343 **Hamilton, G. C., J. J. Ahn, W. Bu, T. C. Leskey, and A. L. Nielsen. 2018.** *Halyomorpha halys*
344 (Stål). In J. E. McPherson (ed.), *Invasive stink bugs and related species (Pentatomoidea):*
345 *biology, higher systematics, semiochemistry, and management.* CRC Press Taylor & Francis
346 Group, Boca Raton, pp 243- 292.
- 347 **Hammann, I., and W. Sirrenberg. 1980.** Laboratory evaluation of SIR 8514, a new chitin
348 synthesis inhibitor of the benzoylated urea class. *Pflanzenschutz-Nachrichten Bayer* 33: 1-
349 34.
- 350 **Herbert, D. A., S. Malone, M. Arrington, and J. Hogue. 2013.** Evaluation of selected foliar
351 insecticides for control of stink bug in soybean, 2012. *Arthropod. Man. Tests* 38
- 352 **Hurley, T. M., and P. D. Mitchell. 2014.** Insect resistance management: Adoption and
353 compliance. In D.W. Onstad (ed.), *Insect Resistance Management.* Academic Press, pp 421-
354 451.

- 355 **Kamminga, K. L., T. P. Kuhar, A. Wimer, and D. A. Herbert. 2012.** Effects of the insect growth
356 regulators novaluron and diflubenzuron on the brown marmorated stink bug. *Plant Health*
357 *Progr.* 13, 2. (doi:10.1094/PHP-2012-1212-01-RS)
- 358 **Kuhar, T. P., and K. L. Kamminga. 2017.** Review of the chemical control research on
359 *Halyomorpha halys* in the USA. *J. Pest. Sci.* 90: 1021-1031.
- 360 **Leskey, T. C., and A. L. Nielsen. 2018.** Impact of the invasive brown marmorated stink bug in
361 North America and Europe: history, biology, ecology, and management. *Annu. Rev.*
362 *Entomol.* 63: 599-618.
- 363 **Leskey, T. C., B. D. Short, B. R. Butler, and S. E. Wright. 2012.** Impact of the invasive brown
364 marmorated stink bug, *Halyomorpha halys* (Stål), in mid-Atlantic tree fruit orchards in the
365 United States: case studies of commercial management. *Psyche* 2012: 1–14
366 (doi:10.1155/2012/535062)
- 367 **Leskey, T. C., B. D. Short, and D. H. Lee. 2014.** Efficacy of insecticide residues on adult
368 *Halyomorpha halys* (Stål) (Hemiptera: Pentatomidae) mortality and injury in apple and
369 peach orchards. *Pest. Man. Sci.* 70: 1097-1104.
- 370 **Martinson, H. M., P. D. Venugopal, E. J. Bergmann, P. M. Shrewsbury, and M. J. Raupp.**
371 **2015.** Fruit availability influences the seasonal abundance of invasive stink bugs in
372 ornamental tree nurseries. *J. Pest. Sci.* 88: 461–468.
- 373 **Marx, J. L. 1977.** Chitin synthesis inhibitors: new class of insecticides. *Science* 197: 1170-1172.
- 374 **Matioli, T. F., O. Z. Zanardi, and P. T. Yamamoto. 2019.** Impacts of seven insecticides on
375 *Cotesia flavipes* (Cameron) (Hymenoptera: Braconidae). *Ecotoxicol.* 28: 1210-1219.
- 376 **Medal, J., T. Smith, and A. Santa Cruz. 2013.** Biology of the brown marmorated stink bug
377 *Halyomorpha halys* (Heteroptera: Pentatomidae) in the laboratory. *Fla. Entomol.* 96: 1209-
378 1212.

- 379 **Pener, M. P., and T. S. Dhadialla. 2012.** An overview of insect growth disruptors; applied aspects.
380 *In* T. S. Dhadialla (ed.), *Advances in insect physiology - Insect growth disruptors*, vol 43.
381 Elsevier, Oxford, pp 1-162.
- 382 **Retnakaran, A., and J. E. Wright. 1987.** Control of insect pests with benzoylphenyl ureas. *In* J. E.
383 Wright, and A. Retnakaran (eds.), *Chitin and benzoylphenyl ureas. Series Entomologica*, vol
384 38. Springer, Dordrecht, pp 205-282.
- 385 **Schneider-Orelli, O. 1947.** Entomologisches praktikum: Einführung in die land-und
386 forstwirtschaftliche Insektenkunde. Sauerländer.
- 387 **Short, B. D., A. Khrimian, and T. C. Leskey. 2017.** Pheromone-based decision support tools for
388 management of *Halyomorpha halys* in apple orchards: development of a trap-based
389 treatment threshold. *J. Pest. Sci.* 90: 1191–1204.
- 390 **Soares, M. A., L. C. Passos, M. R. Campos, L. J. Collares, N. Desneux, and G. A. Carvalho.**
391 **2019.** Side effects of insecticides commonly used against *Tuta absoluta* on the predator
392 *Macrolophus basicornis*. *J. Pest Sci.* 92: 1447-1456.
- 393 **Spomer, N. A., and J. J. Sheets. 2019.** Chitin biosynthesis and inhibitors. *In* P. Jeschke, M.
394 Witschel, W. Krämer, and S. Ulrich, S. (eds.) *Modern crop protection compounds vol. 3:*
395 *Insecticides*. Wiley-VCH Verlag GmbH & Co. Weinheim, pp 1067-1085.
- 396 **Sun, R., C. Liu, H. Zhang, and Q. Wang. 2015.** Benzoylurea chitin synthesis inhibitors. *J. Agric.*
397 *Food. Chem.* 63: 6847-6865.
- 398 **Zhu, K. Y., H. Merzendorfer, W. Zhang, J. Zhang, and S. Muthukrishnan. 2016.** Biosynthesis,
399 turnover, and functions of chitin in insects. *Annu. Rev. Entomol.* 61: 177-196.

400 **Figure Caption**

401 **Fig. 1.** Mortality of *Halyomorpha halys* nymphs exposed for 7, 14 and 21 days to residues of
402 triflumuron (black line) or control (gray line) on potted peach plants. Each panel represent a
403 different aging period of residues in glasshouse conditions. Vertical bars indicate standard errors of
404 the means. Asterisks indicate significant differences between treatments, which were compared
405 within exposure intervals, in the case of significant interaction of treatment * exposure interval
406 detected by GLMM.

407

408 **Fig. 2.** Percentages of fruit with feeding injuries by *Halyomorpha halys* recorded in 2019 in pear
409 orchards in Northern Italy. GLMM detected significant differences between strategies ($p<0.001$)
410 and between fruit locations ($p<0.001$). Vertical bars indicate standard errors of the means.

**Impact of triflumuron on *Halyomorpha halys* (Hemiptera: Pentatomidae):
laboratory and field studies**

Antonio Masetti*, Laura Depalo and Edison Pasqualini

Dipartimento di Scienze e Tecnologie Agro-Alimentari, *Alma mater studiorum* - Università di
Bologna, v.le G. Fanin, 42 – 40127 Bologna, Italy

* Corresponding author, E-mail: antonio.masetti@unibo.it

Abstract

Halyomorpha halys, (the brown marmorated stink bug, BMSB), is a high-concern invasive species causing severe damage to orchards in many countries outside its native Asian range. Control options matching both effectiveness and sustainability are currently lacking. Inhibitors of chitin biosynthesis might be exploited for integrated management programs because of the overall better ecotoxicological profile in comparison with most neurotoxic insecticides used so far against BMSB. In this study, the activity of triflumuron, a benzoylphenyl urea hampering chitin biosynthesis, was tested on BMSB in laboratory and field conditions. In laboratory bioassays, the insecticide was sprayed on potted peach plants (30 cm high) and residues were aged in a glasshouse for 0, 7, 14 and 21 days. Then 3rd instar bugs were placed on the plants and continuously exposed to residues. Mortality was scored after 7, 14 and 21 day exposure. Triflumuron caused significantly higher mortality on BMSB nymphs in comparison with water controls at all aging periods. Moreover, aging of residues up to 21 days did not cause any significant reduction of activity. Field experiments were also carried out in 2019 in eight pear orchards. Injuries to fruits at harvest were compared between plots where triflumuron was added to insecticide sprays against BMSB and control plots managed exactly in the same way but without any triflumuron treatment. An overall mean of $9.99 \pm 1.98\%$ stink bug injured fruits was detected in plots managed with the strategy including triflumuron, whereas $19.45 \pm 3.55\%$ of fruits were injured in plots assigned to controls.

Keywords: Brown marmorated stink bug; Invasive species; Inhibitors of chitin biosynthesis; Integrated pest management; Reduced-risk insecticides.

32 **Introduction**

33 The brown marmorated stink bug (BMSB) - *Halyomorpha halys* (Stål) (Hemiptera: Pentatomidae) -
34 is a serious agricultural pest in most countries where it has been accidentally introduced (Bosco et
35 al. 2017, Leskey and Nielsen 2018). Current management strategies rely mostly on neurotoxic
36 broad-spectrum insecticides that are quite effective in preventing BMSB damages but have a short
37 residual activity (Kuhar and Kamminga 2017). This forces farmers to increase the frequency of
38 insecticide sprays (Leskey et al. 2012) causing the disruption of previously established integrated
39 pest management (IPM) programs and the resurgence of secondary pests (Leskey and Nielsen
40 2018). The identification of more selective insecticides targeting BMSB would significantly reduce
41 the environmental issues associated to its control and could contribute to IPM principles. Besides
42 efficacy on BMSB and less severe side effects, residual activity of insecticides could play a key role
43 in restoring sustainable management programs.

44 Benzoylphenyl ureas (BPUs) are classified by Insecticide Resistance Action Committee (IRAC,
45 <https://www.irac-online.org/modes-of-action>) into group 15 (Inhibitors of chitin biosynthesis, type
46 0). These compounds interfere with chitin biosynthesis, thus hindering insect molts. Detrimental
47 effects on fecundity and egg viability are reported as well (Pener and Dhadialla 2012). Although
48 BPUs have been used as pesticides since the 1970s (Retnakaran and Wright 1987, Spomer and
49 Sheets 2019), the exact mode of action is still quite obscure (Zhu et al. 2016). However, Douris et
50 al. (2016) shed some light on the molecular targets of BPUs by demonstrating a direct interaction
51 with the enzyme chitin synthase 1. These insecticides are persistent and do not show cross-
52 resistance with neurotoxic active ingredients (Doucet and Retnakaran 2012, Arruda et al. 2020).
53 Moreover, BPUs are considered less harmful to beneficial insects and non-target organisms than
54 most nerve poisons (Sun et al. 2015). For these reasons, BPUs are desirable for inclusion in IPM
55 programs in several cropping systems (Dhadialla et al. 2005).

56 Several studies focused on the effects of a number of insecticides on BMSB, but the activity of

BPU has not been investigated extensively. Kamminga et al. (2012) reported promising effects of diflubenzuron and novaluron on BMSB nymphs in laboratory assays carried out in Petri dishes, no more investigations were undertaken. The first objective of this study was to evaluate the activity of triflumuron (the only BPU currently allowed on orchards in EU) on BMSB nymphs by means of laboratory assays. The effects of aged residues were also investigated because triflumuron is known to be persistent on vegetation (Marx 1977, Aplada-Sarlis et al. 1999) and this could be a crucial factor to reduce the frequency of insecticide sprays (Kuhar and Kamminga 2017). Potted peach plants were used in the experiments because this is more representative of real conditions for residues aging. Moreover, the use of living plants allowed a prolonged exposure of insects, which is needed because of the mode action of BPUs (Dhadialla et al. 2005). A second aim of the study was to test, by mean of a field experiment set up in 2019 on pear orchards, if the addition of triflumuron to IPM strategies against BMSB could lead to a decrease in injuries to fruit.

Materials and methods

Insects

BMSB nymphs were obtained from a laboratory colony established in 2015 at the Department of Agricultural and Food Sciences (University of Bologna). Bugs were reared in a walk-in climatic chamber at $25^{\circ}\text{C} \pm 2^{\circ}\text{C}$, 50%–70% RH, with a 14 L:10 D photoperiod. Adults and nymphs were fed twice per week with green beans, carrots and soybean seeds; fresh fruits (pears, apples or kiwi fruits) were provided weekly to adults only.

Insecticide treatments

The insecticide Alsystin 48SC (triflumuron 39.34% = 480 g L⁻¹) was provided by Bayer Crop Sciences (Milan, Italy) and applied at 25 mL hL⁻¹, the maximum field recommended concentration for pome and stone fruits. Potted peach plants (GF-677 rootstock, approximately 30 cm high) were

sprayed using a hand sprayer up to dripping and allowed to dry completely before trials. Control plants were sprayed in the same way with tap water.

Laboratory bioassays

From July 2017 to November 2018, four aging periods (0, 7, 14 and 21 days) of insecticide residues were investigated, each in a separate experiment. Twelve experimental units, each consisting of a potted peach plant, were arranged for each aging period. Six plants were treated with triflumuron and the others, which were assigned to control, with tap water. Overall, 48 experimental units were set up and 480 BMSB nymphs were tested.

Once sprayed outdoor, plants were transferred into a glasshouse box where residues of the treatments aged. Here the plants were maintained for 7, 14 or 21 days at the following conditions:

T_{\max} 30° C, T_{\min} 15° C, 16 L:8 D photoperiod, minimum artificial lighting in photophase 70,000 lx.

In the experiment of 0-day aging of residues, the peach plants were used immediately after treatment droplets dried.

To set up experimental units, potted plants were placed in plexiglas cylinders (diameter 8 cm, height 30 cm), sealed with a fine net on the top and with a hole (diameter 8 cm) closed with a fine net on the base to allow air circulation and avoid mold growth. The soil surface on the pot was covered with nonwoven fabric, which was tightly wrapped to plant stem. Ten BMSB early 3rd instar nymphs were placed in each cylinder. Given that BMSB nymphs must feed on fruit or vegetables to complete development, carrots and green beans were provided ad libitum and changed twice per week. The green beans were hung on the plants to force the nymphs to climb on canopy in order to feed (Supplementary data Fig. SD1).

Experimental units were held at the same conditions reported for BMSB rearing and mortality was checked after 7, 14 and 21 days of continuous exposure of nymphs to residues. Besides the number of dead nymphs also their instars, and therefore the number of molts they successfully completed,

was recorded. Moribund nymphs (i.e. unable to upright themselves when flipped on their back) found at the last exposure interval were reared individually until they died or reached the adult stage (recorded as a recovery). Nymphs were held in ventilated 200-mL plastic jars at the same conditions and with the same food provisions as previously reported.

Field trials

From 21 June to 5 September 2019, field experiments were carried out in eight pear orchards located in Northern Italy, which suffered severe damages by BMSB in the previous years. Each orchard was considered as an experimental block and two contiguous plots (> 0.5 ha) were delimited. One plot was managed according to IPM strategies (here after referred as “farmer’s strategy”) recommended in Northern Italy. The other plot of each orchard underwent the same management practices as the former, but a treatment with Alsystin 48 SC at 25 mL hL^{-1} was also added (hereafter reported as triflumuron strategy). This treatment was carried out between 21 and 30 June targeting the first-generation nymphs of the BMSB. Of course, no triflumuron treatment was allowed in plots managed with farmer’s strategy. The sprays of broad-spectrum insecticides, which included chlorpyrifos-methyl, acetamiprid and pyrethroids (deltamethrin, lambda-cyhalothrin and tau-fluvalinate), varied slightly among orchards, as farmers were allowed to make minor changes in order to adjust for pest pressure and phenology. These sources of variation were accounted in the block factor.

Fruit sampling was carried out in a transect of 3 rows x 15 m. The transects were established in the middle of each plot at least 10 m away from plot edges to avoid border effect from the perimeter of the orchards and to accommodate for drift of triflumuron spray. External injuries were checked at harvesting on a sample of 400 fruits per plot. Two hundred fruits were sampled at eye-level and 200 were sampled above 2.5-m height using a fruit picking ladder, because most severe injuries by BMSB are usually recorded on fruit growing in the upper parts of the trees (Bariselli et al. 2016).

Pears were examined directly on the trees and scored as injured if at least one distinct depression or discoloration on fruit surface compatible with BMSB punctures could be detected (Acebes-Doria et al. 2016). Although the occasional occurrence of Heteropteran bugs other than BMSB could not be ruled out, the overwhelming majority of fruit injuries were likely due to feeding activity by BMSBs. Damages to fruit by Heteropteran bugs have been rarely reported in orchards of Northern Italy and the symptoms used to score injuries to pears are quite specific to BMSB.

Statistical analysis

To assess the activity of triflumuron on BMSB nymphs in laboratory experiments, insecticide-treated plants and relative water controls were compared within each aging period of residues. Data were analyzed by general linear mixed models (GLMM) with first-order autoregressive covariance structure, binomial distribution, probit link function and Kenward–Roger method for estimating degrees of freedom. The number of dead insects out of the total nymphs tested was considered as dependent variable. The treatments (triflumuron and water control) were used as factors, the exposure intervals (7, 14 and 21 days) were included as repeated measures and their interaction (treatment x exposure interval) were tested as well.

To study the possible decrease in the activity of triflumuron residues (i.e. to compare mortality only in insecticide-treated plants among different aging period), nymphs mortality recorded in triflumuron experimental units was corrected by Schneider-Orelli formula (Schneider-Orelli 1947) considering as natural the mean mortality recorded in water controls aged for the same period. Corrected percentages of mortality matched the assumptions of parametric tests and were analyzed by a mixed design ANOVA without any transformation. Aging periods (0, 7, 14 and 21 days) were considered as between-subject factor and exposure intervals (7, 14 and 21) as within-subject factor. The interaction aging period x exposure interval was tested as well.

Pearson χ^2 was used to test the association between the ages of residues and the number of molts that

dead nymphs exposed to triflumuron were able to successfully complete before dying.

A GLMM was carried out to analyze data from the field experiments. Binomial error distribution and log link function were selected, the number of injured fruits out of the total number of fruits checked was considered as dependent variable, treatments (farmer's strategy vs triflumuron strategy) and location of fruits (>2.5 m vs eye level) were considered as fixed factors, and orchards were included as random block factor.

All the analyses were performed using IBM SPSS Statistics (ver. 26).

Results

Laboratory bioassays

GLMMs detected significant effects on mortality by both main factors (i.e., treatment and exposure interval) for all aging periods of residues. Whereas the interaction treatment * exposure interval was significant only for 7- and 14 -day aged residues (Table 1). In other word, triflumuron residues caused overall higher mortality than water controls and longer exposure interval increased mortality (Fig. 1). Because of significant interactions, treatments were compared within each of the exposure intervals for 7-and 14-day old residues. For both aging periods, GLMMs did not detect significant difference between triflumuron and control at 7-day exposure but indicated higher mortality in triflumuron than in water at 14- and 21-day exposures (Table 1).

Mortality increased at longer exposure intervals also in the controls with a certain degree of variation among aging periods. Overall, the lowest number of dead nymphs was counted in the controls of residues tested immediately after drying. Whereas for other aging periods the natural mortality ranged from $16.06 \pm 3.99\%$ to $28.60 \pm 9.80\%$, $23.55 \pm 4.99\%$ to $38.14 \pm 7.30\%$ and $30.07 \pm 4.77\%$ to $47.79 \pm 6.61\%$ for 7-, 14- and 21-day exposure, respectively.

The comparison of corrected percentages of mortality did not show any significant effect of aging periods on the activity of triflumuron residues ($F_{(3; 20)} = 1.34$; $p = 0.29$). The interaction aging

period * exposure interval was not significant too ($F_{(6, 40)} = 1.32$; $p = 0.27$). Therefore, the aging of residues up to 21 days did not significantly decrease the activity of the insecticide.

The number of molts that nymphs exposed to triflumuron were able to complete before dying did not differ among aging periods ($\chi^2 = 6.34$; $df = 6$; $p = 0.39$). Therefore, the ages of residues did not significantly affect the insecticide speed of action. Pooling all aging periods, 29.70% of BMSB nymphs died without completing any molt, 57.43% of individuals successfully molted from 3rd to 4th instar but died before or during the next molt, 12.87% reached the 5th instar but failed to cast the exuviae and died during the molt to adult stage (Supplementary data Fig. SD2).

Field trials

The addition of triflumuron to insecticide treatments allowed in IPM strategy for BMSB led to a significant decrease in the percentage of injured fruits ($F_{(1; 28)} = 106.93$; $p < 0.001$). Fruit location had also a significant effect ($F_{(1; 28)} = 137.92$; $p < 0.001$), and the percentage of injured fruits was higher above 2.5 m than at eye level. The interaction strategy * fruit location was not significant ($F_{(1; 28)} = 0.81$; $p = 0.78$). A mean of $9.99 \pm 1.98\%$ injured fruits was detected in plots managed with triflumuron strategy, whereas $19.45 \pm 3.55\%$ of fruits were injured in plots assigned to farmer's strategy. Overall, $20.19 \pm 3.42\%$ of pears showed symptoms of feeding activity by BMSB above 2.5 m. This percentage dropped to 9.25 ± 1.95 at eye level (Fig. 2).

Discussion

Triflumuron residues on potted peach plants showed a significant activity on BMSB nymphs that died increasingly between progressive nymphal instars. These results agreed with laboratory assays by Kamminga et al. (2012) who reported 90% and 65% mortality of BMSB nymphs fed for 7 days with fresh beans dipped in solutions of novaluron and diflubenzuron, respectively. Significant reduction in the abundance of BMSB and other stink bugs was also described for diflubenzuron

207 applications to soybean in open field conditions (Herbert et al. 2013). On the other hand, Gradish et
208 al. (2019) found neither contact nor residual toxicity of novaluron on BMSB nymphs. However, in
209 this study, mortality was checked only in the short term, whereas prolonged exposure to BPU
210 residues is necessary to detect activity of this group of insecticide (Dhadialla et al. 2005).

211 The activity of triflumuron on BMSB nymphs persisted up to 21 days of residue aging in
212 glasshouse. Overall, the mortality of bugs exposed to the insecticide was higher than in controls for
213 all ages of residues. Moreover, no significant decrease in mortality could be detected on insecticide-
214 treated plants at increasing periods of residues aging. The environmental persistence of BPUs has
215 been well documented and the slow decrease in residue concentration of triflumuron over time
216 (Bayer Crop Sciences, unpublished data) is in line with the steady insecticidal activity of this active
217 ingredient up to the longest aging period. Given that treated plants were maintained in a glasshouse,
218 the only relevant insecticide degradation factor was photolysis. Rainfall, which was not considered
219 in this study, seems indeed a minor issue as BPUs are known to adhere to plant waxes and to
220 undergo to slight wash off (Retnakaran and Wright 1987, Doucet and Retnakaran 2012).

221 A relatively high mortality in the controls of some aging periods was related to the long-term
222 exposure used in the bioassays. Fluctuations in nymphal survival has been often observed in BMSB
223 laboratory colonies. Our findings were in line with Medal et al. (2013) who reported a juvenile
224 mortality from 23% to 50% in BMSB rearing and with Fisher et al. (2020) who recorded overall
225 low nymphal survival when testing the influence of temperature and humidity on BMSB.

226 Progressive poisoning through tarsal adsorption seemed the main way of insecticide contamination
227 because most of BMSB nymphs did not die at the first molt, but many insects completed one or
228 even two molts before succumbing. Ingestion has been usually considered as the major way of
229 uptake for BPUs, but slow penetration through cuticle was also described (Hammann and
230 Sirrenberg 1980, Retnakaran and Wright 1987, Sun et al. 2015). This second way of triflumuron
231 uptake was likely prevalent in our experiments, given that untreated food was used and the bug

232 probing on treated peach plants was negligible.

233 Given that intoxicated nymphs could survive for a couple of weeks, the damage they may cause

234 before succumbing needs to be taken into account. In this context, sub lethal effects may be **worth**

235 of consideration as secondary impacts on pest population (Cira et al. 2017, Depalo et al. 2017). In

236 particular, BPU's could reduce hatch rate, impair feeding activity and increase susceptibility to

237 pathogens on intoxicated BMSB (Catchot et al. 2021).

238 Early BMSB instars are more susceptible to insecticides than adults and several insecticides have

239 demonstrated a significant activity on nymphs (Kuhar and Kamminga 2017, Gradish et al. 2019).

240 Mostly broad-spectrum neurotoxic active ingredients have been so far investigated and, in

241 comparison with these insecticides, BPU's are more compatible with IPM (Dhadialla et al. 2005,

242 Doucet and Retnakaran 2012). Nevertheless, BPU's do not lack any impact on beneficials. Lethal

243 effects have been found on nymphs of predatory bugs (Soares et al. 2019) and on larvae of

244 ladybeetles (Cabrera et al 2017). Some sub-lethal effects have been reported on adult parasitoids

245 either after direct contact with residues (Matioli et al. 2019) or when parasitized eggs of the hosts

246 were treated (Goulart et al. 2012). Therefore, the possible detrimental effects of BPU's on natural

247 enemies of BMSB seem worth of investigations to draw a comprehensive picture of the possible

248 role of these insecticides in IPM scenario.

249 Besides the environmental side effects and the disruption of IPM practices, the overuse of a few

250 neurotoxic insecticides to control BMSB often impaired the rotation of insecticide chemistries upon

251 which insecticide resistance management strategy should rely (Hurley and Mitchell 2014). The

252 potential for insecticide-resistance development in BMSB populations has not been established yet,

253 but it would be wise to avoid over-reliance on neurotoxic active ingredients (Alford et al. 2020).

254 From this standpoint, the inclusion of BPU's in IPM practices could help to restore active ingredient

255 rotation as a strategy to delay the insurgence of resistance (Spomer and Sheets 2019).

256 The long-lasting activity of triflumuron, and possibly other BPU's, could be exploited to reduce

257 damages by BMSB nymphs without the need of frequent treatments. The short residual activity,
258 which lasts less than three days for most insecticides used against BMSB (Leskey et al. 2014), is
259 widely recognized as a major problem (Kuhar and Kamminga 2017). This pest is polyphagous and
260 very mobile; both nymphs and adults have big dispersal abilities and move across the landscape
261 following phenology of crops and uncropped plants (Martinson et al. 2015, Hamilton et al. 2018).
262 Repeated and unpredictable infestations of orchards are common and frequent sprays of short-
263 lasting insecticides are needed.

264 The field experiments, which showed a significant decrease in the percentages of injured pears
265 where triflumuron was added, reinforced the findings of laboratory assays. Field trials pointed out
266 also that in 2019 strategies mainly relying on neurotoxic active ingredients did not achieve a
267 satisfactory control of BMSB. The mean percentage of injured fruit recorded in plots managed by
268 farmer's strategy (19.45 ± 3.55) far exceeded the economic threshold. Although the percentage of
269 injured fruits dropped to 9.99 ± 1.98 in the plots where triflumuron was added, this was still not
270 completely acceptable from an economic standpoint. A refinement of management strategies for
271 BMSB seems therefore urgently needed and the development of treatment thresholds for this stink
272 bug would be a relevant step forward (Short et al. 2017).

273 BPUs lack any lethal effect on adult insects and have no chance to **decrease** the pressure exerted by
274 highly damaging adult bugs that migrate into crops from surrounding areas (Acebes-Doria et al.
275 2016). Therefore, triflumuron and other BPUs might be considered for field applications only as a
276 tool in a wider management strategy. Control methods aimed at preventing the invasion of crops by
277 adult bugs such as exclusion nets (Candian et al. 2018), insecticide netting screens and sprays at
278 orchard perimeter (Blaauw et al. 2015) might take major benefits from few and long-lasting BPU
279 treatments aimed decreasing nymphal populations and hampering reproduction cycle that can occur
280 within the orchards.

281

282 Acknowledgments

283 We are grateful to Elisa Marchetti and Andrea Angelici for technical assistance in BMSB rearing
284 and in bioassays. We also thank personnel from Bayer Crop Science for help in fruit samplings.
285 Part of this study was supported by Bayer Crop Science.

286

287

288 References Cited

- 289 **Acebes-Doria, A. L., T. C. Leskey, and Bergh, J. C. 2016.** Injury to apples and peaches at harvest
290 from feeding by *Halyomorpha halys* (Stål) (Hemiptera: Pentatomidae) nymphs early and
291 late in the season. Crop Prot. 89: 58-65.
- 292 **Alford A., T. P. Kuhar, G. C. Hamilton, P. Jentsch, G. Krawczyk, J. F. Walgenbach, and C.**
293 **Welty. 2020.** Baseline toxicity of the insecticides bifenthrin and thiamethoxam on
294 *Halyomorpha halys* (Hemiptera: Pentatomidae) collected from the eastern United States. J.
295 Econ. Entomol. 113: 1043-1046.
- 296 **Aplada-Sarlis, P. G., G. E. Miliadis, and N. G.Tsiropoulos. 1999.** Dissipation of teflubenzuron
297 and triflumuron residues in field-sprayed and cold-stored pears. J. Agric. Food. Chem. 47:
298 2926-2929.
- 299 **Arruda, L. S., A. R. Rodrigues, N. C. Bermudez, L. M. Ribeiro, J. E. L. Neto, and H. A.**
300 **Siqueira. 2020.** Field resistance of *Plutella xylostella* (Lepidoptera: Plutellidae) to
301 lufenuron: Inheritance and lack of cross-resistance to methoxyfenozide. Crop. Prot. 136:
302 article 105237.
- 303 **Bariselli, M., R. Bugiani, and L. Maistrello. 2016.** Distribution and damage caused by
304 *Halyomorpha halys* in Italy. EPPO Bull. 46: 332-334.
- 305 **Bosco, L., S. T. Moraglio, and L. Tavella. 2017.** *Halyomorpha halys*, a serious threat for hazelnut
306 in newly invaded areas. J. Pest. Sci. 91: 661–670.

- 307 **Blaauw, B. R., D. Polk, and A. L. Nielsen. 2015.** IPM-CPR for peaches: incorporating
 308 behaviorally-based methods to manage *Halyomorpha halys* and key pests in peach. *Pest.*
 309 *Man. Sci.* 71: 1513-1522.
- 310 **Cabrera, P., D. Cormier, and É. Lucas. 2017.** Differential sensitivity of an invasive and an
 311 indigenous ladybeetle to two reduced-risk insecticides. *J. Appl. Entomol.* 141: 690–701.
- 312 **Candian, V., M. G. Pansa, R. Briano, C. Peano, R. Tedeschi, and L. Tavella. 2018.** Exclusion
 313 nets: a promising tool to prevent *Halyomorpha halys* from damaging nectarines and apples
 314 in NW Italy. *Bull. Insectol.* 71: 21-30.
- 315 **Catchot, B. D., F. R. Musser, J. Gore, N. Krishnan, D. R. Cook, S. D. Stewart, G. M. Lorenz,**
 316 **S. Brown, N. Seiter, A. L. Catchot, D. L. Kerns, R. Jackson, and K. S. Knighten. 2021.**
 317 Sublethal impacts of novaluron on Tarnished Plant Bug (Hemiptera: Miridae) adults. *J.*
 318 *Econ. Entomol.* 114: 739-746.
- 319 **Cira, T. M., E. C. Burkness, R. L. Koch, and W. D. Hutchison. 2017.** *Halyomorpha halys*
 320 mortality and sublethal feeding effects following insecticide exposure. *J. Pest. Sci.* 90:
 321 1257–1268.
- 322 **Depalo, L., A. Lanzoni, A. Masetti, E. Pasqualini, and G. Burgio. 2017.** Lethal and sub-lethal
 323 effects of four insecticides on the aphidophagous coccinellid *Adalia bipunctata* (Coleoptera:
 324 Coccinellidae). *J. Econ. Entomol.* 110: 2662-2671.
- 325 **Dhadialla, T. S., A. Retnakaran, and G. Smagghe. 2005.** Insect growth and development
 326 disrupting insecticides. *In* L. I. Gilbert, K. Iatrou, and S. S. Gill (eds.), *Comprehensive*
 327 *insect molecular science*, vol 6. Pergamon Press, Oxford, pp 55–100
- 328 **Doucet, D., and A. Retnakaran. 2012.** Insect chitin: metabolism, genomics and pest management.
 329 *In* T. S. Dhadialla (ed.), *Advances in insect physiology - Insect growth disruptors*, vol 43.
 330 Elsevier, Oxford, pp 437-511.

- 331 **Douris, V., D. Steinbach, R. Panteleri, I. Livadaras, J. A. Pickett, T. Van Leeuwen, R. Nauen,**
332 **and J. Vontas. 2016.** Resistance mutation conserved between insects and mites unravels
333 the benzoylurea insecticide mode of action on chitin biosynthesis. *Proc. Natl. Acad. Sci.*
334 U.S.A. 113: 14692-14697.
- 335 **Fisher, J. J., R. P. Rijal, and F. G. Zalom. 2020** Temperature and humidity interact to influence
336 Brown Marmorated Stink Bug (Hemiptera: Pentatomidae), survival. *Environ. Entomol.* (IN
337 PRESS, <https://doi.org/10.1093/ee/nvaa146>).
- 338 **Goulart, R. M., H. X. Volpe, A. M. Vacari, R. T. Thuler, and S. A. De Bortoli. 2012.** Insecticide
339 selectivity to two species of *Trichogramma* in three different hosts, as determined by
340 IOBC/WPRS methodology. *Pest Man. Sci.* 68: 240-244.
- 341 **Gradish, A. E., H. Fraser and C. D. Scott-Dupree. 2019.** Direct and residual contact toxicity of
342 insecticides to *Halyomorpha halys* (Hemiptera: Pentatomidae). *Can. Entomol.* 151: 209-218.
- 343 **Hamilton, G. C., J. J. Ahn, W. Bu, T. C. Leskey, and A. L. Nielsen. 2018.** *Halyomorpha halys*
344 (Stål). In J. E. McPherson (ed.), *Invasive stink bugs and related species (Pentatomoidea):*
345 *biology, higher systematics, semiochemistry, and management.* CRC Press Taylor & Francis
346 Group, Boca Raton, pp 243- 292.
- 347 **Hammann, I., and W. Sirrenberg. 1980.** Laboratory evaluation of SIR 8514, a new chitin
348 synthesis inhibitor of the benzoylated urea class. *Pflanzenschutz-Nachrichten Bayer* 33: 1-
349 34.
- 350 **Herbert, D. A., S. Malone, M. Arrington, and J. Hogue. 2013.** Evaluation of selected foliar
351 insecticides for control of stink bug in soybean, 2012. *Arthropod. Man. Tests* 38
- 352 **Hurley, T. M., and P. D. Mitchell. 2014.** Insect resistance management: Adoption and
353 compliance. In D.W. Onstad (ed.), *Insect Resistance Management.* Academic Press, pp 421-
354 451.

- 355 **Kamminga, K. L., T. P. Kuhar, A. Wimer, and D. A. Herbert. 2012.** Effects of the insect growth
356 regulators novaluron and diflubenzuron on the brown marmorated stink bug. *Plant Health*
357 *Progr.* 13, 2. (doi:10.1094/PHP-2012-1212-01-RS)
- 358 **Kuhar, T. P., and K. L. Kamminga. 2017.** Review of the chemical control research on
359 *Halyomorpha halys* in the USA. *J. Pest. Sci.* 90: 1021-1031.
- 360 **Leskey, T. C., and A. L. Nielsen. 2018.** Impact of the invasive brown marmorated stink bug in
361 North America and Europe: history, biology, ecology, and management. *Annu. Rev.*
362 *Entomol.* 63: 599-618.
- 363 **Leskey, T. C., B. D. Short, B. R. Butler, and S. E. Wright. 2012.** Impact of the invasive brown
364 marmorated stink bug, *Halyomorpha halys* (Stål), in mid-Atlantic tree fruit orchards in the
365 United States: case studies of commercial management. *Psyche* 2012: 1–14
366 (doi:10.1155/2012/535062)
- 367 **Leskey, T. C., B. D. Short, and D. H. Lee. 2014.** Efficacy of insecticide residues on adult
368 *Halyomorpha halys* (Stål) (Hemiptera: Pentatomidae) mortality and injury in apple and
369 peach orchards. *Pest. Man. Sci.* 70: 1097-1104.
- 370 **Martinson, H. M., P. D. Venugopal, E. J. Bergmann, P. M. Shrewsbury, and M. J. Raupp.**
371 **2015.** Fruit availability influences the seasonal abundance of invasive stink bugs in
372 ornamental tree nurseries. *J. Pest. Sci.* 88: 461–468.
- 373 **Marx, J. L. 1977.** Chitin synthesis inhibitors: new class of insecticides. *Science* 197: 1170-1172.
- 374 **Matioli, T. F., O. Z. Zanardi, and P. T. Yamamoto. 2019.** Impacts of seven insecticides on
375 *Cotesia flavipes* (Cameron) (Hymenoptera: Braconidae). *Ecotoxicol.* 28: 1210-1219.
- 376 **Medal, J., T. Smith, and A. Santa Cruz. 2013.** Biology of the brown marmorated stink bug
377 *Halyomorpha halys* (Heteroptera: Pentatomidae) in the laboratory. *Fla. Entomol.* 96: 1209-
378 1212.

- 379 **Pener, M. P., and T. S. Dhadialla. 2012.** An overview of insect growth disruptors; applied aspects.
 380 *In* T. S. Dhadialla (ed.), *Advances in insect physiology - Insect growth disruptors*, vol 43.
 381 Elsevier, Oxford, pp 1-162.
- 382 **Retnakaran, A., and J. E. Wright. 1987.** Control of insect pests with benzoylphenyl ureas. *In* J. E.
 383 Wright, and A. Retnakaran (eds.), *Chitin and benzoylphenyl ureas. Series Entomologica*, vol
 384 38. Springer, Dordrecht, pp 205-282.
- 385 **Schneider-Orelli, O. 1947.** Entomologisches praktikum: Einführung in die land-und
 386 forstwirtschaftliche Insektenkunde. Sauerländer.
- 387 **Short, B. D., A. Khrimian, and T. C. Leskey. 2017.** Pheromone-based decision support tools for
 388 management of *Halyomorpha halys* in apple orchards: development of a trap-based
 389 treatment threshold. *J. Pest. Sci.* 90: 1191–1204.
- 390 **Soares, M. A., L. C. Passos, M. R. Campos, L. J. Collares, N. Desneux, and G. A. Carvalho.**
 391 **2019.** Side effects of insecticides commonly used against *Tuta absoluta* on the predator
 392 *Macrolophus basicornis*. *J. Pest Sci.* 92: 1447-1456.
- 393 **Spomer, N. A., and J. J. Sheets. 2019.** Chitin biosynthesis and inhibitors. *In* P. Jeschke, M.
 394 Witschel, W. Krämer, and S. Ulrich, S. (eds.) *Modern crop protection compounds vol. 3:*
 395 *Insecticides*. Wiley-VCH Verlag GmbH & Co. Weinheim, pp 1067-1085.
- 396 **Sun, R., C. Liu, H. Zhang, and Q. Wang. 2015.** Benzoylurea chitin synthesis inhibitors. *J. Agric.*
 397 *Food. Chem.* 63: 6847-6865.
- 398 **Zhu, K. Y., H. Merzendorfer, W. Zhang, J. Zhang, and S. Muthukrishnan. 2016.** Biosynthesis,
 399 turnover, and functions of chitin in insects. *Annu. Rev. Entomol.* 61: 177-196.

400 **Figure Caption**

401 **Fig. 1.** Mortality of *Halyomorpha halys* nymphs exposed for 7, 14 and 21 days to residues of
402 triflumuron (black line) or control (gray line) on potted peach plants. Each panel represent a
403 different aging period of residues in glasshouse conditions. Vertical bars indicate standard errors of
404 the means. Asterisks indicate significant differences between treatments, which were compared
405 within exposure intervals, in the case of significant interaction of treatment * exposure interval
406 detected by **GLMM**.

407

408 **Fig. 2.** Percentages of fruit with feeding injuries by *Halyomorpha halys* recorded in 2019 in pear
409 orchards in Northern Italy. GLMM detected significant differences between strategies ($p<0.001$)
410 and between fruit locations ($p<0.001$). Vertical bars indicate standard errors of the means.

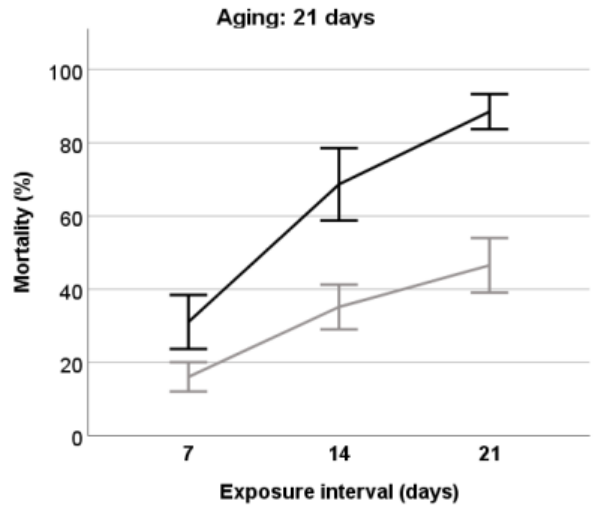
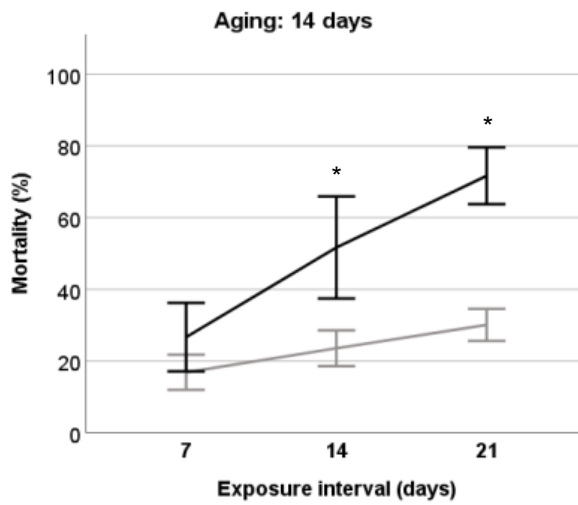
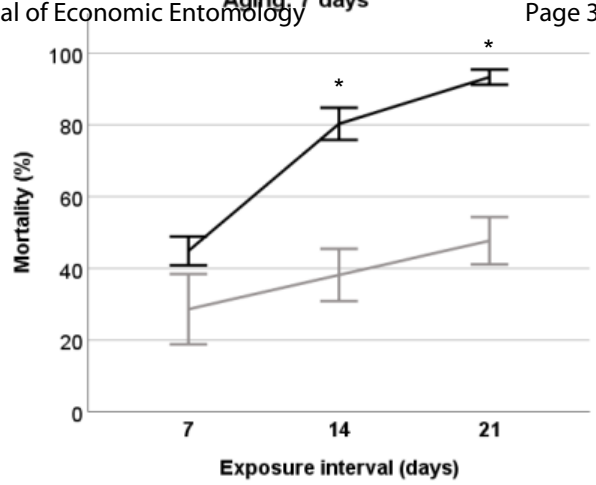
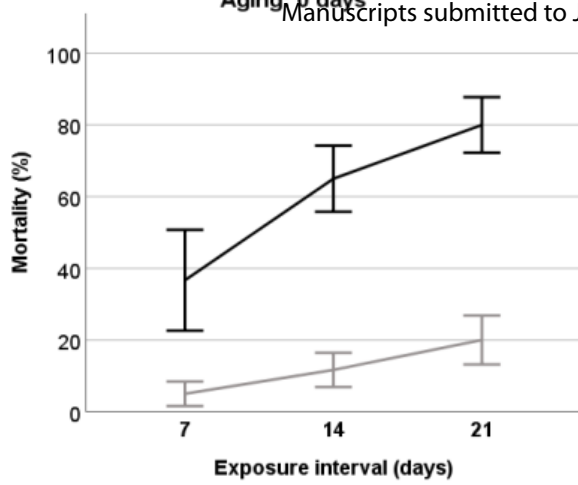
Table 1. Percentages of mortality (mean \pm standard error) of *Halyomorpha halys* nymphs recorded for the different aging periods of residues at increasing exposure intervals. **GLMMs** were carried separately for each aging period and tests of fixed effects are reported. In the case of significant interaction of treatment * exposure interval, treatments were compared within each exposure interval and the *p* level is reported below mortality means.

0-day aged residues		Exposure intervals		
Treatment	7 days	14 days	21 days	
Triflumuron	36.67 \pm 14.06	65.00 \pm 9.22	80.00 \pm 7.72	
Control	5.00 \pm 3.42	11.67 \pm 4.77	20.00 \pm 6.83	
GLMM effects	F	df	p	
Treatment	14.35	1; 9.89	0.004	
Exposure interval	10.58	2; 21.12	0.001	
Treatment * Exposure interval	0.42	2; 21.12	0.661	

7-day aged residues		Exposure intervals		
Treatment	7 days	14 days	21 days	
Triflumuron	44.85 \pm 4.01	80.30 \pm 4.48	93.33 \pm 2.11	
Control	28.60 \pm 9.80	38.14 \pm 7.30	47.69 \pm 6.61	
	<i>p</i> = 0.064	<i>p</i> < 0.001	<i>p</i> < 0.001	
GLMM effects	F	df	p	
Treatment	21.03	1; 10.33	0.001	
Exposure interval	27.88	2; 20.84	<0.001	
Treatment * Exposure interval	7.50	2; 20.84	0.004	

14-day aged residues		Exposure intervals		
Treatment	7 days	14 days	21 days	
Triflumuron	26.67 \pm 9.55	51.67 \pm 14.24	71.67 \pm 7.92	
Control	16.85 \pm 4.90	23.55 \pm 4.99	30.07 \pm 4.77	
	<i>p</i> = 0.327	<i>p</i> = 0.023	<i>p</i> = 0.002	
GLMM effects	F	df	p	
Treatment	6.05	1; 11.09	0.032	
Exposure interval	18.05	2; 21.03	<0.001	
Treatment * Exposure interval	3.82	2; 21.03	0.038	

21-day aged residues		Exposure intervals		
Treatment	7 days	14 days	21 days	
Triflumuron	31.06 \pm 7.38	68.64 \pm 9.88	88.48 \pm 4.79	
Control	16.06 \pm 3.99	35.15 \pm 6.11	46.52 \pm 7.56	
GLMM effects	F	df	p	
Treatment	13.23	1; 11.31	0.004	
Exposure interval	29.13	2; 20.92	<0.001	
Treatment * Exposure interval	2.51	2; 20.92	0.105	



— = triflumuron — = control

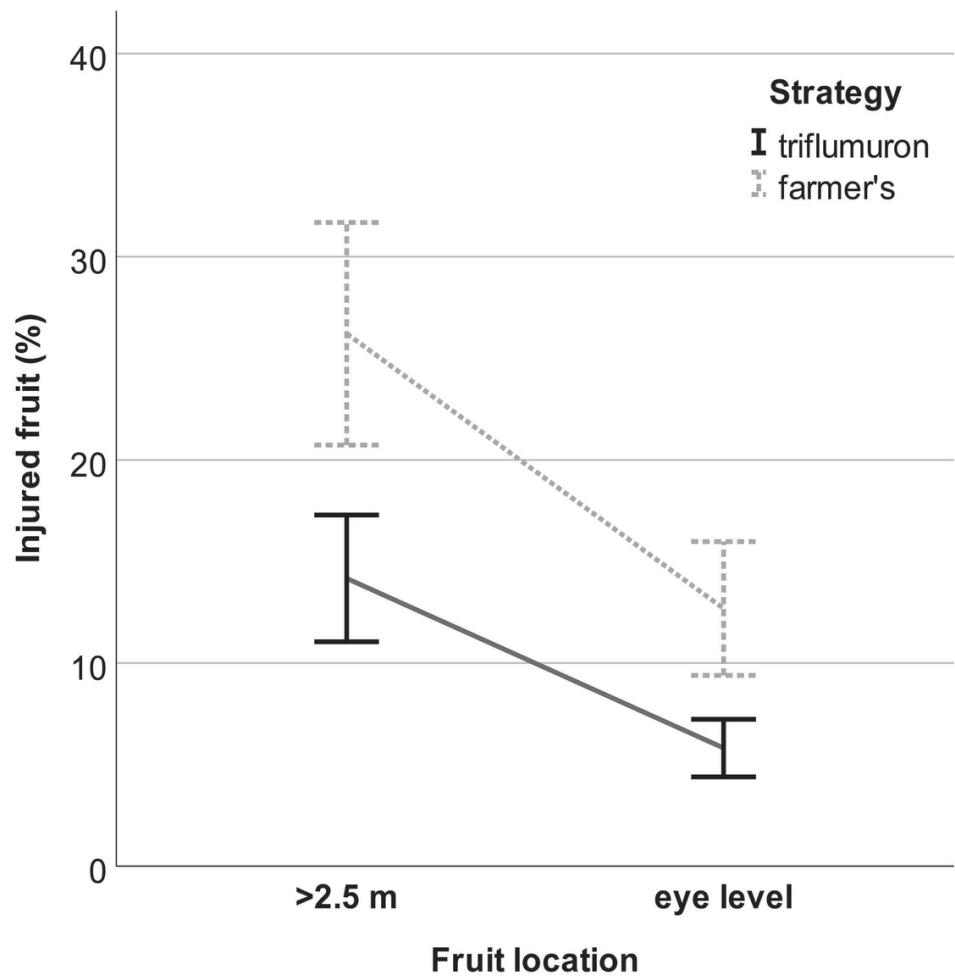


Fig. 2. Percentages of fruit with feeding injuries by *Halyomorpha halys* recorded in 2019 in pear orchards in Northern Italy. GLMM detected significant differences between strategies ($p<0.001$) and between fruit locations ($p<0.001$). Vertical bars indicate standard errors of the means.

99x99mm (300 x 300 DPI)

SUPPLEMENTARY DATA

Impact of triflumuron on *Halyomorpha halys* (Hemiptera: Pentatomidae):

laboratory and field studies

Antonio Masetti, Laura Depalo and Edison Pasqualini



Fig. SD1. Experimental units with potted peach plants inserted in plexiglas cylinders.



N3 / N4



N4 / N5



N5 / adult

Fig. SD1. Individuals of *Halyomorpha halys* died at the ecdysis. Unsuccessful molting is a typical symptom of the activity of benzoylphenyl ureas.