

Alma Mater Studiorum Università di Bologna
Archivio istituzionale della ricerca

Alternative biological sources for extracellular vesicles production and purification strategies for process scale-up

This is the final peer-reviewed author's accepted manuscript (postprint) of the following publication:

Published Version:

Giancaterino S., Boi C. (2023). Alternative biological sources for extracellular vesicles production and purification strategies for process scale-up. BIOTECHNOLOGY ADVANCES, 63, 1-16 [10.1016/j.biotechadv.2022.108092].

Availability:

This version is available at: <https://hdl.handle.net/11585/919037> since: 2023-02-28

Published:

DOI: <http://doi.org/10.1016/j.biotechadv.2022.108092>

Terms of use:

Some rights reserved. The terms and conditions for the reuse of this version of the manuscript are specified in the publishing policy. For all terms of use and more information see the publisher's website.

This item was downloaded from IRIS Università di Bologna (<https://cris.unibo.it/>).
When citing, please refer to the published version.

(Article begins on next page)

Alternative biological sources for extracellular vesicles production and purification strategies for process scale-up

Sara Giancaterino and Cristiana Boi*

Department of Civil, Chemical Environmental and Materials Engineering (DICAM)

University of Bologna, Via Terracini 28, 40131 Bologna, Italy

*Corresponding author: cristiana.boi@unibo.it

Tel.: +39 051 2090432

Abstract

Extracellular vesicles (EVs) are phospholipidic bi-layer enclosed nanoparticles secreted naturally by all cell types. They are attracting increasing attention in the fields of nanomedicine, nutraceuticals and cosmetics as biocompatible carriers for drug delivery, with intrinsic properties beneficial to human health. Scientific work now focuses on developing techniques for isolating EVs that can translate into industrial-scale production and meet rigorous clinical requirements. The science of EVs is ongoing, and many pitfalls must be addressed, such as the requirement for standard, reproducible, inexpensive, and Good Manufacturing Practices (GMP) adherent EV processing techniques. Researchers are exploring the use of alternative sources to EVs derived from mammalian cultures, such as plant EVs, as well as the use of bacteria, algae and milk. Regarding the downstream processing of EVs, many alternative techniques to the ultracentrifugation (UC) protocols most commonly used in the laboratory are emerging. In the context of process scale-up, membrane-based processes for isolation and purification of EVs are the most promising, either as stand-alone processes or in combination with chromatographic techniques. This review discusses current trends on EVs source selection and EVs downstream processing techniques, with a focus on plant-derived EVs and membrane-based techniques for EVs enrichment.

Abbreviations

<i>EVs</i>	<i>Extracellular vesicles</i>
GMP	<i>Good Manufacturing Practices</i>
<i>UC</i>	<i>Ultracentrifugation</i>
ISEV	<i>International Society for Extracellular Vesicles</i>
TFF	<i>Tangential Flow Filtration</i>
OMV	<i>Outer Membrane Vesicles</i>
SEC	<i>Size Exclusion Chromatography</i>
FFF	<i>Flow Field Fractionation</i>
AEX	<i>Anion Exchange Chromatography</i>
AC	<i>Affinity Chromatography</i>
FFF	<i>Field Flow Fractionation</i>
MF	<i>Microfiltration</i>
UF	<i>Ultrafiltration</i>
dgUC	<i>Density Gradient Ultracentrifugation</i>
CQA	<i>Critical Quality Attributes</i>
mAbs	<i>monoclonal Antibodies</i>
UF/DF	<i>Ultrafiltration/Diafiltration</i>
MWCO	<i>Molecular weight cut off</i>
PES	<i>Polyether sulfone</i>
TMP	<i>Transmembrane pressure</i>
TFAC	<i>Tangential Flow for Analyte Capture</i>
dcTFF	<i>Dual cyclic filtration system</i>
Mf-F	<i>Microfluidic filtration</i>
AF4	<i>Asymmetric Flow Field Fractionation</i>
PDEVs	<i>Plant-Derived Extracellular Vesicles</i>

26 **Keywords**

27 Extracellular vesicles; Plant-derived Extracellular vesicles; Drug delivery system; Nanomedicine;
28 Downstream Processing; Membrane-based separation processes; Process scale-up;

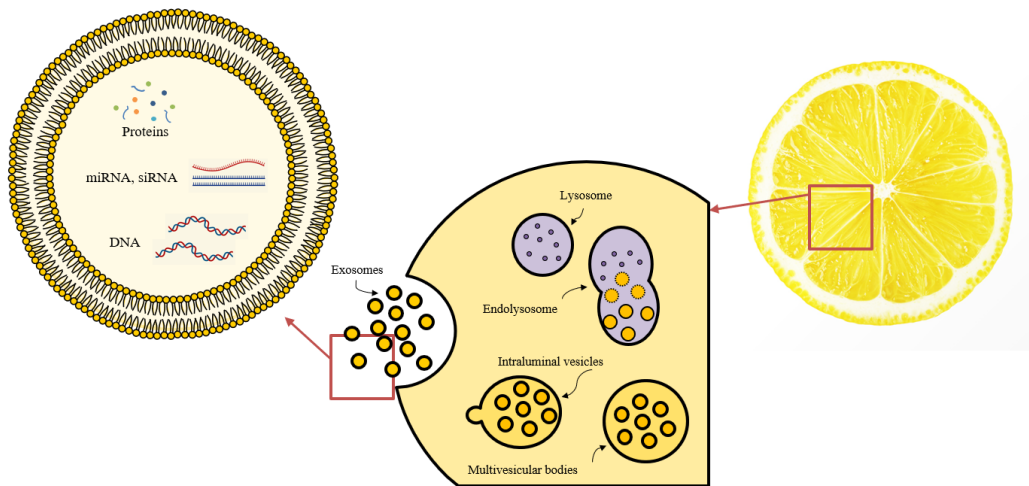
29

30 **1. Introduction**

31

32 EVs are a heterogeneous group of biological nanoparticles naturally released by cells - eukaryotes and
33 prokaryotes. They are characterized by a bi-layer membrane made by phospholipids that encloses the cytosol
34 of the deriving cell, rich in proteins, lipids and nucleic acids (mRNA, microRNA, tRNA, rRNA, DNA). The
35 most popular way to classify EVs is according to their biogenesis mechanism (Figure 1), into exosomes,
36 microvesicles and apoptotic bodies.

37

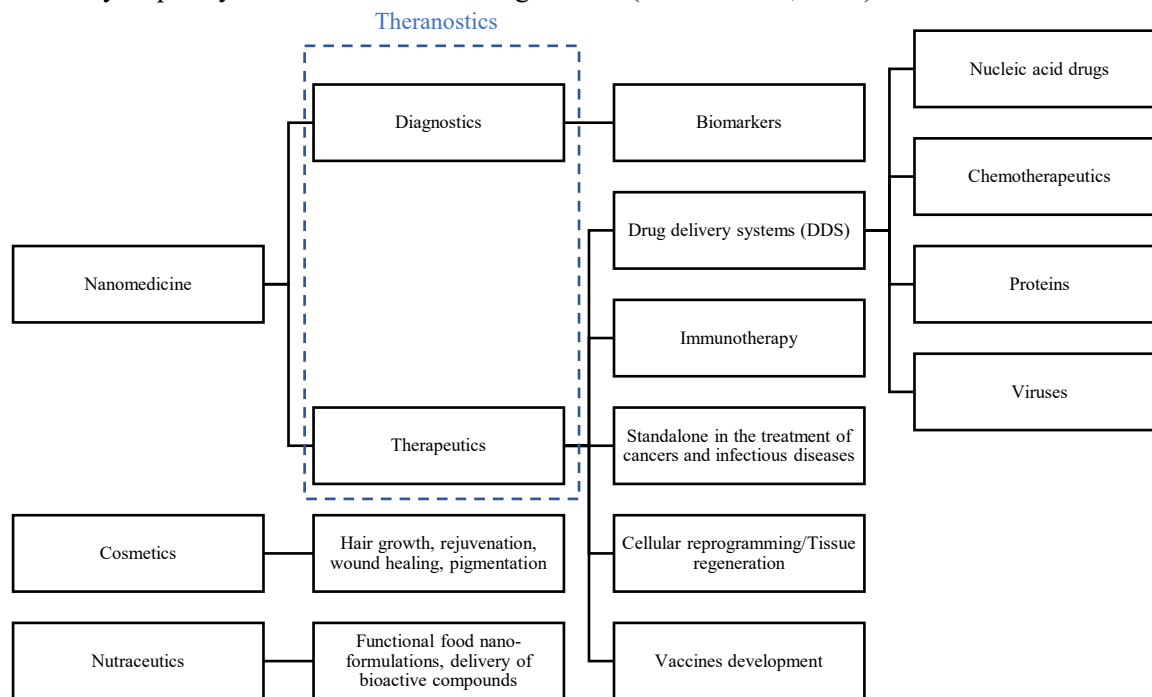


38

39 *Figure 1: All types of cells release EVs, including plant cells. Fruits such as lemons can be exploited as biological source to isolate
40 and purify EVs. These are released by cells through several biogenesis pathways – exosomes (30-150 nm) are produced during the
41 formation of multivesicular bodies (MVB) of endosomal origin. Microvesicles (50–1000 nm) are formed by budding of the plasma
42 membrane. The largest EVs, apoptotic bodies (800-5000 nm) are formed by blebbing of the membrane of apoptotic cells.*

43 Nonetheless, a clear biological distinction between the different populations is missing and the International
44 Society for Extracellular Vesicles (ISEV) recommends the use of “EVs” as blanket-term for “particles
45 naturally released from the cell that is delimited by a lipid bilayer and cannot replicate” (They et al., 2018).
46 EVs represent a “universal, evolutionary conserved mechanism for inter-kingdom and intra-kingdom
47 communication” (Chronopoulos and Kalluri, 2020) and have been defined as “signalosomes, multifunctional
48 signaling complexes for controlling fundamental cellular and biological functions” (Gandham et al., 2020).
49 EV-mediated communication is involved in all the domains of life and in many cellular physiological and
50 pathological processes. EVs contain bioactive cargos upon which they are able to deliver complex biological
51 messages to target cells, leading to the induction and coordination of the immune response, maintenance of
52 cellular integrity and homeostasis, cell development, cell differentiation and angiogenesis (Ramirez et al.,
53 2018). A glaring example of EVs functionalities comes from human diet. The discovery that plants cells do
54 secrete various types of vesicles spontaneously lead to the observation that, as we eat every day, these vesicles
55 are continuously put in contact with our intestinal tract and microbiome (Halperin and Jensen, 1967; Marchant
56 et al., 1967). Recent data suggest that EVs from food and their cargos might have relevant biological role on
57 our digestive tract, contributing to the homeostasis of the whole body through gene regulation (Rome, 2019).
58 Many studies have disclosed EVs role as cross-kingdom modulators, as EV-mediated interactions between
59 mammals, plants, bacteria and parasites (Hou et al., 2019; Ionescu et al., 2014; Rutter and Innes, 2018;

60 Svennerholm et al., 2017; Szempruch et al., 2016). EVs have a promising potential in three main domains -
 61 nanomedicine, cosmetics and nutraceuticals (Figure 2). In nanomedicine EVs can be used as drug-delivery
 62 systems, therapeutics and diagnostic tools. EVs are attractive candidates in clinical applications due to their
 63 *intrinsic* potential based on their specific bioactive cargo or exploiting their unique delivery properties.
 64 Concerning their use as drug delivery vectors evidence suggests a long-range action (e.g. ability to cross the
 65 epithelial endothelial barriers), cargo protection and engineering possibilities. In gene therapy, EVs can be
 66 modified for targeted delivery of nucleic acids-based drugs and viruses, as well as carriers for protein and
 67 small molecules to treat diseases and cancer (Gandham et al., 2020; Konoshenko et al., 2018). As stand-alone
 68 therapeutics, EVs produced by stem cells can be used to induce tissue regeneration, while EVs produced by
 69 dendritic cells and macrophages can regulate immune responses (Robbins et al., 2016). Besides, EVs have
 70 shown therapeutic effects against infectious diseases, diabetes, tumors, neurodegenerative and cardiovascular
 71 diseases (García-Manrique et al., 2018; Liu et al., 2020). The use of body-fluid-derived EVs (e.g. EVs from
 72 blood, urine, semen, and saliva) as non-invasive biomarkers for early diagnosis and prognosis of cancer, via
 73 liquid biopsies, has a revolutionary potential (Pang et al., 2020). EVs are also attractive candidates for the
 74 development of functional cosmetics for skin treatments as wound healing, rejuvenation, pigmentation and
 75 hair growth treatments (Carrasco et al., 2019; Peršurić and Pavelić, 2021). Furthermore, EVs from plants and
 76 animals are very promising to create alternative delivery options for nutraceuticals to enhance the
 77 bioavailability of poorly absorbed active food ingredients (Akuma et al., 2019).



78
 79 *Figure 2: The three main application areas of EVs: nanomedicine, cosmetics and nutraceuticals*

80 The physical and biochemical properties of EVs mirror the mother cell phenotype. Thus, there are notable
 81 differences in the release rate, biochemical composition and size, depending on the state and characteristics of
 82 the cell of origin. Current EVs production is based on vesicles naturally released from a source or EVs obtained
 83 from cell culture conditioned media under a controlled environment. The use of a certain EVs source
 84 automatically implies a better suitability for a particular application. For example, EVs from physiological
 85 fluids are mainly used for diagnostic and prognostic applications. To date, studies of mammalian EVs produced
 86 by cell culture for clinical purposes are widespread. Mesenchymal stem cells (MSC), dendritic cells, tumor
 87 cells, red blood cells and macrophages are among the most frequently used sources of therapeutic EVs (García-
 88 Manrique et al., 2018; Liu et al., 2020). In recent years, interest has grown in the use of alternative sources to
 89 human cells for drug delivery applications, such as animal EVs, plant EVs, bacterial EVs and algal EVs. An
 90 introductory analysis of the current uses, advantages and disadvantages related to the employment of each

91 different EV source is presented in this review. At present, most EVs have been isolated and purified by UC-
92 based methods, but from a manufacturing perspective, UC has many limitations and lacks the potential for
93 scalability. Its use has been reduced in favor of other methods such as filtration techniques, chromatographic
94 separations, polymer precipitation, affinity-based processes and microfluidic technologies. Currently, the field
95 of downstream processing of EVs is limited to laboratory-scale research, and there are many limitations that
96 need to be overcome to move to clinical and industrial-scale research, such as typically low yields, lot-to-lot
97 variability, lack of standardization, and development of cost-effective isolation protocols. Filtration techniques
98 hold great promise as they are already being exploited industrially in the field of liposome and virus production,
99 where tangential flow filtration (TFF) is considered the standard purification method. Membrane processes are
100 flexible, scalable and adaptable to continuous operations, making them the optimal candidates as unit
101 operations for large scale EVs production. Therefore, the second part of this review covers a detailed state-of-
102 the-art of the most widely used membrane techniques for EVs isolation and purification to identify the crucial
103 parameters that enable standardization and reproducibility of EV preparations.

104

105 **2. EVs sources**

106 Regarding biological source selection, EV production cannot rely on a single cell line, biofluid or tissue. Source
107 selection is entirely driven by *the end user application*, as the properties of EVs are closely related to the
108 functions and phenotype of the parent cell. Table 1 provides an overview of the most commonly used sources
109 with the main processing characteristics. So far, most EVs are isolated from human body fluids or produced
110 by different types of human cells, such as stem cells, dendritic cells, macrophages, epithelial cells, and tumor
111 cells. Human cell cultivation requires optimization of several parameters, such as cell isolation and banking,
112 composition of culture media and cell expansion to the desired density and amount. Some of the most crucial
113 aspects in the framework of massive EV production for clinical trials are the low available volume, cost, safety
114 and ethical compliance. In addition, the process of cell senescence and yield limitations resulting from the fact
115 that human cells are generally adherent represent further complications (Paganini et al., 2019). Indeed,
116 although some applications require specific human cell lines and their use cannot be avoided, these
117 complications have encouraged researchers to explore alternative EV sources. Animal, plant, and bacterial
118 sources are recently gaining attention in the field of EV production because they are cheap and highly available,
119 allow EVs to be easily isolated from large volumes of fluid, and lead to better yields. Bacterial and algal cells
120 cultivation has significant advantages over that of eukaryotic cells, especially in terms of proliferation ability
121 and ease of gene editing strategies. Food-derived EVs, such as plant and milk EVs, do not require any cell
122 cultivation, thus their use saves entirely on upstream costs and management. Besides, food-derived EVs are
123 inherently biocompatible, safe and possess many beneficial effects on human health, by being part of our
124 dietary regimen (Ly et al., 2023). Researchers around the world are trying to isolate EVs from many different
125 natural sources in an effort to find the most economically viable and sustainable sources that could translate
126 toward massive EV production. From the perspective of a circular bioeconomy, residues from animals, fruits
127 and vegetables can be potentially employed as sources for EV production. In this context, EVs represent a
128 promising valorization pathway, allowing the conversion of agro- and animal-waste into many EV-based
129 added-value products (Sangiorgio et al., 2020). However, it is crucial to consider that there is still a substantial
130 knowledge gap related to the biological role of EVs from plants and animals, and that the level of maturation
131 of the field, compared to that of mammalian cells, is in its infancy.

132

132 **2.1. Bacterial EVs**

133 EVs are naturally released by both gram-negative and gram-positive bacteria. There are different kinds of
134 bacterial vesicles, but Outer Membrane Vesicles (OMVs) from gram-negative bacteria are the most studied.
135 They are generally smaller than eukaryotic EVs, having dimensions ranging from 20 to 300 nm, and are
136 released through the blebbing of the cell wall. The presence of liposaccharides toxins on OMVs surface is a
137 key molecular feature, besides the presence of outer membrane lipids and proteins, soluble periplasmic

138 components and peptidoglycans (Schwechheimer and Kuehn, 2015). Bacterial EVs are much less studied than
139 those of mammalian origin, but several studies have demonstrated their prominent physiological and
140 pathological role as mediators, in bacteria–bacteria and bacteria–host interactions (Ñahui Palomino et al.,
141 2021). Bacterial EVs are capable of triggering an innate immune response by presenting EV surface ligands –
142 natural or engineered – to the immune cell pattern recognition receptors (Gilmore et al., 2021). Due to their
143 potent immunomodulatory properties, the potential use of bacterial EVs as therapeutics is increasingly being
144 studied, especially as immune adjuvants against infections, platforms for vaccine development and anticancer
145 therapies (Chronopoulos and Kalluri, 2020; Jahromi and Fuhrmann, 2021). Bacterial EVs are extremely
146 promising in vaccine design and development, as they can increase the antibody production by simultaneously
147 carrying multiple viral antigens on their surface, (Cai et al., 2018; Gerritzen et al., 2017; L. Zhang et al., 2016).
148 They are low cost, scalable, easy to manipulate, and their release can be spontaneous in a culture medium or
149 even induced by the use of a chemical detergent (e.g., sodium deoxycholate), heat stress or antibiotics (Momen-
150 Heravi et al., 2013). By genetically engineering donor cells, more efficient recombinant vaccines can be
151 obtained, with further improvements to their safety profile, immunogenicity and yield (Jiang et al., 2019).
152 Gerritzen et al. developed a vaccine platform based on OMVs produced by *Neisseria meningitidis* (Gerritzen
153 et al., 2019). The vaccine’s mechanism of action is based on the expression on heterologous antigens on the
154 OMVs. The release of OMVs was powered by high concentration of oxygen in the culture media, and
155 tangential flow microfiltration was used as a scalable purification strategy. The authors were able to obtain 90
156 mg of OMV proteins per liter of culture.

157 2.2. Algae EVs

158 Several studies have shown that microalgae are promising sources of EVs (Adamo et al., 2021; Kuruvinashetti
159 et al., 2020; Picciotto et al., 2021). Microalgae are a natural, sustainable and renewable bioresource with
160 attractive metabolic properties. Microalgal EVs are obtained under controlled environmental conditions from
161 cultures of microalgal strains, characterized by high growth rates. Picciotto et al. performed microalgal selection
162 and batch culture on seven different strains (Picciotto et al., 2021). After 30 days of incubation and a
163 differential UC purification protocol, they were able to obtain 2×10^9 particles per mL of cultivation medium
164 from *Cyanophora paradoxa*. According to Adamo et al. the production of microalgal EVs is scalable and could
165 be performed in large scale photobioreactors and obtain EVs with comparable yield to other sources (Adamo
166 et al., 2021). Algae EVs can be used to deliver biomolecules, drugs and high-value microalgal substances such
167 as antioxidants, pigments, lipids and complex carbohydrates.

168 2.3. Bovine milk EVs

169 Over the years, milk has been adopted by researchers as the main alternative EV source to human cells. There
170 is a massive amount of literature related to the use of EVs from bovine milk (Betker et al., 2019; Vashisht et
171 al., 2017). Milk is one of the most promising scalable sources of EV for mass production, because it is easily
172 accessible, inexpensive and it requires no cell culture. Several studies on the safety of milk-EVs have shown
173 low toxicity levels and a good in vivo tolerability (Manca et al., 2018). Somiya et al. found that milk-EV
174 administration in mice resulted in the induction of low cytokine levels and the absence of systemic toxicity
175 (Somiya et al., 2018). Matsuda et al. observed developmental toxicity in zebrafish embryos following
176 administration of milk-EVs loaded with RNA at high concentrations, while no acute toxicity was detected
177 (Matsuda et al., 2020). Milk-derived EVs have been shown to increase the oral bioavailability of drugs and are
178 optimal vectors to transport bioactive compounds for nutritional and therapeutics purposes (Carobolante et al.,
179 2020). In cancer therapy, milk EVs can be functionalized with ligands such as folic acid to achieve tumor
180 targeting (Munagala et al., 2016). In addition, milk-derived EVs have shown several therapeutic effects such
181 as a selective interaction with macrophages and induction of intestinal stem cell proliferation (Maghraby et al.,
182 2021). The three main steps involved in the isolation of milk EVs are milk defatting, establishing a method for
183 casein depletion, and EVs enrichment. Somiya et al. concentrated 321 μg of milk-EVs from 1 mL of whey by
184 performing casein removal through centrifugation and UC for EVs purification (Somiya et al., 2018). They
185 also attempted casein removal by acid precipitation and obtained a 20-fold lower yield. Milk-EVs can be

186 obtained from raw milk, commercial milk and dairy industry waste streams. Interestingly, others have found
187 that industrial processing of commercial milk, such as pasteurization, homogenization, and ultra-heat-treated
188 milk, impacts the integrity of milk-EVs, causing changes in their functionalities (Kleinjan et al., 2021). Sukreet
189 et al. tested the enrichment of EVs from cheesemaking byproducts by TFF, resulting in low EV count (10^9
190 particles/mL of milk), but a high protein content (0.65 mg/mL of milk). They found heterogenous EV-enriched
191 populations, which likely include components that escaped precipitation from the complex whey matrix,
192 consisting of lipoproteins, fat globules and casein micelles (Sukreet et al., 2021). Therefore, heterogeneous
193 preparations of milk EVs may be suitable for applications that do not require a high level of purity, given the
194 excellent economic and environmental advantages of using EVs derived from milk waste.



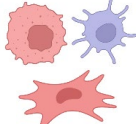


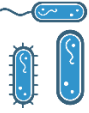
195 **2.4. Plant EVs**

196 Plant EVs are released by vegetable cells and their structure resembles that of vesicles of mammalian origin
197 (Pucci and Raimondo, 2020). To date, vesicles from ginger, grapes, grapefruit, orange, lemons, broccoli, apple,
198 kiwi, tomato, ginseng, coconut, blueberry, and carrot, among many others, have been successfully isolated and
199 observed by TEM microscopy. Over the past decade, the role of plant miRNAs as a functional component of
200 food with therapeutic effects has been investigated by many studies (Díez-Sainz et al., 2021; Sanwlani et al.,
201 2021; Teng et al., 2018). Due to their miRNA content, plant EVs are gaining attention as a new class of cross-
202 kingdom modulators, capable of mediating animal-plant interactions at the molecular level, as well as playing
203 crucial roles in plant physiology in terms of cell proliferation, differentiation and response to environmental
204 stresses (Rome, 2019). Applications of plant EVs in nanomedicine and nutraceuticals are based on their intrinsic
205 biological properties, such as anti-cancer, anti-inflammatory, anti-aging, and anti-Alzheimer's, and on their
206 use as nano-carriers to transport therapeutic biomolecules. Wang et al. demonstrated that grapefruit-derived
207 vesicles can enhance the anti-inflammatory capability of intestinal macrophages, thus alleviating dextran
208 sulfate sodium (DSS)-induced colitis in mice without any toxicity (Wang et al., 2014). Several studies have
209 revealed the role of plant vesicles in inhibiting cancer cell proliferation. Ginger-derived EVs by Zhang et al.
210 demonstrated their anti-tumor action in colitis-associated cancer. They were able to decrease the levels of
211 cancer-associated pro-inflammatory cytokines and suppress the proliferation and apoptosis of intestinal
212 epithelial cell (M. Zhang et al., 2016a). In addition, vesicles isolated from lemons by Raimondo et al. inhibit
213 the growth of several cancer cell types through tumor targeting, reduction of oxidative stress, and activating
214 of a TRAIL-mediated apoptotic cell death mechanism (Raimondo et al., 2015). Concerning the regenerative
215 effects of EVs, Sahin et al. isolated vesicles from wheat grass and investigated their potential use in wound
216 healing through *in-vitro* studies, demonstrating that they induce skin regeneration by triggering proliferation
217 in a dose-dependent manner on epithelial, endothelial, and dermal fibroblasts (Şahin et al., 2019). Furthermore,
218 Zhuang et al. studied the use of ginger-derived EVs to treat alcohol-induced liver damage in mice. These
219 vesicles were seen to contribute to hepatoprotection by suppressing the generation of reactive oxygen species
220 (Zhuang et al., 2015). In the context of industrial production, plant-EVs are extremely promising vectors for
221 drug delivery. The large volumes availability and affordability may provide easier and faster industrial
222 application than that of mammalian EVs. Like milk EVs, they are also potentially obtainable from agricultural
223 wastes and residues. Plant EVs can be loaded, by both passive and active techniques, with therapeutics such
224 as proteins, miRNAs, siRNAs and expression vectors to achieve superior effects against diseases, but also in
225 nutraceuticals and cosmetics, enhancing the beneficial action of natural bioactive phytomolecules (Wang et
226 al., 2014, 2013; M. Zhang et al., 2016). Furthermore, literature data show that plant-derived vesicles can be
227 produced in higher yields (Chen et al., 2019; Lobb et al., 2015). Of course, these comparisons are merely
228 qualitative and do not consider the variability of sources, the influence of upstream processing, the difficult
229 reproducibility of isolation procedures, and the processing of complex and heterogeneous biological matrices.
230 Importantly, it is crucial to fill the relevant knowledge gaps in the fields. More studies on plant EVs biological
231 roles are needed, as well as the determination of specific plant EVs protein markers, *in-vivo* safety, stability
232 and efficacy studies that could translate to clinical studies. There are currently five plant-EVs-based therapies
233 in clinical trials (ClinicalTrials.gov Identifiers: NCT01294072, NCT04879810, NCT01668849,

234 NCT05318898 NCT04698447). These studies are in their early stages and complete results of clinical trials
 235 using plant EVs are missing. Preliminary results have been published in only one study (ClinicalTrials.gov
 236 NCT04698447) on the use of dietary supplements containing nanovesicles derived from citrus lemon juice
 237 (CitraVes®), (Raimondo et al., 2021). The authors recruited 20 healthy volunteers who received 1000 mg/day
 238 EV CitraVes® spray-dried formulation for three months. After 4 weeks they observed a significant reduction
 239 of low-density lipoprotein cholesterol levels, an important risk factor for cardiovascular diseases. It is
 240 noticeable that in all the clinical studies cited above, guidance on EVs dosing strategies, a crucial factor in the
 241 establishing the safety and therapeutic profiles of plant EVs, was omitted.

242
 243

Table 1: Classification of EVs according to sources and their main processing characteristics..

EV classification		Cell sources	Collection/ Upstream processing	Main applications	Cell culture platforms?	Scalability potential	
Eukaryotic	Mammalian EVs	Animal EVs Milk derived EVs 	Bovine milk	Milk collection and pretreatments	Drug delivery, therapeutics, nutraceuticals	No	High due to the large availability of cow's milk
		Human EVs EVs from body fluids 	Blood, saliva, semen, urine, cerebrospinal fluids, bronchoalveolar fluid, amniotic fluid	Physiological fluids collected from the body	Diagnostics (e.g. liquid biopsies)	No	Low: need to find donors and ethical issues
		Human EVs EVs from human cells 	MSC, cancer cells, immune cells, dendritic cells, epithelial cells; cardiac, nerve, muscle, kidney, liver, intestinal cells	Collection of conditioned culture medium from cell culture expansion	Therapeutics with specific targeted functions; drug delivery for cancer therapies	Yes	Medium: high cost of cell cultures
	Plant-derived EVs 	From fruits, rhizomes, apoplasic fluids, seeds, roots	Tissue disruption and juice collection, vacuum infiltration, hydroponic medium collection	Drug delivery, therapeutics, nutraceuticals, cosmetics	No	High: Easy availability and low cost of sources	
	Algae EVs 	From microalgae (e.g. Cyanophora paradoxa, Tetraselmis chuii)	Microalgae strain selection and cultivation	Drug delivery, therapeutics, nutraceuticals, cosmetics	Yes	Medium: requires cell culture, cost lower than human cells	
Prokaryotic	Bacterial EVs 	Gram-positive and gram-negative bacteria	Spontaneous or induced release in a growth medium; possible genetic engineering strategies	Drug delivery, vaccines and cancer therapies	Yes	Medium: requires cell culture, cost lower than human cells	

244
 245

3. Downstream processing of EVs

246 **3.1. State-of-the-art of EVs isolation methods**

247 To date, researchers use several methods for isolating EVs on a laboratory scale. They can be classified
 248 according to the working principle on which they are based as reported in Table 2.

249 *Table 2: Classification of methods used for the isolation of extracellular vesicles according to their working principle.*

Methods based on size and buoyant density

- Ultracentrifugation-based techniques
- Size exclusion chromatography (SEC)
- Microfiltration/Ultrafiltration
- Flow field fractionation (FFF)

Methods based on solubility changes

- Precipitation with polyethylene glycol or protamine or sodium acetate

Methods based on charge

- Anion Exchange Chromatography (AEX)
- Electrophoresis

Methods based on highly specific surface interactions

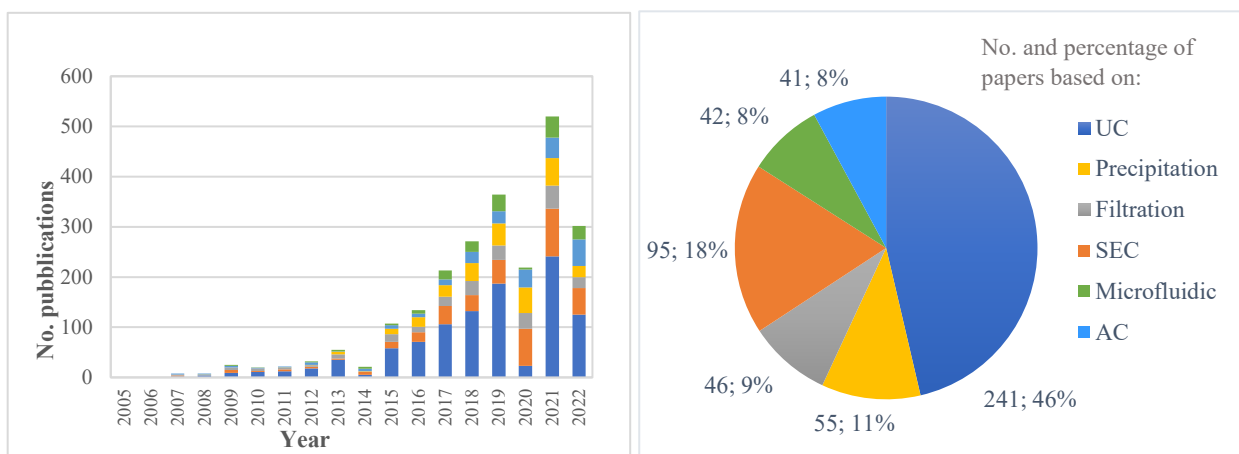
- Immuno-affinity capture
- Affinity Chromatography (AC)

Microfluidic technologies

- Immuno-affinity based microfluidics
- Viscoelastic separation
- Microfluidic filtration
- Acoustic devices

250

251 The traditional methods used for isolating EVs are those based on vesicle size and density, namely UC,
 252 filtration techniques and size exclusion chromatography (SEC). Methods based on EVs solubility changes,
 253 such as chemical precipitation, have emerged later over the years. In addition, numerous methods for isolation
 254 of EVs populations based on highly specific interactions with molecules exposed on the surface of EVs or
 255 microfluidic technologies have recently appeared. The number of publications on the isolation of EVs has
 256 increased exponentially over the past decade, as shown in Figure 3a, where the number of publications found
 257 in PubMed with the search keyword “EVs isolation methods” for the years 2010-July 2022 is shown.



258
 259
 260
 261
 262

Figure 3: (a) Number of publications on the isolation of EVs in recent years. (b) In 2021, the total number of publications on the isolation of EVs was 520. Among them, 241 papers used UC as the primary method of EV isolation, 95 papers used SEC, 55 papers used precipitation techniques, 46 papers used filtration processes, 42 used microfluidics technologies and 41 affinity capture (source PubMed, July 2022).

263 Considering the year 2021, a pie chart that reports the worldwide distribution of different methods used for
 264 EVs primary isolation is shown (Figure 3b). From the figure it can be seen that UC remains the predominant

265 isolation method (46%) adopted by researchers, while the other half of the pie is divided among SEC (18%),
 266 precipitation (11%), filtration techniques (9%), AC (8%) and microfluidic technologies (8%). It should be
 267 emphasized that the above statistics refer only to the “primary” isolation method, whereas usually researchers
 268 use a combination of different techniques to obtain EVs preparations. In fact, according to the 2019 worldwide
 269 survey on the methods for separation and characterization of EVs, more than half (60%) of the respondents
 270 use a combination of different isolation techniques in their protocols (Royo et al., 2020). Each separation
 271 process has resulted in unique characteristics of EVs and has advantages and disadvantages. Table 3 provides
 272 a comprehensive list of the advantages and disadvantages of the currently most widely used techniques for
 273 downstream processing of EVs, considering factors such as process time, potential for scalability, and cost-
 274 effectiveness. Clearly, it is not possible to entrust the entire production of EVs to a single isolation strategy,
 275 and downstream processing is strictly dependent on the desired application and the characteristics of the source
 276 material. The Minimal Information for Studies of Extracellular Vesicles (MISEV2018) conference outlined
 277 the key guidelines for EV research and standardization and proposed a very intuitive distinction between
 278 different EV isolation methods, to be placed on a *specificity vs. recovery* grid (Figure 4), (They et al., 2018).
 279 If EVs are to be used as diagnostics, the need for high EV yields is paramount, whereas high structural integrity
 280 may not be necessary. In contrast, for drug delivery applications, preserving the structure of EVs is a priority.
 281 In the case of highly complex samples such as biofluids, multiple purification steps may be necessary. UC is
 282 considered the golden standard in EV isolation. Ultracentrifuges are widely distributed in non-specialized
 283 laboratories, and the massive amount of literature available on differential UC protocols easily allows
 284 comparison with new separation processes. However, the technique has many limitations, such as the negative
 285 impact on EV integrity and aggregation, co-isolation of non-EV impurities, and low reproducibility. Standard
 286 commercial ultracentrifuges can process up to 400 mL of samples, thus the low sample throughput does not
 287 allow for scalability (Staubach et al., 2021).

288
289

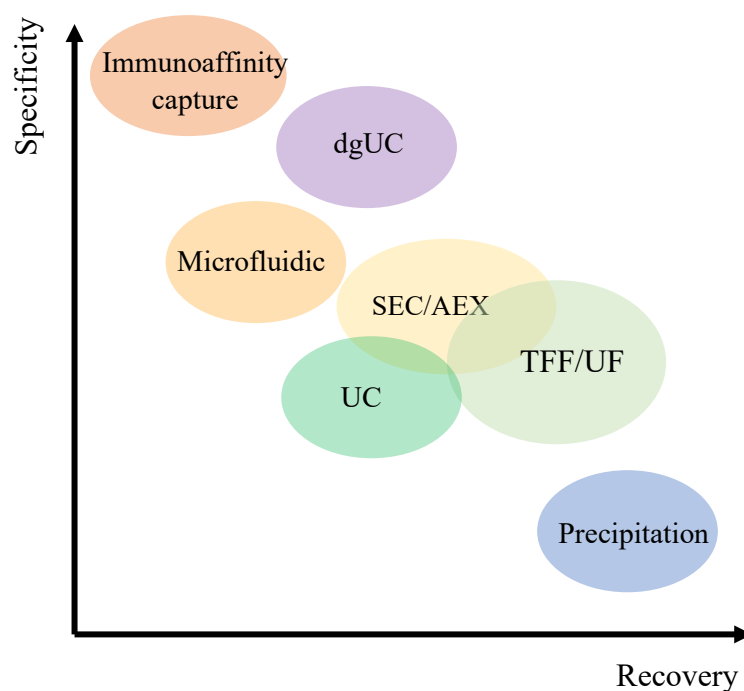
Table 3: Comparison of the most commonly used techniques for EVs isolation.

	Principle	Time	Advantages	Disadvantages	Scalability ¹	Cost ¹
Ultracentrifugation (UC)	Sedimentation of biomolecules according to density using high g-force	140 - 600 min (Greening et al., 2015; Théry et al., 2006)	<ul style="list-style-type: none"> - Easy protocol - No additional chemicals - Most common method in the field for data comparison 	<ul style="list-style-type: none"> - Low throughput - Efficiency affected by many factors - Low reproducibility - Possible damage of EVs - Long duration - Limited to small-scale 	+	€€€
Density gradient ultracentrifugation (dg UC)	Separation according to density in a pre-constructed density gradient medium	250 min – 2 days (Greening et al., 2015)	<ul style="list-style-type: none"> - Higher EVs purity than UC - No additional chemicals 	<ul style="list-style-type: none"> - Complex - Low throughput - Efficiency affected by many factors - Operator-dependent yields - Time consuming - Possible damage of EVs - Limited to small-scale 	+	€€€
Size exclusion chromatography (SEC)	Separates by hydrodynamic volume	1 mL/min (Lobb et al., 2015)	<ul style="list-style-type: none"> - Reproducibility - Reduced contamination - Gentle method - Prevents EV aggregation - No additional chemicals 	<ul style="list-style-type: none"> - Low resolution - Limitations on sample volume - Dilution of EV isolates - Co-isolation of same-size particles 	++	€€

Filtration (MF/UF)	Uses membranes with specific pore sizes	130 min (Salih et al., 2014)	<ul style="list-style-type: none"> - Simple procedure - High throughput - Time efficient - Relatively gentle - No additional chemicals 	<ul style="list-style-type: none"> - Membrane clogging - Loss of sample and aggregation - Low purity - Possible deformation of vesicles. 	++++	€
Flow Field Fractionation (FFF)	Flow modulated by a normal force field	45-60 min (Liangsupree et al., 2021)	<ul style="list-style-type: none"> - Reproducible - Removal of lipoproteins - Non-invasive 	<ul style="list-style-type: none"> - Low input volume 	+	€€
Polymeric precipitation	Solubility changes by adding a crowding agent	8-12 h (Liangsupree et al., 2021)	<ul style="list-style-type: none"> - Inexpensive - Simple - Gentle method - High yield 	<ul style="list-style-type: none"> - Need to remove the crowding agent - High contamination - Time-consuming 	++++	€
Anion exchange chromatography (AEX)	Separation based on charge	180 min (Heath et al., 2018)	<ul style="list-style-type: none"> - Scalability - Short processing time - Structural and biological integrity or EVs 	<ul style="list-style-type: none"> - Co-isolation of other negatively charged biomolecules - Need of a final concentration step. 	+++	€€
Electrophoresis	Separation based on electrophoretic mobility in an electric field	60-120 min (Marczak et al., 2018)	<ul style="list-style-type: none"> - Easy control - Fast and efficient - Non-invasive 	<ul style="list-style-type: none"> - Sample heating - Co-isolation of negatively charged biomolecules - Combination with other techniques may be required 	++	€
Affinity capture (AC)	EVs capture using antibodies or other ligands	240 min (Greening et al., 2015)	<ul style="list-style-type: none"> - High purity - Target specific populations - Great potential in diagnostics 	<ul style="list-style-type: none"> - Costly - Harsh elution - Limited knowledge of EVs markers - Isolation of a subset of EVs - Non-specific binding 	+++	€€€
Microfluidics technologies	Flow manipulation in microscale	60-120 min (Meng et al., 2021)	<ul style="list-style-type: none"> - Specificity and selectivity - Low energy and material requirements - Quick 	<ul style="list-style-type: none"> - Low sample loading - Possible blockage due to system clogging 	++	€€€

290 ¹ Qualitative criteria based on bioprocess engineering knowledge on unit operations and established processes

291



292
293
294

Figure 4: Specificity vs recovery grid; qualitative chart constructed according to ISEV recommendations on EV isolation techniques (They et al., 2018).

295

296 3.2 Main challenges in EV isolation techniques and process scalability

297 Many factors must be considered as essential requirements for the scalability of the EV process. Among them,
 298 the need for reproducible, cost-effective, and high-throughput isolation methods is critical. The methods
 299 chosen must comply to GMP standards in order to support large-scale manufacturing. The main challenge in
 300 GMP of EVs is quality control, and identification of the Critical Quality Attributes (CQAs) that affect the
 301 stability and efficacy over time of preparations, as well as standardization of sample collection, handling and
 302 storage (Chen et al., 2020; Herrmann et al., 2021). Standardization requirements must address several
 303 challenges associated with EV isolation (Table 4). First, the product of interest is present in complex biological
 304 fluids or matrices that contain a myriad of bioparticles. The biological samples contain protein assemblies or
 305 lipoproteins, with similar size and biological properties to EVs. Co-isolates may provide a synergistic effect
 306 to the actions of EVs. Often, when subjected to rigorous characterization, it is not necessary to consider them
 307 as “impurities”, but rather to speak of an EV-enriched secretome as an end product (Wiklander et al., 2019).
 308 This strategy saves the high costs associated with achieving a high level of sample purity in downstream
 309 processing. In addition, a single EV sample contains heterogeneous populations, as EVs from the same source
 310 can be released from parent cells through various biogenesis pathways, leading to the simultaneous presence
 311 of various EVs subpopulations. Therefore, heterogeneity in EVs content can result in intra- and inter-batch
 312 variabilities, which must be taken into account in the isolation procedures.

313
314

Table 4: Overview of key process optimization strategies for EV separation to advance process scale-up.

Main limitations on EVs downstream processing	Process optimization strategy
<i>There is no single best isolation method</i>	<ul style="list-style-type: none"> - Sample and application-driven decisions; - Fit the process constraints to the sample type and the specific purpose.
<i>EVs are heterogeneous in nature</i>	<ul style="list-style-type: none"> - Define a method target;

	- Decide whether to focus on specific EVs properties or general physical/chemical characteristics.
<i>Batch-to-batch variability</i>	- Define and control the most important process parameters.
<i>Regulatory requirements</i>	- Define GMP compliant raw materials; - Define storage and administration strategies; - Identify CQAs; - Define a viral inactivation step.
<i>Difficult characterization of the final product</i>	- Define potency assays; - Establish the product <i>mode of action</i> .
<i>Co-isolation/ impurities</i>	- Characterization of co-isolates is a requirement; - A possible synergic effect between EVs and co-isolates needs to be evaluated; - Prioritize therapeutic efficacy over purity, depending on the application.
<i>Low product yield</i>	- Establish an optimal trade-off between yield and purity; - Switch to the EVs sources with a higher scalability potential; - Optimize the upstream processing technologies; - Switch from lab-scale techniques (e.g. UC) to large scale techniques already exploited in other industrial bioprocesses (e.g. TFF, SEC, AEX, AC).
<i>Throughput limitations</i>	- Use downstream processing technologies that can process several ten or hundred liters of conditioned media / starting material.

315

316

317

318

319

320

321

322

323

324

325

326

327

328

329

330

331

332

333

334

335

336

337

338

339

340

341

342

343

344

A common weakness of current isolation methods is the very low yield of vesicles. According to Haraszti et al. a dose of 10^9 – 10^{11} exosomes per mouse is typically required for a single test in mice models. This quantity is approximately obtained from one liter of conditioned culture medium, with current practices such as UC (Haraszti et al., 2018). The low EV yields severely limit the preclinical and clinical development of EV applications in medicine, as well as their industrial translation to other applications. In this context, considering upstream processing, yield improvements can be achieved by changing the EV source and/or bioreactor system, in case of EVs from cell culture supernatant. As for downstream processing, yield improvements can be achieved by changing and/or optimizing purification techniques. It is essential to take advantage of the knowledge previously gained in the fields of industrial production of liposomes, monoclonal antibodies (mAbs) and viral vectors, thus applying the same downstream processing strategies in the processing of EVs. Liposomes are the synthetic equivalent of EVs, having a comparable phospholipidic bilayer nanostructure. Since TFF is considered the golden standard in the field of industrial liposome production (Paganini et al., 2019), given the similarities between EVs and liposomes, TFF can be considered the most suitable unit operation for large-scale EV production. Viral vectors, in particular enveloped viruses, and EVs share similar properties, such as size, morphology and composition. At the industrial level, virus purification is mainly achieved through a combination of chromatography and membrane-based processes (Staubach et al., 2021). A common platform for downstream processing of viral vectors is based on AEX purification, UF concentration/diafiltration and polishing with SEC. Also in this field, the use of TFF as main capture/purification step for upscaling purposes is increasing (Geraerts et al., 2005). Industrial capture and purification of mAbs relies on the use of sequential chromatographic steps including AC and AEX, as well as the use of centrifugation, depth filtration, and/or microfiltration for clarification. Ultrafiltration/Diafiltration (UF/DF) performed in TFF mode is used as unit operation for concentrating and purifying mAbs solutions (Buyel et al., 2017; Tripathi and Shrivastava, 2019). Indeed, the use of TFF/chromatography multistep processes for EVs isolation is becoming increasingly adopted. Recently, Seo et al. proposed a large-scale purification protocol for EVs preparation using TFF and AEX: they isolated Cytotoxic T-lymphocyte EVs from 4 L of culture supernatant using hollow fiber TFF with 750 kDa polyether sulfone (PES) membranes and AEX. Two distinct subpopulations were observed, exosome-like particles that eluted at low NaCl concentration (2×10^{12} particles/mL), and microvesicle-like particles that eluted at high NaCl concentration (1.5×10^{12} particles/mL). Through AEX, they demonstrated to be able to distinguish between different

345 functional EV subpopulations (Seo et al., 2022). A comparison between EVs, viral vectors and liposomes
 346 actual production systems is reported in Table 5.

347 *Table 5: Comparison between EV, viral vectors and liposome production systems in terms of upstream and downstream processing*
 348 *techniques and product yields.*

		Upstream		Downstream			Yield	Ref.
		Source	Cell culture	Harvest/clarification	Capture	Purification		
Small scale	EVs	Human cells	T-flasks	C	UC	UC		
		HEK293	T75	100 x g 10 min, 1000 x g 10 min, 10000 x g 1 h	100,000 x g 3 h	200,000 x g 3 h		10 ⁸ -10 ⁹ particles/mL of CM (Lee et al., 2019)
Large scale	EVs	Human cells	HF bioreactor	C + MF	UF/DF TFF			
		HEK293 + miRNAs	FiberCell®	1000 x g 30 min + 0.85 µm	500 kDa PES HF module			10 ¹³ particles/mL of CM (Yoo et al., 2018)
	Virus	Human cells + plasmids	T-flasks	Chemical lysis, nuclease	UF/DF	UC	UF/DF, sterile filtration	
		293T	2 x 10-layer cell factory		TFF 100 kDa cassette	76,000 x g, 2h		94 % recovery (Geraerts et al., 2005)
Liposome	Lipids + cargo proteins	Ethanol injection	-	UF TFF	DF TFF	Sterile filtration		
	DPPC + rh-Cu/Zn-SOD protein	Crossflow triple injection	-	PS 100 kDa TFF cassette			3.6 mg entrapped protein/mL (Wagner et al., 2002)	

349 List of abbreviations: C= Centrifugation; DF= Diafiltration; HF = Hollow fiber; CM = Conditioned media; UF = Ultrafiltration; MF =
 350 Microfiltration; PES = Polyethersulfone; PS = Polysulfone; DPPC = Dipalmitoyl-phosphatidylcholine.

351

352 There are several companies emerging in the production of EV-based therapeutics from human cell lines.
 353 Codiak Bioscience is a clinical-stage biopharmaceutical company focused on the development of engineered
 354 EVs-based therapeutics. Their production system is based on fed-batch or perfusion bioreactors, having
 355 volumes up to 2000 L and 500 L, respectively, to cultivate genetically engineered immortalized human cells.
 356 In downstream processing, clarification is performed through filtration steps, while purification is
 357 accomplished through filtration (UF/DF in TFF mode) and different chromatographic steps, such as cation and
 358 anion exchange chromatography (CEX, AEX) and mixed mode chromatography (MMC). All processes, CEX,
 359 AEX and MMC can be also performed with membrane chromatography. They claim to produce amounts of
 360 purified EVs 2000 times more than can be obtained with conventional centrifuges (Bourdeau et al., 2021).
 361 EVOX Therapeutics is a biotechnological company devoted to the development of protein and nucleic acid-
 362 based therapeutics via exosome engineering. Their proprietary exosome manufacturing processes are based on
 363 batch and perfusion bioreactors to cultivate genetically engineered human cell lines, downstream processes
 364 utilizing filtration processes and liquid chromatography (e.g., AEX, SEC). Recently the company patented an
 365 Affinity Chromatography (AC) purification method wherein EVs are engineered to achieve highly specific
 366 binding. In particular, the company invention involves the use of chromatography matrices comprising Fc
 367 domains and the development of engineered EVs presenting Fc binding polypeptides on their surface
 368 (Raymond et al., 2021). EVOX was recently able to scale its production up to 2000 L under GMP conditions.
 369 ExoCoBio is another exosome-based biomedicine company focusing on regenerative medicine and aesthetics.
 370 They developed a technological platform called ExoSCRT™ for the large-scale production of EVs from MSC
 371 derived from adipose tissue entirely based on filtration processes. Briefly, it includes the use of 0.22 µm PES
 372 filters for clarification, concentration and subsequent diafiltration by TFF with a 500 kDa Molecular Weight
 373 Cut Off (MWCO) membrane (Lee et al., 2020).

374 3.3 Recent developments and challenges in affinity technology

375 TFF followed by AC and final polishing steps are the most promising approaches for clinical development of
376 high-purity EVs (Colao et al., 2018). In this context affinity technology holds a remarkable potential for large
377 scale EVs purification, as the technique allows for tunable specificity depending on the adopted ligand.
378 Moreover, this field has recently seen important progress in the development of innovative stationary phases,
379 such as magnetic microbeads, chromatographic membranes, monolithic columns and microfluidic devices.
380 Recent advances in the manufacturing of human EVs (Ströhle et al., 2022), should be also considered in
381 processing EVs originating from alternative sources. As biological knowledge advances, the exploitation of
382 affinity techniques for large-scale purification of EVs from milk, plants, bacteria and algae will become
383 increasingly likely. The use of antibodies that specifically target protein receptors on the surface of human
384 EVs is perhaps the most traditional, with several studies dealing with antibodies targeting the protein markers
385 CD9, CD63, and CD81 on the surface of EVs. However, as in all immunoaffinity techniques, the main
386 drawback is the need for alkaline or acidic elution buffers, which can damage the integrity of EVs (Ströhle et
387 al., 2022). The use of aptamers has emerged as a viable alternative to antibody-based AC. Like antibodies,
388 aptamers have been developed to bind human EVs protein markers. Importantly, they provide for intact EVs,
389 as they require milder elution conditions (e.g., saline solutions). Besides, they offer a greater chemical stability
390 and a higher affinity for EVs, due to genetic modifications of the oligomer filaments (Ströhle et al., 2022). The
391 use of antibody and aptamer ligands requires specific selection and modification strategies and their application
392 on the field of non-human EVs is hindered by the lack of knowledge of EVs markers. To date, Alix,
393 tetraspanins (CD9, CD63, CD81), heat shock proteins (HSP70, HSP90) and annexins are the most frequently
394 used mammalian EVs protein markers (Deng and Miller, 2019). Interestingly, some proteins families are
395 common to different EVs biological domains, such as heat shock proteins and annexins, that have been
396 identified also in plant EVs (Pucci and Raimondo, 2020). To our knowledge, affinity purification strategies
397 applied to plant and algae EVs have not been attempted yet. Concerning OMVs general protein markers have
398 not yet been identified, but OmpA protein in *E.coli* has been explored as target receptor for affinity purification
399 (Alves et al., 2017). Specifically, mutant OmpA-His6 OMVs were created through the incorporation of a non-
400 native histidine amino acid repeat sequence (His-tag). These plasmids were spiked into a culture of native
401 OMVs and purified utilizing immobilized metal affinity chromatography (IMAC). Affinity techniques based
402 on pseudo-ligands, phospholipid membrane properties, and generic biochemical properties have also been
403 developed for the purification of human EVs (Ströhle et al., 2022). These are more versatile approaches,
404 compared to the use of antibodies and aptamers, as they do not require any specific knowledge on the EVs
405 markers, thus they have a relevant potential for the purification of non-mammalian EVs. For example, as
406 certain phospholipids are associated to the membranes of an entire EV population, their recognition allow to
407 purify the whole EVs spectra of a sample rather than specific subpopulations, a matter that is commonly
408 involved with the use of antibodies. Nakai et al. obtained highly purified EVs from conditioned culture media
409 and biofluids by using Tim4, a transmembrane protein that works as a receptor for the phosphatidylserine
410 present on the EVs surface (Nakai et al., 2016). EVs elution is simply achieved by adding a Ca^{2+} chelating
411 buffer, given that Tim4-binding to phosphatidylserine is dependent on Ca^{2+} concentration. Recently,
412 Morozumi et al. carried out a comparative study using membrane-affinity and phosphatidylserine-affinity
413 isolation for cow milk EVs (Morozumi et al., 2021). Membrane affinity was conducted using an exoEasy Maxi
414 Kit (Qiagen), based on a membrane affinity spin column. According to the producers, the method is based on
415 a generic biochemical feature of EVs, to recover all the EV populations present in a sample.
416 Phosphatidylserine-affinity isolation was performed using a MagCapture Exosome Isolation (Fujifilm Wako
417 Pure Chemical Corp). A proprietary substance was applied to the EVs sample, fostering the binding to
418 phosphatidylserine groups on EVs surface, in a calcium dependent manner. Streptavidin magnetic beads were
419 used to immobilize EVs for capture. Overall, the EV preparations isolated with phosphatidylserine-affinity had
420 a higher level of purity compared to those obtained with the membrane affinity isolation. Notably, in both
421 cases, the particle concentration was lower than that obtained with SEC. Following another strategy, Kim et
422 al. exploited the negatively-charged molecules present on plasma EVs surface by using poly-l-lysine coated
423 on magnetic beads (Kim and Shin, 2021). To remove contaminating proteins, they used a buffer having a pH
424 equal to their isoelectric point, which allowed the so-neutralized proteins to be released in solution. Final EVs
425 elution was accomplished through 1 M NaCl, obtaining a 6.6-fold higher yield compared with that of UC.
426 Another interesting affinity strategy is based on the use of heparin, that is a glycosaminoglycan ligand isolated

427 from animal tissues. Heparin is widely used to purify a range of proteins and viruses. It is not dependent on an
428 affinity-tag mechanism and it acts like a cation exchanger. A recent study evaluated the purification of stem
429 cell-derived EVs through TFF and heparin affinity chromatography, the affinity step had a minimum recovery
430 of 68.7% compared to a 39.8% recovery using SEC, based on particle counts, besides an average recovery of
431 98% and 99% of residual proteins and DNA, respectively (Barnes et al., 2022). Heparin AC was also used to
432 separate EVs in distinct subpopulations. Overall, the study found a partial interaction between heparin and
433 EVs, indicating that some populations can bind EVs and others cannot. These affinity differences may be used
434 for fractionation between subpopulations of EVs once the mechanism of interaction between EVs and heparin
435 is better elucidated.

436

437 **4. Membrane based-techniques for EVs isolation**

438 Membrane processes are the most versatile, as they can be exploited for clarification, concentration, and
439 purification of fluids, and they can be used alone or in combination with chromatography. They are scalability-
440 oriented. as modular systems allow the plant to be adapted to handle high volumes of fluids, offering different
441 levels of functionalization and flexibility. This section provides an overview on the main membrane-based
442 techniques used for EVs processing.

443 **4.1. Microfiltration and Ultrafiltration**

444 Filtration is a popular size separation technique used for both volume reduction and purification of EVs.
445 Microfiltration (MF) membranes have pore sizes in the order of micrometers, and when clarifying EVs
446 solutions by MF, filters with pore sizes of 3, 0.8, 0.45, 0.22, and 0.1 μm are typically used (Konoshenko et al.,
447 2018). Ultrafiltration (UF) employs more selective membranes, with defined molecular MWCO ranging from
448 10 to 600 kDa for most applications. Recovery of EVs based on filtration techniques can be accomplished
449 through different isolation protocols. MF and UF are often used in combination with other techniques, for
450 example as a complement to UC protocols or as additional steps in SEC. However, MF and UF are also
451 applicable as stand-alone techniques, as both UF and MF membranes can be exploited in sequential MF/UF
452 isolation protocols: they rely on a series of filtration steps for EV enrichment. First, larger impurities (cells,
453 cell debris, apoptotic bodies) are removed using MF filters, leaving a vesicle-rich permeate. Lower molecular
454 weight impurities (free proteins, contaminants) are then eliminated by using UF membranes with smaller pores
455 than the target EVs (0.22 μm , 0.1 μm , 600 kDa, 500 kDa, 100 kDa); they are able to retain vesicles and remove
456 impurities into a waste permeate. In this way, the EV fraction of a given size is concentrated and purified
457 (Konoshenko et al., 2018). For EVs concentration, their dimension should be larger than the MWCO of the
458 membrane by a factor of 2 to 5 (Scott and Keith, 1995). The selection of a tighter membrane (5) will yield
459 maximum EVs recovery with a lower flowrate. On the other hand, if processing time is a major concern the
460 selection of a loose (2) membrane should be preferred. EVs have heterogeneous dimensions depending on
461 their source, biogenesis and processing conditions. Following this rule of thumb, and assuming a correlation
462 between the EVs diameter (D) and molecular weight (MW) like $D \propto MW^{1/3}$ to isolate small EVs (e.g.,
463 exosomes) having average dimensions of 20 nm, an UF membrane having a MWCO between 200-500 kDa
464 should be selected. This is only a rough estimate; it would be helpful if the average pore size of the membrane
465 could be provided by membrane manufacturers, along with the MWCO. Merchant et al. proposed a MF
466 protocol for urinary exosomes using a 0.1 μm hydrophilized polyvinylidene difluoride filter. They compared
467 the EVs isolated from the membrane-based protocol with standard UC and obtained comparable EVs protein
468 yields and reduced contamination by non-EVs proteins, (Merchant et al., 2010). Heinemann et al. developed
469 an optimized sequential UF/MF protocol for the isolation of EVs from cell culture media or body fluids. The
470 first step involves prefiltration in dead-end mode with a 0.11 μm modified PES membrane, to remove cells
471 and cell debris. Microvesicles larger than 0.1 μm should also pass through the filter because of their flexibility.
472 The second step is based on a 5-times TFF with a 500 kDa MWCO PES membrane to remove free proteins
473 and contaminants and to concentrate the sample. In the final step a filtration with a 0.1 μm track-etched
474 polycarbonate membrane for final enrichment of exosomes is performed at very low pressure to filter out
475 microvesicles larger than 0.1 μm (Heinemann et al., 2014). Based on the sequential UF protocol, many

476 companies have recently developed kits for the isolation of EVs. ExoMir™ from Bio Scientific Corporation
477 uses two membranes (200 nm and 20 nm) both placed in a syringe that allows rapid fractionation of exosomes
478 and larger membrane-bound particles (Doyle and Wang, 2019). ExoTIC (Exosome total isolation chip)
479 developed by Liu et al. is also based on the same principle: it is a solid device that houses a track-etched
480 polycarbonate membrane (30 nm or 50 nm pore size) and a PES filter (200 nm pore size). It enables the
481 purification of intact EVs in the 30–200 nm size range from various biological fluids (Liu et al., 2017). Both
482 kits help make filtration-based exosome isolation a more reproducible and clinically simpler procedure. It is
483 important to note that all the UF techniques mentioned in this section are small scale techniques, relative to
484 the filtration of small sample volumes (< 250 mL). The development of large-scale UF techniques is mainly
485 conducted in TFF mode, and it will be covered in Section 4.3.

486

487 **4.2. Centrifugal UF**

488 In centrifugal UF, the g-force applied on the centrifuge rotor provides the driving force to remove solvents and
489 small molecules through an UF membrane. Centrifugal UF is usually carried out in centrifugal concentrators,
490 centrifuge tubes packed with a membrane filter, usually suitable for small volumes, ranging from 100 µL to
491 200 mL. Cheruvanky et al. demonstrated rapid enrichment of urinary EVs using a centrifugal concentrator
492 with 100 kDa PES membranes by centrifugation at 3000g, (Cheruvanky et al., 2007). Lobb et al. it have shown
493 that centrifugation-based filters recover three times more particles from conditioned media than pressure-
494 driven UF stirred cells. They found that centrifuge-based concentrators work well for small volumes (50-200
495 mL), while pressure-driven concentration is more appropriate with volumes greater than 400 mL, to reduce
496 the gel layer formation by generating a convective crossflow motion across the membrane, (Lobb et al., 2015).
497 The main challenge in UF processes is clogging and entrapment of vesicles on the membrane surface, which
498 slows down the process and causes partial loss and aggregation of the target material. Membrane fouling is
499 common and unavoidable in all filtration operations, but its formation can be limited and controlled through
500 optimization of fluid dynamics, identification of an optimal membrane cut-off and materials, such as those
501 with low non-specific protein adsorption.

502

503 **4.3. Tangential flow filtration (TFF)**

504 In conventional filtration systems, fluid flow is applied perpendicularly to the membrane, which causes particle
505 accumulation, unpredictable change in the hydrodynamic resistance of the membrane, and membrane
506 clogging. In TFF mode, on the other hand, the feed flows tangentially across the membrane, and membrane
507 fouling is significantly limited compared with dead-end mode. It can be controlled by achieving steady
508 conditions that ensure constant flux and cake thickness over time. Depending on the membrane MWCO, TFF
509 can be applied to purify EVs from larger particles or from smaller impurities. In addition, it can be configured
510 as buffer exchange in diafiltration mode or volume reduction to concentrate the product in the retentate stream.
511 Busatto et al. applied TFF to isolate EVs from cell culture medium with a 500 kDa PES hollow fiber membrane.
512 EVs can be concentrated and purified from a scalable sample volume with a high recovery rate in a rapid and
513 sterile manner (Busatto et al., 2018). Comparative assessment of TFF and UC revealed that the former
514 concentrates EVs with comparable physicochemical characteristics, but with 5-fold higher yield, improved
515 batch-to-batch consistency, and less albumin contaminants in half the processing time (1 h). In contrast, the
516 study by Heath et al. underlined that TFF provides EVs with lower purity than UC, detecting co-isolated lipids
517 and proteins, despite having a higher yield (Heath et al., 2018). Moreover, one aspect that should be further
518 evaluated is the potential deformation and lysis of EVs caused by shear forces. Overall, it can be observed that
519 the high degree of flexibility offered by the TFF technique allow to preserve EV integrity through optimization
520 of process conditions (e.g., transmembrane pressure (TMP), agitation speed, feed flowrate, feed
521 concentration). Some authors demonstrated that, under optimal operating conditions, TFF is a gentler method
522 than UC for liposome purification, (Dimov et al., 2017). In this context, the selection of an appropriate TMP

523 appears crucial. The work done by Dehghani et al. offers an example of an optimized TFF isolation protocol
524 for EV concentration from large volumes of fluid that involves standardization of the membrane cleaning step.
525 The authors developed a filtration-based microfluidic system called tangential flow for analyte capture
526 (TFAC), which is a modified version of TFF. In this three-step protocol: particles are first trapped on the
527 surface of a membrane in tangential flow, then washed under the same flow conditions with a cleaning buffer
528 to remove contaminants; finally, the TMP is reversed, releasing the particles from the membrane that are
529 collected downstream (Dehghani et al., 2019). According to the authors, processing human plasma in TFAC
530 mode enabled the capture of EVs with minimal contamination. Conventional TFF systems are single isolation
531 units with only one type of membrane, which does not allow isolation of specific size ranges of EVs. Kim et
532 al. proposed a dual cyclic filtration system (dcTFF) consisting of two TFF modules with 200 and 30 nm
533 membranes, connected to two peristaltic pumps that provide continuous circulation while preventing clogging.
534 The authors created a simultaneous dual flow condition that allowed them to isolate a specific size range of
535 extracellular vesicles (30-200 nm) in a single step. The two modules were assembled to form three chambers:
536 a sampling chamber, an isolation chamber and a waste chamber. They obtained active EVs with 1.3-fold more
537 abundant CD63 exosome marker than a commercial filtration kit (K. Kim et al., 2021). TFF processes are
538 modular and fully adaptable to continuous operations. They can be considered as a hybrid of concentration
539 and purification strategies, which is highly suitable for large-scale EV isolation from diluted samples. In
540 addition, industrial-scale input volumes can be used as crossflow filtration units, as they can hold volumes on
541 the order of liters.
542

543 **4.4. Microfluidic filtration**

544 Recent advances in the science of microfabrication have led to the development of microfluidic devices,
545 compact units composed of a network of microchannels that are intended to control fluid flow at the
546 microscale. Microfluidic devices enable highly efficient and precise separation of micro- or nano-sized
547 particles within a given volume of fluid. Indeed, at the micro- and nano- scale fluids possess distinctive
548 properties, with frictional forces dominating kinetic forces. This offers the possibility of fine tuning and
549 manipulating various process and material-related parameters. These devices are commonly referred to as *Lab-*
550 *on-Chip*, i.e., capable of reproducing different laboratory processes on a single integrated micrometric
551 platform, a *chip*. They thus offer high accuracy and specificity in the isolation of EVs and, compared to other
552 conventional methods, allow a substantial reduction in the number of samples, reagents and time required for
553 experiments, while increasing process automation. The most relevant microfluidic techniques recently
554 developed for EV isolation are microfluidic filtration, immunoaffinity capture, chip centrifugation, acoustic
555 separation, viscoelastic flow, and hydrodynamic flow. Microfluidic filtration (Mf-F) is a very promising tool
556 for continuous separation and enrichment of EVs according to specific EV sizes. Davies et al. developed two
557 types of pressure- and electrophoresis-driven Mf-F devices, that separate cells, debris and small EVs from
558 blood through a nanoporous membrane with an adjustable pore size. The limitation of pressure-driven Mf-F is
559 that the pores become blocked after obtaining approximately 4 μ L of filtrate. Electrophoresis avoids this
560 problem and increases the separation efficiency and purity (Davies et al., 2012). Double microfluidic filtration
561 approaches have also been developed. Liang et al. constructed a Mf-F double-filtration system that includes a
562 filter with a pore size of 200 nm to remove cells and large impurities, and a second filter with 30 nm pore size
563 that allows proteins to pass through. This system achieves high yields, compared with UC, for isolation of 30–
564 200 nm EVs, (Liang et al., 2017). Mf-F small scales are greatly advantageous in terms of reagent use and
565 precise flow control. These features are particularly exploitable in bioprocess development, as they offer the
566 ability to precisely direct process scale-up and scale-out, study and optimize fluid dynamic conditions, and
567 perform quality control. To increase the throughput, microfluidic systems can either be scaled-out or scaled-
568 up. Process scale-out is accomplished through parallelization. Many authors argue that by following this
569 strategy, Mf technologies are indefinitely scalable (Webb et al., 2020). However, these designs are expensive,
570 especially in terms of nanofabrication requirements, as well as requiring separate sets of pumps and controls.

571 In contrast, microfluidics scale-up involves increasing channel size in order to increase product throughput.
572 The key to successful scale-up of a microfluidic process is the creation of a scale-independent process that
573 maintains the optimal flow characteristics created at the microscale on larger scales, regardless of channel size.
574 Webb et al. studied the use of microfluidic devices for continuous production of loaded liposomes, from bench
575 scale (12 mL/min) to GMP volume production (200 mL/min), using different micromixer cartridge designs
576 (Webb et al., 2020). With a particular design (toroidal mixer design) they achieved a scale-independent
577 production process, ensuring homogeneous nanoparticle production over a range of flow rates and volumes
578 using the same process production parameters.

579

580 **4.5. Flow field fractionation (FFF)**

581 Field-Flow Fractionation (FFF) is a size-based isolation technique that has been applied in the field of EVs
582 isolation. Asymmetric Flow Field-Flow Fractionation (AF4) is the most widely used sub-technique of FFF. In
583 AF4 separation is achieved by diffusion of particles flowing in a sub-millimetric thin film of laminar flow
584 confined in a narrow chamber with a membrane at the bottom. A force field is applied perpendicular to the
585 laminar flow and pushes the particles toward the UF membrane, which subsequently permeate according to
586 their size. The feed flow has a parabolic profile because a constant laminar flow is employed (Zhang and
587 Lyden, 2019). In addition, AF4 has a programmable crossflow intensity that can be optimized to increase the
588 separation efficiency, making the process very flexible. Unlike elution in SEC, smaller particles elute first,
589 followed by larger particles. This is because the smaller particles have a higher diffusion coefficient. The main
590 disadvantage of the method is the low volume of sample input, as the field and membrane can be overloaded
591 at high volumes. Usually, these devices are coupled with online detectors such as UV, dynamic light scattering
592 (DLS) and multi-angle light scattering (MALS) for particle size distribution detection (Gandham et al., 2020;
593 Liangsupree et al., 2021). AF4 can successfully separate EVs from lipoproteins and is becoming attractive for
594 fractionation of EV subpopulations. Zhang and colleagues fractionated EVs into distinct subclasses: small
595 exosomes (60–80 nm), large exosomes (90–120 nm) and discovered a new subpopulation of non-membranous
596 nanoparticles that they called “exomeres” (35 nm) from various cell types. According to them, AF4 is a highly
597 reproducible, rapid, simple, label-free and gentle process, (Zhang et al., 2018). Moreover, they isolated
598 different subpopulations of exosomes in a single AF4 run with real-time measurements of various physical
599 parameters of individual particles, showing that AF4 can also be an important additional analytical tool.

600

601 **4.6. Membrane techniques combined with charge-based techniques**

602 One potential isolation strategy could be to combine filtration techniques with charge-based separation
603 methods, taking advantage of the negative surface charge that most EVs possess. Yang et al. recently developed
604 a method for isolating lemon-derived EVs that combines an electrophoretic technique with a dialysis bag of
605 300 kDa MWCO for isolating plant EVs (Yang et al., 2020). With the application of an electric field, impurities
606 and non-vesicular proteins were able to pass through the 300 kDa membrane, while lemon vesicles were
607 retained and thus purified. The electrophoretic buffer was changed every 30 minutes, and the electrophoretic
608 direction was reversed to prevent the membrane pores from being blocked by the vesicles. They obtained a
609 preparation highly enriched in lemon vesicles in only 2.5 hours, demonstrating that the method is efficient for
610 isolating lemon EVs, saving time and without the need for special equipment. The main drawback of
611 electrophoretic separations is the heat generated during the process due to the huge amount of electric field
612 required for efficient separation. This can be potentially detrimental to the vesicles. Marczak et al. addressed
613 this problem by combining electrophoresis with an ion membrane process in a continuous configuration
614 performed in a microfluidic chip. The applied electric field allows EVs to migrate to a cationic membrane. The
615 pores of the agarose gel are in the order of 200-300 nm in size and prevent large particles, such as cell debris,
616 from entering. These are washed away by the continuous flow provided by the pump, minimizing membrane
617 clogging. EVs are concentrated and trapped on the membrane surface, as they do not enter it, as they are both

618 negatively charged (Marczak et al., 2018). The cationic membrane allows the concentration and isolation of
619 exosomes, while electrophoresis allows their purification. A comparison was made with UC and a commercial
620 precipitation reagent kit. The authors found a recovery rate of 70-80%, while in comparison, from the same
621 source, UC and precipitation achieved recoveries of 6% and 11%, respectively.
622

623 **5. Isolation of Plant EVs**

624 The isolation of plant-derived EVs (PDEVs) can be very challenging because plants, fruits, seeds, and roots
625 are complex matrices consisting of different tissues with peculiar physical structures. UC has gained
626 benchmark status in the isolation and purification of EVs from plant and mammalian sources. To date, the UC
627 isolation protocol is mainly applied for the isolation of plant vesicles. The starting point is the extraction of
628 plant juice, which is then subjected to a series of centrifugation steps with gradually increasing speed. At each
629 step, the pellet is discarded and the supernatant is further processed. In the final step, the supernatant undergoes
630 further higher speed UC of at least 100,000 g to obtain a pellet rich in EVs. The pellet containing EVs is
631 subsequently resuspended and washed in a small amount of phosphate buffer. After this basic UC procedure,
632 the resulting product is often contaminated with nucleic acids and protein aggregates (Dad et al., 2021).
633 Therefore, for further purification, the homogenized suspension is subjected to ultracentrifugation in a sucrose
634 gradient (dgUC) at a high speed of more than 150,000 g for 120 minutes.

635 To obtain ultra-pure EVs the high-speed UC cycle can be repeated several times. Although this is advantageous
636 for achieving purity of EVs, it reduces the PDEVs concentration yield. In addition, repeated pelleting of EVs,
637 under the high centrifugal force of differential UC, can compromise the structural integrity of vesicles and
638 cause agglomeration (Dad et al., 2021). A comprehensive overview of the main results obtained so far in the
639 isolation of plant EVs is presented in Table 6. So far, the vast majority of EVs have been isolated by UC
640 methods, and the same drawbacks reported for purification of mammalian EVs also apply here. As an
641 alternative to UC/dgUC for isolation of plant vesicles, Kalarikkal et al. developed a method for purification of
642 ginger EVs based on polyethylene glycol-6000 (PEG6000). Using different concentrations of PEG6000, the
643 authors were able to recover between 60 and 90% of EVs compared with the UC method. PEG-EVs exhibit
644 almost identical composition, size and zeta potential to UC obtained vesicles, (Kalarikkal et al., 2020). PEG
645 precipitation methods can provide a scalable and cost-effective alternative to purify plant EVs with high yields,
646 although contamination by non-EV proteins and the need for additional cleaning steps to remove PEGs are
647 limiting factors (Iravani and Varma, 2019). Bokka et al. explored the use of SEC to purify tomato-derived EVs
648 (Bokka et al., 2020). The authors compared the performance of UC/SEC and UC/dgUC methods for the
649 isolation of tomato EVs and found that while gUC allowed for the collection of distinct subpopulations of EVs,
650 SEC provided a higher level of purity of EV products. You et al. used UF to reduce juice volume and SEC as
651 the main purification step to isolate EVs from different types of cabbage. Interestingly, they compared the
652 yield and purity of cabbage-EVs obtained by UC and precipitation with PEG (You et al., 2021). The authors
653 concluded that the SEC/UF method was superior to the other methods, reporting similar yields (10×10^9
654 particles/ μg of protein for SEC derived EVs) but consistently higher purity values. Of all the methods
655 mentioned, filtration techniques are easy and fast and have a great potential in biomanufacturing of plant
656 vesicles. So far, TFF for isolation of plant EVs has only been used in combination with other techniques such
657 as UC. Kim et al isolated EVs from aloe vera peels by coupling UC and TFF. In particular, they used a standard
658 UC protocol followed by UF using a 0.22 μm filter and a TFF concentration with a 300 kDa membrane. They
659 recovered 5.35×10^9 particles/mL of aloe vera juice, (M. K. Kim et al., 2021). Further work should be directed
660 toward the development of filtration techniques that can be suitable alternatives to UC, and not just additional
661 purification steps.

Table 6: Review of the literature on nanovesicles (NVs) and microvesicles (MVs) of plant origin obtained, reporting the method of isolation, physical and biological properties, yield and particle number (when available).

Source	Part	Isolation method	Diameter [nm]	Yield	Particle Number	Cell uptake	Stability and biological activity	Ref.
Ginger	Rhizome	dUC/gUC	102 – 998 (mean ~386 and ~294)	NA	NA	Uptake by primary Hepatocytes	Very stable in stomach-like and small intestine-like solutions	(Zhuang et al., 2015)
Ginger	Rhizome	PEG precipitation	100-900 (mean ~400)	2-3.8 g/kg	NA	Uptake by the murine macrophages; protects cells from H ₂ O ₂ induced oxidative stress.	/	(Kalarikkal et al., 2020)
Grape	Fruit	dUC/gUC	50-300 (mean 380.5 ± 37.47)	NA	NA	Uptake by mouse intestinal stem cells	/	(Ju et al., 2013)
Grapefruit	Fruit	dUC/gUC	105-390 (mean 210.8 ± 48.62)	NA	NA	Uptake by mouse intestinal macrophages	Very stable at 37 °C	(Wang et al., 2014)
Grapefruit	Fruit	dUC/gUC	180-200	2.21 ± 0.044 g/kg	NA	Uptake by splenic and liver cancer cells lines in mouse models	Very stable at 4 °C for more than one month and loaded with curcumin	(Wang et al., 2013b)
Tomatoes	Fruit	dUC/gUC/SEC	50–500	MVs 35.6 ± 8.6 mg/kg (protein) NVs; 25.8 ± 11.4 mg/kg (protein)	MVs 2.7 x 10 ¹⁶ particles/kg; NVs 3.8 x 10 ¹⁶ particles/kg			(Bokka et al., 2020)
Broccoli	Flower	dUC/gUC	~18 and 118.	NA	NA		Broccoli NVs administration in mice protects from intestinal inflammation and prevent colitis	(Deng et al., 2017)
Apple	Fruit	dUC	100-400	NA	1.6 x 10 ¹³ particles/L	Uptake by Caco.2 cells (intestinal epithelium)	NVs disappear when boiled or sonicated	Fujita et al.(Fujita et al., 2018)
Coconut	Fruit	dUC/MF	10-100 (Mean coconut water 59.72, milk 100)	NA	NA			(Zhao et al., 2018)
Citrus clementina	Fruit	dUC/gUC	75–345 (mean populations at 75, 120, 155)	1.67 x 10 ⁻³ g/L (protein)	1.16 x 10 ¹² particles/L juice		Significant presence of membrane transporters protein	(Stanly et al., 2019)
Citrus sinensis	Fruit	dUC	950, 480 (avg sizes)	0.178 g/L (protein)	NA			(Pocsfalvi et al., 2018)

(sweet orange)								
<i>Citrus paradisis</i> (grapefruit)	Fruit	dUC	255, 350 (avg sizes)	0.134 g/L (protein)	NA			(Pocsfalvi et al., 2018)
<i>Citrus aurantium</i> (bitter orange)	Fruit	dUC	5500, 700 (avg sizes)	0.161 g/L (protein)	NA			(Pocsfalvi et al., 2018)
<i>Citrus limon</i>	Fruit	dUC	820, 460 (avg sizes)	0.409 g/L (protein)	NA			Pocsfalvi et al. (Pocsfalvi et al., 2018)
<i>Citrus limon</i>	Fruit	dUC/MF/g UC	50-70	2.5 x 10 ⁻³ g/L	NA	Uptake by human lung carcinoma cell line and myeloid leukaemia cell line	<i>Citrus</i> NVs inhibit the growth of tumor cell lines inducing TRAIL-mediated cell death.	(Raimondo et al., 2015)
Carrot	Root	dUC/gUC	100-1000	NA	NA	Targeting properties to intestinal macrophages and stem cells	Data suggest that the vesicle size can be altered in a pH-dependent manner	(Mu et al., 2014)
Blueberry	Fruit	dUC/MF	100-900	NA	NA		* miRNA profiling of PDEVs of 11 different fruits and vegetables.	(Xiao et al., 2018)
Hami melon	Fruit	dUC/MF	100-800	NA	NA		*	(Xiao et al., 2018)
Pea	Seed	dUC/MF	100-800	NA	NA		*	(Xiao et al., 2018)
Pear	Fruit	dUC/MF	100-800	NA	NA		*	(Xiao et al., 2018)
Soybean	Seed	dUC/MF	100-700	NA	NA		*	(Xiao et al., 2018)
Orange	Fruit	dUC/MF	100-700	NA	NA		*	(Xiao et al., 2018)
Kiwifruit	Fruit	dUC/MF	10-700	NA	NA		*	(Xiao et al., 2018)
Sunflower	Seed	MF/ dUC	50-200	NA	NA			(Regente et al., 2009)
Strawberry (<i>Fragaria x ananassa</i>)	Fruit	dUC/MF	30-191	18 ± 3 µg/0.25 L juice	NA	Uptake by human MSCs preventing oxidative stress in a dose-dependent manner	Rich content of vitamin C and miRNAs cargo	(Perut et al., 2021)

665

666 6. Conclusions

667 EVs offer many therapeutic opportunities as natural nano-vectors for drug delivery applications. If they are to
668 be exploited industrially, there are several challenges to overcome in moving from the current laboratory-scale
669 research practices to reliable, GMP-compliant technologies for processing EVs on a large scale. The main
670 hurdle facing the bioprocessing of EVs is the lack of analytics, that prevents the identification of specific EVs
671 CQAs, thus hindering process development. There are many recent advances in EVs characterization
672 techniques, and global efforts should be devoted to their implementation in EVs processing protocols. An
673 example of advanced EV surface characterization technique to identify and quantify the expression of identity
674 markers is given by the study of Skovronova et al.; they performed single vesicles imaging on MSC-EVs using

675 super-resolution microscopy, allowing to characterize a large number of EVs at a single EV level. Besides,
676 ExoView chip-based analysis allowed an easy quantification and comparison of MSC-EVs markers, through
677 the evaluation of the number of particles captured on a chip coated with tetraspanins. The authors also
678 performed semiquantitative bead-based flow cytometry using a MACSPlex exosome kit (Skovronova et al.,
679 2021). Sanchez et al. developed Green Fluorescent Protein (GFP)-tagged EVs by engineering Chinese hamster
680 ovary (CHO) cells to express CD81 fused to GFP through a flexible peptide linker. The GFP-tagged EVs can
681 be identified through a fluorescence plate reader and GFP concentration can be estimated based on
682 fluorescence intensity, (Carrillo Sanchez et al., 2022). This fluorescence approach allows to estimate EVs
683 yields and track EVs recovery during purification processes, such as UF and SEC, greatly simplifying process
684 development. There is growing interest in using alternative sources to human cells, as the latter require
685 challenging and expensive cell culture and expansion. Cultivation of bacteria and algae cell is simpler and
686 cheaper, and EVs derived from these sources possess distinctive characteristics that can interest a wide range
687 of applications. EVs derived from foods, such as milk and vegetables, do not require any cell culture system,
688 are widely available, inexpensive, and can be potentially isolated from the waste streams existing industrial
689 plants. However, their use is limited by knowledge gaps and the need for extensive biological characterization
690 (e.g., definition of specific protein markers) and CQAs. To date, food derived EVs are mostly isolated using
691 UC-based protocols, achieving yields and product purity comparable to current mammalian EVs production
692 systems. The use of chromatographic separations, such as gel filtration and ion exchange chromatography, as
693 alternative isolation methods is increasing. They possess a good trade-off between recovery and product purity
694 and they are already being exploited in the field of industrial bioprocessing of mAbs, liposomes and viral
695 vectors. Affinity chromatographic techniques are particularly attractive for large-scale EVs production and
696 their recent advances applied to the purification of human EVs could be exploited in processing EVs from
697 alternative sources. For instance, the use of pseudoligands (e.g., heparin that exploits electrostatic interactions
698 on the EVs surface) or receptors for the membrane's EVs phospholipids (e.g., Tim4 for cow milk vesicles),
699 have good potential, as they guarantee high specificity and do not require knowledge of specific EVs markers.
700 In this field, membrane processes are emerging for both product concentration and purification by diafiltration
701 and have the greatest potential for scalability. They can be used as stand-alone techniques or coupled with
702 others, such as liquid chromatography, UC or polymer precipitation. Filtration processes are flexible in that
703 process parameters can be tuned and membranes can be selected to recover intact, well-defined EV
704 populations. They are fast and inexpensive and offer many opportunities for functionalization (e.g., ionic
705 membranes, affinity membranes). Here, the use of TFF for downstream processing of EVs to achieve high
706 product yield is illustrated. Future efforts should be devoted to minimize membrane fouling through the
707 development of novel filtration apparatuses aimed at optimizing fluid dynamic conditions. In this context,
708 microfluidics techniques are particularly intriguing as emerging tools for understanding and optimizing
709 membrane processes. They enable manipulation of fluid flow at the microscale, resulting in more predictable
710 systems with improved flux and selectivity, exploiting shear-induced phenomena at the membrane surface to
711 reduce particle aggregation and deposition (de Aguiar and Schroën, 2020). In the field of EVs production, *the*
712 *process defines the product* (Rathore and Winkle, 2009) and the mentioned separation techniques should be
713 designed in a product-specific context. Overall, to accelerate progress in the field, early actions are needed to
714 define quality control matrices, as standard platforms for EVs characterization and product potency assays.

715

716 **References**

717

718 Adamo, G., Fierli, D., Romancino, D.P., Picciotto, S., Barone, M.E., Aranyos, A., Božič, D., Morsbach, S.,
719 Raccosta, S., Stanly, C., Paganini, C., Gai, M., Cusimano, A., Martorana, V., Noto, R., Carrotta, R.,
720 Librizzi, F., Randazzo, L., Parkes, R., Capasso Palmiero, U., Rao, E., Paterna, A., Santonicola, P.,
721 Igljč, A., Corcuera, L., Kisslinger, A., di Schiavi, E., Liguori, G.L., Landfester, K., Kralj-Igljč, V., Arosio,
722 P., Pocsfalvi, G., Touzet, N., Manno, M., Bongiovanni, A., 2021. Nanoalgsomes: Introducing

723 extracellular vesicles produced by microalgae. *J Extracell Vesicles* 10.
724 <https://doi.org/10.1002/jev2.12081>

725 Akuma, P., Okagu, O.D., Udenigwe, C.C., 2019. Naturally Occurring Exosome Vesicles as Potential
726 Delivery Vehicle for Bioactive Compounds. *Front Sustain Food Syst.*
727 <https://doi.org/10.3389/fsufs.2019.00023>

728 Alves, N.J., Turner, K.B., DiVito, K.A., Daniele, M.A., Walper, S.A., 2017. Affinity purification of bacterial
729 outer membrane vesicles (OMVs) utilizing a His-tag mutant. *Res Microbiol* 168.
730 <https://doi.org/10.1016/j.resmic.2016.10.001>

731 Barnes, B., Caws, T., Thomas, S., Shephard, A.P., Corteling, R., Hole, P., Bracewell, D.G., 2022.
732 Investigating heparin affinity chromatography for extracellular vesicle purification and
733 fractionation. *J Chromatogr A* 1670. <https://doi.org/10.1016/j.chroma.2022.462987>

734 Betker, J.L., Angle, B.M., Graner, M.W., Anchordoquy, T.J., 2019. The Potential of Exosomes From Cow
735 Milk for Oral Delivery. *J Pharm Sci* 108. <https://doi.org/10.1016/j.xphs.2018.11.022>

736 Bokka, R., Ramos, A.P., Fiume, I., Manno, M., Raccosta, S., Turiák, L., Sugár, S., Adamo, G., Csizmadia,
737 T., Pocsfalvi, G., 2020. Biomanufacturing of Tomato-Derived Nanovesicles. *Foods* 9.
738 <https://doi.org/10.3390/foods9121852>

739 Bourdeau, R., Jang, S.C., Harrison, R., O'Neil, C., Noyes, A., 2021. Methods of producing extracellular
740 vesicles. WO2021062290A1.

741 Busatto, S., Vilanilam, G., Ticer, T., Lin, W.-L., Dickson, D., Shapiro, S., Bergese, P., Wolfram, J., 2018.
742 Tangential Flow Filtration for Highly Efficient Concentration of Extracellular Vesicles from Large
743 Volumes of Fluid. *Cells* 7. <https://doi.org/10.3390/cells7120273>

744 Buyel, J.F., Twyman, R.M., Fischer, R., 2017. Very-large-scale production of antibodies in plants: The
745 biologization of manufacturing. *Biotechnol Adv.*
746 <https://doi.org/10.1016/j.biotechadv.2017.03.011>

747 Cai, W., Kesavan, D.K., Wan, J., Abdelaziz, M.H., Su, Z., Xu, H., 2018. Bacterial outer membrane
748 vesicles, a potential vaccine candidate in interactions with host cells based. *Diagn Pathol.*
749 <https://doi.org/10.1186/s13000-018-0768-y>

750 Carobolante, G., Mantaj, J., Ferrari, E., Vllasaliu, D., 2020. Cow milk and intestinal epithelial cell-
751 derived extracellular vesicles as systems for enhancing oral drug delivery. *Pharmaceutics* 12.
752 <https://doi.org/10.3390/pharmaceutics12030226>

753 Carrasco, E., Soto-Herederó, G., Mittelbrunn, M., 2019. The role of extracellular vesicles in cutaneous
754 remodeling and hair follicle dynamics. *Int J Mol Sci.* <https://doi.org/10.3390/ijms20112758>

755 Carrillo Sanchez, B., Hinchliffe, M., Bracewell, D.G., 2022. GFP-tagging of extracellular vesicles for
756 rapid process development. *Biotechnol J* 17. <https://doi.org/10.1002/biot.202100583>

757 Chen, X., Zhou, Y., Yu, J., 2019. Exosome-like Nanoparticles from Ginger Rhizomes Inhibited NLRP3
758 Inflammasome Activation. *Mol Pharm* 16. <https://doi.org/10.1021/acs.molpharmaceut.9b00246>

759 Chen, Y.S., Lin, E.Y., Chiou, T.W., Harn, H.J., 2020. Exosomes in clinical trial and their production in
760 compliance with good manufacturing practice. *Tzu Chi Med J.*
761 https://doi.org/10.4103/tcmj.tcmj_182_19

762 Cheruvanky, A., Zhou, H., Pisitkun, T., Kopp, J.B., Knepper, M.A., Yuen, P.S.T., Star, R.A., 2007. Rapid
763 isolation of urinary exosomal biomarkers using a nanomembrane ultrafiltration concentrator.
764 *Am J Physiol Renal Physiol* 292. <https://doi.org/10.1152/ajprenal.00434.2006>

765 Chronopoulos, A., Kalluri, R., 2020. Emerging role of bacterial extracellular vesicles in cancer.
766 *Oncogene*. <https://doi.org/10.1038/s41388-020-01509-3>

767 Colao, I.L., Corteling, R., Bracewell, D., Wall, I., 2018. Manufacturing Exosomes: A Promising
768 Therapeutic Platform. *Trends Mol Med*. <https://doi.org/10.1016/j.molmed.2018.01.006>

769 Dad, H.A., Gu, T.W., Zhu, A.Q., Huang, L.Q., Peng, L.H., 2021. Plant Exosome-like Nanovesicles:
770 Emerging Therapeutics and Drug Delivery Nanoplatfoms. *Molecular Therapy*.
771 <https://doi.org/10.1016/j.ymthe.2020.11.030>

772 Davies, R.T., Kim, J., Jang, S.C., Choi, E.J., Gho, Y.S., Park, J., 2012. Microfluidic filtration system to
773 isolate extracellular vesicles from blood. *Lab Chip* 12. <https://doi.org/10.1039/c2lc41006k>

774 de Aguiar, I.B., Schroën, K., 2020. Microfluidics used as a tool to understand and optimize membrane
775 filtration processes. *Membranes (Basel)*. <https://doi.org/10.3390/membranes10110316>

776 Dehghani, M., Lucas, K., Flax, J., McGrath, J., Gaborski, T., 2019. Tangential Flow Microfluidics for the
777 Capture and Release of Nanoparticles and Extracellular Vesicles on Conventional and Ultrathin
778 Membranes. *Adv Mater Technol* 4. <https://doi.org/10.1002/admt.201900539>

779 Deng, F., Miller, J., 2019. A review on protein markers of exosome from different bio-resources and
780 the antibodies used for characterization. *J Histotechnol*.
781 <https://doi.org/10.1080/01478885.2019.1646984>

782 Deng, Z., Rong, Y., Teng, Y., Mu, J., Zhuang, X., Tseng, M., Samykutty, A., Zhang, L., Yan, J., Miller, D.,
783 Suttles, J., Zhang, H.G., 2017. Broccoli-Derived Nanoparticle Inhibits Mouse Colitis by Activating
784 Dendritic Cell AMP-Activated Protein Kinase. *Molecular Therapy* 25.
785 <https://doi.org/10.1016/j.ymthe.2017.01.025>

786 Díez-Sainz, E., Lorente-Cebrián, S., Aranaz, P., Riezu-Boj, J.I., Martínez, J.A., Milagro, F.I., 2021.
787 Potential Mechanisms Linking Food-Derived MicroRNAs, Gut Microbiota and Intestinal Barrier
788 Functions in the Context of Nutrition and Human Health. *Front Nutr*.
789 <https://doi.org/10.3389/fnut.2021.586564>

790 Dimov, N., Kastner, E., Hussain, M., Perrie, Y., Szita, N., 2017. Formation and purification of tailored
791 liposomes for drug delivery using a module-based micro continuous-flow system. *Sci Rep* 7.
792 <https://doi.org/10.1038/s41598-017-11533-1>

793 Doyle, L., Wang, M., 2019. Overview of Extracellular Vesicles, Their Origin, Composition, Purpose, and
794 Methods for Exosome Isolation and Analysis. *Cells* 8. <https://doi.org/10.3390/cells8070727>

795 Fujita, D., Arai, T., Komori, H., Shirasaki, Y., Wakayama, T., Nakanishi, T., Tamai, I., 2018. Apple-Derived
796 Nanoparticles Modulate Expression of Organic-Anion-Transporting Polypeptide (OATP) 2B1 in
797 Caco-2 Cells. *Mol Pharm* 15. <https://doi.org/10.1021/acs.molpharmaceut.8b00921>

798 Gandham, S., Su, X., Wood, J., Nocera, A.L., Alli, S.C., Milane, L., Zimmerman, A., Amiji, M., Ivanov,
799 A.R., 2020. Technologies and Standardization in Research on Extracellular Vesicles. *Trends*
800 *Biotechnol*. <https://doi.org/10.1016/j.tibtech.2020.05.012>

801 García-Manrique, P., Matos, M., Gutiérrez, G., Pazos, C., Blanco-López, M.C., 2018. Therapeutic
802 biomaterials based on extracellular vesicles: classification of bio-engineering and mimetic
803 preparation routes. *J Extracell Vesicles*. <https://doi.org/10.1080/20013078.2017.1422676>

804 Geraerts, M., Micheils, M., Baekelandt, V., Debyser, Z., Gijssbers, R., 2005. Upscaling of lentiviral vector
805 production by tangential flow filtration. *Journal of Gene Medicine* 7.
806 <https://doi.org/10.1002/jgm.778>

807 Gerritzen, M.J.H., Martens, D.E., Wijffels, R.H., van der Pol, L., Stork, M., 2017. Bioengineering
808 bacterial outer membrane vesicles as vaccine platform. *Biotechnol Adv*.
809 <https://doi.org/10.1016/j.biotechadv.2017.05.003>

810 Gerritzen, M.J.H., Salverda, M.L.M., Martens, D.E., Wijffels, R.H., Stork, M., 2019. Spontaneously
811 released *Neisseria meningitidis* outer membrane vesicles as vaccine platform: production and
812 purification. *Vaccine* 37. <https://doi.org/10.1016/j.vaccine.2019.01.076>

813 Gilmore, W.J., Johnston, E.L., Zavan, L., Bitto, N.J., Kaparakis-Liaskos, M., 2021. Immunomodulatory
814 roles and novel applications of bacterial membrane vesicles. *Mol Immunol* 134.
815 <https://doi.org/10.1016/j.molimm.2021.02.027>

816 Greening, D.W., Xu, R., Ji, H., Tauro, B.J., Simpson, R.J., 2015. A protocol for exosome isolation and
817 characterization: Evaluation of ultracentrifugation, density-gradient separation, and
818 immunoaffinity capture methods, in: *Methods in Molecular Biology*.
819 https://doi.org/10.1007/978-1-4939-2550-6_15

820 Halperin, W., Jensen, W.A., 1967. Ultrastructural changes during growth and embryogenesis in carrot
821 cell cultures. *Journal of Ultrastructure Research* 18. [https://doi.org/10.1016/S0022-5320\(67\)80128-X](https://doi.org/10.1016/S0022-5320(67)80128-X)

823 Haraszi, R.A., Miller, R., Stoppato, M., Sere, Y.Y., Coles, A., Didiot, M.C., Wollacott, R., Sapp, E.,
824 Dubuke, M.L., Li, X., Shaffer, S.A., DiFiglia, M., Wang, Y., Aronin, N., Khvorova, A., 2018.
825 Exosomes Produced from 3D Cultures of MSCs by Tangential Flow Filtration Show Higher Yield
826 and Improved Activity. *Molecular Therapy* 26. <https://doi.org/10.1016/j.ymthe.2018.09.015>

827 Heath, N., Grant, L., de Oliveira, T.M., Rowlinson, R., Osteikoetxea, X., Dekker, N., Overman, R., 2018.
828 Rapid isolation and enrichment of extracellular vesicle preparations using anion exchange
829 chromatography. *Sci Rep* 8. <https://doi.org/10.1038/s41598-018-24163-y>

830 Heinemann, M.L., Ilmer, M., Silva, L.P., Hawke, D.H., Recio, A., Vorontsova, M.A., Alt, E., Vykoukal, J.,
831 2014. Benchtop isolation and characterization of functional exosomes by sequential filtration. *J*
832 *Chromatogr A* 1371. <https://doi.org/10.1016/j.chroma.2014.10.026>

833 Herrmann, I.K., Wood, M.J.A., Fuhrmann, G., 2021. Extracellular vesicles as a next-generation drug
834 delivery platform. *Nat Nanotechnol*. <https://doi.org/10.1038/s41565-021-00931-2>

835 Hou, Y., Zhai, Y., Feng, L., Karimi, H.Z., Rutter, B.D., Zeng, L., Choi, D.S., Zhang, B., Gu, W., Chen, X., Ye,
836 W., Innes, R.W., Zhai, J., Ma, W., 2019. A *Phytophthora* Effector Suppresses Trans-Kingdom RNAi
837 to Promote Disease Susceptibility. *Cell Host Microbe* 25.
838 <https://doi.org/10.1016/j.chom.2018.11.007>

839 Ionescu, M., Zaini, P.A., Baccari, C., Tran, S., da Silva, A.M., Lindow, S.E., 2014. *Xylella fastidiosa* outer
840 membrane vesicles modulate plant colonization by blocking attachment to surfaces. *Proc Natl*
841 *Acad Sci U S A* 111. <https://doi.org/10.1073/pnas.1414944111>

842 Iravani, S., Varma, R.S., 2019. Plant-Derived Edible Nanoparticles and miRNAs: Emerging Frontier for
843 Therapeutics and Targeted Drug-Delivery. *ACS Sustain Chem Eng* 7.
844 <https://doi.org/10.1021/acssuschemeng.9b00954>

845 Jahromi, L.P., Fuhrmann, G., 2021. Bacterial extracellular vesicles: Understanding biology promotes
846 applications as nanopharmaceuticals. *Adv Drug Deliv Rev*.
847 <https://doi.org/10.1016/j.addr.2021.03.012>

848 Jiang, L., Schinkel, M., van Essen, M., Schiffelers, R.M., 2019. Bacterial membrane vesicles as promising
849 vaccine candidates. *European Journal of Pharmaceutics and Biopharmaceutics*.
850 <https://doi.org/10.1016/j.ejpb.2019.09.021>

851 Ju, S., Mu, J., Dokland, T., Zhuang, X., Wang, Q., Jiang, H., Xiang, X., Deng, Z. bin, Wang, B., Zhang, L.,
852 Roth, M., Welti, R., Mobley, J., Jun, Y., Miller, D., Zhang, H.G., 2013. Grape exosome-like
853 nanoparticles induce intestinal stem cells and protect mice from DSS-induced colitis. *Molecular*
854 *Therapy* 21. <https://doi.org/10.1038/mt.2013.64>

855 Kalarikkal, S.P., Prasad, D., Kasiappan, R., Chaudhari, S.R., Sundaram, G.M., 2020. A cost-effective
856 polyethylene glycol-based method for the isolation of functional edible nanoparticles from
857 ginger rhizomes. *Sci Rep* 10. <https://doi.org/10.1038/s41598-020-61358-8>

858 Kim, H., Shin, S., 2021. Exocas-2: Rapid and pure isolation of exosomes by anionic exchange using
859 magnetic beads. *Biomedicines* 9. <https://doi.org/10.3390/biomedicines9010028>

860 Kim, K., Park, J., Jung, J.H., Lee, R., Park, J.H., Yuk, J.M., Hwang, H., Yeon, J.H., 2021. Cyclic tangential
861 flow filtration system for isolation of extracellular vesicles. *APL Bioeng* 5.
862 <https://doi.org/10.1063/5.0037768>

863 Kim, M.K., Choi, Y.C., Cho, S.H., Choi, J.S., Cho, Y.W., 2021. The Antioxidant Effect of Small Extracellular
864 Vesicles Derived from Aloe vera Peels for Wound Healing. *Tissue Eng Regen Med* 18.
865 <https://doi.org/10.1007/s13770-021-00367-8>

866 Kleinjan, M., van Herwijnen, M.J.C., Libregts, S.F.W.M., van Neerven, R.J., Feitsma, A.L., Wauben,
867 M.H.M., 2021. Regular Industrial Processing of Bovine Milk Impacts the Integrity and Molecular
868 Composition of Extracellular Vesicles. *Journal of Nutrition* 151.
869 <https://doi.org/10.1093/jn/nxab031>

870 Konoshenko, M.Y., Lekchnov, E.A., Vlassov, A. v., Laktionov, P.P., 2018. Isolation of Extracellular
871 Vesicles: General Methodologies and Latest Trends. *Biomed Res Int*.
872 <https://doi.org/10.1155/2018/8545347>

873 Kuruvinashetti, K., Pakkiriswami, S., Pakkirisamy, M., 2020. Algal extracellular vesicles for therapeutic
874 applications, in: *Proceedings of the IEEE Conference on Nanotechnology*.
875 <https://doi.org/10.1109/NANO47656.2020.9183452>

876 Lee, J.H., Ha, D.H., Go, H.K., Youn, J., Kim, H.K., Jin, R.C., Miller, R.B., Kim, D.H., Cho, B.S., Yi, Y.W.,
877 2020. Reproducible large-scale isolation of exosomes from adipose tissue-derived mesenchymal
878 stem/stromal cells and their application in acute kidney injury. *Int J Mol Sci* 21.
879 <https://doi.org/10.3390/ijms21134774>

880 Lee, S.-S., Won, J.-H., Lim, G.J., Han, J., Lee, J.Y., Cho, K.-O., Bae, Y.-K., 2019. A novel population of
881 extracellular vesicles smaller than exosomes promotes cell proliferation. *Cell Communication*
882 *and Signaling* 17, 95. <https://doi.org/10.1186/s12964-019-0401-z>

- 883 Liang, L.G., Kong, M.Q., Zhou, S., Sheng, Y.F., Wang, P., Yu, T., Inci, F., Kuo, W.P., Li, L.J., Demirci, U.,
884 Wang, S.Q., 2017. An integrated double-filtration microfluidic device for isolation, enrichment
885 and quantification of urinary extracellular vesicles for detection of bladder cancer. *Sci Rep* 7.
886 <https://doi.org/10.1038/srep46224>
- 887 Liangsupree, T., Multia, E., Riekkola, M.L., 2021. Modern isolation and separation techniques for
888 extracellular vesicles. *J Chromatogr A* 1636. <https://doi.org/10.1016/j.chroma.2020.461773>
- 889 Liu, C., Lin, X., Su, C., 2020. Extracellular Vesicles: “Stealth Transport Aircrafts” for Drugs, in:
890 *Theranostics - An Old Concept in New Clothing [Working Title]*.
891 <https://doi.org/10.5772/intechopen.94502>
- 892 Liu, F., Vermesh, O., Mani, V., Ge, T.J., Madsen, S.J., Sabour, A., Hsu, E.C., Gowrishankar, G., Kanada,
893 M., Jokerst, J. v., Sierra, R.G., Chang, E., Lau, K., Sridhar, K., Bermudez, A., Pitteri, S.J., Stoyanova,
894 T., Sinclair, R., Nair, V.S., Gambhir, S.S., Demirci, U., 2017. The Exosome Total Isolation Chip. *ACS*
895 *Nano* 11. <https://doi.org/10.1021/acsnano.7b04878>
- 896 Lobb, R.J., Becker, M., Wen, S.W., Wong, C.S.F., Wiegmans, A.P., Leimgruber, A., Möller, A., 2015.
897 Optimized exosome isolation protocol for cell culture supernatant and human plasma. *J Extracell*
898 *Vesicles* 4. <https://doi.org/10.3402/jev.v4.27031>
- 899 Ly, N.P., Han, H.S., Kim, M., Park, J.H., Choi, K.Y., 2023. Plant-derived nanovesicles: Current
900 understanding and applications for cancer therapy. *Bioact Mater* 22, 365–383.
901 <https://doi.org/10.1016/j.bioactmat.2022.10.005>
- 902 Maghraby, M.K., Li, B., Chi, L., Ling, C., Benmoussa, A., Provost, P., Postmus, A.C., Abdi, A., Pierro, A.,
903 Bourdon, C., Bandsma, R.H.J., 2021. Extracellular vesicles isolated from milk can improve gut
904 barrier dysfunction induced by malnutrition. *Sci Rep* 11. [https://doi.org/10.1038/s41598-021-](https://doi.org/10.1038/s41598-021-86920-w)
905 [86920-w](https://doi.org/10.1038/s41598-021-86920-w)
- 906 Manca, S., Upadhyaya, B., Mutai, E., Desaulniers, A.T., Cederberg, R.A., White, B.R., Zemleni, J., 2018.
907 Milk exosomes are bioavailable and distinct microRNA cargos have unique tissue distribution
908 patterns. *Sci Rep* 8. <https://doi.org/10.1038/s41598-018-29780-1>
- 909 MARCHANT, R., PEAT, A., BANBURY, G.H., 1967. THE ULTRASTRUCTURAL BASIS OF HYPHAL GROWTH.
910 *New Phytologist* 66. <https://doi.org/10.1111/j.1469-8137.1967.tb05433.x>
- 911 Marczak, S., Richards, K., Ramshani, Z., Smith, E., Senapati, S., Hill, R., Go, D.B., Chang, H.C., 2018.
912 Simultaneous isolation and preconcentration of exosomes by ion concentration polarization.
913 *Electrophoresis* 39. <https://doi.org/10.1002/elps.201700491>
- 914 Matsuda, A., Moirangthem, A., Angom, R.S., Ishiguro, K., Driscoll, J., Yan, I.K., Mukhopadhyay, D.,
915 Patel, T., 2020. Safety of bovine milk derived extracellular vesicles used for delivery of RNA
916 therapeutics in zebrafish and mice. *Journal of Applied Toxicology* 40.
917 <https://doi.org/10.1002/jat.3938>
- 918 Meng, Y., Asghari, M., Aslan, M.K., Yilmaz, A., Mateescu, B., Stavakis, S., deMello, A.J., 2021.
919 Microfluidics for extracellular vesicle separation and mimetic synthesis: Recent advances and
920 future perspectives. *Chemical Engineering Journal*. <https://doi.org/10.1016/j.cej.2020.126110>
- 921 Merchant, M.L., Powell, D.W., Wilkey, D.W., Cummins, T.D., Deegens, J.K., Rood, I.M., McAfee, K.J.,
922 Fleischer, C., Klein, E., Klein, J.B., 2010. Microfiltration isolation of human urinary exosomes for
923 characterization by MS. *Proteomics Clin Appl* 4. <https://doi.org/10.1002/prca.200800093>

- 924 Momen-Heravi, F., Balaj, L., Alian, S., Mantel, P.Y., Halleck, A.E., Trachtenberg, A.J., Soria, C.E., Oquin,
925 S., Bonebreak, C.M., Saracoglu, E., Skog, J., Kuo, W.P., 2013. Current methods for the isolation of
926 extracellular vesicles. *Biol Chem*. <https://doi.org/10.1515/hsz-2013-0141>
- 927 Morozumi, M., Izumi, H., Shimizu, T., Takeda, Y., 2021. Comparison of isolation methods using
928 commercially available kits for obtaining extracellular vesicles from cow milk. *J Dairy Sci* 104.
929 <https://doi.org/10.3168/jds.2020-19849>
- 930 Mu, J., Zhuang, X., Wang, Q., Jiang, H., Deng, Z. bin, Wang, B., Zhang, L., Kakar, S., Jun, Y., Miller, D.,
931 Zhang, H.G., 2014. Interspecies communication between plant and mouse gut host cells through
932 edible plant derived exosome-like nanoparticles. *Mol Nutr Food Res* 58.
933 <https://doi.org/10.1002/mnfr.201300729>
- 934 Munagala, R., Aqil, F., Jeyabalan, J., Gupta, R.C., 2016. Bovine milk-derived exosomes for drug
935 delivery. *Cancer Lett* 371. <https://doi.org/10.1016/j.canlet.2015.10.020>
- 936 Ñahui Palomino, R.A., Vanpouille, C., Costantini, P.E., Margolis, L., 2021. Microbiota–host
937 communications: Bacterial extracellular vesicles as a common language. *PLoS Pathog*.
938 <https://doi.org/10.1371/journal.ppat.1009508>
- 939 Nakai, W., Yoshida, T., Diez, D., Miyatake, Y., Nishibu, T., Imawaka, N., Naruse, K., Sadamura, Y.,
940 Hanayama, R., 2016. A novel affinity-based method for the isolation of highly purified
941 extracellular vesicles. *Sci Rep* 6. <https://doi.org/10.1038/srep33935>
- 942 Paganini, C., Capasso Palmiero, U., Pocsfalvi, G., Touzet, N., Bongiovanni, A., Arosio, P., 2019. Scalable
943 Production and Isolation of Extracellular Vesicles: Available Sources and Lessons from Current
944 Industrial Bioprocesses. *Biotechnol J*. <https://doi.org/10.1002/biot.201800528>
- 945 Pang, B., Zhu, Y., Ni, J., Thompson, J., Malouf, D., Bucci, J., Graham, P., Li, Y., 2020. Extracellular
946 vesicles: The next generation of biomarkers for liquid biopsy-based prostate cancer diagnosis.
947 *Theranostics*. <https://doi.org/10.7150/thno.39486>
- 948 Peršurić, Ž., Pavelić, S.K., 2021. Bioactives from bee products and accompanying extracellular vesicles
949 as novel bioactive components for wound healing. *Molecules* 26.
950 <https://doi.org/10.3390/molecules26123770>
- 951 Perut, F., Roncuzzi, L., Avnet, S., Massa, A., Zini, N., Sabbadini, S., Giampieri, F., Mezzetti, B., Baldini,
952 N., 2021. Strawberry-derived exosome-like nanoparticles prevent oxidative stress in human
953 mesenchymal stromal cells. *Biomolecules* 11. <https://doi.org/10.3390/biom11010087>
- 954 Picciotto, S., Barone, M.E., Fierli, D., Aranyos, A., Adamo, G., Božič, D., Romancino, D.P., Stanly, C.,
955 Parkes, R., Morsbach, S., Raccosta, S., Paganini, C., Cusimano, A., Martorana, V., Noto, R.,
956 Carrotta, R., Librizzi, F., Capasso Palmiero, U., Santonicola, P., Iglič, A., Gai, M., Corcuera, L.,
957 Kisslinger, A., di Schiavi, E., Landfester, K., Liguori, G.L., Kralj-Iglič, V., Arosio, P., Pocsfalvi, G.,
958 Manno, M., Touzet, N., Bongiovanni, A., 2021. Isolation of extracellular vesicles from microalgae:
959 Towards the production of sustainable and natural nanocarriers of bioactive compounds.
960 *Biomater Sci* 9. <https://doi.org/10.1039/d0bm01696a>
- 961 Pocsfalvi, G., Turiák, L., Ambrosone, A., del Gaudio, P., Puska, G., Fiume, I., Silvestre, T., Vékey, K.,
962 2018. Protein biocargo of citrus fruit-derived vesicles reveals heterogeneous transport and
963 extracellular vesicle populations. *J Plant Physiol* 229. <https://doi.org/10.1016/j.jplph.2018.07.006>
- 964 Pucci, M., Raimondo, S., 2020. Plant extracellular vesicles: the safe for bioactive compounds.
965 *Advances in Biomembranes and Lipid Self-Assembly*. <https://doi.org/10.1016/bs.abl.2020.04.002>

- 966 Raimondo, S., Naselli, F., Fontana, S., Monteleone, F., lo Dico, A., Saieva, L., Zito, G., Flugy, A., Manno,
967 M., di Bella, M.A., de Leo, G., Alessandro, R., 2015. Citrus limon-derived nanovesicles inhibit
968 cancer cell proliferation and suppress CML xenograft growth by inducing TRAIL-mediated cell
969 death. *Oncotarget* 6. <https://doi.org/10.18632/oncotarget.4004>
- 970 Raimondo, S., Nikolic, D., Conigliaro, A., Giavaresi, G., Sasso, B. lo, Giglio, R.V., Chianetta, R., Manno,
971 M., Raccosta, S., Corleone, V., Ferrante, G., Citarrella, R., Rizzo, M., de Leo, G., Ciaccio, M.,
972 Montalto, G., Alessandro, R., 2021. Preliminary results of citravesTM effects on low density
973 lipoprotein cholesterol and waist circumference in healthy subjects after 12 weeks: A pilot open-
974 label study. *Metabolites* 11. <https://doi.org/10.3390/metabo11050276>
- 975 Ramirez, M.I., Amorim, M.G., Gadelha, C., Milic, I., Welsh, J.A., Freitas, V.M., Nawaz, M., Akbar, N.,
976 Couch, Y., Makin, L., Cooke, F., Vettore, A.L., Batista, P.X., Freezor, R., Pezuk, J.A., Rosa-
977 Fernandes, L., Carreira, A.C.O., Devitt, A., Jacobs, L., Silva, I.T., Coakley, G., Nunes, D.N., Carter,
978 D., Palmisano, G., Dias-Neto, E., 2018. Technical challenges of working with extracellular vesicles.
979 *Nanoscale*. <https://doi.org/10.1039/c7nr08360b>
- 980 Rathore, A.S., Winkle, H., 2009. Quality by design for biopharmaceuticals. *Nat Biotechnol.*
981 <https://doi.org/10.1038/nbt0109-26>
- 982 Raymond, B., Chul, J.S., Rane, H., Conlin, O., Aaron, N., 2021. Method of producing Extracellular
983 Vesicles. EP3700566B1.
- 984 Regente, M., Corti-Monzón, G., Maldonado, A.M., Pinedo, M., Jorrín, J., de la Canal, L., 2009. Vesicular
985 fractions of sunflower apoplasmic fluids are associated with potential exosome marker proteins.
986 *FEBS Lett* 583. <https://doi.org/10.1016/j.febslet.2009.09.041>
- 987 Robbins, P.D., Dorransoro, A., Booker, C.N., 2016. Regulation of chronic inflammatory and immune
988 processes by extracellular vesicles. *Journal of Clinical Investigation*.
989 <https://doi.org/10.1172/JCI81131>
- 990 Rome, S., 2019. Biological properties of plant-derived extracellular vesicles. *Food Funct* 10.
991 <https://doi.org/10.1039/c8fo02295j>
- 992 Royo, F., Théry, C., Falcón-Pérez, J.M., Nieuwland, R., Witwer, K.W., 2020. Methods for Separation and
993 Characterization of Extracellular Vesicles: Results of a Worldwide Survey Performed by the ISEV
994 Rigor and Standardization Subcommittee. *Cells* 9. <https://doi.org/10.3390/cells9091955>
- 995 Rutter, B.D., Innes, R.W., 2018. Extracellular vesicles as key mediators of plant-microbe interactions.
996 *Curr Opin Plant Biol*. <https://doi.org/10.1016/j.pbi.2018.01.008>
- 997 Şahin, F., Koçak, P., Güneş, M.Y., Özkan, İ., Yıldırım, E., Kala, E.Y., 2019. In Vitro Wound Healing Activity
998 of Wheat-Derived Nanovesicles. *Appl Biochem Biotechnol* 188. [https://doi.org/10.1007/s12010-](https://doi.org/10.1007/s12010-018-2913-1)
999 [018-2913-1](https://doi.org/10.1007/s12010-018-2913-1)
- 1000 Salih, M., Zietse, R., Hoorn, E.J., 2014. Urinary extracellular vesicles and the kidney: Biomarkers and
1001 beyond. *Am J Physiol Renal Physiol*. <https://doi.org/10.1152/ajprenal.00128.2014>
- 1002 Sangiorgio, P., Verardi, A., Spagnoletta, A., Balducchi, R., Leone, G.P., Pizzichini, D., Raimondo, S.,
1003 Conigliaro, A., Alessandro, R., 2020. Citrus as a multifunctional crop to promote new bio-
1004 products and valorize the supply chain. *Environ Eng Manag J* 19.
1005 <https://doi.org/10.30638/eemj.2020.179>

- 1006 Sanwlani, R., Fonseka, P., Mathivanan, S., 2021. Are Dietary Extracellular Vesicles Bioavailable and
 1007 Functional in Consuming Organisms?, in: *Subcellular Biochemistry*. [https://doi.org/10.1007/978-](https://doi.org/10.1007/978-3-030-67171-6_21)
 1008 [3-030-67171-6_21](https://doi.org/10.1007/978-3-030-67171-6_21)
- 1009 Schwechheimer, C., Kuehn, M.J., 2015. Outer-membrane vesicles from Gram-negative bacteria:
 1010 Biogenesis and functions. *Nat Rev Microbiol*. <https://doi.org/10.1038/nrmicro3525>
- 1011 Scott, Keith, 1995. *Handbook of industrial membranes*. . Elsevier.
- 1012 Seo, N., Nakamura, J., Kaneda, T., Tateno, H., Shimoda, A., Ichiki, T., Furukawa, K., Hirabayashi, J.,
 1013 Akiyoshi, K., Shiku, H., 2022. Distinguishing functional exosomes and other extracellular vesicles
 1014 as a nucleic acid cargo by the anion-exchange method. *J Extracell Vesicles* 11.
 1015 <https://doi.org/10.1002/jev2.12205>
- 1016 Skovronova, R., Grange, C., Dimuccio, V., Deregibus, M.C., Camussi, G., Bussolati, B., 2021. Surface
 1017 marker expression in small and medium/large mesenchymal stromal cell-derived extracellular
 1018 vesicles in naive or apoptotic condition using orthogonal techniques. *Cells* 10.
 1019 <https://doi.org/10.3390/cells10112948>
- 1020 Somiya, M., Yoshioka, Y., Ochiya, T., 2018. Biocompatibility of highly purified bovine milk-derived
 1021 extracellular vesicles. *J Extracell Vesicles* 7. <https://doi.org/10.1080/20013078.2018.1440132>
- 1022 Stanly, C., Moubarak, M., Fiume, I., Turiák, L., Pocsfalvi, G., 2019. Membrane transporters in citrus
 1023 clementina fruit juice-derived nanovesicles. *Int J Mol Sci* 20.
 1024 <https://doi.org/10.3390/ijms20246205>
- 1025 Staubach, S., Bauer, F.N., Tertel, T., Börger, V., Stambouli, O., Salzig, D., Giebel, B., 2021. Scaled
 1026 preparation of extracellular vesicles from conditioned media. *Adv Drug Deliv Rev*.
 1027 <https://doi.org/10.1016/j.addr.2021.113940>
- 1028 Ströhle, G., Gan, J., Li, H., 2022. Affinity-based isolation of extracellular vesicles and the effects on
 1029 downstream molecular analysis. *Anal Bioanal Chem* 414, 7051–7067.
 1030 <https://doi.org/10.1007/s00216-022-04178-1>
- 1031 Sukreet, S., Braga, C.P., An, T.T., Adamec, J., Cui, J., Tribble, B., Zempleni, J., 2021. Isolation of
 1032 extracellular vesicles from byproducts of cheesemaking by tangential flow filtration yields
 1033 heterogeneous fractions of nanoparticles. *J Dairy Sci* 104. [https://doi.org/10.3168/jds.2021-](https://doi.org/10.3168/jds.2021-20300)
 1034 [20300](https://doi.org/10.3168/jds.2021-20300)
- 1035 Svennerholm, K., Park, K.S., Wikström, J., Lässer, C., Crescitelli, R., Shelke, G. v., Jang, S.C., Suzuki, S.,
 1036 Bandeira, E., Olofsson, C.S., Lötvall, J., 2017. Escherichia coli outer membrane vesicles can
 1037 contribute to sepsis induced cardiac dysfunction. *Sci Rep* 7. [https://doi.org/10.1038/s41598-017-](https://doi.org/10.1038/s41598-017-16363-9)
 1038 [16363-9](https://doi.org/10.1038/s41598-017-16363-9)
- 1039 Szempruch, A.J., Sykes, S.E., Kieft, R., Dennison, L., Becker, A.C., Gartrell, A., Martin, W.J., Nakayasu,
 1040 E.S., Almeida, I.C., Hajduk, S.L., Harrington, J.M., 2016. Extracellular Vesicles from Trypanosoma
 1041 brucei Mediate Virulence Factor Transfer and Cause Host Anemia. *Cell* 164.
 1042 <https://doi.org/10.1016/j.cell.2015.11.051>
- 1043 Teng, Y., Ren, Y., Sayed, M., Hu, X., Lei, C., Kumar, A., Hutchins, E., Mu, J., Deng, Z., Luo, C., Sundaram,
 1044 K., Sriwastva, M.K., Zhang, L., Hsieh, M., Reiman, R., Haribabu, B., Yan, J., Jala, V.R., Miller, D.M.,
 1045 van Keuren-Jensen, K., Merchant, M.L., McClain, C.J., Park, J.W., Egilmez, N.K., Zhang, H.G., 2018.
 1046 Plant-Derived Exosomal MicroRNAs Shape the Gut Microbiota. *Cell Host Microbe* 24.
 1047 <https://doi.org/10.1016/j.chom.2018.10.001>

- 1048 Théry, C., Clayton, A., Amigorena, S., Raposo, and G., 2006. Isolation and Characterization of
1049 Exosomes from Cell Culture Supernatants. *Curr Protoc Cell Biol*.
- 1050 Thery, C., Laviue, G., Martin-Jaular, L., Mathieu, M., Tkach, M., Zivkovic, A.M., Zocco, D., 2018.
1051 Minimal information for studies of extracellular vesicles 2018 (MISEV2018): a position statement
1052 of the International Society for Extracellular Vesicles and update of the MISEV2014 guidelines. *J*
1053 *Extracell Vesicles* 7.
- 1054 Tripathi, N.K., Shrivastava, A., 2019. Recent Developments in Bioprocessing of Recombinant Proteins:
1055 Expression Hosts and Process Development. *Front Bioeng Biotechnol*.
1056 <https://doi.org/10.3389/fbioe.2019.00420>
- 1057 Vashisht, M., Rani, P., Onteru, S.K., Singh, D., 2017. Curcumin Encapsulated in Milk Exosomes Resists
1058 Human Digestion and Possesses Enhanced Intestinal Permeability in Vitro. *Appl Biochem*
1059 *Biotechnol* 183. <https://doi.org/10.1007/s12010-017-2478-4>
- 1060 Wagner, A., Vorauer-Uhl, K., Katinger, H., 2002. Liposomes produced in a pilot scale: Production,
1061 purification and efficiency aspects. *European Journal of Pharmaceutics and Biopharmaceutics* 54.
1062 [https://doi.org/10.1016/S0939-6411\(02\)00062-0](https://doi.org/10.1016/S0939-6411(02)00062-0)
- 1063 Wang, B., Zhuang, X., Deng, Z. bin, Jiang, H., Mu, J., Wang, Q., Xiang, X., Guo, H., Zhang, L., Dryden, G.,
1064 Yan, J., Miller, D., Zhang, H.G., 2014. Targeted drug delivery to intestinal macrophages by
1065 bioactive nanovesicles released from grapefruit. *Molecular Therapy* 22.
1066 <https://doi.org/10.1038/mt.2013.190>
- 1067 Wang, Q., Zhuang, X., Mu, J., Deng, Z. bin, Jiang, H., Xiang, X., Wang, B., Yan, J., Miller, D., Zhang, H.G.,
1068 2013a. Delivery of therapeutic agents by nanoparticles made of grapefruit-derived lipids. *Nat*
1069 *Commun* 4. <https://doi.org/10.1038/ncomms2886>
- 1070 Wang, Q., Zhuang, X., Mu, J., Deng, Z.-B., Jiang, H., Zhang, L., Xiang, X., Wang, B., Yan, J., Miller, D.,
1071 Zhang, H.-G., 2013b. Delivery of therapeutic agents by nanoparticles made of grapefruit-derived
1072 lipids. *Nat Commun* 4, 1867. <https://doi.org/10.1038/ncomms2886>
- 1073 Webb, C., Forbes, N., Roces, C.B., Anderluzzi, G., Lou, G., Abraham, S., Ingalls, L., Marshall, K., Leaver,
1074 T.J., Watts, J.A., Aylott, J.W., Perrie, Y., 2020. Using microfluidics for scalable manufacturing of
1075 nanomedicines from bench to GMP: A case study using protein-loaded liposomes. *Int J Pharm*
1076 582. <https://doi.org/10.1016/j.ijpharm.2020.119266>
- 1077 Wiklander, O.P.B., Brennan, M., Lötvall, J., Breakefield, X.O., Andaloussi, S.E.L., 2019. Advances in
1078 therapeutic applications of extracellular vesicles. *Sci Transl Med*.
1079 <https://doi.org/10.1126/scitranslmed.aav8521>
- 1080 Xiao, J., Feng, S., Wang, X., Long, K., Luo, Y., Wang, Y., Ma, J., Tang, Q., Jin, L., Li, X., Li, M., 2018.
1081 Identification of exosome-like nanoparticle-derived microRNAs from 11 edible fruits and
1082 vegetables. *PeerJ* 2018. <https://doi.org/10.7717/peerj.5186>
- 1083 Yang, M., Liu, X., Luo, Q., Xu, L., Chen, F., 2020. An efficient method to isolate lemon derived
1084 extracellular vesicles for gastric cancer therapy. *J Nanobiotechnology* 18.
1085 <https://doi.org/10.1186/s12951-020-00656-9>
- 1086 Yoo, K.W., Li, N., Makani, V., Singh, R.N., Atala, A., Lu, B., 2018. Large-Scale preparation of
1087 extracellular vesicles enriched with specific microRNA. *Tissue Eng Part C Methods* 24.
1088 <https://doi.org/10.1089/ten.tec.2018.0249>

1089 You, J.Y., Kang, S.J., Rhee, W.J., 2021. Isolation of cabbage exosome-like nanovesicles and
1090 investigation of their biological activities in human cells. *Bioact Mater* 6.
1091 <https://doi.org/10.1016/j.bioactmat.2021.04.023>

1092 Zhang, H., Freitas, D., Kim, H.S., Fabijanic, K., Li, Z., Chen, H., Mark, M.T., Molina, H., Martin, A.B.,
1093 Bojmar, L., Fang, J., Rampersaud, S., Hoshino, A., Matei, I., Kenific, C.M., Nakajima, M., Mutvei,
1094 A.P., Sansone, P., Buehring, W., Wang, H., Jimenez, J.P., Cohen-Gould, L., Paknejad, N., Brendel,
1095 M., Manova-Todorova, K., Magalhães, A., Ferreira, J.A., Osório, H., Silva, A.M., Massey, A.,
1096 Cubillos-Ruiz, J.R., Galletti, G., Giannakakou, P., Cuervo, A.M., Blenis, J., Schwartz, R., Brady, M.S.,
1097 Peinado, H., Bromberg, J., Matsui, H., Reis, C.A., Lyden, D., 2018. Identification of distinct
1098 nanoparticles and subsets of extracellular vesicles by asymmetric flow field-flow fractionation.
1099 *Nat Cell Biol* 20. <https://doi.org/10.1038/s41556-018-0040-4>

1100 Zhang, H., Lyden, D., 2019. Asymmetric-flow field-flow fractionation technology for exomere and
1101 small extracellular vesicle separation and characterization. *Nat Protoc* 14.
1102 <https://doi.org/10.1038/s41596-019-0126-x>

1103 Zhang, L., Wen, Z., Lin, J., Xu, H., Herbert, P., Wang, X.M., Mehl, J.T., Ahl, P.L., Dieter, L., Russell, R.,
1104 Kosinski, M.J., Przysiecki, C.T., 2016. Improving the immunogenicity of a trivalent *Neisseria*
1105 meningitidis native outer membrane vesicle vaccine by genetic modification. *Vaccine* 34.
1106 <https://doi.org/10.1016/j.vaccine.2016.05.049>

1107 Zhang, M., Viennois, E., Prasad, M., Zhang, Y., Wang, L., Zhang, Z., Han, M.K., Xiao, B., Xu, C.,
1108 Srinivasan, S., Merlin, D., 2016a. Edible ginger-derived nanoparticles: A novel therapeutic
1109 approach for the prevention and treatment of inflammatory bowel disease and colitis-associated
1110 cancer. *Biomaterials* 101. <https://doi.org/10.1016/j.biomaterials.2016.06.018>

1111 Zhang, M., Xiao, B., Wang, H., Han, M.K., Zhang, Z., Viennois, E., Xu, C., Merlin, D., 2016b. Edible
1112 ginger-derived nano-lipids loaded with doxorubicin as a novel drug-delivery approach for colon
1113 cancer therapy. *Molecular Therapy* 24. <https://doi.org/10.1038/mt.2016.159>

1114 Zhao, Z., Yu, S., Li, M., Gui, X., Li, P., 2018. Isolation of Exosome-Like Nanoparticles and Analysis of
1115 MicroRNAs Derived from Coconut Water Based on Small RNA High-Throughput Sequencing. *J*
1116 *Agric Food Chem* 66. <https://doi.org/10.1021/acs.jafc.7b05614>

1117 Zhuang, X., Deng, Z. bin, Mu, J., Zhang, L., Yan, J., Miller, D., Feng, W., McClain, C.J., Zhang, H.G., 2015.
1118 Ginger-derived nanoparticles protect against alcohol-induced liver damage. *J Extracell Vesicles* 4.
1119 <https://doi.org/10.3402/jev.v4.28713>

1120

1121

1122

1123