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Eradication of glioblastoma by immuno-virotherapy with a retargeted oncolytic HSV in a preclinical model

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Abstract

Oncolytic herpes simplex viruses are proving to be effective in clinical trials against a number of cancers. Here, R-115, an oncolytic herpes simplex virus retargeted to human erbB-2, fully virulent in its target cells, and armed with murine interleukin-12 was evaluated in a murine model of glioblastoma. We show that a single R-115 injection in established tumors resulted, in about 30% of animals, in the complete eradication of the tumor, otherwise invariably lethal. The treatment also induced a significant improvement in the overall median survival time of mice and a resistance to recurrence from the same neoplasia. Such a high degree of protection was unprecedented; it was not observed before following treatments with the commonly used, mutated/attenuated oncolytic viruses. This is the first study providing the evidence of benefits offered by a fully virulent, retargeted, and armed herpes simplex virus in the treatment of glioblastoma and paves the way for clinical translation.

Introduction

Herpes simplex virus 1 (HSV-1) is among the most studied oncolytic viruses for treating several types of cancers. HSV-1 has relevant characteristics as therapeutic agent: it has a large genome that can accommodate exogenous genes, does not integrate into the host genome, and is not oncogenic [1–5]. Moreover, anti-HSV-1 drugs are available to block virus replication in case of uncontrolled infection while the presence of pre-existing antibodies against the virus does not

seem to impair its infectivity [6, 7]. Oncolytic HSVs (oHSVs) are emerging as one of the efficient therapies to fight malignant neoplasms lacking effective therapeutic alternatives [8]. Clinical trials are ongoing to test the safety and the efficacy of oHSVs against colorectal, breast, and non-resectable pancreatic cancers [9–11]. Recently, T-VEC, an oHSV with tumor-selective replication that expresses granulocyte monocyte colony stimulating factor, has been approved as therapy for nonresectable metastatic melanoma [12, 13].

A noteworthy potential target for oHSV therapy is glioblastoma [14, 15], the most aggressive form of brain tumor and one of the most lethal types of human cancer, classified by WHO as grade IV glioma [16]. The life expectancy of glioblastoma patients has not substantially changed in the past 50 years. The median survival time of patients treated with the standard of care protocol is about 14 months; only 10% of them survive for more than 5 years from diagnosis [17]. The challenging properties of glioblastoma are its tendency to infiltrate the healthy brain parenchyma and its high radio- and chemo-resistance. As a consequence, the incidence of recurrence after surgical resection is high. In this picture, oncolytic viroimmunotherapy may represent a breakthrough since viral spread could potentially hit disseminating cells. Oncolytic viruses have the advantage that they can disseminate to adjacent cancer cells and kill them. In addition, they stimulate the immune system so as to break the tolerance against the

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tumor. Although for a long time the central nervous system (CNS) has been considered a privileged site of immunity, this paradigm is currently disputed and there is growing evidence that activated T cells are able to recognize their targets even when they are located within the CNS parenchyma [18, 19]. oHSVs offer the possibility to be armed with immunostimulatory molecules. In recent years, evidence that the expression of mIL-12 increases the effectiveness of oncolytic viruses by activating the immune response has been provided, thus potentiating the clearance of cancer cells through an immunotherapeutic effect [20–22]. Preclinical trials were conducted with oHSVs armed with mIL-12 for ovarian cancer [23], breast cancer brain metastases [24], and also glioblastoma [25]. An HSV armed with IL-12 is currently under the phase 1 clinical trial [26].

The above studies exploited HSV recombinants attenuated to varying degrees by genetic manipulation, in order to achieve cancer-specific replication. In a strikingly different approach, we recently assayed the safety and the efficacy of a replication-competent tropism-retargeted oHSV, fully virulent in its target cells, named R-LM113. R-LM113 is retargeted at the human receptor tyrosine-protein kinase erbB-2 (hHER2, human epidermal growth factor receptor 2), a receptor expressed in a number of tumors. hHER2 is a clinically relevant target for high-grade gliomas (HGGs) since it is expressed in a large fraction of these tumors, quantified between 15 and 80% depending on the technology employed for detection [27–29]. Malignancy of gliomas positively correlates with hHER2 expression [30, 31], which is particularly high in the CD133-positive, stem cell-like, glioblastoma cells [32]. Furthermore, hHER2 expression is virtually absent in healthy brain parenchyma, confirming its suitability and specificity as target [33]. In a murine model developed in our laboratory, R-LM113 counteracted HGG [34, 35]. Of note, the retargeted oHSVs are intrinsically safer than the attenuated ones, replicate to higher yields, and can be readily engineered so as to target different cancer-specific receptors [36].

Here, for the first time, we report on the efficacy of a fully virulent, retargeted oHSV-1, armed with mIL-12 (R-115 [37]) in counteracting glioma. The effects of treatment were tested against established orthotopic HGGs in a syngeneic glioma model in the BALB/c mouse strain.

Results

Mouse PDGF-driven glioma cells show hallmarks of human glioblastoma and are poorly immunogenic

To study the effects of the mIL-12-armed hHER2-retargeted oHSV (R-115) on HGGs, we took advantage of a model of HGG based on the transduction of platelet-derived growth

factor-B (PDGF-B) in neural progenitor cells derived from BALB/c mice, a mouse strain sensitive to HSV infection [35]. The HGG model was generated by transplanting in adult mouse brains murine neural progenitor cells explanted at embryonic day 14 (E14) and transduced with a PDGF-B overexpressing retroviral vector as described elsewhere [34]. The generated HGGs, hereinafter referred to as mHGG^{pdgf}, express markers typical of oligodendroglial progenitor cells (OPCs) and exhibit histological features typical of human HGGs, as previously described [35]. Here, mHGG^{pdgf} cells were further characterized by RNA sequencing carried on three independent tumor specimens (raw data are available on GEO Dataset. Accession number GSE109614). The analysis showed that mHGG^{pdgf} cells express OPC markers such as Olig2, PDGFR- α , and NG2 as well as stem cell markers, including Sox2, Stat3, and Nestin (Fig. 1a). mHGG^{pdgf} cells showed very low levels of mRNAs for immune system co-stimulatory molecules CD40 and CD80, and the IL-12 and IL15 cytokines. Low levels of expression were also observed for the co-stimulatory molecule CD86. On the contrary, the immunosuppressive gene CD73 [38] was highly expressed. This pattern is consistent with mHGG^{pdgf} being poorly immunogenic, as typically seen in human glioblastomas.

Primary mHGG^{pdgf} were microdissected, cultured, and engineered to express hHER2 receptor. hHER2-transduced cells (hereinafter referred to as mHGG^{pdgf}-hHER2) maintained the tumorigenic potential exhibited by the parental mHGG^{pdgf} cells, and were able to consistently generate tumor masses after intracranial transplantation of 2×10^4 cells in adult immunocompetent mice leading them invariably to death (Supplementary figure 1).

R-115 efficiently infects hHER2-positive glioma cells in vitro, and mediates mIL-12 secretion

To assess if mHGG^{pdgf}-hHER2 are susceptible to R-115 infection in vitro and to compare R-115 with R-LM113, whose efficiency was reported earlier [35], we infected mHGG^{pdgf}-hHER2 cells with R-115 or R-LM113 at the multiplicity of infection (MOI) of 0.5 plaque-forming units (PFU) per cell, as titrated in SK-OV-3 cells. Viral infection was scored as expression of the EGFP reporter gene inserted in the virus genome. After 30 h, we observed an equivalent percentage of infected cells in the R-115- and R-LM113-infected cultures (Fig. 1b, c). To ensure that R-115 exhibited the specific hHER2 tropism typical of R-LM113, the control hHER2-negative mHGG^{pdgf} monolayers were inoculated with R-115. No EGFP-positive-infected cells were detected up to 96 h after exposure to the virus (Fig. 1d). mHGG^{pdgf}-hHER2-expressing cells infected with R-115 and monitored for 72 h after infection produced and secreted mIL-12 in the medium, in a MOI-dependent

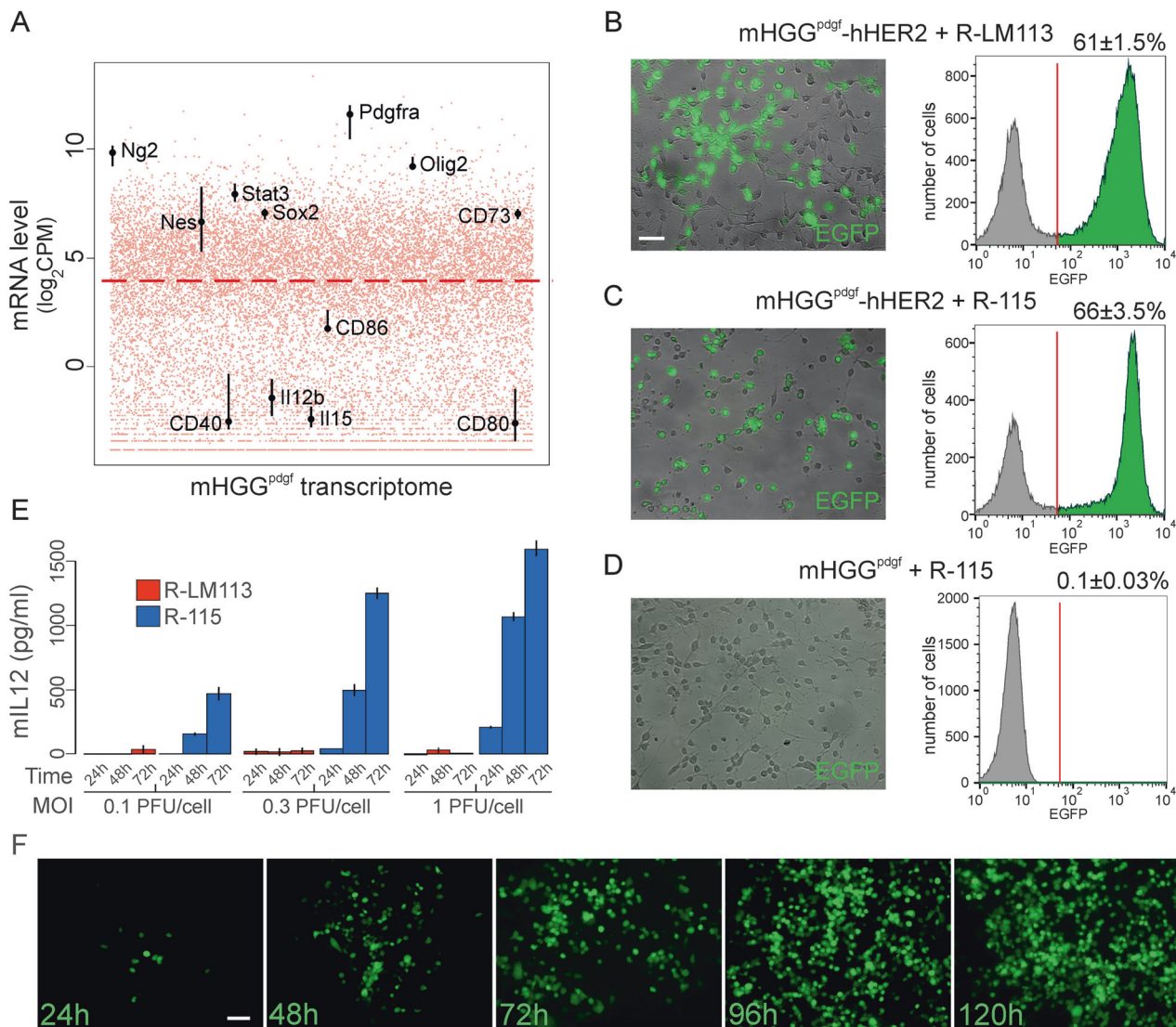


Fig. 1 **a** Expression levels of the whole transcriptome of mHGG^{pdgf} cells as determined by RNA seq analysis from three independent tumors. Genes of interest are highlighted and the level of their expression is represented as a bar summarizing the results from the three samples. Black dots represent the median. Dashed red line represents the overall median expression level across the whole transcriptome. The arrangement of the genes along the *x* axis is randomized. **b** **d** Infection of mHGG^{pdgf} hHER2 cells with 0.5 PFU/cell of R LM113 (**b**) or R 115 (**c**), or of the parental mHGG^{pdgf} cells with 0.5

PFU/cell of R 115 (**d**). Pictures were taken 30 h after infection. Histograms show the corresponding cytofluorimetric quantifications of the cells expressing the EGFP reporter expressed by the viral genome. **e** ELISA quantification of mIL12 concentration in supernatants of mHGG^{pdgf} hHER2 cells infected with R LM113 (red bars) or R 115 (blue bars) at the indicated MOI and times after infection. Error bars indicate standard deviation. **f** Time course of R-115 infection in mHGG^{pdgf} hHER2 monolayers (input MOI 0.01 PFU/cell). Scale bars: 50 μ m

fashion (Fig. 1e). In addition, we monitored the time course of R-115 infection in mHGG^{pdgf}-hHER2 cell monolayers, after infection at low MOI (0.01 PFU/cell, as titrated in SK-OV-3 cells). The collected images showed that R-115 progeny was able to efficiently spread in culture (Fig. 1f).

The ability of R-115 to infect hHER2-expressing cells is not limited to the engineered murine models. We tested R-115 on 11 independent glioma-initiating cell (hGic) cultures obtained from glioblastoma patients [39], code-named hGic-G6, -G7, -G10, -G14, -G17, -G19, -G21, -G24, -G33, -G37, and -G39. We found that they expressed

hHER2 receptor at different levels and were efficiently infected by R-115, with a higher extent in cultures displaying higher HER2 expression levels. Glioma-initiating cell cultures (hGic-G14 and -G33) that did not express hHER-2 were not infected by R-115 (Fig. 2a). In order to verify that oHSV infection leads to cell death, we performed flow cytometry-based analysis of three of hGic cultures (Fig. 2b and data not shown) by evaluating apoptosis and necrosis markers. Compared to the control, we observed the appearance of high percentages of early apoptotic (Annexin V-positive) and necrotic/late apoptotic (Annexin V- and

A

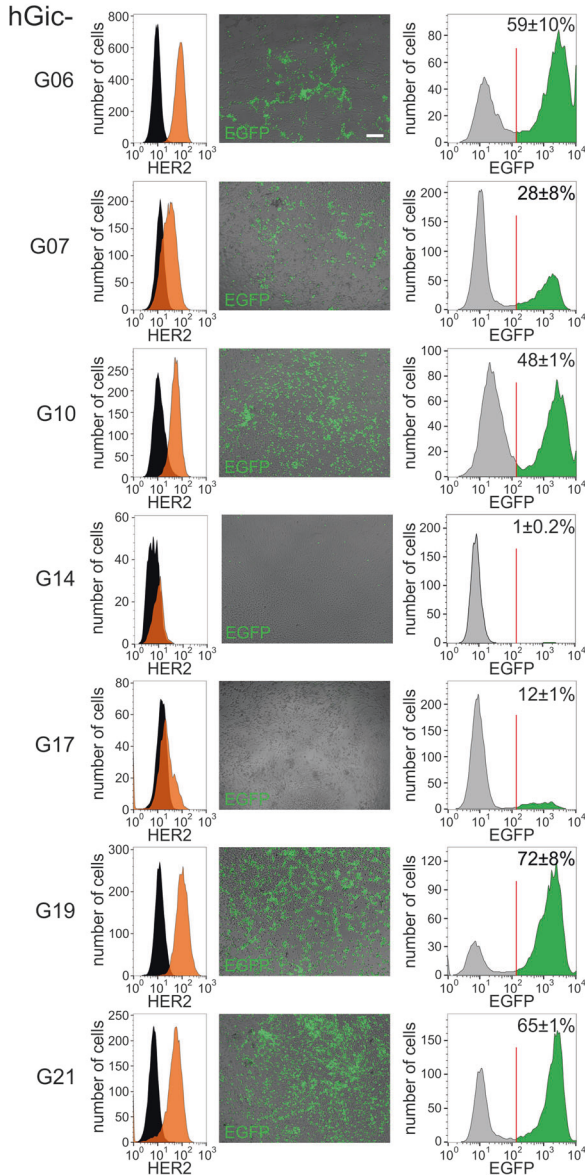
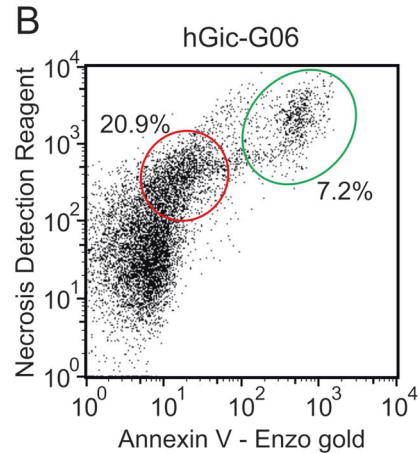
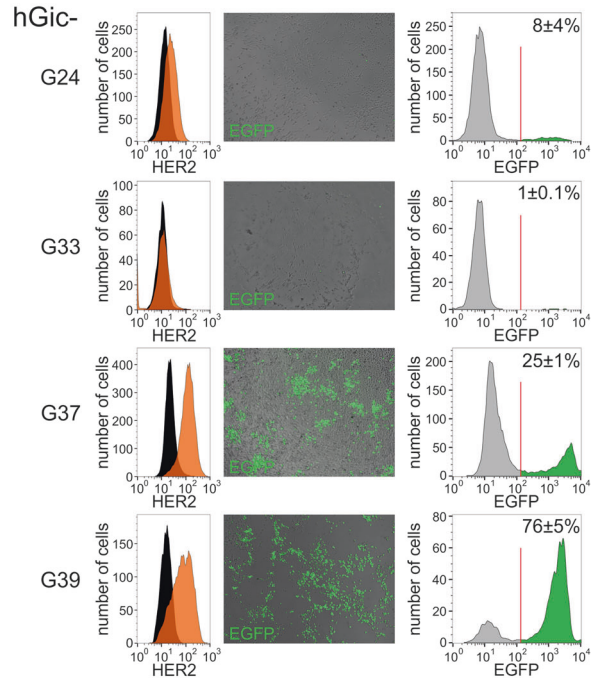


Fig. 2 a Cytofluorimetric quantification of hHER2 expression (orange peaks; negative control in black), of the indicated hGic cultures, micrograph of the corresponding cultures 40 h after infection with 5 PFU/cell of R 115 (Scale bar: 50 μ m) and cytofluorimetric quantifications of the percentage of infected cells based on EGFP reporter

Necrosis Detection Reagent-positive) cells after infection with R-115 (Fig. 2b). This is the typical cell death induced by HSV infection [40].

In vivo inhibition of tumor growth by R-115 in immunocompetent mice and induction of cancer-specific immune memory

The R-115 efficacy in counteracting gliomas was analyzed as ability to inhibit the progressive growth of established



expressed by R 115 (gray and green peaks). **b** Cytofluorimetric analysis of early apoptotic (red circle) and necroptotic/late apoptotic cells (green circle) of hGic G06 culture 120 h after infection with 5 PFU/cell of R 115

mHGG^{pdgf}-hHER2 tumors in vivo. Mice bearing tumors induced by orthotopic transplantation of mHGG^{pdgf}-hHER2 cells in the left cranial hemisphere were randomized and treated by intracranial injection of 2×10^6 PFU of R-115 (R-115 low dose arm), or the same amount of R-LM113 (R-LM113 arm), 21 days after tumor implantation. The control group was injected with an equivalent volume of gamma-irradiated R-LM113 (control arm). In an additional arm, R-115 was injected at a higher concentration (10^8 PFU; R-115 high-dose arm). The animals that did not

survive intracranial injection procedure and died within 3 days from virus injection were excluded from the experiment ($n = 1$). In total, we analyzed 14 mice in the R-115 low-dose arm, 8 in the R-115 high-dose arm, 12 mice in the R-LM113 arm and 10 mice in the control arm. Analysis of survival curves revealed no difference between the R-115 low- and high-dose arms (log rank-test power 1 $\beta \geq 0.8$ for $HR \leq 0.33$, Supplementary figure 2) and therefore the two arms were considered hereafter as a single one (R-115 arm) including 22 mice (Fig. 3a).

The median survival time of animals in the control arm was 13 days from the treatment, while that of mice belonging to the R-LM113 and R-115 arms were respectively 33 and 35 days. Survival time in both treatment arms (R-LM113 and R-115) was significantly different from that in the control arm (log rank-test $p < 0.0001$; Fig. 3a). No statistically significant difference in term of median survival time was detected between the two different oHSVs (log rank-test power 1 $\beta \geq 0.8$ for $HR \leq 0.4$). However, a dramatic difference between the R-LM113 and R-115 arms was evident in terms of long survivors. While in the R-LM113 arm all mice died within 48 days from treatment, 27% of mice treated with R-115 ($n = 6$, 4 belonging to low-dose arm and 2 to high-dose arm) were still alive 100 days after the virus treatment. Two long survivors were sacrificed and resulted tumor-free (Fig. 3b). Other two were implanted with additional 2×10^4 mHGG^{pdgf}-hHER2 cells in the contralateral hemisphere relative to the first injection and received no further virus treatment. In parallel, the same amount of cells was injected into the right hemisphere of four control mice. The animals belonging to the control set died within 26 days after transplantation. Conversely, the two R-115-treated mice were still alive 220 days after the transplant in the contralateral hemisphere (Fig. 3c). The mice were additionally orthotopically transplanted, with 2×10^4 cells from the parental mHGG^{pdgf} culture, lacking hHER2. Even in this case they did not develop gliomas, demonstrating that they had acquired a long-term resistance to the tumor cells, regardless of the presence of the exogenous hHER2 receptor. Control mice ($n = 5$) transplanted with the same cells died, as expected, within 50 days (Fig. 3d).

R-115 elicits production of antibodies and the infiltration of CD4- and CD8-positive cells in the tumor

To provide direct evidence that R-115 elicits a tumor-specific immune response, we transplanted 14 additional mice with mHGG^{pdgf}-hHER2 cells. After 21 days, six mice were injected with R-115 and 5 with R-LM113 (2×10^6 PFU/mouse). After further 20 days, we took the blood from the retro-orbital sinus to check whether the mice had

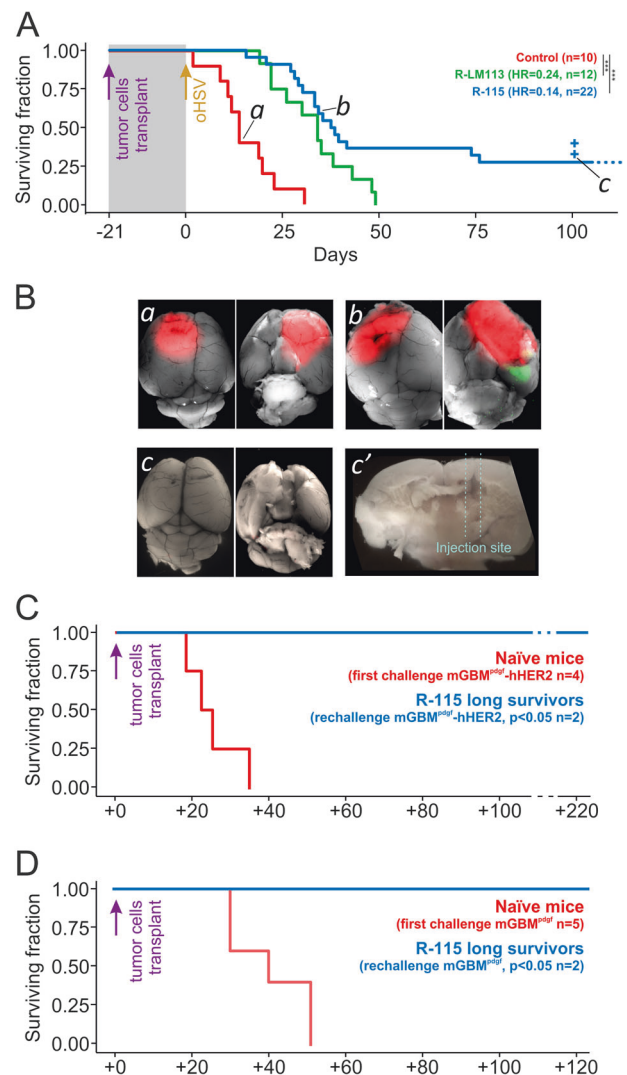


Fig. 3 **a** Kaplan Meyer survival curves of mice transplanted with mHGG^{pdgf} hHER2 and treated after 21 days with R LM113 (green line), R 115 (blue line) and gamma irradiated R LM113 (control; red line). HR are indicated for the indicated arms compared to the control arm. *** Rank test p values < 0.0001 . **b** Representative dorsal and ventral images of brains from mice of the control arm (a), the R 115 arm (b) and a long surviving mouse from R 115 arm, found to be tumor free (c). **c'**: coronal section of the brain of panel c at the level of injection site. In all micrographs, the red channel shows the DsRed fluorescent reporter expressed by mHGG^{pdgf} hHER2 cells, the green channel shows the viral reporter EGFP. **c, d** Kaplan Meyer curves of the long surviving mice from R 115 arm (blue line) rechallenged by a second transplantation of mHGG^{pdgf} hHER2 (c) or by a transplantation of the parental cells mHGG^{pdgf} lacking hHER2 receptor (d) in the contralateral hemisphere, and control naïve mice transplanted with the same cells (red line)

developed antibodies targeting mHGG^{pdgf}-hHER2 antigens. Blood was also drawn from two long-surviving mice from the R-115 arm of a previous experiment, from two non-transplanted mice (naïve), and from three mice transplanted with mHGG^{pdgf}-hHER2 cells and not treated with the virus. The plasma samples were assessed for the presence of

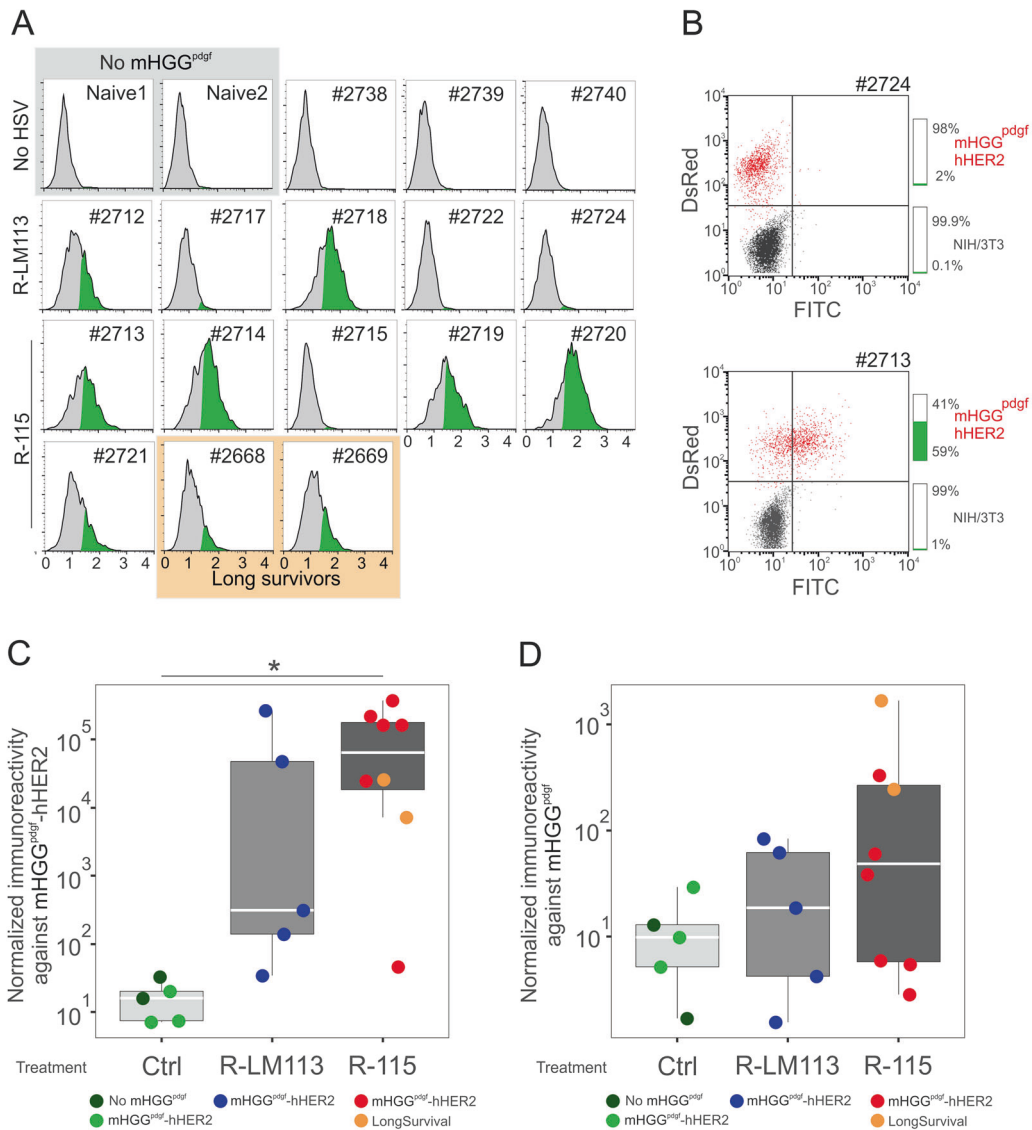


Fig. 4 a Development of antibodies against mHGG^{pdgf} hHER2 cells by mice treated with R LM113 or R 115 oHSV. Plasma samples drawn from mice treated with the indicated virus were evaluated for their ability to label mHGG^{pdgf} hHER2 cells. Green areas represent the events above the fluorescent intensity threshold determined from the negative control. **b** Specificity of the immunoreactivity of the plasma of two representative mice: evaluation of the ability of plasma samples to specifically label mHGG^{pdgf} hHER2 cells in a mixture

containing both mHGG^{pdgf} hHER2 (labeled with DsRed) and unlabeled NIH/3T3 as negative control. **c** Ratio between (specifically) labeled mHGG^{pdgf} hHER2 cells and the (unspecifically) labeled NIH/3T3, each normalized to the size of the respective population, calculated for all the plasma samples from control, R LM113, and R 115 arm (Anova $p < 0.05$), graphed as boxplot. **d** The same assay was repeated using the parental mHGG^{pdgf} cells instead mHGG^{pdgf} hHER2

antibodies against mHGG^{pdgf} hHER2 cells, by checking their reactivity to mHGG^{pdgf} hHER2 cells. The plasma samples were incubated with a cell mixture constituted by DsRed-labeled mHGG^{pdgf} hHER2 cells and unlabeled murine fibroblasts (NIH/3T3), as internal negative control. Following incubation with an FITC-conjugated anti-mouse IgG secondary antibody, the increase in FITC fluorescence of DsRed-labeled cells was measured (Fig. 4a). This analysis showed that no antibodies were detectable in the plasma of naïve (non-tumor-transplanted) mice, nor in those of mice receiving tumor cells but no virus injection. In

contrast, immunoreactivity was clearly detectable in the plasma of two out of five animals from the R-LM113 group, and in five out of six animals from the R-115 group. Immunoreactivity was also detected in the plasma of both long-surviving mice. To evaluate the specificity of immunoreactivity of plasma samples, we took advantage of the internal negative control, which allowed to discriminate between specific and unspecific FITC signal. To do so, we normalized the fraction of mHGG^{pdgf} hHER2 cells above a fixed FITC threshold to the fraction of NIH/3T3 cells above the same FITC threshold (i.e. unspecific signal; Fig. 4b).

This analysis performed on all animals allowed to test by one way Anova followed by a TukeyHSD post-hoc test the levels of immunoreactivity in the three groups. The analysis confirmed that there was a significant difference (Anova $p < 0.05$) and the difference was between R-115 and the control group (Fig. 4c). Next, we carried out the same assay with the parental hHER2-negative mHGG^{pdgf} cells. The reactivity was much lower than that against mHGG^{pdgf}-hHER2, suggesting that the immune response elicited by oHSV infection was mainly targeting the xenogeneic hHER2 receptor. However, three plasma samples exhibited a higher reactivity: they belonged to the two long-survivor mice from the R-115 arm of the first experiment, and to one of the mice that had received R-115 (Fig. 4d).

To further characterize the immune response likely boosted by IL-12 expression, we determined the isotypes of the immunoglobulins targeting glioma cells. Plasma samples drawn from mice belonging to the R-115 arm were incubated with mHGG^{pdgf}-hHER2 cells, and afterward with FITC-conjugated anti-mouse isotype-specific secondary antibodies. While only one sample was found positive for IgG1 antibodies subclass, all of them resulted positive for at least one of the two IgG2 subtypes (Supplementary figure 3), typical, in the mouse, of cytotoxic response mediated by T-helper type 1 (TH1) [41].

IL-12 is known to activate T-cell response [42]. An increase in the number of T-lymphocytes a few days after oHSVs armed with mIL-12 was reported [43]. We analyzed the brains of 11 mice treated with R-LM113 or R-115 for 22 days after virus administration. The brains were analyzed for the presence of CD4- and CD8-positive cells by immunostaining. While the overall number of CD4- and CD8-positive cells did not significantly differ between animals treated with R-LM113 or with R-115, we observed a striking difference in their intratumoral distribution, depending on the injected oHSV. Thus, CD4- and CD8-positive cells accumulated at the edge of the tumors treated with R-LM113, while they deeply infiltrated the tumor masses in the animals treated with the mIL-12 expressing R-115 (Fig. 5a-d). A quantification of the distances of CD4- and CD8-positive cells inside the tumor masses from the nearest edge of the tumor was performed on five independent tumors for each treatment (Fig. 5e, f). *T*-test confirmed a significant difference between R-LM113 and R-115 groups for both CD4 and CD8 cells tumor localization ($p < 10^{-12}$), arguing that the mIL-12 armed oHSV strongly promoted CD4- and CD8-positive infiltration of the tumor masses that, otherwise, tend to exclude lymphocytes.

To investigate whether in our experimental conditions the mIL-12 expression was inducing a similar path to that reported by other authors [25], hence suggesting a common mechanism, we assayed the expression of interferon- γ (IFN- γ) in a subset of samples treated with R-115 and R-LM113.

The immunohistochemical analysis showed that IFN- γ expressing cells were present at a density greater than 5 cells/mm² in 3 out of 4 animals treated with R-115 and only in 1 out of 4 in animals treated with R-LM113 (Supplementary figure 4). This suggests that, analogously to what already reported in other systems, mIL-12 may act by enhancing the expression of IFN- γ .

Discussion

We report that more than one fourth of mice bearing a highly invasive orthotropic high-grade glioma (mHGG^{pdgf}) at advanced stage, treated with a single injection of R-115, a hHER2-retargeted, fully virulent oHSV, armed with mIL-12, underwent permanent and complete remission and exhibited resistance to a distant re-challenge with the same tumor cells, as well as to a re-challenge with the parental mHGG^{pdgf} tumor, lacking hHER2 expression. To our knowledge, this is the first description of a complete tumor eradication achieved in this model of HGG. Even more importantly, the observation that resistance is extended to parental mHGG^{pdgf} population lacking hHER2 indicates that the R-115 treatment, by stimulating an immune response, has good chances to be effective also against tumor that display heterogeneity in hHER2 expression.

The mHGG^{pdgf} model developed in this and previous studies [35] exhibits an immunosuppressive gene expression profile, similar to that shown by embryonically PDGF-induced gliomas [44] and that of human glioblastomas. Like human glioblastomas, it is rather resistant to a number of treatments, including chemotherapy and experimental gene therapy [45], and it invariably leads to animal death. In our experience, all the different treatments tested so far (temozolomide administration, peptide immunotherapy, oHSVs) on mHGG^{pdgf} harboring mice led, at best, to an improvement in the median survival time [34, 35, 45]. So far, one of the most effective treatments was that with the fully virulent retargeted oHSV (R-LM113) [34] i.e. the unarmed parent of R-115 [37]. In the present work, as few as 2×10^6 PFU of R-115 was administered in a single injection session in animals harboring already established tumors. Noteworthy, the timing for treatment was selected to be shortly before the first animals exhibited disease symptoms, as inferred from previously determined survival curves for mHGG^{pdgf}-hHER2. Furthermore, our data show that a 50-fold increase in the amount of administered virus did not lead to higher efficacy, implying that, at least with this administration protocol, as few as 2×10^6 PFU of fully virulent mIL-12 expressing virus is already a saturating amount. Not all treated mice responded uniformly. While some mice fully recovered, others showed only an increased survival time. Heterogeneity in the response to treatment

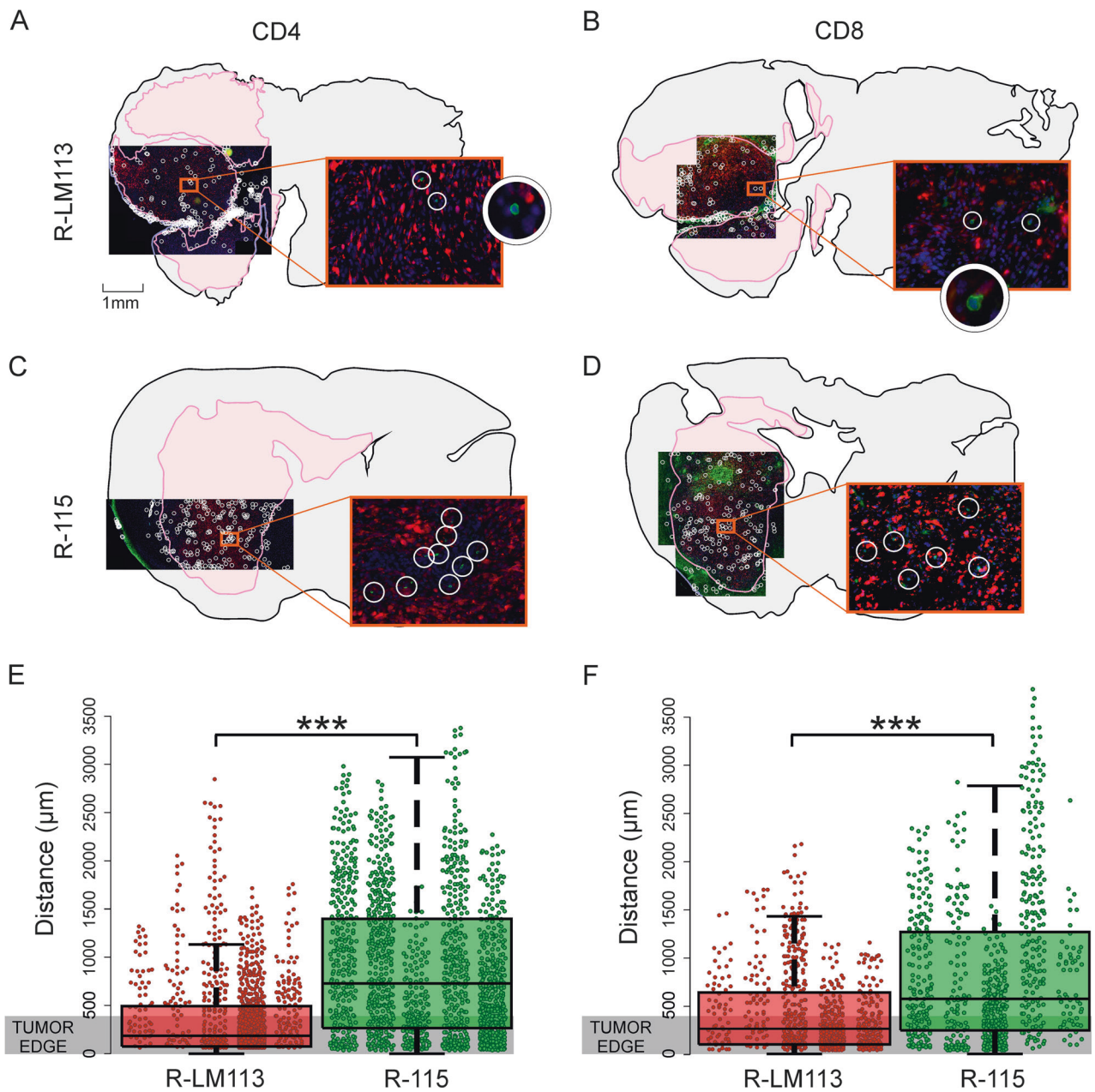


Fig. 5 Brain sections of mice transplanted with mHGG^{pdgf} hHER2 cells and treated with R LM113 (a, b) or with R 115 (c, d) were immunostained for CD4 (a, c) or CD8 (b, d). Positive cells (green) were encircled in white as shown in the inserts. Tumors are represented as pink areas. (e, f) The distance from the nearest edge of the

tumor mass of each CD4 positive cell (e) or CD8 positive cell (f) inside the tumor was measured in sections of 10 independent animals treated with R LM113 or R 115. The results are depicted as bee swarm plots and cumulative box plots (t test $p < 10^{-12}$)

may have different concurring reasons. Among the likeliest are the difference in tumor size at the time of the treatment and the stochastic nature of the immune response. From previous studies, we know that transplanted gliomas grow roughly exponentially by doubling their size in 2-4 days [46]. Therefore, although their survival time is relatively homogeneous (38 ± 8 days in the control group), the tumor size at any given time may vary substantially (from 4 to 8 doublings i.e. up to 250 times). These differences are likely

affecting the percentage of infected cells and, possibly, the intensity of the immune response, which, per se, has been early recognized highly variable even in syngeneic populations subjected to the same stimulus [47]. This concept that has been later corroborated by different studies [48]. The variability of the immune response is well illustrated by the observation that only a subset of R-115-treated mice developed a significant immunoreactivity against the non-hHER2 expressing parental population.

Our observations show that the mIL-12 expressing hHER2 retargeted R-115 enhanced the production of antibodies targeting the transplanted glioma cells. The immunoreactivity was mostly directed to the exogenous human receptor hHER2, expressed by mHGG^{pdgf}-hHER2. A subset of animals (about 1/3, among which the long survivors) showed immunoreactivity also against the parental mHGG^{pdgf} cells, lacking hHER2. Antibodies targeting tumor cells mainly belonged to the IgG2 isotype rather than the IgG1, a feature expected to be observed during a cytotoxic Th1-mediated immune response [41]. This observation fits with the enhancing of the secretion of IFN- γ , observed in tumors treated by virus carrying mIL-12²⁵ and Supplementary Fig. 4.

Our analysis on brain sections did not highlight differences in the total amount of CD4- and CD8-positive T-lymphocytes depending on treatment. However, a striking difference was evident in the distribution of these cells 20 days after virus injection. In mice treated with R-LM113, CD4- and CD8-positive lymphocytes appeared confined at the tumor edges, as observed in cold (i.e. excluded) tumors [49]. In contrast, CD4 and CD8 T-lymphocytes deeply infiltrated the tumor masses in mice treated with R-115 arguing that mIL-12 influenced the immune cell landscape within the immunosuppressive tumor microenvironment, making tumor mass more permissive to cytotoxic lymphocytes infiltration. Data observed are consistent with the observation reported by Cheema et al. that depletion of T-lymphocytes abolishes the ability of IL-12 armed oHSVs in counteracting tumors [25].

The overall median survival time of mice treated with R-115 was doubled in comparison to that of the mock-treated animals, although it was not significantly different from that of the animals treated with the non-armed version of the oHSV (R-LM113). Current findings differ from the reported efficacy study on mIL-12-armed versus non-armed versions of mutated/attenuated oHSVs, which carry the deletion of the $\gamma_{134.5}$ virulence gene [25, 43]. The main difference between our and the former results lies in the higher efficacy of R-LM113 on the overall survival time: indeed, unlike mutated/attenuated oHSVs, fully virulent unarmed R-LM113 was capable of doubling the median survival time even when injected in large neoplastic masses.

It is likely that if the protocol of R-115 administration were similar to that employed in earlier studies, which consisted of repeated sessions of injections or earlier treatments [25], the R-115 efficacy would be much higher. Recently, high percentages of experimental glioma eradication in immunocompetent mice were achieved with a mIL-12 expressing attenuated oHSV [50]. Of note, that extent of eradication was only achieved when the mIL-12 expressing oHSV was employed in simultaneous combination with two immune checkpoint inhibitors (anti-CTLA-

4 and anti-PD1 antibodies); no eradication was achieved with the mIL-12 armed o-HSV alone or the two checkpoint inhibitors in the absence of the o-HSV.

R-115 was recently recognized effective against a model of HER2-Lewis lung carcinoma [51]. This study now provides a proof of concept of the benefits of a retargeted, fully virulent, oHSVs over mutated/attenuated oHSVs to counteract HGGs by overexpressing mIL-12 and suggests they are candidates worth to be explored for oHSV-based glioblastoma treatment, most likely in combination with immune checkpoint inhibitors.

Materials and methods

Animal procedures

Mice were handled in agreement with the guidelines conforming to current Italian regulations for the protection of animals used for scientific purposes (D.lvo 27/01/1992, no. 116). Procedures were approved by the Ethical Committee for Animal Experimentation of the National Institute of Cancer Research and by the Italian Ministry of Health. The experiments were performed with the BALB/c mouse strain using animals between 3 and 6 months of either sex.

Tumors were implanted by injecting a suspension of 2×10^4 cells in adult mouse brains using a Hamilton syringe (Bregma coordinates: anterior posterior, 1.0 mm anterior; lateral, and 2.5 mm below the skull surface). Twenty-one days after tumor implantation, 5 μ l oHSV preparation, containing 2×10^6 or 10^8 PFU of either R-115 or R-LM113 or gamma-irradiated R-LM113 were injected at the same stereotaxic coordinates. Animals were monitored daily and were killed at first signs of neurological symptoms or at the predetermined time point. Their brains were photographed by fluorescence stereomicroscope (Wetzlar, Germany).

Survival curves were determined using Kaplan Meier survival between groups and compared by the log-rank test. The minimum sample size was preliminary established based on log-rank power analysis, assuming an effect size $HR \leq 0.33$ and a power of $1 - \beta \geq 0.8$.

Mice were randomly assigned to the different experimental arms by assigning them a unique ID and using Microsoft Excel pseudorandom generator to sort them. The animals that did not survive intracranial injection procedure and died within 3 days from virus injection were excluded from the experiment.

Up to 0.3 ml of blood was withdrawn 20 days after treatment with oHSV. In addition, we took blood from long-surviving mice, 4 days after the transplantation of mHGG^{pdgf}-hHER2 in the contralateral hemisphere. Heparinized capillaries were used for retro-orbital bleeding procedure. Samples were incubated for 6 h at 60 °C with

heparin. Cells were removed from serum by centrifugation (10 min at 2000 g with refrigerated centrifuge).

Cell cultures and infection

PDGF-B-induced brain tumors expressing DsRed fluorescent reporter were obtained as previously described [35]. Cells derived from primary tumors were maintained onto plates coated with Matrigel matrix (1:200; BD Biosciences, Franklin Lakes, NJ) in Dulbecco's Modified Eagle Medium (DMEM)/F12 (Gibco-Life Technologies, Carlsbad, CA, USA) added with B27 supplement (Gibco-Life Technologies, Carlsbad, CA, USA), recombinant human FGF2 (10 ng/ml; PeproTech, Rocky Hill, USA) and recombinant human EGF (10 ng/ml; PeproTech, Rocky Hill, USA). mHGG^{pdgf} cells were transduced with RAVI-hHER2 retroviral vectors and sorted by FACS based on the expression of the receptor.

Human glioma-initiating cells were produced as described elsewhere [39] and maintained onto plates coated with Matrigel matrix (1:200; BD Biosciences, Franklin Lakes, NJ) in DMEM/F12 and Neurobasal (Gibco-Life Technologies, Carlsbad, CA, USA), supplemented with B27 supplement (Gibco-Life Technologies, Carlsbad, CA, USA), L-glutamine (2 mM, Euroclone, Pero (MI), Italy), penicillin-streptomycin (Euroclone, Pero (MI), Italy), recombinant human basic FGF (10 ng/ml; Peprotech, Rocky Hill, USA), and recombinant human EGF (20 ng/ml; Peprotech, Rocky Hill, USA).

The construction and production of the recombinant oHSVs R-LM113 and R-115, retargeted to hHER2, were described elsewhere [37, 52]. Three technical replicates of cells were infected with R-115 or R-LM113, at the indicated MOI estimated on the basis of titer determined in SK-OV-3 cells. The efficiency of infection was monitored under inverted microscope (EVOS FL Cell Imaging System, ThermoFisher Scientific, Waltham, MA, USA) by means of enhanced green fluorescent protein (EGFP) expression for both viruses. The percentage of infected cells was evaluated measuring the fluorescence intensity of cells by flow cytometry (CyAn ADP, Beckman Coulter, Indianapolis, IN, USA).

mIL-12 was quantified from the supernatants of three independent technical replicates of mHGG^{pdgf}-hHER2 cells infected at 0.1, 0.3 or 1 PFU/cell with R-LM113 or R-115, using the Mouse IL-12 (p70) ELISA Plate kit (ThermoFisher Scientific, Waltham, MA, USA).

Immunostaining and flow cytometry

For flow cytometry and cell sorting, mHGG^{pdgf}-hHER2 cells were stained in suspension with mouse monoclonal antibody against hHER2 (1 µg/10⁶ cells; Santa Cruz

Biotechnology, Santa Cruz, CA, USA, #Sc-08). Binding of primary antibody was revealed with secondary anti-mouse CyTM2-conjugated, (1:50; Jackson ImmunoResearch, Milano, Italy, #115-485-003).

To detect antibodies in murine plasma, tumor cells and NIH/3T3 cells (in ratio 1:1.5) were incubated for 1 h at 4 °C with plasma diluted in saline solution (1:5). After washing in PBS, cells were incubated for 30 min with secondary anti-mouse antibodies (anti IgG: CyTM2 conjugated, Jackson ImmunoResearch, Milano, Italy; anti IgG1, anti IgG2a, anti-IgG2b: Southern Biotech, Birmingham, AL USA, #1071-02, #1081-02, and 1091-02) and analyzed by flow cytometry (CyAn ADP, Beckman Coulter, Indianapolis, IN, USA). Positive control consisted of mouse monoclonal antibody against hHER2 and secondary anti-mouse CyTM2-conjugated. The proportion of DsRed-positive mHGG^{pdgf}-hHER2 cells above a threshold on the FITC-fluorescence axis among the cells were normalized to the mean of the proportion of cells above the same threshold among the DsRed-negative in all samples.

To detect apoptotic/necrotic population, three different hGic cell cultures 6 days after R-115 infection were analyzed by using the GFP-CERTIFIED® Apoptosis/Necrosis Detection Kit (Enzo Life Sciences, Farmingdale, NY, USA). Cells treated for 4 h with staurosporine included in the kit were used as positive control.

FlowJo software v10.0.7 was used for flow cytometry data analyses.

For histological analysis, brains were fixed with 4% paraformaldehyde, cryoprotected in 20% sucrose and sectioned with a Leica CM3050 S cryostat. Immunostainings on brain sections or cultured cells were performed using mouse monoclonal antibody against CD4 (1:500, BD Pharmingen, San Diego, CA, USA, #550280) or CD8 (1:200, Novus Biological, Littleton, USA, #B173835). Binding of primary antibody was revealed with secondary anti-rat Alexa 488-conjugated (1:500; Jackson ImmunoResearch, Milano, Italy, #112-545-003). The shapes of the sections of tumor masses and positions of CD4 and CD8 were acquired by epifluorescence microscope (Axio Imager. M2, Zeiss, Oberkochen, Germany) by using AxioVision Rel. 4.8 (Zeiss, Oberkochen, Germany) and Slide Explorer2 plug-in of Micromanager [53]. A set scripts for R 3.4.0 [54] developed in house (available on request) were used to produce the overlay between pictures and acquired positions of lymphocytes and to calculate their distances from the tumor edges.

Immunohistochemical stainings on brain sections were performed using mouse IFN-γ antibody (1:500, Biolegend, San Diego, CA, USA, #505806). Binding of primary antibody was revealed with secondary anti-rat biotiny-conjugated antibody (1:500, Sigma-Aldrich Saint Louis, USA, # B7139) and then with HRP-conjugated streptavidin

(1:250, R&D Biosystem, Minneapolis, USA, #890803) and revealed with AEC (3-amino-9-ethylcarbazole). Cells nuclei were counterstained with hematoxylin. Images of tumor areas were acquired with Eclipse 80i microscope (Nikon Corporation, Tokyo, Japan). The images obtained with AEC dye were processed using Photoshop CC2018 (Adobe Systems; Tokyo, Japan) to reconstruct the image of tumor areas and then were analyzed with Fiji software [55].

RNA sequencing

For whole-exome sequencing, at least 0.01 µg RNA derived from three ex-vivo primary tumor samples, sorted by FACS for DsRed reporter and harvested in Trizol, was sent to BGI genomics (BGI, Shenzhen, China) and sequenced on BGISEQ-500 RS generating 50 base-pair single-end reads. The high-quality clean tags were mapped to reference genome (mm10) using STAR [56]. To quantify the gene expression level, RSEM analysis was carried out [57], acquiring read count of each gene of each sample, based on the mapping results. Normalization and data trimming were performed by edgeR [58]. Raw and processed data are available on GEO Dataset (accession number GSE109614).

Statistical methods

Sample sizes for each experiment are indicated in the results or in the material and methods sections. Statistical analyses were performed with two-sided *t*-test when between two conditions, one-way ANOVA followed by Tukey HSD post-hoc test when between more conditions. Survival analyses were performed with log-rank test. Error bars represent standard deviations. Numerical values are reported as mean ± standard deviation.

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Compliance with ethical standards

Conflict of interest GCF owns shares in Nouscom. GCF and LM receive equity payments from Amgen. The funding sponsors had no role in the design of the study; in the collection, analyses, or interpretation of data; and in the writing of the manuscript. The other authors declare that they have no conflict of interest.

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