Research Article

Postveraison Shoot Trimming Reduces Cluster Compactness without Compromising Fruit Quality Attributes in Organically-Grown Sangiovese Grapevines

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Abstract: Vine performance following preveraison shoot trimming is well document, the consequences of such treatments later in the season are poorly understood. Therefore, a 4-yr study was conducted in a mature vineyard of Sangiovese (clone 12T) grafted onto Kober 5BB rootstock at a spacing of 1 m x 2.8 m (intra- and interrow) to analyze the influence of postveraison shoot trimming on vine growth characteristics, cluster architecture (cluster compactness), and yield from a physiological viewpoint. The treatments consisted of shoot trimming during postveraison in a randomized block design with eight replications; each replication comprised of six vines. Three shoot trimming treatments consisting of light trimming (14 nodes), severe trimming (10 nodes), and an un-trimmed control were imposed when the soluble solids reached 15 Brix in August (40-45 day before expected harvest). Following the treatments, various vine growth characteristics, cluster morphology, and fruit quality attributes

were measured. Postveraison shoot trimming, especially severe trimming reduced cluster weight along with cluster compactness, productivity, and total yield. In terms of fruit quality attributes, these reductions were manifested as lowering of Brix and pH with minor effects on TA, yeast assimilable nitrogen, the anthocyanin profile, and total anthocyanins. These results demonstrated that postveraison shoot trimming can be a valuable production practice in reducing cluster compactness without compromising overall fruit quality attributes in Sangiovese.

Key words: anthocyanin, Brix, growth, productivity, yield

Introduction

The cultivar Sangiovese is the most important Italian winegrape used for making prestigious Tuscan wines such as Brunello di Montalcino, Nobile di Montepulciano and Chianti. Although grown with both conventional and organic viticultural practices, organically-grown Sangiovese has been increasing for the last few years as organically generated products meet the standard requirements for quality as well as healthiness (Pagliarini et al. 2013). A key reproductive feature of Sangiovese is that regardless of how it is grown, it produces tight clusters, which are grouped as moderately compact, semi-compact, and compact (Nelson-Kluk 2006). Clusters of such morphology incur losses in cuticular barrier properties at the contact surfaces and hence become victims of a host of fungal diseases, mostly *Botrytis* cluster rot during ripening especially of grapevines grown with organic protocols. Other concerns include its tendency to overcrop ensuing from its high fruitfulness of shoots regardless of origin (primary or secondary buds, basal buds) and vigorous procumbent growth habit leading to dense canopies, which generally result in negative effects on fruit quality, wood maturity, and vine size maintenance (Poni et al. 2006).

54 Combatting cluster rot has always been a multifaceted endeavor to have a profitable
55 grape production. For instance, fungicides spray is one obvious option. However, it is an
56 expensive short-term operation as pathogens develop resistance to fungicides overtime and thus
57 it is unlikely that practical control of fungal diseases can be achieved by the use of fungicides
58 alone. It is indeed true in organic viticulture necessitating an alternative and a more
59 environmentally-benign approach as the chemicals that are organically certified add to more
60 environmental problems rather than remedying the situation due to their ineffectiveness
61 (Fragoulis et al. 2009). The other option is to employ various canopy management practices to
62 improve the cluster architecture and subsequently microclimate by loosening the clusters, which
63 minimizes fungal problems. For instance, defoliation decreased physical characteristics, number
64 of berries, and cluster compactness in Trebbiano and Sangiovese (Poni et al. 2006). Other
65 practices that reduced cluster compactness include cluster division (Molitor et al. 2012), type of
66 irrigation method (drip irrigation versus partial rootzone drying), pruning strategy (spur pruning
67 by hand, mechanical hedging or minimal pruning), and combination of irrigation and pruning
68 that affected the number of clusters per vine and the rate of berry maturation (Leong et al. 2006).
69 From a rootstock perspective, cluster compactness, berry skin strength, pedicel strength, and
70 weights of cane and crop are greatly influenced by choice of cultivar (Ferreira and Marais,
71 1987). For instance, clusters of Chenin blanc on Jacquez were significantly less compact than
72 those on Ramsey or 110 Richter (Ferreira and Marais, 1987).

73 Regardless of where it is practiced, the main thrust of organic farming including
74 viticulture is to exploit ecologically balanced protocols deprived of chemical usage in order to
75 avoid any environmental problem. Accordingly, organic grape growers resort to using mostly
76 eco-friendly production practices with the exception that copper may be used only when there is

an immediate risk of infection (Fragoulis et al. 2009). Furthermore, they treat the vineyard as an interactive ecosystem known as *terroir* in viticulture parlance entailing the regional physical (e.g. climate) and cultural domains (e.g. viticultural practices), which are put to best use towards optimizing yield and fruit quality. Because of the need to adhering to the principles of organic viticulture, organic grape growers seek out a wide repertoire of canopy management practices such as shoot topping (Molitor et al. 2015), shoot trimming (Martínez de Toda et al. 2013), pruning and leaf removal (Martin, 1990), hedging (Leong et al. 2006) etc. to maintain healthy vines. Of these, an ideal practice for improving the fruit and yield of organically-grown Sangiovese would be shoot trimming as it is easily mechanized and hence relatively simple to perform. It involves pruning at different locations along the shoot and is generally used to facilitate the movement of manpower and equipment between vineyard rows, to reduce vine vigor, avoid shade from overhanging shoots, and to facilitate harvest in grapevines with upright trellis system (e.g. VSP) in cool viticultural areas (Hatch et al., 2011). Since it is more of a facilitator of cultural operations, little information is available with respect to its impact on vine performance and fruit quality, especially when it is performed late in the season after veraison. Previous studies that dealt with shoot trimming were carried out during the early part of the season i.e. preveraison and examined reproductive growth, yield, ripening, and fruit and wine quality (Keller et al. 1999), all of which widely varied in their responses. Furthermore, none of these studies examined features such as growth characteristics of vine and cluster morphology, which have a strong bearing on the incidence of fungal diseases, especially in organic viticulture. This is the first study that demonstrates positive influence of postveraison shoot trimming in the form of reduced cluster compactness and fruit yield in Sangiovese grown with the norms of organic viticulture, which has been increasing for the last few years.

Materials and Methods

Plant material and experimental layout. The experiment was conducted over four years from 2008-2011 in a mature vineyard of Sangiovese (clone 12T) grafted onto Kober 5BB rootstock at a spacing of 1 m x 2.8 m (intra- and interrow) for a density of 3571 vines/ha located in a hot hilly area of Emilia Romagna Region in Italy (lat. 44°17'7"N; long. 11°52'59"E, elevation 117 m asl). Since 2007, the vineyard established on diverse sub-soils of alluvial deposits has been managed with the principles of organic viticulture in accordance with Reg. EC 834/2007 (EC. 2007). Vines were trained to cordon de Royat training system consisting of a short trunk with a unilateral cordon trained to only one side of the trunk and hence extends from one vine to another. The vines were spur-pruned during winter to two count nodes equating to 12-14 nodes per vine. The noncount shoots (shoots arising from basal buds of the spur) were removed at the beginning and the cluster number was adjusted by cluster thinning. Throughout the experiment, the vines were maintained without any irrigation and fertilization.

The treatments consisted of shoot trimming during postveraison in a randomized block design with eight replications; each replication comprised of six vines (total of 144 vines). Prior to imposing the treatments, first vines were regularly (twice) trimmed to normalize a shoot length of 145 cm in each year. Thereafter, three shoot trimming treatments were imposed when the soluble solids reached 15 Brix in August (40-45 day before expected harvest). These include light trimming (LT of 14 nodes), severe trimming (ST of 10 nodes), and an un-trimmed control (CK 18 nodes). Shoot trimming was performed manually using large pruners. Following trimming, the shoot length in CK, LT and ST vines was 145, 102 and 64 cm, respectively. The amount of leaf area removed following trimming treatments and maintained on vines were

measured in 2009 and 2010 growing seasons using a leaf-area meter (Li-CoR Biosciences, Lincoln, NE) by removing all leaves in single plants per replicate.

Climatic data. During each season, climatic data (mean, maximum and minimum daily air temperatures (T), relative humidity (RH) and total rainfall) were recorded from bud burst to harvest from a meteorological station located close to the vineyard.

2008. In 2008, from bud burst (March 26th) to harvest (September 24th), the growing season was characterized by average T of 19.4 C with a maximum temperature of 34 C occurring at the beginning of August. During the same period, the average RH varied from 45.5% to 89.5%. The total rainfall from bud burst to harvest (243 mm) occurred generally during April-June.

2009. In 2009, the average T recorded from bud burst (April 3rd) to harvest, (September 23rd) was 22 C; maximum T of 38.7 C occurred on July 23rd. The average RH varied from 43.1% to 100%. The total rainfall from bud burst to harvest was 190 mm and predominantly took place in April and during the first week of July.

2010. In 2010, from bud burst to harvest (September 27th), the average daily T was 20 C and the average RH varied from 39.3% to 92.8%; there was an abundance of rainfall (464 mm), which occurred mostly in the spring and prior to harvest date.

2011. Overall, the 2011 growing season was marked by average T well above the seasonal normal with a maximum T of 30 C occurring in the month of August. From bud burst to harvest (September 21st), the average RH varied from 40 to 70%; highest values were observed during spring (92%) and the lowest (38%) during the latter part of August. The total rainfall from

bud burst to harvest was 204 mm, occurred generally during spring and almost none during ripening.

Measurement of growth characteristics, yield, and yield components. Leaf area maintained on vines and removed by trimming, were measured through leaf-area meter (Li-CoR Biosciences, Lincoln, NE), by removing all leaves from one vine per replicate and leaves belonging to a representative shoot in each vine. Cane length and pruning weight were determined during winter. All prunings were weighed fresh. Yield and yield components (yield, number of clusters, and cluster weight) were measured when berries met optimal universal set of criteria (e.g. Brix, TA) to determine the same harvest date for all treatments. This was determined by periodic sampling of berries starting in July. Accordingly, all treatments were harvested when berries attained same maturity levels, i.e. when soluble solid concentration reached a minimal value of 20 Brix, TA of less than 7.0 g/L etc., by taking into account the qualitative (e.g. berry ripening disorders) and sanitary (e.g. cluster rot) status of the clusters. Cluster compactness was determined according to the 1983 OIV classification. Chlorophyll was measured with a SPAD-502 chlorophyll meter (Minolta, Osaka, Japan).

Analysis of fruit composition. At harvest, berry weight, soluble solids (refractometer PAL-1, ATAGO, Tokyo, Japan), titratable acidity (TA) and pH (Crison Compact Titrator, Crison Instruments, Barcelona, Spain) were determined by randomly collecting 100 berries per replication in each treatment. On the same samples, must YAN was determined by the method described by Aerny (1996) and berry skin anthocyanins using HPLC methodology described by Venencie et al. (1997). Briefly, for each sample, skins were peeled from 30 berries, weighed and placed in 20 mL hydroalcoholic solution (EtOH:H₂O, 10:90 v/v) containing 15 mL distilled water and 2.5 g tartaric acid. The samples were homogenized with an Ultraturrax (IKA

Labortechnik, Staufen, Germany) and centrifuged for 5 min at 3000 x g at 20 C. The final solution weight was adjusted to 100 g and pH to 3.6. The samples were then macerated for 24 hr at 4 C in dark and centrifuged (ALC International, PK121R, Italy) for 20 min at 1400 g. Thereafter, the samples were filtered under vacuum with Whatman Filter Paper 40, porosity 8 µm. Skin anthocyanins were determined using a Waters 1525 instrument equipped with a diode array detector (Waters Corporation, Milford, MA) and a reversed-phase column RP18 (Supelco Inc., Bellefonte, PA); 5 µm pore size; 250 mm × 4 mm. Signals were detected at 520 nm. The elution gradient consisted of the following solvents. Solvent A: water/formic acid (10%, v/v) (Romil Ltd., Cambridge, UK); solvent B: acetonitrile (Sigma-Aldrich, Milan, Italy). The binary gradient applied was as follows: 0-7.9 min of 96% A-4% B; 7.9-23.0 min of 85% A-15% B; 23.0-27.0 min of 80% A-20% B; 27.0-43.0 min of 70% A-30 % B; 43.1-45.0 96%A-4% B. Anthocyanins were quantified by measuring peak area at 520 nm. A calibration curve was prepared from standard solutions of purified Oenin Chloride (Sigma-Aldrich, Milan, Italy) with the concentrations between 50 mg/L and 500 mg/L. The anthocyanins were expressed as mg/g of berry skin. For each treatment, eight field replicates (each one deriving from a 6-plant experimental plot) were measured. All field replicates were maintained separately and measured in the laboratory. Therefore the number of field replicates equaled to analytical replicates.

Statistical analysis. Analysis of variance and comparison of means between treatments were done by using SAS 6.04 software (SAS Institute, Cary, NC). Means were compared by the Student-Newman-Keuls test ($P \leq 0.05$). Cluster weight and cluster compactness were subjected to Kruskall Wallis non-parametric test.

Results

Growth, yield and compositional characteristics. The postveraison shoot trimming treatments were imposed on the same vines each year with the consequence that the treatment effects accumulated throughout the experiments resulting alterations in growth and fruit quality attributes. For instance, the crop load (Ravaz index) increased with an increase in shoot trimming (Table 1), which needs to be interpreted with caution. This is because, the trimming treatments, while they altered the yield to pruning weight ratio, it was rather an imposed response, not a developing response. On the other hand, berry weight declined with no differences among the treatments during the first two years of experiment (2008-2009), remained the same in 2010, but differed during the final year i.e. 2011 following the adoption of production practices of organic viticulture in 2007 (Table 1). This indicated that reduction in berry weight following shoot trimming was a cumulative process, nevertheless the possible effects of smaller vine size and relatively warmer season cannot be ignored. Cluster weight and yield per vine declined starting in 2008 (Table 1). The amount of leaf area removed and that remained on the vine following shoot trimming showed differences among the three treatments (Table 1). Once again, such differences primarily stemmed from severity of pruning treatments *per se* and were not the consequence of treatments. Analogous to cluster weight and yield per vine, cluster compactness decreased in response to shoot trimming, especially against the severe one (Table 2). Regarding fruit composition, the Brix values were reduced during the first two seasons (2008-2009), whereas no difference was observed during the latter two years (2010-2011) (Table 3). No difference was observed in pH in 2008 and 2010 whereas the 2009 and 2011 seasons displayed differences (Table 3). In contrast, the TA and YAN except for one year were immune to shoot

trimming treatments (Tables 3). In a similar pattern, the anthocyanin profile (cyanidin, peonidin, petunidin, delphinidin, malvidin) and total anthocyanins remained unaffected except for peonidin and total anthocyanins in 2010, which tended to decline in response to shoot trimming (Supplemental Table 1). Between the two shoot trimming treatments, the severe one in general mostly induced differences (lowered the values) on the measured variables.

Discussion

Shoot trimming, the mainstay of this study, is one of the cultural repertoire of production practices to which grapevines adjust by forming tylose in the xylem vessels below the cut ends as a sealing mechanism to avoid water loss and pathogen entry into the vine (Sun et al. 2006). In the aftermath of shoot trimming following the sealed cut ends, the vine attains an altered state of shoot physiology and growth characteristics. This is particularly true when shoots are trimmed after veraison, the phenological event marking the inception of ripening (Poni and Intrieri, 1996). As a result, the effects of postveraison shoot trimming would be manifested in altered cluster and berry morphologies and eventually fruit composition. These are evident in our study wherein the postveraison shoot trimming reduced cluster compactness by loosening the clusters in Sangiovese, a cultivar well-known for its vigorous growth and large sized compact clusters of varying degree (moderately compact, semi-compact, and compact) (Nelson-Kluk 2006). A similar effect was observed when shoot topping (shoot tips tipped off ~5 cm below the apex) was delayed until four weeks after the end of flowering in Riesling and Pinot gris (Molitor et al. 2015). Other canopy management practices that yielded similar effects include techniques of pruning and leaf removal (Martin 1990), but it was not clear as to what might have contributed to reducing cluster compactness and the consequent effects on fruit composition. As per this study,

it was reduced berry size (weight) and cluster weight that loosened the clusters and the ramification of this physiological activity in terms of productivity was reduced yield per vine, which was expected as berry and cluster weights are a function of productivity. This trade-off is a highly sought after feature by the practitioners of organic viticulture as loosened clusters by virtue of increased epicuticular wax load and cuticle thickness become less susceptible to cluster rots (Martin 1990). Accompanying the loosened clusters were low Brix, pH, and yield. This is a reflection of constrained ripening associated with reduced leaf area following shoot trimming (Stoll et al. 2009). Contrary to our study, these attributes reacted far more negatively (i.e. drastic reductions) when a similar form of severe shoot trimming was performed after fruit set (berry diameter 3-4 mm) by excising the internode just above the distal clusters of Grenache and Tempranillo (Martínez de Toda et al. 2013). Then again, analogous treatments of our study (trimming to the 10th node) performed early in the season (pre-veraison) not only elongated clusters but also increased yield and Brix (Cartechini et al. 2000) except for its combination with nitrogen application, wherein shoot trimming decreased fruit quality (Keller et al. 1999). While the Brix, pH, and yield were reduced, the anthocyanin profile and total anthocyanins remained unaffected by shoot trimmings. Herrera et al. (2015) observed the same phenomenon when Merlot vines were subjected to severe canopy reduction during early stages of ripening. These studies confirm that many of the precursors for anthocyanin development accumulate pre-veraison (Dokoozlian and Kliewer 1996) whereas the postveraison anthocyanin accumulation is mediated by sugars inducing genes for encoding enzymes in the anthocyanin biosynthesis pathway (Conde et al. 2007). If this is the case, then what was the source of sugars for accumulating anthocyanins in trimmed shoots? The obvious one is the reserves, which are stimulated to hydrolyze by pruning activities into sugars so as to remobilize them into the sink

organs-in-need (Clair-Maczulajtys et al. 1996). Conversely, shoot-trimming during pre-veraison (Cartechini et al. 2000) or when shoots were trimmed to two leaves per cluster reduced polyphenols including anthocyanins due to inhibition of anthocyanin biosynthetic enzymes (Wu et al. 2013). The same phenomenon is responsible for anthocyanin losses under conditions of high temperature (Mori et al. 2007), which explains the lower anthocyanin levels in 2011, a relatively much warmer season than the preceding wet seasons receiving high rainfall (190-455 mm) from budbreak to harvest. Such results are a testimony to climatic differences playing a major role in inducing seasonal variations in growth and fruit quality attributes (Herrera et al. 2015). The TA analogous to anthocyanins was not affected either. Such a response was expected of as grape berries complete acid accumulation by synthesizing most of it *in situ* not later than veraison (Conde et al. 2007). In contrast, the accumulation of nitrogen in the form of α -amino acids and ammonia contributing to the pool of YAN increases at the onset of veraison (Bell and Henschke 2008) and requires continuous supply of photoassimilates (Perez and Kliewer 1982). Hence, not surprisingly, the YAN levels declined with the removal of leaves due to shoot trimming.

Since the physiological status of vines at a given phenological event determines morphological features and shapes fruit quality attributes, the next logical proceeding is to analyze cluster architecture and fruit composition of trimmed shoots in the context of vine physiology and growth characteristics to understand the mechanics behind observed changes. In view of that, what ensued from postveraison shoot trimming was an unexpected and accelerated senescence of the shoot system. When this happens, the source:sink ratio declines and subsequently the overall photosynthetic capacity is compromised (Poni et al. 2003). This is simply due to the fact that the most photosynthetically active leaves during ripening are located

at the top of the canopy and those arising from lateral shoots (Candolfi-Vasconcelos et al. 1994). These sources provide abundant supply of assimilates for increasing both yield and Brix (Poni and Intrieri 1996), which did not occur in our study due to severe shoot trimming after veraison. On the other hand, if shoots are trimmed lightly (e.g. shoot topping) early in the season (before veraison), Brix and yield increase due to elimination of competing organs of shoot tip (Cartechini et al. 2000). Based on these observations, it is logical to infer that source limitation reduces Brix and yield. But then again, one may argue that shoot trimming should enhance ripening as it induces lateral growth with more photosynthetically efficient leaves for transporting assimilates into the berry (Koblet et al. 1996). The compensatory lateral growth for supporting berries did not surface in our study as shoots trimmed after veraison induce little regrowth (Poni and Intrieri 1996).

Other consequences of postveraison shoot trimming relates to an imbalanced (decreased) source:sink ratio resulting in a shift in sink strength, the ability to attract photosynthates towards stems and roots limiting partitioning into berries (Kriedemann and Lenz 1972). In this study, roots whose sink strength is generally weakened in the presence of fruits, the organs with the highest sink potential (Minchin and Lacombe 2005), became the next strongest sink as these are the site of cytokinin synthesis and any tissue or organ enriched with cytokinins is known to attract assimilates (Ronzhina 2004). Further support comes from Ravaz index (yield:pruning weight ratio), a measure of crop load and source-sink relationship (Kliewer and Dokoozlian 2005) increased with trimming treatments. However, such evidence needs to be interpreted with caution as the increase in Ravaz index primarily ensued from losses of leaves and stems caused by shoot trimming *per se*. Given the severity of the trimming, the loss in shoot dry matter in our study is estimated to be more than 25%. As a result, most of the assimilates freed by fruits were

mobilized into roots, a cogent course of action not only to avoid death of fine roots but also to replenish the sugars lost to bud break and early summer growth.

The reduced sink strength of berries in the shoot-trimmed clusters is also a reflection of resistance to flow in the phloem pathway (Minchin and Lacointe 2005) due to plugging of sieve plates of the phloem sieve tubes with callose, a polymer of carbohydrate, which is synthesized naturally when vines go into dormancy (Bondada, 2014). Its synthesis can also be induced by abiotic stress (Bondada 2014), hence, it is highly possible that the stress caused by trimming (Candolfi-Vasconcelos et al. 1994) might have accelerated an early synthesis of callose thereby reducing sugar export into the berries and the fact that postveraison berry expansion relies on phloem influx (Bondada et al. 2005), a resistant phloem will reduce berry size by reducing its sink strength. Although these premises lend credible physiological explanations for the lower Brix of shoot-trimmed clusters, they need to be evaluated in future studies.

Conclusions

Grapevines are subjected to many different cultural practices that may or may not favor fruit quality. The results of this study on shoot trimming are in favor of promoting positive effects on cluster morphology and fruit quality i.e. a reduction in cluster compactness and sugar levels, which ensued from source-sink imbalance associated with postveraison shoot trimming. Despite reductions in yield and sugar levels, the overall fruit quality was not lost to shoot trimming. This is a reflection of adaptive nature of grapevine conferring them with adjustments to maintain homeostasis. Grapes with reduced sugar levels is highly attractive to wineries as they can be fermented into balanced wines of reduced alcohol content with relative ease; such qualities are highly desired in today's market and by consumers. From an environmental

perspective, the cultural practice of shoot trimming is ideal to organic viticulture, which aims at promoting and enhancing biodiversity and soil biological activity. Hence, this practice is worthy of deserving recommendation to organic grape growers as an effective cultural practice for reducing sugar levels in tandem with cluster compactness in Sangiovese.

Literature Cited

- Aerny, J. 1996. Composés azotés des moûts et vins. Rev. Suisse Vitic. Arboric. Hortic. 28:161-165.
- Bell, S.J., and P.A. Henschke. 2008. Implications of nitrogen nutrition for grapes, fermentation and wine. Aust. J. Grape Wine Res. 11:242-295.
- Bondada, B.R., M.A. Matthews, and K. A. Shackel. 2005. Functional xylem in grapevine berries. J. Exp. Bot. 56:2949-2956.
- Bondada, B. 2014. Structural and compositional characterization of suppression of uniform ripening in grapevine: A paradoxical ripening disorder of grape berries with no known causative clues. J. Am. Soc. Hort. Sci. 139:567-581.
- Candolfi-Vasconcelos, M.C., M.P. Candolfi, and W. Koblet. 1994. Retranslocation of carbon reserves from the woody storage tissues into the fruit as a response to defoliation stress during the ripening period in *Vitis vinifera* L. Planta 192:567-573.
- Cartechini, A., A. Palliotti, C. Lungarotti. 2000. Influence of timing of hedging on yield and grape quality in some red and white grapevine cultivars. Acta Hort. 512:101-110.
- Clair-Maczulajtys, D., I. Le Disquet, and G. Bory. 1996. Pruning stress: changes in the tree physiology and their effects on the tree health. Acta Hort. 496:317-324.
- Conde, C., P. Silva, N. Fontes, A.C.P. Dias, R.M. Tavares, M.J. Sousa, A. Agasse, S. Delrot, and H. Gerós. 2007. Biochemical changes throughout grape berry development and fruit and wine quality. Food 1:1-22.
- Dokoozlian, N.K., and W.M. Kliewer. 1996. Influence of light on grape berry growth and composition varies during fruit development. J. Am. Soc. Hort. Sci. 121:869-874.

-
- 356 Ferreira, J.H.S., and P.G. Marais. 1987. Effect of rootstock cultivar, pruning method and crop
357 load on *Botrytis cinerea* rot of *Vitis vinifera* cv. Chenin blanc grapes. S. Afr. J. Enol. Vitic. 8:41-
358 44.
- 359
- 360 Fragoulis, G., M. Trevisan, A.D. Guardo, A. Sorce, M.V. Meer, F. Weibel, and E. Capr. 2009.
361 Development of a management tool to indicate the environmental impact of organic viticulture.
362 J. Environ. Qual. 38:826-835.
- 363
- 364 Hatch, T.A., C.C. Hickey, and T.K. Wolf. 2011. Cover crop, rootstock, and root restriction
365 regulate vegetative growth of Cabernet Sauvignon in a humid environment. Am. J. Enol. Vitic.
366 62:298-311.
- 367
- 368 Herrera, J.C., B. Bucchetti, P. Sabbatini, P. Comuzzo, L. Zulin, and A. Vecchione. 2015. Effect
369 of water deficit and severe shoot trimming on the composition of *Vitis vinifera* L. Merlot grapes
370 and wines. Aust. J. Grape Wine Res. 21:254-265.
- 371
- 372 Keller, M., R. M. Pool, and T. Henick-Kling. 1999. Excessive nitrogen supply and shoot
373 trimming can impair colour development in Pinot noir grapes and wine. Aust. J. Grape Wine
374 Res. 5:45-55.
- 375
- 376 Kliewer, M., and N.K. Dokoozlian. 2005. Leaf area/crop weight ratios of grapevines: Influence
377 on fruit composition and wine quality. Am. J. Enol. Vitic. 56:170-181.
- 378
- 379 Koblet, W., M.C. Candolfi-Vasconcelos, and M. Keller. 1996. Stress und Stressbewältigung bei
380 Weinreben. Bot. Helv. 106:73-84.
- 381
- 382 Kriedemann, P.E., and F. Lenz. 1972. The response of vine leaf photosynthesis to shoot tip
383 excision and stem cincturing. Vitis 11:193-197.
- 384
- 385 Leong, S.L., A.D. Hocking, J.I. Pitt, B.A. Kazi, R.W. Emmett, and E.S. Scott. 2006. Australian
386 research on ochratoxigenic fungi and ochratoxin A. Int. J. Food Microbiol. 111:S10-S17.
- 387
- 388 Martin, S.R. 1990. Systematic management to minimize botrytis cluster rot in three victorian
389 vineyards. Aust. New Zealand Wine Indust. J. 5:235-237.
- 390
- 391 Martínez de Toda, F., J.C. Sancha, and P. Balda. 2013. Reducing the sugar and pH of the grape
392 (*Vitis vinifera* L. cvs. 'Grenache' and 'Tempranillo') through a single shoot trimming. S. Afr. J.
393 Enol. Vitic. 34:246-251.
- 394

-
- Minchin, P.E.H., and A. Lacombe. 2005. New understanding on phloem physiology and possible consequences for modelling long-distance carbon transport. *New Phytol.* 166:771-779.
- Molitor, D., M. Behr, L. Hoffmann, and D. Evers. 2012. Impact of grape cluster division on cluster morphology and cluster rot epidemic. *Am. J. Enol. Vitic.* 63:508-514.
- Molitor, D., N. Baron, T. Sauerwein, C.M. Andre, A. Kicherer, J. Doring, M. Stoll, M. Beyer, L. Hoffmann, and D. Evers. 2015. Postponing first shoot topping reduces grape cluster compactness and delays cluster rot epidemics. *Am. J. Enol. Vitic.* 66:164-176.
- Mori, K., N. Goto-Yamamoto, M. Kitayama, M. and K. Hashizume. 2007. Loss of anthocyanins in red wine grape varieties under high temperature. *J. Exp. Bot.* 58:1935-1945.
- Nelson-Kluk, S. 2006. Sangiovese at FPS. FPS Grape Program Newsletter, November, 2006.
- OIV, 1983. Le code des caractères descriptifs des variétés et espèces de *Vitis*. Office International de la Vigne et du Vin, Ed. Dedon, Paris.
- Pagliarini, E., M. Laureati, and D. Gaeta. 2013. Sensory descriptors, hedonic perception and consumer's attitudes to Sangiovese red wine deriving from organically and conventionally grown grapes. *Front. Psychol.* 4:1-7.
- Perez, J.R., and W.M. Kliewer. 1982. Influence of light regime and nitrate fertilization on nitrate reductase activity and concentrations of nitrate and arginine in tissues of three cultivars of grapevines. *Am. J. Enol. Vitic.* 33:86-93.
- Poni, S. and C. Intrieri. 1996. Physiology of grape leaf ageing as related to improved canopy management and grape quality. In *Proceedings of the 9th Australian Wine Industry Technical Conference*. C.S. Stockley, A.N. Sas, R.S. Johnstone and T.H. Lee (eds). pp. 113–122. Adelaide, Australia (Winetitles).
- Poni, S., M. Quartieri, and M. Tagliavini, 2003. Potassium nutrition of Cabernet Sauvignon grapevines (*Vitis vinifera* L.) as affected by shoot trimming. *Plant Soil* 253: 341-351.
- Poni S., L. Casalini, F. Bernizzoni, S. Civardi, and C. Intrieri. 2006. Effects of early defoliation on shoot photosynthesis, yield components, and grape composition. *Am. J. Enol. Vitic.* 57:397-407.

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- Ronzhina, E.S. 2004. Structural and functional rearrangements of mesophyll as a probable basis for the cytokinin-dependent assimilate translocation in detached leaves. *Russian J. Plant Physiol.* 51:333-341.
- Stoll, M., M. Scheidweiler, M. Lafontaine, and H.R. Schultz. 2009. Possibilities to reduce the velocity of berry maturation through various leaf area to fruit ratio modifications in *Vitis vinifera* L. Riesling. *In* Proceedings of 16th international GiESCO Symposium. N. Dokoozlian and J. Wolpert (eds.). pp. 93-96. University of California, Davis, CA, USA.
- Sun, Q., T.L. Rost, and M.A. Matthews. 2006. Pruning-induced tylose development in stems of current-year shoots of *Vitis vinifera* (Vitaceae). *Am. J. Bot.* 93:1567-1576.
- Venencie, C., M.N. Uveira, and S. Guet. 1997. Maturité polyphénolique du raisin mise en place d'une méthode d'analyse de routine. *Rev. Fr. Oenol.* 167:36-41.
- Wu, B.H., N. Niu, J.H. Li, and S.H. Li. 2013. Leaf:fruit ratio affects the proteomic profile of grape berry skins. *J. Am. Soc. Hort. Sci.* 138:416-427.

Table 1 Growth characteristics, crop load, and yield and yield components in control, lightly and severely trimmed Sangiovese vines from 2008-2011.

Year	Treatment	Pruning			Yield (kg/plant)	Ravaz index	Cluster	Berry
		wt (kg/vine)	LA1 (m ²)	LA2 (m ²)			wt (g)	wt (g)
2008	CK	0.47 a	4.5 a	-	5.6 a	11.9 b	332.0 a	2.2
	ST	0.39 b	2.9 b	1.3	5.2 ab	13.3 b	318.4 ab	2.1
	LT	0.25 c	2.4 c	2.5	4.7 b	18.8 a	282.6 b	2.0
	Significance	***	**	***	*	***	*	ns
2009	CK	0.55 a	3.6 a	-	3.4 a	6.0 b	268.1 a	2.6
	ST	0.42 b	2.8 ab	1.1	3.2 ab	7.6 ab	245.9 ab	2.5
	LT	0.29 c	2.1 b	1.6	2.6 b	9. b	210.2 b	2.4
	Significance	***	***	ns	*	**	*	ns
2010	CK	0.66 a	2.9 a	-	5.4 a	8.2 b	294.5 a	2.5
	ST	0.55 a	2.4 b	0.9	4.7 ab	8.5 b	260.5 b	2.5
	LT	0.40 b	1.3 c	0.6	4.0 b	10.0 a	246.8 b	2.5
	Significance	**	***	***	*	*	*	ns
2011	CK	0.42 a	4.9 a	-	2.5 a	6.0 b	216.1 a	2.1 a
	ST	0.32 b	2.7 b	0.8	2.2 a	6.9 ab	191.6 b	1.9 b
	LT	0.21 c	1.9 c	1.2	1.8 b	8.6 a	158.9 b	1.8 b
	Significance	***	***	*	**	*	**	***

*Significant at $p < 0.05$, ** significant at $p < 0.01$, *** significant at $p < 0.001$, ns, not significant. Means within a column followed by different letters are significantly different according to the Student-Newman-Keuls test (pruning weight, Ravaz index, LA, berry weight, yield) and Kruskal-Wallis test (cluster weight). CK, control vines; LT, lightly trimmed vines; ST severely trimmed vines; LA1, leaf area before imposing the treatments; LA2, leaf area after imposing the treatments.

Table 2 Cluster compactness (OIV rating) in control, lightly and severely trimmed Sangiovese vines from 2010 and 2011 experiments.

Treatment	Cluster compactness	
	2010	2011
CK	6.5 a	7.3 a
LT	6.6 a	5.5 b
ST	5.6 b	5.6 b
Significance	*	*

*Significant at $p < 0.05$. Means within a column followed by different letters are significantly different according to the Student- Newman-Keuls test. CK, control vines; LT, lightly trimmed vines; ST, severely trimmed vines.

Table 3 Berry compositional attributes in control, lightly and severely trimmed Sangiovese vines from 2008-2011.

Year	Treatment	TSS (Brix)	pH	TA (g/L tartaric acid)	YAN (mg/L)
2008	CK	22.7 a	3.30	6.8	
	ST	21.5 ab	3.30	7.0	
	LT	20.7 b	3.20	7.1	
	Significance	*	ns	ns	
2009	CK	24.2 a	3.40 a	4.6	165 a
	ST	23.4 b	3.30 ab	4.4	142 b
	LT	23.5 b	3.4 b	4.9	140 b
	Significance	*	***	ns	*
2010	CK	20.9	3.11	7.0	106
	ST	20.5	3.11	7.1	105
	LT	20.6	3.13	6.7	116
	Significance	ns	ns	ns	ns
2011	CK	25.6	3.49 a	7.0	99
	ST	25.4	3.45 b	6.9	91
	LT	25.3	3.41c	6.7	81
	Significance	ns	***	ns	ns

*Significant at $p < 0.05$, ** significant at $p < 0.01$, significant at $p < 0.001$. Means within a column followed by different letters are significantly different according to the Student- Newman-Keuls test. CK, control vines; LT, lightly trimmed vines; ST, severely trimmed vines; TA, titratable acidity; TSS, total soluble solids; YAN, yeast assimilable nitrogen.

Supplemental Table 1 Concentration (mg/g of skin) of individual anthocyanins at harvest in berries of control, lightly, and severely trimmed Sangiovese vines from 2008-2011. Cyanidin-3-glucoside (cn-3G); peonidin-3-glucoside (pn-3G); delphinidin-3-glucoside (dp-3G); petunidin-3-glucoside (pt-3G); malvidin-3-glucoside (mv-3G); and total glycosylated anthocyanin (tot-G).

Year	Treatment	cn-3G	pn-3G	dp-3G	pt-3G	mv-3G	tot-G
2008	CK	0.28	0.21	0.21	0.24	0.71	1.64
	LT	0.26	0.19	0.21	0.24	0.71	1.61
	ST	0.28	0.19	0.20	0.23	0.65	1.55
	<i>Significance</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>
2009	CK	0.34	0.21	0.14	0.16	0.38	1.22
	LT	0.37	0.21	0.16	0.17	0.40	1.30
	ST	0.36	0.20	0.15	0.17	0.41	1.29
	<i>Significance</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>
2010	CK	0.51	0.26 a	0.28	0.29	0.71	2.04 a
	LT	0.43	0.22 b	0.23	0.25	0.59	1.71 b
	ST	0.51	0.25 ab	0.28	0.30	0.63	1.95 ab
	<i>Significance</i>	<i>n.s.</i>	*	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	*
2011	CK	0.17	0.11	0.07	0.08	0.17	0.59
	LT	0.16	0.10	0.07	0.08	0.16	0.56
	ST	0.16	0.10	0.07	0.08	0.16	0.57
	<i>Significance</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>	<i>n.s.</i>

*Significant at $p < 0.05$; ns, not significant. Means within a column followed by different letters are significantly different according to the Student- Newman-Keuls test. CK, control vines; LT, lightly trimmed vines; ST, severely trimmed vines.