

## SUPPORTING INFORMATION

### Assessing the Role of Counterion in Gold Catalyzed Dearomatization of Indoles with Allenamides by NMR Studies

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## 1. General procedures

Complex **1Cl** was synthesized following known method<sup>1</sup> and complex **1TFA** was obtained via halogen metathesis reactions (**1Cl** + AgTFA) as previously described.<sup>2</sup>

Samples of **1OTf** were generated *in situ* within a screw cap NMR tube, by reacting the suitable amount of **1Cl** with an excess of AgOTf in methylene chloride-*d*<sub>2</sub>. 15 minutes after mixing, the solution was filtered into another screw cap NMR tube in order to remove the excess of AgOTf and formed AgCl. Quantitative <sup>1</sup>H and <sup>19</sup>F NMR spectra were performed to assess the purity of the complex (>90%).

Unless specified, NMR samples were prepared in open air by using methylene chloride-*d*<sub>2</sub> dried over CaH<sub>2</sub>. Experiments in exclusion of water and oxygen were performed by loading the suitable amount of solids within a J-Young NMR tube, which was interfaced to an high vacuum Schlenk line (<10<sup>-5</sup> Torr) and dried overnight. Then, dry methylene chloride-*d*<sub>2</sub> was condensed into the NMR tube at 77K and kept at low temperature before inserting the sample into the NMR probe.

1D- and 2D-NMR spectra were measured on a Bruker Avance III HD 400 spectrometer equipped with a <sup>1</sup>H,BB smartprobe. Referencing is relative to solvents (<sup>1</sup>H and <sup>13</sup>C), CCl<sub>3</sub>F (<sup>19</sup>F) and 85% H<sub>3</sub>PO<sub>4</sub> (<sup>31</sup>P). <sup>1</sup>H NOESY NMR experiments were acquired by using the PFG pulse sequence version,<sup>3</sup> <sup>19</sup>F, <sup>1</sup>H HOESY NMR experiments were acquired by using the standard four-pulse sequence or its modified version.<sup>4</sup> The number of transients and the number of data points were chosen according to the sample concentration and to the desired final digital resolution. Semi-quantitative spectra were acquired with 1s relaxation delay and 800ms mixing time.

<sup>1</sup>H PGSE (Pulsed Field gradient Spin Echo) NMR measurements were performed by using a double stimulated sequence, containing 3 spoil gradients and longitudinal encode-decode (dstegp3s1d), on a Bruker Avance III HD NMR spectrometer equipped with a <sup>1</sup>H,BB smartprobe, at 298K without spinning. The dependence of the resonance intensity on a constant waiting time and on a varied gradient strength *G* is described as:

$$\ln \frac{I}{I_0} = (\gamma\delta)^2 D_t \left( \Delta - \frac{\delta}{3} \right) G^2$$

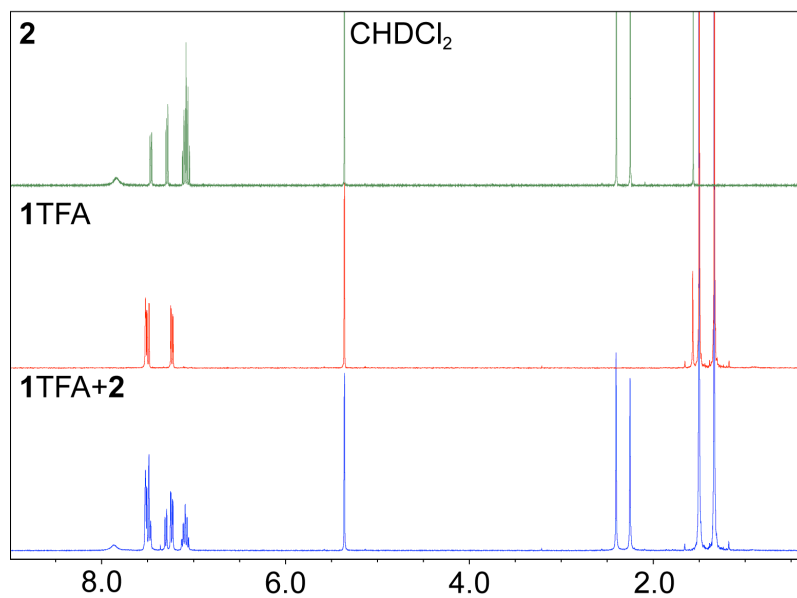
where  $I$  is the intensity of the observed spin echo,  $I_0$  is the intensity of the spin echo in the absence of gradient,  $D_t$  is the self-diffusion coefficient,  $\Delta$  is the delay between the midpoints of the gradients,  $\delta$  is the length of the gradient pulse and  $\gamma$  is the magnetogyric ratio. The shape of the gradients was rectangular, their length  $\delta$  was 4-5 ms and their strength was varied during the experiments. Spectra were acquired for 64k points with a spectral width of 6000 Hz and processed with a line broadening of 1.0. The semi-logarithmic plots of  $\ln(I/I_0)$  versus  $G^2$  were fitted by using a standard linear regression algorithm. Different values of  $\Delta$ ,  $G$ , and number of transients were used for different samples. Hydrodynamic data were derived by using solvent as internal standard as reported elsewhere.<sup>5</sup>

**1TFA** –  $^1\text{H}$  NMR (400.13 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  7.52 (m, H2 P(OR)<sub>3</sub>), 7.49 (dd,  $J_{\text{H,H}}=8.6$  and 1.5 Hz, H5 P(OR)<sub>3</sub>), 7.23 (dd,  $J_{\text{H,H}}=8.6$  and 2.5 Hz, H4 P(OR)<sub>3</sub>), 1.49 (s,  $t\text{-Bu}^1$  P(OR)<sub>3</sub>), 1.33 ppm (s,  $t\text{-Bu}^2$  P(OR)<sub>3</sub>).  $^{19}\text{F}$  NMR (376.5 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  -74.0 ppm (s, TFA).  $^{31}\text{P}\{^1\text{H}\}$  NMR (150.0 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  88.6 ppm (s, P(OR)<sub>3</sub>).

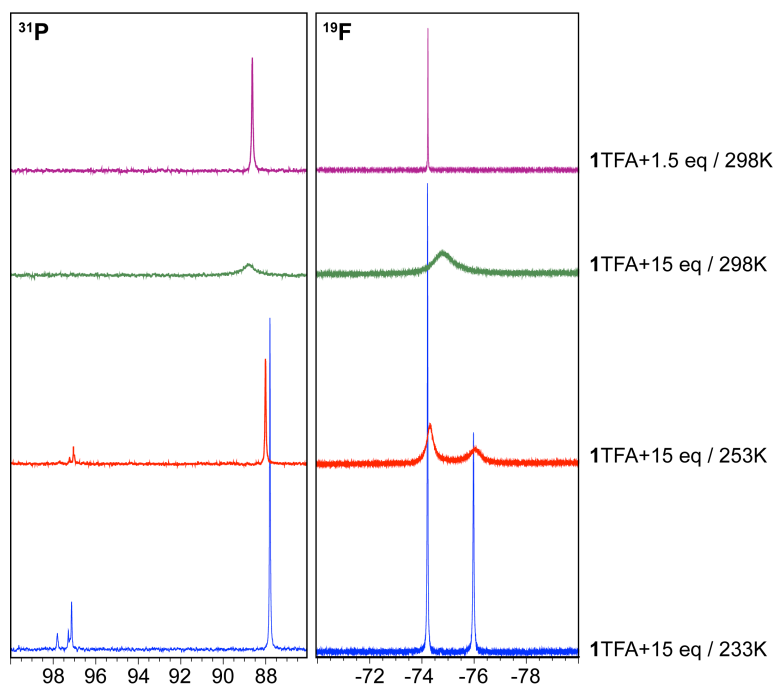
**1OTf** –  $^1\text{H}$  NMR (400.13 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  7.52 (m, H2 P(OR)<sub>3</sub>), 7.44 (d,  $J_{\text{H,H}}=8.6$ , H5 P(OR)<sub>3</sub>), 7.23 (dd,  $J_{\text{H,H}}=8.6$  and 2.4 Hz, H4 P(OR)<sub>3</sub>), 1.49 (s,  $t\text{-Bu}^1$  P(OR)<sub>3</sub>), 1.33 ppm (s,  $t\text{-Bu}^2$  P(OR)<sub>3</sub>).  $^{19}\text{F}$  NMR (376.5 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  -77.7 ppm (s, TFA).  $^{31}\text{P}\{^1\text{H}\}$  NMR (150.0 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  85.4 ppm (s, P(OR)<sub>3</sub>).

## 2. Relevant NMR spectra

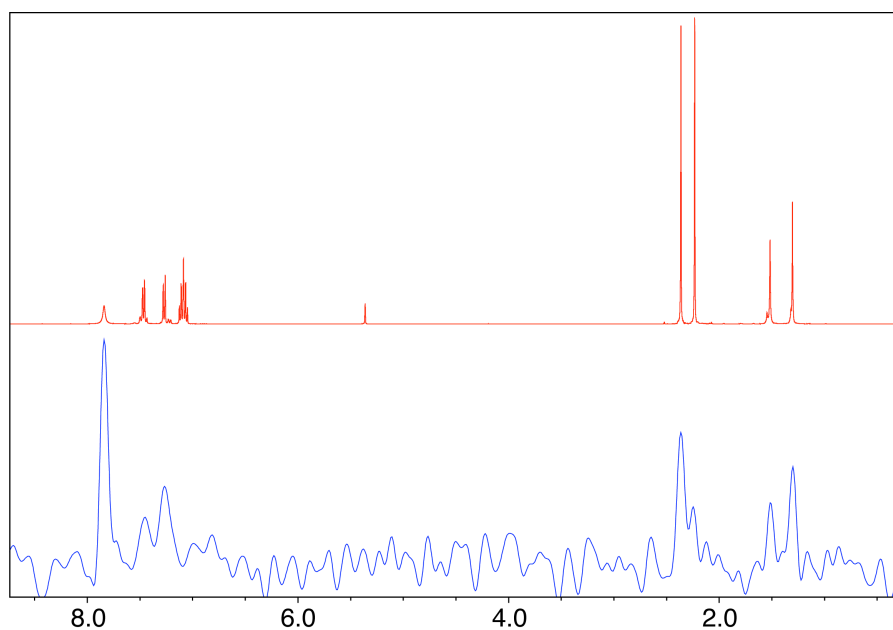
### 2.1 1TFA + 2



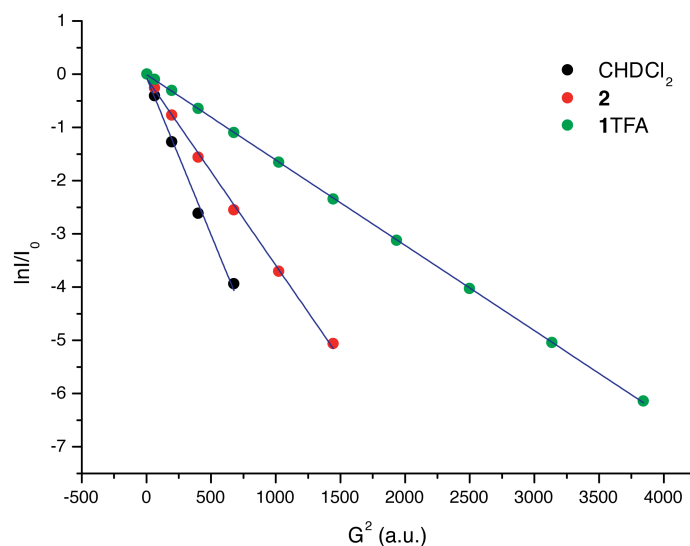
**Figure S1.** Comparison between  $^1\text{H}$  NMR spectra of **1TFA**, **2** and their mixture in anhydrous methylene chloride- $d_2$  at 298K.



**Figure S2.**  $^{31}\text{P}\{^1\text{H}\}$  and  $^{19}\text{F}$  NMR spectra obtained after the addition of different equivalents of **2** to **1TFA** at different temperatures in anhydrous methylene chloride- $d_2$ .

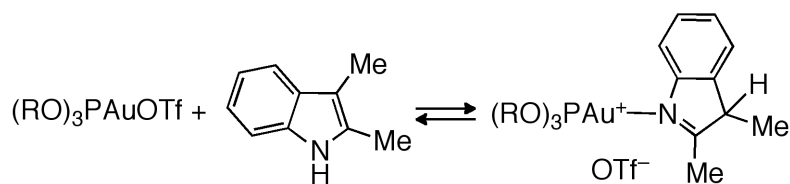


**Figure S3.** Comparison between  $^1\text{H}$  and trace at  $\delta_{\text{F}} = -72.2$  ppm of the  $^{19}\text{F}$ ,  $^1\text{H}$  HOESY NMR spectra of a mixture containing **1TFA** and 15 equivalents of **2** at 233 K in anhydrous methylene chloride- $d_2$ .



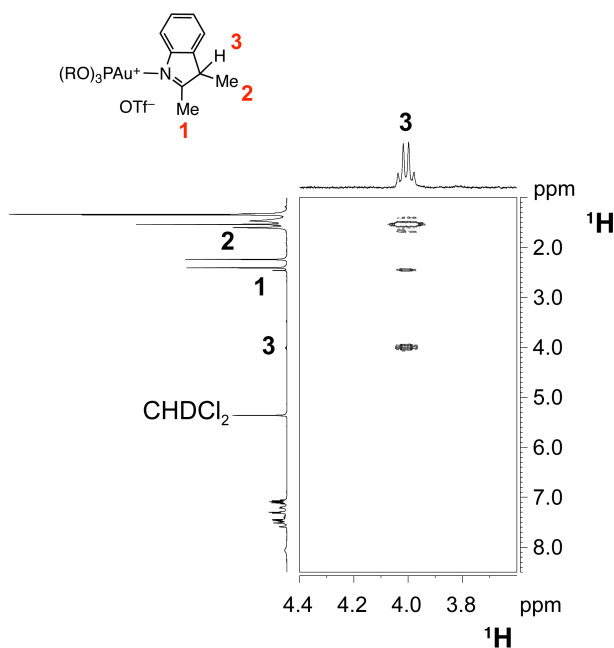
**Figure S4.**  $\ln(I/I_0)$  vs.  $G^2$  plot for a mixture containing **1TFA** and 15 equivalents of **2** at 298 K in anhydrous methylene chloride- $d_2$ . Data analysis gave an hydrodynamic volume value of  $170 \text{ \AA}^3$  for **2** and  $998 \text{ \AA}^3$  for **1TFA**, suggesting that the two species have a low tendency to interact at room temperature.

## 2.2 1OTf + 2

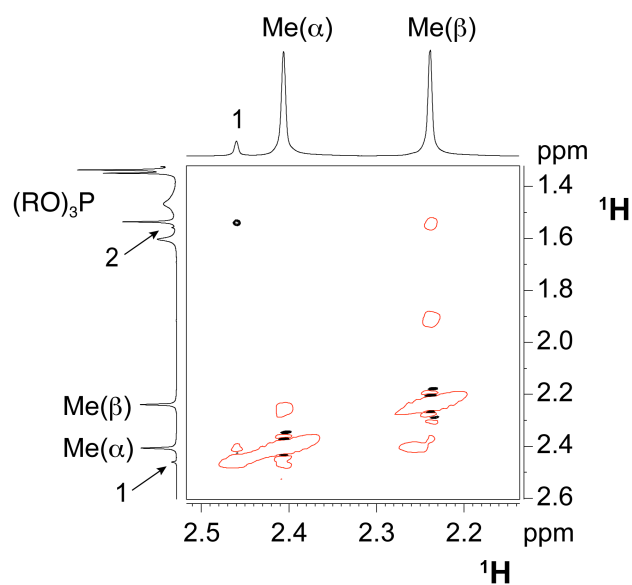


**Scheme S1.** Reaction between **1OTf** and **2**.

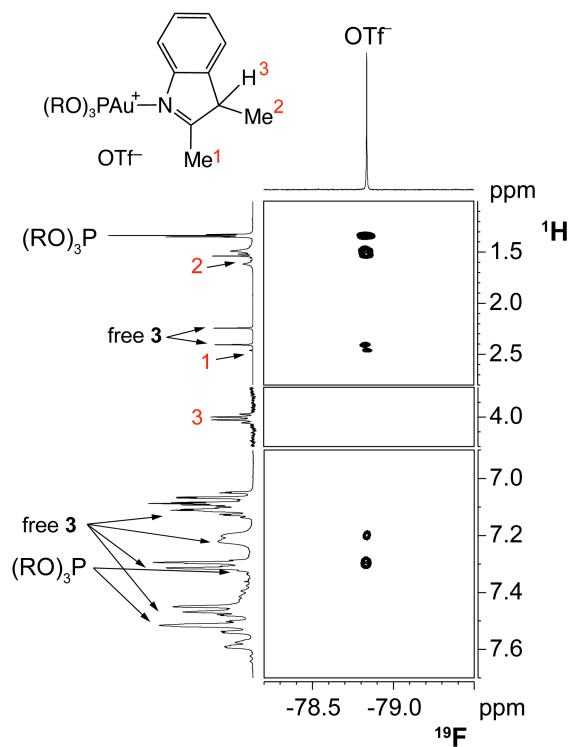
Selected  $^1\text{H}$  NMR resonances (400.13 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  7.58 (m,  $\text{P}(\text{OR})_3$ ), 7.31 (buried under **2**,  $\text{P}(\text{OR})_3$ ), 7.27 (buried under **2**,  $\text{P}(\text{OR})_3$ ), 4.00 (q,  $J_{\text{H,H}}=7.9$  Hz, H3), 2.46 (s, H1), 1.54 (d,  $J_{\text{H,H}}=7.9$  Hz, H2), 1.53 (s,  $\text{P}(\text{OR})_3$ ), 1.35 ppm (s,  $\text{P}(\text{OR})_3$ ). Selected  $^{13}\text{C}\{^1\text{H}\}$  NMR resonances (100.55 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  51.0 (s, C3), 30.9 (s,  $\text{P}(\text{OR})_3$ ), 30.2 (s,  $\text{P}(\text{OR})_3$ ), 19.2 (s, C1), 14.2 ppm (s, C2).  $^{19}\text{F}$  NMR (376.5 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  -78.7 ppm (s,  $\text{OTf}^-$ ).  $^{31}\text{P}\{^1\text{H}\}$  NMR (150.0 MHz, methylene chloride- $d_2$ , 298K)  $\delta$  97.3 ppm (s,  $\text{P}(\text{OR})_3$ ).



**Figure S5.** A section of the  $^1\text{H}$  COSY NMR spectrum obtained after mixing **1OTf** and 2 equivalents of **2** in methylene chloride- $d_2$  showing scalar coupling of H3 with H2 and H1.



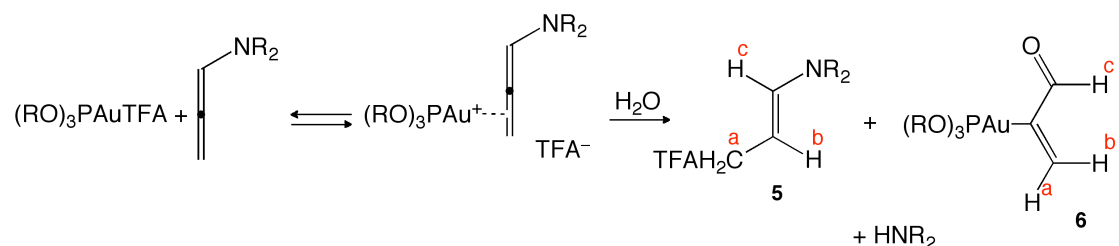
**Figure S6.** A section of the  $^1\text{H}$  NOESY NMR spectrum obtained after mixing **1**OTf and 2 equivalents of **2** in methylene chloride- $d_2$  showing chemical exchange of H1 and H2 with Me( $\alpha$ ) and Me( $\beta$ ) of **2**.



**Figure S7.**  $^{19}\text{F}$ ,  $^1\text{H}$  HOESY NMR spectrum obtained after mixing **1**OTf and 2 equivalents of **2** in methylene chloride- $d_2$  at 298K.

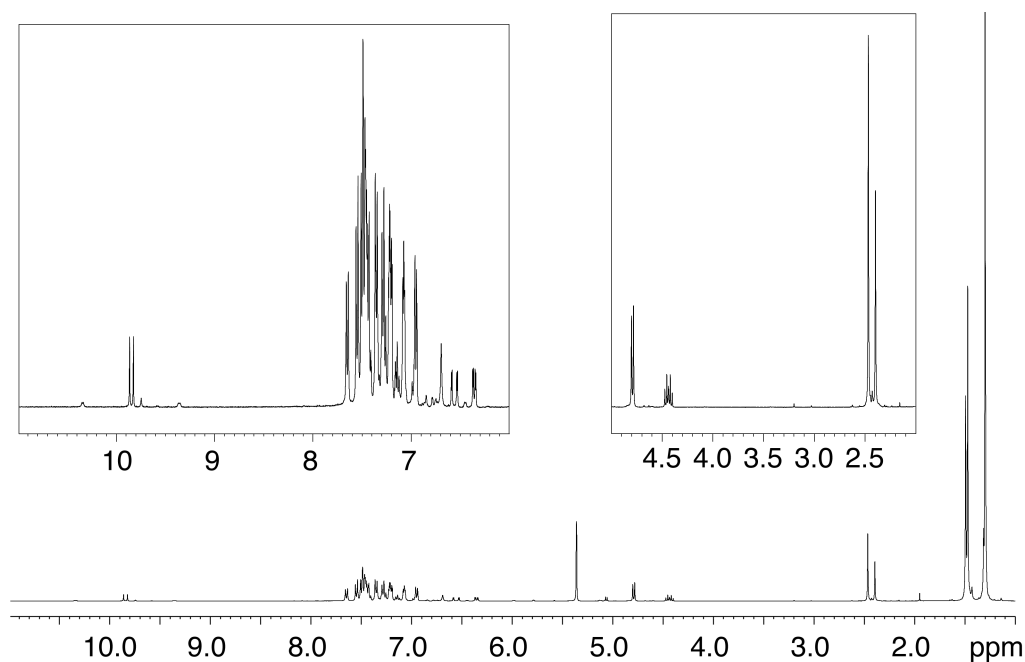
## 2.3 1TFA + 3

a) Reactivity in the presence of traces of water:



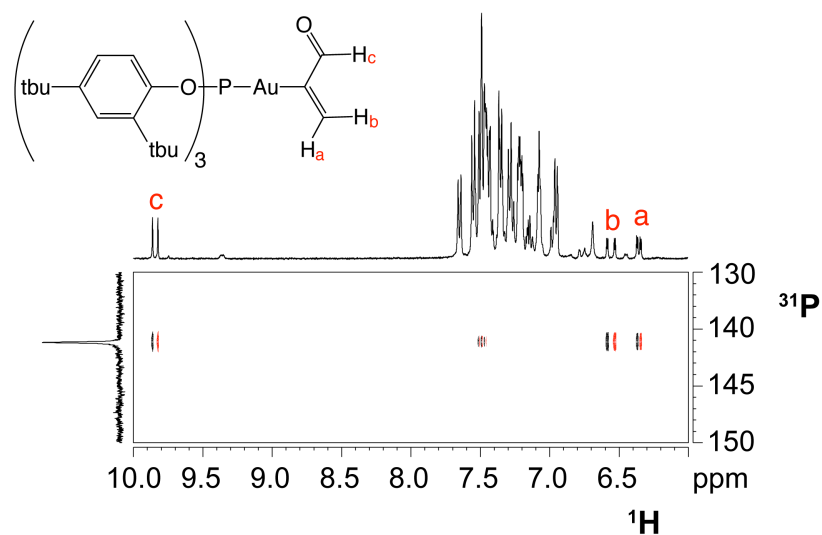
**Scheme S2.** Proposed deactivation pathway of gold-allene species in the presence of  $\text{H}_2\text{O}$  leading to anion transfer to substrate, allene hydrolysis and  $[\text{Au}(\text{I})]$ -acrolein.

Selected  $^1\text{H}$  NMR resonances (400.13 MHz, methylene chloride- $d_2$ , 253K)  $\delta$  9.84 (d,  $J_{\text{H,P}}=15.6$  Hz, Hc of **6**), 7.64 (d,  $J_{\text{H,H}}=8.4$  Hz, Ts of  $\text{HNR}_2$ ), 7.55 (d,  $J_{\text{H,H}}=8.4$  Hz, Ts of **5**), 7.48 (m, Hc of **5** +  $(\text{RO})_3\text{P}$ ), 7.35 (d,  $J_{\text{H,H}}=8.4$  Hz, Ts of **5**), 7.55 (d,  $J_{\text{H,H}}=8.4$  Hz, Ts of **5**), 7.28 (d,  $J_{\text{H,H}}=8.4$  Hz, Ts of  $\text{HNR}_2$ ), 7.21 (m,  $(\text{RO})_3\text{P}$ ), 7.14 (m, Ph of  $\text{HNR}_2$ ), 7.07 (m,  $(\text{RO})_3\text{P}$ ), 6.95 (m, Ph of **5**), 6.70 (s, NH of  $\text{HNR}_2$ ), 6.55 (dd,  $J_{\text{P,H}}=21.9$  Hz,  $J_{\text{H,H}}=3.2$  Hz, Hb of **6**), 6.35 (dd,  $J_{\text{P,H}}=10.0$  Hz,  $J_{\text{H,H}}=3.2$  Hz, Ha of **6**), 4.79 (d,  $J_{\text{H,H}}=8.0$  Hz, Ha of **5**), 4.43 (dt,  $J_{\text{H,H}}=13.8$  Hz,  $J_{\text{H,H}}=8.0$  Hz, Hb of **5**), 2.47 (s, Ts of **5**), 2.39 (s, Ts of  $\text{HNR}_2$ ), 1.47 (s,  $(\text{RO})_3\text{P}$ ), 1.29 ppm (s,  $(\text{RO})_3\text{P}$ ). Selected  $^{13}\text{C}\{^1\text{H}\}$  NMR resonances (100.55 MHz, methylene chloride- $d_2$ , 253K)  $\delta$  203.1 (d,  $J_{\text{P,C}}=4.2$  Hz, Cc of **6**), 149.7 (s, Ca/b of **6**), 145.8 (s, Cc of **5**), 100.8 (s, Cb of **5**), 67.5 (s, Ca of **5**), 31.1 (s,  $(\text{RO})_3\text{P}$ ), 29.9 (s,  $(\text{RO})_3\text{P}$ ), 21.6 ppm (s, Ts of **5** and Ts of  $\text{HNR}_2$ ).  $^{19}\text{F}$  NMR (376.5 MHz, methylene chloride- $d_2$ , 253K)  $\delta$  -75.3 ppm (s, TFA of **5**).  $^{31}\text{P}\{^1\text{H}\}$  NMR (150.0 MHz, methylene chloride- $d_2$ , 253K)  $\delta$  141.2 (s,  $\text{P}(\text{OR})_3$ ), 119.6 ppm (s,  $(\text{RO})_3\text{PAuP}(\text{OR})_3^+$ ).

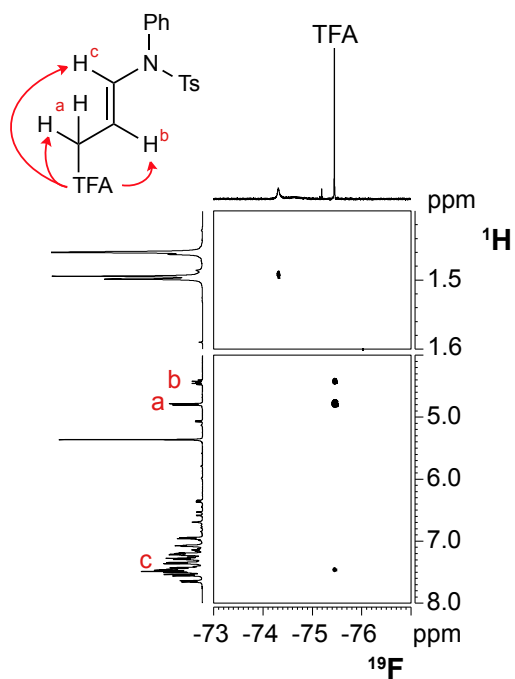


**Figure S8.**  $^1\text{H}$  NMR spectrum obtained after mixing **1TFA** with 1.2 equivalents of **3** in methylene chloride- $d_2$  in the presence of water traces ( $T = 253\text{K}$ ).



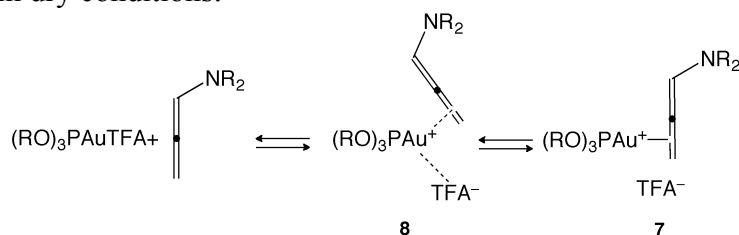


**Figure S9.** A section of the  $^1\text{H}$ ,  $^{31}\text{P}$  phase sensitive HMBC NMR spectrum obtained after mixing **1**TFA with 1.2 equivalents of **3** in methylene chloride- $d_2$  in the presence of water traces ( $T = 253\text{K}$ ).



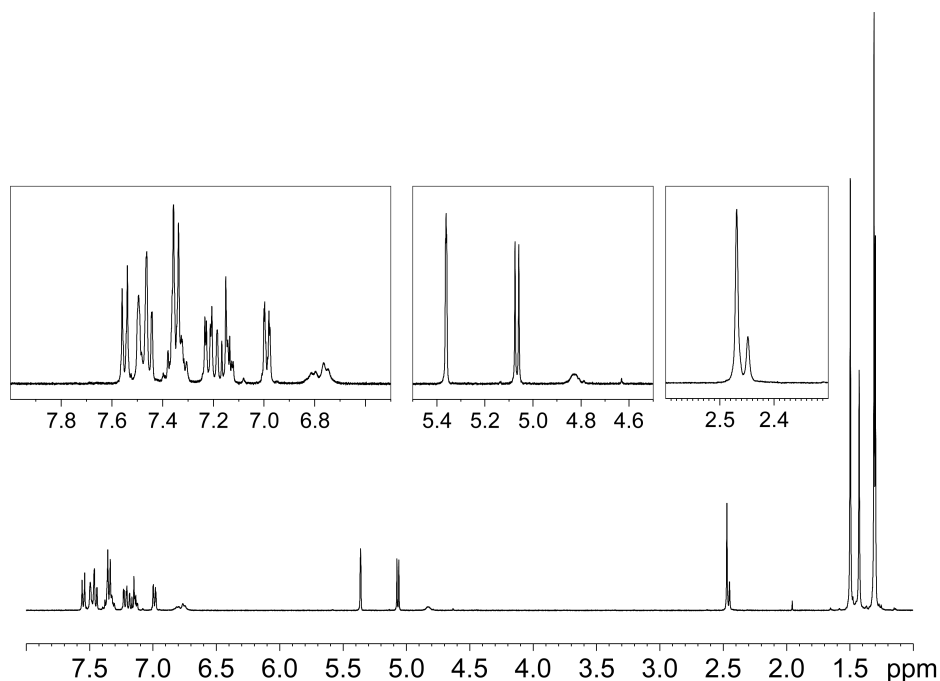
**Figure S10.**  $^{19}\text{F}$ ,  $^1\text{H}$  HOESY NMR spectrum obtained after mixing **1**TFA with 1.2 equivalents of **3** in methylene chloride- $d_2$  in the presence of water traces ( $T = 253\text{K}$ ). Red arrows denote NOEs.

b) Reactivity in dry conditions:

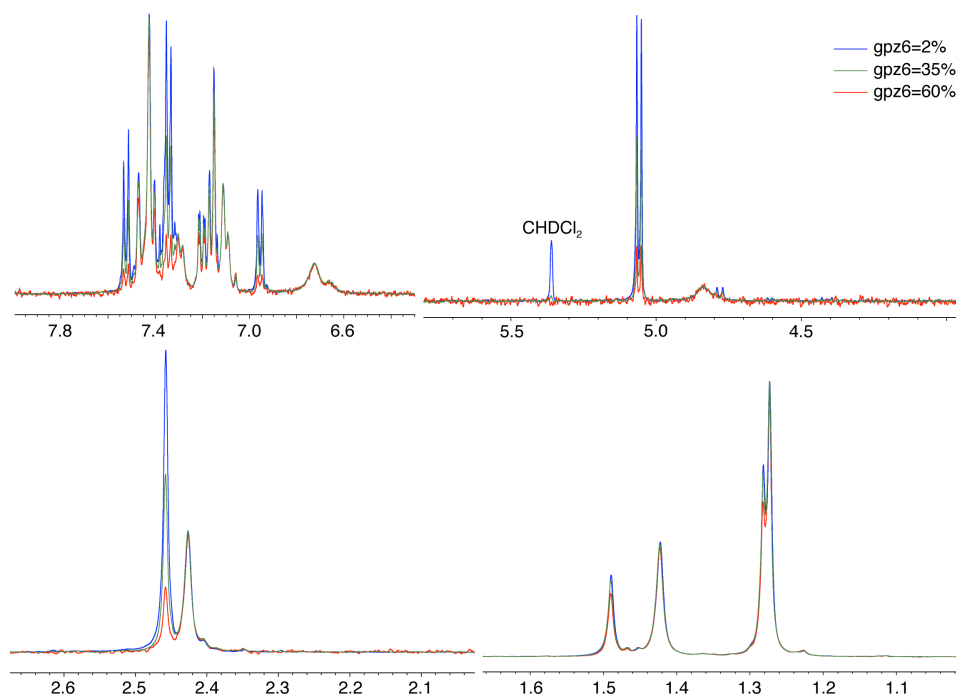


**Scheme S3.** Proposed reactivity between **1TFA** and **3** under rigorous dry conditions in methylene chloride at 233 K.

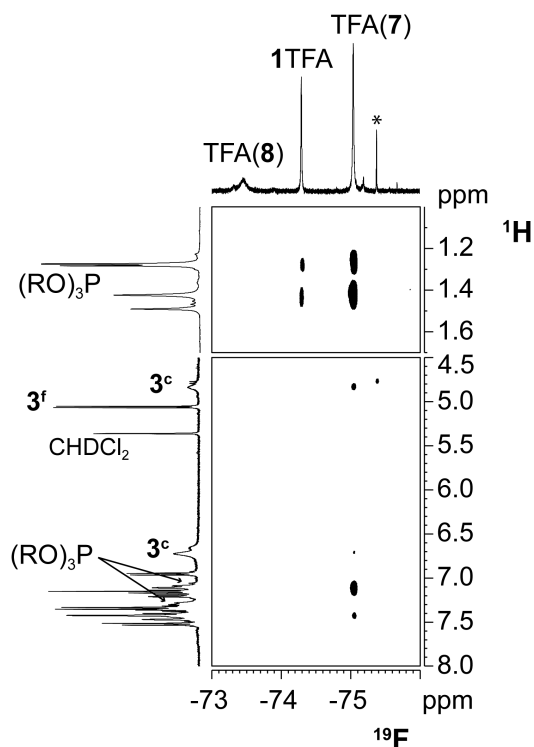
Selected  $^1\text{H}$  NMR resonances (400.13 MHz, methylene chloride- $d_2$ , 233K)  $\delta$  7.52 (d,  $J_{\text{H,H}}=8.3$  Hz, Ts of free **3**), 7.47 (m,  $(\text{RO})_3\text{P}$  of **1TFA**), 7.42 (m,  $(\text{RO})_3\text{P}$  of **7/8**), 7.34 (d,  $J_{\text{H,H}}=8.3$  Hz, Ts of free **3**), 7.29 (brd, Ts of coordinated **3**), 7.20 (m,  $(\text{RO})_3\text{P}$  of **1TFA+7/8**), 7.15 (t,  $J_{\text{H,H}}=6.3$  Hz, CH of free **3**), 6.72 (brd, Ph of coordinated **3**), 5.05 (d,  $J_{\text{H,H}}=6.3$  Hz,  $\text{CH}_2$  of free **3**), 4.83 (brd,  $\text{CH}_2$  of coordinated **3**), 2.45 (s, Ts of free **3**), 2.43 (s, Ts of coordinated **3**), 1.49 (s,  $(\text{RO})_3\text{P}$  of **1TFA**), 1.42 (s,  $(\text{RO})_3\text{P}$  of **7/8**), 1.28 (s,  $(\text{RO})_3\text{P}$  of **1TFA**), 1.27 ppm (s,  $(\text{RO})_3\text{P}$  of **7/8**). Selected  $^{13}\text{C}\{^1\text{H}\}$  NMR resonances (100.55 MHz, methylene chloride- $d_2$ , 233K)  $\delta$  129.8 (s, Ph of coordinated **3**), 125.2 (s,  $(\text{RO})_3\text{P}$  of **7/8**), 123.8 (s, Ts of coordinated **3**), 77.5 (s,  $\text{CH}_2$  of coordinated **3**), 31.2 (s,  $(\text{RO})_3\text{P}$  of **7/8**), 30.1 (s,  $(\text{RO})_3\text{P}$  of **7/8**), 21.8 ppm (s, Ts of coordinated **3**).  $^{19}\text{F}$  NMR (376.5 MHz, methylene chloride- $d_2$ , 233K)  $\delta$  -73.5 (brs, TFA **8**), -74.3 (s, **1TFA**), -75.0 ppm (s, TFA **7**).  $^{31}\text{P}\{^1\text{H}\}$  NMR (150.0 MHz, methylene chloride- $d_2$ , 233K)  $\delta$  139.6 (s,  $\text{P}(\text{OR})_3$  of **7**), 130.1 (s,  $\text{P}(\text{OR})_3$  of **8**), 87.8 ppm (s, **1TFA**).



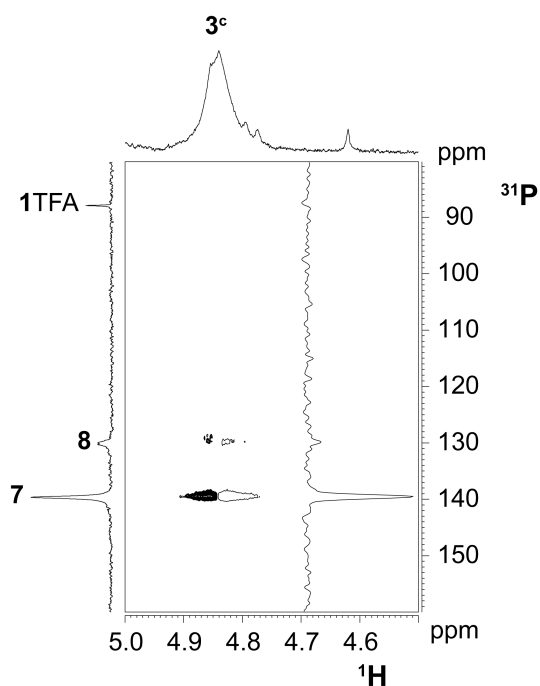
**Figure S11.**  $^1\text{H}$  NMR spectrum obtained after mixing **1TFA** with 1.5 equivalents of **3** in dry methylene chloride- $d_2$  at 233K showing the presence of both free and coordinated allene.



**Figure S12.** Four sections of  $^1\text{H}$  PGSE NMR spectra obtained with 3 different gradient strengths of a mixture containing **1TFA** with 1.5 equivalents of **3** in dry methylene chloride- $d_2$  at 233K. Intensities of spectra were scaled to show the similar diffusion behaviour between coordinated **3** and  $(\text{RO})_3\text{P}$  resonances at  $\delta_{\text{H}}=1.42$  and 1.27 ppm.

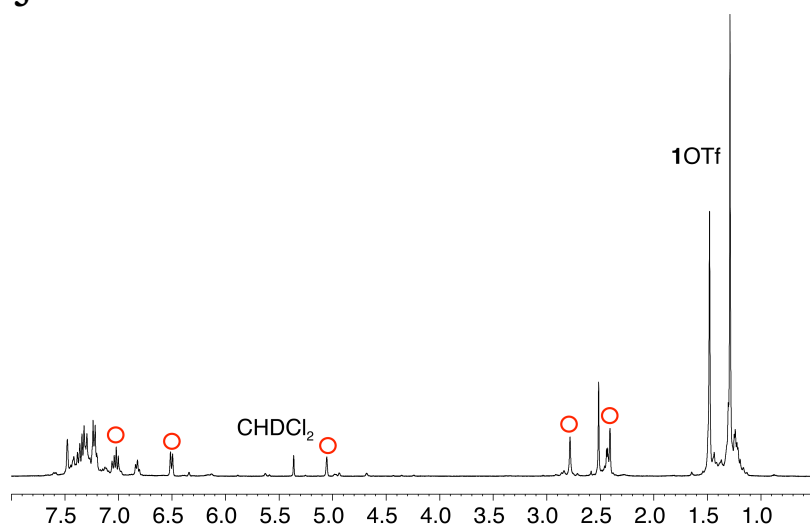


**Figure S13.**  $^{19}\text{F}$ ,  $^1\text{H}$  HOESY NMR spectrum of a mixture of **1TFA** and 1.5 equivalents of **3** in dry methylene chloride- $d_2$  at 233K. Asterisk denotes traces of **5** (see Scheme S2).



**Figure S14.** A section of the  $^1\text{H}$ ,  $^{31}\text{P}$  phase sensitive HMBC NMR spectrum of a mixture of **1TFA** and 8 equivalents of **3** in dry methylene chloride- $d_2$  at 233K.

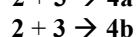
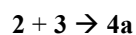
## 2.4 1OTf + 3



**Figure S15.**  $^1\text{H}$  NMR spectrum obtained upon mixing **1OTf** with 2 equivalents of **3** in dry methylene chloride- $d_2$ . Red circles denote main resonances of disubstituted cyclobutane obtained as side product.

### 3. In situ NMR catalytic experiments

Samples for kinetic NMR studies were prepared by loading the suitable amount of **2** and **3** within a screw cap NMR tube and by injecting 0.6 ml of catalyst solution in anhydrous methylene chloride- $d_2$ . The reaction was monitored by  $^1\text{H}$  and  $^{31}\text{P}\{^1\text{H}\}$  NMR spectroscopies over the period of 12 hours at 298 K. Concentration *versus* time plots were obtained from integration of NMR spectra and fitted by means of COPASI software package, by imposing a two independent second order reaction model as follows:

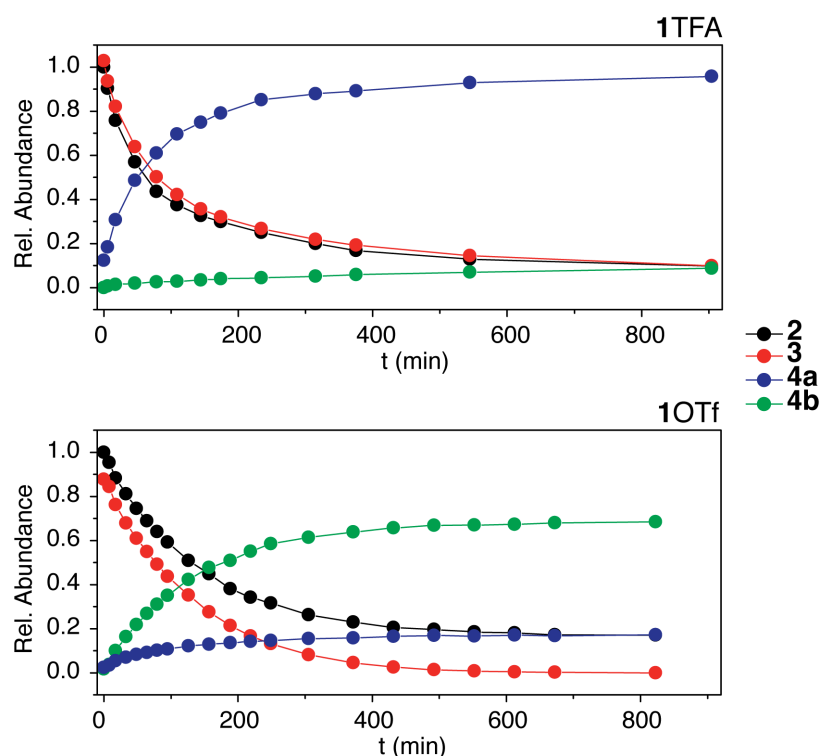


$$\frac{d[\mathbf{2}]}{dt} = -k_{4a}[\mathbf{2}][\mathbf{3}] - k_{4b}[\mathbf{2}][\mathbf{3}]$$

$$\frac{d[\mathbf{3}]}{dt} = -k_{4a}[\mathbf{2}][\mathbf{3}] - k_{4b}[\mathbf{2}][\mathbf{3}]$$

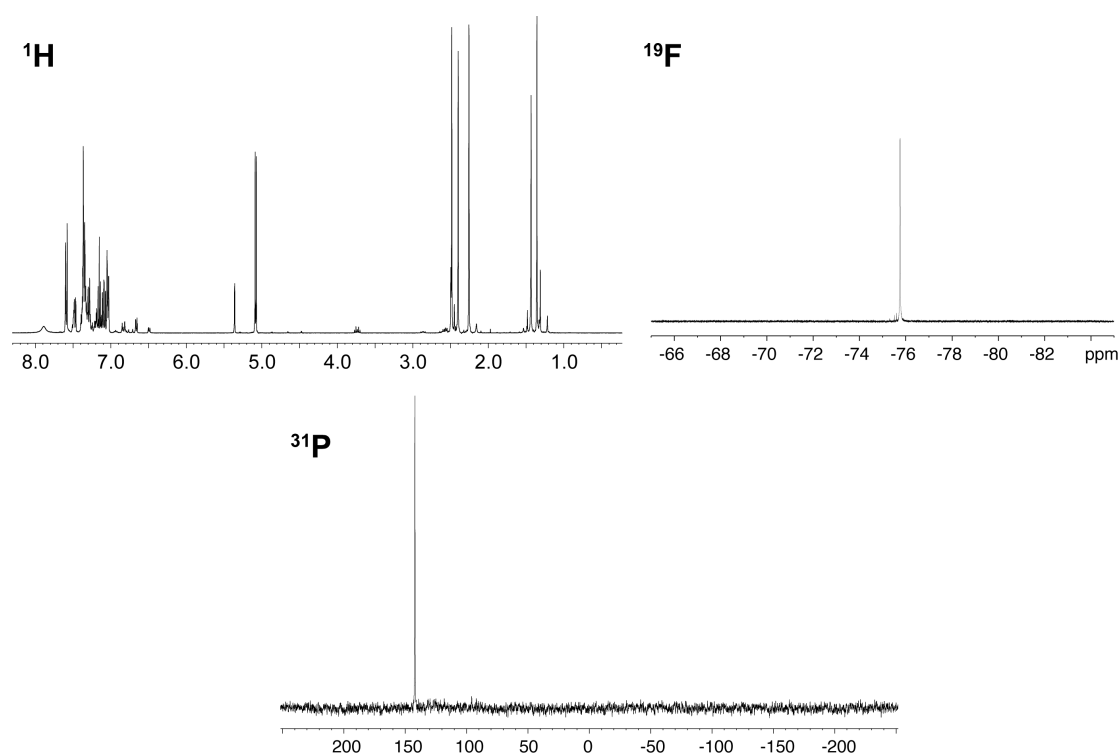
$$\frac{d[\mathbf{4a}]}{dt} = k_{4a}[\mathbf{2}][\mathbf{3}]$$

$$\frac{d[\mathbf{4b}]}{dt} = k_{4b}[\mathbf{2}][\mathbf{3}]$$

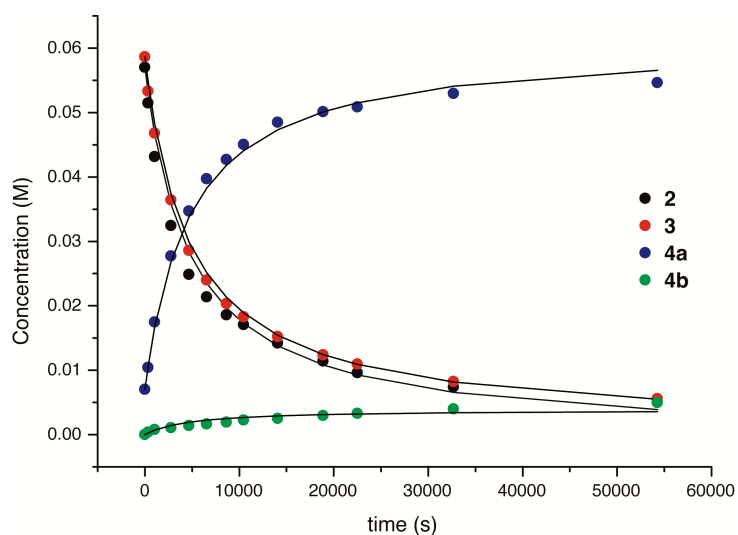


**Figure S16.** Kinetic profiles for dearomatization of **2** with **3** catalyzed by **1TFA** (top) and **1OTf** (bottom) in methylene chloride- $d_2$  at 298K.

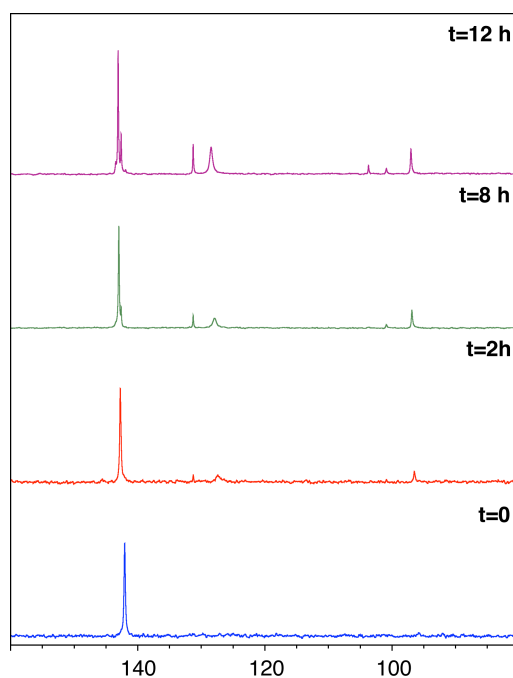
### 3.1 1TFA



**Figure S17.**  $^1\text{H}$ ,  $^{19}\text{F}$  and  $^{31}\text{P}\{^1\text{H}\}$  NMR spectra obtained upon mixing **2**, **3** and **1TFA** (8% mol) in anhydrous methylene chloride- $d_2$  at 298K.

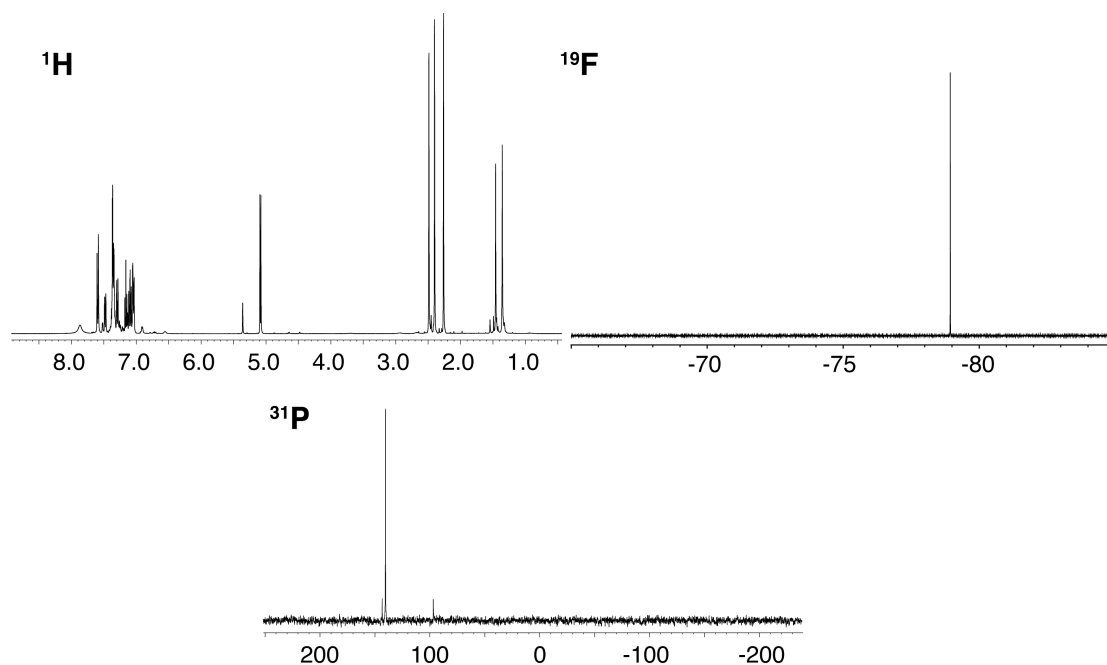


**Figure S18.** Concentration *versus* time plot for dearomatization reaction of **2** with **3** catalysed by **1TFA**. Mathematical fitting by imposing a two independent reaction model (black lines) gave a  $k_{4a}=3.4\times 10^{-3} \text{ M}^{-1} \text{ s}^{-1}$  and  $k_{4b}=2.5\times 10^{-4} \text{ M}^{-1} \text{ s}^{-1}$ .

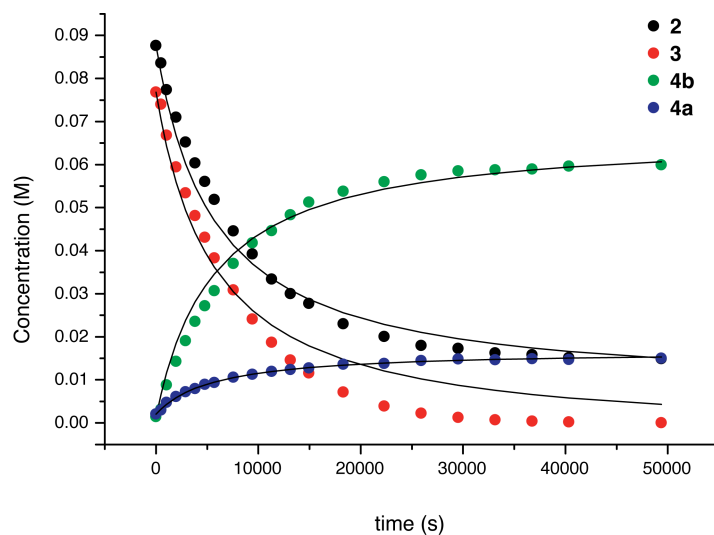


**Figure S19.** Evolution of  $^{31}\text{P}\{^1\text{H}\}$  NMR spectrum with time of the reaction mixture during dearomatization reaction of **2** with **3** catalysed by **1TfA**.

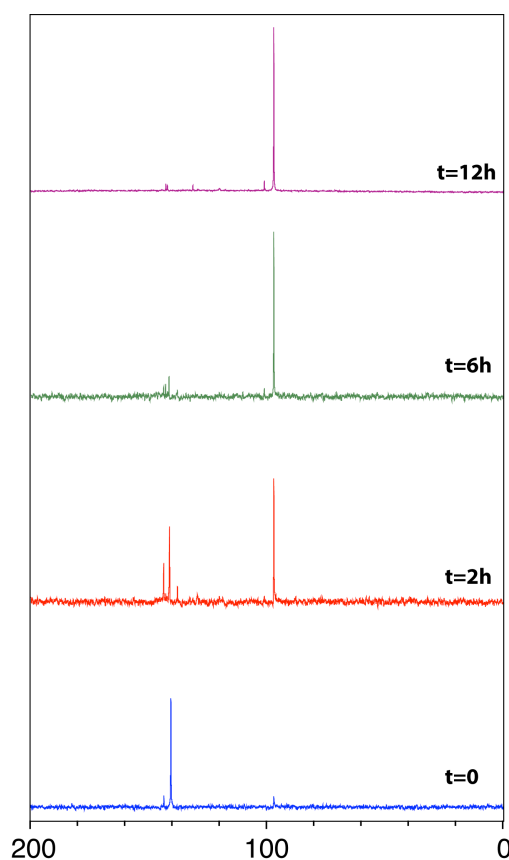
### 3.1 1OTf



**Figure S20.**  $^1\text{H}$ ,  $^{19}\text{F}$  and  $^{31}\text{P}\{^1\text{H}\}$  NMR spectra obtained upon mixing **2**, **3** and **1OTf** (5% mol) in anhydrous methylene chloride- $d_2$  at 298K.

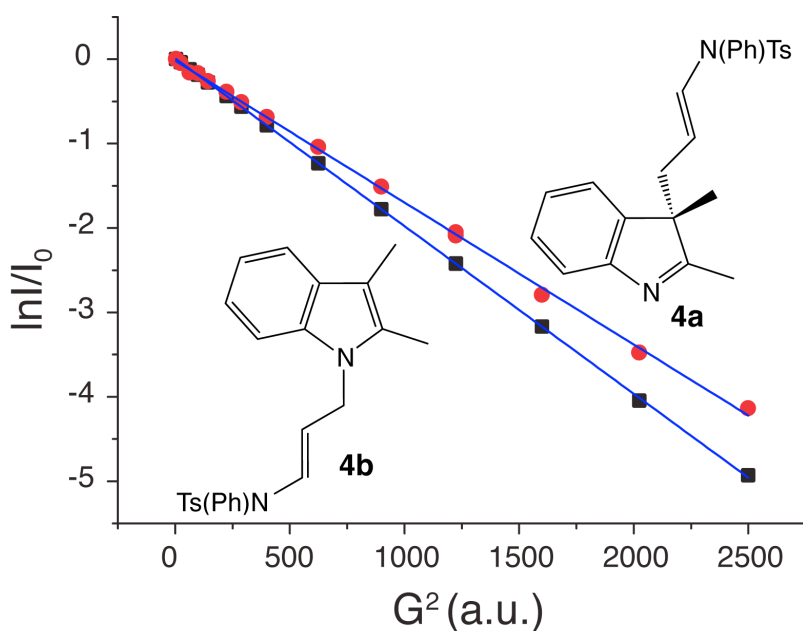


**Figure S21.** Concentration *versus* time plot for dearomatization reaction of **2** with **3** catalysed by 1OTf. Mathematical fitting by imposing a two independent reaction model (black lines) gave a  $k_{4a}=3.9\times 10^{-4} \text{ M}^{-1} \text{ s}^{-1}$  and  $k_{4b}=1.7\times 10^{-3} \text{ M}^{-1} \text{ s}^{-1}$ .



**Figure S22.** Evolution of  $^{31}\text{P}\{^1\text{H}\}$  NMR spectrum with time of the reaction mixture during dearomatization reaction of **2** with **3** catalysed by 1OTf.





**Figure S23.**  $\ln I/I_0$  versus  $G^2$  plot obtained by means of  $^1\text{H}$  PGSE NMR experiments on the reaction mixture deriving from dearomatization of **2** catalysed by **1OTf**.

PGSE NMR CALCULATIONS:

$$\frac{\text{slope}(4b)}{\text{slope}(4a)} = \frac{1.99 \cdot 10^{-3}}{1.68 \cdot 10^{-3}} = 1.185$$

Given that the slope ratio is directly proportional to the self-diffusion coefficients ratio and inversely proportional to the hydrodynamic radii ratio, it can be derived that

$$\frac{D_t(4b)}{D_t(4a)} = \frac{r_H(4a)}{r_H(4b)} = 1.185$$

Therefore, the experimental **4a/4b** hydrodynamic volume ratio is  $(1.185)^3 = 1.662$

Assuming the Van der Waals volumes of **4a** and **1OTf** equal to 370 and 730 Å<sup>3</sup>, respectively, and considering that, at the end of the reaction, **3** equivalents of **4a** are present, if all (RO)<sub>3</sub>PAu<sup>+</sup> cation is bound to **4a** it should be observed that:

$$V_{4a}^* = (x_{\text{free}} V_{4a}) + ((1 - x_{\text{free}}) V_{\text{Au-4a}}) = (0.67 \cdot 370) + (0.33 \cdot 1100) = 610.9 \text{ Å}^3$$

In perfect agreement with the experimental data,  $\frac{V_{4a}^*}{V_{4b}} = \frac{610.9}{370} = 1.651$

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